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3 **Sex-specific strategies of phosphorus acquisition in *Populus cathayana* as affected**
4 **by soil P availability and distribution**

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6 Zhichao Xia ¹, Yue He ¹, Lei Yu ¹, Rubing Lv ¹,

7 Helena Korpelainen ² and Chunyang Li ^{1,*}

8

9 ¹ College of Life and Environmental Sciences, Hangzhou Normal University, Hangzhou
10 310036, Zhejiang, China

11 ² Department of Agricultural Sciences, Viikki Plant Science Centre, University of
12 Helsinki, P.O. Box 27, FI-00014, Finland

13 * Corresponding author: Chunyang Li, E-mail address: lily@hznu.edu.cn

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15 **Highlights**

- 16 • Sexual differences in P acquisition are affected by soil P level and distribution.
17 • *P. cathayana* females benefit more from high P level and heterogeneous distribution.

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23 **Abstract** Soil phosphorus (P) availability and its distribution influence plant growth
24 and productivity, but how they affect the growth dynamics and sex-specific P
25 acquisition strategies of dioecious plant species is poorly understood. In this study, the
26 effects of soil P availability and its distribution on dioecious *Populus cathayana* Rehd.
27 were characterized. *P. cathayana* males and females were grown under three levels of
28 phosphorus (P) supply, and with homogeneous or heterogeneous P distribution.
29 Females had a greater total root length, specific root length (SPL), biomass and foliar
30 P concentration. Under P deficiency, males had a smaller root system than females but
31 a greater exudation of soil acid phosphatase, and a higher colonization rate and
32 arbuscular mycorrhizal hyphal biomass, suggesting a better capacity to mine P and a
33 stronger association with arbuscular mycorrhizal fungi to forage P. Heterogeneous P
34 availability enhanced growth and root length density in females. Female root
35 proliferation in P-rich patches was associated with increased foliar P assimilation.
36 Increased P availability by localized P application did not enhance the biomass
37 accumulation and the morphological plasticity of roots in males, but increased hyphal
38 biomass. Our results suggest that *P. cathayana* displays sexually different P acquisition
39 strategies, which explain the performance of females and males under variable soil P
40 environments.

41 **Keywords:** dioecy; P deficiency; heterogeneous phosphorus supply; rhizosphere
42 processes; root morphological plasticity; mycorrhizal associations.

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44 **Introduction**

45

46 Males and females of dioecious trees meet different reproductive demands and selective
47 pressures. Female plants often allocate more resources to reproductive effort than males,
48 which results in different resource-related trade-offs between males and females
49 (Tognetti 2012; Juvany and Munne-Bosch 2015; Hultine et al. 2016). For instance,
50 *Populus purdomii* females have showed higher reproductive investment, construction
51 cost and payback time when compared to males (Lei et al. 2017). However, sex related
52 differences in physiological and life-history traits can be observed already before
53 reproductive maturity, indicating that many sex-specific differences may be inherently
54 present and not only a consequence of unequal reproductive costs between genders
55 (Montesinos et al. 2012; Randriamanana et al. 2014).

56

57 Functional trait differences of dioecious plant species become greater in responses to
58 stressful conditions (Xu et al. 2008; Graff et al. 2013; Li et al. 2016), when males tend
59 to be more tolerant to both abiotic and biotic stresses (Hultine et al. 2016; Melnikova
60 et al. 2017; Retuerto et al. 2018). Most previous studies have focused on gender
61 responses to nutrient availability (Robakowski et al. 2018; Song et al. 2018). For
62 example, Zhang et al. (2014) found that when *Populus* confronted nutrient shortage,
63 the photosynthetic rate of males was higher than that of females. Males showed a
64 smaller accumulation of starch grains in the chloroplast, indicating that males could
65 better adjust their physiological leaf-level processes to reduce growth damage.
66 However, the responses of different genders to nutrients stress on the root level and

67 related adaptation mechanisms are largely unknown.

68

69 Plants have evolved a suite of adaptive strategies to acquire resources in severely
70 impoverished soils (Lambers et al. 2008; Shen et al. 2011; Zemunik et al. 2015). For
71 instance, plants can develop highly specialized morphological root traits to enhance soil
72 exploration efficiency for phosphorus (P) assimilation, such as increasing the specific
73 root length (SRL), and density and length of lateral roots. (Zhu and Lynch 2004). Also,
74 plants can enhance P availability by mining P in the rhizosphere via the release of Pi-
75 solubilizing root exudates, such as organic acids, enzymes and hydrogen protons, which
76 can liberate Pi from inaccessible soil complexes and phosphorylated compounds
77 (Lambers et al. 2009; Neumann and Martinoia 2002; Richardson et al. 2011). Moreover,
78 a symbiotic relationship between roots and mycorrhizal fungi is the most common
79 strategy (Smith and Read 2008), which increases a plant's acquisition of soil P through
80 the increased soil volume the plant can explore via a hyphal network. Because all
81 strategies require significant amounts of carbon, plants may increase the expression of
82 one mechanism at the expense of the other one (Barber 1995).

83

84 There is a balance or complementarity between the morphological and physiological
85 responses of roots to variable P-limited environments across plant species (Fort et al.
86 2015). For instance, Lyu et al. (2016) found that species with fibrous roots have higher
87 root/shoot biomass ratios than legumes, whereas legumes have a higher degree of
88 carboxylate exudation than species with fibrous roots under decreasing soil P supply.

89 Soil nutrient heterogeneity can greatly affect root plasticity. Plants should exploit and
90 utilize localized soil nutrient patches through root proliferation or mycorrhizal hyphal
91 foraging (Hodge 2004; Wang et al. 2006). In addition, some plant species show
92 physiological responses of roots to localized nutrient enrichment (e.g. an increase in the
93 nutrient uptake capacity per root length unit; Jackson et al., 1990). This is an effective
94 strategy to adapt spatiotemporal variability in P availability, often considered to be a
95 compensatory response (Richardson et al. 2009; Peret et al. 2011). Thus, foraging for
96 nutrients appears determined by root plasticity dynamics in response to soil P variability.
97 Plants' responses and adaptation strategies to the changing availability and distribution
98 of soil P can vary widely across plant species (Zhang et al. 2012; Adams et al. 2013;
99 Hou et al. 2016; McNickle et al. 2016). However, it is unclear, how dioecious plants
100 respond to a P deficiency and varying local distribution of soil P.

101

102 *Populus cathayana* is a typically rapidly growing dioecious species, which plays a very
103 important role in the forestation and maintenance of ecological stability in high altitude
104 regions of China. It also exhibits a strong spatial habitat segregation of the sexes under
105 natural conditions, with males usually being prevalent in nutrient-poor environments
106 while females favor nutrient-rich conditions (Meng et al. 2019). Some studies have
107 indicated that contrasting genotypes dominating in different habitats may have
108 divergent P-capture strategies (Barot et al. 2016): species from nutrient-rich habitats
109 have a strong root proliferation capacity, especially in P-rich zones (Grime 1994),
110 whereas species from nutrient-poor habitats may have the capacity to adapt to P

111 shortage conditions. In the present study, in order to investigate sex-specific adaptive
112 strategies, we conducted experiments with varying P supply levels and with both
113 homogeneous or heterogeneous P distribution to test the hypothesis that females show
114 a more positive response to rich and heterogeneous P environments compared to males,
115 because females have a high capacity for the morphological proliferation of roots,
116 resulting in improved biomass growth and nutrient uptake. In contrast, males would be
117 favored by a low P availability due to their compensatory strategies associated with
118 significant physiological and mycorrhizal plasticity of roots.

