



Mcaleenan, A., Ambrose, P. G., Bhavnani, S. M., Drusano, G. L., Hope, W. W., Mouton, J. W., Higgins, J. P. T., & Macgowan, A. P. (2020). Methodological features of clinical pharmacokinetic–pharmacodynamic studies of antibacterials and antifungals: a systematic review. *Journal of Antimicrobial Chemotherapy*, [dkaa005]. <https://doi.org/10.1093/jac/dkaa005>

Peer reviewed version

Link to published version (if available):  
[10.1093/jac/dkaa005](https://doi.org/10.1093/jac/dkaa005)

[Link to publication record in Explore Bristol Research](#)  
PDF-document

This is the author accepted manuscript (AAM). The final published version (version of record) is available online via Oxford University Press at <https://academic.oup.com/jac/advance-article/doi/10.1093/jac/dkaa005/5743463>. Please refer to any applicable terms of use of the publisher.

## University of Bristol - Explore Bristol Research

### General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available: <http://www.bristol.ac.uk/red/research-policy/pure/user-guides/ebr-terms/>

1 **Methodological features of clinical pharmacokinetic (PK)-pharmacodynamic (PD)**  
2 **studies of antibacterials and antifungals: a systematic review**

3

4 *Running title: Systematic review of clinical PK-PD studies*

5

6 Alexandra MCALEENAN<sup>1</sup>, Paul G AMBROSE<sup>2</sup>, Sujata M BHAVNANI<sup>2</sup>, George L DRUSANO<sup>3</sup>, William  
7 HOPE<sup>4</sup>, Johan W MOUTON<sup>5†</sup>, Julian PT HIGGINS<sup>1</sup>, Alasdair P MACGOWAN<sup>6\*</sup>

8

9 <sup>1</sup>Population Health Sciences, Bristol Medical School, University of Bristol, Canynge Hall, Bristol, BS8  
10 2PS, UK

11 <sup>2</sup>Institute of Clinical Pharmacodynamics, 242 Broadway, Schenectady, New York, 12305, USA

12 <sup>3</sup>Institute for Therapeutic Innovation, Department of Medicine, University of Florida, UF Research  
13 and Academic Center at Lake Nowa, 6550 Sanger Road, Orlando Florida 32827, USA

14 <sup>4</sup>Centre for Antimicrobial Pharmacodynamics, Institute of Translational Medicine, University of  
15 Liverpool, Liverpool, L69 4BX, UK

16 <sup>5</sup>Department of Medical Microbiology & Infectious Diseases, Erasmus Medical Centre, s-  
17 Gravendijkwal 230, 3015 CE Rotterdam, The Netherlands

18 <sup>6</sup>Bristol Centre for Antimicrobial Research & Evaluation, Infection Sciences, Pathology Science  
19 Quarter, North Bristol NHS Trust, Southmead Hospital, Westbury-on-Trym, Bristol, BS10 5NB, UK

20 † Deceased

21 \* *Corresponding author:* Email: [Alasdair.Macgowan@nbt.nhs.uk](mailto:Alasdair.Macgowan@nbt.nhs.uk). Phone: +44 (0) 117 414 6215. Fax:  
22 +44 (0) 117 414 9392.

23

24 **Synopsis**

25 **Background:** Pharmacokinetic (PK)- pharmacodynamic (PD) indices relate measures of drug  
26 exposure to antibacterial effect. Clinical PK-PD studies aim to correlate PK-PD indices with outcomes  
27 in patients. Optimisation of dosing based on pre-clinical studies mean that PK-PD relationships are  
28 difficult to establish, therefore studies need to be designed and reported carefully to validate pre  
29 clinical findings.

30 **Objectives:** To describe the methodological features of clinical antibacterial and antifungal PK-PD  
31 studies that reported the relationship between PK-PD indices and clinical or microbiological  
32 responses.

33 **Methods:** Studies published between 1980 and 2015 were identified through systematic searches.  
34 Methodological features of eligible studies were extracted.

35 **Results:** We identified 85 publications containing 97 PK-PD analyses. Most studies were small, with  
36 fewer than 100 patients. Around a quarter were performed on patients with infections due to a  
37 single specific pathogen. In approximately one third of studies, patients received concurrent  
38 antibiotics/antifungals, and in some other studies patients received other treatments that may  
39 confound the PK-PD – outcome relationship. Most studies measured antimicrobial concentrations in  
40 blood/serum, and only four measured free concentrations. Most performed some form of  
41 regression, time to event analysis or used the Hill/ $E_{max}$  equation to look at the association between  
42 PK-PD index and outcome. Target values of PK-PD indices that predict outcomes were investigated in  
43 52% of studies. Target identification was most commonly done using recursive partitioning or logistic  
44 regression.

45 **Conclusions:** Given the variability in conduct and reporting, we suggest that an agreed set of  
46 standards for the conduct and reporting should be developed.

47 **Introduction**

48 Pharmacokinetics (PK) and pharmacodynamics (PD) are central to an understanding of how best to  
49 employ anti-infective medications. PK and PD considerations ensure that patient outcomes are  
50 optimised, since they inform the choice, dose and frequency of appropriate anti-infectives, the  
51 potential use of combinations, decisions about duration of treatment, and definitions for  
52 categorising the infecting pathogen as susceptible or resistant to therapy.

53 The PK and PD properties of antimicrobials are often considered simultaneously to form PK-PD  
54 indices, for example, the ratio of peak concentration of the antimicrobial ( $C_{max}$ ) to the MIC  
55 ( $C_{max}/MIC$ ).

56 Knowledge of the PK-PD index that best predicts treatment efficacy, and the magnitude of that index  
57 that is associated with optimal outcomes, can be used to guide antimicrobial treatment. This can  
58 maximise the possibility of both successful clinical outcome and pathogen eradication, which in turn  
59 may help to prevent the development of drug resistant pathogens.

60 The study of PK-PD is an iterative process whereby pre-clinical (*in vitro* and *in vivo*) experiments,  
61 population PK models, and *in silico* simulations are used to investigate potential dosing regimens and  
62 PK-PD targets. These studies can be used to inform regimen selection for clinical studies.

63 Numerous clinical PK-PD studies of antimicrobials have been performed. They generally aim to  
64 investigate the relationship between PK-PD indices and either clinical or microbiological response (or  
65 some element of these outcomes, or both) in patients. In some cases, they have sought to  
66 determine the target magnitude of a PK-PD index to increase the probability of successful outcome  
67 during therapy. Relationships between PK-PD indices and response, and PK-PD targets for efficacy  
68 may be difficult to find in clinical PK-PD studies if pre-clinical development has been performed well.

69 Clinical PK-PD studies are required to confirm conclusions from pre-clinical studies and to provide  
70 information for future use to calibrate pre-clinical systems more closely to clinical circumstances. In  
71 addition, there are concerns that many of these clinical PK-PD studies have limitations including:  
72 small size (<100 patients), not limiting analyses to populations with similar pathogens and sites of

73 infection, not measuring free drug concentration, not being designed with a primary  
74 pharmacodynamic end point in mind, and not adjusting analyses for confounders such as source  
75 control or comorbidities. There is also uncertainty over how results should be analysed and how PK-  
76 PD index targets for efficacy should be identified. In addition, there may be a bias in the literature  
77 towards reporting positive results.

78 It is therefore timely to perform a systematic review of the methodological features of clinical PK-PD  
79 studies of antibacterials and antifungals. Here we review articles published between 1980 and 2015  
80 that reported the relationship between PK-PD indices and some form of clinical or microbiological  
81 response. In addition to summarising the methodological features of these studies, we had a  
82 secondary goal of finding examples of good practice that might be used to inform future PK-PD  
83 analyses.

## 84 **Materials and methods**

### 85 **Inclusion and exclusion criteria**

86 We sought clinical PK-PD studies that related a calculated PK-PD index to the probability of clinical or  
87 microbiological cure (or some element of either of these outcomes).

88 We included nested case-control, cohort studies and randomised controlled trials (RCTs) published  
89 since 1980 in English. We excluded conference abstracts, due to the lack of methodological detail,  
90 and case reports, due to the inability to link a PK-PD index with outcome. Participants had to have a  
91 bacterial or fungal infection, although we excluded studies on patients with mycobacterial  
92 infections. We included studies of combination therapy, and studies that pooled patients treated  
93 with different antimicrobials of the same class when performing analyses, but we excluded studies  
94 that pooled different classes of antimicrobials as PK-PD indices and targets are class-specific.

95 Eligible studies had to measure the concentration of an antimicrobial in a biological material for all  
96 patients in the analysis and use this concentration in the calculation of the PK-PD index (directly, by  
97 creating a population PK model, or by using the concentrations in conjunction with a previously

98 constructed PK model). We excluded studies that estimated PK parameters using only values of  
99 covariates in a population PK model to which the included patients did not contribute concentration  
100 measurements. We excluded studies that did not measure microbial susceptibility of an isolated  
101 causative pathogen and instead used, for example, breakpoints for susceptibility or the distribution  
102 of MICs for a particular pathogen. We excluded inhibitory/bactericidal titres in either urine or serum  
103 due to the methodological problems with these assays.

104 Eligible outcomes were any measure of treatment response or failure. We included studies that  
105 analysed statistically or described the association between PK-PD index and outcome. We excluded  
106 studies that had only development of resistance as an outcome.

107

#### 108 **Search strategy**

109 We searched MEDLINE, Embase, Web of Science and BIOSIS up to 22 July 2015 using a combination  
110 of MeSH headings and free-text search terms for PK-PD indices, antibiotics and antifungals, and  
111 treatment outcome. Full search strategies for each database are presented in the Supplementary  
112 data. In addition, we tried to find full journal articles corresponding to relevant conference abstracts  
113 that had been identified in the search, we hand-searched reference lists of included studies, and  
114 PGA, SMB, GLD, WH and JWM ratified the list of included studies and provided any missing studies  
115 that met our inclusion criteria.

116

#### 117 **Selection of studies**

118 One author (Alexandra McAleenan [AM]) sifted the titles and abstracts. At title and abstract stage  
119 conference abstracts and foreign language papers were not excluded as they were used to identify  
120 English language versions or additional studies. Potentially relevant studies were obtained and full-  
121 texts were assessed for eligibility by one author (AM).

122

123 **Data extraction**

124 We extracted data on study characteristics including population, antibiotic analysed, how PK  
125 parameters were derived, how bacterial susceptibility was determined, PK-PD indices calculated,  
126 outcomes and timing of outcome assessment, methods used to explore the relationship between  
127 PK-PD index and outcome, industry funding, and results of the study. Study characteristics were  
128 extracted from each publication without recourse to other cited references. The full list of items  
129 extracted is given the Supplementary data.

130 When a single publication included studies performed on different groups of patients, data on each  
131 cohort were extracted as a separate PK-PD analysis, providing no analysis had been performed on all  
132 patients combined. If patients received therapy with different antimicrobials and separate PK-PD  
133 analyses were performed per antimicrobial, these were also extracted as separate PK-PD analyses.  
134 One author (AM) extracted the data. In all situations, if numbers were not reported they were  
135 calculated from available data where possible.

136 ***Role of the funding source***

137 The funders had no role in data collection and analysis, decision to publish, or preparation of this  
138 manuscript.

139 **Results**

140 ***Study selection and description***

141 We identified 6096 records after deduplication. Of the 348 full-text studies assessed, we included 85  
142 publications. Ten publications included multiple analyses (studies).<sup>1-10</sup> In total, 97 PK-PD analyses  
143 were included in the review. The flow chart for study inclusion is shown in Figure 1. Features of the  
144 included analyses are presented by class in Table 1, and study level details are presented in the  
145 Supplementary data, Tables S1-S5.

