



# Genetic Determinants of Circulating Glycine Levels and Risk of Coronary Artery Disease

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**Background**—Recent studies have revealed sexually dimorphic associations between the carbamoyl-phosphate synthase 1 locus, intermediates of the metabolic pathway leading from choline to urea, and risk of coronary artery disease (CAD) in women. Based on evidence from the literature, the atheroprotective association with carbamoyl-phosphate synthase 1 could be mediated by the strong genetic effect of this locus on increased circulating glycine levels.

**Methods and Results**—We sought to identify additional genetic determinants of circulating glycine levels by carrying out a meta-analysis of genome-wide association study data in up to 30 118 subjects of European ancestry. Mendelian randomization and other analytical approaches were used to determine whether glycine-associated variants were associated with CAD and traditional risk factors. Twelve loci were significantly associated with circulating glycine levels, 7 of which were not previously known to be involved in glycine metabolism (*ACADM*, *PHGDH*, *COX18-ADAMTS3*, *PSPH*, *TRIB1*, *PTPRD*, and *ABO*). Glycine-raising alleles at several loci individually exhibited directionally consistent associations with decreased risk of CAD. However, these effects could not be attributed directly to glycine because of associations with other CAD-related traits. By comparison, genetic models that only included the 2 variants directly involved in glycine degradation and for which there were no other pleiotropic associations were not associated with risk of CAD or blood pressure, lipid levels, and obesity-related traits.

**Conclusions**—These results provide additional insight into the genetic architecture of glycine metabolism, but do not yield conclusive evidence for a causal relationship between circulating levels of this amino acid and risk of CAD in humans. (*J Am Heart Assoc.* 2019;8:e011922. DOI: 10.1161/JAHA.119.011922.)

**Key Words:** causality • coronary artery disease • genome-wide association study • glycine • Mendelian randomization • meta-analysis

Metabolites derived from gut microbiome and hepatic-mediated metabolism of dietary choline and L-carnitine, such as trimethylamine N-oxide and betaine, have recently been shown to be proatherogenic in mice and novel biomarkers of coronary artery disease (CAD) risk in humans.<sup>1–3</sup> In searching for genetic determinants of these metabolites, we identified sexually dimorphic associations between the carbamoyl-

phosphate synthase 1 (*CPS1*) locus and not only plasma trimethylamine N-oxide and betaine levels, but also other intermediates in the metabolic pathway leading from choline to urea.<sup>4</sup> We and others further noted that, of the various other biomarkers/metabolites that had previously been linked to *CPS1*,<sup>5–14</sup> the strongest effect size and most significant association was with circulating glycine levels in women.<sup>4,15–17</sup> Most important, the lead *CPS1* variant also exhibited a strikingly significant female-specific association with decreased risk of CAD.<sup>4</sup> However, the direction of the associations between *CPS1* and the various biomarkers and metabolites was opposite to what would be expected for a variant that decreased risk of CAD.

One explanation for the protective association of *CPS1* with CAD could be the strong genetic effect of this locus on increased circulating glycine levels.<sup>4</sup> For example, previous in vitro and in vivo studies have shown that glycine reduces inflammation and oxidative stress in endothelial cells, activated macrophages, and other leukocytes.<sup>18–22</sup> Furthermore, platelet

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Accompanying Data S1, Figures S1 through S5, and Tables S1 through S5 are available at <https://www.ahajournals.org/doi/suppl/10.1161/JAHA.119.011922>

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## Clinical Perspective

### What Is New?

- The study identifies 12 genetic determinants of circulating glycine levels, 7 of which are novel and not previously known to be involved in the metabolism of this amino acid.
- Biological mechanisms for half of the loci associated with circulating glycine levels are not directly evident.

### What Are the Clinical Implications?

- Although findings from this study provide additional insight into the genetic architecture of glycine metabolism, they do not yield conclusive evidence for a causal relationship between circulating levels of this amino acid and coronary artery disease in humans.

aggregation of both human and rodent platelets can be prevented by glycine in a dose-dependent manner through mechanisms involving the glycine receptor.<sup>23</sup> Interestingly, the same glycine-raising *CPS1* variant has been associated with reduced platelet counts.<sup>24</sup> Alternatively, glycine has been reported to have antihypertensive effects in mice and humans.<sup>25</sup> A recent epidemiological study also demonstrated an inverse relationship between plasma glycine levels and risk of an acute myocardial infarction.<sup>26</sup> Taken together, these observations support the concept that glycine could have atheroprotective properties, but direct evidence for a causal relationship between this amino acid and risk of CAD is lacking.

In the present study, we used a meta-analysis approach with genome-wide association study (GWAS) data to identify additional genetics determinants of circulating glycine levels. The identified loci were then used to investigate the possible causal association between circulating glycine levels and risk of CAD and traditional risk factors. In total, 12 loci were identified for circulating glycine levels, 7 of which were novel and not previously known to be involved in glycine metabolism. However, various analytical approaches with glycine-raising alleles at these loci did not provide conclusive evidence for a causal relationship between circulating glycine and risk of CAD in humans.

## Methods

The statistical methods used in this study will be made available to other researchers for purposes of reproducing the results or replicating the analyses. The summary statistics of the meta-analysis for circulating glycine levels will be made available through the NHGRI-EBI Catalog of published GWASs (<https://www.ebi.ac.uk/gwas/download/s/summary-statistics/>).

## Study Populations

The present analyses included 30 118 subjects of European ancestry from the GeneBank (GB),<sup>4</sup> FINRISK 1997 and 2007 (FR97 and FR07),<sup>27</sup> YFS (Cardiovascular Risk in Young Finns Study),<sup>28</sup> NFBC1966 and NFBC1986 (Northern Finland Birth Cohort),<sup>29</sup> and METSIM (Metabolic Syndrome in Men)<sup>30</sup> studies. Details of subject recruitment and genotyping methodology for each cohort are provided in Data S1. For each cohort, written informed consent was obtained from all participants before being enrolled, and the studies were approved by the institutional review boards of the participating institutions. The present analysis was approved by the institutional review board of USC Keck School of Medicine.

## Measurement of Circulating Glycine Levels

Glycine levels were quantified using stable isotope dilution high-performance liquid chromatography with online electrospray ionization tandem mass spectrometry in the GB study<sup>4</sup> and by quantitative high-throughput NMR in the FR97, FR07, YFS NFBC66, NFBC86, and METSIM cohorts.<sup>31,32</sup>

## Data Harmonization and GWAS Analyses

Circulating glycine levels were first regressed on study-specific covariates chosen by the investigators of each cohort. These included age and sex in GeneBank; age, sex, and time from last meal in FR97, FR07, YFS NFBC66, and NFBC86<sup>31</sup>; and age, age,<sup>2</sup> and body mass index in METSIM.<sup>32</sup> Inverse rank-based normal transformations were carried out on the residuals after adjustment for covariates and used as the outcome in GWAS analyses by linear regression in each study.

## Meta-Analysis for Circulating Glycine Levels

We performed a fixed-effects meta-analysis for circulating glycine levels with 7 487 927 SNPs that were imputed using 1000 Genomes Project data and that were common to all data sets. This analysis was carried out assuming an additive model and after controlling for population structure within each study, as implemented in GWAMA (Genome-Wide Association Meta-Analysis) software.<sup>33</sup> In addition to a combined meta-analysis with all subjects, we also carried out a sex-stratified fixed-effects meta-analysis. The genome-wide threshold for significant association was set at  $P=5.0 \times 10^{-8}$ . A locus was defined as novel if the lead single-nucleotide polymorphism (SNP) was in weak or no linkage disequilibrium ( $r^2 \leq 0.1$ ) with variants at genome-wide significant loci previously reported for circulating glycine levels. Manhattan and quantile-quantile plots were constructed using the “qqman” package in R (R Foundation for Statistical Computing, Vienna, Austria).<sup>34</sup> To examine whether

all novel loci identified in our meta-analysis were also significantly associated with other traits (phenome-wide association studies), we used publicly available databases, such as PhenoScanner,<sup>35</sup> the UCSC Genome Browser (<https://genome.ucsc.edu/>), and the GWAS Catalog (<https://www.ebi.ac.uk/gwas/home>). The significance threshold for phenome-wide association studies analyses was set to  $P=5.0 \times 10^{-8}$  with a linkage disequilibrium cut off of  $r^2 \geq 0.8$  for proxy SNPs.

### Proportion of Phenotypic Variance Explained

The proportion of variation in glycine levels explained by the identified variants was estimated using SumHer software.<sup>36</sup> SNP heritability was calculated using a weighted linkage disequilibrium adjusted kinships model with the 12 glycine-associated SNPs. 1000 Genomes Project–based imputed genotypes in  $\approx 4500$  subjects of European ancestry from the GB cohort were used as a reference panel for linkage disequilibrium ( $r^2$ ) for these estimates.

### Analysis of Variants With Risk of CAD and Traditional Risk Factors

Publicly available summary results from large-scale GWAS in subjects of European ancestry<sup>37–39</sup> were used to determine whether glycine-associated variants were associated with risk of CAD and various lipid-, metabolic-, and blood-pressure–related risk factors. Specifically, we tested associations using 3 analytical strategies with 4 genetic models that were based on various nested combinations of the 12 identified variants. Genetic model 1 included all 12 loci identified for glycine; model 2 was designed to specifically test only the 7 novel loci (*ACADM*, *PHGDH*, *COX18-ADAMTS3*, *PSPH*, *TRIB1*, *PTPRD*, and *ABO*); and model 3 included only the 4 loci known to be related to glycine metabolism (*PSPH*, *PHGDH*, *GLDC*, and *GCSH*). Model 4 was the most restrictive and included only the 2 glycine-associated loci that are known to be directly involved in the catabolism of glycine through the glycine cleavage system (*GLDC* and *GCSH*) and that did not exhibit pleiotropic effects with other traits or metabolites. In the first analytical approach, the average/overall association of CAD and its risk factors with glycine-raising alleles in the 4 genetic models were evaluated by meta-analysis, as implemented in the “meta” R package (<https://cran.r-project.org/web/packages/meta/index.html>). In the second approach, we generated genetic risk scores (GRS) with the identified variants for the same 4 genetic models to evaluate the cumulative joint effects of glycine-raising alleles. Additive multi-SNP GRS associations were estimated using the *grs.summary* function of the “gtx: Genetic ToolboX” R package (<https://cran.r-project.org/web/packages/gtx>). This approach approximates the regression of an intermediate trait or biomarker

onto a GRS, which is based on the weighted sum of the single SNP coefficients derived from the association summary statistics.<sup>40</sup> For the third strategy, we carried out weighted median and inverse variance weighted Mendelian randomization (MR) analyses with the 4 genetic models, as implemented in the “TwoSampleMR” R package.<sup>41</sup> Because the weighted median MR method requires 3 or more variants, only the inverse variance weighted MR test was used for determining association of the 2 SNPs in model 4 with CAD and traditional risk factors.

## Results

### GWAS for Circulating Glycine Levels

To identify novel loci for circulating glycine levels, we carried out a meta-analysis of GWAS summary-level data with 7 487 927 genotyped and imputed SNPs in 30 118 subjects of European ancestry. Table 1 shows the characteristics of the study cohorts and data sets used for these analyses. A GWAS was carried out for circulating glycine levels in each cohort, followed by a fixed-effects meta-analysis. The genomic control factor ( $\lambda$ ) in GB I (0.995), GB II (0.989), and the combination of the FR97, FR07, YFS NFBC66, and NFBC86 cohorts (1.039), and METSIM (1.014) were small or modest, thus decreasing the likelihood of identifying spurious associations attributed to population stratification (Figure S1). To further account for this potential confounder, we also applied genomic control to each study before the meta-analysis. In total, 4934 variants distributed across 12 loci were associated with circulating glycine levels at the genome-wide significance threshold ( $P=5.0 \times 10^{-8}$ ; Figure 1, Table 2, and Table S1). Seven of these loci (*ACADM*, *PHGDH*, *COX18-ADAMTS3*, *PSPH*, *TRIB1*, *PTPRD*, and *ABO*) were novel and identified as being associated with circulating glycine levels for the first time herein (Figure 1, Table 2, and Figure S2). The other 5 loci (*CPS1*, *ALDH1L1*, *PPP1R3B-LOC157273*, *GLDC*, and *GCSH*) have previously been reported for circulating glycine levels, but the association signals became more significant in our meta-analysis because of increased sample size (Figure 1, Table 2, and Figure S2). Overall, the 12 identified loci explained  $\approx 15\%$  of the variation in circulating glycine levels.

Based on previous observations that the *CPS1* locus exhibited a pattern of sexually dimorphic associations with glycine, various other metabolites, and risk of CAD,<sup>4,17</sup> we also carried out meta-analyses in men and women separately. Five and 9 regions were significantly associated with circulating glycine levels in females and males, respectively (Figures S3 and S4), all of which were also observed in the combined GWAS analysis with all subjects (Figure 1). With the exception of the previously observed stronger association

**Table 1.** Description of Cohorts Used in Meta-Analysis for Circulating Glycine Levels

Cohort	No. of SNPs	N (Male/Female)	Metabolomics Platform
GB I	8 986 545	391 (195/196)	HPLC-MS
GB II	8 986 545	885 (602/283)	HILIC-MS
FR97	11 512 433	6631 (3198/3433)	NMR
FR07	11 512 433	4124 (1860/2264)	NMR
YFS	11 512 433	1947 (1052/895)	NMR
NFBC66	11 512 433	4483 (2152/2331)	NMR
NFBC86	11 512 433	3112 (1508/1604)	NMR
METSIM	16 888 882	8545 (8545/0)	NMR

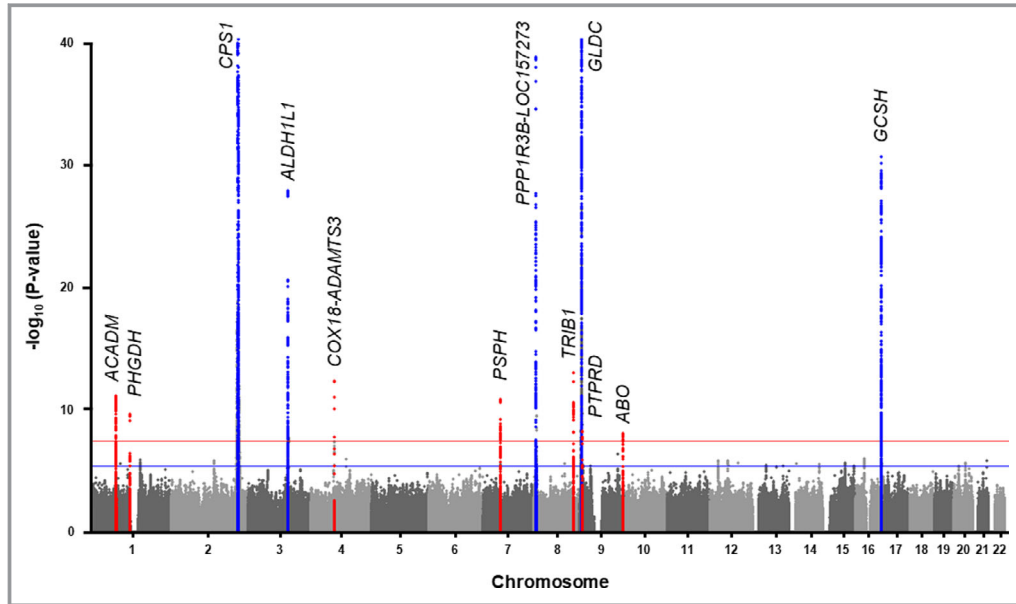
FR97 and FR07 indicates FINRISK; GB, GeneBank; HPLC-MS, high-performance liquid chromatography with mass spectrometry; METSIM, METabolic Syndrome In Men Study; NFBC, Northern Finland Birth Cohort; NMR, nuclear magnetic resonance; SNP, single-nucleotide polymorphism; YFS, Cardiovascular Risk in Young Finns.

signal for glycine levels at the *CPS1* locus in women ( $\beta=0.572$ ;  $P<1.0\times 10^{-300}$ ) compared with men ( $\beta=0.322$ ;  $P=5.9\times 10^{-189}$ ), the effect sizes at the remaining 11 loci were similar in males and females with no significant evidence for heterogeneity (Table S2 and Figure S5). We next carried out a phenome-wide association studies analysis based on publicly available data to determine whether any of the loci for glycine

were associated with other traits. Six of the 12 loci (*ACADM*, *CPS1*, *ALDH1L1*, *PPP1R3B-LOC157273*, *TRIB1*, and *ABO*) exhibited pleiotropic associations with blood cell counts or lipid levels, some of which were even more significant than the association signals for glycine (Table S3). Two other loci (*PSPH* and *PHGDH*) had also been associated with serine and homocysteine levels, which are metabolites related to glycine metabolism (Table S3). However, no genome-wide significant associations have previously been reported for the 4 remaining loci (*COX18-ADAMTS3*, *GLDC*, *PTPRD*, and *GCSH*).

