

UNIVERSITI PUTRA MALAYSIA

THE EFFICACY OF THREE SPECIES OF TRICHODERMA FOR THE CONTROL OF BASAL STEM ROT IN OIL PALM SEEDLINGS

JAYANTHI NAGAPPAN

FPSK(M) 2005 17

THE EFFICACY OF THREE SPECIES OF *TRICHODERMA* FOR THE CONTROL OF BASAL STEM ROT IN OIL PALM SEEDLINGS



By

JAYANTHI NAGAPPAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirements for the Degree of Masters of Science

February 2005



To my beloved parents for their love and patience and my twin sister, Jayasree for her moral support throughout my studies Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science

THE EFFICACY OF THREE SPECIES OF *TRICHODERMA* FOR THE CONTROL OF BASAL STEM ROT IN OIL PALM SEEDLINGS

By

JAYANTHI NAGAPPAN

February 2005

Chairman: Associate Professor Faridah Abdullah, PhD

Faculty: Science

This study evaluated the potential of three *Trichoderma* species, namely *T. harzianum* (isolate BIO T32), *T. longibrachiatum* (BIO T28) and *T. virens* (BIO T128) for the control of *Ganoderma boninense* (EGB 01), the causal pathogen of basal stem rot (BSR) of oil palms in nursery trials. Besides their spore production and antagonistic properties, this study also investigated the growth response of each of the species towards a wide range of temperature and pH conditions. All three species exhibited particular strengths in the growth parameters studied but BIO T32 exhibited consistent and relatively good antagonistic properties and was used as the main inoculant in nursery trials against *G. boninense*. The type and size of wood block were found to influence the success and consistency of the inocula in establishing disease during artificial infection of seedlings. Very low infectivity rates were achieved when inoculum blocks were half to a quarter of the standard 6 x 6 x 12 cm; this size was found to give consistent infection rates leading to approximately 85% mortality. In



nursery trials, seedlings treated with a single inoculum of T. harzianum (T1) gave the lowest and most significant disease severity index (DSI) of 28.34. The conidial drench was stopped at week 14 and the first sign of disease was only observed on week 20. The uninfected and untreated control seedlings gave a DSI of 0 where as, the infected, untreated controls gave a DSI of 86.87. Soils under treatment using a single (T1), two mixed (T2) and three mixed (T3) inocula showed an increase in spore count based on colony forming units (cfu) starting from two weeks after application. When the soil drench was terminated at week 14, the spore count was peak on the 18th, 14th and 10th week for T1, T2 and T3 treatments respectively. Spore counts of BIO T32 were not significantly different on the upper (5 cm) and deeper (15 cm) layer of the treated soils. This study found that when T. harzianum (BIO T32) was used as a single inoculum, it gave the most significant and effective performance as a biological control agent. This was only followed by a mixture of T. harzianum and T. longibrachiatum. Lastly, the use of a combination of three *Trichoderma* species were found to give the poorest disease control, giving a DSI that was not statistically different from the infected, untreated control experiment.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KEUPAYAAN TIGA SPESIS *TRICHODERMA* BAGI PENGAWALAN PENYAKIT REPUT PANGKAL BATANG ANAK POKOK KELAPA SAWIT

Oleh

JAYANTHI NAGAPPAN

Februari 2005

Pengerusi: Profesor Madya Faridah Abdullah, PhD

Fakulti: Sains

Kajian ini menilai potensi tiga spesis *Trichoderma*, terutamanya *T. harzianum* (isolat BIO T32), *T. longibrachiatum* (BIO T28) dan *T. virens* (BIO T128) sebagai kawalan terhadap *Ganoderma boninense* (EGB 01), patogen reput pangkal batang (BSR) pokok kelapa sawit dalam kajian nurseri. Selain penghasilan spora dan ciri-ciri relatif antagonis mereka, kajian ini juga menilai tindakbalas ketiga-tiga spesis ini terhadap julat suhu dan pH yang luas. Ketiga-tiga spesis tersebut mempamerkan ciri-ciri tertentu dalam parameter yang dikaji tetapi BIO T32 mempamerkan ciri relatif antagonis yang baik dan konsisten serta dipilih sebagai inokulan utama dalam kajian nurseri terhadap *G. boninense*. Jenis dan saiz blok kayu didapati mempengaruhi kejayaan dan konsistensi inokula dalam memperkukuhkan penyakit semasa jangkitan secara buatan terhadap anak pokok kelapa sawit. Kadar jangkitan yang amat rendah diperolehi dengan blok inokulum yang bersaiz kecil berbanding dengan saiz 6 x 6 x 12 cm; saiz ini didapati memberi kadar jangkitan yang konsisten sehingga 85% kematian. Dalam kajian



nurseri, anak pokok yang dirawat dengan sejenis aplikasi inokulum T. harzianum (BIO T32) memberikan tahap kemerosotan penyakit (TKP) yang teramat rendah dan signifikasi iaitu sebanyak 28.34. Penggunaan cecair konidia ditamatkan pada minggu ke-14 dan kesan jangkitan hanya diperhatikan pada minggu ke-20. Anak pokok kawalan yang tidak dijangkiti dan tidak dirawat memberi nilai TKP 0, tetapi kawalan yang dijangkiti dan tidak dirawat memberi nilai TKP sebanyak 86.87. Tanah yang dirawat dengan satu (T1), kombinasi dua (T2) dan kombinasi tiga (T3) jenis inokula menunjukkan kenaikan kiraan spora berdasarkan unit pembentukkan koloni (upk) yang bermula dari minggu ke-2 selepas aplikasinya. Apablia aplikasi larutan inokulum ditamatkan pada minggu ke-14, kiraan spora memuncak pada minggu ke 18, 14 dan 10 untuk rawatan T1, T2 dan T3 masing-masing. Kiraan spora BIO T32 tidak signifikan pada tahap atas (5 cm) dan dalam (15 cm) tanah yang dirawat. Kajian ini mendapati apabila sejenis inokulum T. harzianum (BIO T32) digunakan, ia sangat signifikan dan efektif sebagai agen kawalan biologi. Ini diikuti dengan kombinasi T. harzianum dan T. longibrachiatum. Akhirnya, kombinasi ketiga-tiga spesis Trichoderma didapati memberikan pengawalan penyakit yang tidak memuaskan dengan TKP yang tidak signifikan dari anak pokok kawalan yang dijangkiti dan tidak dirawat.

ACKNOWLEDGEMENTS

I would like to express my deepest and sincere gratitude to Prof. Madya Dr. Faridah Abdullah who has guided, supervised and supported my academic work and also thank her for providing financial support during the study from her IRPA project.

I would like to express my heartful thanks to Assoc. Prof. Dr. Umi Kalsom Yusuf and Assoc. Prof. Dr. Zainal Abidin Mior Ahmad for their invaluable advice throughout the research and thesis preparation.

I would like to thank the staff members of the Department of Biology of Soil Science for the use of the greenhouse and Department of Forestry for the cutting of wood blocks.

Special thanks were also extended to lab-mates Dr. G. N. M. Ilias, Mr. Nelson and Mrs. Shamala Sundram for their kind assistance during this study.

Special thanks were reserved for my parents, sister and friends, namely Mrs. Jasmin and Miss Gunavathi for putting up with me during the course of this study.

I certify that an Examination Committee met on to conduct the final examination of Jayanthi Nagappan on her degree thesis entitled "The Efficacy Of Three Species Of *Trichoderma* For The Control Of Basal Stem Rot Of Oil Palm Seedlings" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination committee are as follows:

Professor Madya Dr. Radzali B. Muse, Ph.D. Faculty of Science Universiti Putra Malaysia (Chairman)

Professor Dr. Sariah Meon, Ph.D. Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Dr. Inon Sulaiman, Ph.D. Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Professor Dr. Baharudin Salleh, Ph.D. School of Biological Sciences Universiti Sains Malaysia (External Examiner)

GULAM RUSUL RAHMAT ALI, Ph.D.

