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1 Adaptive radiation and the evolution of nectarivory in a large  
2 songbird clade

3

4 ABSTRACT

5 The accumulation of exceptional ecological diversity within a lineage is a key feature of adaptive  
6 radiation resulting from diversification associated with the subdivision of previously underutilized  
7 resources. The invasion of unoccupied niche space is predicted to be a key determinant of adaptive  
8 diversification, and this process may be particularly important if the diversity of competing lineages  
9 within the area in which the radiation unfolds is already high. Here, we test whether the evolution of  
10 nectarivory resulted in significantly higher rates of morphological evolution, more extensive  
11 morphological disparity, and a heightened build-up of sympatric species diversity in a large  
12 radiation of passerine birds (the honeyeaters, ca. 190 species) that have diversified extensively  
13 throughout continental and insular settings. We find that a large increase in rates of body size  
14 evolution and general expansion in morphological space followed an ancestral shift to nectarivory,  
15 enabling the build-up of large numbers of co-occurring species that vary greatly in size compared to  
16 related and co-distributed non-nectarivorous clades. These results strongly support the idea that  
17 evolutionary shifts into novel areas of niche space play a key role in promoting adaptive radiation in  
18 the presence of likely competing lineages.

19

20 Keywords: character displacement, macroevolution, macroecology, species richness, key  
21 innovations, morphological evolution

22

23

## 24 INTRODUCTION

25 Adaptive radiation describes the scenario in which lineage diversification is coupled with extensive  
26 ecological divergence into a wide variety of niches (Osborn 1902; Huxley 1942; Simpson 1953;  
27 Schluter 2000a). Although some iconic adaptive radiations have been extensively studied by  
28 evolutionary biologists (e.g. Darwin's finches, Hawaiian honeycreepers and Caribbean anoles), our  
29 general understanding of the factors that promote this phenomenon remain incomplete (Schluter  
30 2000a). Ecological opportunity in the form of new and/or underexploited resources is believed to be  
31 a common prerequisite for adaptive radiation, but this may arise in a multitude of ways. For  
32 example, ecological opportunity may emerge as a consequence of (i) the colonization of new  
33 geographic areas, (ii) the appearance of a new resource, (iii) the extinction of competitors/predators,  
34 or (iv) as a result of the evolution of key innovations (Simpson 1953; Schluter 2000a; Losos and  
35 Mahler 2010; Stroud and Losos 2016). Most well-studied adaptive radiations have resulted from the  
36 colonization of geographically isolated areas and are therefore likely to have unfolded in the  
37 absence of competition from closely related lineages (Losos 2010; Soulebeau et al. 2015; Stroud  
38 and Losos 2016). Much less is known about the factors facilitating adaptive radiation when the  
39 levels of species diversity among potentially competing lineages is already high. This scenario is  
40 particularly applicable to radiations occurring throughout continental settings, where the bulk of the  
41 world's species diversity is distributed. In these instances, one important factor is thought to be the  
42 evolution of new morphological and physiological traits that allow lineages to utilize novel  
43 resources, and radiate free from competition with related co-occurring species (Miller 1949;  
44 Simpson 1944; Hunter 1998; Rabosky 2017).

45           The evolution of traits that facilitate access to previously inaccessible resources has  
46 been hypothesized to underlie the evolutionary success of many large radiations, with proposed  
47 examples including the evolution of phytophagy in insects (Mitter *et al.* 1998), or the pharyngeal

48 jaw in labroid fishes (Liem 1973; Galis and Drucker 1996). Under this scenario, lineages that are  
49 able to invade unoccupied niche space, are predicted to undergo increased rates of trait evolution  
50 and exhibit greater ecological disparity compared to related clades, assuming that the available  
51 resources are amenable to further subdivision (Futuyma 1998; Losos and Mahler 2010; Rabosky  
52 2017). Moreover, such clades should also be characterized by a shift in ecological positioning  
53 relative to the background clade, as may be evidenced by the evolution of new traits (or trait  
54 combinations) that facilitate novel patterns of resource utilization (Rabosky 2017). Adaptive  
55 radiations that unfold in this way may also be expected to support higher numbers of species at  
56 smaller spatial scales, as greater ecological divergence facilitates a high degree of sympatry among  
57 the constituent taxa (Schluter 1996). The invasion of underexploited areas of niche space through  
58 the evolution of novel traits has also in some instances been proposed to result in increased lineage  
59 diversification (Mitter et al. 1988; Slowinski and Guyer 1993; Hodges and Arnold 1995; Bond and  
60 Opell 1998). However, this hypothesis remains contentious as the evolution of such traits may  
61 increase the overall diversification of the parent clade (thus raising its accumulated species  
62 richness), without necessarily increasing rates of lineage diversification among the innovative clade  
63 (Rabosky 2017). However, empirical assessments of these predictions, and documentation of the  
64 tempo and mode by which radiations of this nature unfold, are currently limited. We address these  
65 issues by assessing the effect of an ancestral shift in diet on rates of morphological evolution,  
66 lineage diversification and patterns of species co-occurrence within a large clade of passerine birds  
67 that has radiated extensively throughout continental and insular settings.

68           The infraorder Meliphagides is a passerine radiation of approximately 290 species  
69 distributed across Australasia and the Indo-Pacific (Gardner et al. 2010; Marki et al. 2017).  
70 Australasia is thought to represent the ancestral area of songbird (oscine passerines) diversification  
71 (Barker et al. 2004; Jønsson et al. 2011; Moyle et al. 2016), thus providing a contrasting geographic

72 setting to other studies of adaptive radiations that have predominantly assessed these trends in  
73 isolated and species depauperate island environments (Pratt 2005; Grant and Grant 2008; Losos  
74 2009). Ecological and phenotypic diversity is particularly pronounced in the honeyeater subclade  
75 (family Meliphagidae), which comprises ca. 65% (187 species) of the overall species richness of the  
76 infraorder. Honeyeater species possess a number of unique morphological and physiological  
77 adaptations for nectarivory, including structural modifications to the renal system for more efficient  
78 balancing of fluid intake and a brush-tipped protrusible tongue (Paton and Collins 1989; Cassoti  
79 and Richardson 1992; Goldstein and Bradshaw 1998a,b). These adaptations are hypothesized to  
80 have allowed honeyeaters to successfully exploit a novel food source (nectar) and radiate into areas  
81 of ecological niche space that were previously unoccupied in this geographic setting (Keast 1976;  
82 Miller et al. 2017). Together, these factors make the Meliphagides an ideal study system for  
83 investigating the dynamics of adaptive radiation at large geographic scales.

84           Here, we use empirical data to assess core, but largely untested predictions of adaptive  
85 radiation theory following the invasion of novel niche space. First, we test the prediction that  
86 following the evolution of nectarivory honeyeaters should occupy a unique and exceptionally  
87 diverse part of morphological space compared to other co-distributed and closely related passerine  
88 clades. Second, we evaluate whether the macroevolutionary dynamics of trait evolution in  
89 nectarivorous lineages are decoupled from those of non-nectarivorous lineages. Finally, having  
90 established such a link, we examine whether these processes have influenced lineage diversification  
91 dynamics, geographic variation in species richness and the functional diversity of Meliphagoid  
92 assemblages.

93

94 MATERIALS AND METHODS

95 *Phylogenetic, morphological and ecological data*

96 We used the recently published molecular phylogeny of the Meliphagides by Marki et al. (2017) in  
97 all analyses. This phylogeny is nearly complete at the species-level and includes 286 of 289 (99%)  
98 of the currently recognized species according to the IOC World Bird List version 6.2 (Gill and  
99 Donsker 2016). The phylogeny was calibrated using a combination of fossils and secondary  
100 calibration points, and was summarized as a maximum clade credibility (MCC) tree upon which all  
101 comparative analyses were performed, unless otherwise stated .

102 To quantify morphological variation among the Meliphagides, we collected data on  
103 seven ecologically relevant traits that represent major aspects of external avian anatomy, from  
104 museum study skins. We measured tarsus length, hind toe length (including claw), wing length,  
105 Kipp's distance and bill length, width and depth (Table S1). Male specimens were measured where  
106 possible, although in a few cases when these were unavailable in the respective collections, the  
107 measurements for these species were supplemented with those from females or unsexed specimens.  
108 We obtained measurements for a total of 1,245 individual specimens including all but 13 taxa  
109 represented in the phylogeny (the species for which we were not able to obtain morphological data  
110 were *Acanthiza katherina*, *Amytornis ballarae* and *A.dorotheae*, *Aphelocephala pectoralis*,  
111 *Bolemoreus hindwoodi*, *Chenorhamphus campbelli*, *Lichmera monticola*, *Manorina melanotis*,  
112 *Meliphaga cinereifrons* and *M. fordiana*, *Myzomela blasii*, *Ptiloprora mayri*, *Stipiturus mallee* and  
113 *ruficeps*), with an average of  $4.5 \pm 1.9$  SD specimens measured per species. In addition to the  
114 Meliphagides, we also collected morphological measurements for the majority of species within 13  
115 families that are co-distributed with the honeyeaters (Artamidae, Campephagidae, Cinclosomatidae,  
116 Climacteridae, Corvidae, Melanocharitidae, Monarchidae, Oriolidae, Pachycephalidae,  
117 Paradisaeidae, Petroicidae, Ptilonorhynchidae and Rhipiduridae) totaling 398 additional species  
118 (2,085 specimens measured, mean per species =  $5.2 \pm 1.6$  SD). Using ANOVA across the full

119 morphological data set, we found that between-species variance on average accounted for 98%  
120 (range 96 – 99%) of the variance across all seven traits. Consequently, all subsequent analyses were  
121 performed on the log-transformed mean trait values calculated for each individual species. The  
122 MCC tree and the morphological measurements from the individual specimens have been made  
123 available on the Dryad online repository (hyperlink to be provided upon acceptance).

124           We discretely classified individual meliphagoid species according to whether or not  
125 they include nectar in their diets using information from a large database of ecological traits  
126 (Wilman et al. 2014). For species not included in the Wilman et al. (2014) diet database but present  
127 in the phylogeny ( $n = 13$ ), we used the most frequent condition among members of the genus to  
128 represent their dietary category.

