

1 **A possible Cambrian stem-group gnathiferan-chaetognath from the Weeks**
2 **Formation (Miaolingian) of Utah**

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18 **Running Header:** Possible stem-group gnathiferan-chaetognath

19

20 **Abstract**—In recent years the plethora of “weird wonders”, the vernacular for the apparently
21 extinct major bodyplans documented in many of the Cambrian Lagerstätten, has been
22 dramatically trimmed. This is because various taxa have either been assigned to known phyla
23 or at least accommodated in larger monophyletic assemblages. Nevertheless, a number of
24 Cambrian taxa retain their enigmatic status. To this intriguing roster we add *Dakorhachis*
25 *thambus* n. gen. n. sp., from the Miaolingian (Guzhangian) Weeks Formation Konservat-

26 Lagerstätte of Utah. Specimens consist of an elongate body lacking appendages, but which is
27 apparently segmented. A prominent feeding apparatus consists of a circlet of triangular teeth,
28 while posteriorly there are three distinct skeletal components. *D. thambus* n. sp. is interpreted
29 as an ambush predator and may have been partially infaunal. The wider affinities of this new
30 taxon remain conjectural but it is suggested that it may represent a stem-group member of the
31 Gnathifera, today represented by the gnathostomulids, micrognathozoans, rotifers, and
32 possibly with links also to the chaetognaths.

33

34 UUID: <http://zoobank.org/22113E6B-F79E-4D06-9483-144618A61327>

35

36 **Introduction**

37

38 Cambrian Konservat-Lagerstätten (“Lagerstätten” hereafter) provide exceptional insights into
39 early metazoan evolution, not least because of an abundance of lightly sclerotized and soft-
40 bodied taxa. Laurentian examples include the iconic Burgess Shale in British Columbia
41 (Canada), as well as a series of important deposits in Utah (Spence Shale, Wheeler Formation,
42 Marjum Formation; e.g. Muscente et al., 2017; also Fig. 1). In contrast the Weeks Formation
43 (Miaolingian), exposed near Notch Peak, Utah, only more recently has yielded an important
44 Burgess Shale-type fauna (Hesselbo, 1989; Lerosey-Aubril et al., 2013, 2014, 2015, 2018;
45 Lerosey-Aubril, 2015; Ortega-Hernández et al., 2015; Robison et al., 2015). Not only is this
46 latter assemblage important in extending our knowledge of Cambrian life, but its
47 chronological position close to the Miaolingian/Furongian boundary fills a significant gap in
48 the fossil record of non-biomineralizing animals and apparently corresponds to the onset of
49 major biotic changes (Lerosey-Aubril et al., 2018).

50 To the first approximation Burgess Shale-type faunas (e.g. Briggs et al., 1994; Hou et
51 al., 2004) have a well-established identity with a predominance of arthropods (both trilobites,
52 including agnostoids, as well as lightly skeletonized taxa), priapulids (and related
53 scalidophorans), and sponges. Somewhat more occasional there occur such groups as the
54 annelids, vetulicolians, wiwaxiids and other sclerite-bearing taxa. Such faunas remain a focus
55 of attention not only on account of their sheer diversity, but also because a number of hitherto
56 problematic taxa appear to belong to stem-groups that in principle are instrumental in our
57 understanding of the origin of phyla.

58 Not all such taxa, however, can be accommodated in such a fashion and in one way or
59 another a number of them retain their enigmatic status. Broadly these can be divided into
60 three categories, although the boundaries that separate them are by no means absolute. There
61 are those, such as the vetulicolians, that form a relatively diverse clade but whose wider
62 relationships within the deuterostomes continue to be controversial (e.g. Ou et al., 2012;
63 García-Bellido et al., 2014). Then there are such taxa as *Nectocaris* that have deeply
64 polarized opinion, in this case as to whether or not this animal is an early cephalopod (e.g.
65 Kroger et al., 2011; Smith, 2013). Finally there are singletons that for all intents and purposes
66 remain in taxonomic limbo, and it is to this last category we add a remarkable new taxon,
67 *Dakorhachis thambus* n.gen. n.sp. (Fig. 1). These three categories also have the heuristic
68 value of providing a crude metric of relative phylogenetic ignorance, although in each case
69 new fossil finds ultimately will ensure more secure placement within the metazoan tree.
70 Moreover, properly interpreted these enigmatic taxa may help to throw crucial light on key
71 transitions between major groups. At this juncture we are unable to assign *D. thambus* n. sp.
72 with confidence to any known group, but it is evidently a member of the Bilateria. In our
73 opinion this taxon is more likely to fall within the Spiralia (rather than the deuterostomes or

74 ecdysozoans). Below we tentatively suggest that *D. thambus* n. sp. might represent a stem-
75 group gnathiferan.

76

77 **Geological setting**

78

79 The general setting of this Lagerstätte has been reviewed by Lerosey-Aubril et al., (2018). In
80 brief, the Weeks Formation (Miaolingian; Guzhangian) is a relatively deep-water deposit,
81 apart from the upper section (70 m) that records a substantial shallowing of the depositional
82 environment associated with the end of basinal accumulation in the so-called House Range
83 Embayment. Below this transitional interval, lithologies are alternating micrites and
84 calcareous claystones. These are indicative of a low-energy, distal ramp environment, which
85 was periodically disturbed by storm-induced gravity flows and episodes of oxygen depletion.
86 Unlike the Burgess Shale, where much of the biota was introduced into a toxic environment
87 by small turbidity flows (e.g. Conway Morris, 1986), in this Lagerstätte transport was
88 evidently minimal. The exceptional preservation in the Weeks Formation is restricted to a 25-
89 meter-interval about 205 m below the top of the unit. This interval has yielded a diverse
90 fauna (c. 73 species) which according to agnostoids (*Proagnostus gibbus* Zone) and trilobites
91 (*Cedaria* Zone) is of mid-Guzhangian age.

