

Arefin, S., Buchanan, S., Hobson, S., Steinmetz, J., Alsalhi, S., Shiels, P. G., Kublickiene, K. and Stenvinkel, P. (2020) Nrf2 in early vascular ageing: calcification, senescence and therapy. *Clinica Chimica Acta*, 505, pp. 108-118. (doi: <u>10.1016/j.cca.2020.02.026</u>)

The material cannot be used for any other purpose without further permission of the publisher and is for private use only.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

http://eprints.gla.ac.uk/210786/

Deposited on 21 February 2020

Enlighten – Research publications by members of the University of Glasgow <u>http://eprints.gla.ac.uk</u>

Nrf2 in early vascular ageing: calcification, senescence and therapy

Samsul Arefin¹, Sarah Buchanan², Sam Hobson¹, Julia Steinmetz³, Shno Alsalhi^{1,4}, Paul G Shiels², Karolina Kublickiene¹, Peter Stenvinkel^{1,*}

- 1. Division of Renal Medicine, Department of Clinical Science, Karolinska University Hospital, 14186 Stockholm, Sweden
- 2. Institute of Cancer Sciences, Wolfson Wohl CRC, ICS, MVLS, University of Glasgow, Glasgow, UK
- 3. Rheumatology Unit, Dep. of Medicine, Solna, Karolinska Institutet, Karolinska University Hospital, SE-171 76, Stockholm, Sweden
- 4. Research Center, Salahaddin University-Erbil, 44001 Erbil, Kurdistan-Region, Iraq

Correspondence:	Peter Stenvinkel, MD, PhD
	Department of Renal Medicine M99
	Karolinska Institutet
	Karolinska University Hospital at Huddinge
	141 86 Stockholm, Sweden
	E-mail: peter.stenvinkel@.ki.se

*

Abstract

Under normal physiological conditions, free radical generation and antioxidant defences are balanced, and reactive oxygen species (ROS) usually act as secondary messengers in a plethora of biological processes. However, when this balance is impaired, oxidative stress develops due to imbalanced redox homeostasis resulting in cellular damage. Oxidative stress is now recognized as a trigger of cellular senescence, which is associated with multiple chronic 'burden of lifestyle' diseases, including atherosclerosis, type-2 diabetes, chronic kidney disease and vascular calcification; all of which possess signs of early vascular ageing.

Nuclear factor erythroid 2-related factor 2 (Nrf2), termed the master regulator of antioxidant responses, is a transcription factor found to be frequently dysregulated in conditions characterized by oxidative stress and inflammation. Recent evidence suggests that activation of Nrf2 may be beneficial in protecting against vascular senescence and calcification. Both natural and synthetic Nrf2 agonists have been introduced as promising drug classes in different phases of clinical trials. However, overexpression of the Nrf2 pathway has also been linked to tumorigenesis, which highlights the requirement for further understanding of pathways involving Nrf2 activity, especially in the context of cellular senescence and vascular calcification.

Therefore, comprehensive translational pre-clinical and clinical studies addressing the targeting capabilities of Nrf2 agonists are urgently required. The present review discusses the impact of Nrf2 in senescence and calcification in early vascular ageing, with focus on the potential clinical implications of Nrf2 agonists and non-pharmacological Nrf2 therapeutics.

Key words: oxidative stress; Nrf2; early vascular ageing; cellular senescence; calcification

1. Introduction

1.1. The Nrf2 signaling pathway

Redox homeostasis, comprising a balance between a pro-oxidative reactive oxygen species (ROS) production and concomitant antioxidant defenses, is a crucial process for protecting against oxidative stress known to be associated with a number of pathologies related to burden of life style diseases [1]. This balance is a determinant of physiological processes that ensure the maintenance of healthy cellular function in multiple organs, including the cardiovascular system [2]. Increasing evidence indicates that Nuclear factor erythroid 2-related factor 2 (Nrf2) acts as a key player in this process and that modulation of its action could facilitate both control of cellular redox homeostasis and physiological homeostasis in pathways related to the maintenance of cardiovascular health [3, 4].

Nrf2 is encoded by the nuclear factor erythroid-derived 2-like 2 (NFE2L2) gene, which is a basic-leucinezipper (bZIP) like transcription factor consisting of seven NRF2-ECH homology (Neh) domains (Neh1-Neh7), belonging to the cap'n'collar (Cnc) subfamily. Its spectrum of action is very broad, regulating the expression of >250 genes [5]. Although Nrf2 is a stress-responsive transcription factor with antiinflammatory and neuroprotective effects, its major function is to maintain cellular homeostasis by activating genes that encode cytoprotective, antioxidant and phase II detoxifying enzymes, such as NAD(P)H dehydrogenase (quinone)1 (NQO1), heme oxygenase (HO-1) and (HO-2), tryptophan hydroxylase-1 (TPH-1) and glutathione-S-transferase (GST) [6, 7].

Nrf2 is expressed ubiquitously and localized to the cytoplasm under basal conditions. Its expression is maintained at low levels through repression by Kelch-like ECH associated protein1 (Keap1) that functions like a molecular dimmer switch. The interaction between Keap1 and Nrf2 is mediated through the Neh2 domain [8]. Nrf2 activity can be induced by cellular stress, triggering nuclear translocation of Nrf2 and binding to antioxidant response elements (AREs) to orchestrate the transcription of target genes associated with a number of cellular functions including protein homeostasis, redox regulation, iron metabolism, DNA repair and prevention of apoptosis [9, 10].

Nrf2 activity can also be regulated via Keap1 independent, or other pathways (**Fig 1**) [11]. Under basal conditions, Keap1 homodimerizes, and together with the ubiquitination-ligase Cullin-3 (Cul3), inhibits the transcriptional activity of Nrf2 via ubiquitination and proteasomal degradation [12, 13]. Cysteine-rich elements in the protein structure of Keap1 account for its stress sensing activities. In particular, the cysteine C151, C273 and C288 are involved in post-translational modifications, such as oxidation or conjugation to

electrophiles [14]. These cysteine-modifications occurring at its cysteine-thiolate bridge, alleviate the interaction with the Cul3 ligase and therefore diminish Nrf2 proteasomal degradation. Oxidative stress, or Nrf2 activators, enable translocation of Nrf2 from the cytoplasm into the nucleus, where it heterodimerizes with small Maf proteins and transactivates an ARE battery of genes [15].

Recent evidence supports a Keap1 independent mechanism of Nrf2 regulation, whereby the Neh6 domain of Nrf2 plays a crucial role through binding via DSGIS and DSAPGS motifs to β -transducin repeatcontaining protein [16]. Another line of evidence suggests a non-canonical pathway for p62-dependent Nrf2 activation, where p62 sequesters Keap1, leading to transactivation of Nrf2-dependent genes [17]. Glycogen synthase kinase 3 β (GSK-3 β) has also been indicated to modulate the Nrf2 mediated oxidative stress response by promoting Keap1 independent degradation of Nrf2 [18]. In addition, recent evidence show that Nrf2 signaling links endoplasmic reticulum oxidative protein folding and calcium homeostasis in health and disease [19].

Currently, several experimental approaches have evaluated the capacity of Nrf2 to enhance the expression of oxidative stress defense genes and maintain vascular health [3]. However, the role and mode of action of Nrf2 within the complex phenotype of early vascular ageing (EVA) is not well understood. How Nrf2 expression is modulated in response to the structural and functional changes in the cardiovascular system as allostatic load accrues [20], remains to be determined.

For several decades oxidative stress has been recognized as a contributing factor to ageing and ageingassociated pathophysiology [21]. At the cellular level, features of vascular ageing include endothelial cell abnormalities, increased vascular smooth muscle cell (VSMC) growth, vascular inflammation, changes in the quantity and quality of extracellular matrix composition and calcification [22, 23]. Cellular senescence has also been implicated as a hallmark for age related disease [24]. The comprehensive involvement of calcification and senescence in EVA and the role of Nrf2 are discussed below.

1.2. Protective effects of Nrf2

Studies have shown that activation of Nrf2 prior to disease onset maintains general health [25, 26]. Kobayashi *et al.* [27] show that Nrf2 opposes transcriptional upregulation of pro-inflammatory cytokine genes. Calvert *et al.* [28] have demonstrated that hydrogen sulfide (H₂S) mediated Nrf2 expression induces cardioprotective effects, while overexpression of Nrf2 in endothelial cells decreases expression of Interleukin-1 beta (IL-1 β), Tumor necrosis factor (TNF), Vascular cell adhesion protein 1 (VCAM1), and Monocyte chemoattractant protein 1 (MCP-1) [29, 30]. Reduced Nrf2 activity leads to higher expression

of pro-inflammatory chemokines and adhesion molecules in endothelial cells [31]. Additionally, activation of Nrf2 has been shown to neutralize oxidative stress in T-lymphocytes, and prevent ischemia-reperfusion (IR) induced acute kidney injury (AKI) [32]. As Nrf2 dampens IR-induced AKI through its protective effects on resident renal epithelial cells [33], this suggests a novel protective mechanism against AKI. Moreover, supplementation of the Nrf2 agonist sulforaphane reduce environmental nephrotoxicity caused by arsenic in rats [34]. Furthermore, in mice with IR injury, activation of the Nrf2 signaling pathway arrested renal interstitial fibrosis [35]. Zheng *et al.* [36] show that natural products isolated from broccoli and cinnamon activate Nrf2 and reduce diabetic renal damage.