### Association of Loci for Circulating Glycine Levels With CAD and Traditional Risk Factors

We next sought to evaluate association of loci for glycine levels with risk of CAD and traditional risk factors. Of the 12 regions identified, glycine-raising alleles of the lead variants at the *CPS1*, *PSPH*, *TRIB1*, and *ABO* loci individually yielded directionally consistent associations with decreased risk of CAD at the Bonferroni-corrected threshold of  $P=4.2\times 10^{-3}$  for testing 12 loci (0.05/12; Table S4). We next tested 4 genetic models based on various nested combinations of the 12 glycine loci for association with risk of CAD using 3 analytical strategies (details provided in Methods). Consistent with the individual SNP results, meta-analysis or GRS-based joint SNP



**Figure 1.** Results of GWAS meta-analysis for circulating glycine levels. The Manhattan plot shows 7 novel significantly associated loci for circulating glycine levels (red dots) identified through meta-analyses of GWAS data from 30 118 subjects in the GeneBank, FR97, FR07, YFS, NFBC66, NFBC86, and METSIM cohorts. The 5 previously known loci are indicated by blue dots and all increased in significance in the meta-analysis. Genome-wide thresholds for significant ( $P=5.0\times 10^{-8}$ ) and suggestive ( $P=5.0\times 10^{-6}$ ) association are indicated by the horizontal red and dark blue lines, respectively.  $P$  values are truncated at  $-\log_{10}(P)=40$ . FR97 and FR07 indicates FINRISK; GWAS, genome-wide association study; METSIM, METabolic Syndrome In Men Study; NFBC, Northern Finland Birth Cohort; YFS, Cardiovascular Risk in Young Finns.

**Table 2.** Meta-Analysis Identifies 12 Loci Significantly Associated With Circulating Glycine Levels

Locus (Nearest Gene(s))*	Lead SNP	Position (bp) <sup>†</sup>	Effect/Other Allele <sup>‡</sup>	EAF	β (SE)	P Value	Direction <sup>§</sup>
1p31.1 ( <i>ACADM</i> )	rs12126607	76 217 097	A/G	0.27	0.06 (0.01)	$1.1 \times 10^{-11}$	+−++
1p12 ( <i>PHGDH</i> )	rs478093	120 255 126	G/A	0.67	0.06 (0.01)	$3.5 \times 10^{-10}$	++++
2q34 ( <i>CPS1</i> )	rs1047891	211 540 507	A/C	0.34	0.43 (0.01)	$<1.0 \times 10^{-300}$	++++
3q21.3 ( <i>ALDH1L1</i> )	rs2364368	125 905 080	T/A	0.40	0.09 (0.01)	$2.2 \times 10^{-28}$	++++
4q13.3 ( <i>COX18-ADAMTS3</i> )	rs143424675	73 749 419	T/C	0.03	0.19 (0.03)	$7.3 \times 10^{-13}$	+−++
7p11.2 ( <i>PSPH</i> )	rs6955423	56 099 352	A/G	0.81	0.07 (0.01)	$2.3 \times 10^{-11}$	++++
8p23.1 ( <i>PPP1R3B-LOC157273</i> )	rs2126263	9 181 611	G/A	0.15	0.16 (0.01)	$5.8 \times 10^{-44}$	++++
8q24.13 ( <i>TRIB1</i> )	rs28601761	126 500 031	G/C	0.41	0.06 (0.01)	$1.6 \times 10^{-13}$	+−++
9p24.1 ( <i>GLDC</i> )	rs71503800	6 102 648	T/C	0.05	0.46 (0.02)	$8.5 \times 10^{-121}$	−++
9p24.1 ( <i>PTPRD</i> )	rs12003835	8 424 378	T/G	0.03	0.15 (0.03)	$8.2 \times 10^{-9}$	−++
9q34.2 ( <i>ABO</i> ) <sup>  </sup>	rs492488	136 144 960	G/A	0.55	0.05 (0.01)	$1.2 \times 10^{-8}$	−+++
16q23.2 ( <i>GCSH</i> )	rs11860711	81 132 493	C/T	0.80	0.12 (0.01)	$4.2 \times 10^{-31}$	++++

EAF indicates effect allele frequency; SNP, single-nucleotide polymorphism.

\*Novel loci identified in this study are highlighted in gray.

<sup>†</sup>SNP base pair (bp) positions are given according to NCBI build 37 of the reference human genome sequence (hg19).

<sup>‡</sup>Effect allele refers to allele that increases glycine levels.

<sup>§</sup>Direction of betas in the 4 data sets used for meta-analysis are in the following order: GB I, GB II, Combination of FR97-FR07-YFS-NFBC66-NFBC86, and METSIM.

<sup>||</sup>N=27 006 for chromosome 9q34.2 locus.

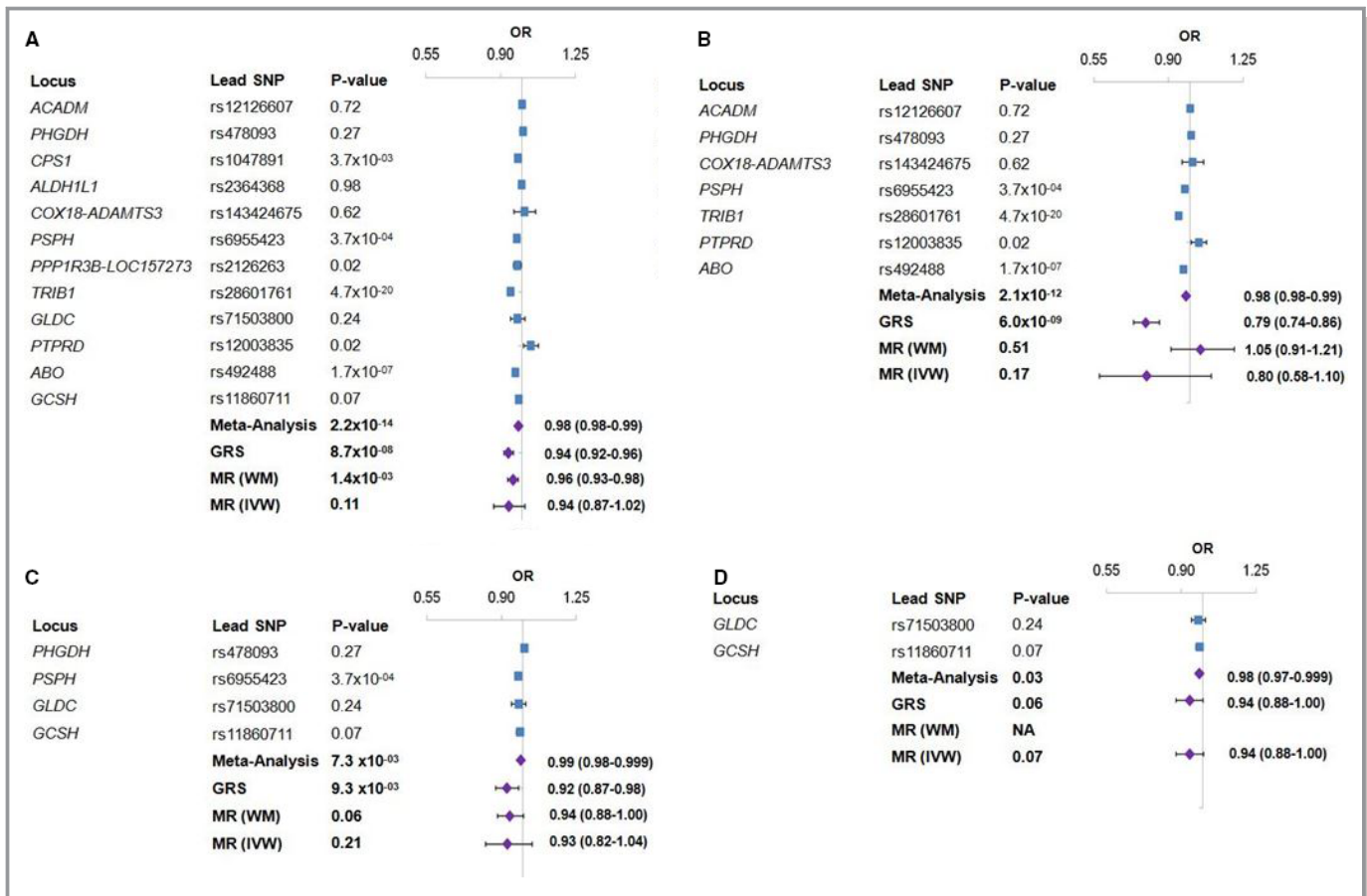
effects analysis of glycine-raising alleles in all 4 genetic models yielded modest, but significant, associations (odds ratios,  $\approx 0.98$ ) with decreased risk of CAD (Figure 2). By comparison, weighted median and inverse variance weighted MR tests yielded much weaker or no evidence for a protective association of glycine-raising alleles with CAD, including the most restrictive model constructed with only variants at the 2 glycine cleavage system loci (Figure 2). We next evaluated whether loci for glycine were associated with blood pressure, lipid levels, and obesity-related traits using the same analytical strategies. Glycine-raising alleles at several loci (*CPS1*, *PPP1R3B-LOC157273*, *TRIB1*, and *ABO*) individually exhibited highly significant associations with decreased blood pressure and lipid levels (Table S5). The meta-analysis and GRS-based joint SNP effects analysis also provided evidence for similar associations with blood pressure and lipid levels, although these were only observed for the genetic models that included either all 12 glycine-associated loci or the 7 novel loci. However, the 2 MR analyses provided no evidence that glycine-raising alleles were causally associated with any of the selected traditional risk factors (Table S5).

## Discussion

In the present study, we used a meta-analysis approach to identify 7 novel genomic regions associated with circulating glycine levels and strengthen the association signals at 5 previously known loci. Among all 12 loci, *CPS1* and *GLDC* were the most strongly associated with glycine levels, with

variants at the remaining 10 loci having anywhere between  $\approx 60\%$  and  $90\%$  lower effect sizes. Furthermore, sex-stratified analyses confirmed the strong effect of *CPS1* on glycine levels in women compared with men, but did not reveal sexually dimorphic associations with any of the remaining 11 loci. Follow-up analyses with the identified loci also yielded evidence that glycine could be causally associated with risk of CAD, although the biological mechanism(s) through which this effect occurs remains to be determined.

Based on what is known about amino acid metabolism, plausible biological links could be inferred between several of the newly identified loci and glycine levels. For example, *PHGDH* and *PSPH* encode phosphoglycerate dehydrogenase and phosphoserine phosphatase, which catalyze the first and last reactions, respectively, in the 3-step process leading to the synthesis of serine from 3-phosphoglycerate.<sup>42</sup> Although the *PHGDH* and *PSPH* loci have both been strongly associated with circulating serine or homocysteine levels,<sup>8,15,16,43–47</sup> they were not known to be associated with glycine levels before the results of our meta-analysis. Interestingly, serine can serve as a substrate for the synthesis of glycine in a reversible reaction catalyzed by *SHMT*,<sup>48</sup> and glycine levels have been reported to be lower in humans deficient for *PHGDH* or *PSPH*.<sup>49–52</sup> With respect to our results, the lead variant at *PHGDH* has yielded several highly significant (*P* values ranging from  $\approx 1.0 \times 10^{-10}$  to  $1.0 \times 10^{-34}$ ) *cis* expression quantitative trait loci where the glycine-raising allele of rs478093 (G) increases *PHGDH* mRNA levels.<sup>35</sup> This would presumably lead to increased production of serine and,



**Figure 2.** Association of loci identified for circulating glycine levels with risk of CAD. Individual associations between glycine-raising alleles at each locus and risk of CAD are shown by blue squares in the forest plots. Purple diamonds indicate combined associations based on meta-analysis, joint SNP effects with a genetic risk score (GRS), and weighted median (WM) or inverse variance weighted (IVW) Mendelian randomization (MR) test. Model 1 included all 12 glycine-associated loci (A), model 2 included the 7 novel loci for glycine in this study (B), model 3 included the 4 loci known to be involved in glycine metabolism (C), and model 4 was constructed with only the 2 loci directly involved in the catabolism of glycine through the glycine cleavage complex (D). CAS indicates coronary artery disease; OR, odds ratio; SNP, single-nucleotide polymorphism.

by extension, glycine, thus providing a directionally consistent molecular mechanism for the observed association of the *PHGDH* locus with circulating glycine levels. However, even when taking into account previously identified associations at loci harboring enzymes involved in either glycine catabolism (*GLDC*, *GCSH*)<sup>53,54</sup> or downstream detoxification through the urea cycle (*CPS1*),<sup>55,56</sup> biological mechanisms for half of the loci associated with circulating glycine levels are not directly evident.