92

93 **Materials and Methods**

94

95 The material consists of nineteen specimens preserved flattened parallel to bedding. One slab
96 (UU.15101.05, 15101.06) bears two specimens, two slabs (UU15101.02, 15101.03, 15101.04
97 and 15101.12, 15101.13, 15104.14) have three specimens each, while another slab
98 (UU15101.07, 15101.08, 15101.09, 15101.10, 15101.11) has five superimposed specimens;

99 other specimens are isolated. This material was examined under a binocular microscope, with
100 a drawing tube employed to prepare camera-lucida interpretative drawings. Specimens
101 UU17122.03, 18056.27, 18056.28 were photographed immersed in dilute ethanol using a
102 Leica IC80 HD camera mounted on a Leica M80 microscope. Specimen UU17122.03 was
103 studied uncoated (low vacuum mode) using a scanning electron microscope (SEM) JEOL
104 JSM-6010LV equipped with an energy-dispersive X-ray spectroscopy (EDS) module JEOL
105 EX-94410T1L11 at the University of New England. Similar SEM and EDS investigations
106 were performed on both entire specimens (UU15101.01, UU15101.07) and polished sections
107 using a QEMSCAN 650F SEM at the University of Cambridge. Lastly, computed
108 tomography (CT) scan of specimen UU15101.01 (holotype) were obtained using a Nikon
109 XTH225 ST CT scanner at the Cambridge Biotomography Centre.

110

111 *Repository and institutional abbreviations.*—Types, figures, and other specimens (including
112 petrographic sections) examined in this study are deposited in the Department of Geology and
113 Geophysics (Research Collections), University of Utah, USA (UU) and Back to the Past
114 Museum, Cancún, Mexico (BPM).

115

116 **Preservation**

117

118 The fossils described here show the same style of preservation as most of the non-
119 biomineralizing taxa of the fauna (Lerosey-Aubril et al., 2018, fig. 3b, c). This is the result of
120 a series of chemical and physical alterations that occurred mostly at a late stage of diagenesis.
121 Such is very much a hall-mark of the Weeks Formation fauna where evidence of diagenetic
122 phosphatization is associated with strong taxonomic and histological controls. Indeed, all
123 known instances of secondarily phosphatization concern organs rich in phosphorus (e.g.

124 arthropod guts) or tissues underneath primary phosphatic structures, such as aglaspidid cuticle
125 or palaeoscolecid plates (Lerosey-Aubril, 2015; Lerosey-Aubril et al., 2012, 2018).

126 In the case of *D. thambus* n. sp. these post-mortem changes include the initial
127 flattening of the carcasses, and much later the replacement of the presumably carbonaceous
128 material with pyrite and subsequent coating of this pyritic layer (now as oxidized
129 pseudomorphs) by chlorite (in a fan-like arrangement) (Fig. 2). This strong diagenetic
130 imprint is related to major igneous intrusions as well as more recent intense weathering.
131 Scanning electron micrographs of specimens of *D. thambus* n. sp. suggest that the trunk is
132 chiefly composed of iron oxides and chlorite (Fig. 2), and this is consistent with
133 compositional (EDS) analyses (Fig. S1).

134 The translucent teeth differ in composition from the trunk, and appear to have a
135 predominantly calcitic composition (Fig. 2.3-2.5). As discussed below whilst an original
136 composition cannot be excluded, it seems as likely that the calcite is also diagenetic. Micro-
137 CT shows moderate 3D preservation of the teeth at the specimen surface (Fig. 5). However,
138 due to the mode of fossil preservation (low density-contrast composition and compression),
139 no further (e.g. internal or subsurface) 3D information was recovered.

140

141 **Systematic paleontology**

142

143 ?Superphylum Spiralia

144 ?Gnathifera-Chaetognatha

145 Family Dakorhachiidae new family

146

147 *Type genus (by monotypy).*—*Dakorhachis* n. gen. from the Miaolingian (Guzhangian) Weeks
148 Formation of the House Range, Utah, USA.

149

150 *Diagnosis.*—Vermiform, segmented body anteriorly bearing prominent ?calcitic teeth.

151

152 *Remarks.*—Chaetognatha is currently treated as a distinct phylum, and recent molecular
153 evidence (Fröbius and Funch, 2017; Marlétaz et al., 2019) links them to the Gnathifera whose
154 component phyla are Gnathostomulida, Micrognathozoa and Rotifera (with parasitic
155 Acanthocephala). Phylum status denotes their morphological distinctiveness, but all these
156 phyla are united by the possession of an anterior basket of chitinous teeth. As discussed
157 below *D. thambus* n. sp. is tentatively interpreted as a stem-group representative of a
158 Gnathifera-Chaetognatha clade (we suggest the colloquial moniker chaetognathiferans).

159

160 *Genus Dakorhachis* new genus

161

162 *Type species (by monotypy).*—*Dakorhachis thambus* n. gen. n. sp.

163

164 *Diagnosis.*—As for type species by monotypy.

165

166 *Etymology.*—A combination of *dakos* (Greek) a biter and *rachis* (Greek) ridge.

167

168 *Remarks.*—A new genus that amongst other taxa from the Cambrian finds no close
169 counterpart, nor appears to have known equivalents elsewhere.

170

171 *Dakorhachis thambus* new species

172

Figures 3–7, S2

173

174 2018 “Enigmatic organism” Lerosey-Aubril et al., fig. 3a–c

175

176 *Holotype*.—Complete specimen (UU15101.01), Department of Geology and Geophysics

177 (Research Collections), University of Utah, Salt Lake City, Utah.

178 *Diagnosis*.—Elongate and robust body. Feeding apparatus comprising at least six hollow

179 teeth, characterized by gently convex outer side with prominent central ridge and concave

180 inner side with narrow ridge-like margins, and in posterior direction associated skeletal

181 elements in form of hook-like elements, inverse v-shaped sclerites, and elongate rods. Trunk

182 composed of 30 segments, gently tapering posteriorly, terminating in blunt tip.

183

184 *Occurrence*.—Exposures in North Canyon, adjacent to Notch Peak, House Range, Utah.

185 Weeks Formation (Cambrian Series 3, Guzhangian).

186

187 *Description*.—Apart from minor preservational variants, the material is united in showing a

188 body consisting of a relatively elongate trunk (Figs. 3, 5, S3) which at its anterior bears a

189 prominent feeding apparatus (Figs. 4, 5.3). Total length can reach 28 mm, and maximum

190 width of trunk 7 mm (Fig. S3). The feeding apparatus bears at least six prominent teeth, of

191 which about half are exposed in outer aspect and the remainder in inner aspect, suggesting

192 that originally they formed a circum-oral circlet (Fig. 4.1,4.3). Each tooth (c. 3 mm long) has

193 a narrow triangular form, and in outer aspect is gently convex and bears a prominent and

194 relatively narrow longitudinal ridge. In inner aspect the tooth is concave, but the margins are

195 defined by very narrow ridges. The teeth have a fibrous texture, while the broken margin of

196 one tooth shows what may be a hollow interior (Fig. 6). Elemental analyses indicate that the

197 teeth to have a predominantly calcitic composition (see above and Dryad file). Posterior to the

198 teeth are three other skeletal components, evidently with a similar composition to the teeth

199 (Fig. 4.2, 4.4). Immediately to the posterior of the teeth are small hook-like structures, while
200 behind them are a series of inverse V-shaped units. Most likely these units also formed
201 circlets. Finally adjacent to, or superimposed on, the anteriormost trunk are rod-like
202 structures, usually straight but occasionally with a sinuous shape.