146 The 97 included analyses comprised 88 analyses of antibacterials and nine analyses of antifungals.

147 The most studied class of antimicrobial were the quinolone (21 studies). For several classes of

148 antimicrobials (polymyxins, macrolides, and polyenes) only one study was identified. The  
149 antimicrobial with the most studies was vancomycin (nine studies). Five studies examined a mix of  
150 antibiotics of the same class: four on aminoglycosides (gentamicin or tobramycin [two studies],<sup>11,12</sup>  
151 gentamicin, tobramycin or amikacin [two studies]),<sup>13,14</sup> and one on  $\beta$ -lactams (ceftriaxone, cefepime  
152 or piperacillin).<sup>15</sup>

153 Overall, 38 studies (39%) reported at least some industry/commercial funding, 39 (40%) reported  
154 that they had no funding or were funded by a non-industrial source, and the funding source was not  
155 reported in 20 studies (21%). It was clear that some PK-PD analyses were performed on the same  
156 patients, although this was not investigated systematically. There was overlap in the populations  
157 used in Peloquin *et al.*,<sup>16</sup> Forrest *et al.*,<sup>17</sup> and Goss *et al.*;<sup>7</sup> in Meinel *et al.*<sup>18</sup> and Forrest *et al.*;<sup>19</sup> in  
158 Passarell *et al.*<sup>20</sup> and Bhavnani *et al.*;<sup>21</sup> and in Ambrose *et al.*<sup>22</sup> and Bhavnani *et al.*<sup>23</sup>

#### 159 ***Population characteristics, site of infection and infecting organisms***

160 Most PK-PD analyses were small, containing 59 patients on average (range 2 to 404).<sup>24,25</sup> Only 15  
161 analyses (15%) studied more than 100 patients. Only two of the included studies made any  
162 comparison between the patients included in the study and patients excluded.<sup>26,27</sup>

163 The analyses included patients with a wide range of infections. Most investigated patients with a  
164 single type of infection, but 19 studies (20%) included patients with a variety of different infections  
165 (categorised as “multiple” infections in Table 1). In general, inclusion into the studies was based  
166 solely on both having an infection (sometimes limited by pathogen) and being treated with a  
167 particular antibiotic. Some studies addressed classes of infection, such as complicated intra-  
168 abdominal infections (cIAI) or complicated skin and skin structure infections (cSSSI), which  
169 encompass several different infections. One study split their analyses by site of infection after  
170 finding in multivariable analyses that differences in outcome were due to site of infection in addition  
171 to antibiotic exposures and PK-PD indices.<sup>28</sup>

172 In 24 studies (25%), analyses were performed on patients with infections due to a particular  
173 pathogen. *Staphylococcus aureus* was the most studied pathogen (13 studies). The other studies of



174 individual pathogens investigated infections due to *Coxiella burnetii*,<sup>29</sup> *Haemophilus influenzae*,<sup>5</sup>  
175 *Pseudomonas aeruginosa* (six studies),<sup>3,30-33</sup> and *Streptococcus pneumoniae* (three studies),<sup>5,6,34</sup> PK-  
176 PD analyses of glycopeptides were most likely of all the antimicrobial classes to be performed on  
177 patients with infection due to a particular pathogen. Other studies only included patients infected  
178 with a restricted number of pathogens (for example infections with *S. pneumoniae* and/or *H.*  
179 *influenzae*<sup>35</sup> or *P. aeruginosa* and/or *Acinetobacter baumannii*),<sup>36</sup> or limited the analysis to patients  
180 infected with a single genus, (for example *Aspergillus*,<sup>1</sup> *Candida*,<sup>37,38</sup> *Haemophilus*,<sup>39</sup> or  
181 Enterobacteriaceae).<sup>21</sup> Several studies subdivided their cohort depending on infecting pathogen, but  
182 were not categorised as being monomicrobial as the subcohorts were subsequently re-combined in  
183 the analyses.<sup>20,40,41</sup> Two studies sequentially combined the subcohorts they had created based on  
184 the infecting pathogen, and found statistically significant relationships between PK-PD indices and  
185 outcomes when certain combinations of patients were pooled, but that these relationships  
186 disappeared when additional patients were added to the analyses.<sup>20,41</sup> They attributed this to the  
187 infecting pathogen becoming so heterogeneous that the relationship between PK-PD indices and  
188 outcomes was occluded.

189 We did not extract specific eligibility criteria for each study, but we observed that several studies  
190 used infection with a resistant organism as an exclusion criterion. Although it would be regarded as  
191 ethical to administer antibiotics with a high probability of successfully treating an infection, if all  
192 patients are infected with susceptible organisms, MICs (or equivalent) will be low and it is therefore  
193 likely that PK-PD indices will be high for all patients. This makes it unlikely that a relationship  
194 between PK-PD index and outcome will be observed.

195 Because microbiological and clinical cure are associated with eradication of pathogens, receipt of  
196 effective antimicrobials in addition to the antimicrobial under investigation may obscure the PK-PD –  
197 outcome relationship of interest. In 20 studies (21%), it was explicitly reported that no patients  
198 received concurrent antibiotics/antifungals or that the receipt of concomitant antibiotics/antifungals  
199 was an exclusion criterion. However, in 33 studies (34%), some patients received concurrent

200 antibiotics or antifungals, and in the remaining 44 studies (45%) it was unclear or not reported.  
201 Some studies included receipt of concurrent antimicrobials as a potential confounder in  
202 multivariable analyses (discussed below).  
203 Depending on the type of infection and on patients' comorbidities, patients could have received  
204 other treatments concurrently with the antimicrobials. Concurrent treatments may also confound  
205 the PK-PD - outcome relationship. Some studies reported that patients had the source of the  
206 infection removed by surgery. This is important for some infections, for example complicated skin  
207 and skin structure infections, complicated intra-abdominal infections, endocarditis, and  
208 bacteraemia/sepsis secondary to central line infection. In some cases it was reported that non-  
209 infectious treatment was "standardised"<sup>31</sup> or that a "standard treatment protocol"<sup>30</sup> was used.  
210 Other studies included concomitant pharmacotherapy and surgery (or probability of a surgical or  
211 radiological procedure being successful in controlling the source of infection) as potential covariates  
212 in multivariable models.  
213 Study level data are presented in Table S1.

#### 214 ***Determination of PK-PD indices***

215 Details of how studies determined PK-PD indices, summarised by antimicrobial class, are presented  
216 in Table 2. We included studies only if they measured antimicrobial concentration in at least one  
217 matrix and these concentrations enabled an estimate of the pharmacokinetics. Nearly all studies (95  
218 [98%]) measured antimicrobial concentration in blood, serum or plasma. In the remaining two  
219 studies the biological source was not reported.<sup>22,40</sup> Antimicrobial concentration was also measured  
220 in sputum (three studies);<sup>33,42,43</sup> bronchoalveolar lavage<sup>44</sup> (using it to calculate PK-PD indices in  
221 epithelial lining fluid); middle ear effusion;<sup>35</sup> urine (two studies<sup>32,33</sup>- which developed population PK  
222 models to characterise the serum concentration and urinary excretion data simultaneously); and  
223 sinus aspirate (two studies<sup>34,45</sup>- in which the plasma and sinus aspirate concentrations were  
224 modelled simultaneously).

225 Most antimicrobials are protein bound to some degree, but only the unbound (free) fraction is  
226 considered active, since antimicrobial binding to serum albumin and other proteins may affect  
227 penetration and the ability of a drug to bind at its site of activity. It has been recommended that all  
228 PK-PD indices should be calculated relative to the unbound fraction of the drug.<sup>46</sup> Four studies  
229 measured free concentrations, using ultrafiltration to isolate unbound drug.<sup>47-50</sup> A further 23 studies  
230 adjusted for protein binding by proportionate scaling of the concentrations measured. However,  
231 proportionate scaling assumes that the fraction of antimicrobial that is protein bound is constant  
232 (i.e. not patient specific). However, changes in a patient's condition may change protein and albumin  
233 levels and alter the free fraction of the drug. One study compared total and free concentrations of  
234 vancomycin.<sup>48</sup> It found that the correlation between free and total concentrations was adequate at  
235 the population level, but the free/total ratio varied between different samples.

236 A population PK model was used in 62 (64%) of the studies. We categorised the population PK  
237 models according to the population from which they were constructed. Eight studies used a  
238 population PK model constructed only from the participants in the analysis. Twenty-two studies used  
239 a population PK model that was constructed from patients from the same cohort or randomized  
240 controlled trial (i.e. treated for the same infection with the same antimicrobial over the same time  
241 period), but not all patients were in the analysis associating the PK-PD index with the outcome  
242 (because, for example, some patients did not have pathogen susceptibility data or outcome data).  
243 Eleven studies used a population PK model that was constructed from patients in the analysis but  
244 also patients with different infections and/or healthy volunteers. Two studies used population PK  
245 models constructed from different populations. Eighteen studies referenced the population PK  
246 model, but the population used to construct the PK model was not described adequately to  
247 categorise it. One study did not describe the population PK model, and it was not referenced.<sup>51</sup>  
248 Some studies used the population PK model for some analyses, but not for all. For example,  
249 Okusanya *et al.*<sup>33</sup> constructed a population PK model for serum and urinary excretion data, and used

250 it to calculate serum AUC, but the sputum AUC on days one and 14 was calculated using the linear  
251 trapezoidal rule. These studies were classified as using a population PK model.

252 Study level data are presented in Table S2.

253 Approximately half of studies reported the strategy adopted if more than one strain/pathogen was  
254 identified. Strategies included excluding patients with polymicrobial infections, calculating PK-PD  
255 indices on a per pathogen basis, using the least susceptible pathogen for calculation of the PK-PD  
256 index, and choosing the pathogen deemed most likely to be responsible for the infection. In many  
257 cases the source of the pathogen and the exact timing of collection were not reported, but we  
258 presumed that pathogens were isolated prior to therapy and from an appropriate source.

259 Study level data are presented in Table S3.

260 Further detail regarding determination of antimicrobial drug concentrations in biological matrices  
261 and determination of antimicrobial drug susceptibility is given in the Supplementary data.

262

263

#### 264 ***PK-PD indices determined***

265 The types of PK-PD indices calculated are presented in Table 3. The PK-PD index that was most  
266 commonly calculated was a ratio of AUC to some measure of microbial susceptibility, but this varied  
267 by antimicrobial class. Of the antimicrobial classes with more than ten studies identified, a ratio of  
268  $C_{max}$  to some measure of microbial susceptibility was the most common PK-PD index calculated for  
269 aminoglycosides, a ratio of AUC to some measure of microbial susceptibility was the most common  
270 PK-PD index calculated for glycopeptides and quinolones, and time that the concentration exceeded  
271 ( $T >$ ) some measure of microbial susceptibility was the most common PK-PD index calculated for  $\beta$ -  
272 lactams. Just under half of the identified studies ( $n=45$  [46%]) calculated more than one index. We  
273 did not observe any particular trend over time in the choice of PK-PD index calculated for each class  
274 of antimicrobial (data not shown). A number of studies investigated other PK-PD indices. The other  
275 PK-PD indices calculated included a ratio of pre-dialysis concentration to MIC,<sup>11</sup> ratio of average

276 concentration to MIC (two studies),<sup>13,25</sup> ratio of concentration at steady state to MIC,<sup>49</sup> intensity  
277 index (AUC above MIC) (two studies),<sup>14,15</sup> and ratio of concentration to MIC or to MBC (three  
278 studies).<sup>24,29,39</sup>

279 Study level data are presented in Table S3.

#### 280 **Outcomes**

281 The outcomes assessed are presented in Table 4. The two most common outcomes were clinical  
282 response (n=48 [49%]) and bacteriological response (n=41 [42%]). We did not examine in detail how  
283 these terms were defined in the individual studies. However, we note the possibility of some  
284 misclassification: for example, in one paper clinical cure was defined as a composite of clinical and  
285 bacteriological cure.<sup>52</sup> Some studies examined specific outcomes that could contribute towards  
286 “clinical cure”.

