

Canterbury Research and Theses Environment

Canterbury Christ Church University's repository of research outputs

http://create.canterbury.ac.uk

Please cite this publication as follows:

Burman, J., Westerberg, L., Ostrow, S., Ryrholm, N., Bergman, Karl-Olof, Winde, I., Nyabuga, F. N., Larsson, M. and Milberg, P. (2016) Revealing hidden species distribution with pheromones: the case of Synanthedon vespiformis (Lepidoptera: Sesiidae) in Sweden. Journal of Insect Conservation, 20 (1). pp. 11-21. ISSN 1366-638X.

Link to official URL (if available):

http://dx.doi.org/10.1007/s10841-015-9835-9

This version is made available in accordance with publishers' policies. All material made available by CReaTE is protected by intellectual property law, including copyright law. Any use made of the contents should comply with the relevant law.

Contact: create.library@canterbury.ac.uk



Revealing hidden species distribution with pheromones: the case of Synanthedon vespiformis
(Lepidoptera: Sesiidae) in Sweden.
Joseph Burman ^{bc} , Lars Westerberg ^a , Suzanne Ostrow ^a , Nils Ryrholm ^d , Karl-Olof Bergman ^a , Inis Winde ^{be} ,
Franklin N. Nyabuga ^{be} , Mattias C. Larsson ^b , Per Milberg ^a
^a IFM Biology, Conservation Ecology Group, Linköping University, SE-581 83 Linköping, Sweden
^b Department of Plant Protection Biology, Swedish University of Agricultural Sciences, Box 102, SE-230 53
Alnarp, Sweden.
^c Ecology Research Group, Canterbury Christ Church University, Canterbury, Kent, England, CT1 1QU
^d University of Gävle, SE-801 76 Gävle, Sweden
^e Department of Biology, Lund University, Sölvegatan 37, 223 62, Lund, Sweden
Corresponding author – Dr. Joseph Burman
joseph.burman@canterbury.ac.uk
01227 767700 ext. 3104
(Office – Ht14) Canterbury Christ Church University, Canterbury, Kent, England, CT1 1QU
Acknowledgements
Stiftelsen Eklandskapet i Linköpings kommun, Marie-Claire Cronstedts Stiftelse, Swedish WWF, the Tranemåla
Foundation, Skogssällskapet, Region Skånes miljövårdsfond, SLU Partnerskap Alnarp, and the IC-E3 Linnaeus
grant (Formas, SLU) to the Division of Chemical Ecology at SLU provided grants for this project. Assistance
was provided by Stefan Ekroth (expert advice, field work and species determination), Henrik Nguyen
(pheromone bait and administrative support); Klas Andersson (field work); Tomas Burén and co-workers at
Kalmar Municipality, Anders Jörneskog at Linköping Municipality and Kjell Antonsson at the County
Administrative Board of Östergotland (various support). Thanks also go to David Heaver at Natural England for
comments on the research from a national monitoring perspective. Finally, a special thanks to all landowners
who gave their permission to set up traps, and their commitment to the historical legacy of their land as well as
its future management.

Abstract

Synanthedon vespiformis L. (Lepidoptera: Sesiidae) is considered a rare insect in Sweden, discovered in 1860, with only a few observations recorded until a sex pheromone attractant became available recently. This study details a national survey conducted using pheromones as a sampling method for this species. Through pheromone trapping we captured 439 specimens in Southern Sweden at 77 sites, almost tripling the number of previously reported records for this species. The results suggest that S. vespiformis is truly a rare species with a genuinely scattered distribution, but can be locally abundant. Habitat analyses were conducted in order to test the relationship between habitat quality and the number of individuals caught. In Sweden, S. vespiformis is thought to be associated with oak hosts, but our attempts to predict its occurrence by the abundance of oaks yielded no significant relationships. We therefore suggest that sampling bias and limited knowledge on distribution may have led to the assumption that this species is primarily reliant on oaks in the northern part of its range, whereas it may in fact be polyphagous, similar to S. vespiformis found as an agricultural pest in Central and Southern Europe. We conclude that pheromones can massively enhance sampling potential for this and other rare Lepidopteran species. Large-scale pheromone-based surveys provide a snapshot of true presences and absences across a considerable part of a species national distribution range, and thus for the first time provide a viable means of systematically assessing changes in distribution over time with high spatiotemporal resolution.

Keywords: Ecology, saproxylic, moth, indicator of species richness, conservation, monitoring.

Introduction

Woodland habitats have undergone significant anthropogenic change in recent centuries, giving way to land use focused on agriculture and housing development (Eliasson & Nilsson, 2002). As well as habitat fragmentation, habitat alteration has also been an issue; whereby open, sunny woodland habitats have been transformed into shady, overgrown habitats less suitable for species dependent on sunlight (Kirby *et al.*, 2005). One such light-dependent species associated with oak woodland habitat and the focus of this paper is the clearwing moth *Synanthedon vespiformis* (L.) (Lepidoptera: Sesiidae), first recorded in Sweden in 1860 (Eliasson, 2007). This species is classified as Vulnerable on the Swedish Red List (Gärdenfors, 2010) and considered 'Nationally Scarce' in the United Kingdom (Greatorex-Davies *et al.*, 2003). The moth could potentially serve as a good indicator for the increasingly rare open woodland habitats. The location and distribution of clearwing moth populations are however difficult to assess, as the species of this family are relatively inconspicuous and frequently mistaken for members of the hymenoptera due to their mimicry (Duckworth & Eichlin, 1974). As a result, knowledge of their true distribution and ecology remains relatively limited.