203 The trunk is relatively featureless and lacks appendages or other extensions. In some
204 specimens the configuration is somewhat sinuous (Fig. 3.3; see also Lerosey-Aubril et al.,
205 2018, fig. 3a), suggesting an original degree of flexibility (also Fig. 3.7). The width is more
206 or less uniform and although most specimens have a rounded termination, it occasionally
207 appears to be acute. Broad transverse folds (c. 0.8 mm) may be surficial annulations but here
208 are interpreted as segments (Figs. 3, 1., 3.2, 5. 1., 5.2, 7). In life these would have totalled
209 about 30. That these structures are original rather than post-mortem (or tectonic) is supported
210 by three lines of evidence. First, these transverse bands are evidently three-dimensional (Fig.
211 S2) and sometimes match a corrugated body margin. In addition, associated specimens with
212 different orientations have folds transverse to their respective bodies rather than parallel to
213 any rock fabric (Fig. S3).

214

215 *Etymology.*—*thambos* (Greek). an astonishment.

216

217 *Materials.*—UU15101.02–15101.15, 17122.03, 18056.27, 18056.28; BPM-1090.

218

219

DISCUSSION

220

221 *Paleoecology and mode of life.*—*D. thambus* n. sp. lacks fins or other anatomical features
222 consistent with a pelagic mode of life and therefore is interpreted as benthic. Co-association
223 of specimens indicates a gregarious habit, although the case of parallel stacking

224 (UU15101.07-15101.11) is most likely post-mortem. Locomotory organs are not evident but
225 presumably this animal could have moved across or within the sea-floor by peristaltic
226 contractions. Given, however, the arrangement of the teeth is in the form of a sort of basket it
227 may have captured its prey as an ambusher and as such the animal may have been semi-
228 sessile and partially concealed in the sea-floor. The attitude of the teeth varies from parallel to
229 an anterior convergence, but in life they presumably opened wider to tackle larger prey. The
230 function of the skeletal elements posterior to the teeth is more conjectural. One suggestion is
231 that they served for insertion of muscles associated with protrusion and subsequent closure of
232 the teeth.

233 *Phylogenetic affinities.*—The wider relationships of *D. thambus* n. sp. are necessarily
234 problematic given its lack of close identity to any known group. Such evidence as there is
235 must look to the feeding apparatus. A potentially important clue might be the calcitic
236 composition of the teeth, although as noted this may well be diagenetic. Certainly amongst
237 metazoans calcitic teeth are unusual, with the most notable instances being in the echinoids
238 (e.g. Wang et al., 1997; Stock et al., 2014) and extinct ophiocistioids (e.g. Reich et al., 2018).
239 Moreover, in the former group they can on occasion show a fibrous microstructure (Reich and
240 Smith, 2009, text-Fig. 9C, D). There is, however, no other feature of *Dakorhachis* n. gen. that
241 would indicate an affinity to either the echinoids or any other echinoderm, especially if the
242 principal teeth totalled six, an obvious departure from the characteristic pentaradial symmetry
243 of this phylum.

244 In passing, it is worth noting that *D. thambus* n. sp. shows some broad similarities in
245 overall shape to the unusual sponge *Takakkawia lineata* from the Burgess Shale, which has
246 marginal “fins” extending from a conical body (Botting, 2012). However, numerous detailed
247 differences in morphology indicate that the resemblance between these taxa is superficial.
248 Specifically, there are differences in the size, shape and annulation of the body in *D. thambus*

249 (which averages 2.1 cm long, is vermiform and has transverse annulations) versus *T. lineata*
250 (which is longer (Botting, 2012, Fig. 1), more vasiform and displays diagnostic lengthwise
251 lineations). Furthermore, there are major differences in the shape and organisation of the
252 teeth of *Dakorhachis* (which are elongate, sharply pointed and restricted to the anterior body
253 margin) versus the “fins” of *T. lineata* (which are wider, flat topped, and accompanied by
254 broad spicules extending down the length of the body). Whilst there is no other reason to
255 interpret *D. thambus* n. sp. as any sort of sponge, the potential complexities of assigning
256 Cambrian taxa to particular groups and the consequent phylogenetic implications are apparent
257 from Botting and Muir’s (2018) proposed linkage of *Takakkawia* to the putative ctenophore
258 *Thaumactena*. That said there is no evidence for comparing *D. thambus* n. sp. to any of the
259 Cambrian ctenophores (e.g. Ou et al., 2015).

260 Notwithstanding such comparisons, *D. thambus* n. sp. is evidently a bilaterian rather
261 than a representative of the diploblasts (let alone a sponge). There appears to be no particular
262 similarity to either the deuterostomes or ecdysozoans. Although, in the latter case, it is true
263 that the priapulids and related scalidophorans typically have an introvert equipped with
264 circlets of teeth, these and associated structures show a complex zonation and diversity of
265 forms (e.g. Smith et al., 2015) that find no counterpart in the array of teeth seen in *D. thambus*
266 n. sp. or its ancillary skeletal structures. Most likely *D. thambus* n. sp. is a member of the
267 Spiralia.

268 Amongst the spiralian the most fruitful comparisons may possibly lie with the
269 Gnathifera. This monophyletic group (e.g. Laumer et al., 2015) comprises the
270 gnathostomulids (e.g. Herlyn and Ehlers, 1997; Sørensen et al., 2006), its sister group the
271 micrognathozoans (e.g. Bekkouche and Worsaae, 2016; Bekkouche et al., 2014), and the
272 syndermatans (the group encompassing the rotifers and endoparasitic acanthocephalans; e.g.
273 Rieger and Tyler, 1995; Wulfken and Ahlrichs, 2012). Gnathiferans are millimetric and

274 typically meiofaunal, but despite this all possess intricate jaw apparatuses that reaches an
275 apogee in the complex array found in the micrognathozoans (e.g. Kristensen and Funch, 2000;
276 De Smet, 2002; Sørensen, 2003). Current phylogenetic schemes place the gnathiferans as
277 sister to all other spiralian (e.g. Bakkouche and Worsaae, 2016; Laumer et al., 2015), which
278 in turn are broadly divided into the “platyzoans” and the more securely identified
279 lophotrochozoans.