Electrophiles derived from polyunsaturated fatty acids, or other organic acids, activate the Nrf2 pathway, predominantly via reversible covalent nucleophile-electrophile modifications on key cysteines in Keap1. Electrophilic nitro-fatty acids inhibit VSMC growth via the activation of the Keap1/Nrf2 pathway [37]. Similar activities have been shown for 15-deoxy- $\Delta_{12,14}$ -PGJ2, a pro-resolving prostaglandin metabolite [38] and the lipid electrophile 4-hydroxynonenal (4HNE), which protected against ischemia-reperfusion injury in rodents and cell culture via Nrf2 activation [39, 40]. Another study show that overexpression of thrombomodulin domain-1 in diabetic mice improved renal function via enhancing the Nrf2 antioxidant pathway, resulting in decreased oxidative stress [41]. Additionally, anti-inflammatory effects can ameliorate diabetic nephropathy (DN) in db/db mice through downregulation of the NF- κ B mediated pathway [42]. The liver protects itself from harmful chemicals and their potentially damaging metabolites through several defense mechanisms, including the Nrf2/ARE pathway. Moreover, Nrf2 has a protective influence on survival rates and lung integrity in mice [43-45]. Taken together, as the balance between oxidants and antioxidants are crucial for maintaining normal cell signaling and function, Nrf2 has emerged as a major modulator of oxidant stress and implicated in a "Nrf2 diseasome" of chronic burden of life style diseases that are characteristically associated with oxidative stress and inflammation [46, 47].

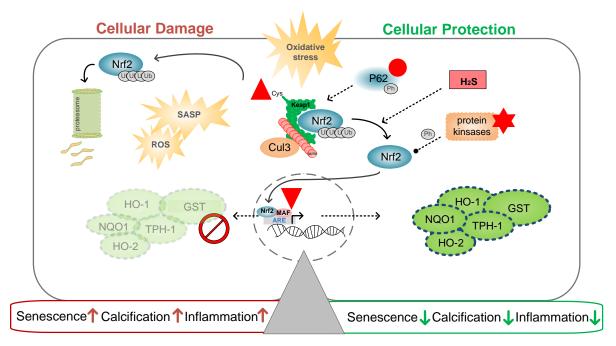


Figure-1: Overview of the role of Nrf2 to regulate the cellular senescence and calcification. Balance of downstream genes of Nrf2 is crucial for the maintenance of cellular homeostasis. Under normal physiological conditions, the cell senses oxidative and inflammatory stress and releases Nrf2 which protects the cell from calcification and cellular senescence via the activation of protective signaling pathways including NQO1, HO-1, HO-2, TPH-1, GST and miRNAs. On the contrary, if the Nrf2 pathway is dysregulated imbalance of Nrf2 response genes occurs and the cell is no longer protected from pro-inflammatory and oxidative stress promoting the development of vascular pathologies. The Nrf2 pathway can be modulated in several steps of the signaling cascade (\blacktriangle cysteine modification in Keap1, • p62 activation, \checkmark ARE activation, *GSK inhibition). **Abbreviations:** Nrf2, nuclear factor erythroid 2-related factor 2; Keap1, kelch-like ECH associated protein1; ROS, reactive oxygen species; H₂S, hydrogen sulfide; NQO1, NAD(P)H dehydrogenase (quinone) 1; HO-1, heme oxygenase-1; HO-2, heme oxygenase-2; TPH-1, tryptophan hydroxylase-1; GST, glutathione-S-transferase; GSK-3, glycogen synthase kinase 3; SASP, senescence associated secretory phenotype; Cul-3, cullin 3; Ub, ubiquitination; Ph-phosphorylation.

2. Nrf2 in pathological conditions

Insufficient Nrf2-dependent gene expression is associated with a number of distinct pathologies associated with ageing. The development of genetically engineered mouse models of human disease has improved our understanding about the importance of Nrf2 signaling in health and disease. Below in Table 1 we present examples of common diseases associated with EVA and status of Nrf2.

Table-1: Nrf2 impacts on various early vascular ageing associated diseases.	Table-1: Nrf2 im	pacts on various	early vascula	r ageing as	sociated diseases.
---	------------------	------------------	---------------	-------------	--------------------

Pathological	Mechanism	Findings	Refs
condition			

Ageing	Process of becoming older	 Nrf2 dysfunction underlying impaired angiogenesis and microvascular rarefaction in aging Nrf2 levels decrease with age Exposure to nano particles induce Nrf2 regulated detoxifying enzymes in young but not older mice cerebellum, liver, and lung Loss of Nrf2 activity in intestinal stem cells accelerates age-related degeneration of the 	[48] [49] [50] [51]
		intestinal epithelium in Drosophila	
Renal disease	Progressive loss of renal function	• Nrf2 knockout mice showed a greater sensitivity to renal damage compared to wild-type mice	[52]
		• Uremic toxin Indoxyl sulfate decrease Nrf2 transcriptional activity in rats	[53]
		• Nrf2 deficiency associated with hypertensive kidney	[52]
		• Impaired Nrf2 activation leads to progression of renal fibrosis	[54]
		• Nrf2 hyperactivation in Keap1 deficient mice showed a bilateral hydronephrosis as indicated by severe bladder swelling	[55]
Atherosclerosis	Narrowing the artery lumen due to build up	• Nrf2 signaling pathway is related with atherosclerosis development	[56]
	plaque	• Nrf2 exhibits both pro- and anti-atherogenic effects in experimental animal models	[56]
Hypertension	Elevated blood pressure	• Selective Nrf2 gene deletion in the rostral ventrolateral medulla (RVLM) evokes hypertension and sympathico-excitation in mice	[57]
		• Impaired Nrf2 regulation of mitochondrial biogenesis in RVLM on hypertension induced by systemic inflammation	[58]

Diabetic Cardio- myopathy	Disorder of the heart muscle in diabetes	 Reduced Nrf2 expression observed in the left ventricle of diabetic patient Nrf2 and its downstream target genes are downregulated in cardiomyocytes from diabetic (db/db) mice 	[59] [60]
Cancers	Uncontrolled cell proliferation	• Nrf2 has both tumors suppressive and tumor- promoting effects in cancers	[61]
Renal cancer	Uncontrolled renal cell proliferation	 Modifications of the Nrf2, Keap1-Cul3 complex allow the activation of Nrf2 to aid the survival of tumor In renal cell cancer, mutations in fumarate 	[62] [63]
		hydratase results uncontrolled upregulations of Nrf2 target genes via Keap1	[64]

3. Nrf2 in early vascular ageing

Vascular ageing develops as a progressive modification of vascular function and structure towards increased arterial stiffening. Early vascular ageing can be described as accelerated or aberrant ageing [65], and is a process associated with impairment of physiological functions and an increased risk of further morbidity and mortality [66]. If undetected, EVA leads to vascular stiffening and earlier development of cardiovascular disease (CVD) [65]. The free radical theory of ageing, developed by Harman in the 1950s [67], states that excessive oxidative stress results in ageing through an accumulation of cellular damage. However, since overexpression of antioxidant enzymes, including zinc superoxide dismutase and catalase, does not extend life span in mice [68], oxidative stress might not be a unique mechanism for triggering the vascular ageing process. Increased ROS production induces macromolecular oxidative modifications that promote oxidative damage. With increasing age, oxidative stress accrues, both in humans and animals [69-71] and is closely associated with vascular aging resulting from a failure to activate ARE-driven gene expression and dysregulation of the Nrf2-ARE pathway [72]. In EVA, increased production of ROS promotes endothelial dysfunction, a pathological phenotype associated with the development of stroke, hypertension, atherosclerosis, myocardial infarction and vascular dementia [73].

