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51 **ABSTRACT**

52

53 **Aim**

54 The sorting of functional traits along environmental gradients is an important driver of  
55 community and landscape scale patterns of functional diversity. However, the significance of  
56 environmental factors in driving functional gradients within biomes and across continents  
57 remains poorly understood. Here, we evaluate the relationship of soil nutrients and climate to  
58 leaf traits in grasses (Poaceae) that are hypothesised to reflect different strategies of resource-  
59 use along gradients of resource availability.

60

61 **Location**

62 Global

63

64 **Taxon**

65 Poaceae

66

67 **Methods**

68 We made direct measurements on herbarium specimens to compile a global dataset of  
69 functional traits and realised environmental niche for 279 grass species that are common in  
70 grassland and savanna biomes. We examined the strength and direction of correlations  
71 between pairwise trait combinations and measured the distribution of traits in relation to  
72 gradients of soil properties and climate, while accounting for phylogenetic relatedness.

73

74 **Results**

75 Leaf trait variation among species follows two orthogonal axes. One axis represents leaf size  
76 and plant height, and we showed positive scaling relationships between these size-related  
77 traits. The other axis corresponds to economic traits associated with resource acquisition and  
78 allocation, including leaf tensile strength (LTS), specific leaf area (SLA) and leaf nitrogen  
79 content (LNC). Global-scale variation in LNC was primarily correlated with soil nutrients,  
80 whilst LTS, SLA and size related traits showed weak relationships to environment.  
81 However, most of the trait variation occurred within different vegetation types, independent  
82 of large-scale environmental gradients.

83

84 **Main conclusions**

85 Our work provides evidence among grasses for relationships at the global scale between leaf  
86 economic traits and soil fertility, and for an influence of aridity on traits related to plant size.  
87 However, large unexplained variance and strong phylogenetic signal in the model residuals  
88 imply that at this scale the evolution of functional traits is driven by factors beyond  
89 contemporary environmental or climatic conditions.

90

91 **Keywords:** functional traits, soil fertility, climate, phylogenetic conservatism, biomes,  
92 grasses

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## 101 INTRODUCTION

102  
103 Functional traits govern competitive interactions and differences in growth and  
104 survival, which are responsible for variation in abundance and distributions across  
105 environmental gradients. Plant functional types (for example evergreen, deciduous, C<sub>3</sub> and  
106 C<sub>4</sub>) have been widely used to group plants from geographically separate communities which  
107 are thought to share response to environmental variation (Woodward, Lomas, & Kelly,  
108 2004). However, there can be significant variation in functional traits within these plant  
109 functional groups (Liu, Edwards, Freckleton, & Osborne, 2012). Crucially, the environmental  
110 responses and biotic interactions of a plant functional type may not apply across all  
111 ecological settings in which it is found (Keith, Holman, Rodoreda, Lemmon, & Bedward,  
112 2007), and there is growing interest in how traits vary within plant functional types, and how  
113 they differ among and within biomes and continents (Lehmann et al., 2014). To address these  
114 issues, we investigate the relationships between functional traits that reflect different  
115 strategies of resource capture and allocation, and investigate whether these correlate with  
116 environmental gradients across the globe, focusing on the grasses that characterise global  
117 grassy biomes (grasslands and savannas).

118 Functional traits of species contribute to ecosystem function according to their  
119 relative abundance / biomass in the community, so that dominant species contribute the most,  
120 and a number of studies have supported this view (Garnier, Navas, & Grigulis, 2015; Grime,  
121 1998). Globally there are ~1,000 species of grass that are dominant within grassy vegetation  
122 types in at least part of their range, and their dominance may reflect the evolution of  
123 particular sets of functional traits that give each species advantages in terms of competition  
124 and survival (Edwards, Osborne, Strömberg, Smith, & Consortium, 2010). Physiological and  
125 morphological constraints mean there are limits to the trait combinations that a species can  
126 deploy, resulting in economic trade-offs between the investment of resources (i.e. water,  
127 light, nutrients and CO<sub>2</sub>) in fast, but cheaply constructed leaves, versus the conservation of  
128 these resources in slow growing, yet long lived tissues (Diaz et al., 2004; Grime et al., 1997;  
129 Ian J. Wright et al., 2004). It has been proposed that trade-offs reflecting differences in the  
130 way plants acquire and allocate resources to growth or conservation of tissues provide  
131 mechanisms that can determine distribution patterns across resource gradients (Fine et al.,  
132 2006; Herms & Mattson, 1992).

133 In environments where resources are limited, species are predicted to have a slow  
134 growth rate, high investment in carbon-based compounds, low leaf nitrogen content (LNC),  
135 long leaf lifespan and low specific leaf area (SLA) (Craine et al., 2002; Grime et al., 1997;  
136 Reich, Walters, & Ellsworth, 1997; Westoby, Falster, Moles, Vesk, & Wright, 2002). These  
137 traits reflect the high cost of tissue loss to for example herbivory, for individuals where  
138 growth is resource limited (Coley, Bryant, & Chapin, 1985; Herms & Mattson, 1992).  
139 Conversely in resource-rich environments (where water, light or nutrients are not limiting),  
140 community assembly is determined by the ability to rapidly acquire and allocate resources to  
141 growth and thereby out-compete neighbouring individuals (Grime, 1977). Traits including  
142 low investment in secondary metabolites, high SLA, high maximum photosynthetic rate,  
143 short leaf lifespan, high relative growth rate and high LNC are predicted to promote  
144 dominance in environments where resource availability does not limit growth (Craine et al.,  
145 2002; Grime et al., 1997; Reich et al., 1997; Westoby et al., 2002) .

146 Functional traits that reflect different strategies of resource acquisition and allocation  
147 correlate strongly with resource availability at the community scale (Katabuchi, Kurokawa,  
148 Davies, Tan, & Nakashizuka, 2012; Kraft & Ackerly, 2010). Recent studies spanning  
149 environmental gradients have shown that similar sorting processes also drive economic trait  
150 distribution in predictable ways at a landscape scale (Asner et al., 2014; Fortunel, Paine, Fine,

151 Kraft, & Baraloto, 2014). However, environmental trait relationships may not vary  
152 predictably across biomes and continents, and can be specific to areas of unique evolutionary  
153 history when considered at larger scales (Knapp et al., 2004; Lehmann et al., 2014). The  
154 relationships between traits and environment at these large scales may, in fact, be in large  
155 part explained by evolutionary history. For example, in tropical forests, trait variation is  
156 phylogenetically partitioned independently of variation in contemporary environmental  
157 conditions (Asner et al., 2014), and yet global-scale analyses rarely consider the role of  
158 evolutionary history when examining the relationships of traits to environment.

159 Current estimates of the global extent of tropical savannas and temperate grasslands  
160 suggest that ~40% of the Earth's terrestrial surface is covered in grassy ecosystems (White,  
161 Murray, & Rohweder, 2000). These store large amounts of carbon, and support livelihoods  
162 and food security globally (Parr, Lehmann, Bond, Hoffmann, & Andersen, 2014). Grassy  
163 biomes are an ideal system for studying the global-scale relationships of functional traits with  
164 environment, as they occur on every continent and most climates. Since these systems are  
165 each typically dominated by relatively few species, these traits are also crucial determinants  
166 of ecosystem function. Here, we investigate the global distribution of functional traits linked  
167 to resource economics in grassy systems, and their relationship to soil fertility and climate,  
168 whilst accounting for the role of evolutionary history in trait distribution. We first measure  
169 the strength and direction of pairwise correlations between traits to test whether co-variation  
170 is consistent with the hypothesis of trade-offs. Secondly, we investigate the distribution of  
171 traits in relation to the environment, testing whether ecological theory explaining the sorting  
172 of species among communities at the landscape scale can be applied to explain the equivalent  
173 sorting along global environmental gradients. Specifically, we test the hypothesis that  
174 dominant species in resource rich grassy environments are characterised by traits associated  
175 with fast acquisition and the maximum allocation of resources to growth. Conversely in  
176 environments where soil nutrients or climate limit plant growth we expect dominant species  
177 to exhibit traits that reflect the conservation of long-lived tissues.

178

## 179 **MATERIALS AND METHODS**

180

### 181 **Species sampling**

182 A global database of the species that characterise grassy biomes was compiled from  
183 regional maps of potential vegetation (Lehmann et al., 2019) using the taxonomy of  
184 Grassbase (Clayton, 2006 onwards). Within the map of Lehmann et al grassy vegetation was  
185 defined as grasslands, savannas or woodland with a continuous grassy underlayer. Grassy  
186 vegetation types and also their characteristic grass species were identified from the  
187 description and metadata associated within original vegetation maps. These species were  
188 therefore based on the expert opinion of vegetation mappers, however, they showed good  
189 correspondence to lists of dominant species generated for each vegetation units from plot  
190 survey datasets (Lehmann et al., 2019). The maps documented 1635 grassy vegetation types,  
191 characterised by, 1154 species of which 841 were identified to species level. This map was  
192 used as the basis for our species sampling (Appendix S1 in Supporting Information).

193 We first generated a randomly ordered list, without replacement, of the 841 globally  
194 dominant grass species. Random draws were weighted by the area over which each species is  
195 common, to ensure that globally important species were represented higher up the list. We  
196 searched the herbarium collection of the Royal Botanic Gardens, Kew, for as many species as  
197 time would feasibly allow, starting from the top of the list and working down. We were able  
198 to search for herbarium specimens for the first 300 species on the list, of which 279 were  
199 present within the herbarium and could therefore be included in our sample. These 279  
200 species represent around one quarter of the world's dominant grass species and can be found

201 within 1012 of the vegetation types (Appendix S2 in Supporting Information). Vegetation  
202 types were characterised by between 1 to 19 dominant species and on average 4. A map  
203 showing the global distribution of our species sample is in Appendix S3.

204

### 205 **Traits measured and their significance**

206 The following traits were selected for study because they reflect different strategies of  
207 resource use along gradients of resource availability and can be measured from herbarium  
208 specimens. SLA is a good predictor of growth rate (Rees et al., 2010) and reflects the return  
209 on previously acquired resources, since there is a trade-off between quickly growing large,  
210 light-capturing, yet vulnerable leaves and producing strong, long-lived leaves (Westoby et al.,  
211 2002; Westoby & Wright, 2006). Maximum leaf size is associated with light capture (Poorter  
212 & Rozendaal, 2008). Plant height declines along gradients of decreasing moisture and/or  
213 nutrient availability (I. J. Wright, Reich, & Westoby, 2001), and is also thought to reflect  
214 different ecological strategies among species in relation to disturbance (Weiher et al., 1999;  
215 Westoby, 1998). Taller plants compete more effectively for light (Cavender-Bares, Ackerly,  
216 Baum, & Bazzaz, 2004; Tilman, 1988), however, smaller plants may be selected for in highly  
217 disturbed environments since there is a trade-off between fast reproduction and competitive  
218 ability (Westoby, 1998). Foliar nitrogen is positively correlated with maximum  
219 photosynthetic rates (Field, Merino, & Mooney, 1983). Leaf tensile strength is an important  
220 form of defence against herbivory (Choong et al., 1992) and is strongly correlated with leaf  
221 life span (Onoda et al., 2011).