287 Clinical PK-PD studies of glycopeptides were most likely to assess the relationship of a PK-PD index  
288 with undesirable outcomes, such as treatment failure or mortality.

289 In 12 studies, at least one outcome was time to some event, such as time to clinical success or time  
290 to bacterial eradication. Of the 91 studies that assessed non time-to-event outcomes, 64 studies  
291 (70%) reported clearly the timing of the outcome assessment. Of the studies which assessed  
292 outcomes at fixed timepoints, just over one third (33 studies) of studies reported assessing at least  
293 one outcome at the end of therapy. Other outcome assessment timing included during therapy, at  
294 test-of-cure (at various time points), and at the end of the study. Some outcomes had timing as part  
295 of the outcome, for example 30-day mortality, or recurrence within 60 days of discontinuation.

296 Study level data are presented in Table S3.

#### 297 **Statistical analyses**

298 The statistical approaches used to link PK-PD index with outcome are presented in Table 5, with  
299 study level data in Table S4. Only three studies (from two publications) reported performing a  
300 sample size calculation,<sup>10,53</sup> although in only one of these publications (two studies)<sup>10</sup> was the

301 calculation based on estimating the association of PK-PD index and outcome (in the other it was  
302 based on the estimated association between MIC and treatment outcome).<sup>53</sup> In addition, in two  
303 studies sample size was a consideration in whether to combine data across cohorts separated by  
304 pathogen, though specific details were not reported.<sup>20,41</sup>

305 Sixty-one studies (63%) performed some form of regression, time to event analysis or used the  
306 Hill/ $E_{max}$  equation to look at the association between PK-PD index and outcome. The most common  
307 form of analysis was logistic regression, performed in 51 studies. We categorised analyses according  
308 to whether linear or non-linear relationships were assumed, and on whether the models were  
309 adjusted for additional variables. Most of the analyses were based on linear models (n=58; we  
310 considered models in which one of the variables was log transformed to be linear), with a non-linear  
311 model being assumed in only seven studies.

312 Linear models may not adequately characterize the relationship between the PK-PD index and the  
313 outcome. For example, Rayner *et al.* examined a variety of linear and non-linear regression models  
314 and found that  $E_{max}$ -type functions (modified Hill equations) best described the relationship between  
315 AUC/MIC and the probability of microbiological eradication and clinical cure at particular sites of  
316 infection.<sup>28</sup> Step functions, in contrast, better described the association between %T>MIC and  
317 probability of microbiological eradication and clinical cure at particular sites of infection. In contrast,  
318 one study explored the possibility of nonlinearity using splines and quadratic functions, but found  
319 that the relationship between the average concentration/MIC ratio versus response was best  
320 described using a linear term.<sup>25</sup> Several other studies explored the relationship between PK-PD  
321 index and outcome using a number of different types of analysis.<sup>17,19</sup> One the papers we included  
322 focussed on whether frequentist or Bayesian logistic regression may impact on the magnitude of the  
323 treatment effect.<sup>22</sup>

324 Many factors may influence the outcome of an infection. For example, general host factors such as  
325 inflammatory response, antibody response, phagocyte function, underlying disease; site of infection  
326 factors such as abscess drainage, sequestrum formation, presence of a foreign body such as an

327 intravascular catheter or a prosthetic cardiac valve; and pathogen factors including toxin production  
328 or other virulence factors, evasion of the host inflammatory response or of antibiotic by entry into a  
329 protected site e.g. an intracellular location. However, fewer than half of the studies (42 studies)  
330 performed or planned a multivariable analysis. The covariates included varied among studies. Thirty-  
331 nine studies examined demographic factors such as age or sex as potential covariates. Thirty-four  
332 studies examined clinical and physiological patient characteristics such as underlying disease and  
333 comorbidities. Thirty studies examined potential covariates related to infection, such as infecting  
334 organism and site/type of infection. Twenty-seven studies examined covariates relating to anti-  
335 infection treatment, such as receipt of concurrent antibiotics and surgery/removal of the infection  
336 focus.

337 Of the 73 studies in which analyses were not performed on a per-pathogen basis, 11 studies  
338 examined the infecting organism as a potential covariate (15%). Of the 33 studies where at least  
339 some proportion of the population received concurrent antimicrobials, 11 studies considered this as  
340 a potential covariate (33%). It was unclear in how many studies surgery was performed, but only  
341 four studies considered surgery, probability of the primary surgical or radiological procedure being  
342 successful in controlling the source of infection, or removal of eradicable focus as a potential  
343 covariate.

344 Eighteen studies did not report planning or performing any form of statistical analysis to examine  
345 the association between PK-PD index and outcome.<sup>8,34,35,42,43,45,49-51,54-62</sup> In these studies there was  
346 either some form of descriptive analysis, for example in one study the AUC/MIC values of strains  
347 that were eradicated and strains that were not eradicated were described,<sup>51</sup> and/or PK-PD indices  
348 and outcomes were tabulated (often per patient or per pathogen).

#### 349 **Study results**

350 Study results are presented in Table S5. Of the studies that performed some sort of statistical  
351 analysis, 56 studies (71%) reported a statistically significant relationship between a PK-PD index

352 (either analysed as a continuous variable or categorised) and at least one of the outcomes  
353 investigated.

354 Over half the studies (n=51, 52%) looked at target values of the PK-PD indices that might be used to  
355 link to clinical outcome. Target values in each study are reported in Table S5. In 20 studies (21%) a  
356 pre-specified target was examined, in 34 studies (35%) a *de novo* target was identified, and three  
357 studies investigated both. The two most common techniques to identify *de novo* targets were  
358 variations on recursive partitioning and logistic regression. Twenty-six studies used a form of  
359 recursive partitioning. The most common method was classification and regression tree (CART)  
360 analysis, which derives optimal 'splits' of the continuous exposure variables in an iterative process.  
361 The split point determined for the PK-PD index is interpreted as a 'target' or 'breakpoint'. When  
362 logistic regression was implemented, it was used to identify the PK-PD value that corresponded to  
363 80% or 90% or 95% probability of cure (four studies). Further techniques for identifying *de novo*  
364 targets included identifying the AUC above dynamic response concentration that achieved  
365 eradication in four days or less based on cohort of patients observed,<sup>8</sup> visual inspection,<sup>63</sup> and  
366 receiver operating characteristic (ROC) curve analysis.<sup>64</sup> In three studies the method used to identify  
367 targets was unclear or not reported.<sup>18,42,65</sup> Two studies identified targets using two different  
368 techniques (CART and logistic regression).<sup>12,66</sup>

369 Many of the studies identified multiple targets by identifying targets for multiple PK-PD indices for  
370 multiple outcomes. In some cases, multiple targets were identified for the same PK-PD index for the  
371 same outcome, resulting in the population being divided into three or more. Some studies reported  
372 infection site-specific or organism-specific targets.

373 Only a few studies reported uncertainty around the PK-PD targets identified, for example using 95%  
374 confidence intervals around the identified target,<sup>2</sup> or 95% confidence intervals around the  
375 probability of being above or below the target.<sup>67</sup> Several studies analysing time-to-event outcomes  
376 reported measures of variability.<sup>28,40,68</sup>



377 Among studies reporting probabilities of a successful outcome above or below a target (or  
378 information sufficient to calculate this probability), the probability of a successful outcome among  
379 individuals with PK-PD index values exceeding the identified target varied. The lowest probability  
380 above a recursive partitioning identified target was 39%, and the highest was 100%. The probability  
381 of a successful outcome above the target was greater than 90% for 36 of the targets identified by  
382 recursive partitioning. In some studies, the probability of the outcome was lower above the CART  
383 identified target than below it. For example, Lodise *et al.* found a probability of a favourable  
384 outcome (not failure, where failure was defined as 30-day mortality, microbiological failure or  
385 recurrence of infection) to be 57% above the target of  $C_{\min 0-24h}/MIC$  (determined by broth  
386 microdilution)  $\geq 14.9$  and 71% below the target.<sup>69</sup> This may have arisen due to increasing  
387 administration of antimicrobial or change in PK parameters in the sickest patients with the poorest  
388 prognosis. These recursive partitioning-based probabilities among individuals above and below an  
389 identified target are averages across values of the PK-PD index. In contrast, logistic regression  
390 estimates the probability of an outcome for each specific value of the index. For instance, Kashuba *et*  
391 *al.* used both CART and logistic regression, finding a target of  $C_{\max}/MIC = 4.7$  using CART, with  
392 success rate of 89% above this threshold. In logistic regression, a  $C_{\max}/MIC$  of 4.7 corresponded to a  
393 68% probability of response while a  $C_{\max}/MIC$  of 23.6 (the highest seen) corresponded to a 99%  
394 probability of response: the value of 89% is the average probability across this range.<sup>12</sup>  
395 Graphical representations of the relationship between PK-PD index and probability of outcome  
396 facilitate judgements by clinicians and policy makers about reasonable targets. Several studies  
397 provided graphs. For example, several studies plotted the probability of success versus a PK-PD index  
398 based on the results of logistic regression, with confidence intervals around the regression  
399 line.<sup>5,23,25,27</sup> Twenty-two studies reported relative risks or odds ratios for outcome, either above and  
400 below a pre-specified target or with increasing levels of PK-PD index.

401 As mentioned above, in 20 studies (21%) a pre-specified target was examined, for example by  
402 seeking to confirm targets identified in another cohort<sup>8</sup> or another outcome.<sup>70</sup> Independent  
403 confirmation of targets is important if targets are going to be used to guide therapy.

#### 404 **Discussion**

405 In a systematic review of clinical PK-PD studies of antimicrobials performed between 1980 and 2015  
406 we found wide variability in conduct and reporting. Many of the concerns that have been voiced  
407 about these studies, such as small size (<100 patients), not limiting analyses to populations with  
408 similar pathogens and sites of infection, not measuring free drug concentration, and not adjusting  
409 analyses for the effect of surgery, appear to be justified. For many of the items we tried to examine,  
410 there were studies which did not report the item or provided descriptions that were unclear.

411 We believe this is the first systematic review of the methods employed in clinical PK-PD studies. We  
412 searched comprehensively for studies reported over a period of over 35 years and examined many  
413 features of their conduct and reporting. Our review has limitations, however. We excluded studies  
414 that did not measure the concentration of an antimicrobial in a biological material for all patients in  
415 the analysis, or which did not use this concentration in the calculation of the PK-PD index, and  
416 therefore we have only reviewed a subset of the clinical PK-PD literature. We did not extract  
417 detailed information on the PK methods used, as we did not aim to assess the PK aspects of the  
418 studies, and we have assumed that these were performed appropriately. We did not extract detailed  
419 information on the definitions of outcomes used in all the papers. Our search did not contain terms  
420 for disease-specific outcomes (for example, improvement in forced expiratory volume in one  
421 second). We therefore may have missed studies with these outcomes if they were not indexed  
422 under the 'Treatment outcome' MeSH heading. Finally, due to resource constraints we excluded  
423 foreign language studies, and sifting of titles and abstracts and extracting the data was performed by  
424 one reviewer. In addition, limiting the review to published, peer reviewed studies introduces the

425 possibility of bias due to non-reporting, where studies with undesirable results are less likely to be  
426 written up by researchers and/or published.

