1	Oestrogenic activities of food supplements and beers as assessed by a yeast
2	bio-reporter assay
3	Iyekhoetin Matthew Omoruyi ^{1,*} and Raimo Pohjanvirta ²
4 5	¹ Department of Basic Sciences, Faculty of Basic and Applied Sciences, Benson Idahosa University, Benin City, Edo State, Nigeria
6 7	² Department of Food Hygiene and Environmental Health, Faculty of Veterinary Medicine, University of Helsinki, Finland
8	*Corresponding author: iomoruyi@biu.edu.ng; +2348062764607
9	
10	
11	
12	
13	
14	
15	
10	
16	
17	
18	
19	
20	

21 Abstract

22 Mounting evidence of the effects of endocrine-disrupting chemicals (EDCs) in humans has led to assaying a vast array of food items (processed or packaged) as possible sources of human 23 24 exposure to oestrogens. In this study, we investigated the current situation in this respect of 25 different food supplements and beer brands. Eleven different food supplements and twenty-four 26 beer brands were obtained from Helsinki, Finland. Sample preparation was carried out by established methods while oestrogenic activities were assessed by a yeast bioluminiscent assay, 27 using two recombinant yeast strains (Saccharomyces cerevisiae BMAEREluc/ERa and S. 28 29 cerevisiae BMA64/luc). All the food supplements as well as 81% of the beer samples tested were 30 found to be oestrogenic, with oestradiol equivalent concentrations of food supplements and beer 31 brands ranging from 7.5 to $11.5 \,\mu$ g/ml and from below detection limits to 43.6 ng/ml, respectively. 32 The oestrogenic activities detected in beer samples were not dependent on the beer's alcoholic 33 content, country of production, or the size of the production brewery. The results of our study 34 imply that both food supplements and beers can be a significant source of human exposure to 35 oestrogens. Therefore, further studies and regular surveillance are warranted.

36

37

38

39

41	Keywords: Endocrine-disrupting chemicals; oestrogenic activity; bioassay; beer; phytoestrogens;
42	isoflavones; food supplements; isoxanthohumol
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	

59 Introduction

60 Food supplements are concentrated sources of nutrients or other substances with nutritional 61 or physiological effect, whose purpose is to supplement the normal diet. They are originally used 62 to correct nutritional deficiencies and to maintain adequate intakes of certain nutrients. In the European Union (EU), food supplements are regulated as foods, and the legislation focuses on 63 vitamins and minerals used as ingredients in food supplements, thus making the toxicological 64 65 safety of food supplements an inconclusive and somewhat controversial issue. However, food 66 supplements have been reported to be of potential public health concern, due to, for example, the presence of and/or contamination by endocrine-disrupting chemicals (EDCs), which may be 67 68 natural (such as phytoestrogens) or synthetic. Recent evidence has shown that food supplements 69 can be contaminated with low quantities of steroids or stimulants (such as ephedrine) not specified 70 on the label. A broad-based investigation of the international nutritional supplement market 71 revealed that between 3 to 25% of dietary food supplements were inadvertently or deliberately 72 contaminated with steroid hormones [1]. In several studies, the presence of pro-hormones 73 (supplements designed to enhance muscle size and strength rapidly) in non-hormonal nutritional 74 supplements has been examined. Results from an international study, involving the European 75 Union and the United States, showed that 14.8% (94 out of 634 samples) of the investigated non-76 hormonal supplements contained one or more anabolic steroids not declared on the label [2].

Another source of compounds with hormonal activity in nutritional supplements is plantderived phytoestrogens. At present, controversy remains as to the health impacts of phytoestrogens. While epidemiological studies have shown that phytoestrogens (such as genistein) may protect against breast cancer and cardiovascular diseases [3] (although genetic composition

seems to play a notable role for breast cancer [4]), experimental studies have suggested they could
enhance the proliferation or metastasis of some types of cancer [5].