Professor/Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee are as follows:

Faridah Abdullah, PhD

Associate Professor Faculty of Science, Universiti Putra Malaysia (Chairman)

Umi Kalsom Yusuf, PhD Associate Professor Faculty of Science, Universiti Putra Malaysia (Member)

Zainal Abidin Mior Ahmad, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

> AINI IDERIS, PhD Professor/Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

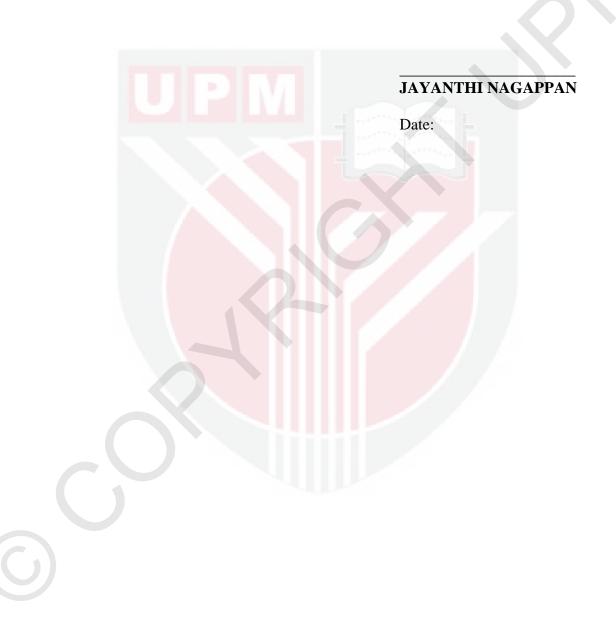


TABLE OF CONTENTS

	Page
DEDICATION	2
ABSTRACT	3
ABSTRAK	5
ACKNOWLEDGEMENTS	7
APPROVAL DECLARATION	8 10
LIST OF TABLES	10
LIST OF FIGURES	14
LIST OF ABBREVIATIONS/NOTATIONS/GLOSSARY OF TERMS	
CHAPTER	
I. INTRODUCTION	21
II. LITERATURE REVIEW	
The oil palm	28
Occurrence of basal stem rot of oil palm	29
Symptoms of basal stem rot of oil palm	30
Taxonomic studies of <i>Ganoderma</i>	31
Pathogenicity studies of <i>Ganoderma boninense</i> Control of basal stem rot	33 33
Biological control	33
Taxonomy and morphology of <i>Trichoderma</i>	38
Distribution and occurrence of <i>Trichoderma</i>	40
Establishment and proliferation of <i>Trichoderma</i> in soil	41
Biological control approaches with <i>Trichoderma</i>	42
Efficacy of <i>Trichoderma</i> as a biological control agent	46
III. BIOLOGICAL CHARACTERISTICS OF T. HARZIANUM	
(RIFAI), T. LONGIBRACHIATUM (RIFAI) AND T. VIRENS	5
(J. MILLER, GIDDENS AND FOSTER) Arx.	
INTRODUCTION	49
METHODOLOGY	.,
Source of test fungi and pathogen	53
Fungal subcultures	53
Morphological characteristics of <i>Trichoderma</i> colony and cul	
Effect of pH: Growth of Trichoderma by mycelial dry weight	
Effect of pH: Radial growth of Trichoderma by mycelial exte	ension 57
Effect of pH: Spore production of Trichoderma on agar medi	a 58
Enumeration of spore counts by heamocytometer	58
Effect of temperature on growth of Trichoderma	59

Growth inhibition of Ganoderma boninense by dual culture	59
Data analysis	61
RESULTS	
Morphological characteristics of Trichoderma colony and cultures	62
Effect of pH: Growth of Trichoderma by mycelial dry weight	66
Effect of pH: Radial growth of Trichoderma on agar media	71
Effect of pH: Spore production of Trichoderma on agar media	74
Effect of temperature on growth of <i>Trichoderma</i>	75
Screening for antagonistic properties	78
DISCUSSION	82

IV. THE EFFECT OF INOCULUM CHARACTERISTICS ON THE PATHOGENICITY OF GANODERMA BONINENSE ON OIL PALM SEEDLINGS

INTRODUCTION	87
METHODOLOGY	
Source of oil palm seedlings, potting media and G. boninense inocula	90
Wood type and its preparation as fungal inocula	90
Method of infecting oil palm seedlings with G. boninense	91
Establishment of disease by two types of inoculum substrate	92
Establishment of disease by smaller sized inoculum substrates	92
Assessment of fungal infection:	93
i. Recognition of Disease Symptoms	93
ii. Scoring by Disease Severity Index (DSI)	94
Data analysis	95
RESULTS	
Disease establishment on 6 x 6 x 12 cm rubber wood inocula	97
Disease establishment on smaller sized inoculum blocks	99
Disease development on seedlings infected by oil palm wood blocks	105
DISCUSSION	108
GREENHOUSE TRIALS: THE APPLICATION OF CONIDIAL	

V. GREENHOUSE TRIALS: THE APPLICATION OF CONIDIAL SOIL DRENCH OF *T. HARZIANUM* AS A SINGLE AND AS A MIXED SPECIES INOCULA AND THE SURVIVAL OF CONIDIA IN TREATED SOILS

INTRODUCTION	111
METHODOLOGY	
Source of fungal cultures	117
Preparation of wood blocks as fungal substrate	117
Preparation of G. boninense substrate inocula	117
Preparation of Trichoderma supplemented mulch	118
Source of oil palm seedlings and potting media	118

	Artificial infection of oil palm seedlings	118
	Preparation of conidial soil drench	119
	Application of Trichoderma conidial soil drench	119
	Estimation of Trichoderma spore counts used as conidial soil drench	121
	Assessment of fungal infection:	121
	i. Recognition of Disease Symptoms	121
	ii. Scoring by Disease Severity Index (DSI)	121
	Air-dry weight of leaves and roots	122
	Determination of soil acidity (pH) and moisture content (MC)	122
	Quantification of <i>Trichoderma</i> species from treated pots using	
	colony forming unit (cfu)/g soil	123
	Data analysis	125
	RESULTS	
	Estimation of <i>Trichoderma</i> spore counts used as conidial soil drench	126
	Disease establishment on experimental control seedlings	127
	Disease establishment on treatment seedlings (T1)	130
	Disease establishment on treatment seedlings (T2)	132
	Disease establishment on treatment seedlings (T3)	134
	Disease severity index	137
	Air-dry weight of leaves and roots	139
	Determination of soil acidity (pH) and moisture content (MC)	140
	Quantification of Trichoderma from soil of treated seedlings by	
	cfu/g soil	142
	DISCUSSION	147
VI	DISCUSSION AND CONCLUSION	155
	REFERENCES	159
	APPENDICES	172
	BIODATA OF THE AUTHOR	218

LIST OF TABLES

Tables		Page
3.2	Concentrations of lactic acid and NaOH used to make up the pH readings in agar media	57
3.7	Percentage inhibition of radial growth (PIRG) of <i>G. boninense</i> (EGB 01) and colony degradation time of <i>Trichoderma</i> species against <i>G. boninense</i>	79
4.1	Disease severity index (DSI) based on modifications for oil palm seedlings (Ilias, 2000)	94
4.2	Sequence of signs and symptoms of disease establishment on oil palm seedlings when using 6 x 6 x 12 cm size rubber wood block inocula	98
4.3	Sequence of signs and symptoms of disease establishment on oil palm seedlings when using 6 x 6 x 6 cm size rubber wood block inocula	100
4.4	Sequence of signs and symptoms of disease establishment on oil palm seedlings when using 3 x 3 x 6 cm rubber wood block inocula	103
4.5	Effect of two types of inoculum substrates on oil palm seedlings at 25 weeks after inoculation	105
4.6	Disease severity index (DSI) based on signs and symptoms on oil palm seedlings for a period of 25 weeks	106
5.1	Mean (\pm standard error) spore counts per ml on heamocytometer of <i>Trichoderma</i> species prior to usage as conidial soil drench for each treatment series for the stated weeks	127
5.2	Sequence of signs and symptoms of disease establishment on oil palm seedlings inoculated with EGB 01 (C2) for a period of 24 weeks	128
5.3	Sequence of signs and symptoms of disease establishment on oil palm seedlings treated with BIO T32 (T1) for a period of 24 weeks	131
5.4	Sequence of signs and symptoms of disease establishment on oil palm seedlings treated with a mixture of two <i>Trichoderma</i> species (T2) for a period of 24 weeks	133
5.5	Sequence of signs and symptoms of disease establishment on oil palm seedlings treated with a mixture of three <i>Trichoderma</i> species (T3) for 24 weeks	135

5.6	A summary of signs and symptoms of oil palm seedlings with five treatment series at week 24 after inoculation with EGB 01	136
5.7	Disease severity index (DSI) based on signs and symptoms on oil palm seedlings for a period of 24 weeks after inoculation with EGB 01.	138
5.8	Mean (\pm standard error) air-dry weight of leaves and roots of oil palm seedlings with treatment series at week 24 after inoculation with EGB 01	139
5.9	Mean (\pm standard error) of soil pH for a period of 22 weeks taken from soils of oil palm seedlings of the five treatment series	140
5.10	Mean (\pm standard error) of soil moisture content (MC) from control and treated soils of oil palm seedlings taken for a period of 22 weeks	141
5.11	Mean (\pm standard error) of colony forming units (cfu) of <i>Trichoderma</i> species prior to treatment on the inoculated and non-inoculated oil palm seedlings for a period of 22 weeks at two different depths	142