129

### 130 *Analyses of trait evolution*

131 To assess the evolutionary origins of nectarivory among the Meliphagides, we reconstructed  
132 ancestral diets using stochastic character mapping (Bollback 2006) implemented in the R package  
133 *phytools* (Revell 2012; R Core Team 2016). To do this, we first compared two models of variation  
134 in transition rates among states by computing the likelihoods of an equal-rates (ER) and an all-rates  
135 different (ARD) model to our data. Likelihood ratio tests indicated that the more complex ARD  
136 model did not represent a significantly better fit than an ER model ( $P = 0.31$ ) and therefore we  
137 consequently estimated 1,000 stochastic character maps using the ER model. To test the hypothesis  
138 that honeyeaters occupy distinct and extended parts of morphological space relative to co-  
139 distributed clades we used a number of different approaches. First, in order to compare the  
140 morphological diversity of honeyeaters ( $n = 180$  species) with that of the four remaining  
141 meliphagoid families ( $n = 93$  species, herein we refer to these clades as the 'background

142 meliphagoids'), we performed a phylogenetic principal component (pPC) analysis upon the  
143 covariance matrix of the seven log-transformed variables (Revell 2009). Second, we also assessed  
144 the morphological space occupied by honeyeaters to that of a broader subset of the  
145 Australasian/Indo-Pacific avifauna that encompassed the background meliphagoids, in addition to  
146 the members of 13 further passerine families present in the region ( $n = 491$  species, herein we refer  
147 to this assemblage of clades as the 'regional passerines'). For this analysis, we used the species  
148 scores generated from a separate principal component analysis of the log-transformed  
149 morphological measurements. Due to the lack of comprehensive molecular phylogeny for this wider  
150 species set, we were unable to correct for the influence of shared ancestry in this analysis.  
151 Combined, PC axes 1-4 explained 95% of the overall variance in both the phylogenetic and non-  
152 phylogenetic PCAs (Table S2-3), and we thus focused our subsequent analyses and interpretations  
153 on these variables. To test whether honeyeaters occupy unique parts of morphological space  
154 compared to the background meliphagoids and the regional passerine fauna, we estimated the four  
155 dimensional hypervolumes of honeyeaters relative to related clades, using the *hypervolume*  
156 methodology (Blonder et al. 2014). We thus performed two sets of comparisons using the first four  
157 PCA axes derived from the separate pPC and PC analyses described above (honeyeaters versus  
158 background meliphagoids, and honeyeaters versus regional passerines). The hypervolumes were  
159 estimated using a multidimensional kernel estimation procedure, and bandwidths that were  
160 determined using the Silverman bandwidth estimator (Blonder et al. 2015). Overlap in the  
161 hypervolumes between the clades in the two sets of analyses was calculated using the Sørensen  
162 index (see Blonder et al. 2015), whereby a value of 0 indicates no overlap between the  
163 hypervolumes, and a value of 1 indicates identical hypervolumes. Finally, to further assess whether  
164 honeyeaters occupy unique parts of morphological space and to define specific traits that  
165 differentiate the groups, we performed a linear discriminant analysis upon the seven original log-



166 transformed morphological measurements, treating the regional passerine clades as both a single  
167 class, and as multiple classes divided by family.

168           The invasion of novel niche space has been predicted to result in a decoupling of rates  
169 of eco-morphological evolution between the invading and non-invading clades (Rabosky 2017). To  
170 test this hypothesis, we compared the relative fit of different models of trait evolution using the R  
171 package *mvMORPH* (Clavel et al. 2015). Specifically, we compared a Brownian motion (BM)  
172 model with a single rate of trait evolution for all lineages (BM1) to a BM model with separate rates  
173 of trait evolution for nectarivorous and non-nectarivorous lineages (BMM). We fit these two models  
174 to each of the 1,000 stochastic character maps. Univariate analyses were run for each of the first  
175 four pPC axes (pPC1-4). Similarly, we also compared models of multivariate evolution (pPC1-4)  
176 across ten evenly sampled stochastic character maps. Model support was assessed using AICc  
177 scores and Akaike weights. To test for the possible influence of phylogenetic uncertainty, we  
178 repeated the above analyses across a posterior distribution of 1,000 Meliphagides trees obtained  
179 from the study of Marki et al. (2017) upon which we first estimated stochastic character maps using  
180 a single simulation per tree. In addition, we also assessed finer scale lineage variation in the tempo  
181 and mode of meliphagoid morphological evolution using a variable rates model as implemented in  
182 BayesTraits v2 (available from <http://www.evolution.rdg.ac.uk/>). This approach uses reversible-  
183 jump Markov chain Monte Carlo algorithms (rjMCMC) and two scaling mechanisms to identify  
184 rate changes along single branches and for whole clades across the phylogeny (Venditti et al. 2014).  
185 We used default priors for the phylogenetic mean ( $\alpha$ ) and Brownian variance ( $\sigma$ ) parameters and ran  
186 a single rjMCMC chain for each of the four pPC axes for 50 million generations, sampling every  
187 5000<sup>th</sup> generation. In addition, we ran a correlated multivariate analysis that assessed the  
188 evolutionary dynamics of all four axes simultaneously, using the same parameters. We assessed  
189 mixing and convergence of the chains, before the first 5 million generations were removed as a

190 burn-in. BayesTraits outputs a posterior distribution of trees in which the branches are scaled by the  
191 rate of evolutionary change that best explain the distribution of the trait values at the tips. Results  
192 were summarized by (i) calculating the mean rate of trait evolution along each branch, considering  
193 the posterior distribution of trees, and (ii) by calculating the posterior probability of branch or clade  
194 shifts over all samples for each node in the tree. To account for uncertainty in the precise location of  
195 rate shifts across posterior samples, we calculated the posterior probability of a rate shift as the sum  
196 of the probability of this having occurred on a focal node, or on either of the descendant nodes  
197 (following Cooney et al. 2017). In addition to BayesTraits, we also investigated another widely  
198 used framework for inferring variable rates of trait evolution across a phylogeny (BAMM v2.5.0;  
199 Rabosky 2014; Rabosky et al. 2014a). The BAMM method attempts to identify the location and  
200 number of distinct macroevolutionary rate regimes on the phylogeny. The number of distinct  
201 regimes are modelled following a Poisson distribution, with rjMCMC used to sample different  
202 regimes that best explain the distribution of trait values at the tips of the tree. We used the R  
203 package *BAMMtools* (Rabosky et al. 2014b) to estimate the prior settings for the phenotypic rates  
204 and for the hyperprior on the Poisson rate prior. The rjMCMC chains were run for 10 million  
205 generations each, sampling every 1000<sup>th</sup> generation. Convergence and mixing of the individual  
206 chains was assessed through visual inspection and by computing effective sample sizes (ESS), with  
207 the first 10% of samples subsequently discarded as a burn-in. We analyzed each of the four pPC  
208 axes calculated for the Meliphagides using the MCC tree as input.

209           To compare model performance between alternative evolutionary methods, we used  
210 the approach outlined in Cooney *et al.* (2017) that builds on the methodological developments of  
211 Pennell et al. (2015) and Chira and Thomas (2016), to calculate log-likelihoods describing the  
212 relative fit of different models of continuous trait evolution to each pPC axis. These analyses were  
213 performed using the *fitContinuous* function in the R package *geiger* (Harmon et al. 2008). We thus

214 calculated the likelihood of three single-process models (Brownian motion (BM), Ornstein-  
215 Uhlenbeck (OU) and early-burst (EB)) fitted to the original untransformed tree, and compared these  
216 to the likelihoods of BM models fit to the mean rate-transformed trees derived from BAMM  
217 (obtained using the function *getMeanBranchLengthTree* in *BAMMtools*) and BayesTraits. Model  
218 comparisons (using delta log-likelihoods) indicated that BayesTraits represented a significantly  
219 better description of the patterns of morphological evolution among the Meliphagides than either  
220 BAMM or any of the single-process models for all pPC axes analyzed (pPC1-4, Table S4).  
221 Consequently, we focus our interpretation and discussion on the BayesTraits results (although those  
222 generated by BAMM were largely congruent, Fig. S1).

223           To test whether the evolution of nectarivory by honeyeaters has led to an increase in  
224 the total volume of eco-morphological space occupied by the Meliphagides (Rabosky 2017), we  
225 assessed the accumulation of morphological disparity and the filling of morphospace through time.  
226 Using maximum likelihood in *phytools* (Revell 2012), we reconstructed ancestral states for each of  
227 the pPC axes using the mean rate-transformed trees from BayesTraits. We then divided the tree in  
228 time slices at 0.5 million year intervals, starting at the root, and for each time slice extracted  
229 ancestral state estimates for all lineages present at a given time. We compared both disparity  
230 accumulation of the individual PC axes, and for all axes combined, by summing the variances  
231 across all four axes. Finally, we compared the empirical accumulation of trait disparity through  
232 time, with that expected under a constant-rate BM model and a variable-rates (VR) model of trait  
233 evolution. Thus, for both null models we simulated 500 replicate datasets for each of the pPC axes  
234 and for pPC1-4 combined, to calculate disparity through time curves.

235

236 *Lineage diversification and spatial diversity patterns*

237 The invasion of novel or unoccupied niche space may in some instances lead to a decoupling in  
238 diversification dynamics between the invading and non-invading clades (e.g. Mitter et al. 1988, but  
239 see Givnish 2015; Rabosky 2017). To test for a decoupling in the dynamics of lineage  
240 diversification between nectarivorous and non-nectarivorous lineages we applied the hidden-state  
241 speciation and extinction framework (HiSSE; Beaulieu and O'Meara 2016). The HiSSE framework  
242 is an extension of the binary-state speciation and extinction model (BiSSE; Maddison et al. 2007)  
243 developed to circumvent issues of high type I error rates associated with this method (Rabosky and  
244 Goldberg 2015). Using HiSSE, we compared the fit of five different models of lineage  
245 diversification (see Table S5 for details of number of parameters and constraints for each model),  
246 accounting for incomplete taxon sampling (3/289 species missing). Given the difficulty in reliably  
247 estimating transition rates in these analyses (Beaulieu and O'Meara 2016), we set transition rates  
248 between diet categories to be equal across all models. Model support was assessed using AICc  
249 scores and Akaike weights, and the results were visualized using model-averaged marginal  
250 reconstructions of diet and net diversification rates.

251 To assess whether increased ecological dispersion among honeyeaters has led to a  
252 heightened accumulation of sympatric species diversity (Schluter 1996), we compared the  
253 geographic species richness patterns of the honeyeaters to that of the background meliphagoids. To  
254 do this, we obtained range maps from a global distributional database (Rahbek and Graves 2001;  
255 Rahbek et al. 2012), with species ranges recorded at a resolution of  $1^\circ \times 1^\circ$ . We then mapped the  
256 species richness of the honeyeaters and the background meliphagoids by overlaying the ranges,  
257 before summing the number of species present in each  $1^\circ$  grid cell. Subsequently, we assessed the  
258 range and standard deviation of the individual pPC axes throughout all grid cells among both  
259 groups. Using linear models, we regressed the grid cell values of the species richness of the  
260 honeyeaters against the background meliphagoids. Finally, we determined how the range and

261 standard deviation of the pPC axes varied among the honeyeaters and background meliphagoids as  
262 a function of the species richness of all grid cells. As the range can be sensitive to outlying values,  
263 we also mapped the 95% quantiles of the range in pPC1-4 to explore the robustness of our results.