83 On the other hand, beer is consumed in increasing volumes globally, and has recently 84 become more controversial as a source of dietary exposure to oestrogens. The oestrogenic activity 85 of beer is due to the presence/use of hops (Humulus lupulus L.) both as a preservative and as a 86 flavouring agent in it [6]. The oestrogenic activity of hops was first attributed to xanthohumol 87 without convincing evidence [7]. Today, it is known that in addition to xanthohumol, hops also 88 contain isoxanthohumol (IX), 6-prenylnaringerin (6-PN) and 8-prenylnaringerin (8-PN) as some 89 of its major constituents. The most potent phytoestrogen of them is 8-PN, with its oestrogenic 90 activity being equal to, or greater than that of other established plant oestrogens [6].

91 Although xanthohumol is the predominant prenylchalcone (prenylated flavonoids found in 92 hops and beer) present in beer, most of it is transformed into IX by thermal isomerization during 93 wort boiling [8]. IX is further converted to 8-PN in the gastrointestinal tract by the intestinal 94 microbe, *Eubacterium limosum*. Thereby, the human exposure to 8-PN may grow more than 10-95 fold [9]. These observations have led to intense research aiming at deciphering the bioactivities of 96 the ultimate metabolite, 8-PN. In vivo, 8-PN showed estrogenic activity [10], inhibited 97 angiogenesis [11] and metastasis [12], exhibited antiandrogenic activity [13], as well as prevented 98 bone loss in rats [14].

A number of yeast bioluminescent assays have been developed in recent times to detect the
 presence of endocrine disrupting chemicals in different matrices. However, the yeast
 bioluminescent assay employed in this study uses two recombinant yeast strains (*Saccharomyces cerevisiae* BMAEREluc/ERα and *S. cerevisiae* BMA64/luc). The yeast strain *S. cerevisiae* BMA64/luc helps to detect the cytotoxic effect of the test compound because it expresses

luciferase constitutively, while the human oestrogen receptor–agonist complex is activated in *S*.
 cerevisiae BMAEREluc/ERα upon binding of an oestrogenic compound, thus resulting in light
 emission. These assays are faster than corresponding colorimetric assays, taking only a few hours
 to be performed.

To shed more light on these issues, the current study sought to determine the oestrogenic activities of a representative selection of food supplements and beer brands marketed in Finland, with a view to furthering our understanding of their potential as a source of human exposure to oestrogens.

- . _ .

128 Materials and methods

129 <u>Sample collection and preparation</u>

130 *Food supplements*

Fifteen different food supplements were randomly purchased from a local shop (Prisma, Viikki) in Helsinki, Finland. All food supplements investigated were packaged as tablets. Samples were prepared for oestrogenic assay by directly dissolving the tablets and capsule in 10 % ethanol. To be sure that the 10 % ethanol used as vehicle was not cytotoxic to the yeast, it was used as a negative control substance in the yeast bioluminescent assay. All food supplement products were readily and completely dissolved in ethanol.

137 *Beer samples:*

A total of twenty-one different brands of beer and 3 controls (unhoped beer samples) were investigated. The selection of beer samples was based on the following grounds: beers produced in the two continents with the highest beer consumption (Europe and the USA), beer production style (e.g. larger, pilsner, ale, etc.), alcoholic content, brewery size, anticipated oestrogenic contents, and hopping style. Our idea was to get an as broad and representative sample of the beer brands currently on the market in Finland as possible regarding all these properties.

Possible oestrogenic compounds in beer samples were extracted by the solid-phase extraction method as previously described [15], with the slight modification that 100 ml of the samples were first mixed with 25 ml of ethanol (extraction solvent) to increase the extraction efficiency of the test system. All beer products mixed readily and completely with ethanol.

In addition to the different beer brands, Menohop (a product developed using hops for the treatmentof menopause-related problems in women) was also analysed for comparison.