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133 **Methods and materials**

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135 *Plant materials and soils*

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137 *Populus cathayana* cuttings were collected from 20 different trees, including 10
138 females and 10 males, sampled from 5 populations (four adult trees per population) in
139 riparian and valley flat habitats (Datong, 35°56'N, 101°35'E) located in the Qinghai
140 Province, China. The cuttings were then grown in a naturally lit greenhouse. After
141 sprouting and growing for 4 weeks, 60 healthy seedlings (30 females and 30 males) of
142 approximately identical crown sizes and equal heights (~20 cm) were selected for the
143 experiments. The used sandy soil was collected from the experimental site at the
144 Hangzhou Normal University, Zhejiang Province, China, air-dried and passed through
145 a 2-mm sieve. Soil properties were as follows: soil organic matter content of 2.82 g kg⁻¹,
146 total N of 0.28 g kg⁻¹, available P of 2.62 mg kg⁻¹ and available K of 90.65 mg kg⁻¹,
147 pH 8.64 (the ratio of soil to CaCl₂ solution was 1:2.5).

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149 *Experimental set-up*

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151 *Experiment 1* In order to investigate how *P. cathayana* males and females respond to
152 changing P availability, a pot experiment was conducted in a glasshouse with two
153 genders and three soil P supply levels: 0 mg kg⁻¹, 50 mg kg⁻¹ and 200 mg kg⁻¹. P was
154 applied as Ca (H₂PO₄)₂ · H₂O. There were 6 treatment combinations, with four replicates

155 per treatment. The pots were filled with 10 kg of air-dried sandy soil. To ensure that the
156 nutrient supply was adequate for plant growth, soil was also fertilized with nutrients as
157 follows (mg per pot): $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ 8040; K_2SO_4 100; $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 130;
158 $\text{MnSO}_4 \cdot \text{H}_2\text{O}$ 50; $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ 75; $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ 15. All pots were arranged in a
159 completely randomized design and additionally randomized weekly. The plants were
160 watered every day to maintain field capacity (18%, w/w). The plants were harvested 6
161 months after transplanting (from March to August 2018) and separated into leaves,
162 stems and roots. Following root excavation, the soil adhering to roots was defined as
163 rhizosphere soil and sub-sampled for pH, soil acid phosphatase and phospholipid fatty
164 acid (PLFA) measurements. Extramatrical arbuscular mycorrhizal (AM) hyphal
165 biomass was estimated using the PLFA biomarker 16:1 ω 5c

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167 *Experiment 2* In order to assess sex-specific variation between *P. cathayana* males and
168 females in response to the spatial distribution of P, we set up a factorial experiment
169 comprising two genders and two P supply patterns, with four replications for each
170 treatment. P was supplied as $\text{Ca}(\text{H}_2\text{PO}_4)_2 \cdot \text{H}_2\text{O}$ in all treatments with the total amount
171 of P being equal. The experiment was conducted using a specific root segregation
172 device made of PVC material (length 20 cm \times width 20 cm \times height 30 cm), divided
173 into two equal compartments by rigid plastic. The partition in the middle prevented the
174 movement of nutrients between the two compartments. Each root device was filled with
175 10 kg soil. A clean 3-cm deep sand layer was placed on the top of the soil as a buffer
176 zone to enable natural root growth in the compartments. For the heterogeneous P

177 treatment, 2 g of P was manually mixed into one side of the root box as a P-rich patch,
178 and the other side was left without added P. For the homogeneous P treatment, 2 g of P
179 was spread evenly throughout the soil of both compartments. To ensure nutrient supply
180 for plant growth, the same amounts of other nutrients as in Experiment 1 were added to
181 the soil, followed by thorough mixing. One cutting of each sex was planted at the center
182 of each root segregation device. All seedlings were harvested at the end of the
183 experiment and divided into leaves, stems and roots (from March to August 2018). In
184 the heterogeneous P treatments, roots grown inside vs outside the P-rich patch were
185 sampled, and the corresponding roots at the same location were sampled in the
186 homogeneous treatments. Following root excavation, the soil was sub-sampled for an
187 extramatrical AM hyphal biomass analysis.

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189 *Root trait measurements*

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191 The root system of each plant was first washed carefully. A part of the roots (15-30
192 pieces of 15-mm long fine roots segments per plant) from each plant was excised from
193 the root system, then analyzed for the AMF colonization rate, as described by Vierheilig
194 et al. (1998). Roots segments (15 mm long) were randomly sampled from each
195 treatment, washed in distilled water, and then immersed in a FAA fixative for 4 h. Root
196 segments were bleached in 10% KOH for 1 h and stained in ink and vinegar (95%
197 vinegar and 5% ink) for 3 min at 90 C. All stained root segments were randomly
198 selected for microscopic observations to calculate the colonization rate (Col).

199 Subsequently, other samples were scanned and analyzed for morphological root
200 parameters. Roots were scanned with an EPSON root scanner at 400 dots per inch
201 (Epson Expression1600 pro, Model EU-35, Tokyo, Japan). The total root length was
202 analyzed using software Win-RHIZO (WinRhizo Pro2004b, version 5.0, Regent
203 Instruments Inc., Quebec, QC, Canada). Afterwards, all roots were collected and dried
204 at 75 °C for 72 h and weighed to calculate the specific root length.

205

206 *Determination of leaf and root P contents*

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208 The phosphorus concentration of shoots and roots was determined after digestion with
209 a mixture of 5 ml of concentrated sulphuric acid and 8 ml of 30% v/v H₂O₂. P was
210 analysed by the molybdovanadophosphate method by spectrophotometry at 440 nm
211 (Varian Vista-Pro CCD; Johnson & Ulrich, 1959).

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213 *Determination of soil pH, acid phosphatase, and extramatrical AM hyphal biomass*

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215 The pH of the rhizosphere solutions was measured using a pH meter. Soil acid
216 phosphatase activity in the rhizosphere soil was analyzed according to Neumann (2006).
217 Briefly, 0.5 mL soil suspension (0.5 g rhizosphere soil mixed with 2 mL deionized
218 water) was placed into 2-mL Eppendorf vials, followed by the addition of 0.4 mL
219 acetate buffer (pH 5.2) and 0.1 mL substrate [pNPP (p-nitrophenylphosphate); Sigma St.
220 Louis, MO, USA]. Vials were gently shaken and incubated for 30 min at 30 °C. The

221 reaction was terminated with 0.5 mL 0.5 M NaOH, and the mixture was centrifuged for
222 10 min at 12,000×g. In the case of controls, NaOH was added before incubation. The
223 absorbance of this solution was measured with a spectrophotometer at 405 nm. For the
224 PLFA analysis, about 5 g of freeze-dried soil was used for fatty acid extraction, and
225 PLFAs of different biomarkers were quantified by gas chromatography (Frostegård and
226 Bååth 1996). Fatty acid 16:1 ω 5c, identified as an AM fungal biomarker in a previous
227 study, was used to measure AM hyphal biomass (Chen et al. 2016).

228

229 *Data analysis*

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231 Biomass, total root length, SPL, root length density (RLD), foliar P concentration, AMF
232 colonization rate, extramatrical AM hyphal biomass, soil pH and acid phosphatase were
233 analyzed with one- or two-way analysis of variance (ANOVA) according to each
234 experiment. All treatments were regarded as fixed factors (i.e. gender, P level, P
235 distribution, as well as the interactions between gender and P level or distribution).
236 Tukey's honestly significant difference test was used for multiple comparisons when
237 ANOVA terms were significant using SPSS 16.0 for Windows (SPSS Inc. Chicago,
238 Illinois, USA).

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242 **Results**

243

244 *Sexual differences in P. cathayana under variable soil P supply (Experiment 1)*

245

246 The shoot and root biomass of *P. cathayana* increased with soil P supply (Fig. 1). The
247 biomass of *P. cathayana* females increased more and was greater than that of males,
248 the largest growth difference being found under 200 mg kg⁻¹ P supply conditions.
249 Similarly, P application significantly increased the leaf P concentration of females (Fig.
250 2). However, the leaf P concentration of males was unaffected by the P supply level.

