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1 Local host-dependent persistence of the entomopathogenic nematode Steinernema carpocapsae

2 used to control the large pine weevil Hylobius abietis

3 Christopher D. Harvey and Christine T. Griffin

4 Abstract

Entomopathogenic nematodes (EPN) applied inundatively to suppress insect pests are more 5 6 likely to persist and establish in stable agroecosystems than in annual crops. We investigated a 7 system of intermediate stability: Tree stumps harbouring the large pine weevil (Hylobius abietis 8 L.; Coleoptera: Curculionidae), a major European forestry pest. We tested whether persistence of 9 EPN Steinernema carpocapsae Weiser (Rhabditida: Steinernematidae) applied around stumps is 10 maintained by recycling of EPN through pine weevils developing within stumps. Steinernema 11 carpocapsae was detected in soil around and under the bark of treated tree stumps up to 2 years, 12 but not 4-5 years after application. Differences in nematode presence between sites were better explained by tree species (pine or spruce) than soil type (mineral or peat). Presence of S. 13 14 carpocapsae in soil was positively correlated with the number of H. abietis emerging from untreated stumps the previous year, which was greater for pine stumps than spruce stumps. 15

16 Keywords: Steinernema; Hylobius; entomopathogenic nematodes; pine weevil; persistence; bark

17 1. Introduction

Entomopathogenic nematodes (EPN) of the genera Steinernema (Rhabditida: Steinernematidae) and Heterorhabditis (Rhabditida: Heterorhabdidae) are lethal pathogens of insects with a wide potential host range (Bathon, 1996; Smits, 1996) that are used against pests in horticulture, agriculture and forestry (Grewal et al. 2005). The free living infective juvenile (IJ) invades the 22 haemocoel of insects and releases symbiotic bacteria that cause toxaemia and/or septicaemia, killing the insect within days (Kaya and Gaugler 1993). EPN are mainly used as inundative 23 biological control agents, with insect suppression being effected by the applied IJs. Following 24 application to soil, numbers of IJs typically decrease rapidly and may reach less than 90% of the 25 original inoculum within days (Glazer 1992; Smits 1996; Griffin 2015). Applied nematodes may 26 27 survive at low numbers for longer periods (Preisser et al. 2005), but longer term persistence of a population depends on recycling - reproduction in target and/or non-target hosts (Campbell et al. 28 1995; Griffin 2015; Koppenhöfer and Fuzy 2009; Peters 1996). Long-term persistence of EPN 29 30 populations therefore crucially depends on the availability of host insects for reproduction, as well as suitable environmental conditions, and hence varies between agronomic systems. Stable 31 ecosystems such as pasture and alfalfa favour long-term persistence and EPN populations can 32 persist for years after application (Koppenhöfer and Fuzy 2009; Shields et al. 1999), while in 33 annual crops, persistence beyond a year is less common (Susurluk and Ehlers 2008). Tree stumps 34 as a breeding resource for certain forestry pests such as the large pine weevil, Hylobius abietis L. 35 (Coleoptera: Curculionidae) represent a moderately stable environment, intermediate between 36 annual and perennial crops. 37

The large pine weevil, H. abietis, is one of the most damaging forestry pests in Europe (Långström and Day 2004; Leather et al. 1999). Development takes place under the bark of recently dead conifers, including stumps of recently felled trees, while adults feed on the bark of saplings planted to restock clear-fell sites, often leading to extensive sapling damage and mortality (Leather et al. 1999; Månsson and Schlyter 2004). Clear-felled coniferous forest plantation sites can support large weevil populations (Leather et al. 1999; Örlander et al. 1997). Stumps can remain suitable for pine weevil oviposition for up to three years after felling

45 (Nordenhem 1989) and emergence of adults occurs within one to two years of oviposition (Leather et al. 1999). Traditionally, seedlings are protected by chemical insecticides, but 46 application of EPN to tree stumps, targeting immature weevils developing within, has shown 47 promise for suppression of adult weevil populations (Brixey et al. 2006; Dillon et al. 2006, 2007 48 and 2008a,b; Torr et al. 2007; Williams et al. 2013). Dillon et al. (2008a) investigated the fate 49 50 over a five year period of four EPN species applied by hand to tree stumps harbouring pine weevil developmental stages. Incidence (percentage of soil cores positive) of all species 51 remained high for the first two years (no difference between months 1, 12 and 24 post 52 53 application), but declined by year three post application (Dillon et al. 2008a). Only Steinernema feltiae Filipjev (Rhabditida: Steinernematidae; native to clearfell sites) was recovered in years 4 54 and 5. Pine weevil larvae can support EPN reproduction, yielding up to 98,000 IJs per insect 55 (Dillon 2003). Dillon et al. (2008a) hypothesized that EPN populations initially remained high 56 due to recycling in the target pest, and that the apparent disappearance after 4-5 years of 57 Steinernenema carpocapsae Weiser and two Heterorhabditis species was due to a concomitant 58 decrease in weevil numbers as stumps degraded. The Dillon et al. (2008a) study was conducted 59 on a single site type: stumps of pine (Pinus sylvestris L. and Pinus contorta Douglas) on a deep 60 61 peat soil.