150 <u>Yeast bioluminescent assay</u>

151 The yeast bioluminescent assay was performed as previously described [16]. Briefly, yeast strains (S. cerevisiae BMAEREluc/ERa and S. cerevisiae BMA64/luc) were grown overnight until the 152 153 optical density at a wave length of $600 (OD_{600})$ reached 0.4 cfu/ml. Necessary dilutions were made, 154 and the yeast strains were further grown at 30°C in a shaker for 2.5 hours until OD600 reached 155 0.6. Different concentrations of the test compounds, the yeast strains, as well as 10 x luciferin solutions were placed in a micro-well-plate and then incubated at 30°C for 2.5 hours before the 156 157 luciferase activity was measured. Oestradiol, genistein and IX served as positive controls, while 158 progesterone and testosterone served as negative controls. The data are given as oestradiol (EEQ) 159 or genistein equivalents.

160 <u>Data analysis</u>

The fold induction, fold induction corrected (FIC), and limit of detection (LOD) were calculated as described previously [17]. Sigmoidal dose–response curves for increasing concentrations of oestradiol, genistein and isoxanthohumol were obtained using the software Prisma 5.0 (GraphPad software Inc. San Diego, CA). The oestradiol, genistein and IX equivalents of food samples showing oestrogenic activity were calculated from probit transformation of the curves.

- 166
- 167
- 168
- 169
- 170
- 171
- 172
- 173

174 Results and Discussion

175 Recent evidence of a decline in male sperm count coupled with increased incidence of 176 various types of cancer amongst young men and women [18], as well as of neurobehavioural 177 diseases observed in the populations of different countries [19] have led researchers to search for 178 possible causes of such inauspicious human conditions. These phenomena have epidemiologically 179 been associated with exposure to EDCs, particularly during the intrauterine phase or during critical 180 periods of postnatal development [20]. More recently, some food items (including those of plant 181 origin) have begun to gain increasing attention as possible sources of human exposure to xeno-182 and phytoestrogens. Thus, our study was aimed at evaluating the current situation of food 183 supplements and beers with respect to their oestrogenic activities.

The oestrogenic activities of food supplements measured are presented in Table 1. All food supplements (15) investigated in this study were positive in the yeast bioluminescent assay, with their oestradiol and genistein equivalent concentrations ranging from 7.9 to 11.5 μ g/ml and 74 to 240 μ g/mg, respectively. These narrow ranges obtained with all samples suggest the presence of a common or similar substance/compound in slightly varying concentrations.

189 Table 2 shows the oestradiol and IX equivalent concentrations of various brands of 190 commercial beer produced in different countries. Seventeen (81%) out of the 21 beer samples 191 examined were positive in the test system. The oestradiol and IX equivalent concentrations of the 192 tested beer samples were not dependent on their alcohol content, brewery size or country of 193 production (Table 2). Beer sample 9 (BS9), with alcoholic content of 4.6% and produced in the 194 Netherlands, had the highest oestrogenic activity (43.6 ng/ml EEQ), followed by BS20 (10.5% 195 ethanol, produced in Denmark), whose oestrogenic activity was 31.8 ng/ml EEQ. Four (BS3, 196 BS12, BS15 and BS19) out of the 21 tested beer samples were negative in the yeast

bioluminescence assay. Although the reason for the low oestrogenicity in these cases is somewhat unclear, the outcome was not associated with their alcoholic content or their country of origin [Finland (3) and Germany (1)]. Interestingly, all unhoped beer samples (3) used as control in this study failed to yield any oestrogenic response, further substantiating the notion that the hop plant is the major source of the oestrogenicity in beer.

202 IX was previously reported to be non-oestrogenic in a similar yeast bioreporter assay [21]. 203 Although Milligan *et al.* [21] did not report the concentration used in that study, we, however, 204 found IX to be oestrogenic at concentrations of 1 mg/ml or greater. Below this concentration, IX 205 was found to lose its oestrogenic activity. Using phenobarbital-induced rat liver microsomal 206 enzyme mix (S9), we therefore investigated the possibility that IX might gain oestrogenicity at 207 low concentrations following metabolic activation; however, we failed to find any (data not 208 shown). The results of this study thus suggest that IX has relatively weak oestrogenic potency and 209 is non-oestrogenic at low concentrations with or without metabolic activation, but it should be kept 210 in mind that the rat liver S9 mix may not contain the enzyme activity critical for IX conversion to 211 8-PN; *i.e.* the reaction that takes place in human distal colon [22].