222

### 223 **Trait measurements from herbarium specimens**

224 Protocols for measuring functional traits usually prescribe the use of fresh leaf  
225 material. However, access to a global range of species was not possible from fresh material,  
226 and so we developed methods for taking measurements from herbarium specimens.  
227 Measurements taken from herbarium specimens have the additional advantage of being from  
228 plants grown in their native range under natural soil and climatic conditions. Prior to  
229 gathering our data, we conducted preliminary tests (Appendix S4 in Supporting Information),  
230 showing that measurements taken from fresh grass leaves correlate strongly with those from  
231 rehydrated herbarium material for SLA ( $r^2 = 0.90$ ) and LTS ( $r^2 = 0.84$ ) (Appendix S4 in  
232 Supporting Information). LNC is typically measured using dried leaf material and can be  
233 estimated directly from dried herbarium samples.

234

### 235 **Trait measurements**

236 Herbarium specimens were only selected for sampling from areas where the species  
237 formed a dominant part of the vegetation. Herbarium sheets were also selected, where  
238 possible, to be distributed along the extent of the range where each species was dominant.

239 *SLA*: A full leaf where possible or, if not, a section of leaf was removed from the  
240 herbarium sheet, weighed using a five-point balance, and rehydrated for 24 hours in distilled  
241 water. The rehydrated leaf was photographed and the one-sided surface area calculated using  
242 image analysis software (WINDIAS, Delta-T Devices Ltd, Cambridge, UK). The SLA  
243 (expressed in  $\text{cm}^2 \text{g}^{-1}$  of dry mass) was calculated by dividing the value of the leaf area by the  
244 dry leaf mass. *LNC*: Leaf material was ground to a fine powder for 15 minutes at 25 beats per  
245 second using a tissue lyser (Tissuelyser II, Qiagen, Netherlands). Between 10-20mg was  
246 weighed into tin capsules and analysed using an elemental analyser (Vario EL Cube,  
247 Elementar, Germany). *LTS*: Leaves collected from herbarium specimens were rehydrated in  
248 distilled water for 24 hours. A section cut away from the midrib was clamped using a texture

249 analyser (Lloyds TA500, AMETEK Test & Calibration Instruments), and the force measured  
 250 at point of tearing (expressed in MPa). Three replicates per species were measured.

251 Values for maximum culm height, leaf length and leaf width were established from  
 252 GrassBase, the Kew taxonomic database (Clayton, 2006 onwards). The former was used as a  
 253 measure of plant height, while maximum leaf length and width were used to estimate leaf  
 254 size, assuming an elliptical shape.

255

256 **Environmental variables**

257 All mapping of environmental variables was implemented in R (Core Development  
 258 Team R, 2016). Global maps of the total topsoil exchangeable bases, soil pH, topsoil and sand  
 259 content (an indicator of drainage), were obtained from the Harmonized World Soils Database  
 260 (IIASA, 2008). These were used to calculate the mean soil pH, percentage topsoil sand content  
 261 and total topsoil exchangeable bases (a measure of fertility, and hereafter referred to as “soil  
 262 nutrients”), across the geographical area in which each species dominated grassy vegetation.  
 263 The total topsoil exchangeable bases is defined as the sum of exchangeable cations, including  
 264 sodium (Na<sup>+</sup>), calcium (Ca<sup>2+</sup>), magnesium (Mg<sup>2+</sup>) and potassium (K<sup>+</sup>).

265 Global data for nineteen climatic variables was obtained from the Worldclim database  
 266 (Hijmans, Cameron, Parra, Jones, & Jarvis, 2005) and summarised as a mean for each species  
 267 across the geographical area in which it dominated vegetation. Climatic variables can be  
 268 highly correlated with one another. We therefore used principal components analysis (PCA)  
 269 to reduce the dimensionality of the nineteen climate variables to axes that describe general  
 270 patterns. The full results of the PCA are in Table 1. In summary, the first six principal  
 271 component (PC) axes accounted for 95% of the total climatic variation and were used in  
 272 multiple regression models of trait ~ soil + climate. PC1 was a gradient relating to  
 273 temperature, PC2 was an axis of dryness and diurnal temperature range. PC3 relates to  
 274 precipitation of the warmest and wettest months. PC4 is an axis of temperature and  
 275 isothermality, PC5 is a gradient of temperature in combination with precipitation, and PC6 is  
 276 a gradient of temperature, moisture and temperature range (Table 1).

277

278

279 **Table 1.** Climate variables with the highest loadings following principal components analysis  
 280 (PCA). Mean diurnal range is the mean of monthly (max temp - min temp) and isothermality  
 281 is the mean diurnal range/ temperature annual range (\*100).

<u>Highest Loading Climate Variables</u>		
	<u>Negatively Loading</u>	<u>Positively Loading</u>
PC1	Mean annual temperature, Min. temperature of coldest Month, Mean temperature of coldest quarter	Temperature seasonality
PC2	Precipitation of Driest Quarter, Precipitation of driest month	Mean Diurnal Range
PC3	Precipitation of Warmest Quarter, Precipitation of Wettest Month	Precipitation of Driest Month
PC4	Mean Temperature of Warmest Quarter	Isothermality
PC5	Precipitation of Warmest Quarter	Precipitation of coldest quarter
PC6	Mean temperature of wettest quarter	Temperature annual range

282

283

284 **Phylogenetic hypothesis**

285 A Bayesian distribution of one hundred phylogenetic trees was constructed of  
 286 hypothesised relationships between all 279 dominant species in this study, including 94 C<sub>3</sub>

287 and 185 C<sub>4</sub> species. Molecular data from 39 genes for all Poaceae species present in Genbank  
288 was downloaded using PHLAWD (Smith & Dunn, 2008) in April of 2014 to build an initial  
289 phylogeny including all grass species with sufficient genetic coverage (Forrestel et al.  
290 unpublished). There was no genetic data available for 66 of the species included in the study,  
291 and these species were therefore included using a set of taxonomic constraints based on  
292 existing expert knowledge of grasses. The phylogeny of Christin et al. (2014) was utilized as  
293 a dated backbone, and the methods of (Jetz, Thomas, Joy, Hartmann, & Mooers, 2012) were  
294 employed to insert taxa for which there was no genetic data available using the “pastis”  
295 package in R (Thomas et al., 2013) . One hundred trees from the final Bayesian distribution  
296 of phylogenies were subsequently pruned down to the 279 species included in our study. The  
297 trees are deposited in the Dryad database.

298

### 299 **Statistical analyses**

300 We first investigated the relationships among traits using a PCA to identify the main  
301 axes of variation between SLA, LTS, LNC, maximum culm height, maximum leaf area and  
302 maximum leaf width. To verify whether trade-offs operate at a global scale in this plant  
303 group, as Reich et al. (1997), Ian J. Wright et al. (2004), and Díaz et al. (2015) have all  
304 shown across all plant groups, we used a phylogenetic generalised least squares (PGLS)  
305 model to determine the strength and direction of correlations between all combinations of  
306 pairwise plant traits. PGLS accounts for phylogenetic autocorrelation in model residuals that  
307 is expected due to common ancestry.

308 We also used a PGLS model to investigate associations between the traits and  
309 environment. We fitted two models; trait ~ nutrients + sand + pH + PC1 + PC2 + PC3 + PC4  
310 + PC5 + PC6 and its results are shown in Tables 3 and 4. Secondly, to evaluate whether traits  
311 differed systematically between continents, we fitted continent as a factor in the PGLS  
312 models trait ~ soil nutrients\*continent + soil pH \*continent + soil % sand\*continent +  
313 PC1\*continent + PC2\*continent + PC3\*continent + PC4\*continent + PC5\*continent +  
314 PC6\*continent (results in Appendix S8). All model residuals were checked for normality and  
315 logarithmically transformed where necessary. PGLS analyses were performed using the R  
316 package “Caper” (Orme et al., 2012)

317 We measured phylogenetic signal in both the residuals of the models and the  
318 individual traits using Pagel’s Lambda ( $\lambda$ ), which estimates how much trait variation depends  
319 on phylogeny according to a Brownian model of evolution. A  $\lambda$  value of 0 implies no  
320 phylogenetic signal, while a value of 1 indicates phylogenetic dependence consistent with a  
321 Brownian motion model.

322 For all phylogenetic analyses, the tree used was randomly selected from the 100  
323 Bayesian distribution of phylogenies. The analyses were repeated on another five randomly  
324 selected trees to assess sensitivity of our statistical models to phylogenetic uncertainty. We  
325 found no difference in any of the results based on using the different trees and so present  
326 results from a single phylogeny.

327 To assess how much of the trait variation occurred within versus between each of the  
328 grassy vegetation types defined by Lehmann et al (2019), we performed variance partitioning  
329 using the lme function ((Trait ~ 1, random = ~ | vegetation type) and the varcomp function  
330 in the R package “nlme” in R (Pinheiro J, 2017).

331

332



## 333 RESULTS

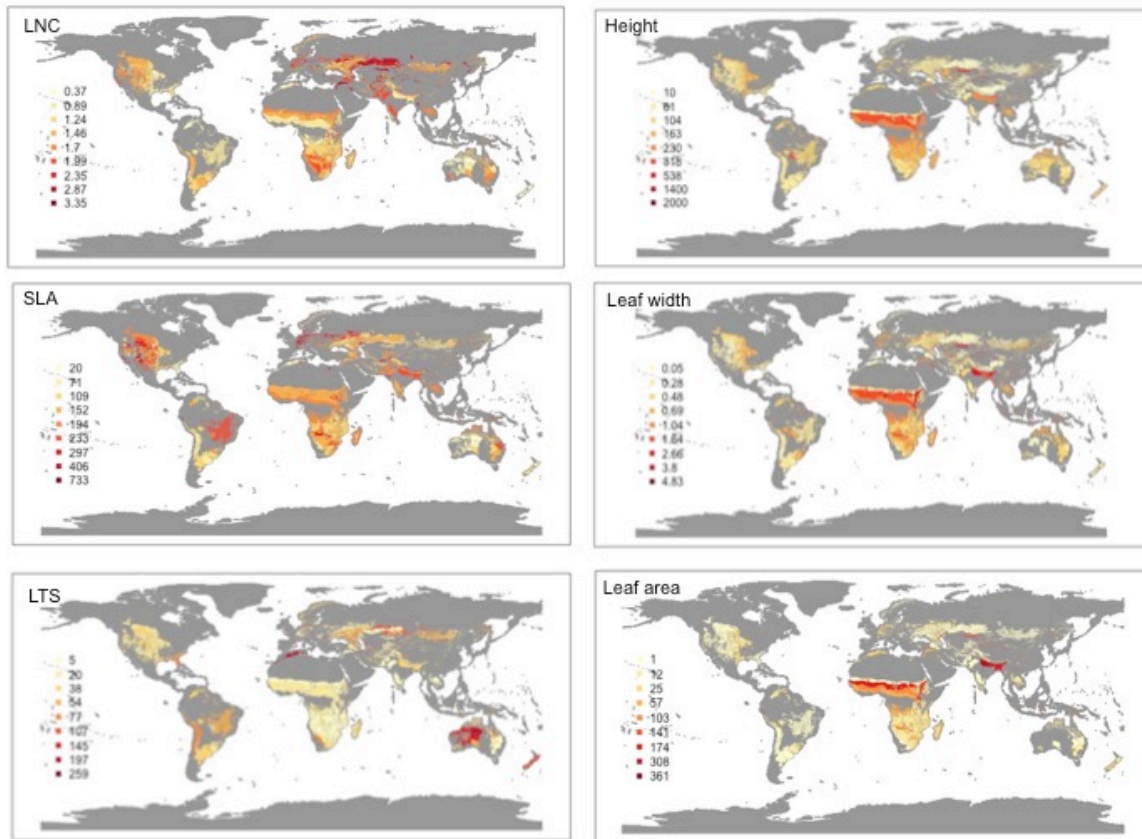
### 334 Geographical and phylogenetic distributions