S. vespiformis is a particularly interesting example of a saproxylic clearwing, as its habitat preferences seems to vary significantly across its geographical range in Europe. In the southern part of its range it is thought to be fairly polyphagous and is in fact considered a pest, attacking numerous tree species including beech (Fagus silvatica L.), oak (Quercus spp.), poplar (Populus spp.), willow (Salix spp.) and fruit crops of blackberry (Rubus fruticosus L.), raspberry (Rubus idaeus L.) and peach (Prunus persica L.) (Szántóné-Veszelka et al., 2010). However, in the northern part of its range where the species is considered threatened and does not occur as an

77 agricultural pest, the primary host of the larvae is assumed to be Quercus robur L. (pedunculate oak) with some 78 suggestions of members of the Rosaceae and Betula spp. as secondary hosts (Eliasson, 2002; Waring & 79 Townsend, 2003). However, the evidence is predominantly anecdotal or based on a small number of 80 observations. Thus a better understanding of its ecology will aid in conservation of this species, particularly in 81 places where it is considered to be in decline. These conservation efforts first require effective and accurate 82 sampling and monitoring methods of the target species, something which has been identified as lacking in 83 modern biodiversity management (Rademaekers et al., 2010). 84 Identifying, protecting and monitoring key areas or habitat types that support a high number of rare or 85 threatened species is essential in conservation (Henle et al., 2013). However, broad systematic surveys are 86 expensive, time-consuming and often reliant on a small number of taxonomic experts (e.g. Horak & Pavlicek 87 2013). Saproxylic habitats in particular are significantly more difficult and expensive to monitor using standard 88 methods, with the number of site visits for establishing species assemblage being at least twice that of 89 equivalent grassland habitats (pers. comm. David Heaver, Natural England). Thus for determining species 90 distribution, systematic surveys of this nature risk generating expensive data of relatively low accuracy and 91 precision. 92 As a result, bioindicator species are often used as a proxy in place of complete surveys in order to locate sites 93 with high species richness and/or conservation value (Fleishman & Murphy, 2009). The most commonly used 94 indicators of terrestrial biodiversity are butterflies, grasshoppers and wild bees, mainly for open environments 95 like grasslands (Rosenberg et al., 1986; Nilsson et al., 1995; Bazelet & Samways, 2011, 2012; Bommarco et al., 96 2012; Gerlach et al., 2013). Bioindicator selection for assessing the quality of woodland or forest habitats is 97 however heavily skewed towards saproxylic beetles due to their prevalence (Grove, 2002), meanwhile 98 Lepidoptera are under-represented in these habitats despite their potential for assessing human impacts on 99 biodiversity (Fiedler & Schultze, 2004). Some woodland moths are rather inconspicuous and are often under-100 reported in survey data despite being sampled regularly (Quinto et al., 2013; Jonason et al., 2013, 2014), 101 possibly due to their predominantly nocturnal behavior, and sampling bias from recorders (Dennis & Thomas, 102 2000). Thus an effective method of standardized recording for these potential bioindicators is highly desirable. 103 In order to provide greater levels of accuracy and sampling power at a lower cost, insect pheromones have been 104 suggested as a supplement to existing sampling methods for insects (Larsson et al., 2003; Tolasch et al., 2007; 105 Larsson et al., 2009; Harvey et al., 2010; Millar et al., 2010; Musa et al., 2013; Andersson et al., 2014). Given 106 their widespread availability already in pest management for Lepidoptera and particularly clearwing moths 107 (Braxton & Raupp, 1995), these tools could be redirected to provide great benefits to biodiversity monitoring of 108 saproxylics. Pheromone monitoring systems in insects are generally species-specific once optimized, although 109 cross-attraction exists, e.g. in some other Lepidoptera (Löfstedt et al., 1991) and some groups of saproxylic 110 beetle species (Hanks et al., 2012). Regardless, this selectivity would be advantageous for a focus on defined 111 indicator species, whilst ultimately a guild of bioindicators would be desirable in order to reflect a wider range 112 of microhabitats within a system. In the present study, we carried out pheromone monitoring of one potential 113 indicator, the clearwing moth S. vespiformis, whose pheromone system has recently been characterized (Levi-114 Zada et al., 2011).

The first general aim of the present study was to establish whether pheromone monitoring could provide more accurate information about the species' distribution than standard monitoring practices. Secondly, we wanted to establish whether the apparent rarity of the species in Sweden was a real phenomenon, or simply a result of poor detectability. Additionally, to conserve threatened species, knowledge of the species' habitat requirements is essential. The type of habitat a species uses can be found by relating particular habitat elements to species abundances or occurrences, with quantity of habitat required being assessed at multiple scales (Bergman *et al.*, 2012; Musa *et al.*, 2013). Once the required type and quantity of that resource has been ascertained, an assessment of areas in the landscape suitable for a species is possible. The third aim of this study was therefore to establish the preferred habitat for *S. vespiformis* by correlating its abundance with habitat characteristics and, by extension, to establish whether it could be used as an indicator for the biodiversity potential of oakdominated open woodlands with which it has traditionally been associated.

Materials & methods

Site selection

The study was conducted in the counties of Östergötland, Skåne, Blekinge and Kalmar where a number of sites with high density of old and/or hollow oaks were selected (Fig.1). These sites generally have high species richness (Nilsson *et al.*, 1995) but have suffered from severe decline and fragmentation over the last 200 years in Sweden, mainly due to the change in ownership of the oaks and shifts in farming and forestry practices (Eliasson & Nilsson, 2002). Included in the selection were a number of sites with lower proportions of oaks, and a higher number of other hardwood species and hollow trees for comparison, covering a total of 251 sites (numbers of traps per county are shown in table 1). Sites selection was also based on a minimum separation distance of 500m between traps, the closest two traps being separated by 512m in this study. In this survey, a total of eight traps had another trap within a 1000m radius, with the majority being separated by many kilometres. Our observations on the attractive range of similar moth pheromones in mark release recapture studies suggest that males may be able to detect lures at a maximum distance of 150-200m (unpublished data). Therefore these distances were maintained in order to rule out the effect of inter-trap competition.

Historical records of S. vespiformis

Pre-existing records were taken from Sweden's nationwide repository for animal, fungi and plant distribution data, the "Swedish Species Gateway" (www.artportalen.se). In order to compare historical records with our own more recent survey efforts, we used data dating back to 1976 when the first database entry for this species occurred from a site in Stockholm. These sightings (often recorded using standard observational methods such as larval/pupal searches) (Fig.1a, b) were then compared to recent inventories we carried out using pheromone lures in 2011-2012 (Fig.1c, d). Artportalen also included a small number of historical pheromone-based sightings between 2005 and 2013 which were excluded from our analyses, but are shown in Figure 1.