The present study complements the Dillon et al. (2008a) report of EPN persistence in a forest ecosystem, focussing on a single EPN species (S. carpocapsae) but extending the investigation to sites with diverse characteristics (soil type and tree species). For this study, nematodes were applied to stumps not by hand, but on a site-wide operational scale using spray nozzles connected to a tank of nematode suspension mounted on a forwarder. Specific objectives are: (1) confirm that the restricted spatial and temporal distribution of EPN reported by Dillon et al

(2008a) for Pinus spp. on peat in small scale trials is also applicable to commercial scale trials on 68 sites with mineral soil and sites planted with Sitka spruce (Picea sitchensis Carr.), the species 69 predominating in Irish and UK plantation forests (Anon. 2003 & 2007). (2) Investigate the 70 occurrence of EPN within the stump. Multiplication of EPN in pine weevils located in or under 71 the bark is expected to release IJs into the space between the bark and the woody material of the 72 73 tree-stump, but this has not been previously reported. (3) Investigate the relationship between EPN incidence in soil around stumps and the size of weevil populations within stumps. If EPN 74 depend mainly on pine weevil as host, a positive correlation between weevil and EPN 75 76 populations across sites is expected.

77 2. Material and methods

78 2.1 Study sites

79 In 2007 and 2008, S. carpocapsae (All strain; Becker Underwood; Littlehampton, England) at a 80 rate of 3.5 million IJs per stump applied in 500 ml water was applied to several clear-fell sites 81 (Table 1) on an operational, site-wide scale (Williams et al. 2013). Steinernema carpocapsae was 82 chosen for the study as it is the only species to date that has been applied operationally against pine weevil (Williams et al. 2013). On each site, a small number of stumps were marked and not 83 treated, to serve as controls. These control stumps allowed an assessment of weevil populations 84 within the stumps, based on the number of adult weevils caught in emergence traps erected over 85 them (Williams et al. 2013). The soil type on each site (peat or mineral soil) was based on 86 records of Coillte Teoranta (the site owner) and confirmed by visual evaluation on site. Conifer 87 forests in northern Europe (including 44% of Irish forests) are frequently planted on former peat 88

bog, having high levels of organic matter; for our purposes, all other soils were classed asmineral soils, having lower organic content.

91 2.2 Soil sampling

Soil and bark samples were taken from stumps spaced at 5 to 15 m intervals along a diagonal 92 transect; for n of stumps sampled see Table 2. Soil cores were collected at four aspects (at right 93 angles) around each sampled stump. One soil core each was taken at the bole (0 cm) and at 20, 94 40 and 60 cm along each aspect, resulting in 16 soil cores per sampled stump. Cores were taken 95 to a depth of approximately 5 cm using a 50 ml plastic tube (2.9 cm inner diameter; Sarstaedt; 96 Nürnbrecht, Germany) which was also used for transport and baiting. Soil cores were baited with 97 final instar waxmoth larvae (Galleria mellonella L; Lepidoptera: Pyralidae) at room temperature 98 99 (Dillon et al. 2006, 2007). Each core was baited twice for 7 days with one bait insect each time. Live insects were incubated at 20°C for a further seven days after removal from soil. Insects that 100 showed signs of Steinernema infection (cadaver consistency and cream/tan colour) were 101 102 incubated at 20°C until IJs emerged. IJs were then measured for length (10 per cadaver) and scored as either S. carpocapsae (mean length: 558µm, length range: 438-650µm) (Adams and 103 104 Nguyen 2002) or a native Steinernema sp. (S. feltiae: mean: 849, range: 736-950; Steinernema kraussei Steiner: mean: 951, range: 797-1102) (Adams and Nguyen 2002). Steinernema feltiae 105 and (rarely) S. kraussei are the only species so far detected in Irish conifer forests (Griffin et al. 106 1991; Gwynn and Richardson 1996; Dillon 2003; C. Harvey unpublished). For samples collected 107 2 years post-application (p.a.) or where no IJs emerged, cadavers were dissected, and spicules of 108 109 male adult nematodes were used for identification (Adams and Nguyen 2002). Cadavers 110 containing no adults or only females were scored as inconclusive and not included in analysis.