280 The disparity of extant gnathiferans, combined with an almost non-existent fossil
281 record (e.g., Poinar and Ricci, 1992; Waggoner and Poinar, 1993; Jha et al., 2011), and their
282 still poorly resolved systematic position within the bilaterians, pose a series of evolutionary
283 questions. Amongst the most problematic is the visualization of a stem-group form and its
284 corresponding recognition in the fossil record. This question may be further exacerbated if
285 the millimetric size of the extant gnathiferans is the result of secondary miniaturization from
286 macroscopic predecessors, rather than a primitive state.

287 Intriguingly there is also phylogenomic evidence for a link between the gnathiferans
288 and chaetognaths (Fröblius and Funch, 2017; Marlétaz et al., 2019). The latter are equipped
289 with a formidable feeding apparatus consisting of prominent grasping spines and associated
290 teeth (e.g. Bone et al., 1991), although at first sight there is no obvious macroscopic
291 connection to any of the considerably more complex gnathiferans jaws. The phylogenetic
292 position of the chaetognaths has long been regarded as basal amongst the bilaterians (Perez et
293 al., 2013), but with conflicting views suggesting either a place amongst the most primitive
294 protostomes (e.g. Marlétaz et al., 2006; Marlétaz and Le Parco, 2008; Shen et al., 2016) as
295 against a position amongst the basal lophotrochozoans (e.g. Matus et al., 2007; Dunn et al.,
296 2008; Bernt et al., 2013).

297 The contribution of the Cambrian fossil record to the early evolution of the
298 chaetognaths and gnathiferans to date has almost entirely focused on the former group. Here

299 the protoconodonts, which apart from occasional fused clusters, are effectively dispersed as
300 small shelly fossils (Szaniawski, 1982, 2002), are complemented by several soft-bodied taxa
301 similar to extant chaetognaths (Chen and Huang, 2002; Hu et al., 2007; Vannier et al., 2007;
302 Shu et al., 2017), and what appear to be two more primitive representatives (*Ankalodus*
303 *sericus* and *Capinator praetermissus*) characterized by supernumerary teeth (Briggs and
304 Caron, 2017) or a multi-jawed morphology (Shu et al., 2017) (Fig. S1). It is now clear,
305 however, that the hitherto enigmatic *Amiskwia* (Conway Morris 1977) possesses a jaw
306 apparatus that supports some sort of connection to the gnathiferans and/or chaetognaths
307 (Vinther and Parry, 2019; Caron and Cheung, 2019).

308 Although the record of relevant soft-bodied taxa (*Amiskwia*, *Ankalodous*, *Capinator*)
309 is meagre, as potential stem-group chaetognathiferans they hint as both morphological
310 disparity and a range of ecologies from swimming to benthic. To this roster we tentatively
311 propose to add *D. thambus* n. sp. As is the case with a number of other controversial
312 Cambrian groups, a convincing phylogenetic analysis is frustrated by the paucity of available
313 character-states and the added possibility that those available for tabulation in reality are
314 convergent. Our assignment relies on a tentative interpretation of the feeding apparatus of *D.*
315 *thambus* n. sp. as a precursor to the much more complex jaws seen in extant gnathiferans as
316 well as the possible equivalent in the chaetognaths. Here, therefore, we sketch a possible set
317 of transitions (Fig. 8) that might link the feeding apparatus of *Dakorhachis* n. gen to those of
318 the gnathiferans and chaetognaths.

319 There is agreement that some of the elements of gnathiferan apparatuses are
320 homologous (e.g., Sørensen, 2002; Sørensen et al., 2006), but nevertheless collectively the
321 clade shows a wide diversity of forms. Interestingly, the more basal gnathostomulids possess
322 a somewhat less elaborate jaw (e.g., Riedl and Rieger, 1972) and within this group there are a
323 number of trends that can be traced from what appears to be the most primitive arrangement

324 (e.g., Sterrer, 1972; Sørensen, 2002). Thus, despite various elaborations, the basic
325 configuration of the jaw is as a forceps-like unit joined to a proximal base and a basal plate.
326 Derivation of this arrangement from something similar to *D. thambus* n. sp. via an amiskwiid
327 (Caron and Cheung, 2019) would, in principle, involve a shift from an effectively radial
328 symmetry to a bilateral configuration, reduction from six teeth to three (along with substantial
329 miniaturization), and possibly incorporation of the more proximal skeletal elements in *D.*
330 *thambus* n. sp. into the jaw apparatus.

331 The likely phylogenetic relationship between chaetognaths and gnathiferans (Fröbius
332 and Funch, 2017; Marlétaz et al., 2019) may also find some support in the morphology
333 exhibited by *D. thambus* n. sp. Whilst there is little obvious similarity between the jaw
334 configurations of the gnathiferans versus chaetognaths, in both cases the principal
335 composition is chitinous (e.g., Bone et al., 1983; Sørensen and Sterrer, 2002). The distinctive
336 rod-like microstructures of most gnathiferan teeth (e.g., Riemann and Ahlrichs, 2008) is
337 presumably a synapomorphy of the group, but in *D. thambus* n. sp. the fibrous microstructure
338 and possible hollow interior find a possible counterpart in the protoconodonts (e.g.
339 Szaniawski, 2002). If there is an evolutionary connection between *D. thambus* n. sp. and the
340 chaetognaths then in parallel to the gnathiferans this would involve a transition between the
341 apparently radial configuration of the teeth in the former taxon to the bilateral arrangement on
342 the chaetognaths. Although very different to the trajectory of the gnathiferans that led
343 towards a meiofaunal existence, this proposed evolutionary path would also be a consequence
344 of a major ecological shift, from a perhaps semi-sessile benthic life style to a more motile
345 pelagic one.

346 It is worth pointing out that whilst the fused clusters of protoconodonts (e.g.,
347 Szaniawski, 1982, 2002) are convincingly compared to the bundles of feeding spines in the
348 chaetognaths, in contrast most protoconodont taxa are never recovered as fused clusters.