Biology of S. vespiformis

Very little is known about the biology of *S. vespiformis*, although it is considered to be saproxylic; feeding on the cortex of its various suggested host plants (Levi-Zada *et al.*, 2011). The adult moth is considered to have a

'moderate' flight range (Van der Meulen & Groenendijk, 2005) taking flight in the afternoon through into the early evening when male moths can be caught by pheromone lure (Levi-Zada *et al.*, 2011). In Sweden the flight period begins in the last week of June and can last until the first week of August, with peak activity taking place in the second week of June in both northern and southern counties (Artportalen, 2015). This species is also known to have an association with the gall-inducing bacterium *Agrobacterium tumefaciens*, which likely facilitates larval feeding in some host plants (Audemard & Vigouroux, 1982).

Pheromone lures

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

We used a modified version of the pheromone blend for *S. vespiformis*, that has been found to be more attractive to Swedish populations than the blend from Levi-Zada *et al.*, (2011) (Ryrholm *unpublished*). Pheromone lures were produced using a pre-prepared blend of four pheromone components dissolved in hexane. The blend was prepared so that 200µl hexane solution contained 300µg E3Z13-18:Ac, 90µg E3Z13-18:OH, 30µg Z3Z13-18:Ac and 3µg E2Z13-18:OH. A blend volume of 200µl was then pipetted onto the surface of a 20mm diameter grey rubber septum (PheroNet, Sweden) and left in a fume cupboard to allow the solvent to fully evaporate. Septa were subsequently stored in the freezer until further use in order to preserve the attractiveness of the lure.

Trap methodology

The whole rationale of the present study was to use a viable and consistent sampling effort to obtain the first semi-quantitative data on presence and abundance across a large number of sites over a wide geographical range, in contrast to the scattered records previously available. For this purpose, we considered sticky traps as the best available option. Lures were hung 2 cm from the roof of a clear plastic delta trap sourced from CSalomon pheromone traps, Budapest, Hungary. Instead of the original sticky bottom inserts, we used cardboard sticky inserts from Oecos ltd, UK, which preserved the morphology of the wings more effectively for subsequent species identification. Studies on destructive sampling have shown that insect populations are robust against lethal sampling methods even when multiple killing traps are used per hectare (Haniotakis & Koutroubas, 1999; Yamanaka et al., 2001; Gezon et al., 2015). Although we did not expect our sampling to affect the populations sampled significantly, we used a reduced sticky area in our traps (about 50% compared to their original size, or 80cm²) to reduce catches. This was a precaution considering that we were working with a rare species and a pheromone of unknown attractiveness. At each site, one trap was placed 1 - 2 m above ground from a nearby tree. Tree selection was based on availability of trees and not individual tree species. Traps were placed throughout the ten day period which commenced on 1st July 2012, and were brought down in a ten day period after 6th August 2012 ensuring that all traps had been placed in the field for a minimum of four weeks. Traps were also left for four weeks during July of 2011 (for a small pilot study in the Västervik region), with the majority of the survey work being carried out in 2012. Subsequent catches were identified and recorded after traps and sticky inserts were removed at the end of this sampling period. Only data from 2012 were used for subsequent habitat analyses.

Statistical analyses

In order to test the relative efficacy of pheromone lures compared to standard historical methods logged on Artportalen, a Kruskal-Wallis test was performed on the catch abundance data. The test was carried out in order 189 significantly different from the median number of catches made through the use of a pheromone lure. 190 The study of habitat characteristics was carried out only in Östergötland, which had the largest proportion of 191 occupied sites. This county also had the largest and most consistent set of tree data in Sweden whilst tree data 192 were partially incomplete in other counties. Thus the 102 sites in this region were deemed most suitable for 193 study of habitat characteristics preferred by S. vespiformis. Tree data used for this study were derived from the 194 most recent survey, which was a 10 year old survey of the region commissioned by the County Administration 195 Board of Östergötland. From these data, tree groups were categorized as oaks or non-oaks in order to test the 196 hypothesis that S. vespiformis is associated more strongly with Quercus spp. in this northern part of its range. 197 European aspen (Populus tremula L.) and silver birch (Betula pendula Roth.) were excluded from the study due 198 to incomplete survey data as well as the following coniferous trees: Norway spruce (Picea abies (L.) H.Karst), 199 larch (Larix sp.), common juniper (Juniperus communis L.), and scots pine (Pinus sylvestris L.). 200 Additionally, trees were classed by a further six groupings as follows; (i) all trees, (ii) trees > 450 cm 201 circumference, (iii) hollow stage > 3 (where stages 1 - 3 have no significant hollows and 4 - 7 have hollows of 202 increasing size categories larger than 10cm in diameter) (Claesson & Ek 2009), (iv) trees located in open areas, 203 (v) trees > 450 cm circumference located in grasslands, (vi) hollow stage > 3 located in grasslands. These 204 groupings would allow us to further determine preferences for different types of oak woodland/semi pasture. A 205 national database on semi-natural grasslands (TUVA) was used to locate targeted trees situated in semi-natural 206 grasslands. The "open areas" category included targeted trees with open canopy cover and open surrounding 207 vegetation. Further explanations of the canopy cover and surrounding vegetation categories can be found in 208 Claesson & Ek (2009). 209 Since saproxylic insect populations can respond to a wide range of geographical scales (Bergman et al. 2012), 210 28 radii, ranging from 30 m to 6,000 m (Fig.2), were used to calculate tree densities around each site (Quantum 211 GIS 1.8.0-Lisboa 2013). The reasoning behind selecting 30 m as the minimum scale was that it needed to be 212 small while still maintaining variation in tree densities among sites. The maximum scale of 6,000 m was 213 selected based on the maximum dispersal range of other moths as well as to retain enough sites without the 214 largest radii overlapping. 215 The scale(s) at which the species responds most strongly to the habitat variables (characteristic scale of 216 response) was estimated for each tree group separately. At each scale, a negative binomial general linear model 217 was run with abundance of S. vespiformis as response and tree counts as predictor variable. The computer 218 program Focus (Geomatics and Landscape Ecology Research Lab, Canada) was used to extract 500 219 combinations of non-overlapping buffers at each radius: no radii overlapped below 500m. The median of the 220 regression results were used to condense the results for one scale and tree group. The characteristic scale of 221 response for a tree group was defined as the scale with the largest absolute Z-value. It is possible to use other 222 variables, and rings instead of circle as buffers, to determine the characteristic scale of response. Tree content in 223 ring buffers are less correlated between scales, and regression coefficients on standardized explanatory variables 224 make comparisons of effect size easier. However, the connection between response to circle buffers and 225 ecological processes is easier to understand. Furthermore, the interpretation of standardized coefficients need a

to determine whether the median number of individuals caught by standard methods from historical survey was