As a partner in crime, chronic, low grade inflammation ("inflammaging") is strongly linked with oxidative stress [74]. Most age-related 'burden of life style' diseases share an underlying inflammatory component [47, 74]. Although inflammaging correlates with reduced health span [66], its etiology remains to be fully determined. Reducing systemic inflammation and associated oxidative stress has been suggested as a means of mitigating the progression of premature ageing processes [75]. Chronic kidney disease (CKD) is a common condition with underlying premature ageing, due to a complex of toxic alterations in the internal milieu [66]. Kooman *et al.* [75] have proposed four major mechanisms underlying premature ageing in CKD; increase in allostatic load, activation of the stress resistance response, activation of age-promoting mechanisms and impairment of anti-ageing pathways. Patients with CKD are especially vulnerable to EVA associated with cellular senescence, vascular calcification (VC) and depressed Nrf2 expression [76].

3.1. Nrf2 and vascular calcification

Vascular calcification is a pathophysiological process characterized by the deposition of calcium-phosphate crystals in the arteries, typically developing in the intima and media of the vascular wall. The presence of vascular calcification is often detected in CKD, diabetes, atherosclerosis, heart failure and other disorders characterized by changes in vascular structure (i.e. stiffening). The development of calcification is considered to be an active response to the environmental stimuli, such as oxidative stress, inflammation, and changes in passive elements of vascular wall, together with increased phosphate and calcium levels [77]. The role of Nrf2 in the calcification process has been increasingly appreciated due to its regulatory function in the antioxidant and anti-inflammatory pathways [27].

The H₂S donor, sodium hydrosulfide (NaHS) ameliorates calciprotein particles-induced calcification *in vitro* via Keap1/Nrf2 activation system [78]. This inhibitory effect on calcification was achieved by increased expression of the downstream NQO1 gene, and the calcification inhibiting effect was lost and NQO1 expression was reduced when Nrf2 was silenced (**Fig 1**) [78]. Whereas silencing of Keap1 alone does not have a strong impact on calcification, NaHS treatment mediated a significant decrease of TNF mRNA in VSMCs, suggesting anti-inflammatory effects for NaHS [78]. An upregulation of Nrf2 and subsequent increase of HO-1 and -2 expressions was reported in rat aorta upon H₂S treatment [79]. Cell culture experiments show that overexpression of Nrf2 attenuates the process of cellular bone differentiation by interfering with runt-related transcription factor 2 (Runx2) [80]. Thus, Nrf2 deletion results in an increased expression of Runx2 [80]. Since *in vitro* assays using VSMCs treated with the Nrf2 agonist resveratrol show a significantly reduced mineralized matrix deposition, the protection against oxidative stress-induced mitochondrial damage and reduced intracellular calcium deposition could be achieved via Nrf2 and Sirtuin1 signaling [81]. As resveratrol increases the mRNA levels of klotho and Nrf2 in VSMCs

after calcification, this drug may improve the anti-oxidative effect of Nrf2 against hyperphosphatemiainduced calcification [81]. Considering that hyperphosphatemia reduces both mRNA and protein expression of Nrf2 in VSMCs culture [81], high phosphate levels may impair the anti-oxidative role of Nrf2. Indeed, elevated phosphate (even within the normal range) has been associated with poorer outcome [82] with coronary atherosclerosis in young healthy adults [83] and microvascular dysfunction [84]. As tertbutylhydroquinone alleviates high phosphate-induced calcification in VSMCs by suppressing ROS production [85] it is evident that the salutary effects of Nrf2 agonists on VC is not restricted to H_2S and resveratrol and may be a class-effect. Indeed, the classic Nrf2 activator, dimethyl fumarate (DMF) significantly attenuated VC in an in vitro ring culture system using mouse thoracic aorta and rat carotid artery [86] under hypercalcemic and hyperphosphatemic conditions. DMF inhibited VC by activating Nrf2 and downregulating osteogenic marker expression in VSMCs [86]. Since Yao et al. [87] demonstrated that the Nrf2-ARE signaling pathway enhance the autophagy of VSMC to reduce hyperphosphatemia-induced VC, several mechanisms contribute to the beneficial effects of Nrf2 agonists. It has been proposed that hydrogen peroxide (H_2O_2) can efficiently guard VSMCs against oxidative stress by preventing development of VC triggered by ROS production through Nrf2-ARE pathway [88]. More studies are warranted to understand the complexity of how Nrf2 contribution could be linked to VC and if pharmacological and/or nutraceutical activation of Nrf2 could have beneficial therapeutic effects in groups with high risk of EVA.

3.2. Nrf2 and Senescence

Senescence is characterized as a state of cellular growth arrest, in which the cells are resistant to apoptosis. In essence, senescence acts as an anti-oncogenic mechanism [89]. Although senescent cells are metabolically active they are not positively physiologically contributory to the tissue or organ in which they reside [90]. External stimuli like ROS, high glucose, fatty acid, DNA damage, oncogenes, inflammation and proteotoxic environment act as triggers to render healthy cells senescent [89]. Senescent cells secret a pro-inflammatory senescence associated secretory phenotype (SASP), which is enriched in pro-inflammatory, pro-fibrotic and matrix degrading factors, consisting of chemokines, cytokines, proteases and growth factors that poison the surrounding tissue and contribute to organ dysfunction [91]. One noticeable feature of the SASP is its capacity to activate senescence in neighboring cells via a bystander effect [89]. We have reported that VC in uremic arteries is characterized by increased *CDKN2A/p16INK4^a* expression indicating senescence [92].

Numerous studies have investigated the mechanisms behind the age-associated decline in Nrf2 expression in number of different cell types, including bronchial epithelial cells [93], vascular cells [94] and cardiomyocytes [95]. Kuosmanen *et al.* [96] demonstrated that miRNAs derived from senescent cells (most

notably miR-126, miR-21 and miR-100) modulate Nrf2 expression in aged endothelial cells; a process mediated by directly targeting on Nrf2 mRNA (**Fig 1**). It is likely just one of many mechanisms that drive Nrf2 depletion. Moreover, Nrf2 deficiency triggers the development of cellular senescence, as aged Nrf2 double knockout (KO) mice present with increased expression of senescence markers p16INK4a (CDKN2A) and p21 (CDKN1A) compared to wild-type aged mice [97]. Additionally, well-established components of the SASP, such as IL-1 β and TNF, are elevated in Nrf2 KO mice [97]. Furthermore, Zhou *et al.* [98] have reported that activation of Nrf2 protects VSMCs against angiotensin II-induced senescence, which was achieved via increased expression of downstream antioxidant genes like HO-1 and NQO1 (**Fig 1**). Although this suggest a bidirectional relationship between Nrf2 expression and senescence [99], further research is needed to fully elucidate how the repression of this adaptive response is induced and regulated during the ageing processes. The effects of Nrf2 deficiency and its relationship with senescence remain to be comprehensively studied in vascular cells [100], in order to asses possible interplay in diseases characterize by EVA.

It is important to stress that the effects of Nrf2 in CVD and other age-related diseases may be cell-specific [100], as exemplified by a discrepancy in gene expression patterns downstream of Nrf2 in different cell types, when comparing young and old *Drosophila* [51]. This is particularly pertinent, given that drivers of the ageing process are subject to antagonistic pleiotropy. In keeping with this consideration, Nrf2 activation in fibroblasts promote re-epithelialization of skin wounds, as well as gene expression profiles associated with tumorigenic activity also stimulated [101]. Taken together, therapeutic targeting of the Nrf2 pathway should be treated with caution, due to potential antagonistic pleiotropic effects.

4. Clinical implications/ therapeutics/interventions

In the multiple disease entities constituting a 'diseasome of ageing' [47, 102], with oxidative stress, mitochondrial dysfunction and persistent inflammation as common underlying features, the Nrf2-Keap1 signaling pathway is frequently disrupted. For example, Ruiz *et al.* [103] demonstrated that CKD-associated diminished antioxidant regulation, was largely caused by impaired activation of Nrf2. Natural Nrf2 activators, such as polyphenols, phytosterols and terpenoids (e.g. Baicalein), mitigate the effects of oxidative stress and reduce inflammation in pre-clinical models of kidney disease, while conversely Nrf2 deletion exacerbated disease pathogenesis and led to autoimmune nephritis [104]. Consequently, restoration and modulation of Nrf2 mediated signaling networks may be effective in retarding CKD progression and EVA [86, 87, 99].

Multiple approaches, such as non-calcium phosphate binders, calcium containing phosphate binders [105, 106], statins [107-109] and vitamin D [110] targeting VC in CKD have been used with varied results and are not curative [111]. A novel approach has been the targeting and removal of senescent cells by senolytic compounds. The use of a senolytic combination (Dasatinib and Quercetin) to specifically remove senescent cells, has been shown to increase the lifespan and improve health span for normatively aged mice [112]. The chronic clearance of senescent cells by combined treatment with Dasatinib and Quercetin has also been shown to alleviate vasomotor dysfunction in normatively aged mice and in mice with established atherosclerosis [113]. This strategy reduced markers of osteogenesis in advanced intimal plaques and ultimately reduced intimal plaque calcification [113]. A recent clinical trial has also reported that treatment with a combination of Dasatinib and Quercetin reduced adipose tissue senescent cell burden in diabetic kidney disease [114].