A primary goal of our study was to test whether glycine is a causal and protective biomarker of CAD risk. To address this question, we used the results of large GWAS meta-analyses to determine whether loci identified for glycine levels were associated with CAD and traditional risk factors. Glycine-raising alleles at 3 of the 7 novel loci (*PSPH*, *TRIB1*, and *ABO*) were individually associated with decreased risk of CAD at the Bonferroni-corrected significance threshold ( $P=4.2 \times 10^{-3}$ ), of

which *TRIB1* and *ABO* had been identified as CAD susceptibility loci in previous GWASs.<sup>37,57,58</sup> Rather than glycine levels, it is likely that association of *TRIB1* and *ABO* with CAD is attributed to their stronger effect sizes on lipid levels and hematological parameters<sup>12,59,60</sup> and, in the case of *ABO*, numerous other CAD-relevant traits.<sup>35</sup> When all 12 loci or only the 7 novel loci were considered in combination, the meta-analyses and joint SNP effects analyses also revealed association of glycine-raising alleles with decreased risk of CAD. Because several of the loci included in these analyses (*CPS1*, *PSPH*, *PPP1R3B-LOC157273*, *TRIB1*, and *ABO*) exhibited associations with other CAD-related traits, either individually or in various combinations, it was not possible based on these results alone to conclude that glycine is the causal biomarker driving the association of these loci with CAD. Therefore, we assessed causality more directly with 2 different MR tests, which provided little to no evidence that

glycine-raising alleles were associated with risk of CAD or lipid levels, blood pressure, and obesity-related traits. In this regard, the results of MR tests with the most restrictive genetic model that included only the 2 loci directly involved in glycine degradation (*GLDC* and *GCSH*) are particularly relevant. For example, no CAD-related traits, aside from glycine levels, are known to be associated with the *GLDC* and *GCSH* loci, thus satisfying the lack of pleiotropy as 1 of the major assumptions in MR analysis. Moreover, the glycine-raising alleles of rs71503800 at the *GLDC* locus and rs1047891 at the *CPS1* locus have nearly equivalent effect sizes on circulating glycine levels. However, none of the analyses with rs71503800 at the *GLDC* locus yielded evidence for association of this variant with risk of CAD or traditional risk factors. Taken together, we conclude that evidence for a causal relationship between circulating glycine and risk of CAD is relatively weak and requires additional studies.

Whereas the present results have revealed novel genetic determinants of circulating glycine levels, our study should also be taken in the context of certain limitations. First, depending on the cohort, metabolomic analysis was carried out using different platforms and glycine was measured in either serum or plasma, some of which were not fasting samples. Although this may have led to identifying fewer significant associations for circulating glycine levels, our relatively large sample size in the meta-analysis still provided sufficient power to detect robust associations at several previously known loci and 7 novel genomic regions. Second, the sex-stratified analyses had approximately half the number of females than males, which likely decreased power to identify loci for circulating glycine levels that were either specific to, or more strongly associated in, 1 sex or the other. Third, all study subjects in our study were of European ancestry, and it is possible that the genetic association results for either circulating plasma glycine levels may not be generalizable to other populations. Last, our evaluation of the causal relationship between glycine and risk of CAD or traditional risk factors may have resulted in biased estimates because of pleiotropic effects, especially in models that included all 12 loci or the 7 newly identified SNPs, or because of weak instruments in nested models that included only the 4 or 2 loci directly involved in glycine metabolism.

In summary, the results of our study provide additional insight into the genetic architecture of glycine metabolism, but a more-complete understanding of the mechanisms through which some of these loci influence circulating levels remains to be determined. Despite these genetic findings, we did not obtain conclusive evidence for a causal relationship between glycine and risk of CAD, raising the possibility that another unknown metabolite or biological pathway is driving the protective association of glycine-raising alleles at the *CPS1* locus with risk of CAD.

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## Disclosures

Z. Wang and Hazen are named as co-inventors on pending and issued patents held by the Cleveland Clinic relating to cardiovascular diagnostics and therapeutics and have the right to receive royalty payment for inventions or discoveries related to cardiovascular diagnostics or therapeutics from Cleveland Heart Lab, Quest Diagnostics, and Procter & Gamble Company. Hazen also reports having been paid as a consultant from Procter & Gamble Company and having received research funds from Procter & Gamble Company and Roche. Kettunen reports owning a modest amount of stock options for Nightingale Health Ltd, a company offering metabolic profiling. The remaining authors have no disclosures to report.

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## References

- Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, Feldstein AE, Britt EB, Fu X, Chung YM, Wu Y, Schauer P, Smith JD, Allayee H, Tang WH, DiDonato JA, Lusis AJ, Hazen SL. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature*. 2011;472:57–63.
- Koeth RA, Wang Z, Levison BS, Buffa JA, Org E, Sheehy BT, Britt EB, Fu X, Wu Y, Li L, Smith JD, DiDonato JA, Chen J, Li H, Wu GD, Lewis JD, Warrier M, Brown JM, Krauss RM, Tang WH, Bushman FD, Lusis AJ, Hazen SL. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat Med*. 2013;19:576–585.
- Tang WH, Wang Z, Levison BS, Koeth RA, Britt EB, Fu X, Wu Y, Hazen SL. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N Engl J Med*. 2013;368:1575–1584.
- Hartiala JA, Tang WH, Wang Z, Crow AL, Stewart AF, Roberts R, McPherson R, Erdmann J, Willenborg C, Hazen SL, Allayee H. Genome-wide association study and targeted metabolomics identifies sex-specific association of CPS1 with coronary artery disease. *Nat Commun*. 2016;7:10558.
- Pare G, Chasman DI, Parker AN, Zee RR, Malarstig A, Seedorf U, Collins R, Erdmann J, Hamsten A, Miletich JP, Ridker PM. Novel associations of CPS1, MUT, NOX4, and DPEP1 with plasma homocysteine in a healthy population: a genome-wide evaluation of 13 974 participants in the Women's Genome Health Study. *Circ Cardiovasc Genet*. 2009;2:142–150.
- Lange LA, Croteau-Chonka DC, Marvelle AF, Qin L, Gaulton KJ, Kuzawa CW, McDade TW, Wang Y, Li Y, Levy S, Borja JB, Lange EM, Adair LS, Mohlke KL. Genome-wide association study of homocysteine levels in Filipinos provides evidence for CPS1 in women and a stronger MTHFR effect in young adults. *Hum Mol Genet*. 2010;19:2050–2058.
- van Meurs JB, Pare G, Schwartz SM, Hazra A, Tanaka T, Vermeulen SH, Cotlarciuc I, Yuan X, Malarstig A, Bandinelli S, Bis JC, Blom H, Brown MJ, Chen C, Chen YD, Clarke RJ, DeGhghan A, Erdmann J, Ferrucci L, Hamsten A, Hofman A, Hunter DJ, Goel A, Johnson AD, Kathiresan S, Kampman E, Kiel DP, Kiemeny LA, Chambers JC, Kraft P, Lindemans J, McKnight B, Nelson CP, O'Donnell CJ, Psaty BM, Ridker PM, Rivadeneira F, Rose LM, Seedorf U, Siscovick DS, Schunkert H, Selhub J, Ueland PM, Vollenweider P, Waeber G, Waterworth DM, Watkins H, Witteman JC, den Heijer M, Jacques P, Uitterlinden AG, Kooren JS, Rader DJ, Reilly MP, Mooser V, Chasman DI, Samani NJ, Ahmadi KR. Common genetic loci influencing plasma homocysteine concentrations and their effect on risk of coronary artery disease. *Am J Clin Nutr*. 2013;98:668–676.
- Williams SR, Yang Q, Chen F, Liu X, Keene KL, Jacques P, Chen WM, Weinstein G, Hsu FC, Beiser A, Wang L, Bookman E, Doherty KF, Wolf PA, Zilka M, Selhub J, Nelson S, Gogarten SM, Worrall BB, Seshadri S, Sale MM; Genomics and Randomized Trials Network; Framingham Heart Study. Genome-wide meta-analysis of homocysteine and methionine metabolism identifies five-one carbon metabolism loci and a novel association of ALDH1L1 with ischemic stroke. *PLoS Genet*. 2014;10:e1004214.
- Kottgen A, Pattaro C, Boger CA, Fuchsberger C, Olden M, Glazer NL, Parsa A, Gao X, Yang Q, Smith AV, O'Connell JR, Li M, Schmidt H, Tanaka T, Isaacs A, Ketkar S, Hwang SJ, Johnson AD, DeGhghan A, Teumer A, Pare G, Atkinson EJ, Zeller T, Lohman K, Cornelis MC, Probst-Hensch NM, Kronenberg F, Tonjes A, Hayward C, Aspelund T, Eiriksdottir G, Launer LJ, Harris TB, Rasmussen L, Mitchell BD, Arking DE, Boerwinkle E, Struchalin M, Cavalieri M, Singleton A, Giallauria F, Metter J, de Boer IH, Haritunians T, Lumley T, Siscovick D, Psaty BM, Zillikens MC, Oostra BA, Feitosa M, Province M, de Andrade M, Turner ST, Schillert A, Ziegler A, Wild PS, Schnabel RB, Wilde S, Munzel TF, Leak TS, Illig T, Klopp N, Meisinger C, Wichmann HE, Koenig W, Zgaga L, Zemunik T, Kolcic I, Minelli C, Hu FB, Johansson A, Igl W, Zaboli G, Wild SH, Wright AF, Campbell H, Elinghaus D, Schreiber S, Aulchenko YS, Felix JF, Rivadeneira F, Uitterlinden AG, Hofman A, Imboden M, Nitsch D, Brandstatter A, Kollerits B, Kedenko L, Magi R, Stumvoll M, Kovacs P, Boban M, Campbell S, Endlich K, Volzke H, Kroemer HK, Nauck M, Volker U, Polasek O, Vitart V, Badola S, Parker AN, Ridker PM, Kardina SL, Blankenberg S, Liu Y, Curhan GC, Franke A, Rochat T, Paulweber B, Prokopenko I, Wang W, Gudnason V, Shuldiner AR, Coresh J, Schmidt R, Ferrucci L, Shlipak MG, van Duijn CM, Borecki I, Kramer BK, Rudan I, Gyllenstein U, Wilson JF, Witteman JC, Pramstaller PP, Rettig R, Hastie N, Chasman DI, Kao WH, Heid IM, Fox CS. New loci associated with kidney function and chronic kidney disease. *Nat Genet*. 2010;42:376–384.
- Choe CU, Atzler D, Wild PS, Carter AM, Boger RH, Ojeda F, Simova O, Stockebrand M, Lackner K, Nabuurs C, Marescau B, Streichert T, Muller C, Lueburg N, De Deyn PP, Benndorf RA, Baldus S, Gerloff C, Blankenberg S, Heerschap A, Grant PJ, Magnus T, Zeller T, Isbrandt D, Schwedhelm E. Homoarginine levels are regulated by L-arginine:glycine amidinotransferase and affect stroke outcome: results from human and murine studies. *Circulation*. 2013;128:1451–1461.
- Kleber ME, Seppala I, Pilz S, Hoffmann MM, Tomaschitz A, Oksala N, Raitoharju E, Lytikainen LP, Makela KM, Laaksonen R, Kahonen M, Raitakari OT, Huang J, Kienreich K, Fahrleitner-Pammer A, Drechsler C, Krane V, Boehm BO, Koenig W, Wanner C, Lehtimaki T, Marz W, Meitner A. Genome-wide association study identifies 3 genomic loci significantly associated with serum levels of homoarginine: the AtheroRemo Consortium. *Circ Cardiovasc Genet*. 2013;6:505–513.
- Global Lipids Genetics Consortium, Willer CJ, Schmidt EM, Sengupta S, Peloso GM, Gustafsson S, Kanoni S, Ganna A, Chen J, Buchkovich ML, Mora S, Beckmann JS, Bragg-Gresham JL, Chang HY, Demirkan A, Den Hertog HM, Do R, Donnelly LA, Ehret GB, Esko T, Feitosa MF, Ferreira T, Fischer K, Fontanillas P, Fraser RM, Freitag DF, Gurdasani D, Heikkila K, Hypponen E, Isaacs A, Jackson AU, Johansson A, Johnson T, Kaakinen M, Kettunen J, Kleber ME, Li X, Luan J, Lytikainen LP, Magnusson PK, Mangino M, Mihailov E, Montasser ME, Muller-Nurasyid M, Nolte IM, O'Connell JR, Palmer CD, Perola M, Petersen AK, Sanna S, Saxena R, Service SK, Shah S, Shungin D, Sidore C, Song C, Strawbridge RJ, Surakka I, Tanaka T, Teslovich TM, Thorleifsson G, Van den Herik EG, Voight BF, Volcik KA, Waite LL, Wong A, Wu Y, Zhang W, Absher D, Asiki G, Barroso I, Been LF, Bolton JL, Bonnycastle LL, Brambilla P, Burnett MS, Cesana G, Dimitriou M, Doney AS, Doring A, Elliott P, Epstein SE, Eyjolfsson GI, Gigante B, Goodarzi MO, Grallert H, Gravitto ML, Groves CJ, Hallmans G, Hartikainen AL, Hayward C, Hernandez D, Hicks AA, Holm H, Hung YJ, Illig T, Jones MR, Kaleebu P, Kastelein JJ, Khaw KT, Kim E, Klopp N, Komulainen P, Kumari M, Langenberg C, Lehtimaki T, Lin SY, Lindstrom J, Loos RJ, Mach F, McArdle TL, Meisinger C, Mitchell BD, Muller G, Nagaraja R, Narisu N, Nieminen VW, Nsubuga RN, Olafsson I, Ong KK, Palotie A, Papamarkou T, Pomilla C, Pouta A, Rader DJ, Reilly MP, Ridker PM, Rivadeneira F, Rudan I, Ruokonen A, Samani N, Scharnagl H, Seeley J, Silander K, Stancakova A, Stirrups K, Swift AJ, Tiret L, Uitterlinden AG, van Pelt LJ, Vedantam S,