427 Future studies of associations between a PK-PD index and a clinical or microbial response may  
428 struggle to identify informative relationships because optimum dosing regimens can often be  
429 derived using pre-clinical data. However, when they are undertaken, they should be designed and  
430 reported carefully so that findings can be attributed to genuine relationships (or lack of a  
431 relationship) rather than to poor conduct, and so that the pre-defined PK-PD indices can be validated  
432 through high clinical success rates. In Table 6 we offer some recommendations for future clinical PK-  
433 PD studies, informed by our deliberations over what we observed while undertaking the review. We  
434 also identified a need for improved reporting of methodology. Studies might consider reporting all  
435 the items that we aimed to extract. In addition, we suggest that studies should compare features  
436 and outcomes of patients included in the PK-PD analysis (because there are data for PK parameters  
437 and MICs for pathogens) and other eligible patients (same infection, same pathogen, same antibiotic  
438 but for some reason do not have PK data or MICs of pathogens) to ensure that there are no material  
439 differences; and that PK-PD studies should plot PK-PD index versus probability of outcome or  
440 amount of improvement so that the exposure-response relationship is clear. Such plots, possibly  
441 with exposures divided into quantiles (an option when regression methods are used) can also allow  
442 individuals to determine their own targets.

443 The results of clinical PK-PD – response studies should be considered in conjunction with results  
444 from pre-clinical PK-PD studies. Obtaining a response rate that correlates with the expectation  
445 operation over the probability of target attainment relationship is powerful evidence.

446

447 **Acknowledgements:**

448 Preliminary findings from this systematic review have been incorporated into a white paper by  
449 COMBACTE's STAT-Net consortium.<sup>71</sup>

450 **Funding:** This research was supported by the Innovative Medicines Initiative Joint Undertaking under  
451 grant agreement no. 115523 (Combatting Bacterial Resistance in Europe projects [COMBACTE]),  
452 resources of which are composed of financial contribution from the European Union's 7th  
453 Framework Programme (FP7/2007±2013) and the European Federation of Pharmaceutical Industries  
454 and Associations (EFPIA) companies' in kind contribution.

455 **Transparency declarations:** Paul G Ambrose and Sujata M Bhavnani are President and Executive Vice  
456 President, respectively, of the Institute for Clinical Pharmacodynamics (ICPD). The ICPD has received  
457 funding from various commercial entities for research and work performed on their behalf, including  
458 A&G Pharmaceutical, Inc., Achaogen Inc., Agility Clinical, AiCuris GmbH, Amplyx, Arixa  
459 Pharmaceuticals, Arsanis Inc., Basilea Pharmaceutica, Boston Pharmaceuticals, B. Braun Medical Inc.,  
460 Cellceutix Corporation, Cembra Pharmaceuticals, Cidara Therapeutics Inc., Contrafect Corporation,  
461 Corcept Therapeutics, Inc., Debiopharm International SA, Discuva Limited, Durata Therapeutics,  
462 Emerald Lake Technologies LLC, Enhanced Pharmacodynamics, Entasis Therapeutics, Genentech,  
463 Geom Therapeutics, Inc., GlaxoSmithKline, Hoffmann-La Roche, Horizon, Indalo Therapeutics,  
464 Insmad Inc., Iterum Therapeutics Limited, Kalyra Pharmaceuticals, KBP BioSciences Co., Ltd., Kedrion  
465 Biopharma Inc., Matinas, The Medicines Company, Meiji Seika Pharma Co., Ltd., Melinta  
466 Therapeutics, Menarini Ricerche S.p.A, Merck Sharpe & Dohme, Nabriva Therapeutics, Naeja RGM  
467 Pharmaceuticals Inc., Nexcida Therapeutics, Inc., Northern Antibiotics, Nosopharm SAS, Novartis  
468 International, NuCana Biomed, Paratek Pharmaceuticals, Pernix Therapeutics, Polyphor Ltd., Profil  
469 Institute for Clinical Research, Inc., Prothena Corporation, PTC Therapeutics, Raptor Pharmaceuticals  
470 Corp., Rempex Pharmaceuticals, Inc., Roche TCRC, ScPharmaceuticals, Scynexis, Shionogi, Inc.,  
471 Sofinnova Ventures Inc., Spero Therapeutics, Synteract, Targanta Therapeutics Corporation, TauRX  
472 Ophan, Theravance Biopharma Pharmaceutica, Tetrphase Pharmaceuticals, Toyama, Turing  
473 Pharmaceuticals, University of Florida, University of Southern California, VenatoRx, Vical, Wockhardt  
474 Ltd., Zavante Therapeutics, Zogenix, Inc., 3-V Biosciences. In addition, Paul G Ambrose and Sujata M  
475 Bhavnani have a patent for Method for Improving Drug Treatments in Mammals (Patent No. US

476 9,345,717 B2). William Hope has received grants from Spero Therapeutics, F2G, AiCuris, Astellas  
477 Pharma, Matinas Biosciences, Antabio, Amplyx, Allecra, Bugworks, NAEJA-RGM, AMR Centre and  
478 Pfizer; and personal fees from Spero Therapeutics, F2G, Astellas Pharma, Amplyx, NAEJA-RGM and  
479 AMR Centre. Johan W Mouton has received grants from Aicuris, Gilead, Wockhardt, VenatorX,  
480 Helperby, Eumedica, Roche, Polyphor, Nordic Pharma and Cidara. Alasdair MacGowan received  
481 grants from the National Institute for Health Research, Achaogen Inc, AiCuris GmbH, F2G,  
482 Infectopharm GmbH, Merck Inc, Paratek, VenatoRx, Wockhardt Ltd and via IMI funding AiCuris,  
483 AstraZeneca, The Medicines Company, Evotech and Nosopharm during the conduct of this study. All  
484 other authors: none to declare.

485 **Contributions of authors:** Alexandra McAleenan designed and performed the literature searches,  
486 selected the studies, extracted the data and wrote the first draft of the manuscript. Paul G Ambrose,  
487 Sujata M Bhavnani, George L Drusano, William Hope and Johan W Mouton examined the list of  
488 included studies and critically commented on the manuscript. Julian PT Higgins gave methodological  
489 guidance and edited the manuscript. Alasdair P MacGowan conceived the study and edited the  
490 manuscript. All authors have read and approved the final manuscript.

491

492 **References**

- 493 1. Liu P, Mould DR. Population pharmacokinetic-pharmacodynamic analysis of voriconazole  
494 and anidulafungin in adult patients with invasive aspergillosis. *Antimicrob Agents Chemother* 2014;  
495 **58**: 4727-36.
- 496 2. Zhang J, Zhang Y, Shi Y, *et al.* Population pharmacokinetic and pharmacodynamic modeling  
497 of norvancomycin. *Eur J Clin Microbiol Infect Dis* 2008; **27**: 275-84.
- 498 3. Burkhardt O, Lehmann C, Madabushi R, *et al.* Once-daily tobramycin in cystic fibrosis: Better  
499 for clinical outcome than thrice-daily tobramycin but more resistance development? *J Antimicrob*  
500 *Chemother* 2006; **58**: 822-9.
- 501 4. Sato R, Tanigawara Y, Kaku M, *et al.* Pharmacokinetic-pharmacodynamic relationship of  
502 arbekacin for treatment of patients infected with methicillin-resistant *Staphylococcus aureus*.  
503 *Antimicrob Agents Chemother* 2006; **50**: 3763-9.
- 504 5. Shi J, Pfister M, Jenkins SG, *et al.* Pharmacodynamic analysis of the microbiological efficacy  
505 of telithromycin in patients with community-acquired pneumonia. *Clin Pharmacokinet* 2005; **44**:  
506 317-29.
- 507 6. Van Wart S, Phillips L, Ludwig EA, *et al.* Population pharmacokinetics and pharmacodynamics  
508 of garenoxacin in patients with community-acquired respiratory tract infections. *Antimicrob Agents*  
509 *Chemother* 2004; **48**: 4766-77.
- 510 7. Goss TF, Forrest A, Nix DE, *et al.* Mathematical examination of dual individualization  
511 principles (II): The rate of bacterial eradication at the same area under the inhibitory curve is more  
512 rapid for ciprofloxacin than for cefmenoxime. *Ann Pharmacother* 1994; **28**: 863-8.
- 513 8. Schentag JJ, Smith IL, Swanson DJ. Role for dual individualization with cefmenoxime. *Am J*  
514 *Med* 1984; **77**: 43-50.
- 515 9. Liu P. Population pharmacokinetic-pharmacodynamic analysis of anidulafungin in adult  
516 patients with fungal infections. *Antimicrob Agents Chemother* 2013; **57**: 466-74.
- 517 10. Pajot O, Burdet C, Couffignal C, *et al.* Impact of imipenem and amikacin  
518 pharmacokinetic/pharmacodynamic parameters on microbiological outcome of Gram-negative  
519 bacilli ventilator-associated pneumonia. *J Antimicrob Chemother* 2015; **70**: 1487-94.
- 520 11. Heintz BH, Thompson IGR, Dager WE. Clinical experience with aminoglycosides in dialysis-  
521 dependent patients: Risk factors for mortality and reassessment of current dosing practices. *Ann*  
522 *Pharmacother* 2011; **45**: 1338-45.
- 523 12. Kashuba ADM, Nafziger AN, Drusano GL, *et al.* Optimizing aminoglycoside therapy for  
524 nosocomial pneumonia caused by gram-negative bacteria. *Antimicrob Agents Chemother* 1999; **43**:  
525 623-9.
- 526 13. Moore RD, Lietman PS, Smith CR. Clinical response to aminoglycoside therapy: Importance  
527 of the ratio of peak concentration to minimal inhibitory concentration. *J Infect Dis* 1987; **155**: 93-9.
- 528 14. Deziel-Evans LM, Murphy JE, Job ML. Correlation of pharmacokinetic indices with  
529 therapeutic outcome in patients receiving aminoglycosides. *Clin Pharm* 1986; **5**: 319-24.
- 530 15. Sadaba B, Azanza JR, Campanero MA, *et al.* Relationship between pharmacokinetics and  
531 pharmacodynamics of beta-lactams and outcome. *Clin Microbiol Infect* 2004; **10**: 990-8.
- 532 16. Peloquin CA, Cumbo TJ, Nix DE, *et al.* Evaluation of intravenous ciprofloxacin in patients with  
533 nosocomial lower respiratory tract infections. Impact of plasma concentrations, organism, minimum  
534 inhibitory concentration, and clinical condition on bacterial eradication. *Arch Intern Med* 1989; **149**:  
535 2269-73.
- 536 17. Forrest A, Nix DE, Ballou CH, *et al.* Pharmacodynamics of intravenous ciprofloxacin in  
537 seriously ill patients. *Antimicrob Agents Chemother* 1993; **37**: 1073-81.
- 538 18. Meinel B, Hyatt JM, Forrest A, *et al.* Pharmacokinetic/pharmacodynamic predictors of time to  
539 clinical resolution in patients with acute bacterial exacerbations of chronic bronchitis treated with a  
540 fluoroquinolone. *Int J Antimicrob Agents* 2000; **16**: 273-80.