LIST OF FIGURES

Figures		Page
3.1	A slide culture chamber.	55
3.2	A seven-day old culture of <i>T. harzianum</i> (isolate BIO T32) on PDA showing the upper surface (L) and lower surface (R).	62
3.3	<i>T. harzianum</i> (isolate BIO T32) culture seen under a microscope at 400X magnification: (a) conidiophore, (b) phialide and (c) a conidium.	63
3.4	A seven-day old culture of <i>T. longibrachiatum</i> (BIO T28) on PDA showing the upper surface (R) and lower surface (L).	64
3.5	<i>T. longibrachiatum</i> (BIO T28) culture seen under a microscope at 400X magnification: (a) conidiophore, (b) phialide and (c) a conidium.	64
3.6	A seven-day old culture of <i>T. virens</i> (BIO T128) on PDA showing the upper surface (L) and lower surface (R).	65
3.7	<i>T. virens</i> (BIO T128) culture seen under a microscope at 400X magnification: (a) conidiophore, (b) phialide and (c) a conidium.	66
3.8	Bars indicate mean mycelial dry weights (g) of the three <i>Trichoderma</i> species cultured in liquid medium at different pH.	67
3.9	Oven-dried BIO T32 mycelia: (a) $L - R$: pH 2.7, 3.0, 4.0 and 5.0 in three replicates; (b) $L - R$: pH 5.11 of control; (c) $L - R$: pH 6.0, 7.0, 7.6 and 8.0 in three replicates.	68
3.10	Oven-dried BIO T128 mycelia: (a) $L - R$: pH 2.7, 3.0, 4.0 and 5.0 in three replicates; (b) $L - R$: pH 5.11 of control; (c) $L - R$: pH 6.0, 7.0, 7.6 and 8.0 in three replicates.	69
3.11	Oven-dried BIO T28 mycelia: (a) $L - R$: pH 2.7, 3.0, 4.0 and 5.0 in three replicates; (b) $L - R$: pH 5.11 of control; (c) $L - R$: pH 6.0, 7.0, 7.6 and 8.0 in three replicates.	70
3.12	Bars indicate mean radial growth (mm/day) of the three <i>Trichoderma</i> species cultured on agar medium at different pH.	72
3.13a	Growth response of isolate BIO T32 in a range of pH media at 28°C.	72
3.13b	Growth response of isolate BIO T128 in a range of pH media at 28°C.	73

3.13c	Growth response of isolate BIO T28 in a range of pH media at 28°C.	73
3.14	Bars indicate spore production (at spores per ml) of three <i>Trichoderma</i> species cultured in a range of pH 2.7 to 8.0 on PDA.	75
3.15	Bars indicate radial growth rate (mm/day) of three <i>Trichoderma</i> species cultured on agar medium at a temperature range of 15 to 40 °C.	76
3.16a	Growth response of BIO T32 at different temperatures.	77
3.16b	Growth response of BIO T128 at different temperatures.	77
3.16c	Growth response of BIO T28 at different temperatures.	78
3.17a	Inhibition of <i>G. boninense</i> by isolate BIO T32.	80
3.17b	Inhibition of <i>G. boninense</i> by isolate BIO T128.	80
3.17c	Inhibition of G. boninense by isolate BIO T28.	81
4.1	The four disease classes: (a) Class 0 - Healthy palms with green a symptomless leaves; (b) Class 1 - Palms stunted and/or leaf discoloration; (c) Class 2 - Appearance of white mycelia at base of stem; (d) Class 3 - Appearance of sporophore(s) at base of stem and (e) Class 4 – Dead palm with dried up and almost/totally brown leaves, with or without sporophore(s).	96
4.2	Plants infected with 6 x 6 x 12 cm rubber wood block showing a DSI of 85 (bottom row).	99
4.3	Plants infected with 6 x 6 x 6 cm rubber wood block showing a DSI of 40 (bottom row).	101
4.4	Plants infected with $3 \times 3 \times 6$ cm rubber wood block showing a DSI of 35 (bottom row).	104
4.5	Disease progress curves of oil palm seedlings infected with different sizes of rubber wood inocula.	104
4.6	Top row: Seedlings infected with 6 x 6 x 12 cm oil palm wood blocks showing no signs of disease.	106
4.7	Oil palm seedlings infected with oil palm wood blocks inocula $(6 \times 6 \times 6 \text{ cm})$ showing no signs of disease development at 25 weeks of inoculation.	107

5.1	Colony forming units (cfu) of the 3 <i>Trichoderma</i> species on Rose Bengal Agar (RBA) showing different colony characteristics: a) dark green of <i>T. harzianum;</i> b) pale yellow of <i>T. longibrachiatum</i> and c) greenishbrown of <i>T. virens</i> .	125
5.2	Up-rooted infected, untreated control seedlings (C2) showing a DSI of 86.87 at 24 weeks after inoculation.	129
5.3	Up-rooted uninfected, untreated control seedlings (C1) showing a DSI of 0 at 24 weeks after inoculation.	129
5.4	Up-rooted oil palm seedlings given a single mode (T1) of application with <i>T. harzianum</i> (BIO T32), giving a DSI of 28.34 at 24 weeks after inoculation.	131
5.5	Up-rooted oil palm seedlings given an application of a mixture of <i>T. harzianum</i> and <i>T. longibrachiatum</i> (T2), showing a DSI of 55 at 24 weeks after inoculation.	134
5.6	Up-rooted oil palm seedlings given an application of a mixture of <i>T. harzianum, T. longibrachiatum</i> and <i>T. virens</i> (T3), showing a DSI of 81.66 at 24 weeks after inoculation.	136
5.7	Disease progress curves of oil palm seedlings infected with different sizes of rubber wood inocula.	138
5.8	Colony forming units (cfu/g soil) of BIO T32 from treatment T1 over a period of 22 weeks at two different depths.	144
5.9	Colony forming units (cfu/g soil) of BIO T32 and BIO T28 from treatment T2 over a period of 22 weeks at two different depths.	145
5.10	Colony forming units (cfu/g soil) of BIO T32, BIO T28 and BIO T128 from treatment T3 over a period of 22 weeks at two different depths.	146

LIST OF ABBREVIATIONS

- G without Ganoderma boninense
- + G with Ganoderma boninense
- BIO T28 Trichoderma longibrachiatum
- BIO T32 Trichoderma harzianum
- BIO T128 Trichoderma virens
- BSR Basal stem rot
- C1 Control 1
- C2 Control 2
- C₆H₈O₇.H₂O Citric Acid
- CFU Colony Forming Unit
- DSI Disease Severity Index
- EGB 01 Isolate of *Ganoderma boninense* (*Elaeis guineensis*-Banting)
- MC Moisture Content
- MEA Malt Extract Agar
- MPOB Malaysian Palm Oil Board
- MW Molecular Weight
- Na₂HPO₄ Sodium Hydrogen Phosphate
- NaOH Sodium Hydroxide
- OPMF Oil Palm Mesocarp Fibre
- PCNB Pentachloronitrobenzene
- PDA Potato Dextrose Agar

PIRG	Percentage Inhibition Radial Growth
PORIM	Palm Oil Research Institute of Malaysia
PP 28	Isolate of Ganoderma boninense from Palm Oil Research Institute of
	Malaysia (PORIM), now know as Malaysian Palm Oil Board (MPOB)
RBA	Rose Bengal Agar
rpm	Rotation per minute
T1	Treatment 1, <i>T. harzianum</i> (BIO T32)
T2	Treatment 2, <i>T. harzianum</i> + <i>T. longibrachiatum</i> (BIO T28)
Т3	Treatment 3, <i>T. harzianum</i> + <i>T. longibrachiatum</i> + <i>T. virens</i> (BIO T128)
UPM	Universiti Putra Malaysia
w.a.i.	Weeks after inoculation
WP	Wettable Powder

C

CHAPTER I

INTRODUCTION

Several million hectares in the world today are planted with commercially important edible oil crops that represent a significant fraction of the resources of the countries concerned (Ariffin and Idris, 2002). Among these species is the oil palm (*Elaeis guineensis*) as an important crop in the topical regions because of its two main raw materials produced, the palm oil and palm kernel oil. Currently, Malaysia is the leading producer of palm oil, with a total production of about 12.5 million tonnes for the year 2004 (MPOB Statistic, 2004) and seeks to maintain dominance in this field.

In order to maintain the current production as well as to strive towards higher yields, every aspect of oil palm cultivation will need to be carefully managed; one of these is in disease management. From seed germination to field planting, the oil palm is prone to attack by various disease-causing organisms, the most common being fungi. Nevertheless, diseases affecting seeds and nursery seedlings are under control in most cases and do not pose a serious threat to the industry. It is diseases of field palms, particularly a basal stem rot (BSR) caused by *Ganoderma* spp. that threaten crop development and requires urgent solution.