251

252 P application significantly enhanced root growth in both sexes (Fig 3a). The females
253 exhibited greater responses in total root length compared with males; the total root
254 length was maximized at 200 mg kg⁻¹ P supply level. In contrast, males exhibited less
255 definite P application effects on the total root length. SPL showed significant
256 differences between the sexes, but not among P supply levels (Fig 3b). Moreover,
257 females possessed higher SRL than did males at all P application levels, the value
258 decreasing with an increasing soil P supply. In contrast, SRL of males remained
259 unchanged.

260

261 Rhizosphere soil pH was significantly lower under P deficiency (Fig. 4a). However,
262 there were no significant differences between the sexes at any P level. By contrast, the
263 acid phosphatase activity in the rhizosphere decreased with an increasing P supply (Fig.
264 4b). Males exhibited significantly higher acid phosphatase activities under P deficiency

265 compared to females.

266

267 P application significantly reduced the colonization rate and AM hyphal biomass of *P.*
268 *cathayana* (Fig. 5a, b). Males had higher colonization rates and AM hyphal biomasses
269 than did females under P shortage, while there were no significant differences in these
270 parameters between the sexes under high P supply (200 mg kg⁻¹ P).

271

272 *The effect of spatial variability in P supply on dioecious P. cathayana (Experiment 2)*

273

274 The shoot biomass of *P. cathayana* was significantly affected by the spatial distribution
275 of P (Fig. 6a). Also sex-related differences in biomass were found. When soil P was
276 supplied heterogeneously, the female biomass was 37% higher than under
277 homogeneous application, whereas males showed a slightly lower biomass in the
278 heterogeneous treatment than in the homogeneous treatment. On the other hand, sex
279 influenced the foliar P concentration of *P. cathayana*, but the spatial distribution of P
280 did not (Fig. 6b).

281

282 RLD of *P. cathayana* females was greater in the P-rich soil in the heterogeneous
283 treatment compared with the homogeneous treatment (Fig.7a). However, RLD of males
284 did not differ between the heterogeneous and homogeneous environment. We used the
285 PLFA biomarker 16:1 ω 5c to represent the external fungal proliferation of AMF. The
286 external biomass of fungi in the two sexes showed no significant differences between

287 the homogeneous and the heterogeneous environment in the P-rich patch (Fig.7b).
288 Overall, RLD of *P. cathayana* in the P-rich patch was significantly influenced by sex
289 and spatial distribution of P, but the external fungal biomass was not affected (Table 1).
290
291 Outside the P-rich soil, RLD of *P. cathayana* was not significantly affected by the
292 spatial distribution of P, but the effect was significant on the external fungal biomass
293 (Fig. 7c-d; Table 1). Unlike females, males showed a higher mycorrhizal hyphal
294 proliferation in the heterogeneous environment outside the P-rich patch when compared
295 to the same position in the homogeneous environment (Fig. 7d).

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303 **Discussion**

304

305 *Populus* trees are fast-growing dioecious species, although males and females display
306 different growth rates under different environmental conditions (Randriamanana et al.
307 2014; Zhang et al. 2014; Chen et al. 2015). In this study, we found that there is sex-
308 specific variation in the growth rates of *P. cathayana* under a different availability and

309 distribution of soil P. Furthermore, under added P conditions, the biomass accumulation
310 of females is greater than that of males, which indicates that females grow more rapidly
311 than males. The growth speed of plants usually depends on the availability of a
312 sufficient supply of nutrients in soil (Chen et al. 2018). Therefore, females are more
313 sensitive to P application, while relatively slowly growing males are more tolerant to
314 low P conditions. Considering the distribution of P in soil, there was no significant
315 difference in male biomass between homogenous and heterogeneous P supply patterns.
316 However, the biomass accumulation of females was much higher under heterogeneous
317 P supply than that in a homogenous P condition, which showed that the heterogeneous
318 P distribution was more conducive to the growth of females.

319

320 Plants can adjust the morphological plasticity of their root systems to adapt to variable
321 soil P environments. In response to P shortage, plants can inhibit the growth of their
322 primary roots and promote the development of lateral roots (Williamson et al. 2001;
323 López-Bucio et al. 2002). We discovered that *P. cathayana* females increase SPL more
324 under P shortage conditions. Having a large contact area between roots and soil is
325 expected to enhance P acquisition, whereas added P allows females to increase the total
326 root length and SRL. A previous study found that plants with a greater root length may
327 possess more lateral roots, which can facilitate nutritional foraging under high nutrient
328 supply conditions (Kavka and Polle 2016; He et al. 2017; Chen et al. 2018). Thus, the
329 greater root length and root surface area of females in comparison with males suggest
330 that females probably have more absorptive roots, which may facilitate P acquisition

331 under high P supply.

332

333 An increase in SRL is not a universal response to a low P supply (Pang et al. 2010; Lyu
334 et al. 2016). Indeed, there was no effect of P deficiency on SRL of males, but males
335 still showed a stronger ability to tolerate P shortage. Strengthening the rhizosphere
336 processes is an important mechanism through which males adapt to soil P deficiency.
337 It has been reported previously that the release of acid phosphatase increases in woody
338 Chinese fir under a low P environment (Chen et al. 2018). Additionally, plants can burst
339 out protons to induce root acidification, which would enhance the hydrolyzation of
340 organic P by increasing phosphatase activity (Yan et al. 2002; Valentinuzzi et al. 2015).
341 In this study, protons and soil acid phosphatase levels increased in the low P
342 environment, but also significant sex-specific differences were found. Males have
343 stronger rhizosphere processes compared to females, thereby having a better capacity
344 to increase the release of soil acid phosphatase to mine organic P in order to compensate
345 the negative impact of P limitation. Moreover, we found that under a low P condition,
346 the AM hyphal biomass of males is higher, which potentially indicates that males may
347 acquire more P through mycorrhizal hyphae proliferation in a complementary fashion.

348

349 A recent study by Wu et al. (2018) found sex-specific responses to mycorrhiza in
350 dioecious *P. cathayana*. The results showed that males have a higher colonization level
351 compared to females. Although studies on several species have found that females show
352 a higher dependence on mycorrhizal fungi in high-resource habitats (Verdú and García-

353 Fayos 2003; Varga and Kytovlita 2008; Varga et al. 2017), *P. cathayana* males perform
354 better than females when subjected to stress and AMF (Li et al. 2015; Wu et al. 2015).
355 Thus, we conclude that sex-specific associations between roots and mycorrhizal fungi
356 depend on the identity of the plant and AM fungal species, as well as on environmental
357 conditions. Taken together, males can adapt to P-impooverished soils through a possible
358 compensatory strategy by changing physiological root features and the function of
359 associated mycorrhizal fungi.

360

361 The growth responses of plants to P distribution often show species-specific effects
362 (Hodge 2004, 2006). However, our study is the first one to report in a dioecious plant
363 that there are gender-related differences in responses to P distribution within the same
364 population. Previously, it has been shown that plants with a larger root biomass often
365 benefit more from a heterogeneous nutrient distribution (Tamme et al. 2016). We
366 discovered that at all P supply levels, females have more root biomass than males and
367 they can acquire more P in nutrient-rich patches, yet depending on their foraging
368 sensitivity or precision. On the other hand, we found a higher biomass of AM hyphae
369 outside P-rich patches in males, which indicates that AM hyphal proliferation
370 compensates for the relative inefficiency to proliferate roots.

371

372 Dioecy effectively increases opportunities of survival and growth in a population.
373 Females have higher reproductive costs than males and need more P to support them
374 (Zhang et al. 2014; Hultine et al. 2016; Lei et al. 2017). Therefore, females produce

375 thin roots (high SRL) with relatively low carbon costs. Consequently, the soil around
376 their roots is not intensively explored, but they build longer roots with a similar C
377 budget compared to males. Plants with high SRL can absorb more P and are better able
378 to adapt to high or heterogeneous P environments, resulting in greater growth benefits
379 (Hodge 2004, 2006). However, this acquisitive strategy may result in a long-term cost,
380 since roots with such characteristics have short lifespans, which is not a good adaptation
381 strategy in infertile soils in a long term (Freschet et al. 2018). In contrast, males with
382 low SPL but a high root tissue density possess high root construction costs, especially
383 under low P conditions.