264

## 265 RESULTS

### 266 *Diet reconstructions and morphological diversity*

267 The ancestral reconstruction of the presence of nectar in the diet of the Meliphagides is strikingly  
268 characterized by a shift from a non-nectarivorous diet to one that can incorporate nectar in the  
269 common ancestor of honeyeaters (Fig. 1a). Nectarivory has also evolved independently among the  
270 pardalotes (family Pardalotidae) and among a handful of species of Australasian warblers (family  
271 Acanthizidae) that are members of the background meliphagoids. Among honeyeaters, loss of  
272 nectarivory has occurred independently on a number of more terminal branches, such as in the  
273 largely frugivorous genera *Melipotes* and *Macgregoria*, as well as in more insectivorous genera  
274 such as *Epthianura* and *Timeliopsis*. A pPCA of the seven log-transformed morphological traits  
275 (Table S1) comparing honeyeaters against background meliphagoids, showed that the first axis  
276 (pPC1) strongly reflected overall size, explaining 65.3% of the total variance in the morphological  
277 measurements (Table S2). The next three axes (pPC2-4) were related to variation in Kipp's distance  
278 (pPC2), bill depth and width (pPC3), and bill length (pPC4) respectively, together explaining 29.8%  
279 of the variance. Visual comparisons of species scores on pPC axes 1-4 highlight the great  
280 morphological disparity and distinctiveness of the honeyeaters. First, the extent and variance of  
281 body sizes (pPC1) exhibited by honeyeaters is much greater than that of the background  
282 meliphagoids (Fig. 2a). Although differences in shape variance are less extensive, honeyeaters  
283 generally have higher values of pPC2, in part, reflecting their greater Kipp's distance values (Fig.

284 2b). In addition, honeyeaters largely cluster separately from the background meliphagoid groups on  
285 pPC4, which is primarily related to differences in relative bill length (Fig. 2b). Results of a second,  
286 phylogenetically uncorrected PCA analysis comparing honeyeaters against the regional passerine  
287 fauna are largely congruent with these findings (Table S3), with honeyeaters exhibiting a high  
288 diversity of body sizes (Fig. 2c) and unique morphologies related to bill shape and length (PC3-  
289 PC4) (Fig. 2d).

290           The four dimensional hypervolume comparisons strongly support the above findings,  
291 with the Sørensen index indicating limited morphological overlap between the honeyeaters and  
292 background meliphagoids (Fig. S2, Sørensen's index = 0.07), and between the honeyeaters and  
293 regional passerines (Fig. S3, Sørensen's index = 0.22). Moreover, honeyeaters were found to occupy  
294 a high fraction of unique morphological space relative to both background meliphagoids and to the  
295 broader regional passerine fauna (0.93 and 0.47 of the overall morphospace respectively). A linear  
296 discriminant analysis of the seven original log-transformed measurements are in congruence with  
297 these results, illustrating that honeyeaters occupy distinct parts of morphological space relative to  
298 other regional passerines, with more than two-thirds of honeyeater species correctly classified as  
299 members of the family. Group means on the single discriminant axis were  $-1.51 \pm 0.93$  for  
300 honeyeaters and  $0.56 \pm 1.03$  for the remaining regional passerine species (Fig. S4). Normalized  
301 canonical coefficients separating the two groups indicate that the distinctiveness is largely driven by  
302 the comparatively long and narrow bills of the honeyeaters relative to other regional passerines  
303 (Table S6). Similar results were obtained from a comparison of honeyeaters against the regional  
304 passerine clades when these were divided by family, with 89% of honeyeaters correctly classified  
305 (Table S7).

306

307 *Morphological evolution*

308 Comparisons of different models of trait evolution using *mvMORPH* provided strong support for a  
309 decoupling of trait diversification dynamics among nectarivorous and non-nectarivorous lineages.  
310 Models with separate rates of trait evolution (BMM) for nectarivorous and non-nectarivorous  
311 lineages represented the best-fitting model for pPC1, pPC2, pPC4 and the multivariate analysis of  
312 pPC1-4, whereas a single-rate BM (BM1) model was the best fit for pPC3 (Table 1). For pPC1,  
313 pPC4 and pPC1-4 nectarivorous lineages were found to have a higher rate of evolution than non-  
314 nectarivorous lineages. For pPC2, nectarivorous lineages were found to have a lower rate of  
315 evolution than non-nectarivorous lineages. These results were largely corroborated when  
316 phylogenetic uncertainty was accounted for (Table S8), although support for a single-rate BM  
317 (BM1) model was only marginally better than a BMM model for the analysis of pPC3.

318           The BayesTraits analyses of the multivariate data (pPC1-4 combined) recovered a  
319 number of rate shifts distributed across the Meliphagides (Fig. S5), including a substantial single-  
320 branch shift on the stem branch of the honeyeaters (PP = 0.73), as well as several rate shifts on  
321 more terminal branches and nodes among both honeyeaters and background meliphagoids.  
322 Deconstructing these trends among the individual pPC axes provided strong support for a clade-  
323 wide shift to higher rates of trait evolution near the base of the honeyeater clade on pPC1 (PP =  
324 0.90; Fig. 1b, S5) and for three species of *Gerygone* among the background meliphagoids (PP =  
325 0.83). No rates shifts in the univariate analysis of pPC2-4 were strongly supported (all PP < 0.7).

326           Analyzing the accumulation of morphological disparity through time, we find that  
327 disparity has steadily accumulated across the Meliphagides when considering all pPC axes  
328 congruently (Fig. S6-8). Focusing on the individual pPC axes, we show that that this pattern is  
329 largely driven by an expansion in size disparity (pPC1) among the honeyeaters (Fig. 1c). Whereas

330 body size disparity has continued to increase throughout the evolutionary history of the honeyeaters,  
331 this has not been the case for the background meliphagoid lineages, which have accumulated more  
332 limited disparity overall (Fig. 1c). Disparity accumulation on pPC2 exhibits a contrasting trend,  
333 however, with an early increase in disparity among the background meliphagoids, followed by two  
334 periods of relative stasis towards the present. Although the background meliphagoids have  
335 accumulated higher total disparity on pPC2 than the honeyeaters, both groups have continued to  
336 accumulate disparity through time on this axis. Disparity accumulation on pPC3 exhibit similar  
337 trends to that of pPC1, being characterized by continual accumulation of disparity towards the  
338 present (Fig. S6-8). For pPC4, the disparity accumulation of the overall Meliphagides is  
339 characterized by an early expansion in disparity, followed by relative stasis, reflecting the  
340 divergence in bill morphology between the honeyeaters and the background meliphagoids.  
341 Following the occupation of unique areas of morphospace, disparity accumulation among the  
342 honeyeaters and background meliphagoids is comparatively less extensive and is dominated by a  
343 largely continuous and constant accumulation of disparity through time. Comparing the above  
344 trends to null expectations based on constant-rate (BM) and variable-rates (VR) models, suggest  
345 that disparity accumulation among meliphagoid lineages is largely consistent with a process of  
346 continuous niche expansion, with the possible exception of overall meliphagoid and background  
347 meliphagoid disparity accumulation on pPC2, and overall meliphagoid disparity accumulation on  
348 pPC4 which for both axes show signatures of slowdowns in disparity and thus niche expansion  
349 towards the present.

350

351 *Lineage diversification and spatial diversity patterns*



352 An analysis of lineage diversification dynamics using HiSSE suggested that a model with speciation  
353 rate variation associated with a hidden trait was the most strongly supported (AIC weight = 0.67,  
354 Table S5). An alternative model where in addition, extinction rates were also free to vary between  
355 the two hidden states also received substantial support (AIC weight = 0.24). Models where  
356 speciation rate variation was associated with diet, received little support (AIC weight < 0.03).  
357 Mapping model-averaged marginal reconstructions of diet and speciation rates onto the  
358 Meliphagides tree suggests that rates of speciation are generally high, with the exception of certain  
359 lineages that have lower rates, including the bristlebirds (Dasyornithidae), goldenface and fernwren  
360 (*Pachycare flavogriseum* and *Oreoscopus gutturalis*), and two species of Sulawesi honeyeaters  
361 (*Myza*) (Fig. S9).

362 Analyzing spatial diversity patterns, we found that honeyeaters exhibit geographic  
363 gradients of species richness that are highly correlated with the overall pattern shown by the  
364 background meliphagoid clades (Fig. 3a and 4a,  $R^2 = 0.65$  where richness of either group  $\geq 1$ ).  
365 Furthermore, both groups almost completely overlap in the range of their overall distribution, with  
366 the highest levels of grid cell richness being found in eastern Australia and New Guinea (Fig. 3a).  
367 However, the absolute richness of the honeyeaters (max richness = 42, mean richness =  $10.5 \pm 7.5$   
368 SD) is substantially higher than that of background meliphagoids (max richness 25, mean richness =  
369  $6.9 \pm 6.2$  SD) across the majority of grid cells in which the groups co-occur. To assess how species  
370 richness patterns compare with those of morphological diversity, we mapped the range and standard  
371 deviation of the individual pPC axes across grid cells (Fig. 3b-c; Fig. S10). First, we find that  
372 honeyeaters have a higher diversity of body sizes (pPC1) across grid cells compared to the  
373 background meliphagoids (Fig. 3b-c), with both continental areas (e.g. eastern Australia and New  
374 Guinea) and islands (e.g. New Caledonia and Manus) standing out as areas harboring exceptional  
375 body size diversity, results that are robust to the exclusion of outliers (Fig. S11). Thus, for a given

376 level of grid cell richness, both the range and standard deviation of body size is greater among the  
377 honeyeaters in contrast to the background meliphagoid groups (Fig 4b-c, Fig. S12). Conversely, for  
378 pPC2 the background meliphagoid groups show a higher range and standard deviation within grid  
379 cells. For pPC3-4, we find that within grid cells, the honeyeaters and background meliphagoid  
380 groups overlap extensively in the range and standard deviation of the values of their co-occurring  
381 species (Fig. S10). Thus, unlike our findings for pPC1, the geographic patterns of range and  
382 standard deviation among pPC2-4 do not reflect the underlying gradients in species richness.

383

## 384 DISCUSSION

385 The invasion of novel ecological niche space has been hypothesized to underlie the adaptive  
386 diversification of a wide range of organismal groups, but the role of this process in generating  
387 species and phenotypic diversity across large geographic scales remains poorly known. In this  
388 study, we tested key predictions of this hypothesis by analyzing the effects of an extensive shift in  
389 diet and resource use among a large continental and insular radiation of passerine birds – the  
390 honeyeaters. By explicitly analyzing these trends in a phylogenetic context that includes the  
391 honeyeaters and their closest relatives, we find strong evidence that the evolution of nectarivory  
392 represented the exploitation of underutilized ecological space that has coincided with substantial  
393 increases in the rate of morphological evolution, leading to the accumulation of extensive  
394 morphological disparity. Analyses of morphological evolution provide evidence for a clade-wide  
395 shift to substantially higher rates of body size evolution within the honeyeaters (Fig. 1B; Fig. S1;  
396 Table 1). The increase in rates of body size evolution followed a major change in diet that evolved  
397 to encompass nectar (Fig. 1A), allowing honeyeaters to enter novel regions of niche space in  
398 comparison to the regional passerine fauna with which they co-occur. However, this significant

399 dietary shift did not lead to a decoupling (i.e. acceleration or deceleration) of the dynamics of  
400 speciation among the honeyeaters and the background meliphagoids (Fig. S9, Table S5).  
401 Conversely, analyses of spatial diversity patterns suggest that despite having converged on  
402 congruent geographic diversity patterns, honeyeaters exhibit consistently higher levels of body size  
403 diversity and species richness than their close relatives within 1° grid cells (Fig. 3-4). These  
404 findings suggest that a shift towards nectarivory positively influenced the capacity of the  
405 honeyeaters to accumulate high sympatric species diversity. Extensive diversification along the  
406 body size axis could enable a greater number of honeyeater species to co-exist, reflecting their entry  
407 into an unoccupied adaptive zone (nectarivory) that allowed honeyeaters to fill vacant ecological  
408 and morphological space. Together, our findings highlight the important role that evolutionary  
409 innovation and the invasion of novel ecological niche space play in generating extensive ecological  
410 diversity and the build-up of sympatric species diversity throughout large geographic areas.