111 2.3 Baiting of bark samples from tree stumps treated with S. carpocapsae

At each sampled stump, approximately 100 cm<sup>2</sup> of bark was stripped from the bole of the stump 112 at the soil horizon at each aspect. Bark from each stump was pooled into a bulk sample, and 113 placed in a 250 ml plastic cup for baiting. Ten wax moth larvae were added per cup and cups 114 were covered with Parafilm (Bemis; Soignies, Belgium) and incubated at 20°C. After three days, 115 bait insect mortality was recorded. For sites A1-A4 in addition, 4 small pieces of bark (approx. 4 116 cm<sup>2</sup> each), one from each of the four sampled aspects, were individually baited with a single wax 117 moth larva. The insect was placed in a well (0.9 cm diam.) and covered by the piece of bark as 118 described by Harvey and Griffin (2012), so that it was in contact with only the under surface of 119 the bark. Since previous studies indicated that infection of insects under bark of tree stumps with 120 121 native steinernematids is extremely rare (Dillon 2003; Dillon et al. 2008; C. Harvey, personal observation), dead insects with cream colouration were scored as infected by S. carpocapsae. 122

123 2.5 Statistics

Statistical analysis was carried out using MiniTab Release 15 (MiniTab Solutions; Coventry, 124 125 UK). To compare the proportion of samples scoring positive for the presence of S. carpocapsae over time (successive years after application) or between sites, these binary data 126 (positive/negative) were compared using 2x2 contingency tables with Pearson's  $\chi^2$  – test or, 127 where the expected count of at least one cell in the table was < 5, with Fisher's exact test ( $\alpha =$ 128 0.05). Yates' correction was used for  $\chi^2$  – tests on 5x2 tables with expected counts <5. 129 Significance levels of multiple pairwise comparisons of binary data between sampling time 130 points and/or sites were adjusted for type-I family error rate after Bonferroni, with the 131 significance level for n pairwise comparisons involving the same data set adjusted to 0.05/n 132

133 (Rice 1989). The Mantel-Haenszel-Cochran (MHC) test was used to calculate odds ratios and 134 detect effects of soil type on presence of EPN (in soil cores, at stumps and under bark) while controlling for the effect of tree stump species and vice versa. To investigate whether S. 135 carpocapsae presence in soil cores and in bark samples was correlated with the size of 136 137 previously recorded weevil populations in stumps, the percentage of soil cores and bark samples 138 scoring positive for S. carpocapsae 2 years p.a. (2009) was regressed in binary logistic regression models for each response variable against the  $log_{10}(x+1)$  of the mean number of adult 139 weevils emerging from untreated control stumps on each of the corresponding sites in 2008 140 141 (predictive variable; see supplementary data, Dillon et al. 2012), which represented an indicator for the size of pine weevil populations within stumps on each site. The Wald-test was used to test 142 the model coefficient for significant difference from 0. Data for spruce and pine stumps at site 143 A1 (Lackenrea) were used separately in the model. Pearson's Goodness-of-Fit test ( $\alpha = 0.05$ ) 144 was used to confirm validity of the Logit link function used in the model and Pearson residuals 145 were tested for normality (Anderson-Darling test,  $\alpha = 0.05$ ). To test for differences between the 146 147 percentage of soil cores scoring positive for S. carpocapsae at four distances from the bole of stumps, a two-way Chi-square test was used on data combined for each distance across sites A1 148 149 to A4.

150 3 Results

151 3.1 Presence of entomopathogenic nematodes in soil samples

Steinernema carpocapsae was detected in soil samples from stumps at all four sites that were sampled up to two years p.a., but not on any of the six sites sampled 4-5 years p.a. (Table 2). Conversely, other Steinernema spp. were found at five out of six sites sampled 4-5 years p.a., but

not earlier (Table 2). At the sites where soil was sampled in each of years 1 and 2 p.a. (A1 and A2), incidence of S. carpocapsae decreased significantly over time in the three samples taken five months, one year and two years after nematode application (A1: cores:  $\chi^2_2 = 8.769$ , P = 0.012; stumps:  $\chi^2_2 = 12.115$ , P = 0.002 [data combined for pine and spruce]; A2: cores  $\chi^2_2 =$ 19.627, P < 0.001; stumps:  $\chi^2_2 = 16.586$ , P < 0.001; Table 2).

Two years p.a., S. carpocapsae was recovered from 0.9 to 5.2% of soil cores, representing 11-160 43% of sampled stumps (Table 2). Differences between sites (A1-A4, treating spruce and pine on 161 A1 separately) were significant based on both cores ( $\chi^2_4$ = 27.311, P <0.001) and stumps (Yates' 162  $\chi^2 = 12.522$ , d.f = 4; p = 0.014). Two years p.a., soil type had a significant effect on the 163 proportion of soil cores scoring positive for EPN when controlling for the effect of stump species 164 165 (MHC = 7.825, odds ratio [peat:mineral] = 2.494, P = 0.005), but the odds ratio for the effect of stump species when controlling for soil type was more than three times as great, with soil cores 166 167 from pine stumps eight times as likely to score positive for S. carpocapsae as cores from spruce stumps (MHC = 17.935, odds ratio [pine:spruce] = 8.009, P < 0.001) (Table 3). When data for 168 stumps were used instead, soil type had no significant effect, though the effect of stump species 169 remained highly significant (soil: MHC = 1.067, odds ratio peat:mineral] = 1.887, P = 0.302; 170 stump species: MHC = 17.680, odds ratio [pine:spruce] =  $\infty$ , P < 0.001) (Table 3). There was a 171 significant positive relationship between the incidence of S. carpocapsae in soil cores and the 172 number of adult weevils emerging the previous year (Wald-test; Coef. = 1.66, Z = 5.62, P < 1.66173 0.001; Fig. 1a). 174