- Wainwright N, Wijmenga C, Wild SH, Willemsen G, Wilsgaard T, Wilson JF, Young EH, Zhao JH, Adair LS, Arveiler D, Assimes TL, Bandinelli S, Bennett F, Bochud M, Boehm BO, Boomsma DI, Borecki IB, Bornstein SR, Bovet P, Burnier M, Campbell H, Chakravarti A, Chambers JC, Chen YD, Collins FS, Cooper RS, Danesh J, Dedoussis G, de Faire U, Feranil AB, Ferrieres J, Ferrucci L, Freimer NB, Gieger C, Groop LC, Gudnason V, Gyllenstein U, Hamsten A, Harris TB, Hingorani A, Hirschhorn JN, Hofman A, Hovingh GK, Hsiung CA, Humphries SE, Hunt SC, Hveem K, Iribarren C, Jarvelin MR, Jula A, Kahonen M, Kaprio J, Kesaniemi A, Kivimaki M, Koener JS, Koudstaal PJ, Krauss RM, Kuh D, Kuusisto J, Kyvik KO, Laakso M, Lakka TA, Lind L, Lindgren CM, Martin NG, Marz W, McCarthy ML, McKenzie CA, Meneton P, Metspalu A, Moilanen L, Morris AD, Munroe PB, Njolstad I, Pedersen NL, Power C, Pramstaller PP, Price JF, Psaty BM, Quertermous T, Rauramaa R, Saleheen D, Salomaa V, Sanghera DK, Saramies J, Schwarz PE, Sheu WH, Shuldiner AR, Siegbahn A, Spector TD, Stefansson K, Strachan DP, Tayo BO, Tremoli E, Tuomilehto J, Uusitupa M, van Duijn CM, Vollenweider P, Wallentin L, Wareham NJ, Whitfield JB, Wolffenbuttel BH, Ordovas JM, Boerwinkle E, Palmer CN, Thorsteinsdottir U, Chasman DI, Rotter JJ, Franks PW, Ripatti S, Cupples LA, Sandhu MS, Rich SS, Boehnke M, Deloukas P, Kathiresan S, Mohlke KL, Ingelsson E, Abecasis GR. Discovery and refinement of loci associated with lipid levels. *Nat Genet.* 2013;45:1274–1283.
13. Danik JS, Pare G, Chasman DI, Zee RY, Kwiatkowski DJ, Parker A, Mileitch JP, Ridker PM. Novel loci, including those related to Crohn disease, psoriasis, and inflammation, identified in a genome-wide association study of fibrinogen in 17 686 women: the Women's Genome Health Study. *Circ Cardiovasc Genet.* 2009;2:134–141.
14. Sabater-Lleal M, Huang J, Chasman D, Naitza S, Dehghan A, Johnson AD, Teumer A, Reiner AP, Folkersen L, Basu S, Rudnicka AR, Trompet S, Malarstig A, Baumert J, Bis JC, Guo X, Hottenga JJ, Shin SY, Lopez LM, Lahti J, Tanaka T, Yanek LR, Oudot-Mellakh T, Wilson JF, Navarro P, Huffman JE, Zemunik T, Redline S, Mehra R, Pulanic D, Rudan I, Wright AF, Kolcic I, Polasek O, Wild SH, Campbell H, Curb JD, Wallace R, Liu S, Eaton CB, Becker DM, Becker LC, Bandinelli S, Raikonen K, Widen E, Palotie A, Fornage M, Green D, Gross M, Davies G, Harris SE, Liewald DC, Starr JM, Williams FM, Grant PJ, Spector TD, Strawbridge RJ, Silveira A, Sennblad B, Rivadeneira F, Uitterlinden AG, Franco OH, Hofman A, van Dongen J, Willemsen G, Boomsma DI, Yao J, Swords Jenny N, Haritunians T, McKnight B, Lumley T, Taylor KD, Rotter JJ, Psaty BM, Peters A, Gieger C, Illig T, Grotevendt A, Homuth G, Volzke H, Kocher T, Goel A, Franzosi MG, Seedorf U, Clarke R, Steri M, Tarasov KV, Sanna S, Schlessinger D, Stott DJ, Sattar N, Buckley BM, Rumley A, Lowe GD, McArdle WL, Chen MH, Tofler GH, Song J, Boerwinkle E, Folsom AR, Rose LM, Franco-Cereceda A, Teichert M, Ikrum MA, Mosley TH, Bevan S, Dichgans M, Rothwell PM, Sudlow CL, Hopewell JC, Chambers JC, Saleheen D, Koener JS, Danesh J, Nelson CP, Erdmann J, Reilly MP, Kathiresan S, Schunkert H, Morange PE, Ferrucci L, Eriksson JG, Jacobs D, Deary IJ, Soranzo N, Witteman JC, de Geus EJ, Tracy RP, Hayward C, Koehnig W, Cucca F, Jukema JW, Eriksson P, Seshadri S, Markus HS, Watkins H, Samani NJ; V.T.E. Consortium, Stroke Consortium, Wellcome Trust Case Control Consortium, C4D Consortium, CARDIOGRAM Consortium, Wallaschofski H, Smith NL, Tregouet D, Ridker PM, Tang W, Strachan DP, Hamsten A, O'Donnell CJ. Multiethnic meta-analysis of genome-wide association studies in >100 000 subjects identifies 23 fibrinogen-associated loci but no strong evidence of a causal association between circulating fibrinogen and cardiovascular disease. *Circulation.* 2013;128:1310–1324.
15. Illig T, Gieger C, Zhai G, Romisch-Margl W, Wang-Sattler R, Prehn C, Altmayer E, Kastenmuller G, Kato BS, Mewes HW, Meitinger T, de Angelis MH, Kronenberg F, Soranzo N, Wichmann HE, Spector TD, Adamski J, Suhre K. A genome-wide perspective of genetic variation in human metabolism. *Nat Genet.* 2010;42:137–141.
16. Suhre K, Shin SY, Petersen AK, Mohny RP, Meredith D, Wagele B, Altmayer E, Deloukas P, Erdmann J, Grundberg E, Hammond CJ, de Angelis MH, Kastenmuller G, Kottgen A, Kronenberg F, Mangion M, Meisinger C, Meitinger T, Mewes HW, Milburn MV, Prehn C, Raffler J, Ried JS, Romisch-Margl W, Samani NJ, Small KS, Wichmann HE, Zhai G, Illig T, Spector TD, Adamski J, Soranzo N, Gieger C. Human metabolic individuality in biomedical and pharmaceutical research. *Nature.* 2011;477:54–60.
17. Mittelstrass K, Ried JS, Yu Z, Krumsiek J, Gieger C, Prehn C, Roemisch-Margl W, Polonikov A, Peters A, Theis FJ, Meitinger T, Kronenberg F, Weidinger S, Wichmann HE, Suhre K, Wang-Sattler R, Adamski J, Illig T. Discovery of sexual dimorphisms in metabolic and genetic biomarkers. *PLoS Genet.* 2011;7:e1002215.
18. Spittler A, Reissner CM, Oehler R, Gornikiewicz A, Gruenberger T, Manhart N, Brodowicz T, Mittlboeck M, Boltz-Nitulescu G, Roth E. Immunomodulatory effects of glycine on LPS-treated monocytes: reduced TNF-alpha production and accelerated IL-10 expression. *FASEB J.* 1999;13:563–571.
19. Wheeler M, Stachlewitz RF, Yamashina S, Ikejima K, Morrow AL, Thurman RG. Glycine-gated chloride channels in neutrophils attenuate calcium influx and superoxide production. *FASEB J.* 2000;14:476–484.
20. Bruck R, Wardi J, Aeed H, Avni Y, Shirin H, Avinoach I, Shahmurov M, Hershkovitz R. Glycine modulates cytokine secretion, inhibits hepatic damage and improves survival in a model of endotoxemia in mice. *Liver Int.* 2003;23:276–282.
21. Hasegawa S, Ichijima T, Sonaka I, Ohsaki A, Okada S, Wakiguchi H, Kudo K, Kittaka S, Hara M, Furukawa S. Cysteine, histidine and glycine exhibit anti-inflammatory effects in human coronary arterial endothelial cells. *Clin Exp Immunol.* 2012;167:269–274.
22. Ruiz-Ramirez A, Ortiz-Balderas E, Cardozo-Saldana G, Diaz-Diaz E, El-Hafidi M. Glycine restores glutathione and protects against oxidative stress in vascular tissue from sucrose-fed rats. *Clin Sci (Lond).* 2014;126:19–29.
23. Schemmer P, Zhong Z, Galli U, Wheeler MD, Xiangli L, Bradford BU, Conzelmann LO, Forman D, Boyer J, Thurman RG. Glycine reduces platelet aggregation. *Amino Acids.* 2013;44:925–931.
24. Polfus LM, Khajuria RK, Schick UM, Pankratz N, Pazoki R, Brody JA, Chen MH, Auer PL, Floyd JS, Huang J, Lange L, van Rooij FJ, Gibbs RA, Metcalf G, Muzny D, Veeraghavan N, Walter K, Chen L, Yanek L, Becker LC, Peloso GM, Wakabayashi A, Kals M, Metspalu A, Esko T, Fox K, Wallace R, Franceschini N, Matijevic N, Rice KM, Bartz TM, Lytikainen LP, Kahonen M, Lehtimaki T, Raitakari OT, Li-Gao R, Mook-Kanamori DO, Lettre G, van Duijn CM, Franco OH, Rich SS, Rivadeneira F, Hofman A, Uitterlinden AG, Wilson JG, Psaty BM, Soranzo N, Dehghan A, Boerwinkle E, Zhang X, Johnson AD, O'Donnell CJ, Johnsen JM, Reiner AP, Ganesh SK, Sankaran VG. Whole-exome sequencing identifies loci associated with blood cell traits and reveals a role for alternative GFI1B splice variants in human hematopoiesis. *Am J Hum Genet.* 2016;99:481–488.
25. El Hafidi M, Perez I, Banos G. Is glycine effective against elevated blood pressure? *Curr Opin Clin Nutr Metab Care.* 2006;9:26–31.
26. Ding Y, Svingen GF, Pedersen ER, Gregory JF, Ueland PM, Tell GS, Nygard OK. Plasma glycine and risk of acute myocardial infarction in patients with suspected stable angina pectoris. *J Am Heart Assoc.* 2016;5:e002621. DOI: 10.1161/JAHA.115.002621.
27. Vartiainen E, Laatikainen T, Peltonen M, Juolevi A, Mannisto S, Sundvall J, Jousilahti P, Salomaa V, Valsta L, Puska P. Thirty-five-year trends in cardiovascular risk factors in Finland. *Int J Epidemiol.* 2010;39:504–518.
28. Raitakari OT, Juonala M, Ronnema T, Keltikangas-Jarvinen L, Rasanen L, Pietikainen M, Hutri-Kahonen N, Taittonen L, Jokinen E, Marniemi J, Jula A, Telama R, Kahonen M, Lehtimaki T, Akerblom HK, Viikari JS. Cohort profile: the cardiovascular risk in Young Finns Study. *Int J Epidemiol.* 2008;37:1220–1226.
29. Rantakallio P. The longitudinal study of the northern Finland birth cohort of 1966. *Paediatr Perinat Epidemiol.* 1988;2:59–88.
30. Laakso M, Kuusisto J, Stancakova A, Kuulasmaa T, Pajukanta P, Lusia AJ, Collins FS, Mohlke KL, Boehnke M. The Metabolic Syndrome in Men study: a resource for studies of metabolic and cardiovascular diseases. *J Lipid Res.* 2017;58:481–493.
31. Kettunen J, Demirkan A, Wurtz P, Draisma HH, Haller T, Rawal R, Vaarhorst A, Kangas AJ, Lytikainen LP, Pirinen M, Pool R, Sarin AP, Soininen P, Tuikainen T, Wang Q, Tiainen M, Tynkkynen T, Amin N, Zeller T, Beekman M, Deelen J, van Dijk KW, Esko T, Hottenga JJ, van Leeuwen EM, Lehtimaki T, Mihailov E, Rose RJ, de Craen AJ, Gieger C, Kahonen M, Perola M, Blankenberg S, Savolainen MJ, Verhoeven A, Viikari J, Willemsen G, Boomsma DI, van Duijn CM, Eriksson J, Jula A, Jarvelin MR, Kaprio J, Metspalu A, Raitakari O, Salomaa V, Slagboom PE, Waldenberger M, Ripatti S, Ala-Korpela M. Genome-wide study for circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA. *Nat Commun.* 2016;7:11122.
32. Teslovich TM, Kim DS, Yin X, Stancakova A, Jackson AU, Wielscher M, Naj A, Perry JRB, Huyghe JR, Stringham HM, Davis JP, Raulerson CK, Welch RP, Fuchsberger C, Locke AE, Sim X, Chines PS, Narisu N, Kangas AJ, Soininen P; Genetics of Obesity-Related Liver Disease Consortium, Alzheimer's Disease Genetics Consortium (ADGC), Diabetes Genetics Replication And Meta-analysis (DIAGRAM), Ala-Korpela M, Gudnason V, Musani SK, Jarvelin MR, Schellenberger GD, Speliotes EK, Kuusisto J, Collins FS, Boehnke M, Laakso M, Mohlke KL. Identification of seven novel loci associated with amino acid levels using single-variant and gene-based tests in 8545 Finnish men from the METSIM study. *Hum Mol Genet.* 2018;27:1664–1674.
33. Magi R, Morris AP. GWAMA: software for genome-wide association meta-analysis. *BMC Bioinformatics.* 2010;11:288.
34. Turner SD. qqman: an R package for visualizing GWAS results using Q-Q and manhattan plots. *J Open Source Software.* 2018;3:731. Available at: <https://doi.org/10.21105/joss.00731>.
35. Staley JR, Blackshaw J, Kamat MA, Ellis S, Surendran P, Sun BB, Paul DS, Freitag D, Burgess S, Danesh J, Young R, Butterworth AS. PhenoScanner: a database of human genotype-phenotype associations. *Bioinformatics.* 2016;32:3207–3209.
36. Speed D, Balding DJ. SumHer better estimates the SNP heritability of complex traits from summary statistics. *Nat Genet.* 2019;51:277–284.
37. van der Harst P, Verweij N. Identification of 64 novel genetic loci provides an expanded view on the genetic architecture of coronary artery disease. *Circ Res.* 2018;122:433–443.
38. Surakka I, Horikoshi M, Magi R, Sarin AP, Mahajan A, Lagou V, Marullo L, Ferreira T, Miraglio B, Timonen S, Kettunen J, Pirinen M, Karjalainen J,