212 The oestrogenic activities of beer samples were not unexpected but warrant attention. The 213 oestrogenicity of beer is associated with the presence of IX and 8-PN [6], of which 8-PN is an 214 exceptionally potent phytoestrogen, being only about 100 times weaker activator of oestrogen 215 receptor- α in a yeast reporter-gene assay than oestradiol [23]. Exposure to 8-PN has previously 216 been linked to menstrual disturbances in female hop workers [7], and it also reduces hot flushes in 217 post-menopausal women. Apart from drugs produced from hops (such as the Menohop used here as a positive control), beer consumption is today the only appreciable source of human exposure 218 219 to IX and 8-PN, since hop-picking is now performed mechanically [6]. Although exposure to

220 especially 8-PN has aroused concern due to the high oestrogenic potency of this compound, 221 Milligan et al. [6] have argued that the concentrations of IX and 8-PN in beer are not sufficient to 222 cause any detrimental health effects. However, Possemiers et al. [22] have contended that intestinal 223 conversion of IX upon moderate beer consumption can lead to 8-PN exposure values that might 224 fall within the range of human biological activity. It is noteworthy that the highest oestrogenic 225 activities of beer brands measured in this study (per l beer) fall only slightly lower than that of 226 Menohop (per g), a widely-used hop-based product for the treatment of menopause-related 227 problems in women.

228 Earlier, Plotan et al. [24] reported that 71–89% of sport supplements exhibited oestrogenic 229 activity in an *in vitro* reporter-gene assay. Some other previous studies have examined isoflavone 230 contents and oestrogenic activities in food supplements intended for alleviating menopausal 231 complaints in women. One of such products was also included in our study (Menohop in Table 1). 232 Reiter et al. [25] reported that only 26.3% (five out of 19) of the high-dose isoflavone preparations 233 they analysed contained the isoflavone content or more specified in the package label. Using a 234 similar oestrogen receptor- α and reporter gene-based yeast assay to that of ours, they recorded 235 oestrogenic activities of up to 200 nmol/g (54 μ g/g) EEQ, which is practically equal to the level 236 measured by us in the Menohop preparation (Table 1). On the other hand, Andres et al. [26] found 237 that the isoflavone supplements they examined contained approximately the amounts of 238 isoflavones claimed by the manufacturers in their product information, and the highest oestrogen 239 receptor-α-mediated EEQ value (obtained by a human embryonic kidney cell-based reporter-gene 240 assay) was 11.6 µg/capsule. Judkins et al. [1] and Gever et al. [2] have also reported hormonal 241 steroid contaminants in various types of food and sport supplements. Interestingly, the contaminants were not listed in the packaging material of the supplements, further calling for 242

serious concern. Controversies exist as to the health impacts of isoflavones. The purported health
benefits are quite variable in different studies. While isoflavones are reported to lower total
cholesterol [27], reduce the incidence of breast cancer [28], and diminish the risk of prostate cancer
[29], they are also known to increase the incidences of goitre and thyroid enlargement in certain
nutrient-deficient individuals [30].

In conclusion, the findings imply that food supplements and commercially produced beer can be significant sources of human exposure to oestrogens. The oestrogenicity of beer was not dependent on the alcoholic content, country of production, or the brewery size. Because of the controversy concerning the effects of phytoestrogens on humans, further studies are warranted.

252

253 **Conflict of interest**

254 The authors declare that there are no conflicts of interest.

255

256 Acknowledgements

We are grateful to Hannu Kause for his insightful contributions and for providing us with the majority of beer brands including the hops-free control beer.