Ganoderma has been known to attack oil palms since the early years when the crop was introduced into this country (Turner, 1981). The disease was recognized since late 1920's (Thompson, 1931; as cited in Ariffin *et al.*, 1996) but was regarded as of

negligible importance since only palms of over 25 years in age were affected. It was not until 1957 that BSR incidence was reported to increase at an alarming rate when younger palms of 10 to 15 years in age were also infected (Turner and Bull, 1967). Gurmit (1991) reported that the disease could set in as early as 12 to 24 months but the effects were only noticeable when they were four to five years old.

Currently, the approaches used to control the disease are mainly by adoption of hygienic cultural practices and the use of chemical control (tridemorph, carboxin, triadimefon, triadimenol, flutriafol, propiconazole and difenoconazole) to a certain degree (Gurmit, 1991). Bayleton[®] is one chemical that has been used in laboratory studies and in field trials as trunk injection (PORIM, 1984). Other fungicides tested in field trials were Benlate[®] T-20, Calixin[®], Bayfidan[®], Thiram[®] and Dazomet[®] but results from these trials were inconclusive (Ariffin and Idris, 1991). Several research institutes have studied this disease and developed means of control but despite many investigations and some 80 years of research no satisfactory solutions in terms of effectiveness, ease of use and cost could be offered.

During the next decade biological control may become an important component of plant disease management practices. The demand for alternatives to chemical control of plant pathogens has become stronger owing to concerns about the safety and environmental impacts of chemicals. The possibility of control of *Ganoderma* should be approached through manipulation of biological agents. Investigations on

the use of fungi such as Trichoderma (Wijesekera et al., 1996; Ilias and Abdullah,

1999), *Aspergillus* (Shukla and Uniyal, 1989) and *Penicillium* (Dharmaputra *et al.*, 1989) as antagonists of *Ganoderma* in culture have been reported. Particular attention is focused on species of *Trichoderma* that may not as yet given any 'wonder drugs' such as penicillin but has the potential to produce enzymes and to attack or inhibit other fungi (Samuels, 1996; Ilias and Abdullah, 1998; Ilias and Abdullah, 1999). Weindling (1932) was the first to discover the antagonistic ability of *Trichoderma* on the plant pathogen *Rhizoctonia solani*. Two major discoveries were reported; the first was that *Trichoderma* killed the pathogen by physical strangulation and the second was by killing them a short distance away through the production of toxic compounds in the media. Ilias and Abdullah (1998) showed situations where the fungal mycelia coiled tightly around the host hyphae resulting in physical strangulation, as well as the formation of hook-like structures by *Trichoderma*, which puncturing the fungal host cells. *Trichoderma* spp. was also found to produce volatile and non-volatile antibiotics (Dennis and Webster, 1971a,b; Ilias and Abdullah, 1998).

The most studied species of *Trichoderma* acting against antagonists of plant pathogens reported were *Trichoderma harzianum* (Wells *et al.*, 1972; Elad *et al.*, 1980; Chamswarng, 1992; Ilias and Abdullah, 1998), *T. virens* (Papavizas and Lewis, 1989; Sariah and Chan, 1999) and *T. longibrachiatum* (Chamswarng *et al.*, 1992; Sreevinasaprasad and Manibushanrao, 1993; Saravanan *et al.*, 2003). An *in vitro* study by Ilias and Abdullah (1999) showed that growth of *Ganoderma boninense* was inhibited using culture filtrates of *T. harzianum* and *T. virens* respectively. Further *in vitro* studies by Abdullah and Jayanthi (1999) found that a metabolite mixture of strains of *T. harzianum*, *T. virens* and *T. longibrachiatum* resulted in a better antagonistic performance against growth of *Ganoderma boninense* than when applied singly.

Trials on using disease-controlling agents are still under explored and play an important role in inducing disease control in oil palm seedlings. A major obstacle towards achieving this objective is the inability to reproduce artificial infection accurately and consistently. Studies by Khairudin (1991), was the most successful and practicable thus far and is the model upon which the present study was based. However, the success in establishing induced disease in oil palm seedlings is meaningful only if the data can be quantified. Many attempts have been made in the earlier years to establish Koch's Postulate, one of which was by Navaratnam and Chee (1965). Khairudin *et al.* (1991) found that the oil palm seedlings were infected by rubber wood inocula but not on oil palm mesocarp fibre (OPMF). It was thus concluded that the type of substrate inocula used determined the success of infection by *G. boninense*. Besides the size, type and age of inoculum may also play an important role in establishing infection by *Ganoderma* (Khairudin, 1994; Abdullah *et al.*, 2001).

Based on *in vitro* experiments by Ilias and Abdullah (1998), a further step was taken to test the antagonist activity in greenhouse trials. An *in vivo* trial carried out by Ilias (2000) found a strain of *T. harzianum* used singly in the form of conidial soil drench gave better results in suppressing diseased in oil palm seedlings than *T. virens*. Abdullah *et al.* (2003b), reported that *T. harzianum* gave a DSI value of 5 after 20 weeks of treatment. Studies using a combination of *Trichoderma* species with other fungal species or chemical adjuvants has not been tried and they could be much more effective than when applied alone. Research in this directions include the control of *R. solani*, causal pathogen of root rot of eggplant, with *T. harzianum* combined with PCNB in soil (Hadar *et al.*, 1979), *Sclerotium rolfsii* Sacc., seedling blight of barley by using seed coating with a combination of *Trichoderma* spp. and *Bacillus* sp. (Chamswarng *et al.*, 1992) and against *Phytophthora erythroseptica*, the causal pathogen of pink rot of potato and root and stem rot of tomato by using a combination of TrichodexTM and *T. virens* (Etebarian *et al.*, 2000).

Trichoderma species have been used in commercial preparations for biological control of fungal induced plant diseases (Samuels, 1996). *T. harzianum* is the active ingredient in TrichodexTM, which is used against post-harvest rot of apple. *T. harzianum* is combined with *T. polysporum* in the product Binab-TTM, which is used in the control of would decay and wood rot (Ricard, 1981). Hence, further trials are needed to determine whether a combination of reagents performed better to arrest the *Ganoderma* lesion.

To assess the efficacy of *Trichoderma* as a biological agent, the disease establishment of the pathogen onto the experimental plants must approach 90%. The state of 'infection' assessed by Navaratnam and Chee (1965) was as 'foliar symptoms'. There was a limitation in this concept. Due to this fact, Khairudin

(1991) assessed the 'infection' of the seedlings by the production of *Ganoderma* sporophores. This study was to establish a good infectivity with more than 80% infection. To document a spectrum of visual signs, from the early to late stages of infection, a disease severity index (DSI) was developed as an aid to quantify the disease establishment. Apart from that, the population dynamics of *Trichoderma* in the soil of treated seedlings was also assessed.

The four main objectives of the study are summarized as follows:

- a) To determine the biological characteristics of the three species-isolates of *Trichoderma* namely, *T. harzianum* (isolate BIO T32), *T. longibrachiatum* (BIO T28) and *T. virens* (BIO T128).
- b) To determine the success of an infectivity with two types of wood blocks (rubber and oil palm) that were tested on their relative suitability as substrate inocula in establishing disease to oil palm seedlings. Apart from that, the effect of smaller sized inoculum blocks in establishing disease on oil palm seedlings was also investigated.

c) To assess the treatments in the control of basal stem rot of oil palm seedlings using single and mixed inocula of *T. harzianum* (BIO T32), *T. longibrachiatum* (BIO T28) and *T. virens* (BIO T128) in greenhouse trials. To monitor the colony forming units (cfu) of *Trichoderma* in the soils of treated seedlings throughout the experiment. The hypothesis of this study was to find out whether *T. harzianum* (BIO T32) with an integration of other *Trichoderma* species (*T. longibrachiatum* and *T. virens*) was able to give better control towards *G. boninense*, causal pathogen of basal stem rot of oil palm.



REFERENCES

Abdullah, F. and Ilias, G. N. M. 2002. Mode of hyperparasitism of *Trichoderma* harzianum on Ganoderma boninense. In Seminar Microscopy: An Update on Light and Electron Microscopy. Institute of Bioscience, 7 - 8 May, 2002, Universiti Putra Malaysia.

Abdullah, F. and Jayanthi, N. 1999. The efficacy of *Trichoderma* metabolites as antagonists of *Ganoderma boninense*. In *Research and Development Priorities in the New Millennium*, Malaysia Science and Technology Congress, Kuala Lumpur, 25-27 October, 1999, Malaysia.

Abdullah, F. and Nelson, M. 2000. Cross pathogenicity of *Ganoderma boninense* and *G. philippi* on oil palm and sentang. *Journal of Bioscience* 11(1&2):11-15.