- Thorleifsson G, Hagg S, Hottenga JJ, Isaacs A, Ladvall C, Beekman M, Esko T, Ried JS, Nelson CP, Willenborg C, Gustafsson S, Westra HJ, Blades M, de Craen AJ, de Geus EJ, Deelen J, Grallert H, Hamsten A, Havulinna AS, Hengstenberg C, Houwing-Duistermaat JJ, Hypponen E, Karssen LC, Lehtimäki T, Lyssenko V, Magnusson PK, Mihailov E, Muller-Nurasyid M, Mpindi JP, Pedersen NL, Penninx BW, Perola M, Pers TH, Peters A, Rung J, Smit JH, Steinthorsdottir V, Tobin MD, Tsernikova N, van Leeuwen EM, Viikari JS, Willems SM, Willemssen G, Schunkert H, Erdmann J, Samani NJ, Kaprio J, Lind L, Gieger C, Metspalu A, Slagboom PE, Groop L, van Duijn CM, Eriksson JG, Jula A, Salomaa V, Boomsma DI, Power C, Raitakari OT, Ingelsson E, Jarvelin MR, Thorsteinsdottir U, Franke L, Ikonen E, Kallioniemi O, Pietiäinen V, Lindgren CM, Stefansson K, Palotie A, McCarthy MI, Morris AP, Prokopenko I, Ripatti S; ENGAGE Consortium. The impact of low-frequency and rare variants on lipid levels. *Nat Genet.* 2015;47:589–597.
39. Loh PR, Kichaev G, Gazal S, Schoech AP, Price AL. Mixed-model association for biobank-scale datasets. *Nat Genet.* 2018;50:906–908.
40. Dastani Z, Hivert MF, Timpson N, Perry JR, Yuan X, Scott RA, Henneman P, Heid IM, Kizer JR, Lyytikäinen LP, Fuchsberger C, Tanaka T, Morris AP, Small K, Isaacs A, Beekman M, Coassin S, Lohman K, Qi L, Kanoni S, Pankow JS, Uh HW, Wu Y, Bidulescu A, Rasmussen-Torvik LJ, Greenwood CM, Ladouceur M, Grimsby J, Manning AK, Liu CT, Kooner J, Mooser VE, Vollenweider P, Kapur KA, Chambers J, Wareham NJ, Langenberg C, Frants R, Willems-Vandijk K, Oostra BA, Willems SM, Lamina C, Winkler TW, Psaty BM, Tracy RP, Brody J, Chen I, Viikari J, Kahonen M, Pramstaller PP, Evans DM, St Pourcain B, Sattar N, Wood AR, Bandinelli S, Carlson OD, Egan JM, Bohringer S, van Heemst D, Kedenko L, Kristiansson K, Nuotio ML, Loo BM, Harris T, Garcia M, Kanaya A, Haun M, Klopp N, Wichmann HE, Deloukas P, Katsareli E, Couper DJ, Duncan BB, Kloppenburg M, Adair LS, Borja JB; DIAGRAM Consortium, MAGIC Consortium, GLGC Consortium, MuTHER Consortium, Wilson JG, Musani S, Guo X, Johnson T, Semple R, Teslovich TM, Allison MA, Redline S, Buxbaum SG, Mohlke KL, Meulenbelt I, Ballantyne CM, Dedoussis GV, Hu FB, Liu Y, Paulweber B, Spector TD, Slagboom PE, Ferrucci L, Jula A, Perola M, Raitakari O, Florez JC, Salomaa V, Eriksson JG, Frayling TM, Hicks AA, Lehtimäki T, Smith GD, Siscovick DS, Kronenberg F, van Duijn C, Loos RJ, Waterworth DM, Meigs JB, Dupuis J, Richards JB, Voight BF, Scott LJ, Steinthorsdottir V, Dina C, Welch RP, Zeggini E, Huth C, Aulchenko YS, Thorleifsson G, McCulloch LJ, Ferreira T, Grallert H, Amin N, Wu G, Willer CJ, Raychaudhuri S, McCarroll SA, Hofmann OM, Segre AV, van Hoek M, Navarro P, Ardlie K, Balkau B, Benediktsson R, Bennett AJ, Blagieva R, Boerwinkle E, Bonnycastle LL, Bostrom KB, Braavender B, Bumpstead S, Burt NP, Charpentier G, Chines PS, Cornelis M, Crawford G, Doney AS, Elliott KS, Elliott AL, Erdos MR, Fox CS, Franklin CS, Ganser M, Gieger C, Grarup N, Green T, Griffin S, Groves CJ, Guiducci C, Hadjadj S, Hassani N, Herder C, Isomaa B, Jackson AU, Johnson PR, Jorgensen T, Kao WH, Kong A, Kraff P, Kuusisto J, Lauritzen T, Li M, Lieverse A, Lindgren CM, Lyssenko V, Marre M, Meitinger T, Midtjell K, Morken MA, Narisu N, Nilsson P, Owen KR, Payne F, Petersen AK, Platou C, Proenca C, Prokopenko I, Rathmann W, Rayner NW, Robertson NR, Rocheleau G, Roden M, Sampson MJ, Saxena R, Shields BM, Shrader P, Sigurdsson G, Sparso T, Strassburger K, Stringham HM, Sun Q, Swift AJ, Thorand B, Tichet J, Tuomi T, van Dam RM, van Haften TW, van Herpt T, van Vliet-Ostaptchouk JV, Walters GB, Weedon MN, Wijmenga C, Witteman J, Bergman RN, Cauchi S, Collins FS, Gloyl A, Gyllenstein U, Hansen T, Hide WA, Hitman GA, Hofman A, Hunter DJ, Hveem K, Laakso M, Morris AD, Palmer CN, Rudan I, Sijbrands E, Stein LD, Tuomilehto J, Uitterlinden A, Walker M, Watanabe RM, Abecasis GR, Boehm BO, Campbell H, Daly MJ, Hattersley AT, Pedersen O, Barroso I, Groop L, Sladek R, Thorsteinsdottir U, Wilson JF, Illig T, Froguel P, van Duijn CM, Stefansson K, Altshuler D, Boehnke M, McCarthy MI, Soranzo N, Wheeler E, Glazer NL, Bouatia-Naji N, Magi R, Randall J, Elliott P, Rybin D, Dehghan A, Hottenga JJ, Song K, Goel A, Lajunen T, Doney A, Cavalcanti-Proenca C, Kumari M, Timpson NJ, Zabena C, Ingelsson E, An P, O'Connell J, Luan J, Elliott A, McCarroll SA, Roccascassa RM, Pattou F, Sethupathy P, Ariyurek Y, Barter P, Beilby JP, Ben-Shlomo Y, Bergmann S, Bochud M, Bonnefond A, Borch-Johnsen K, Bottcher Y, Brunner E, Bumpstead SJ, Chen YD, Chines P, Clarke R, Coin LJ, Cooper MN, Crisponi L, Day IN, de Geus EJ, Delplanque J, Fedson AC, Fischer-Rosinsky A, Forouhi NG, Franzosi MG, Galan P, Goodarzi MO, Graessler J, Grundy S, Gwilliam R, Hallmans G, Hammond N, Han X, Hartikainen AL, Hayward C, Heath SC, Hercberg S, Hillman DR, Hingorani AD, Hui J, Hung J, Kaakinen M, Kaprio J, Kesaniemi YA, Kivimäki M, Knight B, Koskinen S, Kovacs P, Kyvik KO, Lathrop GM, Lawlor DA, Le Bacquer O, LeCoeur C, Li Y, Mahley R, Mangino M, Martinez-Larrad MT, McAttee JB, McPherson R, Meisinger C, Melzer D, Meyre D, Mitchell BD, Mukherjee S, Naitza S, Neville MJ, Orru M, Pakyz R, Paolisso G, Pattaro C, Pearson D, Peden JF, Pedersen NL, Pfeiffer AF, Pichler I, Polasek O, Posthuma D, Potter SC, Pouta A, Province MA, Rayner NW, Rice K, Ripatti S, Rivadeneira F, Rolandsson O, Sandbaek A, Sandhu M, Sanna S, Sayer AA, Scheet P, Seedorf U, Sharp SJ, Shields B, Sigurdsson G, Sijbrands EJ, Silveira A, Simpson L, Singleton A, Smith NL, Sovio U, Swift A, Sydall H, Syvanen AC, Tonjes A, Uitterlinden AG, van Dijk KW, Varma D, Visvikis-Siest S, Vitart V, Vogelzangs N, Waeber G, Wagner PJ, Walley A, Ward KL, Watkins H, Wild SH, Willemssen G, Witteman JC, Yarnell JW, Zelenika D, Zethelius B, Zhai G, Zhao JH, Zillikens MC; DIAGRAM Consortium; GIANT Consortium; Global BPGen Consortium, Borecki IB, Meneton P, Magnusson PK, Nathan DM, Williams GH, Silander K, Bornstein SR, Schwarz P, Spranger J, Karpe F, Shuldiner AR, Cooper C, Serrano-Rios M, Lind L, Palmer LJ, Hu FB, Franks PW, Ebrahim S, Marmot M, Kao WH, Pramstaller PP, Wright AF, Stumvoll M, Hamsten A; Procardis C, Buchanan TA, Valle TT, Rotter JJ, Penninx BW, Boomsma DI, Cao A, Scuteri A, Schlessinger D, Uda M, Ruokonen A, Jarvelin MR, Peltonen L, Mooser V, Sladek R; MAGIC investigators, GLGC Consortium, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M, Pirruccello JP, Chasman DI, Johansen CT, Fouchier SW, Peloso GM, Barbalic M, Ricketts SL, Bis JC, Feitosa MF, Orho-Melander M, Melander O, Li X, Li M, Cho YS, Go MJ, Kim YJ, Lee JY, Park T, Kim K, Sim X, Ong RT, Croteau-Chonka DC, Lange LA, Smith JD, Ziegler A, Zhang W, Zee RY, Whitfield JB, Thompson JR, Surakka I, Spector TD, Smit JH, Sinisalo J, Scott J, Saharinen J, Sabatti C, Rose LM, Roberts R, Rieder M, Parker AN, Pare G, O'Donnell CJ, Nieminen MS, Nickerson DA, Montgomery GW, McArdle W, Masson D, Martin NG, Marroni F, Lucas G, Luben R, Lokki ML, Lettre G, Launer LJ, Lakatta EG, Laaksonen R, Kyvik KO, König IR, Khaw KT, Kaplan LM, Johansson A, Janssens AC, Igl W, Hovingh GK, Hengstenberg C, Havulinna AS, Hastie ND, Harris TB, Haritunians T, Hall AS, Groop LC, Gonzalez E, Freimer NB, Erdmann J, Ejebe KG, Doring A, Dominiczak AF, Demissie S, Deloukas P, de Faire U, Crawford G, Chen YD, Caulfield MJ, Boehmholdt SM, Assimes TL, Quertermous T, Seielstad M, Wong TY, Tai ES, Feranil AB, Kuzawa CW, Taylor HA Jr, Gabriel SB, Holm H, Gudnason V, Krauss RM, Ordovas JM, Munroe PB, Kooner JS, Tall AR, Hegele RA, Kastelein JJ, Schadt EE, Strachan DP, Reilly MP, Samani NJ, Schunkert H, Cupples LA, Sandhu MS, Ridker PM, Rader DJ, Kathiresan S. Novel loci for adiponectin levels and their influence on type 2 diabetes and metabolic traits: a multi-ethnic meta-analysis of 45,891 individuals. *PLoS Genet.* 2012;8:e1002607.
41. Hemani G, Zheng J, Elsworth B, Wade KH, Haberland V, Baird D, Laurin C, Burgess S, Bowden J, Langdon R, Tan VY, Yarmolinsky J, Shihab HA, Timpson NJ, Evans DM, Relton C, Martin RM, Davey Smith G, Gaunt TR, Haycock PC. The MR-Base platform supports systematic causal inference across the human phenome. *Elife.* 2018;7:e34408.
42. de Koning TJ. Amino acid synthesis deficiencies. *J Inherit Metab Dis.* 2017;40:609–620.
43. Rhee EP, Ho JE, Chen MH, Shen D, Cheng S, Larson MG, Ghorbani A, Shi X, Helenius IT, O'Donnell CJ, Souza AL, Deik A, Pierce KA, Bullock K, Walford GA, Vasani RS, Florez JC, Clish C, Yeh JR, Wang TJ, Gerszten RE. A genome-wide association study of the human metabolome in a community-based cohort. *Cell Metab.* 2013;18:130–143.
44. Xie W, Wood AR, Lyssenko V, Weedon MN, Knowles JW, Alkayyali S, Assimes TL, Quertermous T, Abbasi F, Paananen J, Haring H, Hansen T, Pedersen O, Smith U, Laakso M, Dekker JM, Nolan JJ, Groop L, Ferrannini E, Adam KP, Gall WE, Frayling TM, Walker M. Genetic variants associated with glycine metabolism and their role in insulin sensitivity and type 2 diabetes. *Diabetes.* 2013;62:2141–2150.
45. Shin SY, Fauman EB, Petersen AK, Krumsiek J, Santos R, Huang J, Arnold M, Erte I, Forgetta V, Yang TP, Walter K, Menni C, Chen L, Vasquez L, Valdes AM, Hyde CL, Wang V, Ziemek D, Roberts P, Xi L, Grundberg E; Multiple Tissue Human Expression Resource (MuTHER) Consortium, Waldenberger M, Richards JB, Mohny RP, Milburn MV, John SL, Trimmer J, Theis FJ, Overington JP, Suhre K, Brogan MJ, Gieger C, Kastenmuller G, Spector TD, Soranzo N. An atlas of genetic influences on human blood metabolites. *Nat Genet.* 2014;46:543–550.
46. Draisma HHM, Pool R, Kobl M, Jansen R, Petersen AK, Vaarhorst AAM, Yet I, Haller T, Demirkan A, Esko T, Zhu G, Bohringer S, Beekman M, van Klinken JB, Romisch-Margl W, Prehn C, Adamski J, de Craen AJM, van Leeuwen EM, Amin N, Dhurair H, Westra HJ, Franke L, de Geus EJC, Hottenga JJ, Willemssen G, Henders AK, Montgomery GW, Nyholt DR, Whitfield JB, Penninx BW, Spector TD, Metspalu A, Slagboom PE, van Dijk KW, 't Hoen PAC, Strauch K, Martin NG, van Ommen GB, Illig T, Bell JT, Mangino M, Suhre K, McCarthy MI, Gieger C, Isaacs A, van Duijn CM, Boomsma DI. Genome-wide association study identifies novel genetic variants contributing to variation in blood metabolite levels. *Nat Commun.* 2015;6:7208.
47. Long T, Hicks M, Yu HC, Biggs WH, Kirkness EF, Menni C, Zierer J, Small KS, Mangino M, Messier H, Brewerton S, Turpaz Y, Perkins BA, Evans AM, Miller LA, Guo L, Caskey CT, Schork NJ, Garner C, Spector TD, Venter JC, Telenti A. Whole-genome sequencing identifies common-to-rare variants associated with human blood metabolites. *Nat Genet.* 2017;49:568–578.
48. Wang W, Wu Z, Dai Z, Yang Y, Wang J, Wu G. Glycine metabolism in animals and humans: implications for nutrition and health. *Amino Acids.* 2013;45:463–477.
49. Jaeken J, Dethoux M, Van Maldergem L, Foulon M, Carchon H, Van Schaftingen E. 3-Phosphoglycerate dehydrogenase deficiency: an inborn error of serine biosynthesis. *Arch Dis Child.* 1996;74:542–545.
50. Pineda M, Vilaseca MA, Artuch R, Santos S, Garcia Gonzalez MM, Aracil A, Van Schaftingen E, Jaeken J. 3-Phosphoglycerate dehydrogenase deficiency in a patient with West syndrome. *Dev Med Child Neurol.* 2000;42:629–633.
51. van der Crabben SN, Verhoeven-Duif NM, Brilstra EH, Van Maldergem L, Coskun T, Rubio-Gozalbo E, Berger R, de Koning TJ. An update on serine deficiency disorders. *J Inherit Metab Dis.* 2013;36:613–619.