- 541 19. Forrest A, Chodosh S, Amantea MA, *et al.* Pharmacokinetics and pharmacodynamics of oral  
542 grepafloxacin in patients with acute bacterial exacerbations of chronic bronchitis. *J Antimicrob*  
543 *Chemother* 1997; **40**: 45-57.
- 544 20. Passarell JA, Meagher AK, Liolios K, *et al.* Exposure-response analyses of tigecycline efficacy  
545 in patients with complicated intra-abdominal infections. *Antimicrob Agents Chemother* 2008; **52**:  
546 204-10.
- 547 21. Bhavnani SM, Rubino CM, Ambrose PG, *et al.* Impact of different factors on the probability  
548 of clinical response in tigecycline-treated patients with intra-abdominal infections. *Antimicrob*  
549 *Agents Chemother* 2010; **54**: 1207-12.
- 550 22. Ambrose PG, Hammel JP, Bhavnani SM, *et al.* Frequentist and Bayesian pharmacometric-  
551 based approaches to facilitate critically needed new antibiotic development: Overcoming lies, damn  
552 lies, and statistics. *Antimicrob Agents Chemother* 2012; **56**: 1466-70.
- 553 23. Bhavnani SM, Rubino CM, Hammel JP, *et al.* Pharmacological and patient-specific response  
554 determinants in patients with hospital-acquired pneumonia treated with tigecycline. *Antimicrob*  
555 *Agents Chemother* 2012; **56**: 1065-72.
- 556 24. Tascini C, Bongiorno MG, Doria R, *et al.* Linezolid for endocarditis: a case series of 14 patients.  
557 *J Antimicrob Chemother* 2011; **66**: 679-82.
- 558 25. Troke PF, Hockey HP, Hope WW. Observational study of the clinical efficacy of voriconazole  
559 and its relationship to plasma concentrations in patients. *Antimicrob Agents Chemother* 2011; **55**:  
560 4782-8.
- 561 26. Preston SL, Drusano GL, Berman AL, *et al.* Pharmacodynamics of levofloxacin: A new  
562 paradigm for early clinical trials. *JAMA* 1998; **279**: 125-9.
- 563 27. Kimko H, Xu X, Nandy P, *et al.* Pharmacodynamic profiling of ceftobiprole for treatment of  
564 complicated skin and skin structure infections. *Antimicrob Agents Chemother* 2009; **53**: 3371-4.
- 565 28. Rayner CR, Forrest A, Meagher AK, *et al.* Clinical Pharmacodynamics of Linezolid in Seriously  
566 Ill Patients Treated in a Compassionate Use Programme. *Clin Pharmacokinet* 2003; **42**: 1411-23.
- 567 29. Rolain JM, Boulou A, Mallet MN, *et al.* Correlation between ratio of serum doxycycline  
568 concentration to MIC and rapid decline of antibody levels during treatment of Q fever endocarditis.  
569 *Antimicrob Agents Chemother* 2005; **49**: 2673-6.
- 570 30. Munzenberger PJ, Hsu JMC, Holliday SJ. Relationship of ceftazidime pharmacokinetic indices  
571 with therapeutic outcome in patients with cystic fibrosis. *Pediatr Infect Dis J* 1993; **12**: 997-1001.
- 572 31. Mouton JW, Jacobs N, Tiddens H, *et al.* Pharmacodynamics of tobramycin in patients with  
573 cystic fibrosis. *Diagn Microbiol Infect Dis* 2005; **52**: 123-7.
- 574 32. Okusanya OO, Bhavnani SM, Hammel JP, *et al.* Evaluation of the pharmacokinetics and  
575 pharmacodynamics of liposomal amikacin for inhalation in cystic fibrosis patients with chronic  
576 pseudomonal infections using data from two phase 2 clinical studies. *Antimicrob Agents Chemother*  
577 2014; **58**: 5005-15.
- 578 33. Okusanya OO, Bhavnani SM, Hammel J, *et al.* Pharmacokinetic and pharmacodynamic  
579 evaluation of liposomal amikacin for inhalation in cystic fibrosis patients with chronic pseudomonal  
580 infection. *Antimicrob Agents Chemother* 2009; **53**: 3847-54.
- 581 34. Ambrose PG, Anon JB, Owen JS, *et al.* Use of pharmacodynamic end points in the evaluation  
582 of gatifloxacin for the treatment of acute maxillary sinusitis. *Clin Infect Dis* 2004; **38**: 1513-20.
- 583 35. Sugita R. Good transfer of tebipenem into middle ear effusion conduces to the favorable  
584 clinical outcomes of tebipenem pivoxil in pediatric patients with acute otitis media. *J Infect*  
585 *Chemother* 2013; **19**: 465-71.
- 586 36. Narawadeeniamhun, Panomvana D, Pongpech P, *et al.* Pharmacodynamic target associated  
587 with clinical outcome of hospital-acquired pneumonia treatment with cefoperazone/sulbactam. *Int J*  
588 *Pharm Pharm Sci* 2012; **4**: 584-9.
- 589 37. Li CC, Sun P, Dong Y, *et al.* Population pharmacokinetics and pharmacodynamics of  
590 caspofungin in pediatric patients. *Antimicrob Agents Chemother* 2011; **55**: 2098-105.

591 38. Groll AH, Wood L, Roden M, *et al.* Safety, pharmacokinetics, and pharmacodynamics of  
592 cyclodextrin itraconazole in pediatric patients with oropharyngeal candidiasis. *Antimicrob Agents*  
593 *Chemother* 2002; **46**: 2554-63.

594 39. Tran JQ, Ballou CH, Forrest A, *et al.* Comparison of the abilities of grepafloxacin and  
595 clarithromycin to eradicate potential bacterial pathogens from the sputa of patients with chronic  
596 bronchitis: Influence of pharmacokinetic and pharmacodynamic variables. *J Antimicrob Chemother*  
597 2000; **45**: 9-17.

598 40. Rubino CM, Bhavnani SM, Forrest A, *et al.* Pharmacokinetics-pharmacodynamics of  
599 tigecycline in patients with community-acquired pneumonia. *Antimicrob Agents Chemother* 2012;  
600 **56**: 130-6.

601 41. Meagher AK, Passarelli JA, Cirincione BB, *et al.* Exposure-response analyses of tigecycline  
602 efficacy in patients with complicated skin and skin-structure infections. *Antimicrob Agents*  
603 *Chemother* 2007; **51**: 1939-45.

604 42. Cazzola M, Matera MG, Donnarumma G, *et al.* Pharmacodynamics of levofloxacin in patients  
605 with acute exacerbation of chronic bronchitis. *Chest* 2005; **128**: 2093-8.

606 43. Cazzola M, Di Perna F, Boveri B, *et al.* Interrelationship between the pharmacokinetics and  
607 pharmacodynamics of cefaclor advanced formulation in patients with acute exacerbation of chronic  
608 bronchitis. *J Chemother* 2000; **12**: 216-22.

609 44. Athanassa ZE, Markantonis SL, Fousteroi MZF, *et al.* Pharmacokinetics of inhaled  
610 colistimethate sodium (CMS) in mechanically ventilated critically ill patients. *Intensive Care Med*  
611 2012; **38**: 1779-86.

612 45. Ambrose PG, Anon JB, Bhavnani SM, *et al.* Use of pharmacodynamic endpoints for the  
613 evaluation of levofloxacin for the treatment of acute maxillary sinusitis. *Diagn Microbiol Infect Dis*  
614 2008; **61**: 13-20.

615 46. Mouton JW, Dudley MN, Cars O, *et al.* Standardization of  
616 pharmacokinetic/pharmacodynamic (PK/PD) terminology for anti-infective drugs: an update. *J*  
617 *Antimicrob Chemother* 2005; **55**: 601-7.

618 47. Ohki E, Yamagishi Y, Mikamo H. Relationship between the clinical efficacy and AUC/MIC of  
619 intravenous ciprofloxacin in Japanese patients with intraabdominal infections. *J Infect Chemother*  
620 2013; **19**: 951-5.

621 48. Ampe E, Delaere B, Hecq JD, *et al.* Implementation of a protocol for administration of  
622 vancomycin by continuous infusion: Pharmacokinetic, pharmacodynamic and toxicological aspects.  
623 *Int J Antimicrob Agents* 2013; **41**: 439-46.

624 49. Breilh D, Fleureau C, Gordien JB, *et al.* Pharmacokinetics of free ertapenem in critically ill  
625 septic patients: Intermittent versus continuous infusion. *Minerva Anesthesiol* 2011; **77**: 1058-62.

626 50. Benko R, Matuz M, Doro P, *et al.* Pharmacokinetics and pharmacodynamics of levofloxacin in  
627 critically ill patients with ventilator-associated pneumonia. *Int J Antimicrob Agents* 2007; **30**: 162-8.

628 51. Niki Y, Yoshida K, Miyashita N, *et al.* Evaluation of clinical dosage of gatifloxacin for  
629 respiratory tract infections in elderly patients based on pharmacokinetics/pharmacodynamics  
630 (PK/PD). *J Infect Chemother* 2008; **14**: 296-304.

631 52. Di Paolo A, Tascini C, Polillo M, *et al.* Population pharmacokinetics of daptomycin in patients  
632 affected by severe Gram-positive infections. *Int J Antimicrob Agents* 2013; **42**: 250-5.

633 53. Jung Y, Song KH, Cho JE, *et al.* Area under the concentration-time curve to minimum  
634 inhibitory concentration ratio as a predictor of vancomycin treatment outcome in methicillin-  
635 resistant *Staphylococcus aureus* bacteraemia. *Int J Antimicrob Agents* 2014; **43**: 179-83.

636 54. Haeseker M, Havenith T, Stolk L, *et al.* Is the standard dose of amoxicillin-clavulanic acid  
637 sufficient? *BMC Pharmacol Toxicol* 2014; **15**: 38.

638 55. Tascini C, Bongiorno MG, Di Cori A, *et al.* Cardiovascular implantable electronic device  
639 endocarditis treated with daptomycin with or without transvenous removal. *Heart Lung* 2012; **41**:  
640 e24-e30.



641 56. Duszynska W, Taccone FS, Switala M, *et al.* Continuous infusion of piperacillin/tazobactam in  
642 ventilator-associated pneumonia: A pilot study on efficacy and costs. *Int J Antimicrob Agents* 2012;  
643 **39**: 153-8.

644 57. Ohata Y, Tomita Y, Nakayama M, *et al.* Optimal treatment schedule of meropenem for adult  
645 patients with febrile neutropenia based on pharmacokinetic-pharmacodynamic analysis. *J Infect*  
646 *Chemother* 2011; **17**: 831-41.

647 58. Dong H, Wang X, Dong Y, *et al.* Clinical pharmacokinetic/pharmacodynamic profile of  
648 linezolid in severely ill Intensive Care Unit patients. *Int J Antimicrob Agents* 2011; **38**: 296-300.

649 59. Martinkova J, Pokorna P, Zahora J, *et al.* Tolerability and Outcomes of Kinetically Guided  
650 Therapy With Gentamicin in Critically Ill Neonates During the First Week of Life: An Open-Label,  
651 Prospective Study. *Clin Ther* 2010; **32**: 2400-14.

652 60. Sakka SG, Glauner AK, Bulitta JB, *et al.* Population pharmacokinetics and pharmacodynamics  
653 of continuous versus short-term infusion of imipenem-cilastatin in critically ill patients in a  
654 randomized, controlled trial. *Antimicrob Agents Chemother* 2007; **51**: 3304-10.

655 61. Pea F, Di Qual E, Cusenza A, *et al.* Pharmacokinetics and pharmacodynamics of intravenous  
656 levofloxacin in patients with early-onset ventilator-associated pneumonia. *Clin Pharmacokinet* 2003;  
657 **42**: 589-98.

658 62. Hanes SD, Wood GC, Herring V, *et al.* Intermittent and continuous ceftazidime infusion for  
659 critically ill trauma patients. *Am J Surg* 2000; **179**: 436-40.

660 63. Muto C, Liu P, Chiba K, *et al.* Pharmacokinetic-pharmacodynamic analysis of azithromycin  
661 extended release in Japanese patients with common respiratory tract infectious disease. *J*  
662 *Antimicrob Chemother* 2011; **66**: 165-74.