175 3.2 Presence of entomopathogenic nematodes in bark samples

176 Steinernema carpocapsae was recovered from bark one and two years p.a., but no EPN were found there after 4 or 5 years (Table 2). For most of the stumps on sites A2 and A3 where S. 177 carpocapsae was recovered from bulk bark samples it was also detected when only the inside of 178 179 the bark was baited (site A2, one year p.a.: 15/20 stumps; site A2, two years p.a.: 6/11; site A3, two years p.a.: 9/12). The proportion of stumps where bark samples were positive for S. 180 carpocapsae at sites A1 and A2 decreased from one to two years of application (Table 2), 181 significantly so for site A2 ( $\chi^2_1$  = 6.944, P = 0.008; Table 2). Two years p.a., the proportion of 182 stumps with S. carpocapsae in bulk bark samples ranged from 0 to 67%, a highly significant 183 difference between sites (A1-A4, treating spruce and pine on A1 separately) (Yates'  $\chi^2 = 19.203$ , 184 df = 4, P < 0.001). Two years p.a., stump species had a significant effect on the proportion of 185 stumps with EPN detected under their bark (MHC = 15.108, odds ratio [pine:spruce] =  $\infty$ , P < 186 187 0.001), but soil type did not (MHC = 0.204, odds ratio [peat:mineral] = 1.424, P = 0.651) (Table 3). The percentage of stumps with positive bark samples tended to be higher on sites with high 188 weevil emergence compared with sites with low emergence (Fig. 1b). However, link functions of 189 190 binary logistic models regressing the proportion of tree stumps with bark positive for S. carpocapsae against mean number of weevils emerging from untreated stumps in 2008 did not 191 provide an adequate fit for the data (Pearson's Goodness-of-Fit test, P < 0.05). 192

## 193 3.3 Dispersal of EPN from treated stumps

At all sites where S. carpocapsae was recovered, and at all sampling times, the percentage of cores positive for S. carpocapsae tended to be highest directly at the bole of the stump (0 cm distance) and lowest at 60 cm distance from the bole of a stump. Nematodes were detected 60 cm from the stump bole within 5 months p.a. (data not shown). The percentage of soil cores positive for S. carpocapsae (sites A1 to A4) decreased significantly with increasing distance from the bole of the stump two years p.a. ( $\chi^2_3 = 68.57$ , P < 0.001; Fig. 2).

200 4. Discussion

Our results confirm the finding by Dillon et al. (2008a) that S. carpocapsae declines to 201 undetectable levels 4-5 years after application to coniferous tree stumps for pine weevil control. 202 203 They also support Dillon et al.'s (2008a) suggestion that this is due to a concomitant decrease in the availability of weevils for reproducing as stumps degraded. We did find other steinernematid 204 nematodes 4-5 years after application, probably S. feltiae or S. kraussei, the only other 205 Steinernema spp. so far detected in Irish conifer forests or clear-fells (Dillon 2003; Griffin et al. 206 1991; C. Harvey unpublished data). Similarly, Dillon et al. (2008a) found that the only EPN 207 208 recovered 4-5 years after application was S. feltiae, either an indigenous applied strain or a native strain that naturally colonised the site. It is possible that the abundance of these native EPN 209 species is linked to the availability of soil-associated insect hosts, which may increase in 210 211 diversity and number as clear-fell sites proceed through stages of succession (Butterfield 1997; Irwin et al. 2014; Niemelä et al. 1993; Pawson et al. 2006). 212

Though inundatively applied IJs can survive in soil for months (Dillon et al. 2008a; Kung et al. 1990; Poinar and Hom 1986), up to 90 % of them are expected to die within hours of application (Smits 1996). Therefore, most (if not all) of the S. carpocapsae found at our first sampling time five months p.a. and beyond had likely originated from reproducing through insect hosts in the field (Gaugler 1988; Smits 1996; Susurluk and Ehlers 2008). Nematodes were applied in July 2007 to coincide with the occurrence of late instar larvae and pupae of pine weevil in the stumps. Steinernema carpocapsae can reproduce in immature pine weevil in the field and a single 220 infected weevil larva can yield more than 85,000 IJs (Dillon 2003; Pye and Burman 1978). Our 221 data support the hypothesis that S. carpocapsae was reproducing in immature pine weevils: incidence of IJs in soil and bark was positively associated with the size of weevil populations in 222 223 stumps (as indicated by adult weevil emergence from stumps the year previous). Infection rates of pine weevil by S. carpocapsae in the weeks after application are proportionately similar for 224 225 pine and spruce stumps (Brixey et al. 2006; Dillon et al. 2006 and 2008b) and appear not to be affected by weevil population size per stump (Williams et al. 2013). Thus, differences in S. 226 carpocapsae reproduction between spruce and pine stumps are likely driven by long-term 227 228 differences in overall weevil population size between the two stump species, with higher weevil populations in pine (Dillon 2003; Leather et al. 1999; Williams et al. 2013). While soil type is 229 230 more important than tree species in determining effectiveness of S. carpocapsae against the pine weevil (Williams et al. 2013), tree species (as a predictor of weevil populations in stumps) has a 231 greater effect on continued presence of the nematodes in this ecosystem. 232