Abdullah, F., Ilias, G. N. M., Nelson, M. and Umi Kalsom Yusuf. 2003b. Disease assessment and the efficacy of *Trichoderma* as a biocontrol agent of basal stem rot of oil palms. *Bulletin Research Science Putra* 11(2), pp:31-33.

Abdullah, F., Ilias, G. N. M., Sivananthan, M., Zainal, A. M. A. and Umi Kalsom, M. Y. 1999. *In vitro* and *in vivo* studies of *Trichoderma harzianum* as a biocontrol agent of *Ganoderma bonineense*. In 22nd *Microbiology Symposium and JSPS-NRCT/DOST/LIPI/VCC Seminar*, ed. Sijam, A. K., Jalil M. Z. and Halimi, M. S. Malaysian Microbiological Society, pp:98

Abdullah, F., Izzati, M. Z. N. A., Ilias, G. N. M., Nelson, M. and Umi Kalsom Yusuf. 2003a. The efficacy of *Trichoderma harzianum* as a biocontrol agent of basal stem rot of oil palm. In 2003 International Palm Oil Congress – PIPOC (Agriculture), Malaysian Palm Oil Board (MPOB), pp:1056-1060.

Abdullah, F., Jayanthi, N. Ilias, G. N. M. and Nelson, M. 2001. Properties of substrate inocula and plant hosts in disease establishment by *Ganoderma boninense*. In 2001 International Palm Oil Congress - PIPOC (Agriculture), Malaysian Palm Oil Board (MPOB), pp:618-623.

Agosin, E. and Aguilera, J. M. 1998. Industrial production of active propagules of *Trichoderma* for agriculture uses. In *Trichoderma* and *Gliocladium*, ed. Harman, G. E. and Kubicek C. P., 2:205-227.

Ariffin, D and Idris, A. S. 1991. Investigation of the control of *Ganoderma* with dazomet. In *Proceeding of the 1991 International Palm Oil Conference* (Agriculture), ed. Yusof *et al.*, Palm Oil Research Institute of Malaysia (PORIM), pp:424-429.

Ariffin, D. 2000. Major diseases of oil palm. In *Advances in oil Palm Research*. Vol. 1, ed. Yusof Basiron, Jalani, B. S. and Chan, K. W. pp:596-622.

Ariffin, D. and Idris, A. S. 1990. Progress on *Ganoderma* research at PORIM. In *Proceedings of the Ganoderma Workshop*, 11 September 1990, ed. Ariffin, D. and Jalani, S. pp:113-131. Palm Oil Research Institute of Malaysia, Bangi, Selangor, Malaysia.

Ariffin, D. and. Idris, A. S. 2002. Paper 12: Plenary paper: Progress Research on *Ganoderma* basal stem rot of oil palm. In *Seminar on Elevating the National Oil Palm Productivity and Recent Progress in the Management of Peat and Ganoderma*. 6 - 7 May 2002. Hotel Equatorial, Malaysia.

Ariffin, D., Idris, A. S. and Marzuki, A. 1996. Chemically assisted biological control of basal stem rot of oil palm. IRPA (RM6) Project Completion Report. Palm Oil Research Institute of Malaysia, Bangi, Selangor, Malaysia, pp:69.

AWPA. 1986. America Wood-Preserves Association's Standards. Book of Standards, Stevensville, Maryland, USA.

Baker, K. F. and Cook, R. J. 1974. *Biological Control of Plant Pathogens*. W. H. Freeman & Co., San Francisco, CA. pp:433.

Batta, Y. A. 2004a. Effect of treatment with *Trichoderma harzianum* Rifai formulated in invert emulsion on postharvest decay of apple blue mold. *International Journal of Food Microbiology* 96:281-288.

Batta, Y. A. 2004b. Postharvest biological control of apple gray mold by *Trichoderma harzianum* Rifai formulated in an invert emulsion. *Crop Protection* 23:19-26.

Beagle, J. E. and Papavizas, G. C. 1985. Survival and proliferation of propagules of *Trichoderma* spp. and *Gliocladium virens* in soil and in plant rhizospheres. *Phytopathology* 75:729-732.

Bissett, J. 1991. A revision of the genus *Trichoderma* III. Section Pachybasium. *Canadian Journal of Botany* 69:2372-2417.

Black, G. R. 1965. Bulk density: In *Methods of Soil Analysis* (Part II). C. A. Black. American Society of Agronomy, Madison, USA, pp:347-349.

Bruce, A. and Highley, T. L. 1991. Control of growth of wood decay Basidiomycetes by *Trichoderma* spp. and other potential antagonists fungi. *Forest Products Journal* 4 (2):63-67.

Bruce, A. Johnstone, C. J. and McVey, A. P. 1987. Susceptibility of *Lentinus lepideus* (Fr.: Fr.) Fr. to volatiles produced by *Trichoderma* spp. Internat. Research Group on Wood Press Document No: IRG/WP/1316, pp:13.

Bruce, A., Austin, W. J. and King. B. 1984. Control of growth of *Lentinus lepideus* by volatiles from *Trichoderma*. *Transactions of the British Mycological Society* 82:423-428.



Buchanan, P. K. 2001. A taxonomic overview of the genus *Ganoderma* with special reference to species of medicinal and neutriceutical importance. In *International Symposium Ganoderma Science*, Auckland, New Zealand, 27-29 April, 2001.

Burgess, D. R. and Hepworth, G. 1996. Biocontrol of sclerotinia stem rot (*Sclerotinia minor*) in sunflower by seed treatment with *Gliocladium virens*. *Plant Pathology* 45:583-592.

Campbell, C. L. and Madden, L. V. 1990. *Introduction to Plant Disease Epidemiology*. John Wiley & Sons.

Chamswarng, C. 1992. Biological control of sclerotium stem rot of tomato in Thailand. *Journal of Plant Protection In the Tropics* 9 (1):77-83.

Chamswarng, C., Gesnara, W. and Korpraditskul, V. 1992. Control of sclerotium seedling blight of barley by using seeds coated with *Trichoderma* spp. and *Bacillus* sp. *Journal of Plant Protection In the Tropics* 9 (1):69-75.

Chaverri, P., Samuels, G. J. and Stewart, E. L. 2001. *Hypocrea virens* sp. nov., the teleomorph of *Trichoderma virens*. *Mycologia* 93:1113-1124.

Cheah, S. C. 1997. The biotechnology of oil palm. In *The ABSP Global Conference*, held at Asilomar Conference Center, Pacific Grove, California, USA, 28-30 April 1997.

Chet, I. 1987. *Trichoderma* – Application, mode of action and potential as a biocontrol agent of soilborne plant pathogenic fungi. In *Innovative Approaches to Plant Disease Control*, ed. Chet. I., pp:137-160. John Wiley & Sons. Inc. New York, N. Y.

Chet, I. and Baker, R. 1980. Induction of suppressiveness to *Rhizoctonia solani* in soil. *Phytopathology* 70:994-998.

Chinchilla, C. and Richardson, D. L. 1987. Four potentially destructive diseases of the oil palm in Central America. In 1987 International Oil Palm/Palm Oil Conference: Progress and Prospect; Conference I: Agriculture, 23-26 June 1987, ed. Abdul Halim et al. pp:468-470. Palm Oil Research Institute of Malaysia, Bangi, Selangor, Malaysia.

Cook, R. J. and Baker, K. F. 1983. *The Nature and Practice of Biological Control of Plant Pathogens*. The American Phytopathological Society, USA, pp:539.

Danielson, R. M. and Davey, C. B. 1973a. Non-nutritional factors affecting the growth of *Trichoderma* in culture. *Soil Biology and Biochemical* **5**:495-504.

Danielson, R. M. and Davey, C. B. 1973b. Carbon and nitrogen nutrition of *Trichoderma*. *Soil Biology and Biochemical* **5**:505-515.



Danielson, R. M. and Davey, C. B. 1973c. Effects of nutrients and acidity on phialospores germination of *Trichoderma in vitro*. *Soil Biology and Biochemical* **5**:517-524.

Darmono, T. W. 1998. Development and survival of *Ganoderma* sp. In *Oil Palm Tissue*. 1998 International Palm Oil Conference, Nusa Dua, Bali, Sumatera, September 23-25, 1998. pp:613-617.

Dennis, C and Webster, J. 1971a. Antagonistic properties of species groups of *Trichoderma*. I. Production of non-volatile antibiotics. *Transactions of the British Mycological Society* 57 (1):25-39.

Dennis, C. and Webster, J. 1971b. Antagonistic properties of species groups of *Trichoderma*. II. Production of volatile antibiotics. *Transactions of the British Mycological Society* 57 (1):41-48.

Dharmaputra, O. S., Purba, R. Y. and Sipayung, A. 1994. Research activities on the biology and control of *Ganoderma* at Seameo Biotrop and IOPRI Marihat. In *Proceedings of an International Workshop on Perennial Crop Diseases Caused by Ganoderma*, ed. M. Holerness, CABI, 1 - 3 December, Selangor, Malaysia.