188

226 measure of variance which the Z-value already provides. Results from ring buffers were also similar to circle buffers and standardized coefficients scaled with Z-value, thus the alternatives did not change the interpretation. 228 There was generally a weak correlation between oak and non-oak trees which was negative (r = -0.1) for small 229 radii and positive for larger radii (r = 0.1) while correlations within tree-groups of oak or non-oak were higher 230 and positive. Running multiple correlations reduces the pseudoreplication of data points because each iteration uses spatially independent sites (Holland et al. 2004). However, the repeated analysis uses the same data so 232 pseudoreplication may not be entirely avoided. The Focus program therefore allows for optimization of the data 233 available and an increase in the power of the analysis. All analysis and data management was done in the 234 statistical software R (R core team, 2014). **Results** 236 In total 439 individual specimens of S. vespiformis were caught across the four counties, accounting for 77 237 newly identified localities (Fig.1c, d). We also recorded a total of 174 sites where the species was not found. 238 The breakdown of records per county is shown in Table 1. The sampling effort from this study almost tripled the 239 number of reported localities for this species during only two field seasons (47 previously reported sites found in 240 Artportalen were increased to 124 occupied sites in total as a result of sites located by pheromone lure), after decades of reports by classical methods of surveying (Fig. 3). The result is a slight apparent expansion of S. 242 vespiformis range due to increased sampling effort/accuracy, and a much higher density of occupied sites noted 243 within the existing range (Fig.1). 244 The new inventory of the species revealed a scattered pattern of distribution amongst woodland habitats sampled 245 in southern Sweden. In some cases moths were locally abundant, but their absence is notable across significant 246 areas of most counties despite the extensive and systematic sampling effort and high efficacy of the traps. Comparison of the median number of individuals caught by standard vs. pheromone methods however showed 248 significantly higher numbers to be observed via pheromone trapping overall (H = 18.58 d.f. = 1 p < 0.001). 249 The median catch using standard method was one individual, compared to a median of two individuals for 250 pheromone method. Habitat analyses in Östergötland (Fig.4), showed no significant relationship between the numbers of individuals caught in traps, with any of the habitat variables included at any scale. No absolute median Z-values were larger 253 than 1.96 which is the 5% significance level. Focusing on effect size there was a relatively larger positive response to hollow oaks close to the trap (30-200m), while other tree species in general, and hollow trees in 255 particular, seemed to have a negative effect on species presence. The reversed relationship between oak and 256 non-oak was not a result from explanatory variables being highly correlated at lower scales (r was approximately -0.1 between 30 and 200 m). 258 In addition to catches of S. vespiformis, some traps in Skåne and Östergötland also caught another clearwing

227

231

235

241

247

251

252

254

257

259

260

261

species, Pennisetia hylaeiformis Laspeyres. This species was present at fifteen sites in Skåne and nine in

Östergötland, with the number of individuals totaling 128 and 23 across all traps in these regions respectively.

Discussion

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

S. vespiformis is considered a rare insect in Sweden, first recorded in 1860, and with just a few observations logged until the sex pheromone was identified and its synthetic constituents became available more recently. To our knowledge, our approach to addressing this situation represents the first systematic application of pheromones for any regional survey of a lepidopteran conservation target species, although large-scale pheromone surveys have previously targeted pests among the Lepidoptera (e.g., Tobin et al., 2007) and other orders, and also the threatened click beetle Elater ferrugineus (Kadej et al., 2015). This study shows that pheromone-based monitoring of woodland moths can provide a significant increase in accuracy and sampling effort of surveys targeting saproxylic insects. By use of sex pheromones an additional 77 sites with S. vespiformis were found; thereby increasing the known localities from 47 to 124. This represents a near tripling of the number of reported sites in just two field seasons. In addition, the number of individuals caught represents a significant increase in sampling efficiency. Previous site visits often report single insects, whereas we uncovered a total of 439 individuals across our survey sites, with the highest number of individuals at one site being 41 (a site in Västervik municipality, Kalmar county). This increased efficiency is further supported by a significant difference noted in the median number of insects caught when comparing recording based on observation to recording using a pheromone lure. A minor limitation which was noted during this study was the tendency for this pheromone blend to cause crossattraction to Pennisetia hylaeiformis; a similar looking sesiid which could potentially lead to misidentification of S. vespiformis sites. It was also noted that some potential predation had occurred in traps left for a period of time longer than a few weeks, which may have even led to an under-sampling at certain sites. Thus is suggested that for the inventory of this species, voucher specimens should be retained for taxonomic determination, and traps checked at regular intervals where possible. We have not yet studied the specific accuracy of our pheromone lures in detecting populations of S. vespiformis, e.g. through recapture experiments or repeated sampling of known populations, but we would argue that empty pheromone traps in our study usually represent truly unoccupied sites, lacking reproducing populations. The generally high efficiency of these types of classical female-produced sex pheromones in detecting their target species has been well established in lepidopteran pest species (Zhang & Schlyter 1996) and in many other insect groups (Östrand & Anderbrant, 2003), including the threatened click beetle E. ferrugineus (Svensson et al., 2011; Zauli et al., 2014). In most cases it appears highly unlikely that a monitoring trap would fail to detect any males over a full flight season at a site with a local reproducing population. Absence data generated from sex pheromone lures have been shown to be much more reliable than stochastic methods such as unbaited pitfall or window traps in establishing presence or absence (Andersson et al., 2014; Zauli et al., 2014). The data generated by this methodology can therefore allow for better resolution in ecological studies, which rely strongly on accurate presence and absence data to characterize not only the species' habitat, but also its 'nonhabitat'. Future studies of the use of pheromones would benefit from a full assessment of their monitoring accuracy in order to assess exactly how precise moth pheromone systems are for determining absence, similar to studies on emerging eDNA technology for species assessment of great crested newt (Biggs et al., 2015).