411           Character displacement resulting from interspecific competition for resources is  
412 believed to be the main driver of ecological and phenotypic disparification in adaptive radiation  
413 (Simpson 1953; Schluter 2000a,b; Losos and Mahler 2010). For honeyeaters, size-related  
414 aggression and displacement within flowering trees is a well-known phenomenon and assumed  
415 driver of body size evolution (Paton and Ford 1983; Diamond et al. 1989). This hypothesis provides  
416 a possible explanation for the tight congruence between the shift towards a nectarivorous diet and  
417 the increase in rates of body size evolution and disparity accumulation in the group. Honeyeaters  
418 are notorious for their aggressiveness, and even Alfred Russel Wallace noted how friarbirds would  
419 ferociously defend flowering trees against potential competitors (Wallace 1869). Although mimicry  
420 may be one tactic to avoid attack from larger species (Diamond 1982; Prum 2014), positive  
421 selection for smaller body size may represent another viable scenario, as small birds may be able to  
422 utilize resources that are inaccessible or not easily monopolized by larger birds (e.g., on small

423 terminal twigs in outer parts of a tree), thus avoiding aggressive attacks (Diamond et al. 1989).  
424 Interestingly, our findings of rapid and extensive body size evolution among honeyeaters are in  
425 stark contrast to the two other major nectarivorous clades of birds – the hummingbirds and sunbirds  
426 – which exhibit comparatively limited body size diversity, but greater overall phenotypic  
427 specialization for interaction with their flower resources (Stiles 1981; Fleming and Muchhala 2008;  
428 Zanata et al. 2017). Fleming and Muchhala (2008) attributed the among-clade differences in  
429 nectarivory specialization and body size diversity to variation in floral resource predictability  
430 among major regions, ranging from highest in the Neotropics to comparatively low in Australia. In  
431 concordance with this hypothesis, we suggest that strong competition for a valued resource, which  
432 can be highly unpredictable in its spatial and temporal occurrence, has been the prominent driver of  
433 body size evolution among the Australasian honeyeaters. In addition to increased rates of body size  
434 evolution, the transition to a nectarivorous diet appears to have had a profound influence on bill  
435 evolution among honeyeaters. Our results thus suggest that honeyeaters have unique bill  
436 morphology (i.e. longer and narrower) compared to the background meliphagoids and other  
437 regional passerines (Fig. S4; Table S6-7), whereas nectarivorous meliphagoid lineages are also  
438 found to have a higher rate of bill (pPC4) evolution than non-nectarivorous lineages (Table 1).  
439 Taken together, our results suggest that the evolution of nectarivory among honeyeaters have had  
440 important consequences for both rates of morphological evolution (i.e. body size) and  
441 morphological adaptations (i.e. bill size and shape) in this clade.

442           The extensive and continuous accumulation of morphological disparity among  
443 honeyeaters relative to the background meliphagoids, could be caused in part by recent  
444 morphological evolution into further novel and unoccupied areas of niche-space (Simpson 1944;  
445 Slater 2015; Cooney et al. 2017). Examples of this include the genera *Macgregoria* and *Melipotes*  
446 that have transitioned to a largely fruit-based diet that is also reflected in their generally shorter and

447 straighter bills relative to most other honeyeaters. Alternatively, this pattern could reflect the  
448 outcome of strong ecological character displacement, whereby interspecific competition among  
449 recently separated taxa selects for rapid phenotypic divergence (Brown and Wilson 1956; Schluter  
450 2000b). Many island species such as the two sympatric New Zealand honeyeater taxa  
451 *Prothemadera* and *Anthornis* may represent an extreme example of this process, as they display  
452 high levels of recent body size divergence, which is also consistent with the expectation of greater  
453 character displacement among species in depauperate environments (Schluter 2000b). Thus, both  
454 character displacement and diversification into further available and unoccupied niche space are  
455 probable explanations that likely contributed to the continual accumulation of disparity in the case  
456 of honeyeaters.

457           Although transitions into new adaptive zones (and adaptive radiation more generally)  
458 need not always result in increased rates of lineage diversification, increases in ecological diversity  
459 of the adaptively radiating clade may be predicted to facilitate the build-up of extensive sympatric  
460 species diversity (Givnish 1997; Losos and Mahler 2010; Stroud and Losos 2016; Givnish 2015;  
461 Rabosky 2017). Consistent with these predictions, we find that whereas there is no evidence of a  
462 decoupling of diversification dynamics among nectarivorous and non-nectarivorous meliphagoid  
463 lineages (Fig. S9; Table S5), the evolution of nectarivory appears to have influenced the build-up of  
464 extensive sympatric species richness among the predominantly nectarivorous honeyeaters. Thus,  
465 although honeyeaters and the other families within the Meliphagides share very similar  
466 distributional extents and geographic diversity gradients (Fig. 3 and 4), honeyeaters exhibit much  
467 higher levels of species richness within the same grid cells compared to that of the background  
468 meliphagoids. Although honeyeaters might be expected to accumulate higher grid cell richness than  
469 the background meliphagoids due to their higher overall species diversity, a null explanation such  
470 as this is unlikely to be sufficient in accounting for the strong correlations between grid cell

471 richness, body-size disparity and the trends of trait evolution. The evolution of nectarivory among  
472 the honeyeaters may thus represent an intriguing example of how evolutionary innovations may  
473 positively influence the build-up of species diversity without necessarily having direct effects on  
474 rates of lineage diversification (Rabosky 2017).

475           A number of non-mutually exclusive mechanisms may underlie the increased  
476 sympatric species diversity of honeyeaters, including elevated ecological diversity (Keast 1976;  
477 Miller et al. 2017), and increased dispersal capabilities. The association between sympatric species  
478 richness and body size diversity recovered here suggest either that diversity drives ecological  
479 divergence by character displacement, or alternatively, that expansion into unoccupied niche space  
480 allows more species of honeyeaters to coexist through relaxed ecological filtering. Whereas  
481 substantial expansion in morphological space of other regional clades may have been constrained  
482 by the presence of ecologically similar lineages, honeyeaters appear to have been able to expand  
483 more freely due to the general absence of competing nectarivores. Although Australasia and the  
484 Indo-Pacific is inhabited by some other nectarivorous birds, including non-passerine parrots such as  
485 the lorries and lorikeets (family Psittacidae: tribe Loriinae), this group is thought to have radiated  
486 considerably later than the honeyeaters, with most of the diversification having taken place in the  
487 last 5 million years (Schweizer et al. 2015). In comparison with honeyeaters, this group is  
488 characterized by comparatively low levels of sympatric species diversity (Schweizer et al. 2015),  
489 which could suggest that the ecological diversification of lorries and lorikeets has itself been  
490 constrained by the more ecologically diverse honeyeaters. Lorries and lorikeets appear to be less  
491 ecologically diverse than honeyeaters, exhibiting a comparatively reduced diversity of bill shapes  
492 and adaptation to a narrower range of habitats, dietary resources and foraging modes. However, in  
493 the absence of detailed ecological and morphological data for the lorries and lorikeets, these  
494 hypotheses necessitate formal testing. Finally, a number of nectarivorous bats also inhabit the

495 Australasian/Indo-Pacific region (family Pteropodidae), but as these are primarily nocturnal, direct  
496 competition with the diurnal honeyeaters is unlikely to have been pervasive.

497           Under a model of allopatric speciation, for character displacement to occur,  
498 genetic/reproductive differentiation must first accumulate in geographic isolation before subsequent  
499 range shifts into sympatry (Price 2008). The rate at which this process occurs is at least partly  
500 contingent on the dispersal propensity of the organisms in question, as this positively influences the  
501 rate at which lineages achieve range overlap (Pigot and Tobias 2015). A lack of positive selective  
502 pressures on factors that directly facilitate dispersal may thus help to explain why some adaptive  
503 radiations are notably species-poor (Losos and Mahler 2010; Givnish 2015). Among honeyeaters,  
504 good dispersal abilities are a well-established characteristic of many species and this is likely to  
505 have enabled frequent colonization and exchange between geographic regions (Keast 1968; Marki  
506 et al. 2017). The irregular, unpredictable and often highly disjunct occurrence of many nectar  
507 sources may have exposed honeyeaters to significant positive selection for increased dispersal  
508 capabilities as evidenced by the major seasonal and nomadic movements of many species (Keast  
509 1968; Pyke 1980; Wooller 1981). Our findings support this, with honeyeaters having on average  
510 longer and more projected wing tips compared to background meliphagoids, suggesting high  
511 dispersal capacity (Fig. S13; Claramunt et al. 2012). Thus, increased dispersal abilities among the  
512 many nectar-dependent honeyeaters may have been an additional factor promoting the build-up of  
513 species diversity by increasing the rates at which new populations are founded, and their subsequent  
514 transitions back into sympatry following differentiation (Pigot and Tobias 2015).

515           The utilization of previously inaccessible resources has been hypothesized to underlie  
516 the adaptive radiation of a wide range of organismal groups. Here, we have shown that an ancestral  
517 shift to a nectarivorous diet is correlated with rapid body size evolution and the accumulation of  
518 extensive size disparity within the speciose radiation of Australasian honeyeaters. Importantly, our

519 findings suggest that the rapid invasion of novel and previously unoccupied ecological space can  
520 positively affect the build-up of species and functional diversity across different spatial scales, even  
521 in the presence of related and likely competing lineages. Overall, these results highlight the  
522 important role of ecological opportunity in facilitating the generation of morphological and species  
523 diversity across large geographic areas.