- 260
- 261

262 **References**

- [1] Judkins C, Teale P and Hall DJ. The role of banned substance residues analysis in the control
 of dietary supplement contamination. *Drug Test Anal* 2010; 2:417–420.
- 265 [2] Geyer H, Parr MK, Mareck U, Reinhart U, Schrander Y and Schanzer W. Analysis of non-
- hormonal nutritional supplements for anabolic–androgenic steroids Results of an
 international study. *J Sports Medic Physic Fitness* 2004; 25:124–129.
- [3] Pilsakova L, Riecansky I and Jagla F. The physiological actions of isoflavones phytoestrogens.
 Physiol Rev 2010; 59:651–664.
- [4] Chen M, Rao Y, Zheng Y, Wei S, Li Y, Guo T and Yin P. Association between soy isoflavone
 intake and breast cancer risk for pre- and post-menopausal women: a meta-analysis of
 epidemiological studies. *PLoS One* 2014; 20;9(2):e89288.
- [5] Martínez-Montemayor MM, Otero-Franqui E, Martinez J, De La Mota-Peynado A, Cubano
 LA and Dharmawardhane S. Individual and combined soy isoflavones exert differential
- effects on metastatic cancer progression. *Clin Exp Metastasis* 2010;27:465–480.
- [6] Milligan SR, Kalita JC, Heyerick A, Rong H, De Cooman L and De Keukeleire D.
 Identification of a potent phytoestrogen in hops (*Humulus lupulusL.*) and beer. *J Clin Endocrinol Metab* 1999; 84:2249-2252.
- [7] Verzele M. 100 years of hop chemistry and its relevance to brewing. *J Inst Brew* 1986; 92: 3248.
- [8] Santos MC, Salvador AC, Domingues FM, Cruz JM and Saraiva JA. Use of high hydrostatic
 pressure to increase the content of xanthohumol in beer wort. *Food Biop Technol* 2013;
 6:2478-2485.

284	[9] Possemiers SI, Heyerick A, Robbens V, De Keukeleire D and Verstraete W. Activation of
285	proestrogens from hops (Humulus lupulus L.) by intestinal microbiota; conversion of
286	isoxanthohumol into 8-prenylnaringenin. J Agric Food Chem 2005; 53:6281-6288.

- [10] Diel P, Thomae RB, Caldarelli A, Zierau O, Kolba S, Schmidt S, Schwab P, Metz P and
 Vollmer G. Regulation of gene expression by 8-prenylnaringenin in uterus and liver
 ofWistar rats. *Planta Med* 2004;70: 39-44.
- [11] Pepper MS, Hazel SJ, Humpel M and Schleuning WD. 8-prenylnaringenin, a novel
 phytoestrogen, inhibits angiogenesis *in vitro* and *in vivo*. *J Cell Physiol* 2004; 199: 98-107.

[12] Rong HJ, Boterberg T, Maubach J, Stove C, Depypere H, Van Slambrouck S, Serreyn R, De

292

- Keukeleire D, Mareel M and Bracke M. 8-Prenylnaringenin, the phytoestrogen in hops and beer, upregulates the function of the E-cadherin/catenin complex in human mammary carcinoma cells. *Eur J Cell Biol* 2001; 80: 580-585.
- Zierau O, Morrissey C, Watson RWG, Schwab P, Kolba S, Metz P and
 VollmerG.Antiandrogenic activity of the phytoestrogens naringenin, 6-(1,1 dimethylally(l)naringenin and 8-prenylnaringenin. *Planta Med* 2003; 69: 856-858.
- [14] Miyamoto M, Matsushita Y,Kiyokawa A, Fukuda C, Iijima Y, Sugano M and Akiyama T.
 Prenylflavonoids: A new class of nonsteroidal phytoestrogen (part 2). Estrogenic effects of
 8-isopentenylnaringenin on bone metabolism. *Planta Med* 1988; 64: 516-519.
- 302 [15] Omoruyi IM and Pohjanvirta R. Oestrogenic activities of wastewater, bottled waters and tap
 303 water in Finland as assessed by a yeast bio-reporter assay. *Scandin J Publ Health* 2015;
 304 43: 770-775