335 Global patterns in the distribution of traits are shown in Figure 1. Mapping the mean  
336 trait values for dominants in each of the vegetation types revealed clear geographic patterns in  
337 trait values. The lowest values of LNC occurred across areas of the tropics (Fig. 1), areas  
338 characterised by very low soil nutrients, low pH, high rainfall and consistently high  
339 temperatures. The highest LNC occurred across the Eurasian Steppe (Fig. 1), a dry region with  
340 high soil fertility and seasonally low temperatures. Interestingly, regions with notably high  
341 SLA included both parts of the North American Great Plains, where there is a continental  
342 climate and high soil fertility, as well as the Brazilian Cerrado where the climate is tropical and  
343 soil nutrients very low. Despite being characterised by high SLA, areas of the Cerrado also  
344 exhibited very low values of LNC (Fig. 1). The toughest-leaved plants were in areas of  
345 Australia and the Eurasian Steppe (Fig. 1), where SLA was also the lowest (Fig. 1). The tallest  
346 and largest-leaved plants were in areas of the tropics, but particularly tropical Africa (Fig.1).

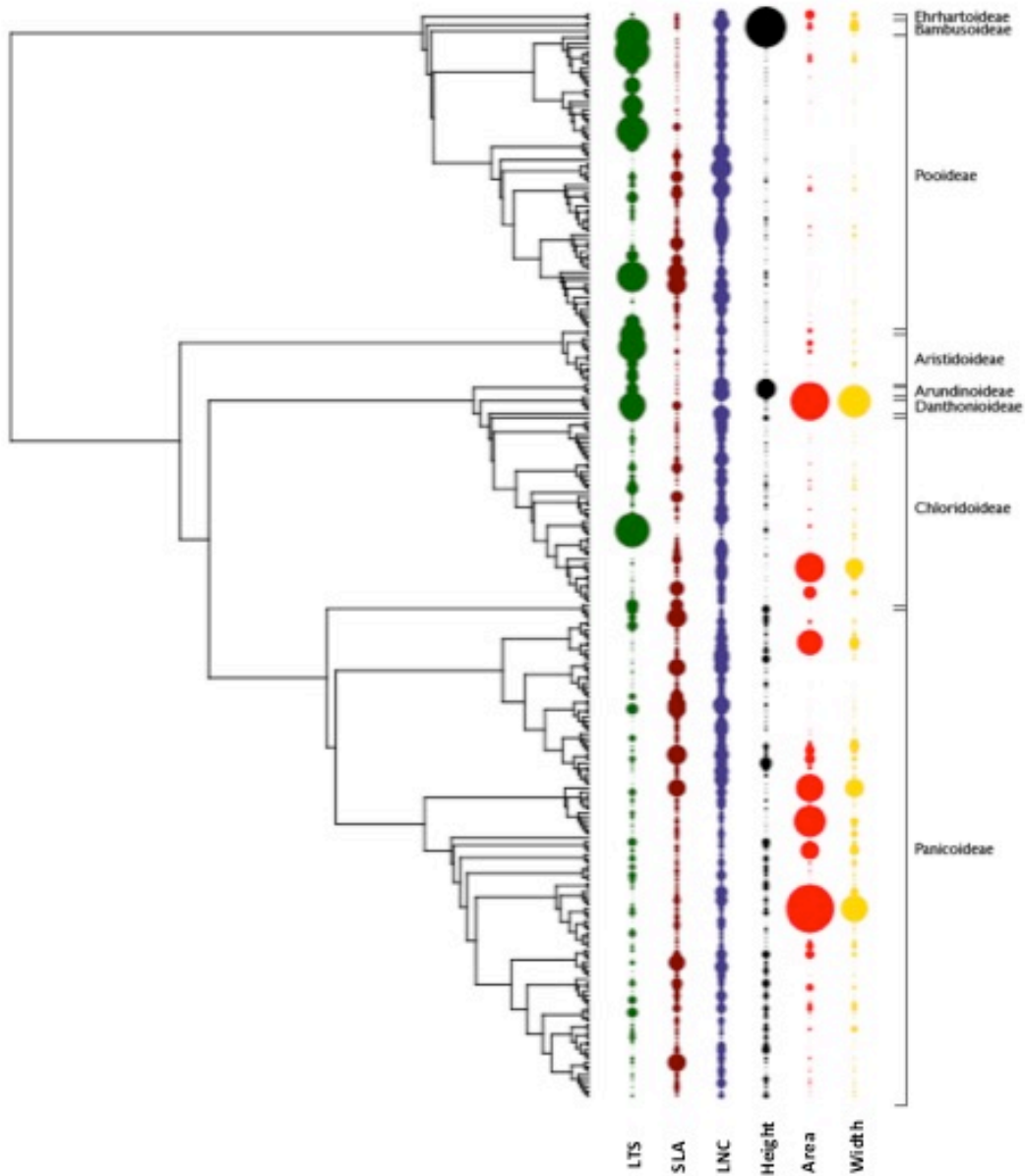
347 The phylogenetic distributions of trait values are shown in Figure 2 and reveal  
348 important differences in traits values between lineages. Individual traits including LNC, LTS,  
349 leaf width and height all showed strong and significant phylogenetic signals ( $P < 0.001$  for  $\lambda =$   
350 0), with  $\lambda$  values ranging from 0.57 to 0.96 (see Table 2). SLA displayed weaker, but  
351 statistically significant phylogenetic dependence ( $\lambda = 0.14$ ). However, maximum leaf area  
352 showed no evidence of a phylogenetic signal ( $\lambda = 0.24$ ).

353 Differences in clade mean height resulted from the divergence between dominant  
354 grasses in the Chloridoideae and Panicoideae lineages with Chloridoideae species being  
355 shortest. Panicoideae and Pooideae lineages were also significantly different in height with  
356 Panicoideae being taller ( $P < 0.001$ ) (Fig. 2; Appendix S5 in Supporting Information). Differences  
357 in clade mean trait values for LTS were most distinct for Danthonioideae species, which are  
358 characterised by the toughest leaves (Fig. 2; Appendix S5). Significant differences in LNC  
359 were also observed between grasses from Pooideae and Panicoideae clades, with Pooideae  
360 grasses having the highest LNC (Fig. 2; Appendix S5). Smaller lineages also contributed to the  
361 phylogenetic signal in all traits (Fig. 2; Appendix S5).

362



363  
 364 **Figure 1.** Global distributions of functional traits. Traits are: (LNC) leaf nitrogen content  
 365 (%), (SLA) specific leaf area ( $\text{cm}^2/\text{g}$ ), (LTS) leaf tensile strength (MPa), (Height) maximum  
 366 culm height (cm), (Leaf width) maximum leaf width (cm), and (Leaf area) maximum leaf  
 367 area ( $\text{cm}^2$ ). Mapping is based on the mean trait values for dominant species in each of the  
 368 grassy vegetation types determined by Lehmann et al. (2019).



369  
 370 **Figure 2.** Functional trait values mapped across the phylogenetic tree. From left to right,  
 371 traits are: leaf tensile strength, LTS (green), SLA (burgundy), % leaf nitrogen content, LNC  
 372 (blue), maximum culm height (black), maximum leaf area (orange) and maximum leaf width  
 373 (yellow) mapped across the phylogenetic tree. A full list of species in the tree is in Appendix  
 374 S1 in supporting information. Dot sizes represent trait values and are scaled to fit the figure,  
 375 which means scaling differs between traits.

376  
 377  
 378  
 379  
 380  
 381  
 382  
 383  
 384

385 **Table 2.** Pagel's  $\lambda$  for the individual traits. All traits except maximum leaf area showed  
 386 strong and significant phylogenetic signal based on a likelihood ratio test against  $\lambda=0$  with 1  
 387 degree of freedom. \* $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ ; *ns* not significant

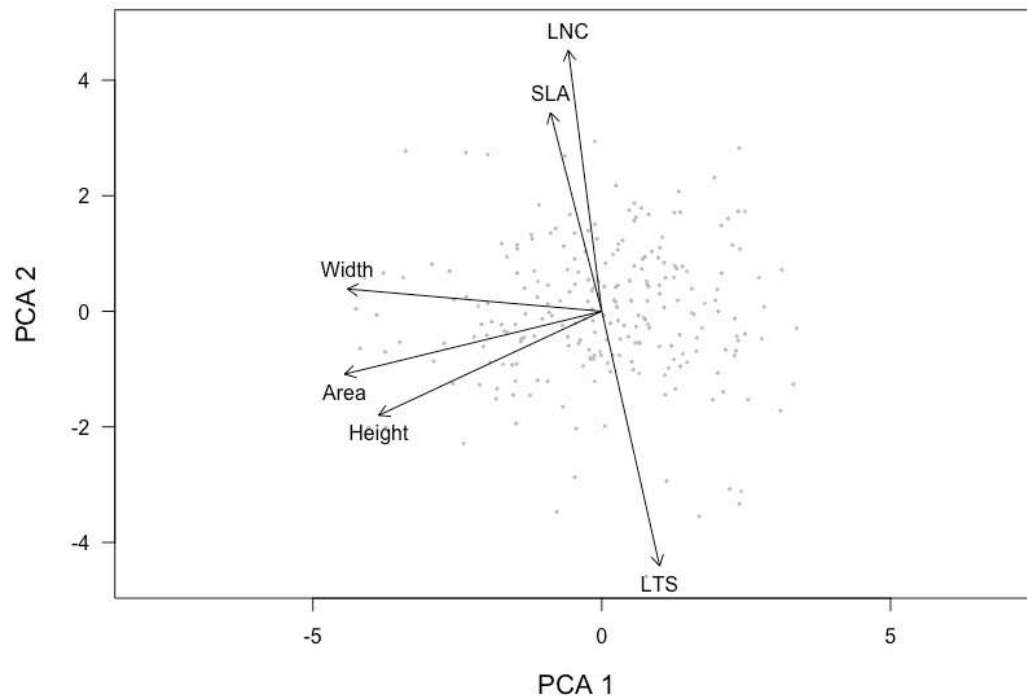
Trait	$\lambda$	P ( $\lambda = 1$ )
LTS	0.77	***
N	0.59	***
SLA	0.14	*
Height	0.96	***
Leaf Area	0.24	N.S
Leaf Width	0.80	***