524

525

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704 **Tables**

705 Table 1. Comparisons of evolutionary models testing for a decoupling of rates of trait evolution  
 706 between nectarivorous and non-nectarivorous lineages. The best-fitting models are highlighted in  
 707 bold. Shown are the mean and standard deviations of delta AICc values, AICc weights, and  
 708 Brownian variance ( $\sigma^2$ ) as estimated across 1,000 (univariate analyses of pPC1-4) and 10  
 709 (multivariate analysis of pPC1-4) stochastic character maps of the evolutionary history of diets  
 710 among the Meliphagides.

	pPC1	pPC2	pPC3	pPC4	pPC1-4
<b>BM1</b>					
Delta AICc	7.8±1.8	8.5±1.1	<b>0.00±0.00</b>	9.1±1.9	17.1±3.8
AICc weight	0.03±0.02	0.02±0.01	<b>0.71±0.02</b>	0.02±0.02	0.00±0.00
$\sigma^2$	0.022±0.000	0.007±0.000	<b>0.002±0.000</b>	0.001±0.000	0.032±0.000
<b>BMM</b>					
Delta AICc	<b>0.0±0.0</b>	<b>0.0±0.0</b>	1.8±0.2	<b>0.0±0.0</b>	<b>0.0±0.0</b>
AICc weight	<b>0.97±0.03</b>	<b>0.98±0.01</b>	0.29±0.02	<b>0.98±0.02</b>	<b>1.00±0.00</b>
$\sigma^2$ (nectarivorous)	<b>0.026±0.000</b>	<b>0.005±0.000</b>	0.002±0.000	<b>0.002±0.000</b>	<b>0.035±0.000</b>
$\sigma^2$ (non-nectarivorous)	<b>0.014±0.001</b>	<b>0.009±0.000</b>	0.002±0.000	<b>0.001±0.000</b>	<b>0.026±0.001</b>

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720 **Supplementary tables**

721 Table S1. Description of morphological traits measured using a calliper and taken to the nearest 0.1  
 722 mm (tarsus, hind toe and bill measurements), or using a wing ruler taken to the nearest 1 mm (wing  
 723 length and Kipp’s distance).

Trait	Description
Tarsus length	Length of the tarsometatarsus as measured from the tibiotarsus joint to the base of the toes, which is represented by the last undivided scute.
Hind toe	Length of the hallux and claw as measured on dorsal side.
Bill length	Total culmen length as measured from the tip of bill to the base of the skull
Bill depth	Vertical height of the bill as measured at the proximal edge of the nostrils
Bill width	Horizontal width of bill as measured at the proximal edge of the nostrils
Wing length	Length of the wing as measured from the carpal joint to the longest primary measured on a flattened wing.
Kipp’s distance	The difference in wing length as measured above, and the length from the carpal joint to the first secondary feather measured on a flattened wing

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732 Table S2. Correlation coefficients and proportion of variance explained by each of the phylogenetic  
 733 principal component (pPC) axes for the analysis of the Meliphagides dataset ( $n = 273$  species).

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<b>Trait</b>	<b>pPC1</b>	<b>pPC2</b>	<b>pPC3</b>	<b>pPC4</b>	<b>pPC5</b>	<b>pPC6</b>	<b>pPC7</b>
Tarsus length	-0.869	0.212	0.237	-0.246	-0.042	-0.279	0.066
Hind toe	-0.862	0.294	0.316	-0.110	0.055	0.229	0.051
Bill length	-0.794	0.349	0.114	0.477	0.030	-0.071	0.008
Bill depth	-0.832	0.222	-0.452	-0.076	0.219	-0.006	0.027
Bill width	-0.812	0.277	-0.330	-0.009	-0.386	0.072	0.028
Wing length	-0.955	-0.056	0.058	-0.077	-0.005	-0.008	-0.276
Kipp's distance	-0.678	-0.733	0.003	0.040	-0.004	0.009	0.036
Proportion of variance	0.653	0.196	0.063	0.039	0.022	0.018	0.009
Cumulative proportion of variance	0.653	0.849	0.912	0.951	0.973	0.991	1.000

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738 Table S3. Trait loadings and proportion of variance explained by each of the principal component  
 739 axes for the analysis of the full passerine dataset ( $n = 671$  species).

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<b>Trait</b>	<b>PC1</b>	<b>PC2</b>	<b>PC3</b>	<b>PC4</b>	<b>PC5</b>	<b>PC6</b>	<b>PC7</b>
Tarsus length	0.240	0.373	-0.338	0.511	0.214	-0.604	-0.130
Hind toe	0.306	0.325	-0.311	0.239	-0.380	0.633	-0.320
Bill length	0.345	0.275	-0.306	-0.816	-0.009	-0.197	-0.082
Bill depth	0.433	0.212	0.479	0.004	0.671	0.289	-0.058
Bill width	0.346	0.081	0.642	0.030	-0.582	-0.322	-0.134
Wing length	0.373	0.043	-0.107	0.104	-0.137	0.080	0.901
Kipp's distance	0.531	-0.791	-0.205	0.066	0.054	-0.045	-0.202
Proportion of variance	0.822	0.117	0.029	0.018	0.007	0.004	0.003
Cumulative proportion of variance	0.822	0.939	0.968	0.986	0.993	0.997	1.000

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745 Table S4. Comparison of model fit for different models of morphological evolution. Delta log-  
 746 likelihoods values are shown for alternative models of morphological evolution. Values for the  
 747 BayesTraits and BAMM were generated by estimating the likelihoods of a BM model fit to the  
 748 mean rate-transformed trees.

	pPC1	pPC2	pPC3	pPC4
BayesTraits	0.0	0.0	0.0	0.0
BAMM	32.9	33.1	28.3	32.9
BM	65.8	47.4	85.1	66.8
OU	65.8	47.4	61.4	66.8
EB	65.8	47.4	85.1	66.8

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750 Table S5. Comparisons of lineage diversification models using HiSSE.

<b>Model</b>	<b>Parameter constraints</b>	<b>No. of parameters</b>	<b>Delta AICc</b>	<b>AICc weight</b>
BiSSE null	Speciation, extinction and transition rates equal	3	5.3	0.05
HiSSE 1	Transition rates equal	5	2.1	0.24
<b>HiSSE 2</b>	<b>Extinction and transition rates equal</b>	<b>4</b>	<b>0</b>	<b>0.67</b>
BiSSE 1	Transition rates equal	5	8.0	0.01
BiSSE 2	Extinction and transition rates equal	4	6.0	0.03

751

752 Table S6. Normalized canonical coefficients separating honeyeaters and other regional passerines  
 753 on the basis of the seven original log-transformed variables.

<b>Trait</b>	<b>Tarsus</b>	<b>Hind toe</b>	<b>Bill length</b>	<b>Bill depth</b>	<b>Bill width</b>	<b>Wing length</b>	<b>Kipp's distance</b>
Coefficient	1.141	0.564	-5.525	-0.519	5.842	0.760	-0.932

754

755 Table S7. Classification of passerine species based on the linear discriminant analysis.

Family	Acanthizidae	Artamidae	Campephagidae	Cinclosomatidae	Climacteridae	Corvidae	Dasyornithidae	Maluridae	Melanocharitidae	Meliphagidae	Monarchidae	Oriolidae	Pachycephalidae	Paradisaeidae	Pardalotidae	Petroicidae	Ptilonorhynchidae	Rhipiduridae	Total number of species	Classification accuracy
Acanthizidae	51	0	0	0	0	0	1	6	0	1	1	0	2	0	0	0	0	0	62	0.82
Artamidae	0	17	2	0	0	4	0	0	0	0	0	0	0	0	0	0	0	0	23	0.74
Campephagidae	0	2	39	0	0	0	0	0	0	3	0	0	1	0	0	3	0	0	48	0.81
Cinclosomatidae	1	0	0	4	0	0	2	0	0	2	0	0	1	0	0	1	0	0	11	0.36
Climacteridae	0	0	0	0	7	0	0	0	0	0	0	0	0	0	0	0	0	0	7	1.00
Corvidae	0	0	0	0	0	18	0	0	0	0	0	0	0	0	0	0	0	0	18	1.00
Dasyornithidae	0	0	0	0	0	0	3	0	0	0	0	0	0	0	0	0	0	0	3	1.00
Maluridae	7	0	0	0	0	0	3	14	0	0	0	0	0	0	0	0	0	0	24	0.58
Melanocharitidae	2	0	0	0	0	0	0	0	0	4	2	0	0	0	0	1	0	0	9	0.00
Meliphagidae	2	0	1	0	0	1	0	0	0	161	1	0	2	4	0	6	0	2	180	0.89
Monarchidae	0	0	0	0	0	0	0	0	0	7	50	0	4	0	0	2	0	3	66	0.76
Oriolidae	0	1	6	0	0	0	0	0	0	0	2	6	1	0	0	0	2	0	18	0.33
Pachycephalidae	0	0	0	0	0	0	0	1	0	2	2	0	41	0	0	0	1	1	48	0.85
Paradisaeidae	0	0	0	0	0	0	0	0	0	6	0	1	0	33	0	1	0	0	41	0.80
Pardalotidae	0	0	0	0	0	0	0	0	0	0	0	0	0	0	4	0	0	0	4	1.00
Petroicidae	2	0	1	4	0	0	0	1	0	2	7	0	0	0	0	27	0	4	48	0.56
Ptilonorhynchidae	0	0	0	0	0	0	0	0	0	0	0	0	2	0	0	2	21	0	25	0.84
Rhipiduridae	0	0	1	0	0	0	0	0	0	1	6	0	0	0	0	6	0	22	36	0.61

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758

759 Table S8. Comparisons of evolutionary models testing for a decoupling of rates of trait evolution  
760 between nectarivorous and non-nectarivorous lineages that accounted for phylogenetic uncertainty.  
761 The best-fitting models are highlighted in bold. Shown are the mean and standard deviations of  
762 delta AICc values, AICc weights, and Brownian variance ( $\sigma^2$ ) as estimated across 1,000 (univariate  
763 analyses of pPC1-4) and 10 (multivariate analysis of pPC1-4) stochastic character maps of the  
764 evolutionary history of nectarivorous diet among the Meliphagides. The character maps were  
765 generated by running a single simulation across each tree in the posterior distribution of 1,000 trees  
766 obtained from the study of Marki et al. (2017).

	pPC1	pPC2	pPC3	pPC4	pPC1-4
BM					
Delta AICc	7.9±3.7	16.0±33.9	<b>1.4±8.9</b>	9.8±6.0	21.7±21.2
Akaike weight	0.06±0.11	0.05±0.06	<b>0.59±0.20</b>	0.04±0.11	0.05±0.12
$\sigma^2$	0.023±0.003	0.008±0.006	<b>0.002±0.001</b>	0.001±0.000	0.035±0.005
BMM					
Delta AICc	<b>0.0±0.2</b>	<b>0.0±0.0</b>	1.2±0.8	<b>0.0±0.2</b>	<b>0.0±0.0</b>
Akaike weight	<b>0.94±0.11</b>	<b>0.95±0.06</b>	0.40±0.20	<b>0.96±0.11</b>	<b>0.95±0.12</b>
$\sigma^2$ (nectarivorous)	<b>0.028±0.003</b>	<b>0.005±0.001</b>	0.002±0.001	<b>0.002±0.000</b>	<b>0.037±0.004</b>
$\sigma^2$ (non-nectarivorous)	<b>0.015±0.003</b>	<b>0.012±0.016</b>	0.002±0.001	<b>0.001±0.000</b>	<b>0.030±0.006</b>