384

385 In the present study, males showed stronger physiological or mycorrhizal but weaker
386 morphological responses of roots compared to females when adapting to low P
387 environments. This represents an economic strategy, because it is generally believed
388 that the cost of spending C on higher metabolic rates is less than the construction of
389 longer roots (Le Roux et al. 2009; Funayama-Noguchi et al. 2015). On the other hand,
390 species with low SPL but a high degree of root colonisation by mycorrhizal fungi can
391 better resist herbivore and pathogen attacks, which may be beneficial in unfertile soils
392 (Laliberte et al. 2015). Actually, this may be another reason for the relatively good male
393 performance.

394

395 Sexual dimorphism results in niche divergence, variation in sex ratios, and spatial
396 segregation of the sexes (SSS) across environmental gradients (Eppley 2006; Li et al.

397 2007). Previously, it has been showed that habitats with female-biased sex ratios are
398 less stressful than those with male-biased sex ratios, while males are more abundant
399 than females under adverse environmental conditions (Juvany and Munne-Bosch 2015;
400 Hultine et al. 2016; Melnikova et al. 2017). Sex ratio biases across resource gradients
401 may reflect trade-offs between resource exploration strategies (Lei et al. 2017),
402 whereby females are selected for a higher resource uptake to meet the higher cost of
403 reproduction through root morphology (Eppley et al. 2009), and males are selected for
404 an effective storage of nutrients to maximize stress tolerance and mating opportunities
405 in low-resource habitats through root physiology to enhance the acquisition of soil
406 organic P or AM hyphal proliferation (Hultine et al. 2016; Graff et al. 2018). However,
407 our studies were conducted in greenhouse conditions with inevitable limitations, and
408 field work is being carried out to further verify our conclusions.

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415 **Conclusions**

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417 This study provided new insights into the growth consequences of sexual dimorphism
418 in dioecious *P. cathayana* to cope with a variable soil P level and distribution. Males
419 are not sensitive to the P availability and distribution, and they have a good ability to

420 regulate physiological and mycorrhizal processes of roots, thereby increasing the
421 acquisition of P in low P conditions. On the other hand, females have larger root
422 systems, which can increase the foraging region of roots and P acquisition. Therefore,
423 when the amount of soil P is high and heterogeneously distributed, females grow better
424 when compared to males.

425

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439 **References**

440

441 Adams TS, McCormack ML, Eissenstat DM (2013) Foraging strategies in trees of
442 different root morphology: the role of root lifespan. *Tree Physiol* 33: 940-948

443

444 Barber S (1995) Soil nutrient bioavailability: a mechanistic approach, 2nd edn. Wiley,
445 New York

446

447 Barot S, Bornhofen S, Boudsocq S, Raynaud X, Loeuille N, Schweitzer J (2016)
448 Evolution of nutrient acquisition: when space matters. *Funct Ecol* 30: 283-294

449

450 Chen J, Dong T, Duan B, Korpelainen H, Niinemets U, Li C (2015) Sexual competition
451 and N supply interactively affect the dimorphism and competitiveness of opposite sexes
452 in *Populus cathayana*. *Plant Cell Environ* 38: 1285-1298

453

454 Chen W, Koide RT, Adams TS, DeForest JL, Cheng L, Eissenstat DM (2016) Root
455 morphology and mycorrhizal symbioses together shape nutrient foraging strategies of
456 temperate trees. *Proc Natl Acad Sci* 113: 8741-8746

457

458 Chen Y, Nguyen THN, Qin J, Jiao Y, Li Z, Ding S, Lu Y, Liu Q, Luo Z-B (2018)
459 Phosphorus assimilation of Chinese fir from two provenances during acclimation to
460 changing phosphorus availability. *Environ Exp Bot* 153: 21-34

461

462 Eppley SM (2006) Females make tough neighbors: sex-specific competitive effects in
463 seedlings of a dioecious grass. *Oecologia* 146: 549-554

464

465 Eppley SM, Mercer CA, Haaning C, Graves CB (2009) Sex-specific variation in the
466 interaction between *Distichlis spicata* (Poaceae) and mycorrhizal fungi. *Am J Bot* 96:
467 1967-1973
468

469 Freschet GT, Violle C, Bourget MY, Scherer-Lorenzen M, Fort F (2018) Allocation,
470 morphology, physiology, architecture: the multiple facets of plant above- and below-
471 ground responses to resource stress. *New Phytol* 219: 1338-1352
472

473 Frostegård Å, Bååth E (1996) The use of phospholipid fatty acid analysis to estimate
474 bacterial and fungal biomass in soil. *Biol Fertil Soils* 22:59-65
475

476 Funayama-Noguchi S, Noguchi K, Terashima I (2015) Comparison of the response to
477 phosphorus deficiency in two lupin species, *Lupinus albus* and *L. angustifolius*, with
478 contrasting root morphology. *Plant Cell Environ* 38: 399-410
479

480 Graff P, Aguiar MR, Almeida RJ (2018) Females engage in stronger relationships:
481 positive and negative effects of shrubs are more intense for *Poa ligularis* females than
482 for males. *Ann Bot-London* 122: 435-443
483

484 Graff P, Rositano F, Aguiar MR, Wilson S (2013) Changes in sex ratios of a dioecious
485 grass with grazing intensity: the interplay between gender traits, neighbour interactions
486 and spatial patterns. *J Ecol* 101: 1146-1157

487

488 Grime JP (1994) The role of plasticity in exploiting environmental heterogeneity. In:
489 Caldwell MM, Pearcy RW (eds) Exploitation of environmental heterogeneity by plants:
490 ecophysiological processes above- and belowground. Academic Press, San Diego, pp
491 1-19

492

493 He H, Peng Q, Wang X, Fan C, Pang J, Lambers H, Zhang X (2017) Growth,
494 morphological and physiological responses of alfalfa (*Medicago sativa*) to phosphorus
495 supply in two alkaline soils. Plant Soil 416: 565-584

496

497 Hodge A (2004) The plastic plant: root responses to heterogeneous supplies of nutrients.
498 New Phytol 162: 9-24

499

500 Hodge A (2006) Plastic plants and patchy soils. J Exp Bot 57: 401-411

501

502 Hou X, Tigabu M, Zhang Y, Ma X, Cai L, Wu P, Liu A, Wang C, Qiu H (2016) Root
503 plasticity, whole plant biomass, and nutrient accumulation of *Neyraudia reynaudiana*
504 in response to heterogeneous phosphorus supply. J Soil Sediment 17: 172-180

505

506 Hultine KR, Grady KC, Wood TE, Shuster SM, Stella JC, Whitham TG (2016) Climate
507 change perils for dioecious plant species. Nat Plants 2: 16109

508

509 Johnson CM, Ulrich A (1959) Analytical methods for use in plant analysis. Berkeley,
510 CA, USA: University of California, Agricultural Experiment Station.