663 64. Zhou QT, He B, Zhang C, *et al.* Pharmacokinetics and pharmacodynamics of meropenem in  
664 elderly chinese with lower respiratory tract infections: Population pharmacokinetics analysis using  
665 nonlinear mixed-effects modelling and clinical pharmacodynamics study. *Drugs Aging* 2011; **28**: 903-  
666 12.

667 65. Tanigawara Y, Kaku M, Totsuka K, *et al.* Population pharmacokinetics and  
668 pharmacodynamics of sitafloxacin in patients with community-acquired respiratory tract infections. *J*  
669 *Infect Chemother* 2013; **19**: 858-66.

670 66. Tam VH, McKinnon PS, Akins RL, *et al.* Pharmacodynamics of cefepime in patients with  
671 Gram-negative infections. *J Antimicrob Chemother* 2002; **50**: 425-8.

672 67. Tanigawara Y, Nozawa K, Tsuda H. Optimal dose finding of garenoxacin based on population  
673 pharmacokinetics/pharmacodynamics and Monte Carlo simulation. *Eur J Clin Pharmacol* 2012; **68**:  
674 39-53.

675 68. Gawronski KM, Goff DA, Jack B, *et al.* A stewardship program's retrospective evaluation of  
676 vancomycin auc<sub>24</sub>/mic and time to microbiological clearance in patients with methicillin-resistant  
677 staphylococcus aureus bacteremia and osteomyelitis. *Clin Ther* 2013; **35**: 772-9.

678 69. Lodise TP, Drusano GL, Zasowski E, *et al.* Vancomycin exposure in patients with methicillin-  
679 resistant Staphylococcus aureus bloodstream infections: How much is enough? *Clin Infect Dis* 2014;  
680 **59**: 666-75.

681 70. Lodise TP, Preston S, Bhargava V, *et al.* Pharmacodynamics of an 800-mg dose of  
682 telithromycin in patients with community-acquired pneumonia caused by extracellular pathogens.  
683 *Diagn Microbiol Infect Dis* 2005; **52**: 45-52.

684 71. de Kraker MEA, Sommer H, de Velde F, *et al.* Optimizing the Design and Analysis of Clinical  
685 Trials for Antibacterials Against Multidrug-resistant Organisms: A White Paper From COMBACTE's  
686 STAT-Net. *Clin Infect Dis* 2018; **67**: 1922-31.

687 72. Sherwin CMT, Svahn S, Van Der Linden A, *et al.* Individualised dosing of amikacin in  
688 neonates: A pharmacokinetic/ pharmacodynamic analysis. *Eur J Clin Pharmacol* 2009; **65**: 705-13.

689 73. Tod M, Minozzi C, Beaucaire G, *et al.* Isepamicin in intensive care unit patients with  
690 nosocomial pneumonia: Population pharmacokinetic-pharmacodynamic study. *J Antimicrob*  
691 *Chemother* 1999; **44**: 99-108.

692 74. Duszynska W, Taccone FS, Hurkacz M, *et al.* Therapeutic drug monitoring of amikacin in  
693 septic patients. *Critical Care* 2013; **17**: 10.

694 75. Falcone M, Russo A, Cassetta MI, *et al.* Variability of pharmacokinetic parameters in patients  
695 receiving different dosages of daptomycin: Is therapeutic drug monitoring necessary? *J Infect*  
696 *Chemother* 2013; **19**: 732-9.

697 76. Casapao AM, Lodise TP, Davis SL, *et al.* Association between vancomycin day 1 exposure  
698 profile and outcomes among patients with methicillin-resistant *Staphylococcus aureus* infective  
699 endocarditis. *Antimicrob Agents Chemother* 2015; **59**: 2978-85.

700 77. Joo J, Yamaki J, Lou M, *et al.* Early response assessment to guide management of Methicillin-  
701 Resistant *Staphylococcus aureus* bloodstream infections with vancomycin therapy. *Clin Ther* 2013;  
702 **35**: 995-1004.

703 78. Brown J, Brown K, Forrest A. Vancomycin AUC<sub>24</sub>/MIC ratio in patients with complicated  
704 bacteremia and infective endocarditis due to methicillin-resistant *Staphylococcus aureus* and its  
705 association with attributable mortality during hospitalization. *Antimicrob Agents Chemother* 2012;  
706 **56**: 634-8.

707 79. Chong YP, Park SJ, Kim HS, *et al.* Persistent staphylococcus aureus bacteremia: A prospective  
708 analysis of risk factors, outcomes, and microbiologic and genotypic characteristics of isolates.  
709 *Medicine (Baltimore)* 2013; **92**: 98-108.

710 80. Hidayat LK, Hsu DI, Quist R, *et al.* High-dose vancomycin therapy for methicillin-resistant  
711 *Staphylococcus aureus* infections: Efficacy and toxicity. *Arch Intern Med* 2006; **166**: 2138-44.

712 81. Bhavnani SM, Passarell JA, Owen JS, *et al.* Pharmacokinetic-pharmacodynamic relationships  
713 describing the efficacy of oritavancin in patients with *Staphylococcus aureus* bacteremia. *Antimicrob*  
714 *Agents Chemother* 2006; **50**: 994-1000.

715 82. Hong Y, Shaw PJ, Nath CE, *et al.* Population pharmacokinetics of liposomal amphotericin B in  
716 pediatric patients with malignant diseases. *Antimicrob Agents Chemother* 2006; **50**: 935-42.

717 83. Rubino CM, Ambrose P, Cirincione B, *et al.* Pharmacokinetics and pharmacodynamics of  
718 gatifloxacin in children with recurrent otitis media: application of sparse sampling in clinical  
719 development. *Diagn Microbiol Infect Dis* 2007; **59**: 67-74.

720 84. Drusano GL, Preston SL, Fowler C, *et al.* Relationship between fluoroquinolone area under  
721 the curve:minimum inhibitory concentration ratio and the probability of eradication of the infecting  
722 pathogen, in patients with nosocomial pneumonia. *J Infect Dis* 2004; **189**: 1590-7.

723 85. Dowzicky M, Nadler H, Dorr MB, *et al.* Comparison of the in vitro activity of and pathogen  
724 responses to sparfloxacin with those of other agents in the treatment of respiratory tract, urinary  
725 tract, and skin and skin-structure infections. *Clin Ther* 1999; **21**: 790-805.

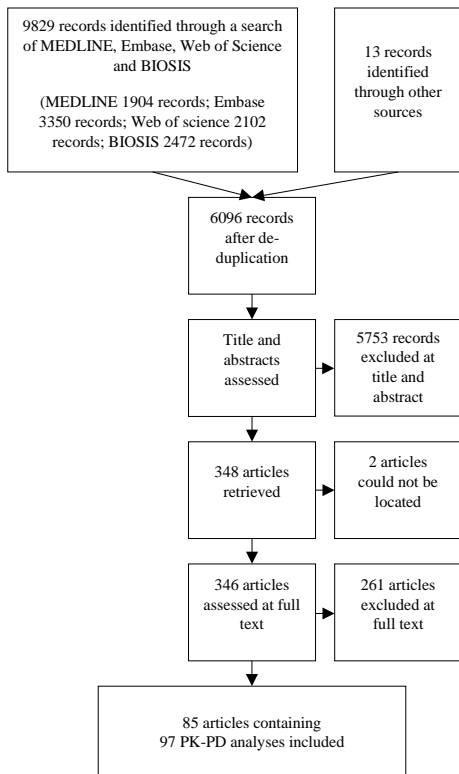
726 86. Gao L, Zhu Y, Lyu Y, *et al.* A pharmacokinetic and pharmacodynamic study on intravenous  
727 cefazedone sodium in patients with community-acquired pneumonia. *Chin Med J* 2015; **128**: 1160-4.

728 87. Du XL, Li CH, Kuti JL, *et al.* Population pharmacokinetics and pharmacodynamics of  
729 meropenem in pediatric patients. *J Clin Pharmacol* 2006; **46**: 69-75.

730 88. Royston P, Altman DG. Regression Using Fractional Polynomials of Continuous Covariates -  
731 Parsimonious Parametric Modeling. *J R Stat Soc C-Appl* 1994; **43**: 429-67.

733 **Figures and Tables**

734 *Figure 1: Flow chart for study inclusion. Reasons for exclusion at full text are given in the Supplementary data.*



735

Table 1: Features of the included clinical PK-PD studies, by antimicrobial class.

Antimicrobial class	Number of studies	Industry funded, n (%)	Specific antibiotic (number of studies)	Average number of people (range)	Number of studies with ≥100 participants, number of studies (%)	Infections (number of studies)	Single pathogen, number of studies (%)	Concomitant antibiotics or antifungals, number of studies (%)*
Aminoglycoside <sup>3,4,10-14,31-33,59,72-74</sup>	16	2 (13%)	Amikacin (5), Arbekacin (2), Gentamycin (1), Isepamicin (1), Tobramycin (3), Multiple aminoglycosides (4)	52 (13-236)	1 (6%)	Bacteraemia (1), Infections in cystic fibrosis (5), Multiple (4), Nosocomial pneumonia (including intensive care unit acquired and ventilator associated pneumonia) (3), Sepsis (3)	7 (44%)	11 (69%)
Cyclic lipopeptide <sup>52,55,75</sup>	3	0 (0%)	Daptomycin (3)	27 (6-41)	0 (0%)	Cardiovascular implantable electronic device endocarditis (1), Multiple (2)	0 (0%)	1 (33%)
Cyclic peptide/ polymyxin <sup>44</sup>	1	0 (0%)	Colistin (1)	20 (20-20)	0 (0%)	Ventilator associated tracheobronchitis (1)	0 (0%)	0 (0%)
Echinocandin (Antifungal) <sup>1,9,37</sup>	5	5 (100%)	Anidulafungin (4), Caspofungin (1)	44 (11-100)	1 (20%)	Invasive aspergillosis (1), Candidiasis (1), Oesophageal candidiasis (1), Invasive candidiasis, including candidemia (1), Mucosal candidiasis (1)	0 (0%)	1 (20%)
Glycopeptide <sup>2,48,53,68,69,76-81</sup>	12	4 (33%)	Norvancomycin (2), Oritavancin (1), Vancomycin (9)	73 (20-139)	3 (25%)	Bloodstream infections/bacteraemia (5), Complicated bacteraemia; Infective endocarditis (1), Concomitant MRSA bacteraemia and MRSA osteomyelitis (1), Infective endocarditis (1), Multiple (4)	9 (75%)	8 (67%)
Ketolide <sup>5,70</sup>	4	3 (75%)	Telithromycin (4)	85 (22-115)	2 (50%)	Community acquired pneumonia (4)	3 (75%)	0 (0%)
Macrolide <sup>63</sup>	1	1 (100%)	Azithromycin (1)	101 (101-101)	1 (100%)	Respiratory tract infections (1)	0 (0%)	0 (0%)
Oxazolidinone <sup>24,28,58</sup>	3	1 (33%)	Linezolid (3)	83 (2-239)	1 (33%)	Endocarditis (1), Multiple (2)	1 (33%)	1 (33%)
Polyene (Antifungal) <sup>82</sup>	1	0 (0%)	Amphotericin B (liposomal) (1)	9 (9-9)	0 (0%)	Multiple (1)	0 (0%)	0 (0%)
Quinolone <sup>6,7,16-19,26,34,39,42,45,47,50,51,61,65,67,83-85</sup>	21	8 (38%)	Ciprofloxacin (4), Garenoxacin (3), Gatifloxacin (3), Grepafloxacin (3), Levofloxacin	58 (4-216)	3 (14%)	Chronic bronchitis (1), Multiple (3), Nosocomial lower respiratory tract infections (1), Nosocomial	2 (10%)	3 (14%)