Since weevil populations in stumps may remain high for 2-3 years after nematodes are applied 233 (Leather et al. 1999) they may facilitate repeated cycles of reproduction of applied S. 234 carpocapsae under the bark. Longer term presence of S. carpocapsae in the soil (1-2 years p.a.) 235 may thus be explained by IJs migrating into the soil following reproduction under stump bark. 236 Nematode populations fluctuate with the target pest population (Campbell et al. 1995), but 237 238 reproduction in non-target insects may also be important. For example, Hodson et al. (2012) found a positive correlation between persistence of S. carpocapsae in pistachio orchards and 239 240 pitfall catches of non-target tenebrionid beetles. Plantation forests and clear-fell sites can harbour a considerable diversity and number of potential non-target insects (Dillon et al. 2012; Fahy and 241 Gormally 1998; Niemelä and Koivula 2007; Sippola et al. 2002). However, Dillon et al. (2012) 242

found no evidence of an impact of EPN on non-target beetle populations in a study that included some of the sites sampled for this study (sites A1 to A4). Consequently, reproduction of S. carpocapsae through such non-target hosts is unlikely to contribute significantly to the EPN persistence we observed.

247 On all of the sites in our study, S. carpocapsae was applied within two years of felling, when the 248 number of weevils per tree stump tends to peak (Dillon et al. 2007; Leather et al. 1999; Williams et al. 2013). As weevils emerge and stumps degrade, the pest population inevitably drops to a 249 level that no longer supports large nematode populations (Leather et al. 1999). Thus, the 250 ephemeral and contained nature of weevil populations in tree stumps should provide a natural 251 252 limit to EPN presence on clear-fell sites, especially if reproduction in non-target insects is 253 infrequent. We detected S. carpocapsae up to 60 cm from the bole of treated tree stumps, but incidence dropped steeply with increasing distance, a trend similar to previous studies (Torr et al. 254 255 2007; Dillon et al. 2008a). On the clear-fell sites we sampled, the absence of a stable pool of 256 suitable hosts for reproduction outside of tree stumps may have prevented S. carpocapsae from establishing. However, there is potential for local pockets of nematode recycling where dead-257 wood with susceptible longhorn beetles occurs (Harvey et al. 2012). 258

Dispersal and prolonged persistence of control agents is usually not a desired outcome of inundative biological control, mainly because of the increased risk of non-target impacts (Bathon 1996; Smits 1996; van Lenteren et al. 2003). Where EPN reproduction is primarily restricted to the target pest habitat, however, as appears to be the case in our studied system, recycling in the pest may enhance and extend the controlling effect, thereby reducing the need for repeated application while minimising damage to non-target hosts outside of this habitat (Klein and Georgis 1992; Smits 1996). We conclude that, for the large pine weevil the persistence of

266	inundatively applied EPN is dependent on the target pest population, resulting in limited risk of
267	dispersal and longer term establishment while at the same time potentially enhancing control
268	efficacy. The same may be true of other pests with transient populations occurring in cryptic
269	habitats.
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275	References
276	Adams BJ, Nguyen, KB (2002) Taxonomy and systematics. In: Gaugler R (ed)
277	Entomopathogenic Nematology, 1 <sup>st</sup> edn. CAB publishing, Wallingford, England, pp 1-35.
278	Anon. (2003) National inventory of woodland and trees. Forestry Commission, Wexford, Ireland
279	Anon. (2007) National forest inventory Republic of Ireland. Forest Service, Midlothian, UK
280	Bathon H (1996) Impact of entomopathogenic nematodes on non-target Hosts. Biocontrol Sci
281	Technol 6:421–434
282	Brixey JM, Moore R, Milner AD (2006) Effect of entomopathogenic nematode (Steinernema
283	carpocapsae Weiser) application technique on the efficacy and distribution of infection of