388  
 389

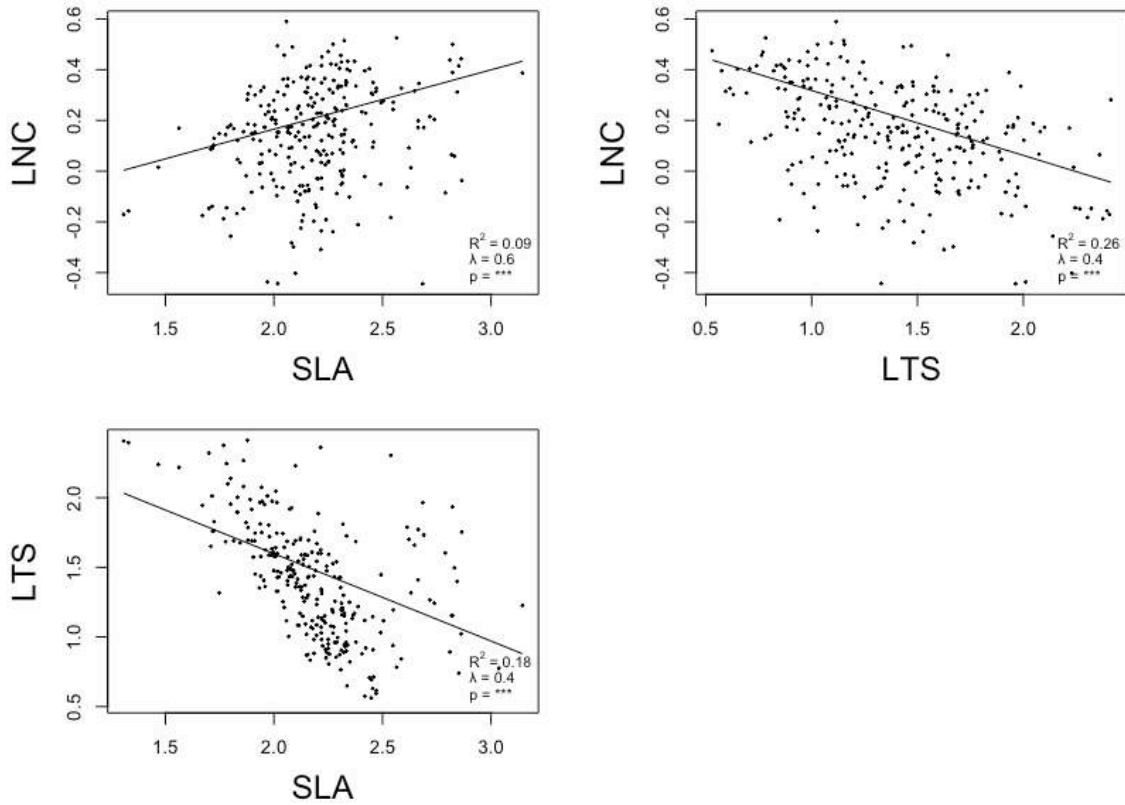
390 **Trait coordination**

391 Traits were separated on two orthogonal axes of variation (Fig. 3). One was identified  
 392 as an axis corresponding to size-related traits including maximum culm height, maximum  
 393 leaf width and maximum leaf area (Fig. 3). Orthogonal to this axis was an axis of resource  
 394 capture and usage, and ranged from low to high SLA and LNC and high to low LTS, all traits  
 395 corresponding to the leaf economic spectrum (Fig. 3). PC1 accounted for 35% of the total  
 396 variance, PC2 accounted for 29%, PC3 14%, PC 4 10% and PC5 6% (Appendix 6a in  
 397 Supporting Information). The loadings of traits on each axis are reported in Appendix 6b  
 398 (Supporting Information).

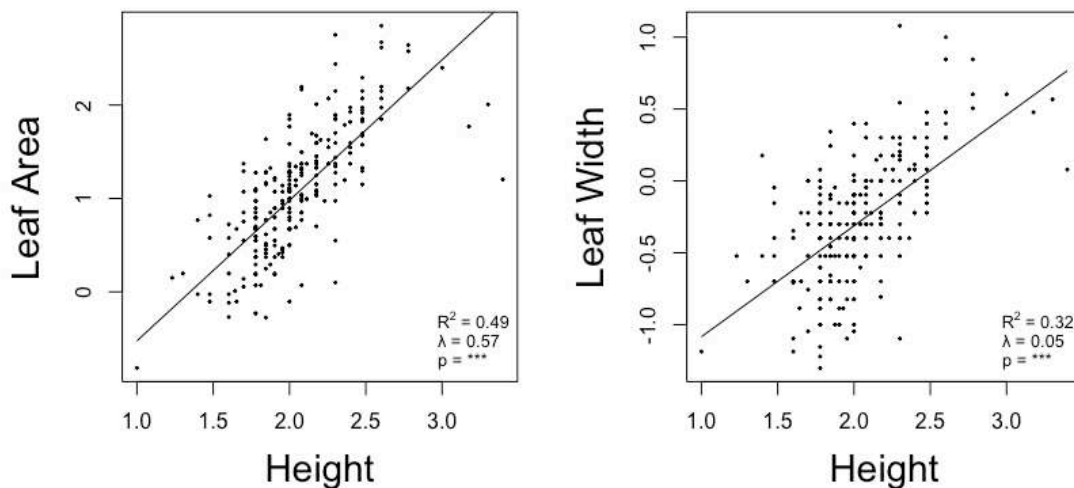
399 Leaf economic traits all showed a statistically significant association with each other  
 400 in the PGLS analysis. SLA and LNC were positively correlated (Fig. 4; Appendix S7  
 401 Supplementary Information). In contrast, SLA and LNC were negatively correlated with LTS  
 402 (Fig. 4; Appendix S7). There was also a strong association among size-related traits, which  
 403 all showed positive relationships (Fig. 5; Appendix S7). A weaker relationship was observed  
 404 between leaf width and SLA, LNC and LTS, and between LTS and maximum culm height  
 405 (Appendix S7).



406  
 407 **Figure 3** Principal components analysis (PCA) of traits, including specific leaf area (SLA),  
 408 leaf tensile strength (LTS), leaf nitrogen content (LNC), maximum culm height (Height),  
 409 maximum leaf area (Area) and maximum leaf width (Width). Orthogonal axes of trait  
 410 variation are identified involving leaf economic traits and traits relating to size. Arrows  
 411 represent the direction of increase of the trait values.  
 412



413  
 414 **Figure 4** Relationship between pairwise combinations of the leaf economic traits, specific  
 415 leaf area (SLA), leaf tensile strength (LTS) and leaf nitrogen content (LNC). Regression lines  
 416 result from PGLS models of pairwise traits. All trait values are logarithmically transformed.  
 417 Lambda values range between 0 and 1. Values closer to 1 indicate higher phylogenetic signal  
 418 in the residuals of the models.  
 419



420  
 421 **Figure 5** Relationship between pairwise combinations of the size traits: maximum leaf area,  
 422 maximum culm height and maximum leaf width. Regression lines result from PGLS models  
 423 of pairwise traits. All trait values are logarithmically transformed. Lambda values range  
 424 between 0 and 1. Values closer to 1 indicate higher phylogenetic signal in the residuals of the  
 425 models.

## 426 **Traits and environment**

427 ANOVA revealed that, with the exception of the relationship between precipitation and LNC,  
428 there was no significant difference in the slopes of the linear model fits for the different  
429 continents (Appendix 8 in Supporting Information). The interaction was therefore dropped  
430 from all subsequent analyses. The only leaf economic trait that showed moderately strong  
431 and significant associations with climate and soil was LNC ( $r^2 = 0.20$ ) (Table 3). Soil  
432 nutrients made a significant contribution to the explanatory power of the model for LNC  
433 ( $P < 0.001$ ) as did PC axes 2 (dryness and diurnal temperature), PC5 (temperature in  
434 combination with precipitation) and PC 4 (temperature and isothermality) ( $P < 0.05$ ) (Table.  
435 3). The remaining traits show hardly any relationship to environment, with very little of the  
436 variation in LTS and SLA explained by environmental gradients ( $r^2 = 0.05$  and  $0.03$   
437 respectively). Only soil nutrients significantly contributing to the explanatory power of the  
438 model for LTS. Soil pH and % sand were significant predictors of SLA although this  
439 association was again very weak (Table 3). Size-related traits were barely associated with  
440 environment, maximum culm height ( $r^2 = 0.05$ ), maximum leaf width ( $r^2 = 0.08$ ) and  
441 maximum leaf area ( $r^2 = 0.06$ ), PC2 (dryness and diurnal temperature) was a significant  
442 predictor of the variation in both height and leaf width (Table 3). Leaf area and leaf width  
443 were significantly influenced by PC 1 (temperature) and PC 3 (precipitation and temperature)  
444 (Table 4). There were strong phylogenetic signals in the residuals of the model for all of the  
445 leaf economic spectrum traits, including LNC ( $\lambda = 0.64$ ), LTS ( $\lambda = 0.60$ ), SLA ( $\lambda = 0.42$ )  
446 (Table 3), as well as height ( $\lambda = 0.69$ ), maximum leaf area ( $\lambda = 0.55$ ) and maximum leaf  
447 width ( $\lambda = 0.59$ ) (Table 4).

448 Variance partitioning was used to compare how much of the trait variation occurred  
449 within and between each of the grassy vegetation types defined by Lehmann et al. (2019).  
450 This showed that 60% of variation in LTS occurred within rather than between vegetation  
451 types, and a large amount of variation within the vegetation types was also evident for SLA  
452 (95%), LNC (64%), maximum culm height (55%), maximum leaf area (83%) and maximum  
453 leaf width (81%), suggesting that global- and regional-scale changes in environment are not  
454 key drivers of variation in grass traits.