Commented [A1]: DO NOT REMOVE - removing comment box will cause problem with table formatting

Antimicrobial class	Number of studies	Industry funded, n (%)	Specific antibiotic (number of studies)	Average number of people (range)	Number of studies with ≥100 participants, number of studies (%)	Infections (number of studies)	Single pathogen, number of studies (%)	Concurrent antibiotics or antifungals, number of studies (%)*
			(6), Sitafloxacin (1), Sparfloxacin (1)			pneumonia (including ventilator associated pneumonia) (3), Otitis media (1), Respiratory tract infections (5), Secondary infections of chronic respiratory diseases (1), Acute exacerbation of chronic bronchitis (3), Intraabdominal infections (1), Acute maxillary sinusitis (2)		
Tetracycline <sup>20-23,29,40,41</sup>	7	6 (86%)	Doxycycline/hydroxychloroquine (1), Tigecycline (6)	62 (15-123)	1 (14%)	Community acquired pneumonia (1), Complicated skin and skin-structure infections (1), Hospital acquired pneumonia (2), Q fever endocarditis (1), Complicated intra-abdominal infections (2)	1 (14%)	1 (14%)
Triazole (Antifungal) <sup>1,25,38</sup>	3	2 (67%)	Cyclodextrin/Itraconazole (1), Voriconazole (2)	151 (23-404)	1 (33%)	Invasive aspergillosis (1), Oropharyngeal candidosis (1), Invasive fungal infections (1)	0 (0%)	1 (33%)
β-lactam <sup>7,8,10,15,27,30,35,36,43,49,54,56,57,60,62,64,66,86,87</sup>	20	6 (30%)	Amoxicillin/clavulanic acid (1), Cefaclor advanced formulation (1), Cefazedone (1), Cefepime (1), Cefmenoxime (3), Cefoperazone/sulbactam (1), Ceftazidime (2), Ceftobiprole (1), Ertapenem (1), Imipenem (or imipenem/cilastatin) (2), Meropenem (3), Piperacillin/tazobactam (1), Tebipenem pivoxil (1), Multiple beta-lactams (1)	42 (6-309)	1 (5%)	Acute otitis media (1), Acute pulmonary exacerbations of cystic fibrosis (1), Community acquired pneumonia (1), Complicated skin and skin-structure infections (1), Febrile neutropenia (1), Lower respiratory tract infections (1), Meningitis (1), Multiple (3), Nosocomial pneumonia (including ICU acquired and ventilator associated pneumonia) (6), Respiratory tract infections (1), Sepsis (1), Ventilator associated pneumonia and signs of severe sepsis or septic shock	1 (5%)	6 (30%)

Antimicrobial class	Number of studies	Industry funded, n (%)	Specific antibiotic (number of studies)	Average number of people (range)	Number of studies with ≥100 participants, number of studies (%)	Infections (number of studies)	Single pathogen, number of studies (%)	Concurrent antibiotics or antifungals, number of studies (%)*
						(1), Acute exacerbation of chronic bronchitis (1)		
<b>Total</b>	<b>97</b>	<b>38 (39%)</b>		<b>59 (2-404)</b>	<b>15 (15%)</b>		<b>24 (25%)</b>	<b>33 (34%)</b>

\* Concurrent antibiotics when antibiotic under investigation; concurrent antifungals when antifungal under investigation.

737  
738

Table 2: Data used to determine PK-PD indices in clinical PK-PD studies, by antimicrobial class.

Antimicrobial class	Number of studies	Pharmacokinetics							Antimicrobial susceptibility			
		Biological material sampled (number of studies)	Mean number of samples (range)*	Sample collection timing reported, number of studies (%) <sup>†</sup>	Method for determining drug concentration reported, number of studies (%)	Free-drug concentration measured, number of studies (%) <sup>‡</sup>	Protein binding adjustment, number of studies (%) <sup>§</sup>	Population PK model used, number of studies (%) <sup>**</sup>	Measure of antimicrobial susceptibility (number of studies)	Method of measurement reported, number of studies (%)	Measurement in accordance with guidelines/ recommendation, number of studies (%) <sup>††</sup>	Strategy if more than one pathogen/ strain, number of studies (%)
Aminoglycoside	16	Blood/serum (16), Sputum (1), Urine (2)	7 (1 to 24)	14 (88%)	14 (88%)	0 (0%)	2 (13%)	10 (63%)	MIC (16)	14 (88%)	5 (31%)	8 (50%)
Cyclic lipopeptide	3	Blood/serum (3)	4 (2 to 9)	2 (67%)	3 (100%)	0 (0%)	0 (0%)	1 (33%)	MIC (3), MBC (1)	1 (33%)	0 (0%)	1 (33%)
Cyclic peptide/ polymyxin	1	Blood/serum (1), BAL fluid (1)	7 (7 to 7)	1 (100%)	1 (100%)	0 (0%)	0 (0%)	0 (0%)	MIC (1)	1 (100%)	0 (0%)	0 (0%)
Echinocandin (Antifungal)	5	Blood/serum (5)	6 (3 to 9)	4 (80%)	1 (20%)	0 (0%)	0 (0%)	5 (100%)	MIC (5)	4 (80%)	4 (80%)	1 (20%)
Glycopeptide	12	Blood/serum (12)	2 (1 to 11)	5 (42%)	5 (42%)	1 (8%)	3 (27%)	7 (58%)	MIC (12)	11 (92%)	8 (67%)	8 (67%)
Ketolide	4	Blood/serum (4)	2 (1 to 3)	3 (75%)	4 (100%)	0 (0%)	0 (0%)	4 (100%)	MIC (4)	4 (100%)	3 (75%)	4 (100%)
Macrolide	1	Blood/serum (1)	8 (8 to 8)	1 (100%)	1 (100%)	0 (0%)	0 (0%)	1 (100%)	MIC (1)	1 (100%)	1 (100%)	1 (100%)
Oxazolidinone	3	Blood/serum (3)	8 (3 to 18)	1 (33%)	2 (67%)	0 (0%)	0 (0%)	1 (33%)	MIC (2), MBC (1)	2 (67%)	0 (0%)	0 (0%)
Polyene (Antifungal)	1	Blood/serum (1)	13 (13 to 13)	1 (100%)	1 (100%)	0 (0%)	0 (0%)	1 (100%)	MIC (1)	1 (100%)	0 (0%)	0 (0%)
Quinolone	21	Blood/serum (21), Sputum (1), Sinus aspirate (2)	6 (1 to 12)	16 (76%)	20 (95%)	2 (10%)	5 (26%)	15 (71%)	MIC (18), MIC midpoint (3)	17 (81%)	11 (52%)	10 (48%)
Tetracycline	7	Blood/serum (5), Not reported (2)	2 (1 to 5)	4 (57%)	2 (29%)	0 (0%)	3 (43%)	5 (71%)	MIC (7)	5 (71%)	3 (43%)	5 (71%)
Triazole (Antifungal)	3	Blood/serum (3)	7 (1 to 16)	1 (33%)	2 (67%)	0 (0%)	1 (33%)	1 (33%)	MIC (3)	2 (67%)	2 (67%)	0 (0%)
β-lactam	20	Blood/serum (20), Middle	5 (1 to 9)	12 (60%)	19 (95%)	1 (5%)	9 (47%)	11 (55%)	MIC (17), MIC midpoint (1),	18 (90%)	4 (20%)	9 (45%)

Antimicrobial class	Number of studies	Pharmacokinetics							Antimicrobial susceptibility			
		Biological material sampled (number of studies)	Mean number of samples (range)*	Sample collection timing reported, number of studies (%) <sup>†</sup>	Method for determining drug concentration reported, number of studies (%)	Free-drug concentration measured, number of studies (%) <sup>‡</sup>	Protein binding adjustment, number of studies (%) <sup>§</sup>	Population PK model used, number of studies (%) <sup>**</sup>	Measure of antimicrobial susceptibility (number of studies)	Method of measurement reported, number of studies (%)	Measurement in accordance with guidelines/recommendation, number of studies (%) <sup>††</sup>	Strategy if more than one pathogen/strain, number of studies (%)
		ear effusion (1), Sputum (1)							Dynamic response (2)			
<b>Total</b>	<b>97</b>		<b>5 (1 to 24)</b>	<b>65 (67%)</b>	<b>75 (77%)</b>	<b>4 (4%)</b>	<b>23 (25%)</b>	<b>62 (64%)</b>		<b>81 (84%)</b>	<b>41 (42%)</b>	<b>47 (48%)</b>

740 \* Mean number of samples taken per patient as reported in results. If range only reported, lower number taken. If number taken not reported but the timing of samples  
741 was reported, the number of samples taken was estimated by counting the number of timepoints. If number taken not reported and timing not reported, but it was clear  
742 than concentration of antibiotic in biological material had been measured, it was assumed that one sample had been taken. NB If the study created a population PK model,  
743 the number of samples was the number of samples per patient who contributed to the population PK model.

744 <sup>†</sup> Had to report more than just "peak" or "trough"

745 <sup>‡</sup> Stated that free-drug concentration measured

746 <sup>§</sup> % of studies which did not measure free-drug concentrations

747 <sup>\*\*</sup> Had to explicitly state that a population PK model used or judged by Alasdair McGowan to have used/constructed a population PK model

748 <sup>††</sup> For example, Clinical & Laboratory Standards Institute (CLSI), National Committee for Clinical Laboratory Standards (NCCLS), British Society for Antimicrobial  
749 Chemotherapy (BSAC) or Japan Society of Chemotherapy guidelines.



Table 3: Type and number of PK-PD indices determined in clinical PK-PD studies, by antimicrobial class.

Antimicrobial class	Number of studies	Number of studies (%) that calculated some form of:				Other PK-PD indices (number of studies)	≥ 2 indices, number of studies (%)	≥ 3 indices, number of studies (%)	≥ 4 indices, number of studies (%)
		AUC/measure of microbial susceptibility	C <sub>max</sub> /measure of microbial susceptibility	C <sub>min</sub> /measure of microbial susceptibility	T> measure of microbial susceptibility				
Aminoglycoside	16	10 (63%)	13 (81%)	2 (13%)	3 (19%)	Pre-dialysis concentration/MIC (1), Average concentration/MIC (1), Intensity index (1)	10 (63%)	5 (31%)	0 (0%)
Cyclic lipopeptide	3	2 (67%)	1 (33%)	0 (0%)	0 (0%)	None	0 (0%)	0 (0%)	0 (0%)
Cyclic peptide/polymyxin	1	1 (100%)	1 (100%)	0 (0%)	1 (100%)	None	1 (100%)	1 (100%)	0 (0%)
Echinocandin (Antifungal)	5	5 (100%)	1 (20%)	1 (20%)	0 (0%)	None	1 (20%)	1 (20%)	0 (0%)
Glycopeptide	12	9 (75%)	1 (8%)	4 (33%)	3 (25%)	None	4 (33%)	1 (8%)	0 (0%)
Ketolide	4	4 (100%)	3 (75%)	0 (0%)	0 (0%)	None	3 (75%)	0 (0%)	0 (0%)
Macrolide	1	1 (100%)	0 (0%)	0 (0%)	0 (0%)	None	0 (0%)	0 (0%)	0 (0%)
Oxazolidinone	3	2 (67%)	0 (0%)	0 (0%)	2 (67%)	Concentration/MBC (1)	2 (67%)	0 (0%)	0 (0%)
Polyene (Antifungal)	1	1 (100%)	1 (100%)	0 (0%)	0 (0%)	None	1 (100%)	0 (0%)	0 (0%)
Quinolone	21	20 (95%)	11 (52%)	3 (14%)	6 (29%)	Concentration/MIC (1)	11 (52%)	6 (29%)	3 (14%)
Tetracycline	7	6 (86%)	0 (0%)	0 (0%)	0 (0%)	Concentration/MIC (1)	0 (0%)	0 (0%)	0 (0%)
Triazole (Antifungal)	3	2 (67%)	1 (33%)	1 (33%)	1 (33%)	C <sub>avg</sub> /MIC (1)	1 (33%)	1 (33%)	1 (33%)
β-lactam	20	10 (50%)	7 (35%)	5 (25%)	19 (95%)	Free concentration at steady state/MIC (1), AUC above MIC (1)	11 (55%)	8 (40%)	3 (15%)
<b>Total</b>	<b>97</b>	<b>73 (75%)</b>	<b>40 (41%)</b>	<b>16 (16%)</b>	<b>35 (36%)</b>		<b>45 (46%)</b>	<b>23 (24%)</b>	<b>7 (7%)</b>

Table 4: Outcomes investigated in clinical PK-PD studies, by antimicrobial class.