52. Byers HM, Bennett RL, Malouf EA, Weiss MD, Feng J, Scott CR, Jayadev S. Novel report of phosphoserine phosphatase deficiency in an adult with myeloneuropathy and limb contractures. *JIMD Rep*. 2016;30:103–108.
53. Kikuchi G, Motokawa Y, Yoshida T, Hiraga K. Glycine cleavage system: reaction mechanism, physiological significance, and hyperglycinemia. *Proc Jpn Acad Ser B Phys Biol Sci*. 2008;84:246–263.
54. Lamers Y, Williamson J, Gilbert LR, Stacpoole PW, Gregory JF III. Glycine turnover and decarboxylation rate quantified in healthy men and women using primed, constant infusions of [1,2-(13)C]glycine and [(2)H3]leucine. *J Nutr*. 2007;137:2647–2652.
55. Freeman JM, Nicholson JF, Schimke RT, Rowland LP, Carter S. Congenital hyperammonemia. Association with hyperglycinemia and decreased levels of carbamyl phosphate synthetase. *Arch Neurol*. 1970;23:430–437.
56. Colombo JP, Bachmann C, Schrammli A. Inborn defects of the mitochondrial portion of the urea cycle. *Ann N Y Acad Sci*. 1986;488:109–117.
57. Reilly MP, Li M, He J, Ferguson JF, Stylianou IM, Mehta NN, Burnett MS, Devaney JM, Knouff CW, Thompson JR, Horne BD, Stewart AF, Assimes TL, Wild PS, Allayee H, Nitschke PL, Patel RS, Martinelli N, Girelli D, Quyyumi AA, Anderson JL, Erdmann J, Hall AS, Schunkert H, Quertermous T, Blankenberg S, Hazen SL, Roberts R, Kathiresan S, Samani NJ, Epstein SE, Rader DJ, Qasim AN, DerOhannessian SL, Ou L, Cappola TP, Chen Z, Matthai W, Hakonarson HH, Wilensky R, Kent KM, Lindsay JM, Pichard AD, Satler L, Waksman R. Identification of ADAMTS7 as a novel locus for coronary atherosclerosis and association of ABO with myocardial infarction in the presence of coronary atherosclerosis: two genome-wide association studies. *Lancet*. 2011;377:383–392.
58. Nikpay M, Goel A, Won HH, Hall LM, Willenborg C, Kanoni S, Saleheen D, Kyriakou T, Nelson CP, Hopewell JC, Webb TR, Zeng L, Dehghan A, Alver M, Armasu SM, Auro K, Bjonnes A, Chasman DI, Chen S, Ford I, Franceschini N, Gieger C, Grace C, Gustafsson S, Huang J, Hwang SJ, Kim YK, Kleber ME, Lau KW, Lu X, Lu Y, Lyytikäinen LP, Mihailov E, Morrison AC, Pervjakova N, Qu L, Rose LM, Salfati E, Saxena R, Scholz M, Smith AV, Tikkanen E, Uitterlinden A, Yang X, Zhang W, Zhao W, de Andrade M, de Vries PS, van Zuydam NR, Anand SS, Bertram L, Beutner F, Dedoussis G, Frossard P, Gauguier D, Goodall AH, Gottesman O, Haber M, Han BG, Huang J, Jalilzadeh S, Kessler T, König IR, Lannfelt L, Lieb W, Lind L, Lindgren CM, Lokki ML, Magnusson PK, Mallick NH, Mehra N, Meitinger T, Memon FU, Morris AP, Nieminen MS, Pedersen NL, Peters A, Rallidis LS, Rasheed A, Samuel M, Shah SH, Sinisalo J, Stirrups KE, Trompet S, Wang L, Zaman KS, Ardisino D, Boerwinkle E, Borecki IB, Bottinger EP, Buring JE, Chambers JC, Collins R, Cupples LA, Danesh J, Demuth I, Elosua R, Epstein SE, Esko T, Feitosa MF, Franco OH, Franzosi MG, Granger CB, Gu D, Gudnason V, Hall AS, Hamsten A, Harris TB, Hazen SL, Hengstenberg C, Hofman A, Ingelsson E, Iribarren C, Jukema JW, Karhunen PJ, Kim BJ, Kooner JS, Kullo IJ, Lehtimäki T, Loos RJ, Melander O, Metspalu A, Marz W, Palmer CN, Perola M, Quertermous T, Rader DJ, Ridker PM, Ripatti S, Roberts R, Salomaa V, Sanghera DK, Schwartz SM, Seedorf U, Stewart AF, Stott DJ, Thiery J, Zalloua PA, O'Donnell CJ, Reilly MP, Assimes TL, Thompson JR, Erdmann J, Clarke R, Watkins H, Kathiresan S, McPherson R, Deloukas P, Schunkert H, Samani NJ, Farrall M; CARDIoGRAM+C4D Consortium. A comprehensive 1,000 Genomes-based genome-wide association meta-analysis of coronary artery disease. *Nat Genet*. 2015;47:1121–1130.
59. Teslovich TM, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M, Pirruccello JP, Ripatti S, Chasman DI, Willer CJ, Johansen CT, Fouchier SW, Isaacs A, Peloso GM, Barbalic M, Ricketts SL, Bis JC, Aulchenko YS, Thorleifsson G, Feitosa MF, Chambers J, Orho-Melander M, Melander O, Johnson T, Li X, Guo X, Li M, Shin Cho Y, Jin Go M, Jin Kim Y, Lee JY, Park T, Kim K, Sim X, Tsee-Hee Ong R, Croteau-Chonka DC, Lange LA, Smith JD, Song K, Hua Zhao J, Yuan X, Luan J, Lamina C, Ziegler A, Zhang W, Zee RY, Wright AF, Witteman JC, Wilson JF, Willemssen G, Wichmann HE, Whitfield JB, Waterworth DM, Wareham NJ, Waeber G, Vollenweider P, Voight BF, Vitart V, Uitterlinden AG, Uda M, Tuomilehto J, Thompson JR, Tanaka T, Surakka I, Stringham HM, Spector TD, Soranzo N, Smit JH, Sinisalo J, Silander K, Sijbrands EJ, Scuteri A, Scott J, Schlessinger D, Sanna S, Salomaa V, Saharinen J, Sabatti C, Ruukonen A, Rudan I, Rose LM, Roberts R, Rieder M, Psaty BM, Pramstaller PP, Pichler I, Perola M, Penninx BW, Pedersen NL, Pattaro C, Parker AN, Pare G, Oostra BA, O'Donnell CJ, Nieminen MS, Nickerson DA, Montgomery GW, Meitinger T, McPherson R, McCarthy MI, McArdle W, Masson D, Martin NG, Marroni F, Mangino M, Magnusson PK, Lucas G, Luben R, Loos RJ, Lokki ML, Lettre G, Langenberg C, Launer LJ, Lakatta EG, Laaksonen R, Kyvik KO, Kronenberg F, König IR, Khaw KT, Kaprio J, Kaplan LM, Johansson A, Jarvelin MR, Cecile JWA, Ingelsson E, Igl W, Kees Hovingh G, Hottenga JJ, Hofman A, Hicks AA, Hengstenberg C, Heid IM, Hayward C, Havulinna AS, Hastie ND, Harris TB, Haritunians T, Hall AS, Gyllenstein U, Guiducci C, Groop LC, Gonzalez E, Gieger C, Freimer NB, Ferrucci L, Erdmann J, Elliott P, Ejebe KG, Doring A, Dominiczak AF, Demissie S, Deloukas P, de Geus EJ, de Faire U, Crawford G, Collins FS, Chen YD, Caulfield MJ, Campbell H, Burt NP, Bonnycastle LL, Boomsma DI, Boekholdt SM, Bergman RN, Barroso I, Bandinelli S, Ballantyne CM, Assimes TL, Quertermous T, Altshuler D, Seielstad M, Wong TY, Tai ES, Feranil AB, Kuzawa CW, Adair LS, Taylor HA Jr, Borecki IB, Gabriel SB, Wilson JG, Holm H, Thorsteinsdottir U, Gudnason V, Krauss RM, Mohlke KL, Ordovas JM, Munroe PB, Kooner JS, Tall AR, Hegele RA, Kastelein JJ, Schadt EE, Rotter JJ, Boerwinkle E, Strachan DP, Mooser V, Stefansson K, Reilly MP, Samani NJ, Schunkert H, Cupples LA, Sandhu MS, Ridker PM, Rader DJ, van Duijn CM, Peltonen L, Abecasis GR, Boehnke M, Kathiresan S. Biological, clinical and population relevance of 95 loci for blood lipids. *Nature*. 2010;466:707–713.
60. Astle WJ, Elding H, Jiang T, Allen D, Ruklisa D, Mann AL, Mead D, Bouman H, Riveros-Mckay F, Kostadima MA, Lambourne JJ, Sivapalaratnam S, Downes K, Kundu K, Bomba L, Berentsen K, Bradley JR, Daugherty LC, Delaneau O, Freson K, Garner SF, Grassi L, Guerrero J, Haimel M, Janssen-Megens EM, Kaan A, Kamat M, Kim B, Mandoli A, Marchini J, Martens JH, Meacham S, Megy K, O'Connell J, Petersen R, Sharifi N, Sheard SM, Staley JR, Tuna S, van der Ent M, Walter K, Wang SY, Wheeler E, Wilder SP, Iotchkova V, Moore C, Sambrook J, Stunnenberg HG, Di Angelantonio E, Kaptoge S, Kujipers TW, Carrillo-de-Santa-Pau E, Juan D, Rico D, Valencia A, Chen L, Ge B, Vasquez L, Kwan T, Garrido-Martin D, Watt S, Yang Y, Guigo R, Beck S, Paul DS, Pastinen T, Bujold D, Bourque G, Frontini M, Danesh J, Roberts DJ, Ouwehand WH, Butterworth AS, Soranzo N. The allelic landscape of human blood cell trait variation and links to common complex disease. *Cell*. 2016;167:1415–1429.e19.

# Supplemental Material

## Data S1.

### Detailed Description of Cohorts

**GeneBank Study:** The Cleveland Clinic GeneBank study is a single site sample repository generated from consecutive patients undergoing elective diagnostic coronary angiography or elective cardiac computed tomographic angiography with extensive clinical and laboratory characterization and longitudinal observation. Subject recruitment occurred between 2001 and 2006. Ethnicity was self-reported and information regarding demographics, medical history, and medication use was obtained by patient interviews and confirmed by chart reviews. All clinical outcome data were verified by source documentation. Coronary artery disease (CAD) was defined as adjudicated diagnoses of stable or unstable angina, myocardial infarction (MI) (adjudicated definition based on defined electrocardiographic changes or elevated cardiac enzymes), angiographic evidence of  $\geq 50\%$  stenosis of one or more major epicardial vessel, and/or a history of known CAD (documented MI, CAD, or history of revascularization). The GeneBank Study has been used previously for discovery and replication of novel genes and risk factors for atherosclerotic disease<sup>1-4</sup>. Plasma glycine levels were measured in blood samples obtained upon entry into GeneBank. Genome-wide genotyping was carried out on 3031 GeneBank subjects of European ancestry using the Affymetrix Genome-Wide Human Array 6.0 SNP chip. After conversion of genomic coordinates to GRCh37/hg19, exclusion of SNPs with duplicates, call rates  $<97\%$ , minor allele frequencies (MAFs)  $<1\%$ , and without chromosome and base pair position, and exclusion of 44 subjects with genotype call rates  $<90\%$ , 642,766 were available for imputation in 2972 participants. Imputation was carried out on the forward (+)

strand using the University of Michigan Imputation Server (<https://imputationserver.sph.umich.edu>) and data from the 1000 Genomes Project (Phase 3, Version 5). Application of the same quality control filters described above to the 46,180,700 imputed SNPs, with the addition of excluding SNPs with Hardy-Weinberg equilibrium p-values  $<0.0001$  and imputation R<sub>sq</sub> scores  $<0.3$ , resulted in 8,986,545 autosomal SNPs that were available for analysis in 1276 GeneBank subjects for whom plasma glycine levels were also available. All patients provided written informed consent prior to being enrolled in GeneBank and the study was approved by the Institutional Review Board of the Cleveland Clinic.

**FINRISK:** FINRISK (FR) surveys are cross-sectional, population-based studies conducted every five years since 1972 to monitor risk of chronic diseases. For each survey, a representative random sample was selected from 25- to 74-year-old inhabitants of different regions in Finland. The survey included a questionnaire and a clinical examination, at which a blood sample was drawn, with linkage to national registries of cardiovascular disease and other health outcomes. The study protocol has been described elsewhere<sup>5</sup>. Study participants were followed up through December 31, 2012. Eligible individuals from FINRISK surveys conducted in 1992, 1997, 2002, and 2007 (total n=27 838) were genotyped in three separate batches and analyzed separately to avoid batch effects, followed by a meta-analysis for glycine levels as described previously<sup>6</sup>. Genome-wide genotyping was carried out on an Illumina core-exome chip. After quality controls, including SNP call rates  $\geq 95\%$ , minor allele frequencies (MAFs)  $\geq 1\%$ , and sample call rates  $\geq 95\%$ , identity-by-descent (IBD)  $\leq 0.1$ , without sex mismatches, duplicates, and heterozygosity outliers by eye from distribution, 273,113 SNPs was available for imputation. IMPUTE2 was used for imputation based on 1000 Genomes Project March 2012 version.

Further exclusions included  $p$  for Hardy–Weinberg equilibrium  $\leq 1.0 \times 10^{-6}$  and imputation info  $\leq 0.4$ <sup>6</sup>.

**Cardiovascular Risk in Young Finns Study (YFS):** The Cardiovascular Risk in Young Finns Study (YFS) is a population based prospective cohort study. It was conducted at five medical schools in Finland (Turku, Helsinki, Kuopio, Tampere and Oulu) with the aim of studying the levels of cardiovascular risk factors in children and adolescents in different parts of the country. The latest follow-up was conducted in 2007 at which serum samples were used for metabolomics analyses. The study and data collection protocols have been described in detail previously<sup>7</sup>. Genome-wide SNP data were generated from a custom Illumina BeadChip containing 670,000 SNPs and CNV probes. The custom content on the custom 670K array replaced some poor performing SNPs on the Human610 BeadChip and added more CNV content, and includes 546,677 SNPs passing QC from 594,210 SNPs on the chip. The custom 670K chip shares 562,643 SNPs in common with the Illumina Human610 BeadChip. Genotypes were called using Illumina's clustering algorithm. A total of 2,556 samples were genotyped. After initial clustering, we removed 2 subjects for poor call rates ( $CR < 0.90$ ), and 54 samples failed subsequent QC filters (i.e., duplicated samples, heterozygosity, low call rate, or custom SNP fingerprint genotype discrepancy). The following filters were then applied to the remaining data: MAF 0.01, GENO 0.05, MIND 0.05, and HWE  $1 \times 10^{-6}$ . Three individuals were removed for low genotyping (MIND > 0.05), 11,766 markers were excluded based on HWE test ( $P \leq 1 \times 10^{-6}$ ), 7,746 SNPs failed missingness test (GENO > 0.05), 34,596 SNPs failed frequency test (MAF < 0.01), and one individual failed gender check. A final list of 546,677 SNPs passed QC and allele frequency filters<sup>8</sup>. IMPUTE2 was used for imputation based on 1000 Genomes

Project March 2012 version. Further exclusions included  $p$  for Hardy–Weinberg equilibrium  $\leq 1.0 \times 10^{-6}$  and imputation info  $\leq 0.4^6$ .

**Northern Finland Birth Cohort (NFBC):** The Northern Finland Birth Cohorts were initiated 20 years apart in 1966 (NFBC66) and 1986 (NFBC86) to examine risk factors involved in pre-term birth and intrauterine growth retardation, and the consequences of these early adverse outcomes on subsequent morbidity and mortality, as described in detail previously<sup>9</sup>. Mothers living in the two northern-most provinces of Finland (Oulu and Lapland) were invited to participate if they had expected delivery dates during 1966 or 1986. Individuals still living in the Helsinki area or Northern Finland were asked at age 31 to participate in a detailed biological and medical examination as well as a questionnaire. GWAS analyses for circulating glycine levels, as measured by NMR, were carried out in 4,483 and 3,112 from the NFBC66 and 1986 NFBC86 studies, respectively. Genomic DNA was extracted from whole blood using standard methods and samples were genotyped on the Illumina Infinium 370cnvDuo array at the Broad Institute Biological Sample Repository. All individuals in the study were genotyped with call rates  $>95\%$ . Individuals with discrepancy between their reported sex and the sex determined from the X chromosome were excluded from analysis. The identity-by-descent (IBD) analysis option of PLINK45 was used to determine possible relatedness among sample subjects and identify sample duplications and sample contamination (the latter identified as individuals who seemed to be related to nearly everyone in the sample). If the sample duplication issue could not be resolved by external means, both samples were excluded. All apparently contaminated samples were also excluded. For pairs of individuals identified to be related at the level of half-sibs or closer in the IBD analysis, the subject with less complete genotyping was excluded. Variants were excluded from the analysis if the call rate in the final sample was  $<95\%$ , if the  $P$  value from a test of



Hardy-Weinberg Equilibrium (HWE) was  $<0.0001$ , or if the MAF was  $<1\%$ <sup>10</sup>. This resulted in 335,118 SNPs that were available for imputation. IMPUTE2 was used for imputation based on 1000 Genomes Project March 2012 version, with further exclusions for p for Hardy–Weinberg equilibrium  $\leq 1.0 \times 10^{-6}$  and imputation info  $\leq 0.4$ <sup>6</sup>.