511

512 Juvany M, Munne-Bosch S (2015) Sex-related differences in stress tolerance in
513 dioecious plants: a critical appraisal in a physiological context. *J Exp Bot* 66: 6083-
514 6092

515

516 Kavka M, Polle A (2016) Phosphate uptake kinetics and tissue-specific transporter
517 expression profiles in poplar (*Populus × canescens*) at different phosphorus
518 availabilities. *BMC Plant Biol* 16: 206

519

520 Laliberte E, Lambers H, Burgess TI, Wright SJ (2015) Phosphorus limitation, soil-
521 borne pathogens and the coexistence of plant species in hyperdiverse forests and
522 shrublands. *New Phytol* 206: 507-521

523

524 Lambers H, Mougél C, Jaillard B, Hinsinger P (2009) Plant-microbe-soil interactions
525 in the rhizosphere: an evolutionary perspective. *Plant Soil* 321: 83-115

526

527 Lambers H, Raven JA, Shaver GR, Smith SE (2008) Plant nutrient-acquisition
528 strategies change with soil age. *Trends Ecol Evol* 23: 95-103

529

530 Le Roux MR, Khan S, Valentine AJ (2009) Nitrogen and carbon costs of soybean and

531 lupin root systems during phosphate starvation. *Symbiosis* 48: 102-109

532

533 Lei Y, Jiang Y, Chen K, Duan B, Zhang S, Korpelainen H, Niinemets U, Li C (2017)

534 Reproductive investments driven by sex and altitude in sympatric *Populus* and *Salix*

535 trees. *Tree Physiol* 37: 1503-1514

536

537 Li C, Xu G, Zang R, Korpelainen H, Berninger F (2007) Sex-related differences in leaf

538 morphological and physiological responses in *Hippophae rhamnoides* along an

539 altitudinal gradient. *Tree Physiol* 27: 399-406

540

541 Li Y, Duan B, Chen J, Korpelainen H, Niinemets U, Li C (2016) Males exhibit

542 competitive advantages over females of *Populus deltoides* under salinity stress. *Tree*

543 *Physiol* 36: 1573-1584

544

545 Li Z, Wu N, Liu T, Chen H, Tang M (2015) Sex-related responses of *Populus cathayana*

546 shoots and roots to AM fungi and drought stress. *PloS one* 10: e0128841.

547

548 López-Bucio J, Hernández-Abreu E, Sánchez-Calderón L, Nieto-Jacobo MF, Simpson

549 J, Herrera-Estrella L (2002) Phosphate availability alters architecture and causes

550 changes in hormone sensitivity in the *Arabidopsis* root system. *Plant Physiol* 129: 244-

551 256

552

553 Lyu Y, Tang HL, Li HG, Zhang FS, Rengel Z, Whalley WR, Shen JB (2016) Major crop
554 species show differential balance between root morphological and physiological
555 responses to variable phosphorus supply. *Front Plant Sci* 7: 15
556

557 McNickle GG, Deyholos MK, Cahill JF, Schweitzer J (2016) Nutrient foraging
558 behaviour of four co-occurring perennial grassland plant species alone does not predict
559 behaviour with neighbours. *Funct Ecol* 30: 420-430
560

561 Melnikova NV, Borkhert EV, Snezhkina AV, Kudryavtseva AV, Dmitriev AA (2017)
562 Sex-specific response to stress in *Populus*. *Front Plant Sci* 8: 6
563

564 Meng Z, X W, Su G, Li D, Dong T, Peng J, Li X, Gong X, L N, X X (2019) Spatial
565 distribution of male and female *Populus cathayana* populations and its drivers in
566 Xiaowutai Mountains, Hebei, China. *Chin J Plant Ecol*, 42: 1145-1153
567

568 Montesinos D, Villar-Salvador P, Garcia-Fayos P, Verdu M (2012) Genders in *Juniperus*
569 *thurifera* have different functional responses to variations in nutrient availability. *New*
570 *Phytol* 193: 705-712
571

572 Neumann G (2006) Quantitative determination of acid phosphatase activity in the
573 rhizosphere and on the root surface. In: Jones, D.L. (Eds.), 4.2 Biochemistry. In: Luster,
574 J., Finlay, R. (Eds.), *Handbook of Methods used in Rhizosphere Research-Online*

575 Edition

576

577 Neumann G, Martinoia E (2002) Cluster roots - an underground adaptation for survival
578 in extreme environments. Trends Plant Sci 7: 162-167

579

580 Pang J, Ryan MH, Tibbett M, Cawthray GR, Siddique KH, Bolland MD, Denton MD,
581 Lambers H (2010) Variation in morphological and physiological parameters in
582 herbaceous perennial legumes in response to phosphorus supply. Plant Soil 331: 241-
583 255

584

585 Peret B, Clement M, Nussaume L, Desnos T (2011) Root developmental adaptation to
586 phosphate starvation: better safe than sorry. Trends Plant Sci 16: 442-450

587

588 Randriamanana TR, Nybakken L, Lavola A, Aphalo PJ, Nissinen K, Julkunen-Tiitto R
589 (2014) Sex-related differences in growth and carbon allocation to defence in *Populus*
590 *tremula* as explained by current plant defence theories. Tree Physiol 34: 471-487

591

592 Retuerto R, Vilas JS, Varga S (2018) Sexual dimorphism in response to stress. Environ
593 Exp Bot 146: 1-4

594

595 Richardson AE, Hocking PJ, Simpson RJ, George TS (2009) Plant mechanisms to
596 optimise access to soil phosphorus. Crop Pasture Sci 60: 124-143

597

598 Richardson AE, Lynch JP, Ryan PR, Delhaize E, Smith FA, Smith SE, Harvey PR, Ryan
599 MH, Veneklaas EJ, Lambers H, Oberson A, Culvenor RA, Simpson RJ (2011) Plant
600 and microbial strategies to improve the phosphorus efficiency of agriculture. *Plant Soil*
601 349: 121-156

602

603 Robakowski P, Pers-Kamczyc E, Ratajczak E, Thomas PA, Ye ZP, Rabska M, Iszkulo
604 G (2018) Photochemistry and antioxidative capacity of female and male *Taxus baccata*
605 L. acclimated to different nutritional environments. *Front Plant Sci* 9: 13

606

607 Shen JB, Yuan LX, Zhang JL, Li HG, Bai ZH, Chen XP, Zhang WF, Zhang FS (2011)
608 Phosphorus dynamics: from soil to plant. *Plant Physiol* 156: 997-1005

609

610 Smith SE, Read DJ (2008) *Mycorrhizal symbiosis*, 3rd edn. Academic Press, London

611

612 Song H, Lei Y, Zhang S (2018) Differences in resistance to nitrogen and phosphorus
613 deficiencies explain male-biased populations of poplar in nutrient-deficient habitats. *J*
614 *Proteomics* 178: 123-127

615

616 Tamme R, Gazol A, Price JN, Hiiesalu I, Pärtel M (2016) Co-occurring grassland
617 species vary in their responses to fine-scale soil heterogeneity. *J Veg Sci* 27: 1012-1022

618

619 Tognetti R (2012) Adaptation to climate change of dioecious plants: does gender
620 balance matter? *Tree Physiol* 32: 1321-1324
621

622 Valentinuzzi F, Pii Y, Vigani G, Lehmann M, Cesco S, Mimmo T (2015) Phosphorus
623 and iron deficiencies induce a metabolic reprogramming and affect the exudation traits
624 of the woody plant *Fragaria* × *ananassa*. *J Exp Bot* 66: 6483-6495
625

626 Varga S, Kytöviita MM (2008) Sex-specific responses to mycorrhiza in a dioecious
627 species. *Am J Bot* 95: 1225-1232
628

629 Varga S, Vega-Frutis R, Kytöviita MM, Franken P (2017) Competitive interactions are
630 mediated in a sex-specific manner by arbuscular mycorrhiza in *Antennaria dioica*. *Plant*
631 *Biology* 19: 217-226
632

633 Verdú M, García-Fayos P (2003) Frugivorous birds mediate sex-biased facilitation in a
634 dioecious nurse plant. *J Veg Sci* 14: 35-42
635

636 Vierheilig, H., Coughlan, A., Wyss, U., Piche, Y., 1998. Ink and vinegar, a simple
637 staining technique for arbuscular-mycorrhizal fungi. *Appl Environ Microbiol.* 64:
638 5004-5007
639

640 Wang LX, Mou PP, Jones RH (2006) Nutrient foraging via physiological and

641 morphological plasticity in three plant species. *Can J Forest Res* 36: 164-173

642

643 Williamson LC, Ribrioux SP, Fitter AH, Leyser HO (2001) Phosphate availability

644 regulates root system architecture in *Arabidopsis*. *Plant Physiol* 126: 875-882