349 Whilst this disaggregation may be the consequence of standard processing of samples by acid
350 digestion, it seems equally possible that in such taxa the arrangement of the feeding apparatus
351 was either more open and/or arranged as a multiple series (Shu et al., 2018). An alternative
352 option might be that some of these feeding spines actually belonged to animals closer to *D.*
353 *thambus* n. sp., where the teeth were not clustered, but radially organised around a terminal
354 mouth. In terms of similarities of the teeth of *D. thambus* n. sp. and supposed
355 protoconodonts, two possible candidates are some specimens of *Protohertzina robusta* (Pyle
356 et al., 2006, fig. 6.8) and an unnamed taxon described by Kouchinsky et al. (2015, fig. 53M,
357 their ‘undetermined form 4’). Our knowledge of early chaetognath evolution may also be
358 incomplete. Thus the otherwise distinctive coelocerodonts (Szaniawski, 2015) have a
359 chaetognath-like arrangement of the teeth, while the possible protoconodont *Huayuanodontus*
360 has a tooth histology distinct from other taxa (Dong, 2007).

361 If we are correct in regarding *D. thambus* n. sp. as a sister-taxon of the clade
362 gnathiferans-chaetognaths, this suggests that their common ancestor was macroscopic, semi-
363 sessile and segmented. Thus, the miniaturization and largely meiofaunal existence would
364 have been secondarily acquired in the evolutionary history of gnathiferans, in contrast to the
365 general assumption that it is a plesiomorphic condition for the group (e.g., Laumer et al.,
366 2015). As to the chaetognaths, our discovery cannot resolve more precisely their position to
367 other early bilaterians (e.g. Marlétaz et al., 2008, 2019; Shen et al., 2016). It supports,
368 however, the idea, that notwithstanding subsequent loss and redeployment (Blair, 2008),
369 segmentation amongst the bilaterians is primitive. Moreover, in extant chaetognaths the
370 progenitor neural cells of the trunk are not only highly organized but form 30-35 rows (Perez
371 et al., 2013), comparable to the segment total in *D. thambus* n. sp.. Primitive chaetognaths
372 such as *Ankalodous* (Shu et al., 2017) may have also had relatively limited motility, but
373 overall there was evidently a shift to a much more active mode of life (e.g. Vannier et al.,

374 2007). Evidence for a migration to a pelagic mode of life (Vannier et al., 2007; Casenove et
375 al., 2011; Hu et al., 2007) is supported both by the evolution of chaetognath musculature
376 (Casanova and Duvert, 2002) and molecular data (Papillon et al., 2006). Significantly this
377 shift may have been via benthoplanktonic intermediates, although the few truly benthic
378 chaetognaths extant are very derived (Casanova and Duvert, 1996) and show no significant
379 similarities to *D. thambus* n. sp. This transition to the pelagic realm would also have been
380 marked by the separation of the teeth into two separate grasping bundles (along with smaller
381 teeth adjacent to the mouth), changes in the patterns of their replacement (Moreno and Kapp,
382 2003), and loss of mineralization to assist buoyancy. This would have been combined with
383 extensive re-organization of the head musculature. Further changes would have included
384 narrowing of the body, reduction to an oligomeric (tripartite) segmentation (Balavoine and
385 Adoutte, 2003), as well as the development of prominent fins and complex eyes.

386

387 **Accessibility of supplemental data**

388

389 Data available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.p5hqbzkkz>

390

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392

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399

400 **References**

401

402 Balavoine, G., and Adoutte, A., 2003, The segmented Urbilateria: A testable scenario:

403 Integrative and Comparative Biology, v. 43, p. 137–147.

404 Bekkouche, N., and Worsaae, K., 2016, Nervous system and ciliary structures of

405 Micrognathozoa (Gnathifera): Evolutionary insight from an early branch in Spiralia:

406 Royal Society Open Science, v. 3, e160289.

407 Bekkouche, N., Kristensen, R.M., Hejnol, A., Sørensen, M.V., and Worsaae, K., 2014,

408 Detailed reconstruction of the musculature in *Limnognathia maerski* (Micrognathozoa)

409 and comparison with other Gnathifera: Frontiers in Zoology, v. 11, e71.

410 Bernt, M. et al., 2013, A comprehensive analysis of bilaterian mitochondrial genomes and

411 phylogeny: Molecular Phylogenetics and Evolution, v. 69, p. 352–364.

412 Blair, S.S., 2008, Segmentation in animals: Current Biology, v. 18, R991-R995 (2008).

413 Bone, Q., Ryan, K.P., and Pulsford, A.L., 1983, The structure and composition of teeth and

414 grasping spines of chaetognaths: Journal of the Marine Biological Association of the

415 United Kingdom, v. 63, p. 929–939.

416 Bone, Q., Kapp, H., and Pierrot-Bults, A.C. eds., 1991, The Biology of Chaetognaths: Oxford

417 University Press, Oxford, 173 p.

418 Botting, J.P., 2012, Reassessment of the problematic Burgess Shale sponge *Takakkawia*

419 *lineata* Walcott, 1920: Canadian Journal of Earth Sciences, v. 49, 1087–1095.

420 Botting, J.P., and Muir, L.A., 2018, Early sponge evolution: A review and phylogenetic

421 framework: Palaeoworld, v. 27, p. 1-29.