284	the large pine weevil (Hylobius abietis L.) in stumps of Sitka spruce (Picea sitchensis Carr.)
285	created at different times. For Ecol Manage 226:161–172
286	Butterfield J, (1997) Carabid community succession during the forestry cycle in conifer
287	plantations. Ecography (Cop.) 20:614-625
288	Campbell JF, Lewis E, Yoder F, Gaugler RJ (1995) Entomopathogenic nematode
289	(Heterorhabditidae and Steinernematidae) seasonal population dynamics and impact on
290	insect populations in turfgrass. Biol Control 5:598-606
291	Dillon AB (2003) Biological control of the large pine weevil, Hylobius abietis L., (Coleoptera:
292	Curculionidae) using entomopathogenic nematodes. PhD thesis submitted at National
293	University of Ireland Maynooth, Ireland
294	Dillon AB, Ward D, Downes MJ, Griffin CT (2006) Suppression of the large pine weevil
295	Hylobius abietis (L.) (Coleoptera: Curculionidae) in pine stumps by entomopathogenic
296	nematodes with different foraging strategies. Biol Control 38:217-226
297	Dillon AB, Downes MJ, Ward D, Griffin CTJ (2007) Optimizing application of
298	entomopathogenic nematodes to manage large pine weevil, Hylobius abietis L.
299	(Coleoptera:Curculionidae) populations developing in pine stumps, Pinus sylvestris. Biol
300	Control 40:253–263
301	Dillon AB, Rolston AN, Meade CV, Downes MJ, Griffin CT (2008a) Establishment, persistence,
302	and introgression of entomopathogenic nematodes in a forest ecosystem. Ecol Appl 18:735-
303	747

304	Dillon AB, Moore CP, Downes MJ, Griffin CT (2008b) Evict or infect? Managing populations
305	of the large pine weevil, Hylobius abietis, using a bottom-up and top-down approach. For
306	Ecol Manage 255:2634–2642
307	Dillon AB, Foster A, Williams CD, Griffin CT (2012) Environmental safety of
308	entomopathogenic nematodes – effects on abundance, diversity and community structure of
309	non-target beetles in a forest ecosystem. Biol Control 63: 107-114
310	Fahy O, Gormally M (1998) A comparison of plant and carabid beetle communities in an Irish
311	oak woodland with a nearby conifer plantation and clearfelled site. For Ecol Manage
312	110:263–273
313	Gaugler R (1988) Ecological considerations in the biological control of soil-inhabiting insects
314	with entomopathogenic nematodes. Agric Ecosyst Environ 24:351-360
315 316	Glazer I (1992) Survival and efficacy of Steinernema carpocapsae in an exposed environment. Biocontrol Sci Technol 2:101–107
317 318	Grewal PS, Ehlers RU, Shapiro-Ilan DI (eds) (2005) Nematodes as biocontrol agents, 1 <sup>st</sup> edn. CAB Publishing, Wallingford, England
319	Griffin CT, Moore JF, Downes MJ (1991) Occurrence of insect-parasitic nematodes
320	(Steinernematidae, Heterorhabditidae) in the Republic of Ireland. Nematologica 37:92-100
321	Griffin CT (2015) Behaviour and population dynamics of entomopathogenic nematodes
322	following application. In: Campos Herrera R (ed) Nematode pathogenesis of insects and
323	other pests – ecology and applied technologies for sustainable plant and crop protection, $1^{st}$
	15

324	edn. Springer,	Berlin, Germany,	pp 57-95.
		, , ,	11

325	Gwynn RL, Richardson PN (1996) Incidence of entomopathogenic nematodes in soil samples
326	collected from Scotland, England and Wales. Fundam appl Nematol 19:427–431
327	Harvey CD, Alameen KM, Griffin CT (2012) The impact of entomopathogenic nematodes on a
328	non-target, service-providing longhorn beetle is limited by targeted application when
329	controlling forestry pest Hylobius abietis. Biol Control 62:173-182
330	Harvey CD, Griffin CT (2012) Host activity and wasp experience affect parasitoid wasp foraging
331	behaviour and oviposition on nematode-infected larvae of the forestry pest Hylobius abietis.
332	Ecol Entomol 37:269–282
333	Hodson AK, Siegel JP, Lewis EE (2012) Ecological influence of the entomopathogenic
334	nematode, Steinernema carpocapsae, on pistachio orchard soil arthropods. Pedobiologia
335	55:51–58
336	Irwin S, Pedley S, Coote L, Dietzsch A, Wilson M, Oxbrough A, Sweeney O, Moore K, Martin
337	R, Kelly D, Mitchell FG, Kelly T, O'Halloran J (2014) The value of plantation forests for
338	plant, invertebrate and bird diversity and the potential for cross-taxon surrogacy. Biodivers
339	Conserv 23:697–714
340	Kaya HK, Gaugler R (1993) Entomopathogenic nematodes. Annu Rev Entomol 38:181–206
341	Klein MG, Georgis R (1992) Persistence of control of Japanese Beetle (Coleoptera:
342	Scarabaeidae) larvae with steinernematid and heterorhabditid nematodes. J Econ Entomol
343	85:727-730