645

646 Wu N, Li Z, Liu H, Tang M (2015) Influence of arbuscular mycorrhiza on

647 photosynthesis and water status of *Populus cathayana* Rehder males and females under

648 salt stress. *Acta Physiol Plant* 37: 183

649

650 Wu Q, Tang Y, Dong T, Liao Y, Li D, He X, Xu X (2018) Additional AM fungi

651 inoculation increase *Populus cathayana* intersexual competition. *Front Plant Sci* 9: 607

652

653 Xu X, Yang F, Xiao X, Zhang S, Korpelainen H, Li C (2008) Sex-specific responses of

654 *Populus cathayana* to drought and elevated temperatures. *Plant Cell Environ* 31: 850-

655 860

656

657 Yan F, Zhu Y, Müller C, Zörb C, Schubert S (2002) Adaptation of H⁺-pumping and

658 plasma membrane H⁺ ATPase activity in proteoid roots of white lupin under phosphate

659 deficiency. *Plant Physiol* 129: 50-63

660

661 Zemunik G, Turner BL, Lambers H, Laliberté E (2015) Diversity of plant nutrient-

662 acquisition strategies increases during long-term ecosystem development. *Nat Plants* 1:

663 15050

664

665 Zhang S, Jiang H, Zhao H, Korpelainen H, Li C (2014) Sexually different physiological
666 responses of *Populus cathayana* to nitrogen and phosphorus deficiencies. *Tree Physiol*
667 34: 343-354

668

669 Zhang Y, Zhou Z, Yang Q (2012) Genetic variations in root morphology and phosphorus
670 efficiency of *Pinus massoniana* under heterogeneous and homogeneous low
671 phosphorus conditions. *Plant Soil* 364: 93-104

672

673 Zhu J, Lynch JP (2004) The contribution of lateral rooting to phosphorus acquisition
674 efficiency in maize (*Zea mays*) seedlings. *Funct Plant Biol* 31: 949

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681 **Figure legends**

682

683 **Figure 1** Effects of P application on the performance of *P. cathayana* females and males.

684 Bars with the same letter are not significantly different at $P < 0.05$, according to

685 ANOVA, followed by Tukey HSD tests.

686

687 **Figure 2** Effects of P application on the foliar P concentration of *P. cathayana* females
688 and male. Columns with the same letter are not significantly different at $P < 0.05$
689 according to ANOVA, followed by Tukey HSD tests.

690

691 **Figure 3** The total root length (a) and specific root length (b) of *P. cathayana* females
692 and males supplied with three levels of P. Columns with the same letter are not
693 significantly different at $P < 0.05$ according to ANOVA, followed by Tukey HSD tests.

694

695 **Figure 4** Soil pH (a) and acid phosphatase activity (b) in the rhizosphere of *P.*
696 *cathayana* females and males supplied with three levels of P. Columns with the same
697 letter are not significantly different at $P < 0.05$ according to ANOVA, followed by
698 Tukey HSD tests.

699

700 **Figure 5** Colonization rates (a) and PLFA biomarkers 16:1 ω 5c representing
701 extramatrical mycorrhizal hyphal biomass (b) of *P. cathayana* females and males
702 supplied with three levels of P. Columns with the same letter are not significantly
703 different at $P < 0.05$ according to ANOVA, followed by Tukey HSD tests.

704

705 **Figure 6** Effects of homogeneous vs heterogeneous phosphorus (P) supply on the shoot
706 biomass (a) and foliar P concentration (b) of *P. cathayana* females and males. Columns

707 with the same letter are not significantly different at $P < 0.05$ according to ANOVA,
708 followed by Tukey HSD tests.

709

710 **Figure 7** Effects of homogeneous/heterogeneous phosphorus (P) supply on the root
711 length density and PLFA biomarkers 16:1 ω 5c representing extramatrical mycorrhizal
712 hyphal biomass of *P. cathayana* females and males within (a-b) and outside P enriched
713 soil (c-d). Columns with the same letter are not significantly different at $P < 0.05$
714 according to ANOVA, followed by Tukey HSD tests.

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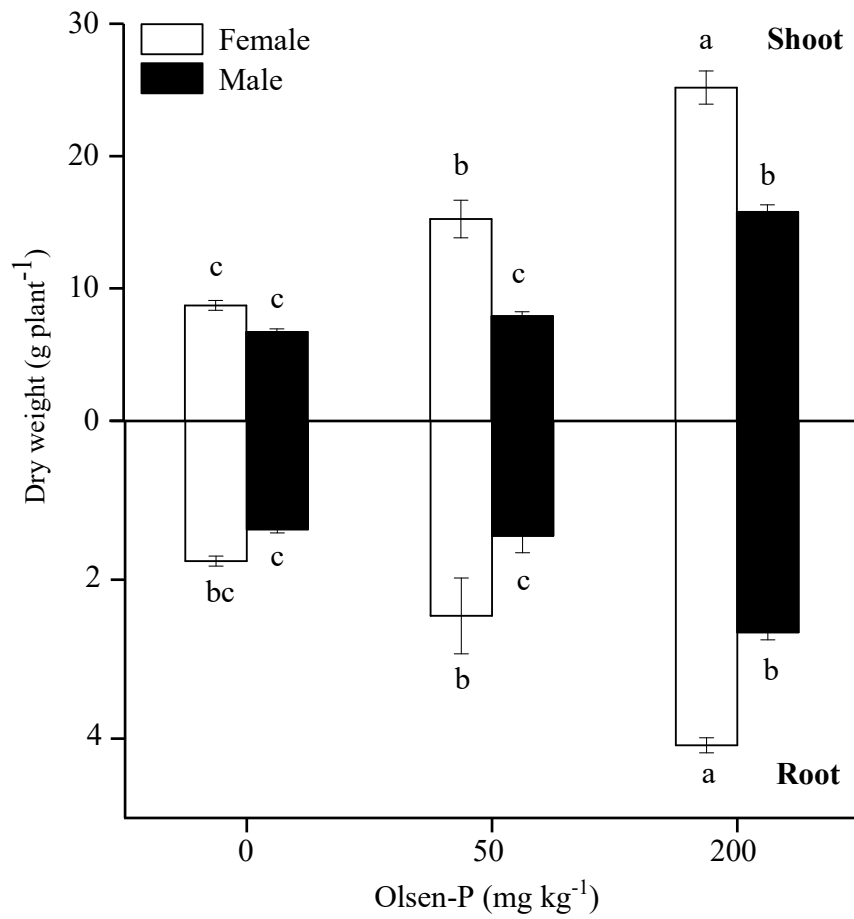
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726 **Figure 1**



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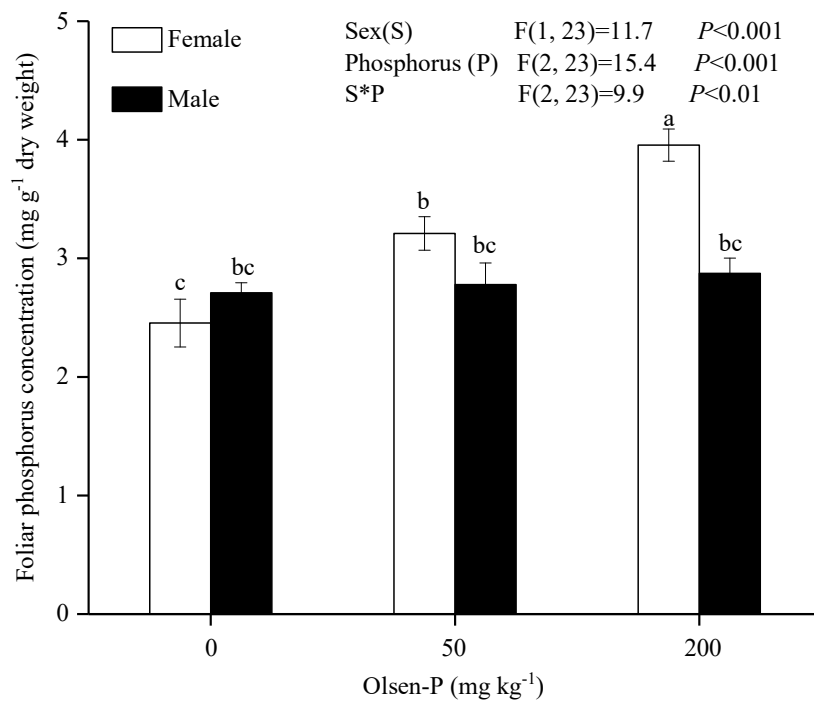
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738 **Figure 2**