**The Metabolic Syndrome in Men (METSIM) Study.** METSIM is a population-based study that recruited 10,197 Finnish men from the city of Kuopio in Eastern Finland between 2005-2010. The aims of METSIM are to investigate nongenetic and genetic factors associated with the risk of type 2 diabetes and cardiovascular disease, and with cardiovascular risk factors<sup>11</sup>. The protocol included a detailed phenotyping of the participants, an oral glucose tolerance test, fasting laboratory measurements, including proton NMR measurements, mass spectrometry metabolomics, as well as adipose tissue biopsies and stool samples in a subset of participants. Participants were genotyped on the Human OmniExpress-12v1\_C BeadChip (OmniExpress) and Infinium HumanExome-12 v1.0 BeadChip (Exome Chip) platforms. Quality controls included sample-level controls for sex and relatedness confirmation, sample duplication, and detection of sample genetic ancestry outliers using principal component analysis. Based on these quality control measures, 14 samples with sex chromosome anomalies, 18 with evidence of participant duplication, 12 population outliers, and 9 samples with non-Mendelian inheritance inconsistencies were removed. In addition, one individual from each of seven monozygotic twin pairs was removed. Variants with low mapping quality of probes to genome build GRCh37, low genotype completeness ( $<95\%$  and  $<98\%$  for the OmniExpress and ExomeChip, respectively), or Hardy-Weinberg equilibrium  $P < 10^{-6}$  were also filtered out. OmniExpress variants passing quality control with SHAPEIT v2 were phased and imputed using minimac v2. For imputation, a

reference panel of 20.9M variants from the GoT2D study (including SNVs, indels and large deletions) based on the whole genome sequence of 2874 Europeans, including 1004 Finnish individual, was used. Following imputation, variants directly genotyped on the ExomeChip were added. In cases of common markers between imputed and genotyped variants, the directly genotyped calls from the ExomeChip were used. Subsequently, 16,607,533 variants with high imputation quality (i.e. minimac RSQ0.3) were carried forward for single-variant association testing. GWAS analyses for circulating glycine levels, as measured by NMR, were carried out in a subset of 8545 non-diabetic men as described previously<sup>12</sup>. The institutional review boards of the University of Kuopio and Kuopio University approved the METSIM study. Written informed consent was obtained from each participant.

**Supplemental Table Legends (see Excel file):**

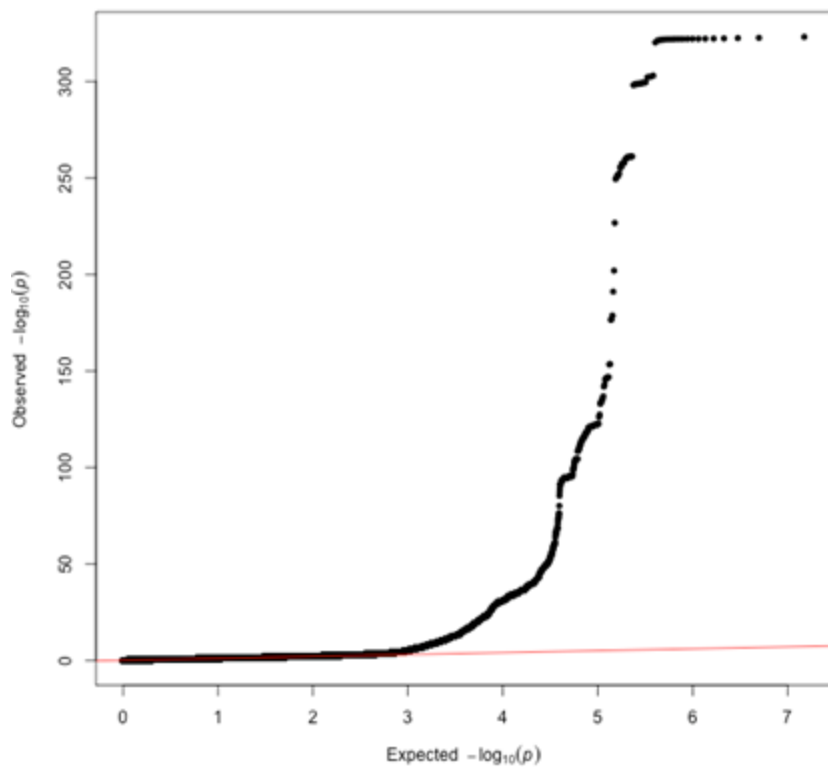
**Table S1.** Results of 12 Loci Significantly Associated with Circulating Glycine Levels Stratified by Metabolomics Platform.

**Table S2.** Results of 12 Loci Significantly Associated with Circulating Glycine Levels Stratified by Sex.

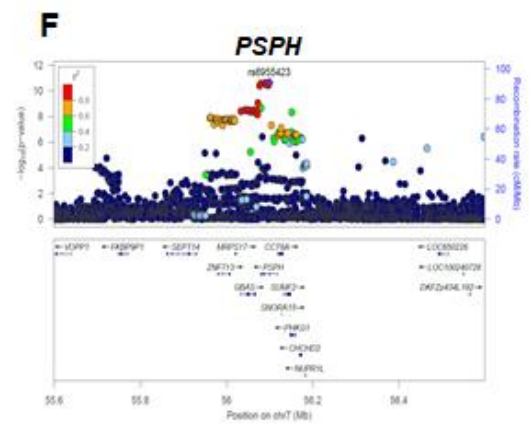
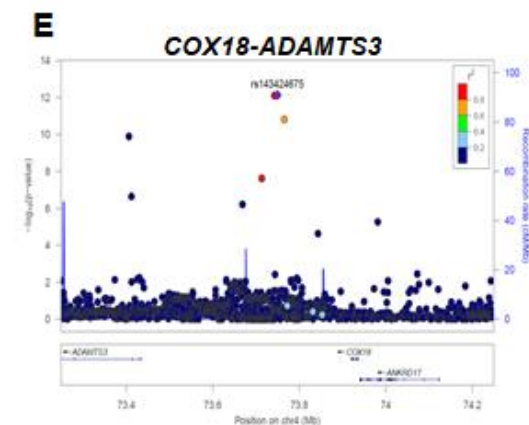
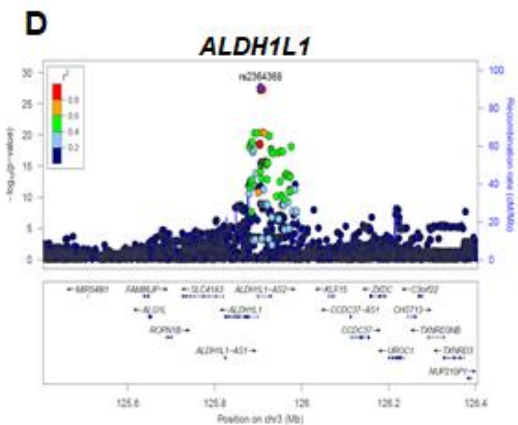
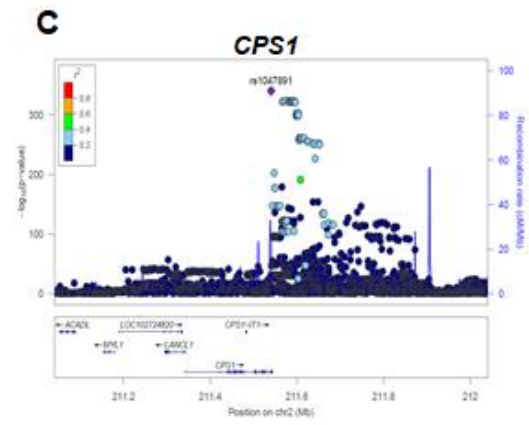
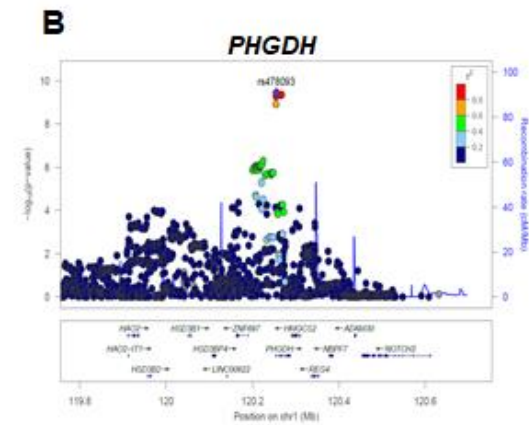
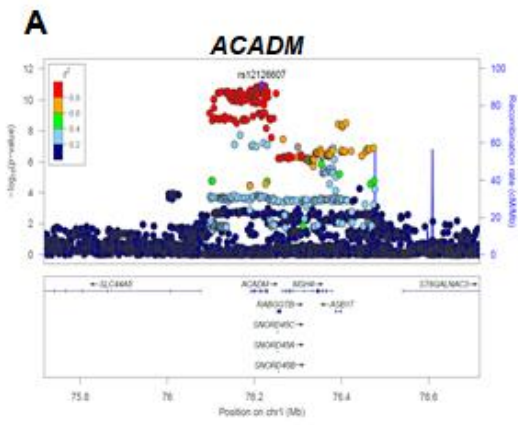
**Table S3.** PheWAS Results for 12 Loci Significantly Associated with Circulating Glycine Levels.

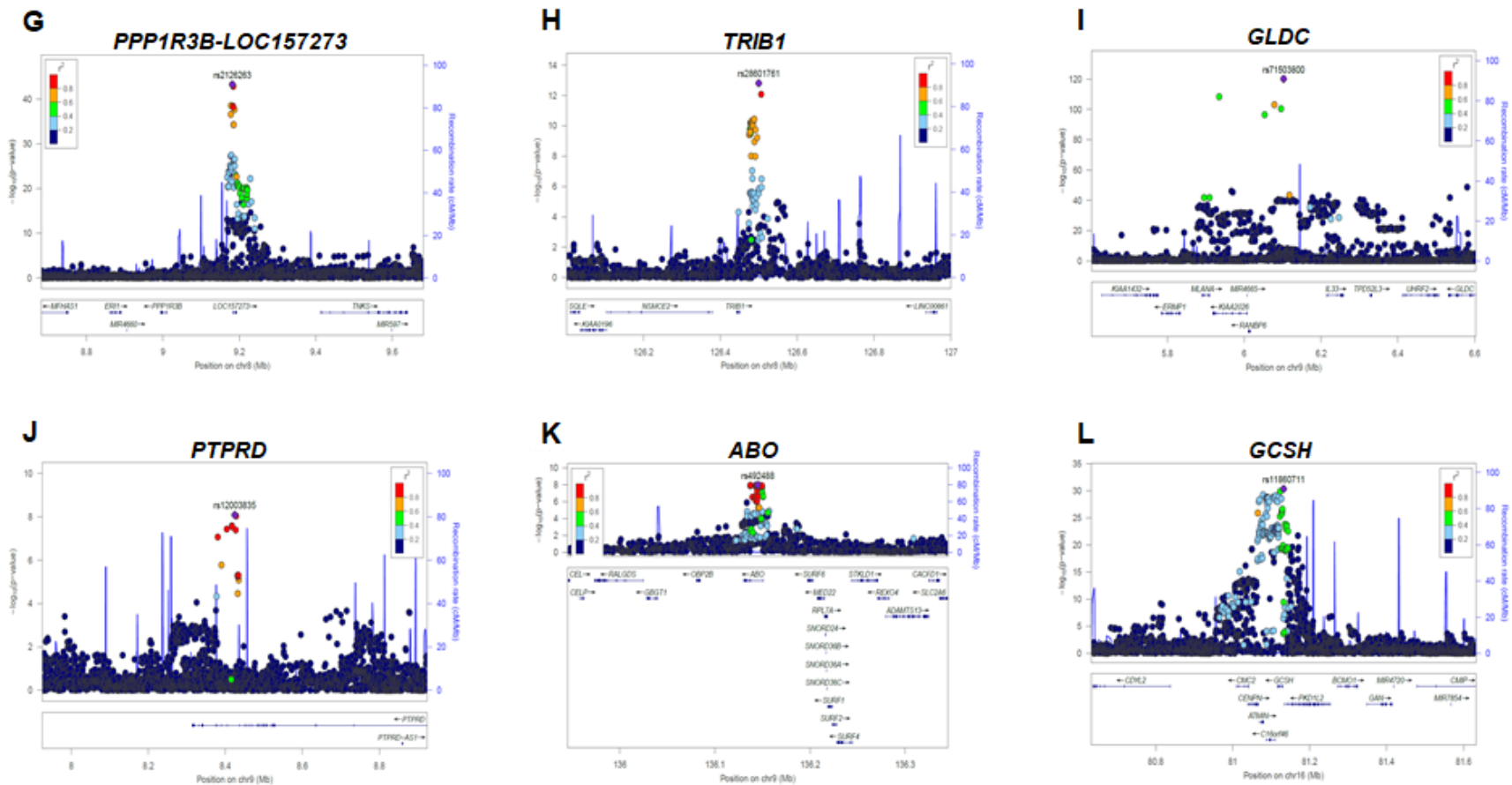
**Table S4.** Association of 12 Glycine-associated Loci with CAD in CARDIoGRAM+C4D and UK Biobank.

**Table S5.** Individual and Joint SNP Effect Associations and Mendelian randomization analysis of Glycine-associated Loci with Traditionally CAD Risk Factors.

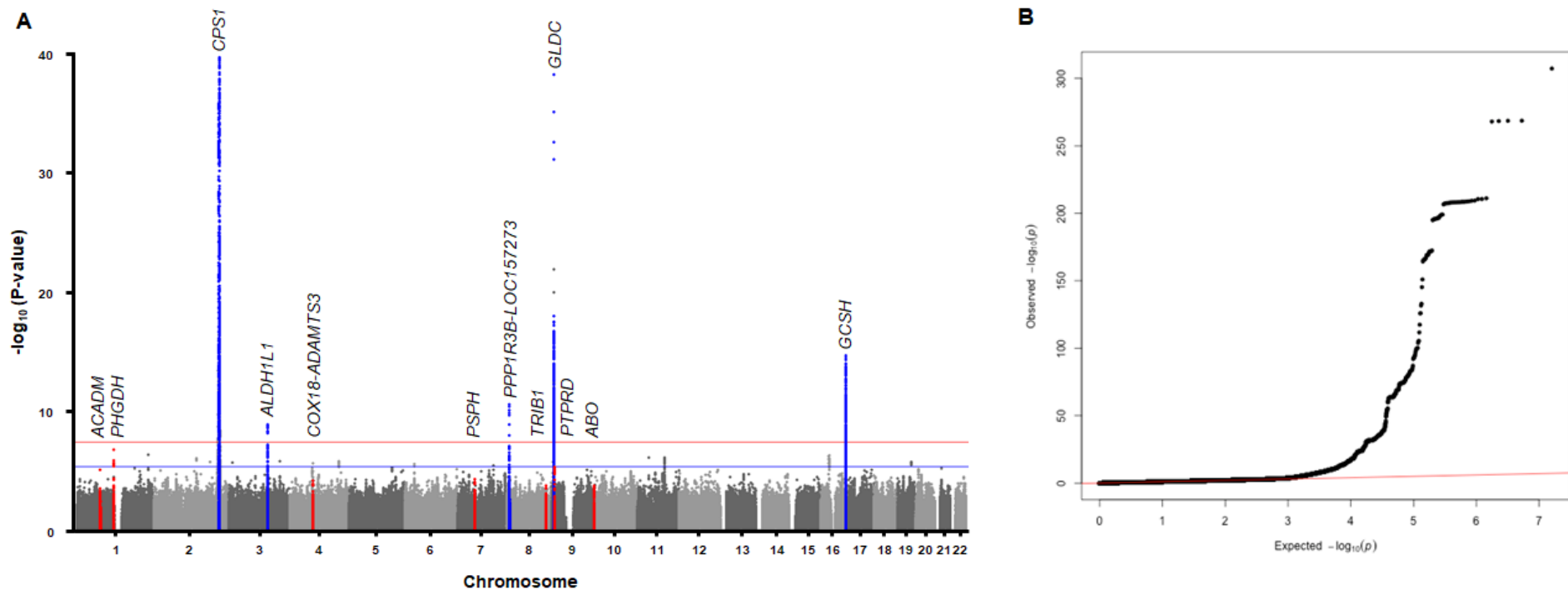


**Figure S1. Quantile-quantile (Q-Q) plot of GWAS meta-analysis results for circulating glycine levels in 30,118 subjects.** The observed versus the expected p-values from the meta-analyses for glycine levels are shown in the Q-Q plot. These analyses yielded a genomic inflation factor ( $\lambda$ ) of 1.035, indicating that the GWAS meta-analyses were not confounded by underlying population stratification.