Dharmaputra, O. S., Tjitrosomo, H. S. and Abadi, A. L. 1989. Antagonistic effect of four fungal isolates to *Ganoderma boninense*, the causal agent of basal stem rot of oil palm. *Biotropia* 3:41-49.

Doi, S. and Mori, M. 1993. Antifungal properties of metabolites produced by *Trichoderma* isolates from sawdust media of edible fungi against wood decay fungi. *Material u. Organismen* 28:143-151.

Domsch, K. H., Gams, W. and Anderson, T. H. 1980. *Compedium of Soil Fungi*. Vol.1, London: Academic. pp:859.

Elad, Y. 2000. Biological control of foliar pathogens by means of *Trichoderma* harzianum and potential modes of action. Crop Protection 19:709-714.

Elad, Y. and Chet, I. 1983. Improved selective medium for isolation of *Trichoderma* or *Fusarium* spp. *Phytoparasitica* 11:55-58.

Elad, Y. and Kapat, A. 1999. The role of *Trichoderma harzianum* protease in the biocontrol of *Botrytis cinerea*. *European Journal of Plant Pathology* 105:177-189.

Elad, Y. Zimand, G., Zaqs, Y., Zuriel, S. and Chet, I. 1993. Use of *Trichoderma harzianum* in combination or alteration with fungicides to control cucumber grey mould (*Botrytis cinerea*) under commercial greenhouse conditions. *Plant Pathology* 42:324-332.

Elad, Y., Chet, I. and Henis, Y. 1981. A selective medium for improving quantitative isolation of *Trichoderma* spp. from soil. *Phytoparasitica* 9:59-67.



Elad, Y., Chet, I. and Katan, J. 1980. *Trichoderma harzianum*: A biocontrol agent effective against *Sclerotium rolfsii* and *Rhizoctonia solani*. *Phytopathology* 70:119-121.

Elad, Y., Chet, I., Boyle, P. and Henis, Y. 1983. Parasitism of *Trichoderma* spp. on *Rhizoctonia solani* and *Sclerotium rolfsii* – Scanning electron microscopy and fluorescence microscopy. *Phytopathology* 73:85-88.

Etebarian, H. R., Scott, E. S. and Wicks, T. J. 2000. *Trichoderma harzianum* T39 and *T. virens* DAR 74290 as potential biological control agents for *Phytophthora erythroseptica*. *European Journal of Plant Pathology* 106:329-337.

Garret, S. D. 1956. *Biology of Root-infecting Fungi*. Cambridge Universiti Press, Cambridge, UK. pp:293.

Ghisalberti, E. L. and Rowland, C. Y. 1993. Antifungal metabolites from *Trichoderma harzianum. Journal of Natural Products* 56(10): 1799-1804.

Gurmit, S. 1990. *Ganoderma* – The scourge of oil palms in the costal areas. In *Proceedings of the Ganoderma Workshop*, 11 September 1990. ed. Ariffin, D. and Jalani, S. pp: 113-131. Palm Oil Research Institute of Malaysia, Bangi, Selangor, Malaysia.

Gurmit, S. 1991. *Ganoderma*: The scourge of oil palm in coastal areas. *The Planter* 67(786).

Hadar, Y., Chet, I. and Henis, Y. 1979. Biological control of *Rhizoctonia solani* damping-off with wheat bran culture of *Trichoderma harzianum*. *Phytopathology* 69:64-68.

Hanson, L. E. 2000. Plant pathology and nematology: Reduction of *Verticillium* wilt symptoms in cotton following seed treatment with *Trichoderma virens*. *The Journal of Cotton Science* 4:224-231.

Hardon, J. J. and Thomas, R. L. 1968. Breeding and selection of the oil palm in Malaya. *Oleagineux* 23(2):85-89.

Hartley, C. W. S. 1988. The botany of oil palm. In *The Oil Palm* (3rd edition), pp:47-94, Longman, London.

Hartley. C. W. S. 1977. *The Oil Palm* (2nd edition), Tropical Agriculture Series. Longman, London.

Hassan, Y. and Turner, P. D. 1998. The comparative importance of different oil palm tissues as infection sources for basal stem rot in replantings. *The Planter* 74(864):119-135.

Hjeljord, L. G. and A. Tronsmo. 1998. *Trichoderma* and *Gliocladium* in biological control: An overview. In *Trichoderma and Gliocladium*. Vol. 2, ed. Harman, G. E. and C. P. Kubicek, pp:131-151.



Hjeljord, L. G., Stensvand, A. and Tronsmo, A. 2000. Effect of temperature and nutrient stress on the capacity of commercial *Trichoderma* products to control *Botrytis cinerea* and *Mucor piriformis* in greenhouse strawberries. *Biological Control* 19:149-160.

Ho, C. T. and Khairuddin, H. 1997. Usefulness of soil mounding treatments in prolonging productivity of prime-age *Ganoderma* infected palms. *The Planter* 73(854):239-244.

Ho, Y. W. and Nawawi, A. 1985. *Ganoderma boninense* Pat. from basal stem rot of oil palm (*Elaeis guineensis*) in Peninsular Malaysia. *Pertanika* 8(3):425-428.

Howell, C. R. 2003. Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: The history and evolution of current concepts. *Plant Disease* 4 (1):4-10.

Ilias, G. N. M. 2000. *Trichoderma* and its efficacy as a biocontrol agent of basal stem rot of oil palm (*E. guineensis*), Ph.D. Thesis, Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

Ilias, G. N. M. and Abdullah, F. 1998. The inhibitory effect of diffusible metabolites of *Trichoderma harzianum* on two strains of *Ganoderma boninense* by agar bilayer technique. In *Proceedings of the Asia-Pacific Mycology Conference on Biodiversity and Biotechnique*, Hua Hin, Thailand.

Ilias, G. N. M. and Abdullah, F. 1999. Effect of culture filtrates of *Trichoderma* harzianum and *T. virens* against *Ganoderma boninense*. In 5th International Conference on Plant Protection in the Tropics, 15-18 March, 1999, Kuala Lumpur, Malaysia.

Jinantana, J. 1995. Evaluation of Malaysian isolates of *Trichoderma harzianum*, (Rifai) and *Gliocladium virens* (Miller, Giddens and Foster) for the biological control of *sclerotium* foot rot of chili. Ph.D. Thesis, Universiti Putra Malaysia, Malaysia.

Jinantana, J. and Sariah. M. 1995. Antagonistic effect of Malaysian isolates of *Trichoderma harzianum* and *Gliocladium virens* on *Sclerotium rolfsii*. *Pertanika Journal of Agriculture Science* 20(1):35-41.

Johnson, L. F. 1957. Effect of antibiotics on the number of bacteria and fungi isolated from soil by the dilution plate method. *Phytopathology* 47:630-631.

Jollands, P. 1983. Laboratory investigations of fungicides and biological agents to control three diseases of rubber and oil palm and their potential applications. *Tropical Pest Management* 29 (1):33-38.

Khairudin, H. 1990. Basal Stem Rot of Oil Palm: Incidence, Etiology and Control. Master of Agriculture Science Thesis. Universiti Pertanian Malaysia, Selangor, Malaysia.



Khairudin, H. 1991. Pathogenicity of three *Ganoderma* species on oil palm seedlings. *Journal of Perak Planters Association* pp:43-49.

Khairudin, H. 1993. Basal stem rot of oil palm caused by *Ganoderma boninense*: An Update. In *1993 PORIM International Palm Oil Conference* (Agriculture). September 20-25, 1993.

Khairudin, H. 1994. Influence of size and age of inoculum of infection of oil palm seedlings by *G. boninense* Pat. In 4^{th} International Conference on Plant Protection in the Tropics, March 28-31, 1994. Kuala Lumpur, Malaysia.

Khairudin, H. Lim, T. K. and Abdul Rahman, A. R. 1991. Pathogenicity of *Ganoderma boninense* Pat. on oil palm seedlings. In *1991 PORIM International Palm Oil Conference*, Malaysia.

Krauss, U. and Soberanis, W. 2001. Biocontrol of cocoa pod diseases with mycoparasite mixtures. *Biological Control* 22:149-158.

Kredics, L., Antal, Z., Manczinger, L., Szekeres, A., Kevei, F. and Nagy, E. 2003. Influence of environmental parameters on *Trichoderma* strains with biocontrol potential. *Food Technology Biotechnology* 41(1):37-42.

Kubicek, C. P. and Harman, G. E. 1998. Basic Biology, Taxanomy and Genetics In *Trichoderma and Gliocladium*. Vol 1. Taylor & Francis, London.

Latiff, A. 2000. The biology of the genus *Elaeis*. In *Advances in Oil Palm Research* Vol. 1, ed. Yusof Basiron, Jalani, B. S. and Chan, K. W. pp:19-38.