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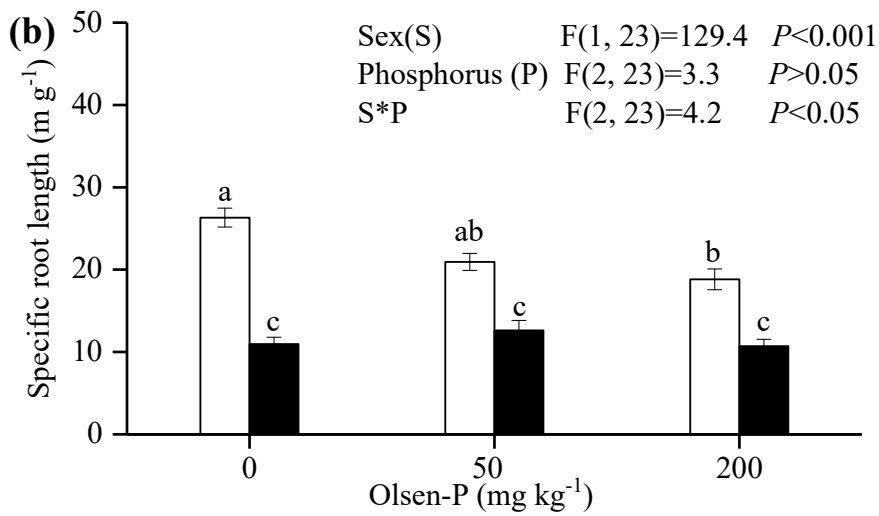
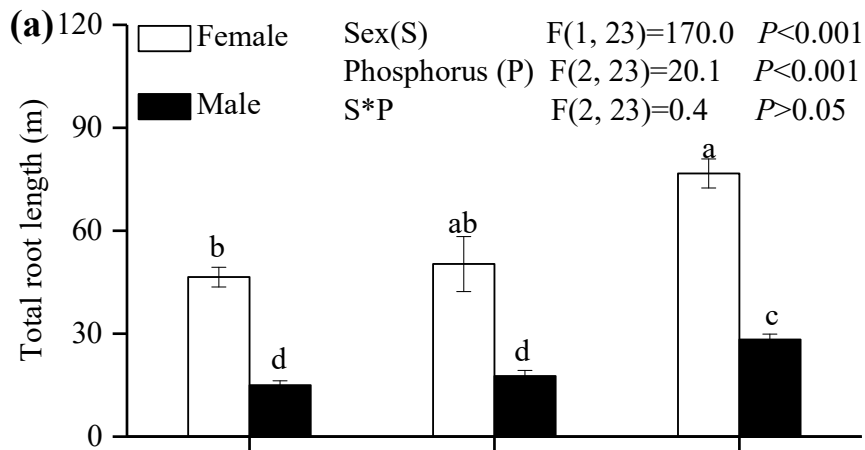
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753 **Figure 3**



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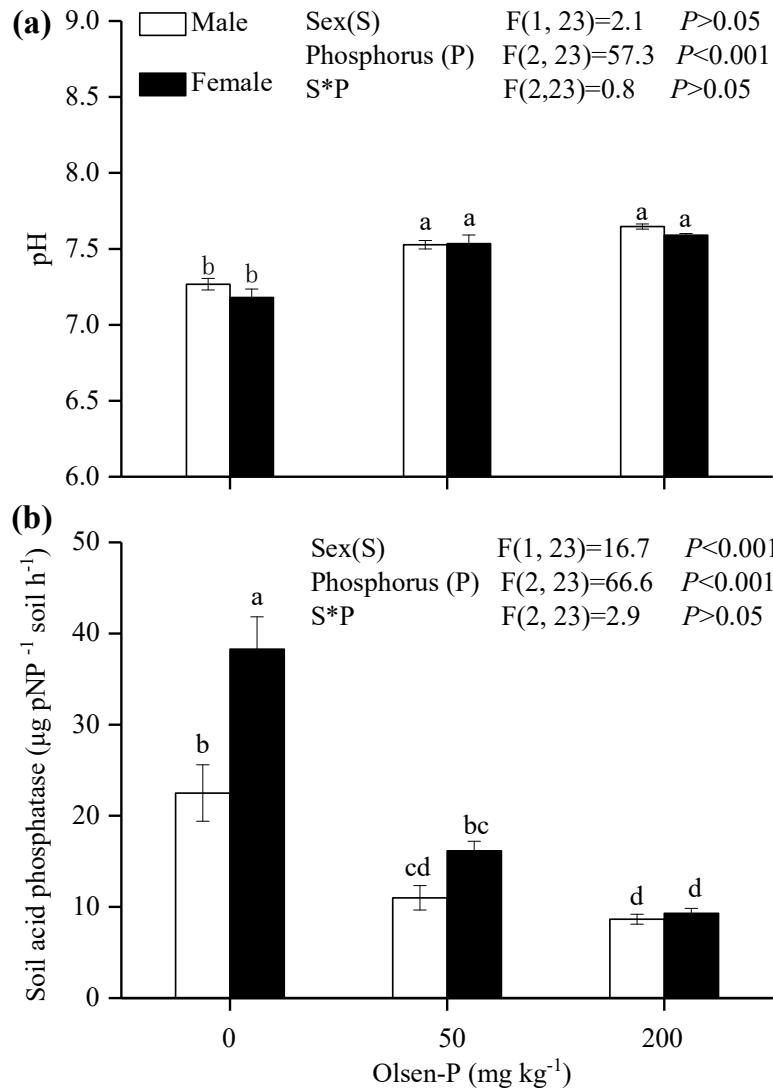
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764 **Figure 4**



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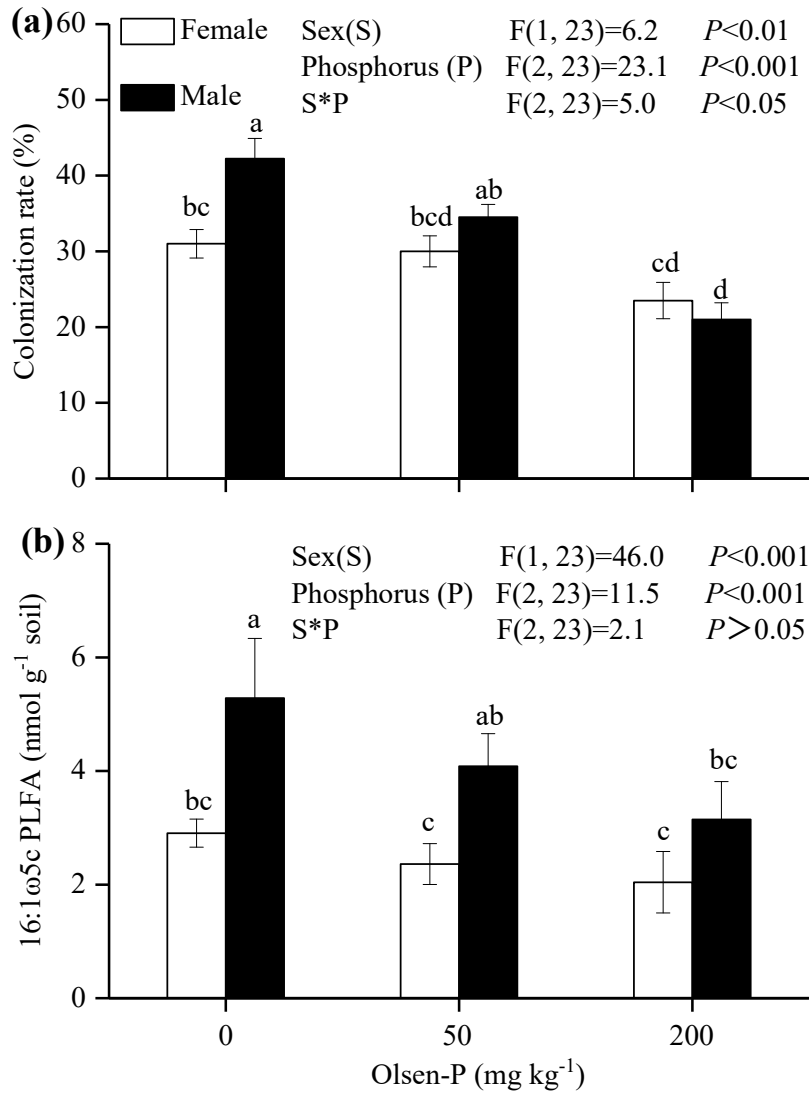
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774 **Figure 5**