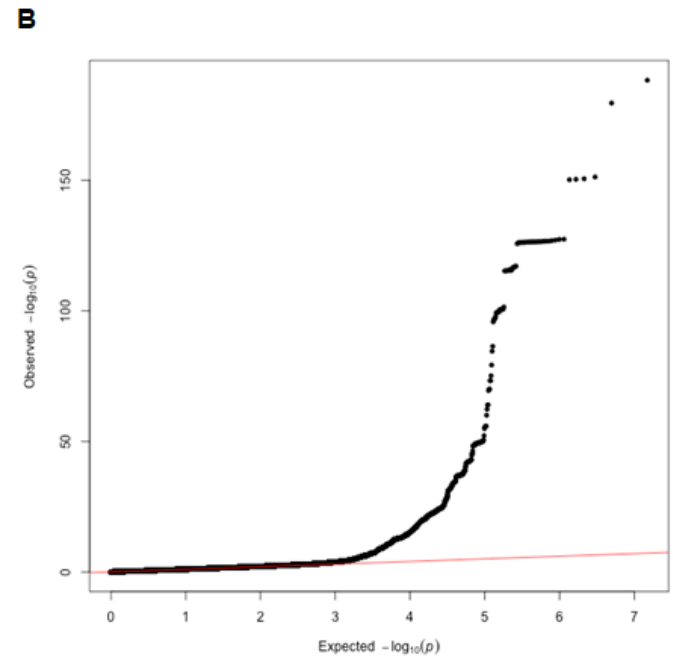
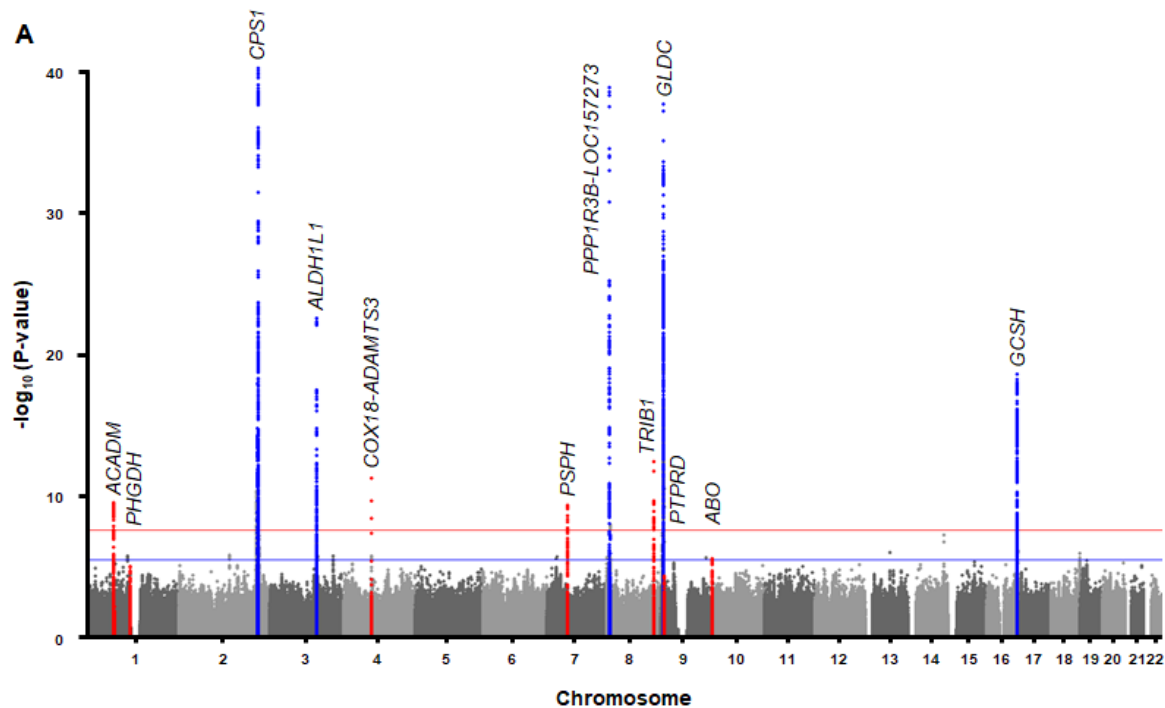




**Figure S2. Twelve loci identified for circulating glycine associated levels.** Regional plots for the *ACADM*, *PHGDH*, *CPS1*, *ALDH1L1*, *COX18-ADAMTS3*, *PSPH*, *PPP1R3B-LOC157273*, *TRIB1*, *GLDC*, *PTPRD*, *ABO*, and *GCSH* loci are shown in panels A-L. Each region is centered on the lead SNP (purple diamond) and the genes in the interval are indicated in the bottom panel. The degree of linkage disequilibrium (LD) between the lead SNP and other variants is shown as  $r^2$  values according to the color-coded legend in the box.

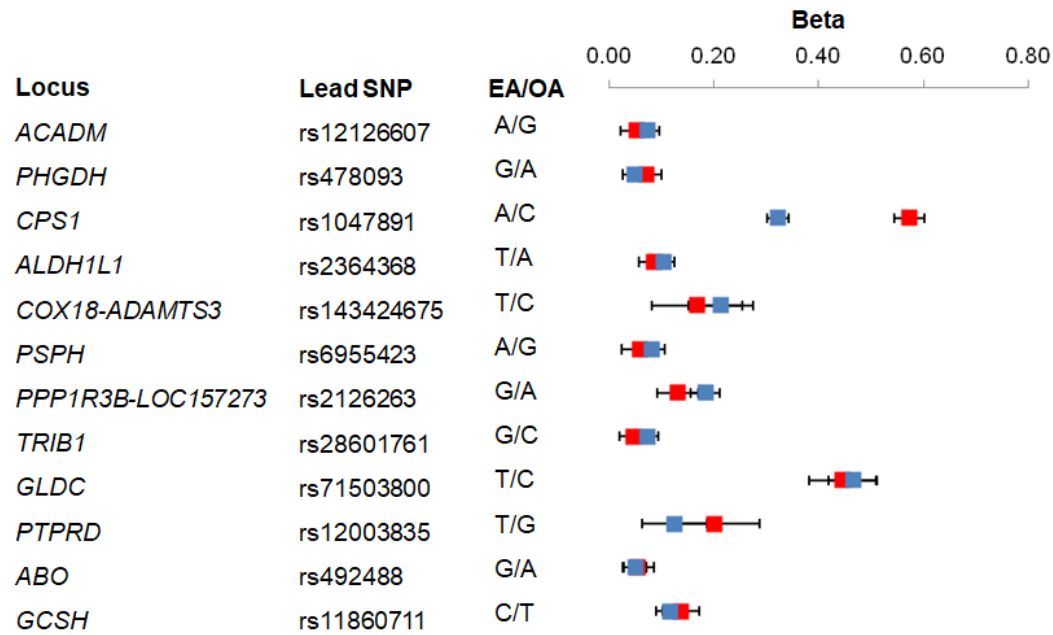


**Figure S3. Results of GWAS meta-analysis for circulating glycine levels in women.** (A) The Manhattan plot shows five previously identified loci significantly associated with circulating glycine levels (blue dots) in a stratified GWAS analysis with 10,886 women. Red dots indicate association signals for the seven novel identified in our meta-analysis with all 30,118 subjects, all of which were only suggestively associated in women. Genome-wide thresholds for significant ( $P=5.0 \times 10^{-8}$ ) and suggestive ( $P=5.0 \times 10^{-6}$ ) association are indicated by the horizontal red and dark blue lines, respectively. P-values are truncated at  $-\log_{10}(P)=40$ . (B) The Q-Q plot shows the observed versus the expected p-values from the meta-analyses for glycine levels in women. These analyses yielded a genomic inflation factor ( $\lambda$ ) of 1.002, indicating that the GWAS meta-analyses were not confounded by underlying population stratification.



**Figure S4. Results of GWAS meta-analysis for circulating glycine levels in men.** (A) The Manhattan plot shows nine loci significantly associated with circulating glycine levels in a stratified GWAS analysis with 19,004 men. The five loci identified in previous studies are indicated by blue dots. The red dots indicate association signals at the seven novel identified by our meta-analysis with all 30,118 subjects, of which four were also significant in only men. Genome-wide thresholds for significant ( $P=5.0 \times 10^{-8}$ ) and suggestive ( $P=5.0 \times 10^{-6}$ ) association are indicated by the horizontal red and dark blue lines, respectively. P-values are truncated at  $-\log_{10}(P)=40$ . (B) The Q-Q plot shows the observed versus the expected p-values from the meta-analyses for glycine levels in men. These analyses yielded a genomic inflation factor ( $\lambda$ ) of 1.035, indicating that the GWAS meta-analyses were not confounded by underlying population stratification.





**Figure S5. Sex-stratified results for 12 loci identified for circulating glycine levels.** Effect sizes for the lead SNPs at the 12 loci identified for circulating glycine levels are shown in men (blue) and women (red) separately. With the exception of *CPS1*, which is associated with approximately two-fold higher glycine levels in women compared to men, effect sizes at the 11 other loci were similar in males and females. EA, effect allele; OA, other allele.

## Supplemental References:

1. Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, Feldstein AE, Britt EB, Fu X, Chung YM, Wu Y, Schauer P, Smith JD, Allayee H, Tang WH, DiDonato JA, Lusis AJ and Hazen SL. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature*. 2011;472:57-63.
2. Koeth RA, Wang Z, Levison BS, Buffa JA, Org E, Sheehy BT, Britt EB, Fu X, Wu Y, Li L, Smith JD, DiDonato JA, Chen J, Li H, Wu GD, Lewis JD, Warrier M, Brown JM, Krauss RM, Tang WH, Bushman FD, Lusis AJ and Hazen SL. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat Med*. 2013;19:576-85.
3. Tang WH, Wang Z, Levison BS, Koeth RA, Britt EB, Fu X, Wu Y and Hazen SL. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N Engl J Med*. 2013;368:1575-84.
4. Hartiala JA, Tang WH, Wang Z, Crow AL, Stewart AF, Roberts R, McPherson R, Erdmann J, Willenborg C, Hazen SL and Allayee H. Genome-wide association study and targeted metabolomics identifies sex-specific association of CPS1 with coronary artery disease. *Nat Commun*. 2016;7:10558.
5. Vartiainen E, Laatikainen T, Peltonen M, Juolevi A, Mannisto S, Sundvall J, Jousilahti P, Salomaa V, Valsta L and Puska P. Thirty-five-year trends in cardiovascular risk factors in Finland. *Int J Epidemiol*. 2010;39:504-18.
6. Kettunen J, Demirkan A, Wurtz P, Draisma HH, Haller T, Rawal R, Vaarhorst A, Kangas AJ, Lyytikainen LP, Pirinen M, Pool R, Sarin AP, Soinen P, Tukiainen T, Wang Q, Tiainen M, Tynkkynen T, Amin N, Zeller T, Beekman M, Deelen J, van Dijk KW, Esko T, Hottenga JJ, van Leeuwen EM, Lehtimaki T, Mihailov E, Rose RJ, de Craen AJ, Gieger C, Kahonen M, Perola M, Blankenberg S, Savolainen MJ, Verhoeven A, Viikari J, Willemsen G, Boomsma DI, van Duijn CM, Eriksson J, Jula A, Jarvelin MR, Kaprio J, Metspalu A, Raitakari O, Salomaa V, Slagboom PE, Waldenberger M, Ripatti S and Ala-Korpela M. Genome-wide study for circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA. *Nat Commun*. 2016;7:11122.
7. Raitakari OT, Juonala M, Ronnema T, Keltikangas-Jarvinen L, Rasanen L, Pietikainen M, Hutri-Kahonen N, Taittonen L, Jokinen E, Marniemi J, Jula A, Telama R, Kahonen M, Lehtimaki T, Akerblom HK and Viikari JS. Cohort profile: the cardiovascular risk in Young Finns Study. *Int J Epidemiol*. 2008;37:1220-6.
8. Smith EN, Chen W, Kahonen M, Kettunen J, Lehtimaki T, Peltonen L, Raitakari OT, Salem RM, Schork NJ, Shaw M, Srinivasan SR, Topol EJ, Viikari JS, Berenson GS and Murray SS. Longitudinal genome-wide association of cardiovascular disease risk factors in the Bogalusa heart study. *PLoS Genet*. 2010;6:e1001094.
9. Rantakallio P. The longitudinal study of the northern Finland birth cohort of 1966. *Paediatr Perinat Epidemiol*. 1988;2:59-88.
10. Sabatti C, Service SK, Hartikainen AL, Pouta A, Ripatti S, Brodsky J, Jones CG, Zaitlen NA, Varilo T, Kaakinen M, Sovio U, Ruokonen A, Laitinen J, Jakkula E, Coin L, Hoggart C, Collins A, Turunen H, Gabriel S, Elliot P, McCarthy MI, Daly MJ, Jarvelin MR, Freimer NB and Peltonen L. Genome-wide association analysis of metabolic traits in a birth cohort from a founder population. *Nat Genet*. 2009;41:35-46.

11. Laakso M, Kuusisto J, Stancakova A, Kuulasmaa T, Pajukanta P, Lusi AJ, Collins FS, Mohlke KL and Boehnke M. The Metabolic Syndrome in Men study: a resource for studies of metabolic and cardiovascular diseases. *J Lipid Res.* 2017;58:481-493.
12. Teslovich TM, Kim DS, Yin X, Stancakova A, Jackson AU, Wielscher M, Naj A, Perry JRB, Huyghe JR, Stringham HM, Davis JP, Raulerson CK, Welch RP, Fuchsberger C, Locke AE, Sim X, Chines PS, Narisu N, Kangas AJ, Soininen P, Genetics of Obesity-Related Liver Disease Consortium TAsDGCTDGR, Meta a, Ala-Korpela M, Gudnason V, Musani SK, Jarvelin MR, Schellenberg GD, Speliotes EK, Kuusisto J, Collins FS, Boehnke M, Laakso M and Mohlke KL. Identification of seven novel loci associated with amino acid levels using single-variant and gene-based tests in 8545 Finnish men from the METSIM study. *Hum Mol Genet.* 2018;27:1664-1674.