455 **Table 3.** Relationship between species means of leaf economic traits relating to resource capture and release and environmental predictors of  
 456 geographical trait variation. The full model is defined as trait ~ soil nutrients +soil pH + soil % sand + PC1+PC2+PC3+PC4+PC5+PC6. Data  
 457 were logarithmically transformed before tests. \* $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ ; *ns* not significant  
 458

	N			SLA			LTS		
	Slope	S.E	P	Slope	S.E	P	Slope	S.E	P
Soil TEB	0.02	0.01	***	0.02	0.01	**	-0.03	0.01	*
Soil pH	-0.07	0.03	*	-0.13	0.05	**	0.04	0.07	ns
Soil %Sand	0.00	0.00		0.01	0.00	**	-0.01	0.00	ns
PC1	0.00	0.01	ns	0.00	0.01	ns	0.01	0.01	ns
PC2	0.02	0.01	**	0.00	0.01	ns	-0.02	0.02	ns
PC3	-0.01	0.01	ns	-0.01	0.01	ns	0.00	0.02	ns
PC4	0.02	0.01	*	0.01	0.02	ns	-0.04	0.02	ns
PC5	-0.03	0.01	**	0.02	0.02	ns	-0.02	0.03	ns
PC6	0.00	0.01	ns	0.00	0.02	ns	0.03	0.03	ns
Lambda	0.64			0.42			0.60		
r2	0.20			0.03			0.05		

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471 **Table 4** Relationship between variation in species means of traits relating to size and environmental predictors of traits variation from the model  
 472 trait ~ soil fertility +soil pH + soil % sand + PC1+PC2+PC3+PC4+PC5+PC6. Data were logarithmically transformed before tests. \* $P < 0.05$ ; \*\*  
 473  $P < 0.01$ ; \*\*\*  $P < 0.001$ ; *ns* not significant  
 474

	Height			Leaf Width			Leaf Area		
	Slope	S.E	P	Slope	S.E	P	Slope	S.E	P
Soil nutrients	-0.01	0.01	ns	0.00	0.01	ns	-0.01	0.02	ns
Soil pH	-0.08	0.05	ns	-0.04	0.07	ns	-0.10	0.12	ns
Soil %Sand	0.00	0.00	ns	0.00	0.00	ns	0.00	0.01	ns
PC1	-0.02	0.01	ns	-0.03	0.01	**	-0.05	0.02	**
PC2	0.04	0.01	***	0.04	0.02	*	0.05	0.03	ns
PC3	-0.02	0.01	ns	-0.05	0.02	**	-0.07	0.03	*
PC4	0.00	0.02	ns	-0.01	0.02	ns	-0.04	0.04	ns
PC5	0.00	0.02	ns	0.01	0.02	ns	0.02	0.04	ns
PC6	0.01	0.02	ns	0.03	0.03	ns	0.04	0.06	ns
$\lambda$	0.69			0.59			0.55		
r <sup>2</sup>	0.05			0.08			0.06		

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## 476 **DISCUSSION**

477 Using a global comparative analysis of traits from around one quarter of the globally  
478 dominant grass species, we demonstrate that leaf nitrogen content is significantly correlated  
479 with soil nutrients and to a lesser extent climate within the world's grassy biomes. However,  
480 overall, global gradients in the abiotic environment explain a relatively small amount of  
481 variation in commonly measured traits that are thought to reflect trade-offs in the acquisition  
482 and allocation of resources across grassy biomes. Instead, our analyses reveal that large  
483 amounts of trait variation occurs within as opposed to between vegetation types, and we show  
484 strong phylogenetic patterns in the distribution of traits. We were only able to sample a subset  
485 of the dominant grass species and vegetation types across the globe. However, our sample  
486 included multiple species from every continent, which represent a global latitudinal, climatic  
487 and soil gradient including species from all of the major and most minor grass phylogenetic  
488 lineages.

### 489 **Trait relationships**

491 We wanted to establish how leaf economic and size traits are coordinated among  
492 species. The traits of species dominating grassy biomes vary at the global scale along  
493 orthogonal axes of variation previously predicted by theory (Díaz et al., 2015; Grime, 1977;  
494 Sandel, Monnet, & Vorontsova, 2016) providing further evidence for trade-offs being a  
495 fundamental mechanism underlying plant functional strategies at a global scale. One axis  
496 revealed trade-offs between traits associated with the rapid acquisition of resources and  
497 allocation to growth, and traits linked to the conservation of resources in well-defended  
498 tissues, a relationship which is concurrent with other work (Coley et al., 1985; Díaz et al.,  
499 2004; Herms & Mattson, 1992; Reich et al., 1997; Westoby et al., 2002). An orthogonal axis  
500 of variation was identified relating to size and this is also consistent with previous studies that  
501 were conducted across broad taxa (Díaz et al., 2004; Sandel et al., 2016; Westoby, 1998).  
502 Interestingly, our study reveals that traits that are tightly correlated do not necessarily share  
503 the same relationships to environment.

### 504 **Leaf economic traits and environment**

506 We wanted to determine whether the trade-offs underlying plant strategies sort  
507 according to abiotic gradients of soil nutrients and climate at the global scale, and if this  
508 depends upon continent, since the magnitude of trait-environment relationships may vary  
509 between regions with different evolutionary histories (Lehmann et al., 2014). There was a  
510 marginally significant difference between continents in the responses of LNC to  
511 precipitation. However, for all other traits and environmental variables this was non-  
512 significant, showing that relationships between traits and environment are at the global scale  
513 independent of geographical location.

514 Although we found little evidence that continents differed in their trait relationships  
515 with environment, we did find strong evidence that evolutionary history shapes the  
516 distribution of traits. All leaf economic traits corresponding to trade-offs associated with a  
517 long leaf life span (SLA, LNC and LTS) exhibited strong phylogenetic signals in both the  
518 individual traits and the residuals of the models. This finding is consistent with previous work  
519 showing phylogenetic signals in both the traits and habitat associations of grasses (Liu et al.,  
520 2012; Visser, Woodward, Freckleton, & Osborne, 2012).

521 LNC was associated with soil fertility and also climate, with high values of LNC  
522 found in dry climate regions with high diurnal temperature range – i.e. semi-arid or desert  
523 regions. Ordoñez et al. (2009), also showed using published data consisting of mostly trees  
524 and shrubs, that globally an increase in LNC corresponds with increasing soil nutrients, and  
525 that soil influences traits more than climate. Species that are distributed across drier sites are

526 known to have higher LNC, which may be a mechanism for improving leaf water-use  
527 efficiency by increasing investment in photosynthetic proteins and raising CO<sub>2</sub>-fixation for a  
528 given stomatal conductance (Schulze et al., 1998; I. J. Wright et al., 2001). Arid regions also  
529 often coincide with areas of high soil fertility, which exerted a stronger effect on the variation  
530 in LNC in our analysis than climate. The observed increase in LNC with increasing soil  
531 nutrients may therefore be a plastic response to resource availability in the environment, as  
532 opposed to an adaptive strategy. However, we note that, in general, LNC varies more  
533 between species than within them (Kichenin, Wardle, Peltzer, Morse, & Freschet, 2013).  
534 Furthermore, our results show that LNC is highly conserved across the phylogeny, indicating  
535 that this pattern is driven by evolutionary adaptations of species that reflect historical  
536 processes rather than by the contemporary environment.

537 Variation in LTS and SLA were barely associated with soil properties and not at all  
538 with climate. A previous global scale analysis of leaf mechanical properties, which included  
539 forest as well as grassland species, showed the influence of mean annual precipitation on  
540 mechanical properties of leaves to be minimal but did not consider properties of soil (Onoda  
541 et al., 2011). Soil fertility has previously been linked to toughness in leaves at smaller scales  
542 (J Read, Sanson, & Lamont, 2005), which is consistent with theory that predicts better  
543 defended leaves in resource-limited habitats (Coley et al., 1985). Toughening of the leaves  
544 caused by lignin production is commonly observed in plants from arid habitats (Jennifer  
545 Read, Sanson, de Garine-Wichatitsky, & Jaffre, 2006), and it was therefore surprising that the  
546 PC axis describing precipitation did not have a significant effect upon LTS at a global scale  
547 and that the relationship with soil fertility were not stronger.

548 SLA showed the weakest relationship of all the leaf economic traits to environment  
549 and was barely explained by a combination of all measured soil properties. Although we did  
550 sample individuals that represented as much of the geographical range of each species as  
551 possible, we were unable to incorporate intraspecific trait variation into our study. This may  
552 account for some of the weak relationships observed. Traits, and in particular SLA can be  
553 highly plastic in response to environment, and this may explain some of the weakness of our  
554 correlations. However, since interspecific variation contributes less to trait variation than  
555 interspecific variation and in the case of SLA mirrors interspecific differences across  
556 environmental gradients (Carlucci, Debastiani, Pillar, & Duarte, 2015) we believe our results  
557 to be robust. Soil pH and sand content were significant predictors of SLA but did not explain  
558 variation in any other traits, and the relationship of LNC to soil nutrients was much stronger  
559 than that of SLA. This shows, importantly, that highly correlated traits do not necessarily  
560 share the same responses to environmental predictors.

561 SLA had the weakest relationship to environment but also had the highest amount of  
562 variation within vegetation types. Variance partitioning showed that 95% of variance in SLA,  
563 64% of variance in LNC and 40% of variance in LTS, occurred within-vegetation type. A  
564 large part of the variance in these traits therefore occurs at finer scales (i.e. landscape and  
565 habitat patch) than can be explained by climate, a pattern also observed in other studies  
566 which included a broader range of taxa and biomes (Freschet, Cornelissen, van Logtestijn, &  
567 Aerts, 2010; Ian J. Wright et al., 2004). Unexplained variation within vegetation types may  
568 result from phylogenetically correlated environmental factors, as indicated by the strong  
569 phylogenetic signal, that vary at the landscape or community scales and could result from  
570 changes in woody plant cover, fire, herbivory or microsite variation in soil properties and  
571 moisture. Soil nutrients and hydrological properties can vary over small spatial scales that  
572 would not be captured by the resolution of our gridded soil data e.g. (Fridley, Grime, Askew,  
573 Moser, & Stevens, 2011). Furthermore, our measure of soil nutrients does not necessarily  
574 reflect plant nutrient uptake, which can be influenced by other factors including soil structure  
575 and compaction. It is possible that our data may capture broad scale patterns but

576 underestimate fine scale relationships between traits and soils. However, He et al. (2010)  
577 provide evidence that in grasslands soil does not explain trait variation that is unexplained by  
578 climate.

579

### 580 **Size related traits and environment**

581 There was strong allometric scaling between maximum culm height, leaf area and leaf  
582 width, and all size-related traits were weakly correlated with climate, but not soil. Leaves  
583 perform several functions including light capture, water transport and defence, and optimal  
584 leaf size and shape therefore depends on environmental factors such as irradiance, energy  
585 balance, water availability and water loss, as well as biotic interactions such as competition  
586 and herbivory. Smaller leaves have higher major vein density which contributes to drought  
587 tolerance by directing water around blockages caused by drought-induced xylem embolism,  
588 and helping to protect the hydraulic system from damage (Sack et al., 2012). We found  
589 smaller and narrower leaves in drier habitats and larger, wider leaves in warm, humid  
590 regions. Aridity and diurnal temperature range were weakly associated with both maximum  
591 culm height and leaf width. Taller plants were found in the wettest regions, which are also the  
592 most productive areas of the world. Height is an important component of competition as taller  
593 plants are better competitors for light and cast shade on neighbouring individuals. Increased  
594 stature can therefore confer dominance in wet, productive areas where competition is likely  
595 to be most intense. Shorter plants and narrower leaves were found in the driest areas with a  
596 high temperature diurnal range, indicating semi-arid or desert climates. It was therefore  
597 surprising that relationships between size and climate were not stronger over global scales.  
598 As with the leaf economic traits, we found a large amount of small-scale variation in size-  
599 related traits (between 55 and 83% of variation in size-related traits occurred within rather  
600 than between vegetation types). It is likely that unexplained variation in size related traits is  
601 driven by predictors which vary over smaller scales than climate, for example herbivory or  
602 fire.