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775 **Figure legends**

776 Figure 1. Diet and body size evolution among the Meliphagides. (a) Phylogeny of the Meliphagides  
777 with ancestral estimation of the presence (red) or absence (yellow) of nectar in the diet.  
778 Reconstructions were performed using stochastic character mapping and summarized using the  
779 function *densityMap* in the R package *phytools*. (b) The phylogeny with branch lengths scaled by  
780 the mean rate of body size (pPC1) evolution as estimated using the variable-rates model in  
781 *BayesTraits*. Branch coloring reflects the relative rate of evolution. (c) Accumulation of size  
782 disparity through time for the overall radiation (black), honeyeaters (red) and background  
783 meliphagoids (blue). The black triangles show the stem branch of honeyeaters. Illustrations are  
784 watercolors by Jon Fjeldså showing (clockwise from top) crow honeyeater (*Gymnomyza aubryana*),  
785 mao (*Gymnomyza samoensis*), Meyer's friarbird (*Philemon meyeri*), cardinal myzomela (*Myzomela*  
786 *cardinalis*) white-throated grasswren (*Amytornis woodwardi*), variegated fairywren (*Malurus*  
787 *lamberti*), large-billed gerygone (*Gerygone magnirostris*), white-browed scrubwren (*Sericornis*  
788 *frontalis*), western spinebill (*Acanthorhynchus superciliosus*), tui (*Prothemadera*  
789 *novaeseelandiae*), gibberbird (*Ashbyia lovensis*), MacGregor's honeyeater (*Macgregoria pulchra*),  
790 orange-cheeked honeyeater (*Oreornis chrysogenys*), and Belford's melidectes (*Melidectes belfordi*).  
791

792 Figure 2. Morphospace of Australasian passerine birds. Morphological diversity of honeyeaters ( $n =$   
793 180 species) (a, b) compared to that of the four background meliphagoid families ( $n = 93$  species),  
794 as well as 13 additional Australasian passerine families ( $n = 398$  species) (c, d) as visualized using  
795 the four first axes of variation from a phylogenetic and standard principal component analysis,  
796 respectively. Principal components for the two sets of comparisons were generated separately.  
797

798 Figure 3. Spatial diversity patterns of honeyeaters compared to that of background meliphagoids in  
799  $1^\circ \times 1^\circ$  grid cells. Comparisons between honeyeaters (left) and background meliphagoids (right) for  
800 differences in species richness (*a*), range and standard deviation of pPC1 (*b* and *c*) are shown.

801

802 Figure 4. Results of linear models examining the relationships between spatial diversity patterns.  
803 The panels show the relationships between (*a*) grid cell richness of the honeyeaters and background  
804 meliphagoids, grid cell richness of both groups and their range (*b*) or standard deviation (*c*) of  
805 pPC1. Points represent the values in each  $1^\circ \times 1^\circ$  grid cell. Line in (*a*) is the 1:1 line, whereas lines  
806 in (*b*) and (*c*) are the least-squares regression fits.

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821 **Supplementary figure legends**

822 Figure S1. Mean shift configurations of BAMM analysis of pPC1-4.

823

824 Figure S2. Pairwise plots showing the estimated four-dimensional pPC hypervolumes for  
825 honeyeaters (red points) and background meliphagoids (blue). Solid points reflect the empirical  
826 data, whereas translucent points represent the stochastic points sampled from the estimated  
827 hypervolumes. Large points represent the hypervolume centroids.

828

829 Figure S3. Pairwise plots showing the estimated four-dimensional pPCA hypervolumes for  
830 honeyeaters (red points) and regional passerines (blue). Solid points reflect the empirical data,  
831 whereas translucent points represent the stochastic points sampled from the estimated  
832 hypervolumes. Large points represent the hypervolume centroids.

833

834 Figure S4. Distribution of discriminant scores for honeyeaters (top panel) and other regional  
835 passerines (bottom panel). Large negative scores reflect species with long and narrow (width) bills,  
836 and characterize honeyeaters relative to other groups.

837

838 Figure S5. Results from the BayesTraits variable-rates analysis of pPC1-4. Branch lengths are  
839 scaled by the mean rate of evolution with branch coloring reflecting the relative rate of evolution.  
840 Colored circles show rate shifts on individual internal branches, whereas colored triangles indicate

841 support for a whole-clade shift in rate of trait evolution. The relative size of the circles and triangles  
842 indicate the support (posterior probability) for a rate shift.

843

844 Figure S6. Accumulation of morphological disparity through time (pPC1-4) for the Meliphagides  
845 (solid black line), with separate lines for the honeyeaters (solid red line) and background  
846 meliphagoids (solid blue line). Shading shows the expected accumulation under a constant-rate BM  
847 model of trait evolution.

848

849 Figure S7. Accumulation of morphological disparity through time (pPC1-4) for the Meliphagides  
850 (solid black line), with separate lines for the honeyeaters (solid red line) and background  
851 meliphagoids (solid blue line). Shading shows the expected accumulation under a variable-rates  
852 model of trait evolution.

853

854 Figure S8. Phenograms of morphological disparity through time (pPC1-4) for the Meliphagides  
855 with separate coloration for the honeyeaters (red) and background meliphagoids (black).

856

857 Figure S9. Model-averaged speciation rates among the Meliphagides as inferred using the hidden-  
858 state speciation and extinction (HiSSE) framework. Ancestral estimation of diet is represented by  
859 white and black branches for nectarivorous and non-nectarivorous lineage respectively. The inset  
860 histogram shows the density distribution of speciation rates in the phylogeny.

861

862 Figure S10. Spatial diversity patterns of honeyeaters compared to that of background meliphagoids  
863 in  $1^\circ \times 1^\circ$  grid cells. Comparisons between honeyeaters (left) and background meliphagoids (right)  
864 for differences in the range and standard deviation of pPC2-4 are shown.

865

866 Figure S11. Spatial diversity patterns of honeyeaters compared to that of background meliphagoids  
867 in  $1^\circ \times 1^\circ$  grid cells. Comparisons between honeyeaters (left) and background meliphagoids (right)  
868 for differences in the 95% quantile range of pPC2-4 are shown.

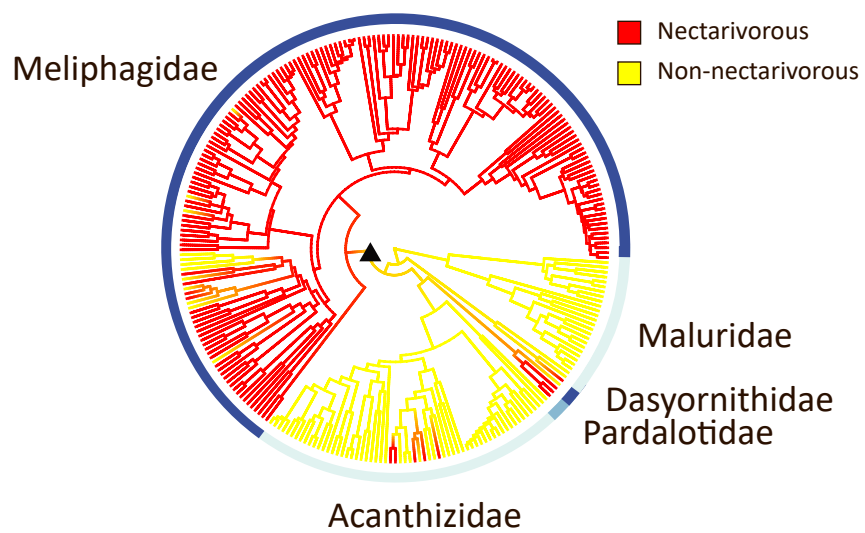
869

870 Figure S12. Results of linear models examining the relationships between spatial diversity patterns.  
871 The panels show the relationship between species richness and range (left) and standard deviations  
872 (right) of pPC2-4 for each of the two groups. Points represent  $1^\circ \times 1^\circ$  grid cell values. Lines are the  
873 least-squares regression fits.