Under-reporting of occupancy is a significant issue in biodiversity monitoring where sampling methods show low detection probability (Pellet & Schmidt, 2005), but this could be easily resolved with the high levels of detectability that pheromones provide (Andersson et al., 2014). The previous lack of occupancy records are likely contributors to our view of S. vespiformis as a species with a very sparse distribution in Sweden. Our findings suggest this species is perhaps a little less rare than previously thought, and occasionally abundant locally despite notable long-term declines in the type of Swedish oak forest considered to be its primary habitat. The slight expansion of the species range, and higher density of known sites within its range seen in Figure 1 is likely to be reflective of increased sampling efficacy rather than actual recent expansion of the species itself. However, the distribution of S. vespiformis is still relatively sparse, with a large proportion of sites where the species was absent despite an apparent abundance of saproxylic resources, suggesting the species may still be under threat or that very little is known about its ecological requirements. During the course of the study, S. vespiformis was found at a number of sites outside of its 'expected' habitat type. Singleton catches outside of the expected habitat may be a result of chance migrants caught nearby a 'true' locality. However, sampling bias has also been highlighted as a limiting factor in insect surveying particularly with Lepidoptera (Dennis et al., 1999), and this may contribute towards the oak-biased recordings seen in previous survey data. Resource-limited county administrative boards and field entomologists can tend to focus their efforts on known sites, or sites similar to those where the species has been found already. This circular approach may obscure hidden biodiversity and niches that might otherwise be uncovered by a less biased search effort in the field. Our analyses of S. vespiformis' habitat further highlight this sampling bias when considering S. vespiformis' preferences in northern Europe. In Sweden, the species is considered to be reliant on oaks (Waring & Townsend, 2003; Eliasson, 2007), but our results showed only weak relationships with old oaks and oaks with holes for number of moths caught and oak and non-oak tree abundance (Fig.4). This is in stark contrast to results from the click beetle E. ferrugineus using similar methodology in relation to similar veteran tree habitats, which demonstrated a very strong association between the beetle and its nominal habitat (Musa et

al. 2013). This casts some doubt on the previous assumptions made regarding host specificity and preference in
S. vespiformis. Our findings were also echoed in a recent Swedish inventory, where it was noted that

"Pheromone lures placed on several premises in Skåne and Öland and in eastern Småland have shown that the species is not as tied to large veteran trees as previously assumed, but is reliant on this habitat when others are

not available." (Palmqvist 2014).

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

325

326

327

328

329

330

331

332

333

334

335

336

337

In the south of its range *S. vespiformis* is found to be rather polyphagous, yet its visibility in these regions is undoubtedly much higher due to its high population numbers in agricultural crops (Levi-Zada *et al.*, 2011), and thus the likelihood of observing the species on alternative hosts is higher. Based on these findings we suggest that Swedish *S. vespiformis* could be more polyphagous than previously expected, possibly in line with its southern siblings. This hypothesis could be further tested by using pheromones to survey in regions significantly outside of its known range (for example Småland), in habitats which are not typical oak woodland and contain higher numbers of "secondary" host species. Incidentally, a recent study on saproxylic beetles also showed that most were polyphagous (Milberg *et al.*, 2014), and the presumed oak-specificity was most likely due to sampling bias. It is therefore suggested that sesiids would benefit from a significant re-inventory in order to assess their true host relationships and conservation status.

In principle, the methods applied here could encompass all previous observations and attempts at surveying S. vespiformis in Sweden and surpass them several times over, in a single season, with comparably little effort. The high quality and potentially low bias of data generated by pheromone monitoring is also important, as knowledge on species distribution needs to inform decision making at both a local, national and international level (Pereira & Cooper, 2006). This increased sampling efficacy has the potential to solve a number of issues highlighted by the European Commission in their efforts to mitigate biodiversity loss by 2020. Firstly, it has been suggested that one of the major challenges in meeting this target is development of effective and standardized methods of monitoring for national or international biodiversity (Pereira & Cooper, 2006; Henry et al., 2008). Additionally such international monitoring efforts must be viable within a restricted pool of resources (Bates et al., 2007), since cost effectiveness is a significant consideration in any environmental monitoring system (Hauser et al., 2006; Lovett et al., 2006). In light of the reduced or unstable funding for conservation/biodiversity monitoring in many member states (Lindenmayer et al., 2012), pheromone attractants could provide an essential tool for insect inventories across Europe. Not only are the pheromones relatively cost effective (costs are relatively low for these simple compounds where synthesis method is already established), but the relative sampling effort in terms of working hours for survey is also greatly reduced (Burman & Thackery, unpublished data). This is particularly promising, as time is also a limiting resource in biodiversity monitoring (Yoccoz et al., 2001). Pheromones could allow those with limited resources the opportunity to spread their efforts further afield, and thereby improve the quality of data obtained. We demonstrate in this study that national level insect species surveys are feasible using sex pheromones, covering large spatial and potentially temporal scales to provide data relevant to international monitoring (Pereira & Cooper, 2006). Largescale pheromone-based surveys provide a snapshot of presences and absences across a considerable part of a species national distribution range, and thus for the first time provide a viable means of systematically assessing changes in distribution over time with high spatiotemporal resolution.

In the present study destructive monitoring of *S. vespiformis* was a prerequisite for obtaining comparable semiquantitative occupancy data with high resolution over an extensive geographical range, and was carried out in agreement with conservation authorities although neither the species nor many of the sites are protected. The species is believed to generally have a two-year life cycle in Sweden (Eliasson, 2007), which would mitigate any risk to individual populations. Whilst destructive monitoring of the kind carried out in this study should be avoided year on year, it does provide a new snapshot of species distribution previously unavailable to conservationists. This includes a significant number of previously unreported sites for this species, many of which receive no statutory protection or management at the time of writing, but which could now be considered for their conservation value. Repeated future monitoring of these sites over extended time intervals perhaps combined with more regular use of pheromone live-trapping at a limited number of sites, could provide regular insights into how these habitats are faring in the longer term in response to different management regimes.