Antimicrobial class	Number of studies	Composite measure of treatment failure or success, number of studies (%)	Bacteriological response, number of studies (%)	Time to bacteriological response, number of studies (%)	Clinical response, number of studies (%)	Time to clinical response, number of studies (%)	Mortality, number of studies (%)	Other outcomes (number of studies)	Timing of outcome assessment reported, number of studies (%)*	At least one outcome at the end of therapy, number of studies (%)*
Aminoglycoside	16	2 (13%)	2 (13%)	0 (0%)	6 (38%)	0 (0%)	3 (19%)	SOFA score>3 at day 7 (1), Duration of mechanical ventilation (1), Development of acute kidney injury (1), Improvement/change in pulmonary function tests (including forced expiratory volume in 1s and forced vital capacity) (5), Change in inflammatory parameters (CRP, leukocyte count and IgG) (2), Time (days) to temperature resolution (1), Time (days) to leukocyte count resolution (1), Change in the number of colony forming units (2).	12 (75%)	6 (38%)
Cyclic lipopeptide	3	0 (0%)	0 (0%)	0 (0%)	2 (67%)	0 (0%)	1 (33%)	None	2 (67%)	1 (33%)
Cyclic peptide/polymyxin	1	0 (0%)	1 (100%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	None	1 (100%)	1 (100%)
Echinocandin (Antifungal)	5	2 (40%)	0 (0%)	0 (0%)	1 (20%)	0 (0%)	1 (20%)	Global response (1), Endoscopic response (1).	5 (100%)	3 (60%)
Glycopeptide	12	7 (58%)	2 (17%)	1 (8%)	3 (25%)	0 (0%)	3 (25%)	Recurrence of MRSA bacteraemia within 60 days of discontinuation of therapy (1), length of stay (1), infection-related length of stay (1), alteration of therapy from vancomycin (1), recurrent MRSA bacteraemia within 90 days (1), hospital readmission within 30 days (1), Resolving bacteraemia (<3 days of bacteraemia with all subsequent blood cultures documented to be negative after the initial positive blood culture) versus persistent bacteraemia (bacteraemia for ≥7 days while receiving appropriate antibiotic therapy) (1).	6 (55%)	3 (27%)
Ketolide	4	0 (0%)	4 (100%)	0 (0%)	1 (25%)	0 (0%)	0 (0%)	None	1 (25%)	0 (0%)
Macrolide	1	0 (0%)	1 (100%)	0 (0%)	1 (100%)	0 (0%)	0 (0%)	None	1 (100%)	0 (0%)
Oxazolidinone	3	0 (0%)	2 (67%)	1 (33%)	3 (100%)	0 (0%)	0 (0%)	None	1 (33%)	1 (33%)
Polyene (Antifungal)	1	0 (0%)	0 (0%)	0 (0%)	1 (100%)	0 (0%)	0 (0%)	None	0 (0%)	0 (0%)
Quinolone	21	0 (0%)	16 (76%)	5 (24%)	13 (62%)	1 (5%)	0 (0%)	Time to 70% decrease in sputum volume from peak (1), Time to 25% decrease in the percentage of neutrophils in sputum from peak (1), Time to 60% decrease in coughs per day from peak (1), Time to equilibration of sinus aspirate neutrophil elastase, and	11 (61%)	5 (27%)

								myeloperoxidase concentration with that of plasma, and time to postnasal drip resolution (1), Time to clinical resolution of individual signs and symptoms of maxillary sinusitis (sinus pain, dental pain, sinus tenderness, purulent nasal discharge, headache, face pressure, nasal congestion, postnasal drip, sore throat, cough) (1).		
Tetracycline	7	0 (0%)	5 (71%)	0 (0%)	6 (86%)	0 (0%)	0 (0%)	Time to defervescence (fever resolution) (1), Response (if the clinical signs and symptoms had regressed along with a >2-fold decrease in antibody titre (IgG and/or IgA) to phase I antigen at the completion of a yearlong course of treatment) versus patients in whom the regression of clinical signs and symptoms was not associated with a fall in antibody titre to phase I antigen (1).	7 (100%)	0 (0%)
Triazole (Antifungal)	3	0 (0%)	0 (0%)	0 (0%)	1 (33%)	0 (0%)	1 (33%)	Global response (1), Change in oropharyngeal candidiasis score (1).	3 (100%)	3 (100%)
β-lactam	20	2 (10%)	8 (40%)	3 (15%)	10 (50%)	0 (0%)	2 (10%)	SOFA score>3 at day 7 (1), Duration of mechanical ventilation (1), CRP normalisation (1), Number of admission days (1), Time to defervescence (1), Brasfield score (1), Pulmonary function score (1), Clinical score (1), General score (1).	14 (78%)	10 (56%)
<b>Total</b>	<b>97</b>	<b>13 (13%)</b>	<b>41 (42%)</b>	<b>10 (10%)</b>	<b>48 (49%)</b>	<b>1 (1%)</b>	<b>11 (11%)</b>		<b>64 (70%)</b>	<b>33 (36%)</b>

753 \* The fields "timing of outcome assessment reported" and "at least one outcome at the end of therapy" exclude the six studies which exclusively reported time-to-event  
754 outcomes.<sup>8,18,34,45,79</sup>

755 Abbreviations: CRP, C-reactive protein; IgA Immunoglobulin A; IgG Immunoglobulin G; SOFA, Sequential Organ Failure Assessment.

Table 5: Statistical analyses performed in clinical PK-PD studies, by antimicrobial class.

Antimicrobial class	Number of studies	Sample size calculation performed, number of studies (%)	Analysis of 2x2 tables, number of studies	Comparison of means, number of studies	Correlation, number of studies	Receiver operating characteristic (ROC) curve, number of studies	Recursive partitioning, number of studies	Type of analysis (number of studies)	Standard and/or non-linear (number of studies)	Adjusted and/or unadjusted (number of studies)	No statistical analyses, number of studies
Aminoglycoside	16	1 (6%)	3	4	5	1	4	Logistic regression (9), Time to event analysis (1), Hill equation/ $E_{max}$ model (1), Repeated-measures mixed effects (1)	Standard (10), Non-linear (1)	Unadjusted (7), Adjusted (7)	1
Cyclic lipopeptide	3	0 (0%)	1	0	0	0	0	Logistic regression (1)	Standard (1)	Adjusted (1)	1
Cyclic peptide/ Polymyxin	1	0 (0%)	0	0	1	0	0				0
Echinocandin (Antifungal)	5	0 (0%)	0	0	0	0	0	Logistic regression (5)	Standard (5)	Unadjusted (2), Adjusted (4)	0
Glycopeptide	12	1 (8%)	7	8	0	0	7	Logistic regression (9), Poisson regression (1), Time to event analysis (1)	Standard (11)	Unadjusted (2), Adjusted (11)	0
Ketolide	4	0 (0%)	0	0	0	0	4	Logistic regression (4)	Standard (4)	Unadjusted (4), Adjusted (1)	0
Macrolide	1	0 (0%)	0	0	0	0	0	Logistic regression (1)	Standard (1)	Adjusted (1)	0
Oxazolidinone	3	0 (0%)	1	0	0	0	1	Logistic regression (1), Time to event analysis (1), Hill equation/ $E_{max}$ model (1)	Standard (1), Non-linear (1)	Adjusted (1)	1
Polyene (Antifungal)	1	0 (0%)	0	1	0	0	0				
Quinolone	21	0 (0%)	2	3	1	1	6	Logistic regression (9), Time to event analysis (4), Hill equation/ $E_{max}$ model (3)	Standard (11), Non-linear (3)	Unadjusted (12), Adjusted (8)	6
Tetracycline	7	0 (0%)	4	1	0	2	5	Logistic regression (6), Time to event analysis (1)	Standard (6)	Unadjusted (6), Adjusted (5)	0
Triazole (Antifungal)	3	0 (0%)	0	0	0	0	0	Logistic regression (2), Hill equation/ $E_{max}$ model (1)	Standard (2), Non-linear (2)	Unadjusted (3), Adjusted (2)	0
$\beta$ -lactam	20	1 (5%)	4	3	4	1	4	Logistic regression (4), Linear regression (1), Time to event analysis (1)	Standard (6)	Unadjusted (5), Adjusted (1)	9
<b>Total</b>	<b>97</b>	<b>3 (3%)</b>	<b>22 (23%)</b>	<b>20 (21%)</b>	<b>11 (11%)</b>	<b>5 (5%)</b>	<b>31 (32%)</b>				<b>18 (19%)</b>

**Suggested guidelines for conducting studies.**

1. Consider sample size calculations to determine whether an exposure-response relationship has a reasonable likelihood of being identified. Conducting analyses on too few patients is unlikely to yield meaningful results.
2. Consider reporting confidence or credible intervals measuring the level of uncertainty in the results to avoid over-interpretation when PK-PD index- response relationships are not identified.
3. Ensure the population is as homogeneous as possible with respect to infection and infecting pathogen, or control for these factors. This will facilitate the detection of PK-PD index- response relationships. Similarly, it may be desirable to standardise concurrent treatments.
4. Derive the PK parameters of the antimicrobial in a robust manner from sufficient samples (the sampling framework may be derived using Stochastic Optimal Design or similar methodology) and using an appropriate population PK model. If free concentrations of antimicrobials are important, then these should be measured rather than adjusting for protein binding using a fixed rate to allow for the fact that protein binding may vary. For some infections, concentration at the site of infection may be important and this should be measured if possible.
5. Perform antimicrobial susceptibility testing of infecting pathogens before the start of therapy, and determine them using standardised methodology. If possible, it may be preferable to store strains to test concurrently in one laboratory.
6. Consider having standardised outcomes and timing of outcomes. The outcomes should be relevant to the patient. PK-PD index-response relationships may be more likely if continuous rather than dichotomous outcomes are used.
7. Follow a pre-specified analysis plan. The most appropriate way of statistically analysing the

relationship between PK-PD indices and outcomes needs to be further investigated. Explicit modelling of the PK-PD index, for example using fractional polynomials,<sup>88</sup> may be preferable to recursive partitioning, if the sample size is sufficient. It may be advisable to produce a standardised list of covariates that should be assessed to see if they are associated with outcome, for example severity of illness and presence of comorbidities. This list may vary by indication. The investigation of the influence of covariates on the PK-PD relationship for efficacy using multivariable analyses and through the findings of interactions with the PK-PD index may help further the understanding of which subsets of patients are at increased risk of suboptimal drug exposure.

8. The evaluation of PK-PD indices achieved among patients relative to non-clinical PK-PD targets for efficacy is useful. Such information provides dose selection support, especially if PK-PD indices achieved are on the upper plateau of the non-clinical PK-PD relationship for efficacy.