Specific targeting of the SASP has also proven beneficial. Hegner et al. [115] reported that the pharmacological blocking of pro-inflammatory cytokines reduces uremia-induced calcification in vascular progenitor cells in vitro. However, several safety and efficacy issues around the removal of senescent cell needs to be addressed. These include the potential for an acceleration of stem cell exhaustion, a hallmark of ageing. In keeping with this, Jeon et al. [116] have demonstrated that senescent cells reappeared after the cessation of senolytic treatment in a model of osteoarthritis. This is intuitive, as senescent cells are expected to be generated continuously over the life course in response to exposome stress. Additionally, there is also the possibility that removal of senescent cells without targeting the causes of their accumulation might limit the longer-term benefits of senolytics. The clinical population in which senolytics are targeted also needs to be considered, as it will necessarily include aged and or infirm patients with limited physiological reserve. Without clearance of apoptotic bodies, secondary necrosis could result in the release of pro-inflammatory, signals, further exacerbating the underlying chronic inflammation that occurs in such a population [24]. Another consideration is when in the life-course senolytics can be used. Under the aegis of antagonistic pleiotropy, removal of senescent cells may be appropriate in the later stages of the lifecourse, but not in the early stages where their requirement for wound healing processes may be desirable [112, 117].

Exploring alternative approaches to target the detrimental effects of senescence without resorting to senolytics therefore need to continue. One such approach would be targeting the SASP. Candidates to suppress or modulate the SASP include rapamycin, NF- κ B, or p38 inhibitors [118-121]. Side effects of this approach could include blunting the senescent response, immunosuppression or exacerbation of the accumulation of senescent cells [24].

With the growth in the discovery of Nrf2 activators and regulators, the pharmacological targeting of the regulation of the Nrf2-Keap1 pathway however remains one of the most promising areas of research with multiple drugs or nutraceuticals currently in different stages of clinical trials, (table 2) most notably DMF, bardoxolone-methyl, sulforaphane and curcumin.

Compound	Class	Modulatory	Clinical Trial	Clinical Trials.gov
		effect		identifier
Dimethyl fumarate	Fumaric acid ester	Activator	I, II, III	NCT02784834
				NCT02546440
				NCT00810836
Bardoxolone-methyl	Synthetic	Activator	I, II, III	NCT00550849
	triterpenoids			NCT00811889
				NCT01351675
Oltipraz	Organosulfur	Activator	I, III	NCT00006457
	compound			NCT02068339
Ursodio	Biliary acid	Activator	I, II, III, IV	NCT02033876
				NCT00200343
				NCT01510860
Sulforaphane	Isothiocyanate	Activator	I, II, III, IV	NCT01008826
				NCT02880462
				NCT02801448
				NCT03220542
Curcumin	Stilbene	Activator	I, II, III, IV	NCT02104752
				NCT01225294
				NCT01052025
Resveratrol	(E)-Stilbene	Activator	I, II, III, IV	NCT01677611
	derivative			NCT01504854
				NCT00743743
				NCT02475564

Table-2: Nrf2 Modulators in clinical development

Sulforadex	Sulforaphane/alpha cyclodextrin complex	Activator	I, II	NCT01228084
Ebselen	-	Activator	I, II	NCT03013400
Complexa/ CXA-10	-	Activator	I, II	NCT02248051 NCT03449524 NCT03422510

4.1. Nrf2 pharmacological agonists and inhibitors, the "drugome"

The activation of Nrf2 has been demonstrated to be effective in inhibiting VC in animal models [86, 87, 99]. Various Nrf2 agonists (e.g. Bardoxolone–methyl), have anti-atherogenic and reno-protective effects [122-125], via the upregulation of Nrf2 responsive genes. The main mechanism regulating Nrf2 activity is the control of protein stabilization by Keap1 and most known inducers are electrophilic molecules that covalently modify Keap1 cysteine residues [126].

A range of clinical regulators are in development or trailing [127, 128]. These include DMF which is currently in Phase III trials for the treatment of Multiple sclerosis (MS)s [129]. Activation of Nrf2 by DMF in the central nervous system has been demonstrated in a mouse model of MS. These effects were not observed in Nrf2 null mice, suggesting DMF acts exclusively via the Nrf2 pathway [130]. Bardoxolone methyl, a potent Nrf2 activator and NF- κ B suppressor has shown promising results in clinical trials [131], providing enhanced kidney function and delayed onset of ESRD in patients with type-2 diabetes and stage 4 CKD.

Nrf2 has not been considered just a target for treatment, but also for prevention of diseases like cancer thus the role of Nrf2 inhibitors are equally important. In laboratory trials many promising Nrf2 inhibitors have been tested. Different compounds of natural origin have been described to inhabit Nrf2 activity. Alkaloid trigonelline which can be retrieved from coffee beans, is one of these, which has been demonstrated to reduce Nrf2 accumulation into nucleus and ultimately inhibits Nrf2-driven genes transcription [132]. Evergreen shrub *Brucea javanica* extract brusatol is an agent that boosts ubiquitination of Nrf2 and accordingly reduces cytoplasmic Nrf2 levels [133]. Another natural compound mycotoxin ochratoxin A can prevent Nrf2 translocation [134]. Compared with Nrf2 activators data on Nrf2 inhibitors are still preliminary, more basic research and clinical trials are needed to estimate the definite outcome of Nrf2 inhibition.

4.1.1. KEAP1 independent drugs to target Nrf2

Whilst the vast majority of current drugs have typically focused on the interaction of Nrf2 with Keap1, Nrf2 is regulated on multiple levels: transcriptional, epigenetic, covalent protein modification and by proteasome degradation [135]. Several proteins can regulate the Nrf2-ARE pathway, mainly by phosphorylation, and Nrf2 comprises several sites for phosphorylation [136]. Attucks *et al.* [137] have demonstrated *in vitro* that modulation of the transcriptional repressor broad complex-tramtrack-bric a brac and Cap'n'collar homology (BACH) inhibited binding to some ARE-driven genes, independently of Keap1. A failure of redox homeostasis is a hallmark of neurodegenerative diseases, such as Alzheimer's disease (AD) and Parkinson's disease (PD). Several drugs alter the Keap1 independent regulation of Nrf2 in both PD and AD and have been employed in clinical trials with various levels of success [35, 138]. Chen *et al.* subsequently demonstrated that the overexpression of tryptophan hydroxylase-1 (TPH-1), an enzyme involved in metabolite 5-methoxytryptophan, (5-MTP), synthesis, reduced renal injury by attenuating renal inflammation and fibrosis, in a mouse model of CKD via the augmentation of Nrf2 independently of Keap1 [35].

4.1.2. Repurposing of drugs

The use of established therapeutics for new indications has been an area gaining recent attention, especially in the context of cancer treatment [139]. DMF, previously used as a MS and psoriasis drug has demonstrated improvement of MS symptoms in a murine model, via activation of Nrf2 [86, 130, 140]. Metformin, sulfuraphane, both involved in glucose metabolism, statins and GSK-3 inhibitors, have all been earmarked for the treatment of pathologies linked to Nrf2 [141]. Lithium treatment in adulthood or later in life, has been shown to extend lifespan in *Drosophila* via inhibition of GSK-3 and activation of Nrf2 independently of Keap1[142]. Recently Fujiki *et al.* have reported that Tolvaptan, a vasopressin type 2 receptor antagonist can regulate Nrf2 activity via the activation of the Nrf2/HO-1 antioxidant pathway, through phosphorylation of protein kinase RNA-like endoplasmic reticulum kinase [143].

4.2. Challenges and considerations of potential Nrf2 therapeutics

4.2.1. The role of Nrf2 in tumorigenesis

While diminished Nrf2 activity is a hallmark of the diseasome of ageing, its expression is elevated in tumors. As such, the biology of Nrf2 in cancer is complex and context dependent, with Nrf2 demonstrating both anti and pro-tumourgenic properties. It remains to be established if its activity also exhibits hormesis. In non-malignant cells, Nrf2 activation enhances cellular defenses, increasing resistance to oxidant-induced

genetic damage and chemical and physical carcinogens. The activation of a Nrf2 response results in the maintenance of ROS levels below those for signaling proteins critical for tumorigenesis e.g. PI3K, MAPKs and NF-κB. In the early stages of development, due to their enhanced ability to adapt to hostile microenvironment, or high ROS levels, malignant cells with constitutively active Nrf2 in conjunction with various oncogenic pathways, are positively selected. Somatic loss of function mutations in Keap1 or gain of function mutations in NFE2L2 are also common in several tumor types, promoting Nrf2 stability in cancer cells and resulting in unrestrained and sustained Nrf2 activation, [139, 144, 145]. The activation and augmentation of activity of Nrf2 by therapeutics (both synthetic or naturally occurring compounds), is often incongruous with this state of affairs, as such activity is both pulsatile and temporary. Some Nrf2 agonists may also have additional targets with anti-tumorigenic effects [146]. It is, however, encouraging that in the phase III trial of DMF, no difference in cancer rates between placebo and treatment groups was detected [129]. However, it is apparent that any Nrf2 therapeutic treatments will need a safe therapeutic window and require careful monitoring in order to assess potential cancer risk [139, 147].