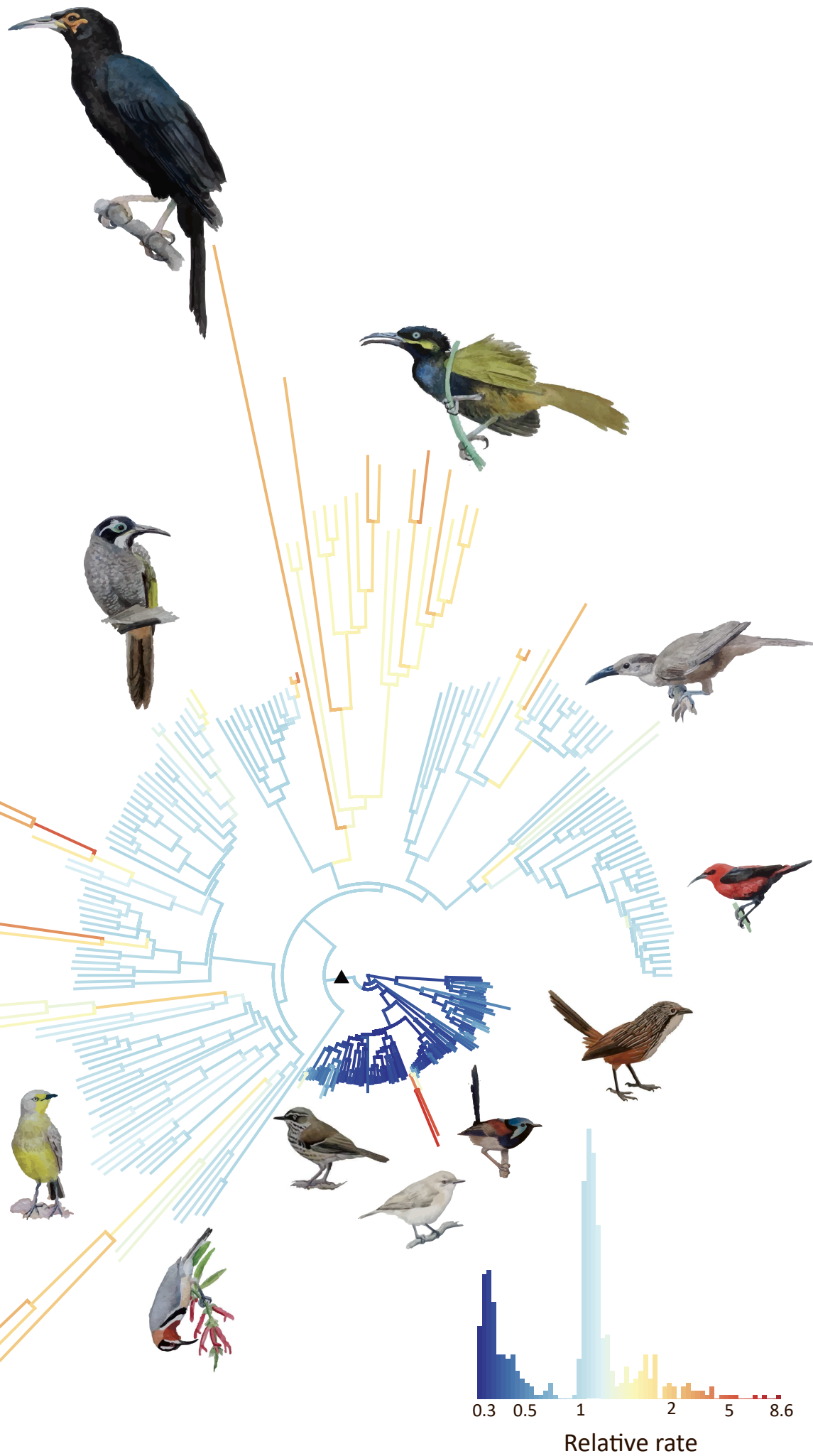
874

875 Figure S13. Relationship between pPC2 and the log-transformed Kipp's distance values.

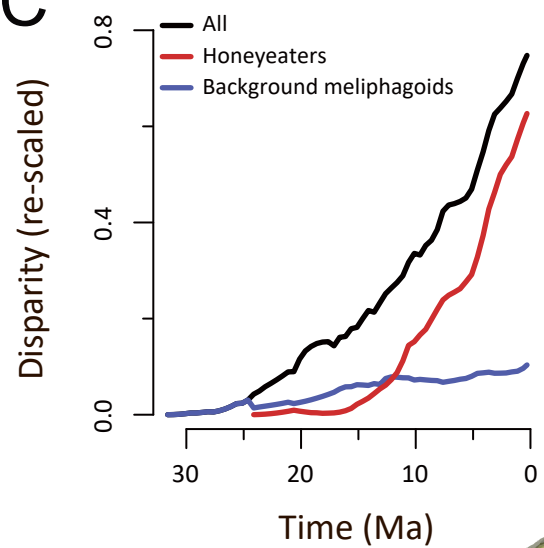
A

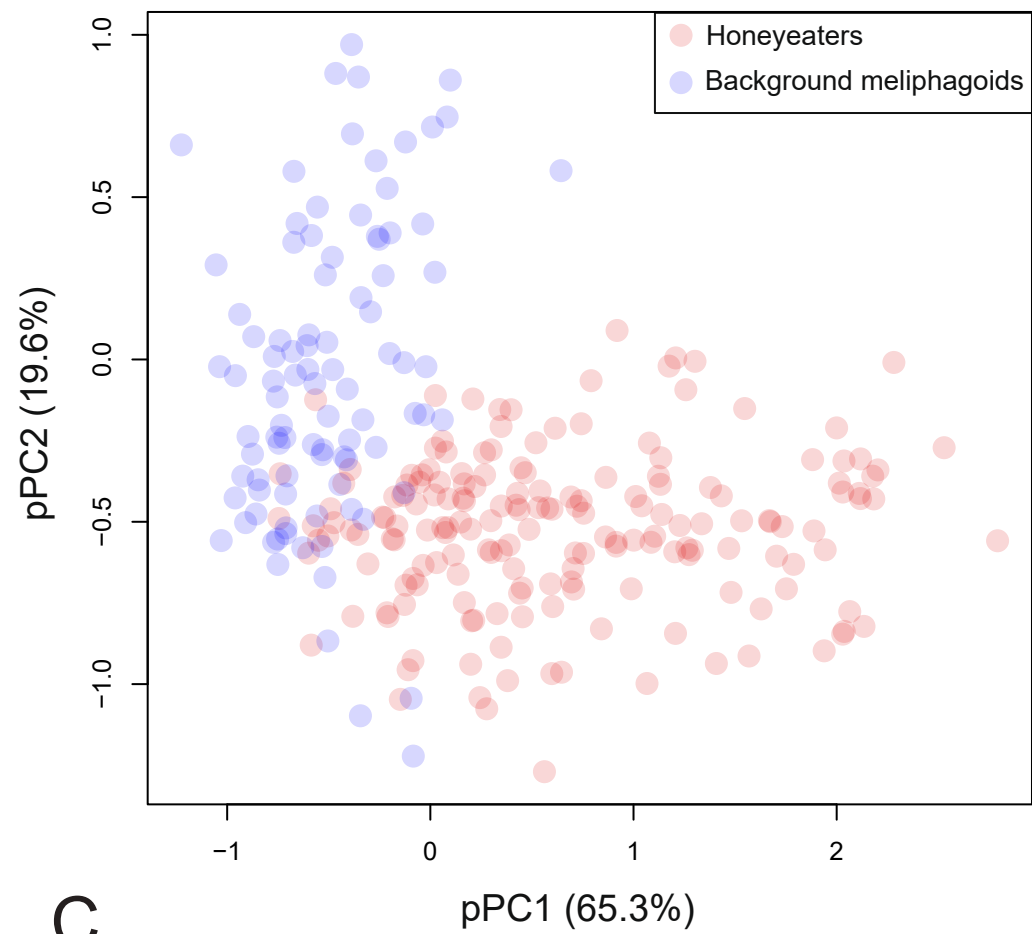
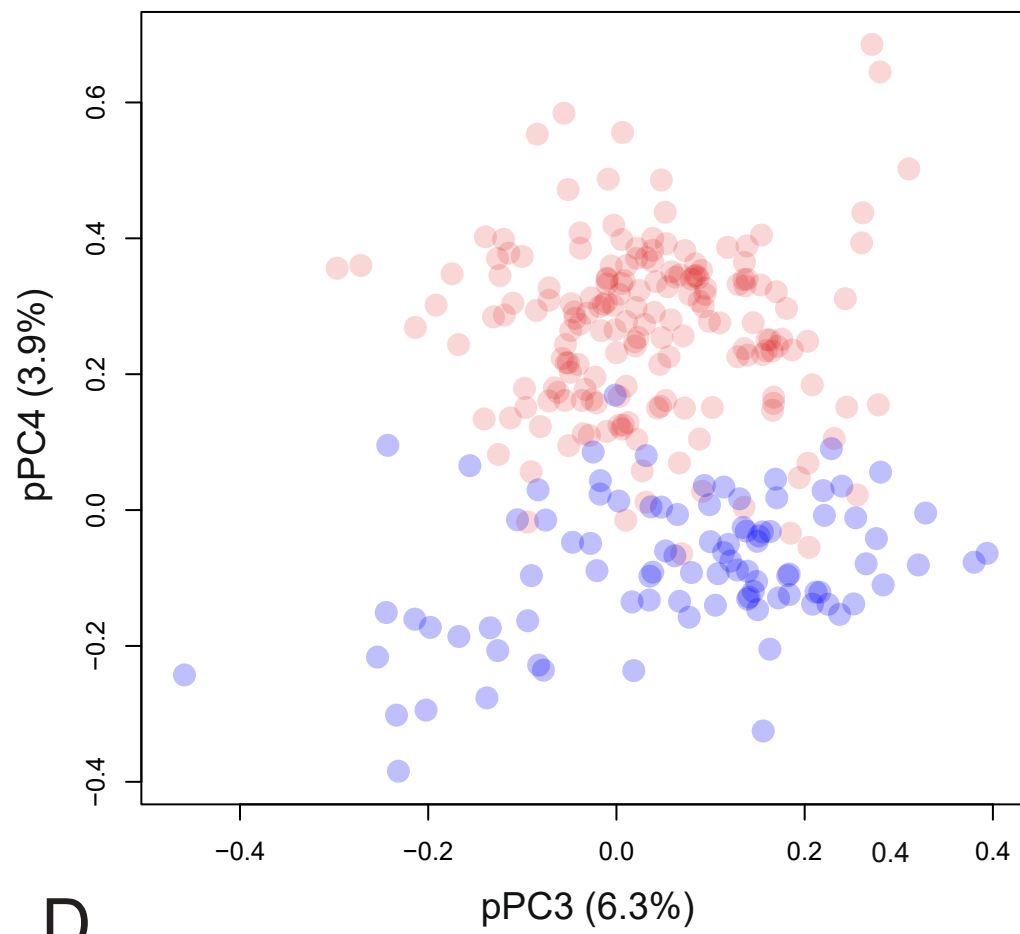
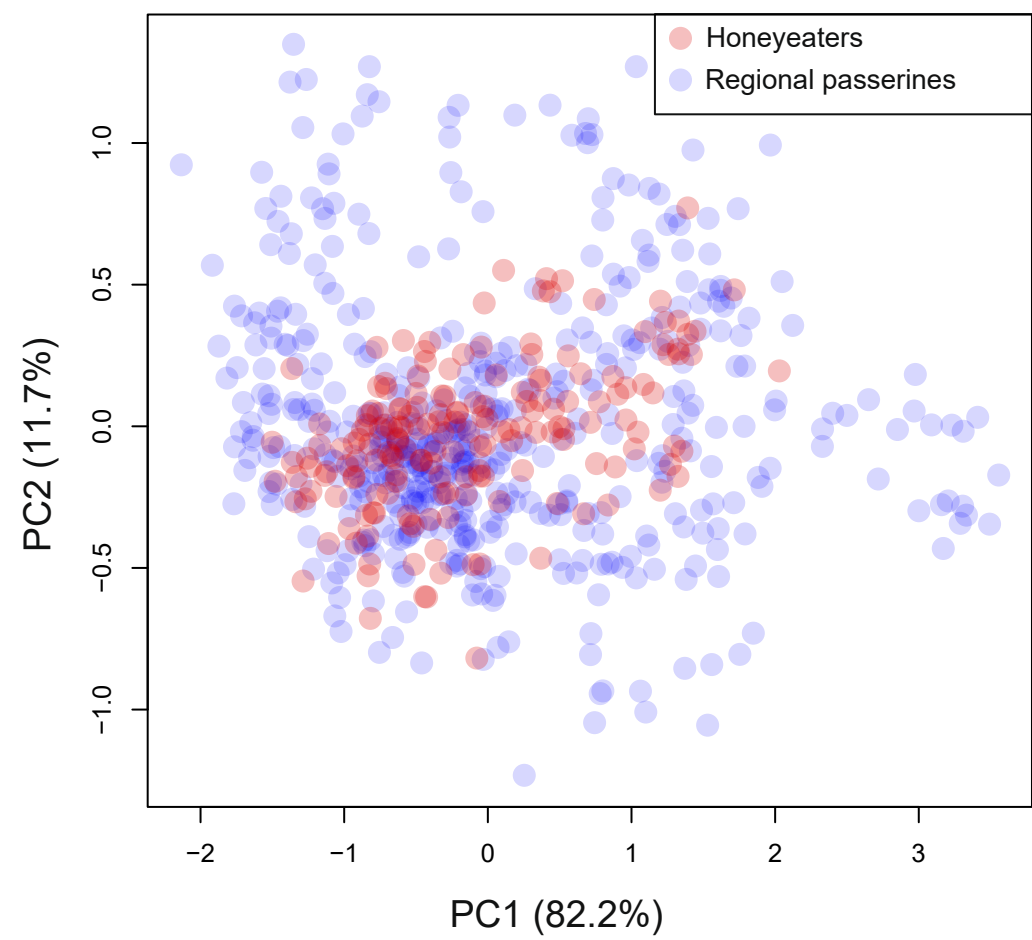
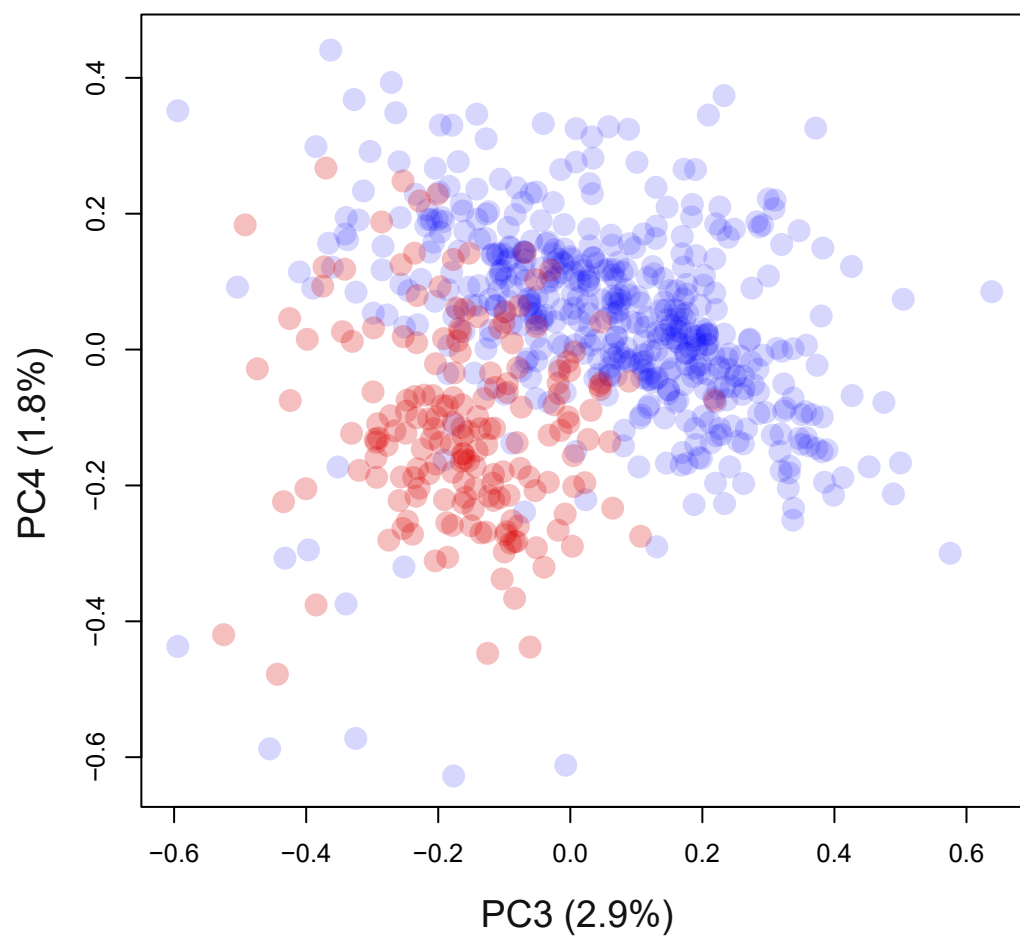