Lewis, J. A. and Lumsden, R. D. 2001. Biocontrol of damping-off of greenhousegrown crops caused by *Rhizoctonia solani* with a formulation of *Trichoderma* spp. *Crop Protection* 20:49-56.

Lewis, J. A. and Papavizas, G. C. 1983. Production of chlamydospores and conidia by *Trichoderma* spp. in liquid and solid growth media. *Soil Biology and Biochemical* 15(3):351-357.

Lewis, J. A. and Papavizas, G. C. 1984. A new approach to stimulate population proliferation of *Trichoderma* species and other potential biocontrol fungi introduced into natural soils. *Phytopathology* 74:1240-1244.

Lim, K. H., Chuah, J. H. and Ho, C. Y. 1993. Effects of soil heaping on *Ganoderma* infected oil palms. In *1993 International Palm Oil Congress* (PORIM) - Update and Vision (Agriculture), pp:447-485.

Lim, T. K. and Teh, B. K. 1990. Antagonistics *in vitro* of *Trichoderma* species against several Basidiomycetes soil borne pathogens and *Sclerotia rolfsii*. *Journal of Plant Disease and Protection* 97:33-41.

165

Lim, T. K., Hamm, R. T. and Mohamad, R. 1990. Persistency and volatile behavior of selected chemical in treated soil against three Basidiomycetes root disease pathogens. *Tropical Pest Management* 36(1):23-26.

Lo, C. T., Nelson, E. B. and Harman, G. E. 1997. Improved biocontrol efficacy of *Trichoderma harzianum* (1295-22) for foliar phases of turf diseases by use of spray applications. *Plant Disease* 81:1132-1138.

Lockwood, J. L. 1977. Fungistasis in soils. *Biological Review* 52:1-43.

Loh, C. F. 1976. Preliminary evaluation of some systemic fungicides for *Ganoderma* control and phytotoxicity to oil palm. *Malay Agriculture Journal* 32:223-230.

Lorito, M., Harman, G. E., Hayes, C. K., Broadway, R. M., Tronsmo, A., Woo, S. L. and Di Pietro, A. 1993. Chitinolytic enzymes produced by *Trichoderma harzianum*: Antifungal activity of purified endochitinase and chitobiosidase. *Molecular Plant Pathology* 83:302-307.

Lumsden, R. D., Locke, J. C., Adkins, S. T., Walter, J. F. and Ridout, C. J. 1992. Isolation and localization of the antibiotic gliotoxin produced by *Gliocladium virens* from alginate prill in soil and soilless media. *Phytopathology* 82:230-235.

Lutz, M. P., Wenger, S., Maurhofer, M., Defago, G. and Duffy, B. 2004. Signaling between bacterial and fungal biocontrol agents in a strain mixture. Federation of European Microbiological Societies (FEMS) *Microbiology Ecology* 48:447-455.

Malaysian Palm Oil Board (MPOB) Statistic. 2004. 23rd edition. (2004).

Martin, J. P. 1950. Use of acid, rose bengal and streptomycin in the plate method for estimating soil fungi. *Soil Science* 69:215-232.

Miller, J. H., Giddens, J. E. and Foster, A. A. 1957. A survey of the fungi of the forest and cultivated soils of Georgia. *Mycologia* 49:779-808.

Moncalvo, J. M. and Ryvarden, L. 1997. A nomenclatural study of the Ganodermataceae Donk. *Synopsis Fungorum* 11:1-114.

Naseby, D. C., Pascual, J. A. and Lynch, J. M. 2000. Effect of biocontrol strains of *Trichoderma* on plant growth, *Pythium ultimum* populations, soil microbial communities and soil enzyme activities. *Journal of Applied Microbiology* 88:161-169.

Navaratnam, J. S. and Chee, L. K. 1965. Root inoculation of oil palm seedlings with *Ganoderma* spp. *Plant Disease Reporter* 49 (12):1011-1031.

Navaratnam, S. J. 1964. Basal stem rot of oil palms on ex-coconut estates. *The Planter* 40:256-259.



Nawawi, A. and Ho, Y. W. 1990. Effect of temperature and pH on growth pattern of *Ganoderma boninense* from oil palm in peninsular Malaysia. *Pertanika* 13 (3):303-307.

Nieto, L. E. 1995. Incidence of oil palm stems rots in Colombia. *Palmas* 16:227-232.

Okigbo, R. N. and Ikediugwu, F. E. O. 2000. Studies on biological control of postharvest rot in yams (*Dioscorea* spp.) using *Trichoderma viride*. *Journal of Phytopathology* 148:351-355.

Ortiz, A. and Orduz, S. 2000. *In vitro* evaluation of *Trichoderma* and *Gliocladium* antagonism against the symbiotic fungus of the leaf-cutting ant *Atta cephalotes*. *Mycopathologia* 150:53-60.

Otieno, W., Termorshuizen, A., Jeger, M. and Othieno, C.O. 2003. Efficacy of soil solarization, *Trichoderma Harzianum* and coffee pulp amendment against *Armillaria* sp. *Crop Protection* 22:325-331.

Papavizas, G. C. 1985. *Trichoderma* and *Gliocladium*: Biology, ecology and potential for biology control. *Annual Review Phytopathology* pp:23-54.

Papavizas, G. C. and Lewis, J. A. 1989. Effect of *Gliocladium* and *Trichoderma* on damping-off and blight of snap beans caused by *Sclerotium rolfsii* in the greenhouse. *Plant Pathology* 38:277-286.

Papavizas, G. C. and Lumsden, R. D. 1980. Biological control of soilborne fungal propagules. Annual Review of *Phytopathology* 18:389-413.

Paulitz, I. C. 2000. Population dynamics of biocontrol agents and pathogens in soils and rhizospheres. *European Journal of Plant Pathology* 106:401-413.

Pechprome, S. and Soytong, K. 1996. In *Proceedings of the 1st International Symposium on Biopesticides*, pp:228-233.

Perello, A., Monaco, C. Simon, M. R. Sisterna, M. and Dal Bello, G. 2003. Biocontrol efficacy of *Trichoderma* isolates for tan spot of wheat in Argentina. *Crop Protection* 22:1099-1106.

Peries, O. S. and Liyange, A. De S. 1985. *Hevea* diseases of economic importance and integrated methods of control. In *International Rubber Conference*, Kuala Lumpur RRIM, pp:14.

Phillips, A. J. L. 1986. Factors affecting the parasitic activity of *Gliocladium virens* on sclerotia of *Sclerotinia sclerotiorum* and notes on its host range. *Phytopahology* 116:211-220.

PORIM. 1984. Laporan Tahunan 1984, Institusi Penyelidikan Minyak Kelapa Sawit Malaysia.



Prasad, R. D., Rangesawaran, R., Hegde, S. V. and Anuroop, C. P. 2002. Effect of soil and seed application of *Trichoderma harzianum* on pigeonpea with caused by Fusarium udum under field conditions. *Crop Protection* 21:293-297.

Purseglove, J. W. 1972. *Tropical Crops. Monocotyledons*. Longman, London. pp:607.

Ramasamy, S. 1972. Cross infectivity and decaying of *Ganoderma* spp. Unpublished Bachelor Agriculture Science Project Paper, Faculty of Agriculture, Universiti Malaya.

Reid, T. C. 2002. Use of fungicides and biological controls in the suppression of Fusarium crown and root rot of asparagus under greenhouse and growth chamber conditions. *Plant Disease* 86:493-498.

Ricard, J. L. 1981. Commercialization of a *Trichoderma* based mycofungicides some problems and solutions. *Biocontrol News and Information* 2:95-98.

Rifai, M. A. 1969. Revision of the genus *Trichoderma*. *Mycological Paper* 116:1-56.

Ristaino, J. B., Perry, K. B. and Lumsden, R. D. 1999. Effect of solarization and *Gliocladium virens* on sclerotia of *Sclerotium rolfsii*, soil microbiota and the incidence of southern blight of tomato. *Phytopathology* 81:1117-1124.

Roberts, D. P., Lohrke, S. M., Meyer, S. L. F., Buyer, J. S., Bowers, J. H., Baker, C. J., Li, W., de Souza, J. T., Lewis, J. A. and Chung, S. 2005. Biocontrol agents applied individually and in combination for suppression of soilborne disease of cucumber. *Crop Protection* 24:141-155.

Roiger, D. J. and Jeffers, S. N. 1991. Evaluation of *Trichoderma* spp. for biological control of *Phytophthora* crown rot and root rot of apple seedlings. *Phytopathology* 81(8):910-917.

Roy, G. V. D. and Thomas Jr., S. B. (1996). *Biological Control*. Chapman and Hall, An International Thomas Publishing Company.