344	Koppenhöfer AM, Fuzy EM (2009) Long-term effects and persistence of Steinernema scarabaei
345	applied for suppression of Anomala orientalis (Coleoptera: Scarabaeidae). Biol Control
346	48:63-72
347	Kung SP, Gaugler R, Kaya HK (1990) Soil type and entomopathogenic nematode persistence. J
348	Invertebr Pathol 55:401–406
349	Långström B, Day KR (2004) Damage, control and management of weevil pests, especially
350	Hylobius abietis. In: Leutier F (ed) Bark and wood boring insects in living trees in
351	Europe, a synthesis, 1 <sup>st</sup> edn. Springer, Berlin, Germany, pp 415-444.
352	Leather SR, Day KR, Salisbury AN (1999) The biology and ecology of the large pine weevil,
353	Hylobius abietis (Coleoptera: Curculionidae): a problem of dispersal? Bull Entomol Res
354	89:3–16
355	Månsson PE, Schlyter F (2004) Hylobius pine weevils adult host selection and antifeedants:
356	feeding behaviour on host and non-host woody scandinavian plants. Agr Forest Entomol
357	6:165-171
358	Niemelä J, Langor D, Spence JR (1993) Effects of clear-cut harvesting on boreal ground-beetle
359	assemblages (Coleoptera: Carabidae) in Western Canada. Conserv Biol 7:551–561
360	Niemelä J, Koivula MKDJ (2007) The effects of forestry on carabid beetles (Coleoptera:
361	Carabidae) in boreal forests. J Insect Conserv 11:5-18
362	Nordenhem, H (1989) Age, sexual development, and seasonal occurrence of the pine weevil
363	Hylobius abietis (L.). J Appl Entomol 108:260-270

364	Örlander G, Nilsson U, Nordlander G (1997) Pine weevil abundance on clear-cuttings of
365	different ages: A 6-year study using pitfall traps. Scand J For Res 12:225–240
366	Pawson SM, Brockerhoff EG, Norton DA, Didham RK (2006) Clear-fell harvest impacts on
367	biodiversity: past research and the search for harvest size thresholds. Can J For Res 36:
368	1035–1046
369	Peters A (1996) The natural host range of Steinernema and Heterorhabditis spp. and their impact
370	on insect populations. Biocontrol Sci Technol 6:389–402
371	Poinar GO, Hom A (1986) Survival and horizontal movement of infective stage Neoaplectana
372	carpocapsae in the field. J Nematol 18:34–36
373	Preisser EL, Dugaw CJ, Dennis B, Strong DR (2005) Long-term survival of the entomopathogenic
374	nematode Heterorhabditis marelatus. Env Entomol 34:1501–1506
375	Pye AE, Burman M (1978) Neoaplectana carpocapsae: Infection and reproduction in large pine
376	weevil larvae, Hylobius abietis. Exp Parasitol 46:1–11
377	Rice WR (1989) Analyzing tables of statistical tests. Evolution 43:223-225
378	Shields EJ, Testa A, Miller JM, Flanders KL (1999) Field efficacy and persistence of the
379	entomopathogenic nematodes Heterorhabditis bacteriophora Oswego and H. bacteriophora
380	NC on Alfalfa Snout Beetle larvae (Coleoptera: Curculionidae). Environ Entomol 28:128-
381	136
382	Sippola AL, Siitonen J, Punttila P (2002) Beetle diversity in timberline forests: a comparison
383	between old-growth and regeneration areas in Finnish Lapland. Ann Zool Fenn 39:69–86

384	Smits PH (1996) Post-application persistence of entomopathogenic nematodes. Biocontrol Sci
385	Technol 6:379–388
386	Susurluk A, Ehlers R-U (2008) Field persistence of the entomopathogenic nematode

387 Heterorhabditis bacteriophora in different crops. BioControl 53:62
--

- Torr P, Heritage S, Wilson MJ (2007) Steinernema kraussei, an indigenous nematode found in 388 coniferous forests: efficacy and field persistence against Hylobius abietis. Agric For 389 Entomol 9:181–188 390
- Van Lenteren JC, Babendreier D, Bigler F, Burgio G, Hokkanen HMT, Kuske S, Loomans AJM, 391
- 392 Menzler-Hokkanen I, van Rijn PCJ, Thomas MB, Tommasini MG, Zeng QQ (2003)
- Environmental risk assessment of exotic natural enemies used in inundative biological 393
- 394 control. BioControl 48:3-38
- Williams CD, Dillon AB, Girling RD, Griffin CT (2013) Organic soils promote the efficacy of 395 entomopathogenic nematodes, with different foraging strategies, in the control of a major
- forest pest: A meta-analysis of field trial data. Biol Control 65:357-364 397

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Table 1: Coniferous clear-fell sites located in the Republic of Ireland with stumps treated with

407 Steinernema carpocapsae on a site-wide scale (i.e. all stumps treated). SS = Sitka Spruce, LP =

408 lodgepole pine. For N see Table 2.