4.2.2. Animal models, comparative biology and progeroid syndromes

Nrf2 may potentially cause or exacerbate age-related pathology [148]. Modeling the dynamics of Nrf2 in normative ageing is challenging, though Nrf2 null mice share remarkable similarities to old animals [149], including elevated levels of cellular senescence and features of inflammaging. Much insight into the complexity and regulation of the Nrf2 signaling network, can also be gained by research into the structure and function of Nrf2 across species, [47, 102, 150].

4.3. Non-pharmacological Nrf2 therapeutics

4.3.1. Nutrition and Nrf2

Hormesis describes an adaptive, non-monotonic biphasic dose response following an initial disruption in homeostasis. Inflammaging, can be regarded as hormetic stress. Evidence suggests that vitamins, minerals and phytochemicals can act in a hormetic manner [151-153]. Good nutrition can therefore be regarded as a powerful tool to redress the imbalance in pro and anti-inflammatory mediators via the modulation of the Nrf2 network (table 3). Martucci *et al.* [151] have demonstrated this in a project featuring a Mediterranean diet rich in poly unsaturated fatty acids and vegetables rich in nitrate and nitrite, which contribute to endogenous nitro-fatty acid formation and containing polyphenolic Nrf2 activators. Participants showed decreased inflammatory markers and an improved lipid profile via Nrf2 regulation of vitagene and heat shock proteins (Hsp) proteins [151, 154]. These data support growing evidence for the impact of nutrition

on Nrf2-Keap1 signaling pathway [151, 155-157]. Since high salt-loading down-regulates Nrf2 expression in kidney collecting duct cells [158] this suggest that high salt diets should be avoided.

Numerous naturally occurring chemicals derived from plants have anti-inflammatory and antioxidant properties regulated via Nrf2 (table 3). Though generally regarded as weaker Nrf2 agonists compared to synthetic chemicals, several natural compounds have been shown to be potent activators and can induce significant clinical effects [47, 159, 160]. Urolithin A, a metabolite derived from polyphenolics, has been demonstrated to upregulate tight junctions and reduced colitis via Nrf2 agonism [161]. Curcumin has also been shown to be effective in various chronic ageing diseases [162]. Sulforaphane has similarly been shown to have a reno-protective role via Nrf2 activation in diabetic rats subject to oxidative damage, and in Type-2 diabetes patients where it reduced fasting blood glucose levels [36, 151, 163].

Compound	Class	Mechanism	Food
Sulforaphane	Isothiocyanate	Keap1 cysteine modification	Broccoli
		stabilize Nrf2	Brussel
			Sprouts
			Cabbage
			Cauliflower
Curcumin	Diferuloylmethane	Keap1 thiol modification	Turmeric
		increase in expression of HO-1	
Epigallocatechin	-	Kinase phosphorylation upstream of	Green tea
gallate		Nrf2 increased HO-1 expression	
Allyl sulfides	Organosulfur compounds	Keap1 cysteine modification	Garlic
Resveratrol	Polyphenol: allyl sulfides	Kinase phosphorylation upstream of	Grapes
		Nrf2	Red wine
Lycopene	Phytochemical:	Increase Nrf2/HO-1 expression	Tomatoes
	tetraterpene carotenoid		Carrots
Caspaicin	Phytochemical	Inhibition of NQO1	Chilies
Fisetin	Flavonoid	Multiple e.g.	Strawberries
		increased glutothione expression	Apples
		neutralisation of reactive oxygen species	Onions
		disruption of the PI3K/AKT pathway	Cucumber

TILL ON NT PO	• . •	4 11	•	1
Table-3: Nrf2	agonists in	naturally	occurring	comnounds
1 abic-3. 14114	agomsts m	maturany	occurring	compounds

Quercitin	Flavonoid	Increased HO-1 and NQO1 expression	Red kidney
			bean
			Caper
			Radish
			onion
Cinnamaldehyde	Unsaturated aldehyde	Increased Nrf2 expression	Cinnamon
		Increased Nrf2 nuclear translocation	
		Suppressed NF-KB activation	

4.3.2. The gut microbiome and Nrf2 activation

CKD is characterized by an altered gut microbiome, resulting in an accumulation of uremic toxins such Pcresyl sulfate (PCS) and indoxyl sulfate (IS) [157, 164]. PCS and IS are uremic toxins that are directly related to accelerated progression of CKD, and both are derived from the colonic bacterial fermentation of dietary protein, [160]. Mafra *et al.* [165] have demonstrated that the dysbiosis in the gut microbiome in CKD leads to an increase in the bacteria that generate the uremic toxins IS, PCS, Indole-3-acetic acid, (IAA), and Trimethylamine (TMA), leading to inflammation. Uremic toxins affect gene expression via NF- κ B and Nrf2 regulated pathways [53]. Lau *et al.* [166] have demonstrated that uremic toxins from gut microbiota accentuate the Nrf2/NF- κ B imbalance and have proposed the use of probiotics, prebiotics and symbiotics to reduce toxin levels in CKD patients, and hence their risk of EVA.

Urolithin A, is a major microbial metabolite which displays anti-inflammatory, anti-oxidative, and anticellular ageing activities, that has been shown to upregulate gut barrier epithelial tight junction proteins via Nrf2 mediated activity [161]. Alteration of the composition of gut microbiota via diet/supplements could therefore be an effective non-pharmacological means of modulating the level of oxidative stress via Nrf2 activation. This has been demonstrated by the restriction of protein in the diet of CKD patients. A low protein diet reduce levels of oxidative stress and inflammation via the modulation of Nrf2 expression in non-dialysis CKD patients [156, 160, 167], and has been hypothesized to modulate the gut microbiota and reduce the generation of uremic toxins, such as PCS and IS [160]. The source of protein within the diet has also been shown to be significant, with plant protein intake being associated with lower production of uremic toxins and lower serum phosphorus and warrants further exploration, [168, 169].

4.3.3. Exercise

Exercise is an effective modulator of Nrf2. Exercise induces ROS, increasing the level of oxidative stress, which results in an increased dissociation of Nrf2 from Keap1 [170]. Several groups have demonstrated an

increase in Nrf2 expression in old rats following exercise [171, 172]. Subsequently Abreu *et al.* [173] have demonstrated Nrf2 induction and inhibition of NF-κB following resistance exercise in CKD patients undergoing haemodialysis. The Nrf2 response to exercise however varies according to training modality, duration and age [95]. Although acute exercise increased Nrf2 protein levels in peripheral blood mononuclear cells in young and older men, nuclear accumulation of Nrf2 was observed only in the young group. This indicates that ageing *per se* is accompanied by a reduced nuclear import of Nrf2 [174, 175]. Exercise has also been shown to induce epigenetic changes, predominantly altering the methylation pattern of the promoters of genes in the adaptive antioxidant response, [176]. Overall, exercise is a realistic intervention to improve endogenous antioxidant defenses via Nrf2 activation, though this must be viewed in the context of antagonistic pleiotropy to yield maximum benefit.

5. Conclusions

Premature vascular aging is a common problem for both general populations and patients with chronic inflammatory diseases like CKD or atherosclerosis. Two major factors that drive healthy cells towards early vascular aging are calcification and senescence, both arising from oxidative stress and persistent low-grade inflammation. These conditions are characterized by a failure to activate ARE-driven gene expression and a dysregulation of the Nrf2-ARE pathway. As reviewed here, a multitude of studies suggest that modulating Nrf2 activation is beneficial for targeting the burden of lifestyle diseases. Nrf2 activation might be induced by pharmacological treatments or non-pharmacological therapeutics such as diet and exercise. Targeting Keap1 pharmacologically, thereby controlling Nrf2 protein stabilization, is proposed as the most auspicious intervention. Several clinical regulators currently in clinical development or trialing for example DMF and Bardoxolone methyl, have shown promising results. Besides the direct interaction with Keap1, Nrf2 is regulated on multiple levels, including transcriptional, epigenetic, covalent protein modification and proteasome degradation. A few Keap1 independent targets have been described to modulate the Nrf2-ARE pathway, including the transcriptional repressor BACH, or TPH-1. Despite the promising results in preclinical research and clinical trials, safety concerns regarding overexpression have surfaced in recent years, regarding the role of Nrf2 in cancer development and potential side effects of Nrf2 activation in a complex tumor environment. These concerns require additional address. Nevertheless, Nrf2 constitutes a powerful target for intervention in premature ageing, whether as sole treatment or as adjuvant treatment in combination with selective therapies targeting of senescence or calcification.