305	[16] Omoruyi IM, Kabiersch G and Pohjanvirta R. Commercial processed food may have
306	endocrine disrupting potential: soy-based ingredients making the difference. Food Addit
307	Contam 2013;30: 1722-1727.
308	[17] Leskinen P, Michelini E, Picard D, et al. Bioluminescentyeast assays for detecting estrogenic
309	and androgenic activity in different matrices. Chemosphere 2005;61:259-66.
310	[18] Feki NC, Abid N, Rebai A, Sellami A, Ayed BB, Guermazi M, Bahloul A, Rebai T and
311	Ammar LK. Semen quality decline among men in infertile relationships: experience over
312	12 years in the South of Tunisia. J Androl 2009;30: 541-547.
313	[19] Bellanger, M, Demeneix, B, Grandjean, P, Zoeller RT, Trasande L. Neurobehavioral
314	deficits, diseases, and associated costs of exposure to endocrine-disrupting chemicals in
315	the European Union. J Clin Endocrinol Metab. 2015;100(4):1256-66.
316	[20] Vaiserman, A. Early-life Exposure to Endocrine Disrupting Chemicals and Later-life Health
317	Outcomes: An Epigenetic Bridge? Aging Dis. 2014; 5(6): 419-429.
318	[21] Milligan SR, Kalita JC, Pocock V, Vab De Karter V, Stevens JF, Deinzer ML, Rong H and
319	De Keukeleire D. The endocrine activities of 8-prenylnaringenin and related hop (Humulus
320	lupulus L) flavonoids. J Clin Endocrinol Metab 2000; 85:4912-4915.
321	[22] Possemiers S1, Bolca S, Grootaert C, Heyerick A, Decroos K, Dhooge W, De Keukeleire D,
322	Rabot S, Verstraete W and Van de Wiele T. The prenylflavonoidisoxanthohumol from hops
323	(Humulus lupulus L.) is activated into the potent phytoestrogen 8-prenylnaringenin in vitro
324	and in the human intestine. J Nutr 2006;136:1862-1867.
325	[23] BoveeTF, Helsdingen RJ, Rietjens IM, Keijer J, Hoogenboom RL. Rapid yeast estrogen
326	bioassays stably expressing human estrogen receptors alpha and beta, and green fluorescent

- 327 protein: a comparison of different compounds with both receptor types. *J Steroid Biochem* 328 *Mol Biol* 2004;91(3):99-109.
- [24] Plotan M, Elliott CT, Scippo ML, Muller M, Antignac JP, Malone E, Bovee TF, Mitchell S,
 Connolly L.The application of reporter gene assays for the detection of endocrine
- disruptors in sport supplements. *Anal Chim Acta* 2011;700(1-2):34-40.
- Reiter E, Beck V, Medjakovic S, Mueller M and Jungbauer A. Comparison of hormonal
 activity of isoflavone-containing supplements used to treat menopausal complaints.
 Menopause 2009;16: 1049-1060.
- [26] Andres S, Hansen U, Niemann B, Palavinskas R and Lampen A. Determination of the
 isoflavone composition and estrogenic activity of commercial dietary supplements based
 on soy or red clover. *Food Funct* 2015;6:2017-2025.
- 338 [27] Anderson JW, Johnstone BM and Cook N.Meta-analysis of the effects of soy protein intake
 339 on serum lipids. *New Eng J Med* 1995; 33:276–82.
- [28] Shu XO, Jin F, Dai Q, Wen W, Potter JD, Kushi LH, Ruan Z, Gao YTandZheng W.
 Soyfoodintake during adolescence and subsequent risk of breast cancer among Chinese
 women. *Cancer Epidemiol Biomarkers Prev* 2001;10:483–8.
- [29] Nagata Y, Sonoda T, Mori M, Miyanaga N, Okumura K, Goto K, Naito S, Fujimoto K and
 Hirao Y. Dietary isoflavones may protect against prostate cancer in Japanese men. *J Nutr* 2007;137:1974–1979.
- [30] Xiao CW. Health effects of soy protein and isoflavones in humans. *J Nutri* 2008;136: 12441249.