Conclusions

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

In this study we have shown that hidden biodiversity and species preferences can be substantial due to sampling bias and less effective sampling methodology. The vast majority of species sampled by standard observational methods are likely to be underreported, and as a result the drop in data quality can lead to problems in conducting ecological studies to uncover true habitat characteristics (Pellet & Schmidt, 2005). By comparison,

- 377 pheromone-based methods can significantly increase the sampling accuracy and give a much more reliable idea
- of a species' distribution. S. vespiformis appears to be genuinely quite rare still, despite a significant number of
- new sites being located. In addition, the increased resolution of data, showed no particular association for large
- oaks, despite previous assumptions.
- We believe as a result, that pheromone-baited traps bring much promise for conservation, both for surveying
- and monitoring targeted species, and could be used as a powerful tool to achieve the EU's optimistic goals of
- halting insect biodiversity loss by 2020, as well as an invaluable resource for carrying out landscape level
- 384 ecological studies.

385 References

- Andersson, K., Bergman, K.O., Andersson, F., Hedenström, E., Jansson, N., Burman, J., Winde, I., Larsson,
- M.C. and Milberg, P. (2014) High-accuracy sampling of saproxylic diversity indicators at regional scales with
- pheromones: The case of *Elater ferrugineus* (Coleoptera, Elateridae). Biol Conserv, 171, 156-166.
- 389 Artportalen (2015) Swedish Species Gateway. http://www.artportalen.se/ 3rd November 2015.
- 390 Audemard, H., & Vigouroux, A. (1982) Une curieuse association parasitaire sur Pecher: Sesie (Synanthedon
- 391 *vespiformis*) et tumeur bacterienne du collet (*Agrobacterium tumefaciens*). Phytoma.
- Bates, C.R., Scott, G. & Tobin, M. (2007) Weighing the costs and benefits of reduced sampling resolution in
- biomonitoring studies: perspectives from the temperate rocky intertidal. Biol Conserv, 137, 617-625.
- Bazelet, C.S. & Samways, M.J. (2011) Identifying grasshopper bioindicators for habitat quality assessment of
- ecological networks. Ecol Indic, 11, 1259-1269.
- Bazelet, C.S. & Samways, M.J. (2012) Grasshopper and butterfly local congruency in grassland remnants. J
- 397 Insect Conserv, 16, 71-85.
- Bergman, K.-O., Jansson, N., Claesson, K., Palmer, M.W. & Milberg, P. (2012) How much and at what scale?
- 399 Multiscale analyses as decision support for conservation of saproxylic oak beetles. Forest Ecol Manag, 265,
- 400 133-141.
- 401 Biggs, J., Ewald, N., Valentini, A., Gaboriaud, C., Dejean, T., Griffiths, R. A., Foster, J., Wilkinson, J.W.,
- 402 Arnell, A., Brotherton, P., Williams, P. & Dunn, F. (2015). Using eDNA to develop a national citizen science-
- based monitoring programme for the great crested newt (Triturus cristatus). Biol Conserv, 183, 19-28.
- Bommarco, R., Lundin, O., Smith, H.G. & Rundlöf, M. (2012) Drastic historic shifts in bumble-bee community
- 405 composition in Sweden. P Roy Soc B-Biol Sci, 279, 309-315.
- Braxton, S. M. & Raupp, M. J. (1995) An annotated checklist of clearwing border pests of ornamental plants
- trapped using commercially available pheromone lures. J. Arboriculture, 21, 177-180.
- Claesson, K., & Ek, T. (2009). Skyddsvärda träd i Östergötland 1997-2008. Länsstryrelsen Östergötland. 1-12.

- Dennis, R.L.H., Sparks, T.H. & Hardy, P.B. (1999) Bias in butterfly distribution maps: the effects of sampling
- 410 effort. J Insect Conserv, 3, 33-42.
- Dennis, R.L.H. & Thomas, C.D. (2000) Bias in butterfly distribution maps: the influence of hot spots and
- recorder's home range. J Insect Conserv, 4, 73-77.
- Duckworth, W.D. & Eichlin, T.D. (1974) Clearwing moths of Australia and New Zealand (Lepidoptera:
- 414 Sesiidae). Smithsonian Institution Press, USA.
- Eliasson, P. & Nilsson, S.G. (2002) 'You should hate young oaks and young noblemen': the environmental
- 416 history of oaks in eighteenth-and nineteenth-century Sweden. Environ Hist, 7, 659-677.
- 417 Eliasson, C.U. (2007) Synanthedon vespiformis. Fact sheet. ArtDatabanken, Swedish University of Agricultural
- 418 Sciences, Uppsala.
- Fiedler, K., & Schulze, C.H. (2004) Forest modification affects diversity (but not dynamics) of speciose tropical
- 420 pyraloid moth communities. Biotropica, 36, 615-627.
- Fleishman, E., & Murphy, D.D. (2009). A realistic assessment of the indicator potential of butterflies and other
- 422 charismatic taxonomic groups. Conserv Biol, 23, 1109-1116.
- Gärdenfors, U. (2010) The 2010 Red List of Swedish Species. Artdatabanken, SLU, Uppsala, Sweden.
- 424 Gerlach, J., Samways, M. & Pryke, J. (2013) Terrestrial invertebrates as bioindicators: an overview of available
- 425 taxonomic groups. J Insect Conserv, 17, 831-850.
- 426 Gezon, Z.J., Wyman, E.S., Ascher, J.S., Inouye, D.W. & Irwin, R.E. (2015). The effect of repeated, lethal
- sampling on wild bee abundance and diversity. Met Ecol Evol
- 428 Greatorex-Davies, N., Sparks, T., & Woiwod, I. (2003). Changes in the Lepidoptera of Monks Wood NNR. Ten
- years of change: Woodland research at Monks Wood NNR, 90.
- 430 Grove, S.J. (2002) Saproxylic insect ecology and the sustainable management of forests. Annu Rev Ecol Syst,
- 431 33, 1-23.
- Haniotakis, G.E., Koutroubas, A., Sachinoglou, A. & Lahlou, A. (1999). Studies on the response of the leopard
- moth, Zeuzera pyrina I (Lepidoptera: Cossidae) to pheromones in apple orchards. IOBC wprs Bulletin, 22, 105-
- 434 114.
- Hanks, L.M., Millar, J.G., Mongold-Diers, J.A., Wong, J.C., Meier, L.R., Reagel, P.F. & Mitchell, R.F. (2012)
- 436 Using blends of cerambycid beetle pheromones and host plant volatiles to simultaneously attract a diversity of
- 437 cerambycid species. Can J Forest Res, 42, 1050-1059.
- Harvey, D.J., Hawes, C.J., Gange, A.C., Finch, P., Chesmore, D. & Farr, I. (2010) Development of non-invasive
- monitoring methods for larvae and adults of the stag beetle, Lucanus cervus. Insect Con Diver, 4, 4-14.