Figure legends

Figure-1: Overview of the role of Nrf2 to regulate the cellular senescence and calcification. Balance of downstream genes of Nrf2 is crucial for the maintenance of cellular homeostasis. Under normal physiological conditions, the cell senses oxidative and inflammatory stress and releases Nrf2 which protects the cell from calcification and cellular senescence via the activation of protective signaling pathways including NQO1, HO-1, HO-2, TPH-1, GST and miRNAs. On the contrary, if the Nrf2 pathway is dysregulated imbalance of Nrf2 response genes occurs and the cell is no longer protected from pro-inflammatory and oxidative stress promoting the development of vascular pathologies. The Nrf2 pathway can be modulated in several steps of the signaling cascade (▲ cysteine modification in Keap1, • p62 activation, ▼ ARE activation, *GSK inhibition). Abbreviations: Nrf2, nuclear factor erythroid 2-related factor 2; Keap1, kelch-like ECH associated protein1; ROS, reactive oxygen species; H₂S, hydrogen sulfide; NQO1, NAD(P)H dehydrogenase (quinone) 1; HO-1, heme oxygenase-1; HO-2, heme oxygenase-2; TPH-1, tryptophan hydroxylase-1; GST, glutathione-S-transferase; GSK-3, glycogen synthase kinase 3; SASP, senescence associated secretory phenotype; Cul-3, cullin 3; Ub, ubiquitination; Ph-phosphorylation.

Table-1: Nrf2 impacts on various early vascular ageing associated diseases.

 Table-2: Nrf2 Modulators in clinical development.

Table-3: Nrf2 agonists in naturally occurring compounds.

Author Contributions: S.A, S.B., S.H, J.S. and S.A. searched the literature and drafted the review. K.K., P.G.S. and P.S. contributed important intellectual content and editing during manuscript drafting and revision.

Acknowledgments: This work was supported by grants from Swedish Research Council grant no. 2018–00932; Strategic Research Program in Diabetes at Karolinska Institutet, Sweden; funding from the European Union (INTRICARE and CaReSyAn); CIMED (centrum för innovativ medicin); Njurfonden, Sweden; Hjärt lungfonden, Sweden. The content of this article reflects only the views of its authors.

Conflicts of Interest: Peter Stenvinkel is in the scientific advisory board of REATA.

Other authors have no conflict of interest to declare.

References

[1] N. Sinha, P.K. Dabla, Oxidative stress and antioxidants in hypertension-a current review, Curr Hypertens Rev 11(2) (2015) 132-42.

[2] M.N. Sack, F.Y. Fyhrquist, O.J. Saijonmaa, V. Fuster, J.C. Kovacic, Basic Biology of Oxidative Stress and the Cardiovascular System: Part 1 of a 3-Part Series, J Am Coll Cardiol 70(2) (2017) 196-211.

[3] T. Cui, Y. Lai, J.S. Janicki, X. Wang, Nuclear factor erythroid-2 related factor 2 (Nrf2)mediated protein quality control in cardiomyocytes, Front Biosci (Landmark Ed) 21 (2016) 192-202.

[4] A. Alfieri, S. Srivastava, R.C. Siow, M. Modo, P.A. Fraser, G.E. Mann, Targeting the Nrf2-Keap1 antioxidant defence pathway for neurovascular protection in stroke, J Physiol 589(17) (2011) 4125-36.

[5] J.D. Hayes, A.T. Dinkova-Kostova, The Nrf2 regulatory network provides an interface between redox and intermediary metabolism, Trends Biochem Sci 39(4) (2014) 199-218.
[6] W. Li, A.N. Kong, Molecular mechanisms of Nrf2-mediated antioxidant response, Mol Carcinog 48(2) (2009) 91-104.

[7] M. Yu, M. Xu, Y. Liu, W. Yang, Y. Rong, P. Yao, H. Yan, D. Wang, L. Liu, Nrf2/ARE is the potential pathway to protect Sprague-Dawley rats against oxidative stress induced by quinocetone, Regul Toxicol Pharmacol 66(3) (2013) 279-85.

[8] K.I. Tong, Y. Katoh, H. Kusunoki, K. Itoh, T. Tanaka, M. Yamamoto, Keap1 recruits Neh2 through binding to ETGE and DLG motifs: characterization of the two-site molecular recognition model, Mol Cell Biol 26(8) (2006) 2887-900.

[9] A.T. Dinkova-Kostova, W.D. Holtzclaw, T.W. Kensler, The role of Keap1 in cellular protective responses, Chem Res Toxicol 18(12) (2005) 1779-91.

[10] M. Dodson, M.R. de la Vega, A.B. Cholanians, C.J. Schmidlin, E. Chapman, D.D. Zhang, Modulating NRF2 in Disease: Timing Is Everything, Annu Rev Pharmacol Toxicol 59 (2019) 555-575.

[11] S.M. Ahmed, L. Luo, A. Namani, X.J. Wang, X. Tang, Nrf2 signaling pathway: Pivotal roles in inflammation, Biochim Biophys Acta Mol Basis Dis 1863(2) (2017) 585-597.

[12] S.B. Cullinan, J.D. Gordan, J. Jin, J.W. Harper, J.A. Diehl, The Keap1-BTB protein is an adaptor that bridges Nrf2 to a Cul3-based E3 ligase: oxidative stress sensing by a Cul3-Keap1 ligase, Mol Cell Biol 24(19) (2004) 8477-86.

[13] A. Kobayashi, M.I. Kang, H. Okawa, M. Ohtsuji, Y. Zenke, T. Chiba, K. Igarashi, M. Yamamoto, Oxidative stress sensor Keap1 functions as an adaptor for Cul3-based E3 ligase to regulate proteasomal degradation of Nrf2, Mol Cell Biol 24(16) (2004) 7130-9.

[14] D.D. Zhang, M. Hannink, Distinct cysteine residues in Keap1 are required for Keap1dependent ubiquitination of Nrf2 and for stabilization of Nrf2 by chemopreventive agents and oxidative stress, Mol Cell Biol 23(22) (2003) 8137-51.

[15] J.D. Hayes, M. McMahon, S. Chowdhry, A.T. Dinkova-Kostova, Cancer chemoprevention mechanisms mediated through the Keap1-Nrf2 pathway, Antioxid Redox Signal 13(11) (2010) 1713-48.

[16] S. Chowdhry, Y. Zhang, M. McMahon, C. Sutherland, A. Cuadrado, J.D. Hayes, Nrf2 is controlled by two distinct β -TrCP recognition motifs in its Neh6 domain, one of which can be modulated by GSK-3 activity, Oncogene 32(32) (2013) 3765-81.

[17] M. Komatsu, H. Kurokawa, S. Waguri, K. Taguchi, A. Kobayashi, Y. Ichimura, Y.S. Sou, I. Ueno, A. Sakamoto, K.I. Tong, M. Kim, Y. Nishito, S. Iemura, T. Natsume, T. Ueno, E. Kominami, H. Motohashi, K. Tanaka, M. Yamamoto, The selective autophagy substrate p62 activates the stress responsive transcription factor Nrf2 through inactivation of Keap1, Nat Cell Biol 12(3) (2010) 213-23.

[18] P. Rada, A.I. Rojo, S. Chowdhry, M. McMahon, J.D. Hayes, A. Cuadrado, SCF/{beta}-TrCP promotes glycogen synthase kinase 3-dependent degradation of the Nrf2 transcription factor in a Keap1-independent manner, Mol Cell Biol 31(6) (2011) 1121-33.

[19] V. Granatiero, C. Konrad, K. Bredvik, G. Manfredi, H. Kawamata, Nrf2 signaling links ER oxidative protein folding and calcium homeostasis in health and disease, Life Sci Alliance 2(5) (2019).

[20] C. López-Otín, M.A. Blasco, L. Partridge, M. Serrano, G. Kroemer, The hallmarks of aging, Cell 153(6) (2013) 1194-217.

[21] B.M. Hybertson, B. Gao, S.K. Bose, J.M. McCord, Oxidative stress in health and disease: the therapeutic potential of Nrf2 activation, Mol Aspects Med 32(4-6) (2011) 234-46.