422 Botting, J.P., and Peel, J.S., 2016, Early Cambrian sponges of the Sirius Passet biota, North

423 Greenland: Papers in Palaeontology, v. 2, p. 463–487.

- 424 Briggs, D.E.G., and Caron, J-B., 2017, A large Cambrian chaetognath with supernumerary
425 grasping spines: *Current Biology*, v. 27, p. 2536–2543.
- 426 Briggs, D.E.G., Erwin, D.H., and Collier, F.J., 1994, *The Fossils of the Burgess Shale*.
427 Smithsonian Institution Press, Washington D.C.
- 428 Casanova, J-P., and M. Duvert, M., 1996, Biodiversity and evolutionary trends in the phylum
429 Chaetognatha: *Bulletin de Société zoologique de France*, v. 121, p. 77–80.
- 430 Casanova, J-P., and Duvert, M., 2002, Comparative studies and evolution of muscles in
431 chaetognaths: *Marine Biology*, v. 141, p. 925–938.
- 432 Casenove, D., Oji, T., Goto, T., 2011, Experimental taphonomy of benthic chaetognaths:
433 Implications for the decay process of Paleozoic chaetognath fossils: *Paleontological*
434 *Research*, v. 15, p. 146–153.
- 435 Chen, J-Y., and Huang, D-Y., 2002, A possible Lower Cambrian chaetognath (arrow worm):
436 *Science*, v. 298, p. 187.
- 437 Conway Morris, S., 1977, A redescription of the Middle Cambrian worm *Amiskwia*
438 *sagittiformis* Walcott from the Burgess Shale of British Columbia: *Paläontologische*
439 *Zeitschrift*, v. 51, p. 271–287.
- 440 Conway Morris, S., 1986, The community structure of the Middle Cambrian phyllopod bed
441 (Burgess Shale) fauna, British Columbia: *Palaeontology*, v. 29, p. 423–467.
- 442 De Smet, W.H., 2002, A new record of *Limnognathia maerski* Kristensen & Funch, 2000
443 (Micrognathozoa) from the sub-Antarctic Crozet Islands, with redescription of the
444 trophi: *Journal of Zoology*, v. 258, p. 381–393.
- 445 Caron, J-B., and Cheung, B., 2019, *Amiskwia* is a large Cambrian gnathiferan with complex
446 gnathostomulid-like jaws: *Communications Biology*, v. 2, e164.

447 Dong, X-P., 2007, Study on the histology and comparative histology of some protoconodonts,
448 paraconodonts and earliest euconodonts: *Acta Micropaleontologica Sinica*, v. 24, p.
449 113–124.

450 Dunn, C.W. et al., 2008, Broad phylogenomic sampling improves resolution of the animal
451 tree of life: *Nature*, v. 452, p. 745–749.

452 Fröblius, A.C., and Funch, P., 2017, Rotiferan *Hox* genes give new insights into the evolution
453 of metazoan bodyplans: *Nature Communications*, v. 8, e20.

454 García-Bellido, D.C. et al., 2014, A new vetulicolian from Australia and its bearing on the
455 chordate affinities of an enigmatic Cambrian group: *BMC Evolutionary Biology*, v. 14,
456 e214.

457 Hagadorn, J.W., 2002, Burgess Shale-type localities: the global picture, *in* Bottjer, D.J. et al.,
458 eds., *Exceptional Fossil Preservation: A Unique View on the Evolution of Marine Life*:
459 Columbia University Press, New York, p. 91–116.

460 Herlyn, H., and Ehlers, U., 1997, Ultrastructure and function of the pharynx of
461 *Gnathostomula paradoxa* (Gnathostomulida): *Zoomorphology*, v. 117, p. 135–145.

462 Hesselbo, S.P., 1989, The aglaspidid arthropod *Beckwithia* from the Cambrian of Utah and
463 Wisconsin: *Journal of Paleontology*, v. 63, p. 636–642.

464 Hou, X-G., et al., 2004, *The Cambrian fossils of Chengjiang China: The Flowering of Early*
465 *Animal Life*. Blackwell, Oxford.

466 Hu, S-X., et al., 2007, Diverse pelagic predators from the Chengjiang Lagerstätte and the
467 establishment of modern-style pelagic ecosystems in the early Cambrian:
468 *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 254, p. 307–316.

469 Jha, N., Kumar, P., Aggarwal, N., Bhattacharyya, D.D., and Pande, A.C., 2011, The oldest
470 bdelloid Rotifera from early Permian sediments of Chamba Valley: A new discovery:
471 *International Journal of Geology, Earth and Environmental Sciences*, v. 1, p. 23–29.

472 Kouchinsky, A., Bengtson, S., Clausen, S., and Vendrasco, M.J., 2015, An early Cambrian
473 fauna of skeletal fossils from the Emyaskin Formation, northern Siberia: *Acta*
474 *Palaeontologica Polonica* v. 60, p. 421–512.

475 Kristen, R.M., and Funch, P., 2000, Micrognathozoa: A new class with complicated jaws like
476 those of Rotifera and Gnathostomulida: *Journal of Morphology*, v. 246, p. 1–49.

477 Kröger, B., Vinther, J., and Fuchs, D., 2011, Cephalopod origin and evolution: A congruent
478 picture emerging from fossils, development and molecules: Extant cephalopods are
479 younger than previously realised and were under major selection to become agile, shell-
480 less predators: *BioEssays*, v. 33, p. 602–613.

481 Laumer, C.E. et al., 2015, Spiralian phylogeny informs the evolution of microscopic lineages:
482 *Current Biology*, v. 25, p. 2000–2006.

483 Lerosey-Aubril, R., 2015, *Notchia weugi* gen. et sp. nov: A new short-headed arthropod from
484 the Weeks Formation Konservat-Lagerstätte (Cambrian; Utah): *Geological Magazine*,
485 v. 152, p. 351–357.

486 Lerosey-Aubril, R., Hegna, T., Babcock, L.E., Bonino, E., and Kier, C., 2014, Arthropod
487 appendages from the Weeks Formation Konservat-Lagerstätte: New occurrences of
488 anomalocaridids in the Cambrian of Utah, USA: *Bulletin of Geosciences*, v. 89, p. 262–
489 282.

490 Lerosey-Aubril, R., Ortega-Hernández, J., Kier, C., and Bonino, E., 2013, Occurrence of the
491 Ordovician-type aglaspidid *Tremaglaspis* in the Cambrian Weeks Formation (Utah,
492 USA): *Geological Magazine*, v. 150, p. 945–951.

493 Lerosey-Aubril, R., et al., 2012, Controls on gut phosphatisation: The trilobites from the
494 Weeks Formation Lagerstätte (Cambrian, Utah): *PLoS ONE*, v. 7, e32934.

495 Lerosey-Aubril, R. et al., 2018, The Weeks Formation Konservat-Lagerstätte and the
496 evolutionary transition of Cambrian marine life: *Journal of the Geological Society*
497 (London), v. 175: p. 705–715.

498 Marlétaz, F. et al., 2006, Chaetognath phylogenomics: a protostome with deuterostome-like
499 development: *Current Biology*, v. 16, p. R577–R578.

500 Marlétaz, F., Le Parco, Y., 2008, Careful with understudied phyla: the case of chaetognath:
501 *BMC Evolutionary Biology* v. 8, e251.

502 Marlétaz, F., Peijnenburg, K.T.C.A., Goto, T., Satoh, N., and Rokhsar, D.S. 2019, A new
503 spiralian phylogeny places the enigmatic arrow worms among gnathiferans: *Current*
504 *Biology*, v. 29, p. 312–318.e3.