- Hauser, C.E., Pople A.R. & Possingham H.P. (2006) Should managed populations be monitored every year?
- 441 Ecol Appl, 16,807-819.
- Henle, K., Bauch, B., Auliya, M., Külvik, M., Pe'er, G., Schmeller, D.S. & Framstad, E. (2013) Priorities for
- biodiversity monitoring in Europe: A review of supranational policies and a novel scheme for integrative
- prioritization. Ecol Indic, 33, 5-18.
- Henry, P.Y., Lengyel, S., Nowicki, P., Julliard, R., Clobert, J., Čelik, T., Gruber, B., Schmeller, D.S., Babij, V.
- 446 & Henle, K. (2008) Integrating ongoing biodiversity monitoring: potential benefits and methods. Biodivers
- 447 Conserv, 17, 3357-3382.
- Horák, J., Vodka, Š., Pavlíček, J., & Boža, P. (2013). Unexpected visitors: flightless beetles in window traps. J
- 449 Insect Conserv, 17, 441-449.
- Holland, J.D., Bert, D.G. & Fahrig, L. (2004). Determining the Spatial Scale of a Species' Response to Habitat.
- 451 BioScience, 54, 227-233.
- Jonason, D., Franzén, M., & Pettersson, L.B. (2013) Transient peak in moth diversity as a response to organic
- 453 farming. Basic Appl Ecol, 14, 515-522.
- Jonason, D., Franzén, M. & Ranius, T. (2014) Surveying Moths Using Light Traps: Effects of Weather and
- 455 Time of Year. PloS one, 9, e92453.
- Kadej, M., Zajac, K., Ruta, R. Gutowski, J.M., Tarnawski, D., Smolis, A., Olbrycht, T., Malkiewicz, A.,
- 457 Myskow, E., Larsson, M.C., Andersson, F. & Hedenström, E. (2015) Sex pheromones as a tool to overcome the
- Wallacean shortfall in Conserv Biol: a case of *Elater ferrugineus* Linnaeus, 1758 (Coleoptera: Elateridae). J
- 459 Insect Conserv, 19, 25-32.
- 460 Kirby, K.J., Smart, S.M., Black, H.I.J., Bunce R.G.H., Corney, P.M. & Smithers, R.J. (2005) Long-term
- Ecological Change in British Woodlands (1971-2001): A Re-survey and Analysis of Change Based on the 103
- Sites in the Nature Conservancy 'Bunce 1971' Woodland Survey. English Nature.
- Larsson, M.C., Hedin, J., Svensson, G.P., Tolasch, T. & Francke, W. (2003) Characteristic odor of Osmoderma
- 464 *eremita* identified as a male-released pheromone. J Chem Ecol, 29, 575-587.
- Larsson, M.C., Svensson, G.P. & Ryrholm, N. (2009) Monitoring rare and threatened insects with pheromone
- 466 attractants. In: Samways MJ, New T, McGeoch M (eds) Insect Conservation: A Handbook of Approaches and
- Methods. Oxford University Press, Oxford, UK, pp.114-116.
- Levi-Zada, A., Ben-Yehuda, S., Dunkelblum, E., Gindin, G., Fefer, D., Protasov, A., Kuznetsowa, T., Manulis-
- 469 Sasson, S. & Mendel, Z. (2011) Identification and field bioassays of the sex pheromone of the yellow-legged
- dearwing Synanthedon vespiformis (Lepidoptera: Sesiidae). Chemoecology, 21, 227-233.
- Lindenmayer, D.B., Gibbons, P., Bourke, M.A.X., Burgman, M., Dickman, C.R., Ferrier, S., Fitzsimons, J.,
- Freudenberger, D., Garnett, S.T., Groves, C., Hobbs, R.J., Kingsford, R.T., Krebs, C., Legge, S., Lowe, A.J.,

- 473 McLean, R., Montambault, J., Possingham, H., Radford, J., Robinson, D., Smallbone, L., Thomas, D., Varcoe,
- 474 T., Vardon, M., Wardle, G., Woinarski, J. & Zerger, A. (2012) Improving biodiversity monitoring. Austral Ecol,
- 475 37, 285-294.
- Lovett, G.M., Burns, D.A., Driscoll, C.T., Jenkins, J.C., Mitchell, M.J., Rustad, L., Shanley, J.B., Likens, G.E.
- 477 & Haeuber R. (2007) Who needs environmental monitoring? Front Ecol Environ, 5, 253-260.
- Löfstedt, C., Herrebout, W.M., & Menken, S.B. (1991). Sex pheromones and their potential role in the evolution
- of reproductive isolation in small ermine moths (Yponomeutidae). Chemoecology, 2, 20-28.
- 480 Milberg, P., Bergman, K.-O., Johansson, H. & Jansson, N. (2014) Low host-tree preferences among saproxylic
- beetles: a comparison of four deciduous species. Insect Conserv Diver, in press.
- 482 Millar, J.G., McElfresh, J.S., Romero, C., Vila, M., Mari-Mena, N. & Lopez-Vaamonde, C. (2010)
- Identification of the sex pheromone of a protected species, the Spanish moon moth Graellsia isabellae. J Chem
- 484 Ecol, 36, 923-932.
- Musa, N., Andersson, K., Burman, J., Andersson, F., Hedenström, E., Jansson, N., Paltto, H., Westerberg, L.,
- Winde, I., Larsson, M.C., Bergman, K.-O. & Milberg, P. (2013) Using sex pheromone and a multi-scale
- 487 approach to predict the distribution of a rare saproxylic beetle. PLoS ONE, 8, e66149.
- Nilsson, S.G., Arup, U., Baranowski, R. & Ekman, S. (1995) Tree-dependent lichens and beetles as indicators in
- 489 conservation forests. Conserv Biol, 9, 1208-1215.
- 490 Palmqvist., G (2014) Intressanta fynd av storfjärilar (Macrolepidoptera) i Sverige 2013. [Remarkable records of
- 491 Macrolepidoptera in Sweden 2013]. Entomologisk Tidskrift, 135, 63-76.
- 492 Pellet, J. & Schmidt, B.R. (2005) Monitoring distributions using call surveys: estimating site occupancy,
- detection probabilities and inferring absence. Biol Conserv, 123, 27-35.
- 494 Pereira, H.M., & Cooper, D.H. (2006) Towards the global monitoring of biodiversity change. Trends Ecol Evol,
- 495 21, 123-129.
- 496 Quinto, J., Marcos-García, M.A., Brustel, H., Galante, E. & Micó, E. (2013) Effectiveness of three sampling
- 497 methods to survey saproxylic beetle assemblages in Mediterranean woodland. J Insect Conserv, 17, 765-776.
- 498 R Core Team (2014) R: A language and environment for statistical computing. R Foundation for Statistical
- Computing, Vienna, Austria. http://www.R-project.org/ 25th November 2015.
- Rademaekers, K., Braat, L.C., Eichler, L., Widerberg, O. & Jones-Walters, L. (2010) Costs and benefits
- assessment of monitoring approaches for measuring progress towards the EU 2020 biodiversity target. Available
- at http://edepot.wur.nl/173781
- Rosenberg, D.M., Danks, H.V. & Lehmkuhl, D.M. (1986) Importance of insects in environmental impact
- assessment. Environ Manag, 10, 773-783.