603

### 604 **Conclusions**

605 Our results demonstrate that leaf traits of the dominant species of grassy biomes vary  
606 along orthogonal axes relating to size, and to resource capture and allocation. Trait  
607 correlations along these axes provide further evidence for trade-offs being a fundamental  
608 mechanism that underlie plant functional strategies at a global scale, however, correlated  
609 traits do not necessarily share the same response to environment. With the exception of LNC,  
610 traits linked to resource economics are barely correlated with global gradients in soil  
611 nutrients. Size-related traits are weakly correlated with climate. After accounting for global  
612 environmental gradients, there remain robust phylogenetic patterns in leaf and size traits,  
613 demonstrating that the trait combinations of dominant grass species depend strongly on their  
614 evolutionary history. There is considerable trait variation among the dominant species within  
615 grassy biomes, such that most trait variation occurs within rather than between different  
616 vegetation types. In combination, these patterns suggest that mechanisms of co-existence and  
617 phylogenetically linked environmental correlates varying over small spatial scales are  
618 important determinates of species occurrence.

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620

### 621 **BIOSKETCH**

622 **Emma Jardine** has worked at Oxford University and at the German Centre for Integrative  
623 Biodiversity Research (iDiv)/FSU. Her research interests focus on understanding variation in  
624 plant functional traits, how plant strategies are associated with different environments and the

625 processes that determine their distribution. This work formed a component of her PhD in the  
626 Animal and Plant Sciences department at the University of Sheffield.

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### 793 **SUPPORTING INFORMATION**

- 794 Appendix S1 Supplementary Methods
- 795 Appendix S2 Grass species included in this study
- 796 Appendix S3 Map showing the distribution of species sampling
- 797 Appendix S4 Correlation between traits measured from fresh leaves and rehydrated  
 798 herbarium specimens
- 799 Appendix S5 Clades with significantly different mean trait values
- 800 Appendix S6 Results of principal components analysis of traits
- 801 Appendix S7 Table showing all pairwise trait correlations
- 802 Appendix S8 ANOVA table comparing slopes of the full model with continent as an  
 803 interaction.  
 804



## SUPPORTING INFORMATION

### Appendix S1 Supplementary methods

The map that was the basis for our species sampling was produced by Lehmann et al (2019), which integrated and reclassified 20 national and regional vegetation maps. These original maps were the products of a combination of botanical surveys, geographic analysis and expert opinion. References for these maps are listed in Lehmann et al (2019). From these data Lehmann et al identified global grassy vegetation types. In total 1635 grassy vegetation types were identified. Grassy vegetation was defined as having > 50% of the relative ground cover or biomass composed of grasses so that the classification of deserts and areas with sparse vegetation cover was not problematic. Vegetation units were considered grassy deserts where the total above-ground biomass was either <50 g m<sup>2</sup>, or where total ground cover was <25%, throughout the year. Any region where grasses were the dominant component of the ground layer, irrespective of tree cover were also included in the map so as to include tropical savannas and woodlands which behave functionally as savanna due to a continuous grassy layer. Areas identified as mosaics of open and closed canopy but with a continuous grassy layer e.g. across the Steppe region of Russia were also classified as grassy. For areas of the world where no other maps were available, the WWF Ecoregions map was used and assessed by the above criteria to re-define units as grassy or otherwise. Artificial vegetation units (ie agricultural units or those planted by humans) were excluded from the map.

The species which characterise each vegetation type were also identified from the same maps and species that were invasive in vegetation types were excluded for the purposes of our study.

### Appendix S2 List of all species included in the study following the taxonomy of GrassBase.

Species	Author
<i>Acroceras macrum</i>	Stapf
<i>Aeluropus lagopoides</i>	(L.) Thw.
<i>Aeluropus littoralis</i>	(Gouan) Parl.
<i>Agropyron cristatum</i>	(L.) J. Gaertn.
<i>Agrostis capillaris</i>	L.
<i>Agrostis leptotricha</i>	E. Desv.
<i>Alloteropsis semialata</i>	(R. Br.) Hitchcock
<i>Andropogon bicornis</i>	L.
<i>Andropogon brazzae</i>	Franch.
<i>Andropogon gayanus</i>	Kunth
<i>Andropogon lateralis</i>	Nees
<i>Andropogon lima</i>	(Hack.) Stapf
<i>Andropogon schirensis</i>	Hochst.
<i>Andropogon selloanus</i>	(Hack.) Hack.
<i>Andropogon tectorum</i>	Schum. & Thonn.
<i>Anthephora argentea</i>	Goossens
<i>Anthephora pubescens</i>	Nees
<i>Anthoxanthum odoratum</i>	L.
<i>Apluda mutica</i>	L.

<i>Aristida adscensionis</i>	L.
<i>Aristida contorta</i>	F. Muell.
<i>Aristida diffusa</i>	Trin.
<i>Aristida jubata</i>	(Arech.) Herter
<i>Aristida junciformis</i>	Trin. & Rupr.
<i>Aristida murina</i>	Cav.
<i>Aristida pallens</i>	Cav.
<i>Aristida purpurea</i>	Nutt.
<i>Aristida rhiniochloa</i>	Hochst.
<i>Aristida rufescens</i>	Steud.
<i>Aristida similis</i>	Steud.
<i>Aristida stricta</i>	Michx.
<i>Arundinella mesophylla</i>	Nees ex Steud.
<i>Arundo donax</i>	L.
<i>Astrebla lappacea</i>	(Lindl.) Domin
<i>Axonopus canescens</i>	(Nees) Pilger
<i>Axonopus compressus</i>	(Sw.) Beauv.
<i>Axonopus fissifolius</i>	(Raddi) Kuhlmann.
<i>Axonopus purpusii</i>	(Mez) Chase
<i>Bambusa polymorpha</i>	Munro
<i>Bambusa tulda</i>	Roxb.
<i>Blepharoneuron tricholepis</i>	(Torr.) Nash
<i>Bothriochloa ischaemum</i>	(L.) Keng
<i>Bothriochloa saccharoides</i>	(Sw.) Rydb.
<i>Bouteloua curtipendula</i>	(Michx.) Torr.
<i>Bouteloua megapotamica</i>	(Spreng) Kuntze
<i>Brachiaria deflexa</i>	(Schum.) C. E. Hubb. ex Robyns
<i>Brachiaria nigropedata</i>	(Fic. & Hiern.) Stapf
<i>Brachiaria serrata</i>	(Thunb.) Stapf
<i>Brachypodium pinnatum</i>	(L.) Beauv.
<i>Briza brizoides</i>	(Lam.) Kuntze
<i>Briza subaristata</i>	Lam.
<i>Bromus auleticus</i>	Trin. ex Nees
<i>Bromus sclerophyllus</i>	Boiss.
<i>Bromus speciosus</i>	Nees
<i>Bromus tectorum</i>	L.
<i>Calamagrostis arundinacea</i>	(L.) Roth
<i>Calamagrostis epigeios</i>	(L.) Roth
<i>Calamagrostis rubescens</i>	Buckl.
<i>Calamagrostis varia</i>	(Schrad.) Host
<i>Calamovilfa longifolia</i>	(Hook.) Scribn.
<i>Cenchrus biflorus</i>	Roxb.
<i>Cenchrus ciliaris</i>	L.
<i>Centropodia glauca</i>	(Nees) T. A. Cope
<i>Chionochloa flavescens</i>	Zotov
<i>Chionochloa pallens</i>	Zotov

<i>Chionochloa rubra</i>	Zotov
<i>Chloris virgata</i>	Sw.
<i>Chondrosum eriopodum</i>	Torr.
<i>Chondrosum gracile</i>	H. B. & K.
<i>Chondrosum hirsutum</i>	(Lag.) Sweet
<i>Chrysopogon aciculatus</i>	(Retz.) Trin.
<i>Chrysopogon fulvus</i>	(Spreng.) Chiov.
<i>Chrysopogon nigritanus</i>	(Benth.) Veldkamp
<i>Cleistogenes squarrosa</i>	(Trin.) Keng
<i>Cortaderia jubata</i>	(Lemoine) Stapf
<i>Ctenium newtonii</i>	Hack.
<i>Cymbopogon caesius</i>	(Hook. & Arn.) Stapf
<i>Cymbopogon distans</i>	(Nees) W. Watson
<i>Cymbopogon flexuosus</i>	(Nees) W. Watson
<i>Cymbopogon giganteus</i>	Chiov.
<i>Cymbopogon nardus</i>	(L.) Rendle
<i>Cymbopogon nervatus</i>	(Hochst.) Chiov.
<i>Cymbopogon pospischilii</i>	(K. Schum.) C. E. Hubb.
<i>Cynodon dactylon</i>	(L.) Pers.
<i>Cynodon incompletus</i>	Nees
<i>Dactylis glomerata</i>	L.
<i>Dactyloctenium aegyptium</i>	(L.) Willd.
<i>Dactyloctenium giganteum</i>	B. S. Fisher & Schweickerdt
<i>Dactyloctenium radulans</i>	(R. Br.) Beauv.
<i>Danthonia californica</i>	Boland.
<i>Dendrocalamus strictus</i>	(Roxb.) Nees
<i>Deschampsia cespitosa</i>	(L.) Beauv.
<i>Deschampsia flexuosa</i>	(L.) Trin.
<i>Desmostachya bipinnata</i>	(L.) Stapf
<i>Dichanthium fecundum</i>	S. T. Blake
<i>Dichanthium foveolatum</i>	(Delile) Roberty
<i>Dichanthium sericeum</i>	(R. Br.) A. Camus
<i>Digitaria abyssinica</i>	(A. Rich.) Stapf
<i>Digitaria brazzae</i>	(Franch.) Stapf
<i>Digitaria californica</i>	(Benth.) Henrard
<i>Digitaria debilis</i>	(Desf.) Willd.
<i>Digitaria eriantha</i>	Steud.
<i>Digitaria macroblephara</i>	(Hack.) Paoli
<i>Digitaria milaniana</i>	(Rendle) Stapf
<i>Diheteropogon amplexans</i>	(Nees) Clayton
<i>Echinochloa colona</i>	(L.) Link
<i>Echinochloa haploclada</i>	(Stapf) Stapf
<i>Echinochloa pyramidalis</i>	(Lam.) Hitchc. & Chase
<i>Echinolaena inflexa</i>	(Poir.) Chase
<i>Eleusine coracana</i>	(L.) Gaertn.
<i>Elionurus muticus</i>	(Spreng.) Kuntze