505 Matus, D.Q., Halanych, K.M., and Martindale, M.Q. 2007, The *Hox* gene complement of a
506 pelagic chaetognath *Flaccisagitta enflata*: *Integrative and Comparative Biology*, v. 47,
507 p. 854–864.

508 Moreno, I., and Kapp, H., 2003, Structures of grasping spines and teeth in three species of
509 chaetognaths from Antarctic waters: *Polar Biology*, v. 26, p. 143–150.

510 Muscente, A.D. et al., 2017, Exceptionally preserved fossil assemblages through geologic
511 time and space: *Gondwana Research*, v. 48: p. 164–188.

512 Nanglu, K., and Caron, J-B., 2018, A new Burgess Shale polychaete and the origin of the
513 annelid head revisited: *Current Biology*, v. 28, p. 319–326, e1.

514 Ortega-Hernández, J., Lerosey-Aubril, R., Kier, C. and Bonino, E., 2015, A rare non-trilobite
515 artiopodan from the Guzhangian (Cambrian Series 3) Weeks Formation Konservat-
516 Lagerstätte in Utah, USA: *Palaeontology*, v. 58: p. 265–276.

517 Ou, Q. et al., 2012, Evidence for gill slits and a pharynx in Cambrian vetulicolians:
518 implications for the early evolution of deuterostomes: *BMC Biology*, v. 10, e81.

519 Ou, Q. et al., 2015, A vanished history of skeletonisation in Cambrian comb jellies: *Science*
520 *Advances*, v. 1, e1500092.

521 Papillon, D. et al., 2006, Systematics of Chaetognatha under the light of molecular data, using
522 duplicated ribosomal 18S DNA sequences: *Molecular Phylogenetics and Evolution*, v.
523 38, p. 621–634.

524 Perez, Y., Rieger, V., Martin, E., Müller, C.H.G., and Harzsch, S., 2013, Neurogenesis in an
525 early protostome relative: Progenitor cells in the ventral nerve center of chaetognath
526 hatchlings are arranged in a highly organized geometrical pattern: *Journal of*
527 *Experimental Zoology B: Molecular and Developmental Evolution*, v. 320, p. 179.

528 Poinar, G.O., and Ricci, C., 1982, Bdelloid rotifers in Dominican amber: Evidence for
529 parthenogenetic continuity: *Experientia*, v. 48, p. 408–410.

530 Pyle, L.J., Narbonne, G.M., Nowlan, G.S., Xiao, S-H., and James, N.P., 2006, Early
531 Cambrian metazoan eggs, embryos, and phosphatic microfossils from northwestern
532 Canada: *Journal of Paleontology*, v. 80, p. 811–825.

533 Reich, M., and Smith, A.B., 2009, Origins and biomechanical evolution of teeth in echinoids
534 and their relatives: *Palaeontology*, v. 52, p. 1149–1168.

535 Reich, M., Stegemann, T.R., Hausmann, I.M., Roden, V.J., and Nützel, A., 2018, The
536 youngest ophiocistioid: a first Palaeozoic-type echinoderm group representative from
537 the Mesozoic: *Palaeontology*, v. 61, p. 803–811.

538 Riedl, R., and Rieger, R.M., 1972, New characters observed on isolated jaws and basal plates
539 of the family Gnathostomulidae (Gnathostomulida): *Zeitschrift für Morphologie der*
540 *Tiere*, v. 72, p. 131–172.

541 Rieger, R.M., and Tyler, S., 1995, Sister-group relationship of Gnathostomulida and Rotifera-
542 Acanthocephala: *Invertebrate Biology*, v. 14, p. 186–188.

543 Riemann, O., and Ahlrichs, W.H., 2008, Ultrastructure and function of the mastax in
544 *Dicranophorus forcipatus* (Rotifera: Monogononta): Journal of Morphology, v. 269, p.
545 698–712.

546 Robison, R.A., Babcock, L.E., and Gunther, V.G., 2015, Exceptional Cambrian fossils from
547 Utah: a window into the age of trilobites. Utah Geological Survey, Miscellaneous
548 Publications, v. 15-1, p. 1–97.

549 Shen, X. et al., 2016, Phylomitogenomic analyses strongly suggest the sister relationship of
550 the Chaetognatha and Protostomia: Zoologica Scripta, v. 45, p. 187–199.

551 Shu, D-G., et al., 2017, Multi-jawed chaetognaths from the Chengjiang Lagerstätte
552 (Cambrian, Series 2, Stage 3) of Yunnan, China: Palaeontology, v. 60, p. 763–772.

553 Smith, M.R., 2013, Nectocaridid ecology, diversity and affinity: Early origin of a cephalopod-
554 like body plan: Paleobiology, v. 39, p. 297–321.

555 Smith, M.R., and Caron, J-B., 2010, Primitive soft-bodied cephalopods from the Cambrian:
556 Nature, v. 465: p. 469–472.

557 Smith, M.P., Harvey, T.H.P., and Butterfield, N.J., 2015, The macro- and microfossil record
558 of the Cambrian priapulid *Ottoia*: Palaeontology, v. 58, p. 705–721.

559 Sørensen, M.V., 2002a, On the evolution and morphology of the rotiferan trophi, with a
560 cladistic analysis of Rotifera: Journal of Zoological Systematics and Evolutionary
561 Research, v. 40, p. 129–154.

562 Sørensen, M.V., 2002b, Phylogeny and jaw evolution in Gnathostomulida, with a cladistic
563 analysis of the genera: Zoologica Scripta, v. 31, p. 461–480.

564 Sørensen, M.V., 2003, Further structures in the jaw apparatus of *Limnognathia maerski*
565 (Micrognathozoa), with notes on the phylogeny of the Gnathifera: Journal of
566 Morphology, v. 255, p. 131–145.

567 Sørensen, M.V., and Sterrer, W. 2002, New characters in the gnathostomulid mouth parts
568 revealed by scanning electron microscopy: *Journal of Morphology*, v. 253, p. 310–334.

569 Sørensen, M.V., Sterrer, W., and Giribet, G., 2006, Gnathostomulid phylogeny inferred from
570 a combined approach of four molecular loci and morphology: *Cladistics*, v. 22, p. 32–
571 58.

572 Sterrer, W., 1972, Systematics and evolution within the Gnathostomulida: *Systematic*
573 *Zoology*, v. 21, p. 151–173.