- 505 Svensson, G.P., Liedtke, C., Hedenström, E., Breistein, P., Bång, J. & Larsson, M.C. (2012) Chemical ecology
- and insect conservation: optimizing pheromone-based monitoring of the threatened saproxylic click beetle
- 507 Elater ferrugineus. J Insect Conserv, 16, 549-555.
- 508 Szántóné-Veszelka, M., Poós, B., & Szőcs, G. (2010) Blackberry and raspberry, new hosts of the yellow legged
- clearwing moth, Synanthedon vespiformis: what can the recently developed sex attractant offer in monitoring
- and beyond. In IOBC Working Group, Integrated Plant Protection in Fruit Crops Subgroup "Soft Fruits", 7th
- workshop on Integrated Soft Fruit Production (pp. 20-23).
- Tobin, P.C., Liebhold, A.M. & Roberts, E. A. (2007) Comparison of methods for estimating the spread of a non-
- 513 indigenous species. J Biogeogr, 34, 305-312.
- Tolasch, T., Von Fragstein, M. & Steidle, J.L.M. (2007) Sex pheromone of *Elater ferrugineus* L. (Coleoptera:
- 515 Elateridae). J Chem Ecol, 33, 2156-2166.
- Van der Meulen, J., & Groenendijk, D. (2005) Assessment of the mobility of day-flying moths: an ecological
- approach. Proc Exp Appl Entomol, 16, 37-50.
- Waring, P. & Townsend, M. (2003) Field Guide to the Moths of Great Britain and Ireland. British Wildlife
- Publishing Ltd, UK.
- Zauli, A., Chiari, S., Hedenström, E., Svensson, G.P. & Carpaneto, G.M. (2014) Using odour traps for
- 521 population monitoring and dispersal analysis of the threatened saproxylic beetles Osmoderma eremita and
- 522 Elater ferrugineus in central Italy. J Insect Conserv, 18, 801-813.
- 523 Zhang, Q.-H., Schlyter, F. (1996) High recaptures and long sampling range of pheromone traps for fall web
- worm moth *Hyphantria cunea* (Lepidoptera: Arctiidae) males. J Chem Ecol, 22, 1783-1796.
- Yamanaka, T., Satoda, S., Senda, S. & Tatsuki, S. (2001). Mass-trapping trials of the fall webworm, *Hyphantria*
- 526 cunea (Drury)(Lepidoptera: Arctiidae), with synthetic sex pheromone in urban street trees. Jpn J Appl Entomol
- 527 Z, 12, 175-183.
- 528 Yoccoz, N.G., Nichols, J.D. & Boulinier, T. (2001) Monitoring of biological diversity in space and time. Trends
- 529 Ecol Evol, 16, 446-453.
- Östrand, F. & Anderbrant, O. (2003) From where are insects recruited? A new model to interpret catches of
- attractive traps. Agr Forest Entomol, 5, 163-171.

533

532

534

535

536

537 Figure captions 538 ¹Fig.1 a) Map showing pre-2010 records (www.artportalen.se) of *S. vespiformis* in Southern Sweden. b) 539 Recordings of S. vespiformis as of 2013 without pheromone survey. c) Reported sites in 2013 after inclusion of 540 pheromone survey data. d) Map showing the distribution of pheromone traps in southern Sweden in the Skåne, 541 Blekinge, Kalmar and Östergötland counties (Swedish "Län" = county). Filled and empty circles indicate sites 542 where S. vespiformis. was recorded and was not recorded, respectively 543 ¹Fig.2 Map of S. vespiformis sampling area in Östergötland, Sweden. The black dots represent trees, the large 544 grey circles represent the largest scale (6,000 m) used to determine tree abundance, and the small white circles 545 with (occupied) and without (empty) black crosses represent trap locations 546 ²Fig.3 Frequency of site reports of *S. vespiformis* since 1976 (www.artportalen.se). The implementation of 547 pheromone lures contributes significantly towards the greatly increased numbers of reported sites in 2011 and 548 2012 549 ³Fig.4 The relationship (median Z-value from 500 negative binomial regressions at each scale and tree group) 550 between abundance of S. vespiformis and amount of a) oak trees with various characteristics at multiple scales 551 and b) non-oak deciduous trees at multiple scales. Light and dark gray circles denote negative and positive 552 relationships respectively, and ring size above 1.96 is significant at 5% level. 553 ¹Figure produced in ARCGIS 554 ²Figure produced in Microsoft Excel 555 ³Figure produced in R version 3.1.2 556

County	Number of moths caught	Number of sites with species present	Number of sites with species not present	Number of traps placed
Skåne	5	2	35	37
Blekinge	12	6	20	26
Kalmar	308	26	46	72
Östergötland	114	43	73	116
Total	439	77	174	251

Table 1. Pheromone based recordings of *S. vespiformis* across four counties in Southern Sweden.