[22] M.M. Bachschmid, S. Schildknecht, R. Matsui, R. Zee, D. Haeussler, R.A. Cohen, D. Pimental, B. Loo, Vascular aging: chronic oxidative stress and impairment of redox signaling-consequences for vascular homeostasis and disease, Ann Med 45(1) (2013) 17-36.

[23] C.M. Shanahan, Mechanisms of vascular calcification in CKD-evidence for premature ageing?, Nat Rev Nephrol 9(11) (2013) 661-70.

[24] D. McHugh, J. Gil, Senescence and aging: Causes, consequences, and therapeutic avenues, J Cell Biol 217(1) (2018) 65-77.

[25] O. Al-Sawaf, T. Clarner, A. Fragoulis, Y.W. Kan, T. Pufe, K. Streetz, C.J. Wruck, Nrf2 in health and disease: current and future clinical implications, Clin Sci (Lond) 129(12) (2015) 989-99.

[26] B.M. Hybertson, B. Gao, Role of the Nrf2 signaling system in health and disease, Clin Genet 86(5) (2014) 447-52.

[27] E.H. Kobayashi, T. Suzuki, R. Funayama, T. Nagashima, M. Hayashi, H. Sekine, N. Tanaka, T. Moriguchi, H. Motohashi, K. Nakayama, M. Yamamoto, Nrf2 suppresses macrophage inflammatory response by blocking proinflammatory cytokine transcription, Nat Commun 7 (2016) 11624.

[28] J.W. Calvert, S. Jha, S. Gundewar, J.W. Elrod, A. Ramachandran, C.B. Pattillo, C.G. Kevil, D.J. Lefer, Hydrogen sulfide mediates cardioprotection through Nrf2 signaling, Circ Res 105(4) (2009) 365-74.

[29] J.E. Teasdale, G.G. Hazell, A.M. Peachey, G.B. Sala-Newby, C.C. Hindmarch, T.R. McKay, M. Bond, A.C. Newby, S.J. White, Cigarette smoke extract profoundly suppresses TNF α -mediated proinflammatory gene expression through upregulation of ATF3 in human coronary artery endothelial cells, Sci Rep 7 (2017) 39945.

[30] L.H. Chen, Q. Huang, L. Wan, L.Y. Zeng, S.F. Li, Y.P. Li, X.F. Lu, J.Q. Cheng, Expression, purification, and in vitro refolding of a humanized single-chain Fv antibody against human CTLA4 (CD152), Protein Expr Purif 46(2) (2006) 495-502.

[31] W. Takabe, E. Warabi, N. Noguchi, Anti-atherogenic effect of laminar shear stress via Nrf2 activation, Antioxid Redox Signal 15(5) (2011) 1415-26.

[32] S. Noel, M.N. Martina, S. Bandapalle, L.C. Racusen, H.R. Potteti, A.R. Hamad, S.P. Reddy, H. Rabb, T Lymphocyte-Specific Activation of Nrf2 Protects from AKI, J Am Soc Nephrol 26(12) (2015) 2989-3000.

[33] H. Saito, Toxico-pharmacological perspective of the Nrf2-Keap1 defense system against oxidative stress in kidney diseases, Biochem Pharmacol 85(7) (2013) 865-72.

[34] S. Thangapandiyan, M. Ramesh, S. Miltonprabu, T. Hema, G.B. Jothi, V. Nandhini, Sulforaphane potentially attenuates arsenic-induced nephrotoxicity via the PI3K/Akt/Nrf2 pathway in albino Wistar rats, Environ Sci Pollut Res Int 26(12) (2019) 12247-12263. [35] D.Q. Chen, G. Cao, H. Chen, C.P. Argyopoulos, H. Yu, W. Su, L. Chen, D.C. Samuels, S. Zhuang, G.P. Bayliss, S. Zhao, X.Y. Yu, N.D. Vaziri, M. Wang, D. Liu, J.R. Mao, S.X. Ma, J. Zhao, Y. Zhang, Y.Q. Shang, H. Kang, F. Ye, X.H. Cheng, X.R. Li, L. Zhang, M.X. Meng, Y. Guo, Y.Y. Zhao, Identification of serum metabolites associating with chronic kidney disease progression and anti-fibrotic effect of 5-methoxytryptophan, Nat Commun 10(1) (2019) 1476.
[36] H. Zheng, S.A. Whitman, W. Wu, G.T. Wondrak, P.K. Wong, D. Fang, D.D. Zhang, Therapeutic potential of Nrf2 activators in streptozotocin-induced diabetic nephropathy, Diabetes 60(11) (2011) 3055-66.

[37] L. Villacorta, J. Zhang, M.T. Garcia-Barrio, X.L. Chen, B.A. Freeman, Y.E. Chen, T. Cui, Nitro-linoleic acid inhibits vascular smooth muscle cell proliferation via the Keap1/Nrf2 signaling pathway, Am J Physiol Heart Circ Physiol 293(1) (2007) H770-6.

[38] M. Kobayashi, L. Li, N. Iwamoto, Y. Nakajima-Takagi, H. Kaneko, Y. Nakayama, M. Eguchi, Y. Wada, Y. Kumagai, M. Yamamoto, The antioxidant defense system Keap1-Nrf2 comprises a multiple sensing mechanism for responding to a wide range of chemical compounds, Mol Cell Biol 29(2) (2009) 493-502.

[39] Y. Zhang, M. Sano, K. Shinmura, K. Tamaki, Y. Katsumata, T. Matsuhashi, S. Morizane, H. Ito, T. Hishiki, J. Endo, H. Zhou, S. Yuasa, R. Kaneda, M. Suematsu, K. Fukuda, 4-hydroxy-2-nonenal protects against cardiac ischemia-reperfusion injury via the Nrf2-dependent pathway, J Mol Cell Cardiol 49(4) (2010) 576-86.

[40] Y. Huang, W. Li, A.N. Kong, Anti-oxidative stress regulator NF-E2-related factor 2 mediates the adaptive induction of antioxidant and detoxifying enzymes by lipid peroxidation metabolite 4-hydroxynonenal, Cell Biosci 2(1) (2012) 40.

[41] S.M. Yang, S.M. Ka, H.L. Wu, Y.C. Yeh, C.H. Kuo, K.F. Hua, G.Y. Shi, Y.J. Hung, F.C. Hsiao, S.S. Yang, Y.S. Shieh, S.H. Lin, C.W. Wei, J.S. Lee, C.Y. Yang, A. Chen, Thrombomodulin domain 1 ameliorates diabetic nephropathy in mice via anti-NF-κB/NLRP3 inflammasome-mediated inflammation, enhancement of NRF2 antioxidant activity and inhibition of apoptosis, Diabetologia 57(2) (2014) 424-34.

[42] S.M. Ka, Y.C. Yeh, X.R. Huang, T.K. Chao, Y.J. Hung, C.P. Yu, T.J. Lin, C.C. Wu, H.Y. Lan, A. Chen, Kidney-targeting Smad7 gene transfer inhibits renal TGF- β /MAD homologue (SMAD) and nuclear factor κB (NF-κB) signalling pathways, and improves diabetic nephropathy in mice, Diabetologia 55(2) (2012) 509-19.

[43] T.L. Adair-Kirk, J.J. Atkinson, G.L. Griffin, M.A. Watson, D.G. Kelley, D. DeMello, R.M. Senior, T. Betsuyaku, Distal airways in mice exposed to cigarette smoke: Nrf2-regulated genes are increased in Clara cells, Am J Respir Cell Mol Biol 39(4) (2008) 400-11.

[44] T. Iizuka, Y. Ishii, K. Itoh, T. Kiwamoto, T. Kimura, Y. Matsuno, Y. Morishima, A.E. Hegab, S. Homma, A. Nomura, T. Sakamoto, M. Shimura, A. Yoshida, M. Yamamoto, K. Sekizawa, Nrf2-deficient mice are highly susceptible to cigarette smoke-induced emphysema, Genes Cells 10(12) (2005) 1113-25.

[45] N.F. Voelkel, H.J. Bogaard, A. Al Husseini, L. Farkas, J. Gomez-Arroyo, R. Natarajan, Antioxidants for the treatment of patients with severe angioproliferative pulmonary hypertension?, Antioxid Redox Signal 18(14) (2013) 1810-7.

[46] Q. Ma, Role of nrf2 in oxidative stress and toxicity, Annu Rev Pharmacol Toxicol 53 (2013) 401-26.

[47] P. Stenvinkel, C.J. Meyer, G.A. Block, G.M. Chertow, P.G. Shiels, Understanding the role of the cytoprotective transcription factor nuclear factor erythroid 2-related factor 2-lessons from evolution, the animal kingdom and rare progeroid syndromes, Nephrol Dial Transplant (2019).