574 Stock, S.R., Ignatiev, K., Lee, P., and Almer, J.D., 2014, Calcite orientations and composition
575 ranges within teeth across Echinoidea: *Connective Tissue Research*, v. 55 (S1), p. 48–
576 52.

577 Szaniawski, H., 1982, Chaetognath grasping spines recognized among Cambrian
578 protoconodonts: *Journal of Paleontology*, v.56, p. 806.

579 Szaniawski, H., 2002, New evidence for the protoconodont origin of chaetognaths: *Acta*
580 *Palaeontologica Polonica*, v. 47, p. 405.

581 Szaniawski, H., 2015, New group of the Early Palaeozoic conodont-like fossils: *Estonian*
582 *Journal of Earth Sciences*, v. 64, p. 91–94.

583 Vannier, J., Steiner, M., Renvoisé, E., Hu, S-X., and Casanova, J-P., 2007, Early Cambrian
584 origin of modern food webs: evidence from predator arrow worms: *Proceedings of the*
585 *Royal Society of London, B*, v. 274, p. 627–633.

586 Vinther, J., and Parry, L.A., 2019, Bilateral jaw elements in *Amiskwia sagittiformis* bridge the
587 morphological gap between gnathiferans and chaetognaths: *Current Biology*, v. 29, p.
588 881-888.

589 Waggoner, B.M., and Poinar, G.O., 1993, Fossil habrotrochid rotifers in Dominican amber:
590 *Experientia*, v. 49, p. 354–357.

- 591 Wang, R., Addadi, L., and Weiner, S., 1997, Design strategies of sea urchin teeth: Structure,
592 composition and micromechanical relations to function: *Philosophical Transactions of*
593 *the Royal Society of London, B*, v. 352, p. 469–480.
- 594 Wulfken, D., and Ahlrichs, W.H., 2012, The ultrastructure of the mastax of *Filinia longiseta*
595 (Flosculariaceae, Rotifera): Informational value of the trophi structure and mastax
596 musculature: *Zoologischer Anzeiger*, v. 251, p. 270–278.
- 597 Zhang, H. et al., 2015, Armored kinorhynch-like scalidophoran animals from the early
598 Cambrian: *Scientific Reports*, v. 5, e16521.
- 599 Zhang, H., Maas, A., and Waloszek, D., 2018, New material of scalidophoran worms in
600 Orsten-type preservation from the Cambrian Fortunian Stage of South China: *Journal of*
601 *Paleontology*, v. 92, p. 14–25.
- 602 Zhao, F. et al., 2017, *Orthrozanclus elongata* n. sp. and the significance of sclerite-covered
603 taxa for early trochozoan evolution: *Scientific Reports*, v. 7, e16304.

604

605 **FIGURE CAPTIONS**

606

607 **Figure 1.** Stratigraphic occurrences of *Dakorhachis thambus* (in color version, red) and the
608 oldest chaetognaths (in color version, green) and protoconodonts (in color version, blue).
609 Cambrian chaetognaths *Eognathacantha*, *Protosagitta* and USNM 199540.

610

611 **Figure 2.** *Dakorhachis thambus* n.gen. n.sp. from the Weeks Formation (Miaolingian,
612 Guzhangian), Utah, USA. Scanning electron micrographs in backscatter mode of polished
613 sections (uncoated) of UU15101.07 (1, 2) and UU15101.08 (3-6). (1) Fossil body composed
614 of radiating fans of a chloritic mineral with pseudomorphs of pyrite across upper surface. (2)
615 Detail of fossil body and pseudomorphs. (3) Tooth, composed of calcite. (4) Tooth, and

616 surrounding fossil body. (5) Two teeth, surrounding fossil body including pyrite
617 pseudomorphs. (6) Fossil body with stacked chloritic mineral. Scale bar for all figures 50
618 μm .

619

620 **Figure 3.** *Dakorhachis thambus* n. gen. n. sp. from the Weeks Formation (Miaolingian,
621 Guzhangian), Utah, USA. (1) UU15101.02 (upper) and UU15101.03 (lower); (2)
622 UU15101.04; (3) UU15101.05 (upper) and UU15101.06 (lower); (4) BPM1090; (5)
623 UU15101.01 (holotype); (6) UU18056.27; (7) UU17122.03; (8) UU18056.28. Specimens
624 photographed (1-5) dry or (6-8) immersed in dilute ethanol. Scale bars are (1, 5) 5 mm; (2-4,
625 6-8) 2 mm.

626

627 **Figure 4.** Feeding apparatus of *Dakorhachis thambus* n. gen. n. sp. in specimens (1)
628 UU15101.01 (holotype; CT images, Fig.5) and (3) UU15101.02 with (2, 4) corresponding
629 camera lucida drawings. Body (blue/light gray), teeth exterior view (red/very dark gray),
630 interior view (pink/fairly dark gray), V-shaped units (green/dark gray), rods (yellow/very pale
631 gray), adhesive (grey/darkish gray), oxides (hatched), sediment (white). Scale bars (1, 3) are 1
632 mm.

633

634 **Figure 5.** Holotype (UU15101.01) of *Dakorhachis thambus* n. gen. n. sp. (1) micro-CT
635 volume rendering, false color represents specimen density. (2) Rotated view showing 3-
636 dimensional transverse banding on the trunk, perpendicular to the long-axis. (3) Detail of
637 teeth. (4) Simplified reconstruction. Scale bar is 5 mm.

638

639 **Figure 6.** Electron micrographs of the feeding apparatus of the holotype (UU15101.01) of
640 *Dakorhachis thambus* n. gen. n. sp.. (1) overview and (2) detail showing the hollow tooth
641 interior and fibrous microstructure. Scale bar is 500 μ m.

642

643 **Figure 7.** SEM of the body trunk surface of *Dakorhachis thambus* n. gen. n. sp. specimen
644 UU15101.01 showing iron oxides layer (black arrow) and the imprints of pseudomorphs of
645 iron oxides after pyrite on the segmented chloritic surface (white arrows). Scale bar 0.5 mm.

646

647 **Figure 8.** Hypothetical transitions between the jaw apparatus of *Dakorhachis thambus* n. gen.
648 n. sp. and (a) those of the chaetognaths (and protoconodonts) via forms similar to *Ankalodous*
649 *sericus* Shu et al. and (b) the gnathiferans (as represented by the gnathostomulids) via forms
650 similar to *Amiskwia sagittiformis* Walcott.