<i>Enneapogon desvauxii</i>	Beauv.
<i>Entolasia imbricata</i>	Stapf
<i>Eragrostis biflora</i>	Hack. ex Schinz
<i>Eragrostis ciliaris</i>	(L.) R. Br.
<i>Eragrostis curvula</i>	(Schrad.) Nees
<i>Eragrostis cylindriflora</i>	Hochst.
<i>Eragrostis lugens</i>	Nees
<i>Eragrostis neesii</i>	Trin.
<i>Eragrostis obtusa</i>	Munro ex Ficalho & Hiern
<i>Eragrostis superba</i>	Peyr.
<i>Eriochloa fatmensis</i>	(Hochst. & Steud.) Clayton
<i>Exothea abyssinica</i>	(Hochst.) Anderss.
<i>Festuca caprina</i>	Nees
<i>Festuca idahoensis</i>	Elmer
<i>Festuca lenensis</i>	Drobov
<i>Festuca novae-zealandiae</i>	(Hack.) Cockayne
<i>Festuca ovina</i>	L.
<i>Festuca pratensis</i>	Huds.
<i>Festuca quadriflora</i>	Honck.
<i>Festuca valesiaca</i>	Schleich. ex Gaud.
<i>Fingerhuthia africana</i>	Lehm.
<i>Helictotrichon desertorum</i>	(Less.) Pilger
<i>Heteropogon contortus</i>	(L.) Beauv. ex Roem. & Schult.
<i>Heteropogon melanocarpus</i>	(Ell.) Benth.
<i>Hyparrhenia anthistirioides</i>	(Hochst.) Anderss. ex Stapf
<i>Hyparrhenia cymbaria</i>	(L.) Stapf
<i>Hyparrhenia dichroa</i>	(Steud.) Stapf
<i>Hyparrhenia diplandra</i>	(Hack.) Stapf
<i>Hyparrhenia familiaris</i>	(Steud.) Stapf
<i>Hyparrhenia filipendula</i>	(Hochst.) Stapf
<i>Hyparrhenia hirta</i>	(L.) Stapf
<i>Hyparrhenia newtonii</i>	(Hack.) Stapf
<i>Hyparrhenia nyassae</i>	(Rendle) Stapf
<i>Hyparrhenia schimperi</i>	(Hochst.) Anderss. ex Stapf
<i>Hyparrhenia smithiana</i>	(Hook.) Stapf
<i>Hyparrhenia subplumosa</i>	Stapf
<i>Hyperthelia dissoluta</i>	(Nees) Clayton
<i>Imperata cylindrica</i>	(L.) Raeusch.
<i>Ischaemum afrum</i>	(J. F. Gmel.) Dandy
<i>Koeleria glauca</i>	(Spreng.) DC.
<i>Koeleria macrantha</i>	(Ledeb.) Schult.
<i>Leersia hexandra</i>	Sw.
<i>Leptochloa fusca</i>	(L.) Kunth
<i>Leptocoryphium lanatum</i>	(HBK.) Nees
<i>Leymus cinereus</i>	(Scribn. & Merr.)
<i>Leymus racemosus</i>	(Lam.) Tsvelev

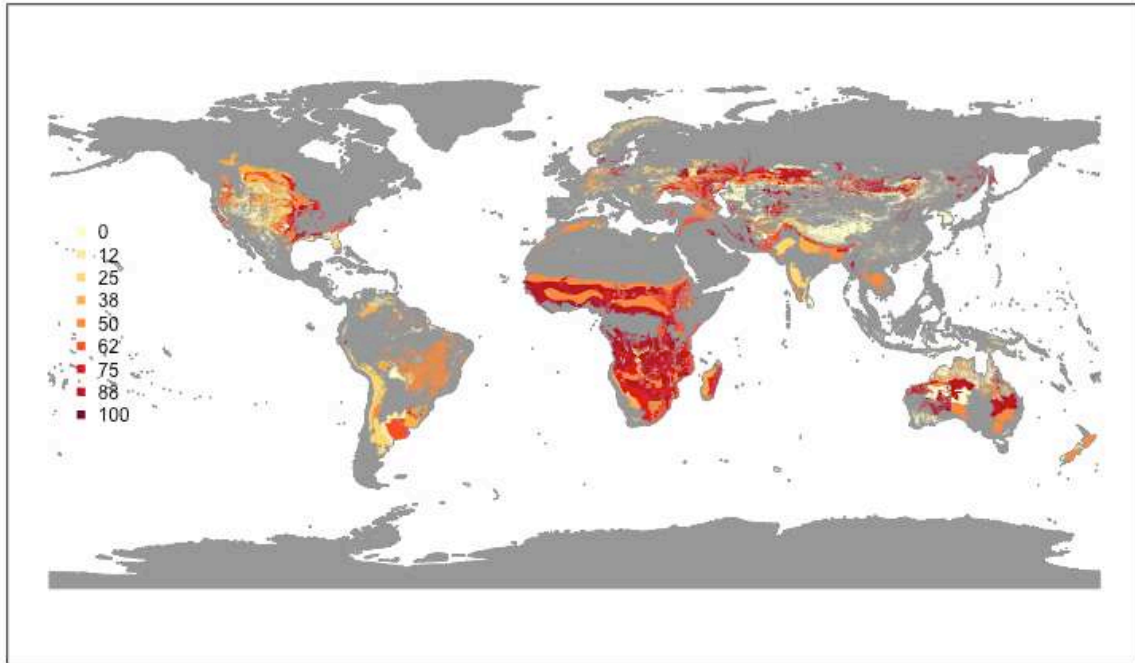
<i>Leymus triticoides</i>	(Buckl.) Pilger
<i>Loudetia arundinacea</i>	(A. Rich) Hochst. ex Steud.
<i>Loudetia phragmitoides</i>	(Peter) C. E. Hubb.
<i>Loudetia simplex</i>	(Nees) C. E. Hubb.
<i>Melica brasiliana</i>	Ard.
<i>Melica minuta</i>	L.
<i>Melica nutans</i>	L.
<i>Melica picta</i>	C. Koch
<i>Melinis amethystea</i>	(Franchet) G. Zizka
<i>Melinis minutiflora</i>	P. Beauv.
<i>Mesosetum loliiforme</i>	(Steud.) Hitchcock
<i>Mesosetum penicillatum</i>	Mez
<i>Microchloa caffra</i>	Nees
<i>Milium effusum</i>	L.
<i>Monocymbium cerasiiforme</i>	(Nees) Stapf
<i>Muhlenbergia richardsonis</i>	(Trin.) Rydb.
<i>Nardus stricta</i>	L.
<i>Nassella charruana</i>	(Arech.) M. E. Barkworth
<i>Nassella neesiana</i>	(Trinius & Ruprecht) M. E. Barkworth
<i>Nassella pulchra</i>	(A. Hitchc.) M. E. Barkworth
<i>Nassella viridula</i>	(Trin.) M. E. Barkworth
<i>Neyraudia reynaudiana</i>	(Kunth) Keng ex Hitchcock
<i>Oryza longistaminata</i>	A. Chevalier & Roehrich
<i>Panicum kalaharensense</i>	Mez
<i>Panicum lanipes</i>	Mez
<i>Panicum maximum</i>	Jacq.
<i>Panicum phragmitoides</i>	Stapf
<i>Panicum repens</i>	L.
<i>Panicum virgatum</i>	L.
<i>Paspalum dilatatum</i>	Poir.
<i>Paspalum notatum</i>	Fluegge
<i>Paspalum scrobiculatum</i>	L.
<i>Paspalum vaginatum</i>	Sw.
<i>Pennisetum massaicum</i>	Stapf
<i>Pennisetum mezianum</i>	Leeke
<i>Pennisetum orientale</i>	Rich.
<i>Pennisetum polystachion</i>	(L.) Schult.
<i>Pennisetum purpureum</i>	Schum.
<i>Pennisetum sphacelatum</i>	(Nees) T. Dur. & Schinz
<i>Pennisetum stramineum</i>	Peter
<i>Pennisetum unisetum</i>	(Nees) Benth.
<i>Phleum alpinum</i>	L.
<i>Phleum phleoides</i>	(L.) Karst.
<i>Phragmites australis</i>	(Cav.) Trin. ex Steud.
<i>Phragmites vallatorius</i>	(Pluk.) J. F. Veldkamp
<i>Piptatherum microcarpum</i>	(Pilg.) Tsvelev

<i>Poa bonariensis</i>	(Lam.) Kunth
<i>Poa bulbosa</i>	L.
<i>Poa cita</i>	E. Edgar
<i>Poa hiemata</i>	Vickery
<i>Poa labillardieri</i>	Steud.
<i>Poa lanuginosa</i>	Poir.
<i>Poa nemoralis</i>	L.
<i>Poa pratensis</i>	L.
<i>Poa secunda</i>	J. & C. Presl
<i>Pogonarthria squarrosa</i>	(Roem. & Schult.) Pilger
<i>Pseudoraphis spinescens</i>	(R. Br.) Vickery
<i>Puccinellia gigantea</i>	(Grossh.) Grossheim
<i>Rytidosperma oreoboloides</i>	(F. Muell.) H. P. Linder
<i>Saccharum bengalense</i>	Retz.
<i>Saccharum spontaneum</i>	L.
<i>Schizachyrium sanguineum</i>	(Retz.) Alston
<i>Schizachyrium scoparium</i>	(Michx.) Nash
<i>Schizachyrium spicatum</i>	(Spreng.) Herter
<i>Schizachyrium tenerum</i>	Nees
<i>Schmidtia kalahariensis</i>	Stent
<i>Schmidtia pappophoroides</i>	Steud.
<i>Sehima ischaemoides</i>	Forsk.
<i>Sehima nervosum</i>	(Rottler) Stapf
<i>Setaria incrassata</i>	(Hochst.) Hack.
<i>Setaria sphacelata</i>	(Schumach.) Stapf & C. E. Hubb. ex Moss
<i>Sorghastrum nutans</i>	(L.) Nash
<i>Sorghum arundinaceum</i>	(Desv.) Stapf
<i>Sorghum purpureosericeum</i>	(A. Rich.) Schweinf. & Aschers.
<i>Spartina patens</i>	(Ait.) Muhl.
<i>Sporobolus airoides</i>	(Torr.) Torr.
<i>Sporobolus compositus</i>	(Poir.) Merrill
<i>Sporobolus contractus</i>	Hitchcock
<i>Sporobolus cubensis</i>	Hitchcock
<i>Sporobolus indicus</i>	(L.) R. Br.
<i>Sporobolus ioclados</i>	(Trin) Nees
<i>Stenotaphrum secundatum</i>	(Walt.) Kuntze
<i>Stipa arabica</i>	Trin. & Rupr.
<i>Stipa barbata</i>	Desf.
<i>Stipa capillata</i>	L.
<i>Stipa caucasica</i>	Schmalh.
<i>Stipa comata</i>	Trin. & Rupr.
<i>Stipa dasyphylla</i>	(Lindem.) Czern. ex Trautv.
<i>Stipa eremophila</i>	Reader
<i>Stipa hohenackeriana</i>	Trin. & Rupr.
<i>Stipa ichu</i>	(Ruiz & Pav) Kunth

<i>Stipa krylovii</i>	Roshev.
<i>Stipa lessingiana</i>	Trin. & Rupr.
<i>Stipa neaei</i>	Nees ex Steud.
<i>Stipa pulcherrima</i>	C. Koch
<i>Stipa richteriana</i>	Kar. & Kir.
<i>Stipa sareptana</i>	Beck.
<i>Stipa speciosa</i>	Trin. & Rupr.
<i>Stipa tenacissima</i>	L.
<i>Stipa thurberiana</i>	Piper
<i>Stipa tirsia</i>	Stev.
<i>Stipa trichophylla</i>	Benth.
<i>Stipa turkestanica</i>	Hack.
<i>Stipa zaleskii</i>	Wilensky
<i>Stipagrostis ciliata</i>	(Desf.) de Winter
<i>Stipagrostis uniplumis</i>	(Licht.) de Winter
<i>Themeda anathera</i>	(Nees) Hack.
<i>Themeda arundinacea</i>	(Roxb.) A. Camus
<i>Themeda tremula</i>	(Nees) Hack.
<i>Themeda triandra</i>	Forsk.
<i>Themeda villosa</i>	(Lam.) A. Camus
<i>Trachypogon spicatus</i>	(L.) Kuntze
<i>Tragus berteronianus</i>	Schult.
<i>Tragus koelerioides</i>	Aschers.
<i>Tragus racemosus</i>	(L.) All.
<i>Triodia basedowii</i>	E. Pritzel
<i>Triodia longiceps</i>	J. M. Black
<i>Triodia pungens</i>	R. Br.
<i>Triodia wiseana</i>	C. A. Gardner
<i>Tristachya leiostachya</i>	Nees
<i>Urochloa mosambicensis</i>	(Hack.) Dandy
<i>Vossia cuspidata</i>	(Roxb.) Griff.