Samuels, G. J. 1996. Centenary Review: *Trichoderma*: A review of biology and systematic of the genus. *Mycologia Research* 100 (8):923-935.

Saravanan, T., Muthusamy, M. and Marimuthu, T. 2003. Development of integrated approach to manage the fusarial wilt of banana. *Crop Protection* 22:1117-1123.

Sariah, M. 2003. The potential of biological management of basal stem rot of oil palm: Issues, challenges and constraints. *Oil Palm Bulletin* 47:1-5.

Sariah, M. and Chan, S. F. 1999. Biological seed treatment for the control of *Rhizoctonia* damping-off of coy sam. In *Symposium on Biological Controls in the Tropics*, Mardi Training Center, Serdang, Selangor, Malaysia, March 18-19, 1999.

C

Sariah, M. and Jinantana, J. 1998. Potential for biological control of *Sclerotium* foot rot of chili by *Trichoderma* spp. *Pertanika Journal of Tropical Agricultural Science* 21(1):1-10.

Sariah, M., Hussin, M. Z., Miller, R. N. G. and Holderness, M. 1994a. Pathogenicity of *Ganoderma boninense* tested by inoculation of oil palm seedlings. *Plant Pathology* 43:507-510.

Sariah, M., Zakaria, M. H. Miller, R. N. G. and M. Holderness. 1994b. Sequential development in the establishment and colonization of *Ganoderma boninense* on oil palm seedlings. In 4th International Conference on Plant Protection in the Tropics. March 28-31, 1994, Kuala Lumpur, Malaysia. pp:417-418.

Schoeman, M. W., Webber, J. F. and Dickinson, D. J. 1996. The effect of diffusible metabolites of *Trichoderma harzianum* on *in vitro* interactions between Basidiomycete isolates at two different temperatures regimes. *Mycologia Research* 100(12):1454-1458.

Shamala, S., Abdullah, F., Zainal, A. M. A. and Umi Kalsom Yusuf. 2003. The use of single and mixed strains of *Trichoderma harzianum* as biological control agents on oil palm seedlings. In 2003 International Palm Oil Congress (PIPOC)-(Agriculture), Malaysian Palm Oil Board (MPOB), pp:1029-1033.

Sharon, E., Bar-Eyal, M., Chet, I., Herrera-Estrella, A., Kleifeld, O. and Spiegel, Y. 2001. Biological control of the root-knot nematode *Meloidogyne javanica* by *Trichoderma harzianum. Phytopathology* 91:687-693.

Sharples, A. 1928. Palm Diseases in Malaysia. *Malaya Agriculture Journal* 16:313-360.

Sherwood, R. T. and Hagedorn, D. J. 1958. Determining the common root rot potential of pea fields. Bulletin 531. Agricultural Experiment, University of Wisconsin, pp:12

Shukla, A. N. and Uniyal, K. 1989. Antagonistic interactions of *Ganoderma lucidum* (lyss.) Karst. against some soil microorganisms. *Current Science* 58:265-267.

Sid Ahmed, A., Perez-Sanchez, C., Egea, C. and Candela, M. E. 1999. Evaluation of *Trichoderma harzianum* for controlling root rot caused by *Phytophthora capsici* in pepper plants. *Plant Pathology* 48:58-65.

Skidmore, A. M. and Dickson, C. H. 1976. Colony interaction and hyphal interference between *Septoria nodorum* and phyllophane fungi. *Transactions of the British Mycological Society* 66(1):57-64.

Sodsa-Art, P. and Soytong, K. 1998. In *Proceedings of the 24th Congress on Science* and *Technology of Thailand*, pp:858-859.

Soytong, K., Usuman, P., Kanokmedulkhul, S., Kanokmedulkhul, K., Kukongviriyapan, V. and Isobe, M. 1999. Integrated biological control of

 \bigcirc

Phytophthora rot of sweet orange using mycofungicides in Thailand. In 5th *International Conference on Plant Protection in the Tropics*. March 15-18, 1999. Kuala Lumpur, Malaysia, pp:329-331.

Sreenivasaprasad, S. and Manibushanrao, K. 1993. Efficacy of *Gliocladium virens* and *Trichoderma longibrachiatum* as biological control agents of groundnut root and stem rot diseases. *International Journal of Pest Management* 39:167-171.

Srinivasan, U. Staines, H. and Bruce, A. 1992. Influence of media type on antagonistic modes of *Trichoderma* spp. against wood decay Basidiomycetes. *Material u. Organismen* 27:301-321.

Thangavelu, R., Palaniswami, A. and Velazhahan, R. 2004. Mass Production of *Trichoderma harzianum* for managing fusarium wilt of banana. *Agriculture, Ecosystems and Environment* 103:259-263.

Tummakate, A. and Likhitakaraj, S. 1994. The situation of *Ganoderma* on oil palm in Thailand. In *Proceedings of the 1st International Workshop on Perennial Crop Diseases Caused by Ganoderma*. 28 Nov. – 3 Dec. 1994. ed. Holderness, M., Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

Turner, P. D. 1965. The oil palm and *Ganoderma* IV. Avoiding disease in new plantings. *The Planter* 41:331-333.

Turner, P. D. 1968. The use of surgery as a method of treating basal stem rot in oil palms. *The Planter* 44:303-308.

Turner, P. D. 1981. *Oil Palm Disease and Disorders*, pp:88-110. Oxford University Press.

Turner, P. D. and Gillbanks, R. A. 1974. *Oil Palm Cultivation and Management*. (1st edition), Kuala Lumpur, Incorporated Society of Planters, pp:672.

Turner, P. D. and Gillbanks, R. A. 2003. Field diseases and disorders of oil palm. In *Oil Palm Cultivation and Management* (2nd edition). The Incorporated Society of Planters, Kuala Lumpur, Malaysia, pp:625-727.

Turner, P. D. and. Bull, R. A. 1967. *Disease and Disorders of the Oil Palm in Malaysia*. Incorporated Society of Planters, Kuala Lumpur, pp:452-461.

Uhl, N. W. and Dransfield, J. 1987. Genera Palmarium. In *A Classification of Palms*-based on the Work of Harold E. Moore, Jr .Lawrence, Kansas, Allan Press, pp:610.

Umar, A. Kusnadi, M. and Ollagnier, M. 1971. Influence of the type of planting material and mineral nutrition on oil palm stem rot due to *Ganoderma*. *Oleagineux* 26:527-534.

Varghese, G. 1972. Soil micro-flora of plantations and natural rain forest of West Malaysia. *Mycopathologia* 48:43-61.

C

Varghese, G., Chew, P. S. and. Lim, J. K. 1975. Biology and chemically assisted biological control of *Ganoderma*. In *International Rubber Conference*, Kuala Lumpur, Malaysia. 3:278-292.

Wakefield, E. M. 1920. Diseases of the oil palm in West Africa. *Kew Bulletin* pp: 306-308.

Weindling, R. 1932. *Trichoderma lignorum* as a parasitic of other soil fungi. *Phytopathology* 22:837-845.

Wells, D. H., Durham, K. B. and Jaworski, C. A. 1972. Efficacy of *Trichoderma* harzianum as a biocontrol for *Sclerotium rolfsii*. *Phytopathology* 62:442-447.

Whitemore, T. C. 1973. The Palms of Malaya. Longman, Malaysia.

Wijesekera H. T. R., Wijesundera, R. L. C. and Rajapakse, C. N. K. 1996. Hyphal interactions between *Trichoderma viridae* and *Ganoderma boninense* Pat., the causal of coconut root and bole rot. *Journal of the National Science Sri Lanka* 24 (3):217-219.

Wolffhechel, H. and Jensen, D. F. 1992. Use of *Trichoderma harzianum* and *Gliocladium virens* for the biological control of post-emergence damping-off and root rot of cucumbers caused by *Pythium ultimum*. *Journal of Phytopathology* 136:221-230.

Worasatit, N., Sivasithamparam, K., Ghisalberti, E. L. and Rowland, C. 1994. Variation in pyrone production, lytic enzymes and control of Rhizoctonia root rot of wheat among single-spore isolates of *Trichoderma koningii*. *Mycological Research* 98(12):1357-1363.

Yonnes, H. and Flood, J. 2003. Colonization of rubber wood and oil palm blocks by monokaryons and dikaryons of *Ganoderma boninense* – Implications to infection in the field. *The Planter* 79(922):31-38.

Zaharah, J. 1992. Kajian perbandingan ke atas *Ganoderma* daripada tunggul dan pokok kelapa sawit. BSc. Thesis, Universiti Putra Malaysia, Serdang, Malaysia.

Zakaria, M. H. 1989. Some aspects of the biology and chemically assisted biocontrol of *Ganoderma* in Malaysia, Ph.D. Thesis, Universiti Putra Malaysia, Serdang, Malaysia.