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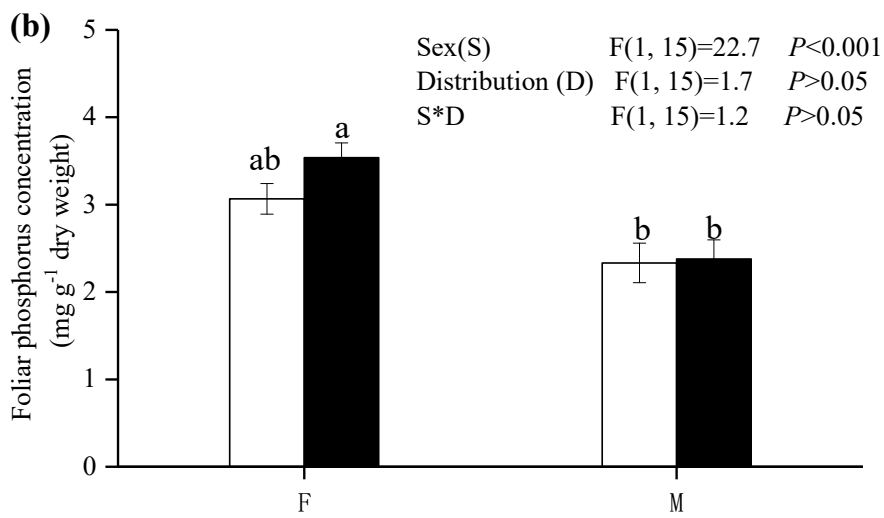
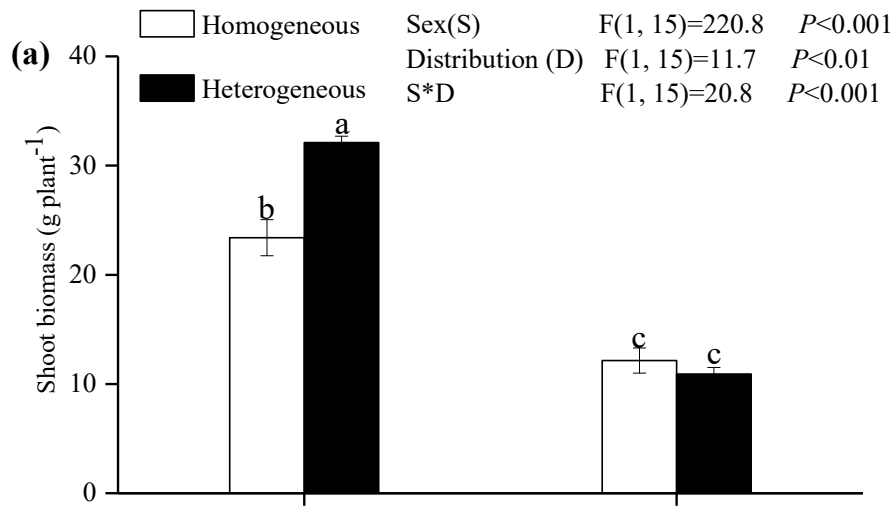
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784 **Figure 6**



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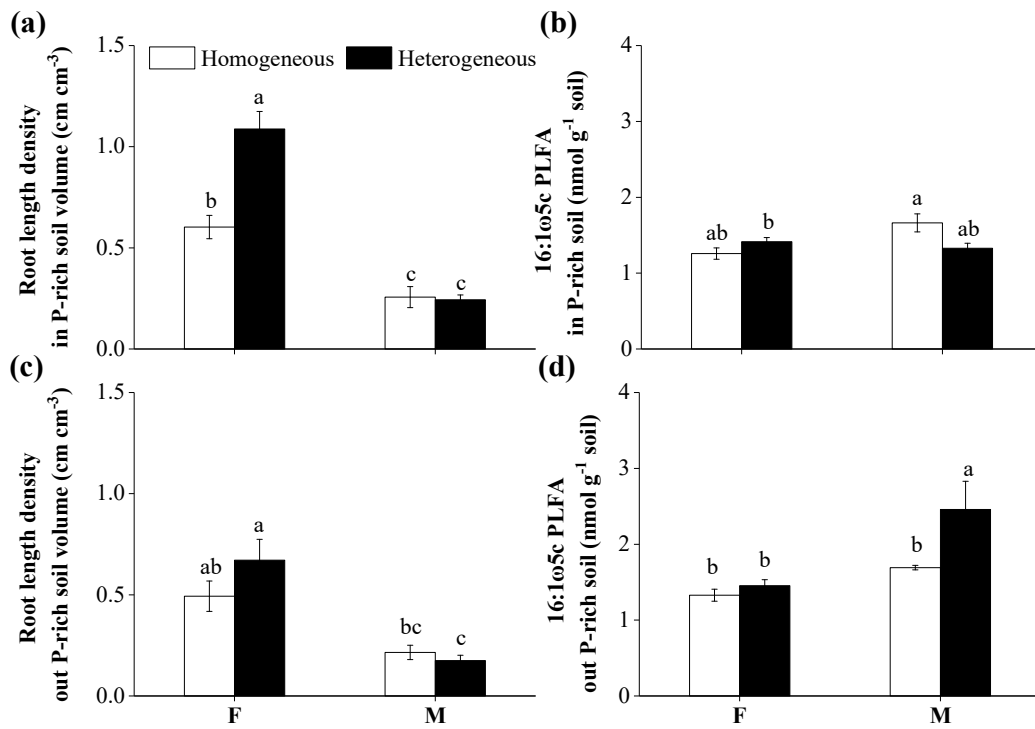
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795 **Figure 7**



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809 **Table 1** Effects of soil P distribution and sex on root length density and PLFA

810 biomarkers 16:1 ω 5c representing extramatrical mycorrhizal hyphal biomass of *P.*
 811 *cathayana* inside and outside P-rich soil.

Factors	Root length density in soil (cm cm ⁻³)				16:1 ω 5c PLFA in soil (nmol g ⁻¹)			
	Inside P-rich patch		Outside P-rich patch		Inside P-rich patch		Outside P-rich patch	
	F	<i>P</i>	F	<i>P</i>	F	<i>P</i>	F	<i>P</i>
Sex	99.5	<0.001	33.1	<0.001	3.8	>0.05	22.2	<0.01
Distribution	15.6	<0.01	1.1	>0.05	1.2	>0.05	7.6	<0.05
Sex \times Distribution	17.4	<0.01	2.7	>0.05	8.8	<0.05	2.5	>0.05

812 F-values from ANOVA are given with their significance levels. Statistically significant

813 P-values are shown in bold.