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**Appendix S3** The global extent of grassy vegetation used as the basis for sampling in this study. Polygons, which make up the map represent different vegetation types. Polygons/vegetation types are coloured by the percentage of the total species per polygon/vegetation that were sampled in this study. The numbers of dominant species for vegetation types ranged from 1 to 19, with a mean of 4.

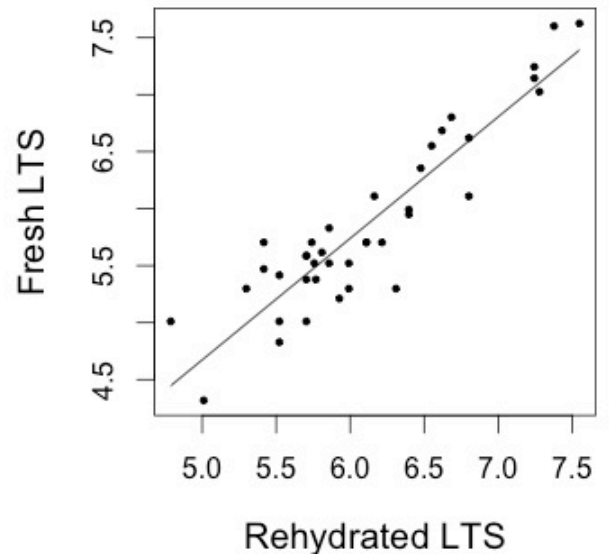
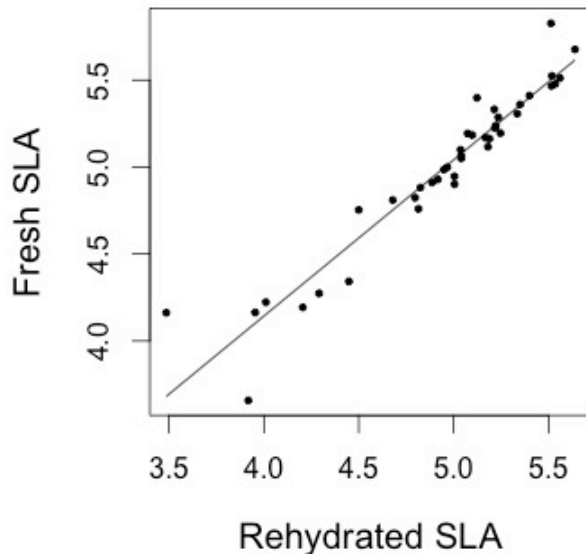


### **Measuring traits from herbarium specimens**

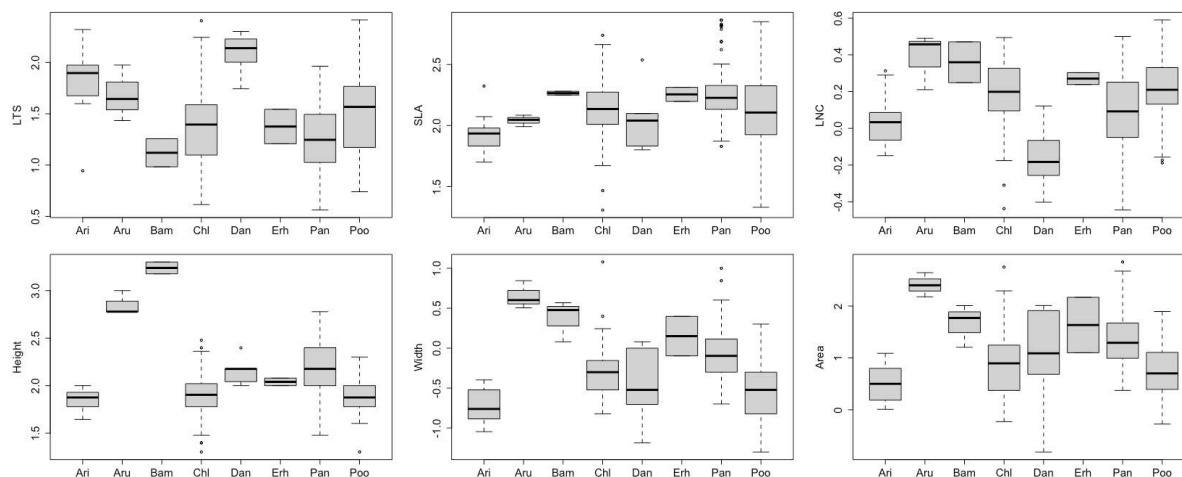
We measured the SLA and LTS on the fresh leaves of 39 grass species using standard protocols (Cornelissen, Lavorel et al. 2003). Leaf area was determined using image analysis software (WINDIAS, Delta-T Devices Ltd, Cambridge, UK) and dry leaf weight using a five-point balance. Leaves were then dried in herbarium presses for 10 days, weighed, rehydrated in de-ionised water for 24 hours before being scanned and then measured again. Using linear regression we showed strong correlations between the fresh and rehydrated trait measurements for SLA ( $r^2 = 0.90$ ,  $P < 0.001$ ) (Figure S1) and LTS ( $r^2 = 0.84$ ,  $P < 0.001$ ).



**Appendix S4** The relationship between trait values measured on fresh leaves and the same leaves that had been subjected to drying in herbarium presses then rehydrated for (a) specific leaf area (SLA) ( $r^2 = 0.90$ ,  $P < 0.001$ ) and (b) leaf tensile strength (LTS) ( $r^2 = 0.84$ ,  $P < 0.001$ ). All data were logarithmically transformed.



**Appendix S5** Comparison of trait values for Aristoideae (Ari), Arundoideae (Aru), Bambusoideae (Bam), Chloridoideae (Chl), Danthonioideae (Dan), Erhartoideae (Erh), Panicoideae (Pan) and Pooideae (Poo). Solid lines show the median and solid circles the mean for each clade. Outliers are unfilled circles. All traits were logarithmically transformed.



**Appendix 6a** The proportion of variance explained by each axis of a principal components analysis of the traits specific leaf area (SLA), leaf tensile strength (LTS), leaf nitrogen content (LNC), leaf carbon content (LCC), maximum culm height, maximum leaf area and maximum leaf width.

	PC1	PC2	PC3	PC4	PC5	PC6
Standard deviation	1.60	1.23	0.91	0.83	0.59	0.24
Proportion of Variance	0.43	0.25	0.14	0.11	0.58	0.01
Cumulative Proportion	0.43	0.68	0.82	0.93	0.99	1.00

**Appendix 6b** Loadings of each traits on each of the PC axis identified following principal components analysis of the traits specific leaf area (SLA), leaf tensile strength (LTS), leaf nitrogen content (LNC), maximum culm height, maximum leaf area and maximum leaf width.

	PC1	PC2	PC3	PC4	PC5	PC6
Height	-0.52	-0.24	0.04	-0.70	-0.81	-0.14
Leaf Area	-0.59	-0.14	0.01	-0.03	0.29	0.73
Leaf Width	-0.59	0.05	-0.12	0.02	0.47	-0.65
LNC	-0.08	0.60	-0.29	-0.73	-0.10	0.07
SLA	-0.11	0.46	0.88	0.04	-0.01	-0.01
LTS	0.13	-0.59	0.36	-0.68	0.19	-0.11

**Appendix S7** Pairwise relationships between all combinations of species mean traits.  $\lambda$  values are for the residuals in the PGLS model.  $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ ; *ns* not significant. Trait data were logarithmically transformed before tests

	Slope	S.E	$\lambda$	$r^2$	$P$
N~LTS	-0.256	0.027	0.40	0.26	***
N~SLA	-0.233	0.044	0.60	0.09	***
N~Height	-0.014	0.043	0.54	0	ns
N~Leaf Width	0.096	0.031	0.48	0.03	**
LTS~SLA	-0.640	0.085	0.40	0.18	***
SLA~Height	-0.036	0.056	0.32	0	ns
SLA~Leaf Width	0.107	0.042	0.30	0.02	*
SLA~Leaf Area	0.019	0.027	0.32	0	ns
Height~LTS	0.120	0.047	0.50	0.02	*
LTS~Leaf Area	0.052	0.040	0.50	0	ns
LTS~Leaf Width	-0.149	0.062	0.49	0.02	*
Leaf Width~Height	0.425	0.039	0.05	0.32	***
Leaf Area~Height	0.330	0.021	0.57	0.49	***

**Appendix S8** ANOVA comparing the slopes of the full model with the continent that each species was dominant in fitted as an interaction. The full model is defined as trait ~ soil nutrients\*continent + soil pH \*continent + soil % sand\*continent + PC1\*continent +PC2\*continent +PC3\*continent +PC4\*continent +PC5\*continent +PC6\*continent.  $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ ; *ns* not significant. Trait data were logarithmically transformed before tests

	<u>LC</u>			<u>LTS</u>			<u>SLA</u>			<u>Height</u>			<u>Leaf Area</u>			<u>Leaf Width</u>		
	DF	F value	P	DF	F value	P	DF	F value	P	DF	F value	P	DF	F value	P	DF	F value	P
Soil TEB	4	1.83	ns	4	0.52	ns	4	2.30	ns	4	0.47	ns	4	1.31	ns	4	1.48	ns
Soil pH	4	0.46	ns	4	0.96	ns	4	0.93	ns	4	1.22	ns	4	2.27	ns	4	0.89	ns
Soil % Sand	4	1.14	ns	4	0.42	ns	4	3.08	ns	4	0.59	ns	4	2.03	ns	4	0.33	ns
PC1	4	0.85	ns	4	0.84	ns	4	0.74	ns	4	0.43	ns	4	1.14	ns	4	1.89	ns
PC2	4	0.78	ns	4	0.69	ns	4	0.76	ns	4	0.63	ns	4	1.53	ns	4	1.46	ns
PC3	4	0.86	ns	4	1.10	ns	4	0.47	ns	4	0.21	ns	4	1.15	ns	4	1.83	ns
PC4	4	2.28	ns	4	2.26	ns	4	0.86	ns	4	2.37	ns	4	2.13	ns	4	1.26	ns
PC5	4	4.57	**	4	0.86	ns	4	2.20	ns	4	0.60	ns	4	1.22	ns	4	0.37	ns
PC6	4	1.74	ns	4	1.58	ns	4	0.80	ns	4	0.43	ns	4	2.04	ns	4	2.02	ns