B



C



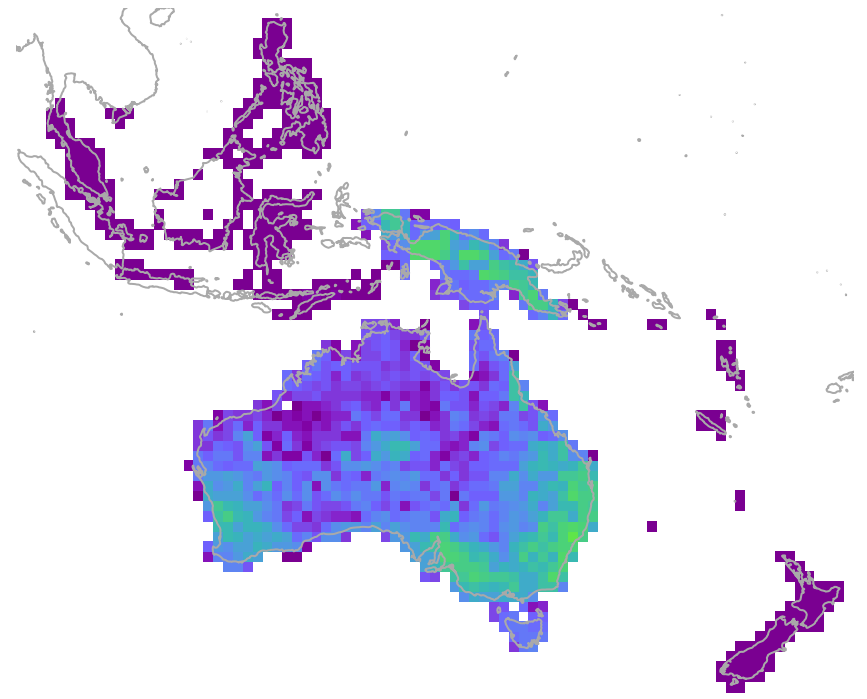
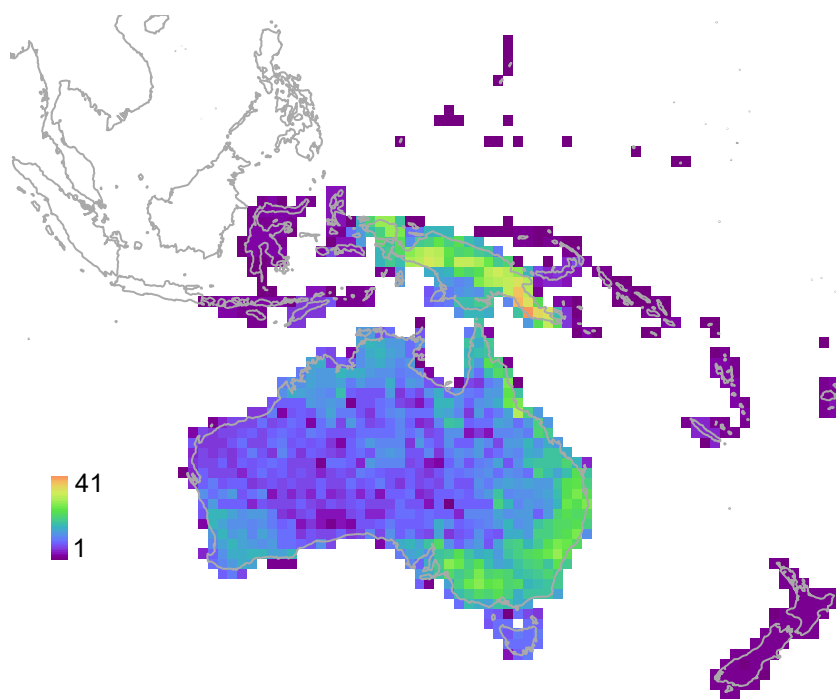
**A****B****C****D**

# Honeyeaters

# Background meliphagoids

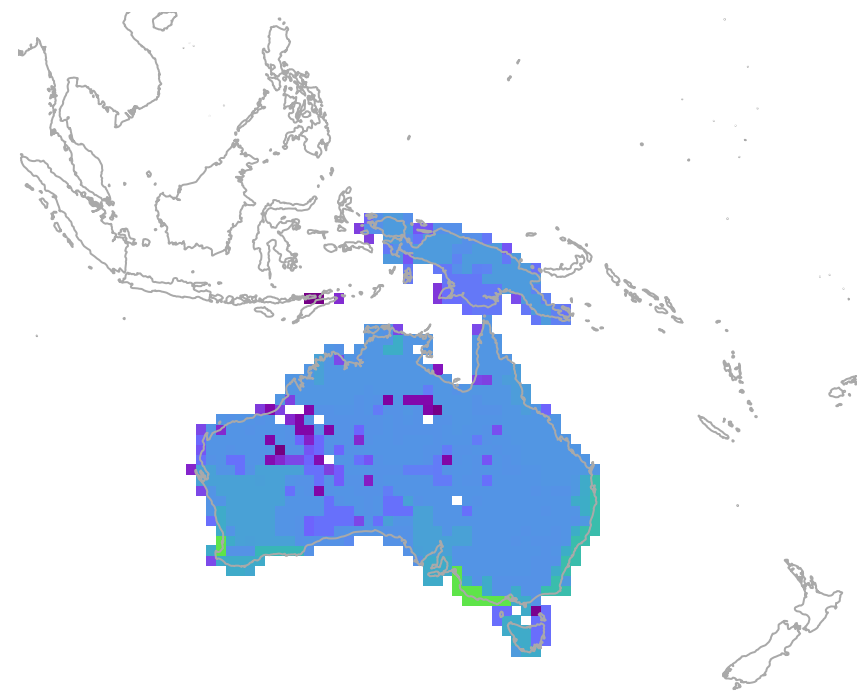
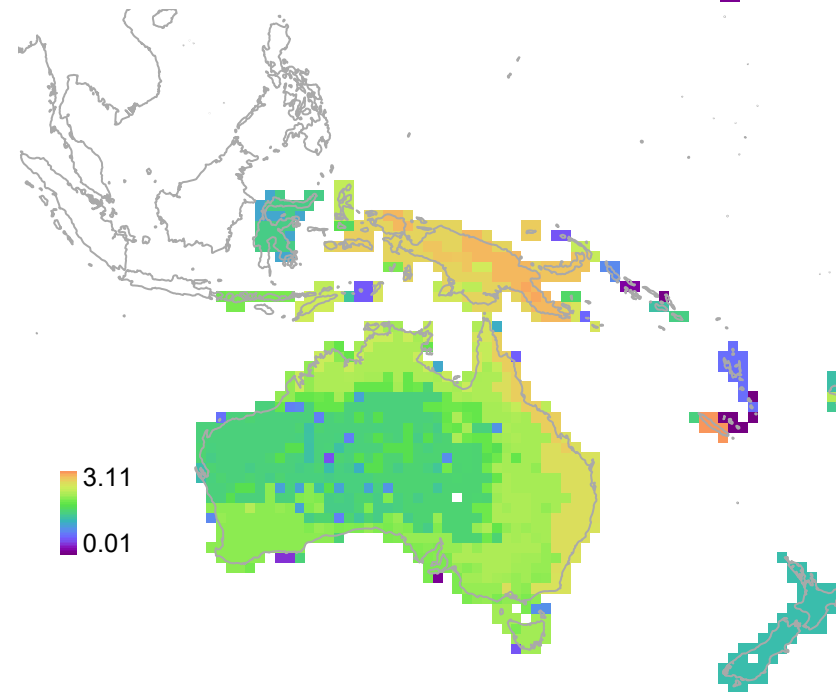
## A

Species richness



## B

Range of pPC1



## C

SD of pPC1