	Dimensions						
Site		Site	of site	Year	Year of	Soil	Stump
number	Site	location	(m approx.)	Felled	application	type	species
		52°08'N					SS, LP
	Lackenrea	007°48'W	400 x 400	2005	2007	Mineral	(mixed
A1		53 m					stand)
		53°05'N					
	Glendine	007°34'W	500 x 400	2005	2007	Peat	LP
A2		458 m					
		52°18'N					
	Ballymacshaneboy	008°36'W	200 x 200	2005	2007	Mineral	LP
A3		311 m					
		52°12'N					
	Knockeen	007°10'W	200 x 100	2005	2007	Peat	SS
A4		79 m					
		53°09'N					
	Deerpark	006°12'W	300 x 300	2005	2007	Mineral	SS
A5		319 m					
		53°14'N					
	Featherbed	006°19'W	500 x 500	2007	2008	Mineral	SS
A6		361 m					
		52°18'N					
	Raheenkyle	008°34'W	250 x 150	2006	2008	Mineral	SS
A7		426 m					

Table 2: Results of soil and bark sampling on seven coniferous clear-fell sites treated with S. 410 carpocapsae to control H. abietis. Percentages indicate proportion of all soil cores from the site 411 that contained S. carpocapsae (cores), proportion of tree stumps sampled on a site with at least 412 413 one of the soil cores collected at the stump containing S. carpocapsae (stumps), proportion of sampled stumps with bulk bark samples containing S. carpocapsae (bulk) and proportion of 414 sampled stumps with S. carpocapsae detected under the bark (under). For sites A1 and A2, 415 values for a site within a column that share the same letter are not significantly different from 416 each other ( $\chi^2$  or Fisher's Exact test; for multiple comparisons on core and stump data, 417 Bonferroni  $\alpha = 0.017$ ). LP = lodgepole pine, SS = Sitka spruce. Site details in Table 1.p.a. = 418 post-application. 419

	Presence of S carnocansae						Presence of other		
			1	Steinernema sp					
Percentage (number)							Percentage (number)		
	Time p.a.	N (cores;	SO	il	b	oark			
Site	(years)	stumps)	cores	stumps	bulk	under	cores	stumps	
	0.42	320;20	2.2 (7)a	30 (6)a	-	-	0	0	
	1	240;15	3.3 (8)a	40 (6)a	7 (1)	7 (1)	0	0	
ALL	2	240;15	1.3 (3)a	13 (2)a	7 (1)	7 (1)	0	0	
	5	240;15	0	0	0	0	2.9 (7)	33 (5)	
	0.42	160;10	3.1 (5)a	40 (4)a	-	-	0	0	
ALSS	1	240;15	3.3 (8)a	53 (8)a	7 (1)	7 (1)	0	0	
111 55	2	240;15	0 b	0 b	0	0	0	0	
	5	240;15	0	0	0	0	0.8 (2)	13.3 (2)	
A21P	0.42	480;30	11.0 (53)a	77 (23)a	-	-	0	0	
A2 LP	1	480;30	13.5 (65)a	87 (26)a	67 (20)a	60 (18)a	0	0	

	2	480;30	5.2 (25)b	43 (13)b	37 (11)b	27 (8)b	0	0
	5	480;30	0	0	0	0	0	0
A3LP	2	480;30	2.9 (14)	37 (11)	40 (12)	33 (10)	0	0
	5	480;30	0	0	-	-	1.3 (6)	20.0 (6)
A4 SS	2	432;27	0.9 (4)	11 (3)	0	0	0	0
A5 SS	5	480;30	0	0	0	0	2.3 (11)	23.3 (7)
A6 SS	4	480;30	0	0	0	0	1.7 (8)	23.3 (7)
A7 SS	4	480;30	0	0	0	0	1.9 (9)	16.7 (5)

Table 3: Summary of S. carpocapsae presence in soil around stumps (proportion and number of
soil cores and stumps scoring positive) and under bark of stumps (proportion and number of bulk
bark samples scoring positive) in 2009, two years after application. Data for sites A1 to A4
pooled by soil type and stump species.

	Data pooled		soil		bulk bark
427	by		cores	stumps	samples
428	Soil type	Mineral	1.9 (17/920)	21.7 (13/60)	21.7 (13/60)
420		Peat	3.2 (29/912)	22.8 (13/57)	19.3 (11/57)
430	Tree species	Lodgepole Pine	3.5 (42/1200)	34.7(26/75)	32.0 (24/75)
431		Sitka Spruce	0.6 (4/672)	0 (0/42)	0 (0/42)

**Fig. 1** Scatterplots for the percentage of soil cores scoring positive for S. carpocapsae (a) or the percentage of stumps with bark samples scoring positive for S. carpocapsae (b) plotted against the  $\log_{10} (x+1)$  of the mean number of pine weevils emerging from untreated control stumps on sampled sites A1-A4 (spruce and pine from site A1 used separately). Black points show spruce sites, grey points show pine sites. Horizontal error bars give standard error of mean weevil emergence

Fig. 2 Presence of S. carpocapsae in soil cores with increasing distance from bole of sampled
stumps 2 years after S. carpocapsae application. Bars represent percentage of soil cores positive
for S. carpocapsae at each distance (data combined at each distance for sites A1-A4; total n =
468 cores for each distance).