• • •					
350	Supplement (code)	EEQ (µg/ml)	GEQ (µg/g)		
351	FS1	7.9	76		
352	FS2	11.5	240		
353	FS3	9.3	110		
354	FS4	10.7	190		
355	FS5	7.5	74		
356	FS6	10.2	170		
357	FS7	8.4	80		
358	FS8	7.9	76		
359	FS9	9.7	140		
360	FS10	10.9	200		
361	FS11	10.1	170		
362	FS12	8.1	79		
363	FS13	10.7	190		
364	FS14	10.1	170		
365	FS15	9.8	140		
366	Menohop	52.4 (µg/g)	NA		

Table 1: Oestrogenic activities in different food supplements expressed as oestradiol (EEQ) andgenistein (GEQ) equivalent concentrations

Key: EEQ: Oestradiol equivalent concentration; GEQ: Genistein equivalent concentration; NA:
Not applicable.

375	Beer (code)	Country	% OH	EEQ (ng/ml)	IXEQ (mg/ml)
376 377	Beer (code)	of production	70 OH		
378					
379	BS1	Finland	4.5	8.4	1.8
380	BS2	Finland	4.6	2.3	0.1
381	BS3	Finland	4.5	Nil	NA
382	BS4	USA		5.9	1.0
383	BS5	Germany		7.0	1.3
384	BS6	Czech Republic	4.4	21.4	4.1
385	BS7	Britain	3.5	29.6	4.9
386	BS8	Belgium	0.0	2.8	0.2
387	BS9	The Netherlands	4.6	43.6	7.4
388	BS10	Scotland	5.0	19.8	3.6
389	BS11	Mexico	4.5	14.8	2.7
390	BS12	Germany	4.7	Nil	NA
391	BS13	Ireland	4.2	2.8	0.2
392	BS14	Finland	5.2	17.4	3.1
393	BS15	Finland	7.2	Nil	NA
394	BS16	Denmark	7.2	10.8	1.1
395	BS17	Belgium	9.0	18.4	3.3
396	B18	Sweden	5.3	18.2	3.3
397	BS19	Finland	7.0	Nil	NA
398	BS20	Denmark	10.5	31.8	6.4
399	BS21	USA	7.3	14.5	2.0
400	Control beer 1*		Nil	Nil	Nil
401	Control beer 2		Nil	Nil	Nil
402	Control beer 3		Nil	Nil	Nil

Table 2: Oestrogenic activities in different beer samples expressed as oestradiol (EEQ) and IX
 (IXEQ) equivalent concentrations

403 <u>Kev</u>: OH: Alcohol; EEQ: Oestradiol equivalent concentration; IXEQ: IX equivalent concentration; NA; Not applicable; * The control beers were made without hops

407 408 409 410	BEER CODE	NAME OF BEER	COUNTRY OF PRODUCTION	PERCENTAGE ALCOHOL	TYPE OF BEER
411	BS1	Laitilan kukko	Finland	4.5	Pils
412	BS2	Karhu II	Finland	3.5	Lager
413	BS3	Koff III	Finland	4.5	Lager
414	BS4	Budweiser III	USA	5.0	Lager
415	BS5	Warsteiner P.	Germany	0.0	Lager
416	BS6	Pilsner Urquell	Czech Republic	4.4	Lager
417	BS7	Chiswick	Britain	3.5	English Bitter
418	BS8	Rainbow al.	Belgium	0.0	Lager
419	BS9	Heineken	The Netherlands	4.6	Lager
420	BS10	5.A.M. Saint	Scotland	5.0	Ale
421	BS11	SOL lager	Mexico	4.5	Lager
422	BS12	Stortebeker	Germany	4.7	Pils
423	BS13	Guinness-d	Ireland	4.2	Stout
424	BS14	Prykmestar	Finland	5.2	Pils
425	BS15	Koff porter	Finland	7.2	Porter
426	BS16	Carlsberg E	Denmark	7.2	Strong Lager
427	BS17	Houblon Chouffe	Belgium	9.0	Ale
428	B18	St. Eriks	Sweden	5.3	Lager
429	BS19	Beer Hunter's	Finland	7.0	Ale
430	BS20	Hr Fredriksen	Denmark	10.5	Stout
431	BS21	West coast	USA	7.3	Ale

406	Supplementary Table 1:	Summary of the characteristic	s of the beer brands analyzed
-----	------------------------	-------------------------------	-------------------------------