[48] M.N. Valcarcel-Ares, T. Gautam, J.P. Warrington, L. Bailey-Downs, D. Sosnowska, R. de Cabo, G. Losonczy, W.E. Sonntag, Z. Ungvari, A. Csiszar, Disruption of Nrf2 signaling impairs angiogenic capacity of endothelial cells: implications for microvascular aging, J Gerontol A Biol Sci Med Sci 67(8) (2012) 821-9.

[49] C.J. Schmidlin, M.B. Dodson, L. Madhavan, D.D. Zhang, Redox regulation by NRF2 in aging and disease, Free Radic Biol Med 134 (2019) 702-707.

[50] H. Zhang, H. Liu, K.J. Davies, C. Sioutas, C.E. Finch, T.E. Morgan, H.J. Forman, Nrf2regulated phase II enzymes are induced by chronic ambient nanoparticle exposure in young mice with age-related impairments, Free Radic Biol Med 52(9) (2012) 2038-46.

[51] C.E. Hochmuth, B. Biteau, D. Bohmann, H. Jasper, Redox regulation by Keap1 and Nrf2 controls intestinal stem cell proliferation in Drosophila, Cell Stem Cell 8(2) (2011) 188-99.

[52] J. Chang, J.Z. Ma, Q. Zeng, S. Cechova, A. Gantz, C. Nievergelt, D. O'Connor, M. Lipkowitz, T.H. Le, Loss of GSTM1, a NRF2 target, is associated with accelerated progression of hypertensive kidney disease in the African American Study of Kidney Disease (AASK), Am J Physiol Renal Physiol 304(4) (2013) F348-55.

[53] D. Bolati, H. Shimizu, M. Yisireyili, F. Nishijima, T. Niwa, Indoxyl sulfate, a uremic toxin, downregulates renal expression of Nrf2 through activation of NF-κB, BMC Nephrol 14 (2013) 56.

[54] C.J. Oh, J.Y. Kim, Y.K. Choi, H.J. Kim, J.Y. Jeong, K.H. Bae, K.G. Park, I.K. Lee, Dimethylfumarate attenuates renal fibrosis via NF-E2-related factor 2-mediated inhibition of transforming growth factor- β /Smad signaling, PLoS One 7(10) (2012) e45870.

[55] T. Suzuki, S. Seki, K. Hiramoto, E. Naganuma, E.H. Kobayashi, A. Yamaoka, L. Baird, N. Takahashi, H. Sato, M. Yamamoto, Hyperactivation of Nrf2 in early tubular development induces nephrogenic diabetes insipidus, Nat Commun 8 (2017) 14577.

[56] J. Mimura, K. Itoh, Role of Nrf2 in the pathogenesis of atherosclerosis, Free Radic Biol Med 88(Pt B) (2015) 221-232.

[57] L. Gao, M.C. Zimmerman, S. Biswal, I.H. Zucker, Selective N*rf2* Gene Deletion in the Rostral Ventrolateral Medulla Evokes Hypertension and Sympathoexcitation in Mice, Hypertension 69(6) (2017) 1198-1206.

[58] K.L.H. Wu, C.W. Wu, Y.M. Chao, C.Y. Hung, J.Y.H. Chan, Impaired Nrf2 regulation of mitochondrial biogenesis in rostral ventrolateral medulla on hypertension induced by systemic inflammation, Free Radic Biol Med 97 (2016) 58-74.

[59] Y. Tan, T. Ichikawa, J. Li, Q. Si, H. Yang, X. Chen, C.S. Goldblatt, C.J. Meyer, X. Li, L. Cai, T. Cui, Diabetic downregulation of Nrf2 activity via ERK contributes to oxidative stressinduced insulin resistance in cardiac cells in vitro and in vivo, Diabetes 60(2) (2011) 625-33.

[60] P.V. Dludla, C.J. Muller, E. Joubert, J. Louw, M.F. Essop, K.B. Gabuza, S. Ghoor, B. Huisamen, R. Johnson, Aspalathin Protects the Heart against Hyperglycemia-Induced Oxidative Damage by Up-Regulating Nrf2 Expression, Molecules 22(1) (2017).

[61] M. Rojo de la Vega, E. Chapman, D.D. Zhang, NRF2 and the Hallmarks of Cancer, Cancer Cell 34(1) (2018) 21-43.

[62] H. Kitamura, H. Motohashi, NRF2 addiction in cancer cells, Cancer Sci 109(4) (2018) 900-911.

[63] K. Trpkov, O. Hes, A. Agaimy, M. Bonert, P. Martinek, C. Magi-Galluzzi, G. Kristiansen, C. Lüders, G. Nesi, E. Compérat, M. Sibony, D.M. Berney, R. Mehra, F. Brimo, A. Hartmann, A. Husain, N. Frizzell, K. Hills, F. Maclean, B. Srinivasan, A.J. Gill, Fumarate Hydratasedeficient Renal Cell Carcinoma Is Strongly Correlated With Fumarate Hydratase Mutation and Hereditary Leiomyomatosis and Renal Cell Carcinoma Syndrome, Am J Surg Pathol 40(7) (2016) 865-75.

[64] A. Ooi, J.C. Wong, D. Petillo, D. Roossien, V. Perrier-Trudova, D. Whitten, B.W. Min, M.H. Tan, Z. Zhang, X.J. Yang, M. Zhou, B. Gardie, V. Molinié, S. Richard, P.H. Tan, B.T. Teh, K.A. Furge, An antioxidant response phenotype shared between hereditary and sporadic type 2 papillary renal cell carcinoma, Cancer Cell 20(4) (2011) 511-23.

[65] P.G. Cunha, P. Boutouyrie, P.M. Nilsson, S. Laurent, Early Vascular Ageing (EVA): Definitions and Clinical Applicability, Curr Hypertens Rev 13(1) (2017) 8-15.

[66] P. Stenvinkel, T.E. Larsson, Chronic kidney disease: a clinical model of premature aging, Am J Kidney Dis 62(2) (2013) 339-51.

[67] D. HARMAN, Aging: a theory based on free radical and radiation chemistry, J Gerontol 11(3) (1956) 298-300.

[68] V.I. Pérez, H. Van Remmen, A. Bokov, C.J. Epstein, J. Vijg, A. Richardson, The overexpression of major antioxidant enzymes does not extend the lifespan of mice, Aging Cell 8(1) (2009) 73-5.

[69] A. Csiszar, Z. Ungvari, J.G. Edwards, P. Kaminski, M.S. Wolin, A. Koller, G. Kaley, Aging-induced phenotypic changes and oxidative stress impair coronary arteriolar function, Circ Res 90(11) (2002) 1159-66.

[70] A.J. Donato, I. Eskurza, A.E. Silver, A.S. Levy, G.L. Pierce, P.E. Gates, D.R. Seals, Direct evidence of endothelial oxidative stress with aging in humans: relation to impaired endothelium-dependent dilation and upregulation of nuclear factor-kappaB, Circ Res 100(11) (2007) 1659-66.

[71] P.E. Gates, M.L. Boucher, A.E. Silver, K.D. Monahan, D.R. Seals, Impaired flow-mediated dilation with age is not explained by L-arginine bioavailability or endothelial asymmetric dimethylarginine protein expression, J Appl Physiol (1985) 102(1) (2007) 63-71.

[72] J.H. Suh, S.V. Shenvi, B.M. Dixon, H. Liu, A.K. Jaiswal, R.M. Liu, T.M. Hagen, Decline in transcriptional activity of Nrf2 causes age-related loss of glutathione synthesis, which is reversible with lipoic acid, Proc Natl Acad Sci U S A 101(10) (2004) 3381-6.

[73] Z. Ungvari, G. Kaley, R. de Cabo, W.E. Sonntag, A. Csiszar, Mechanisms of vascular aging: new perspectives, J Gerontol A Biol Sci Med Sci 65(10) (2010) 1028-41.

[74] C. Franceschi, J. Campisi, Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases, J Gerontol A Biol Sci Med Sci 69 Suppl 1 (2014) S4-9.
[75] J.P. Kooman, M.J. Dekker, L.A. Usvyat, P. Kotanko, F.M. van der Sande, C.G. Schalkwijk, P.G. Shiels, P. Stenvinkel, Inflammation and premature aging in advanced chronic kidney disease, Am J Physiol Renal Physiol 313(4) (2017) F938-F950.

[76] C. Liu, E.K. Gidlund, A. Witasp, A.R. Qureshi, M. Söderberg, A. Thorell, G.A. Nader, P. Barany, P. Stenvinkel, F. von Walden, Reduced skeletal muscle expression of mitochondrialderived peptides humanin and MOTS-C and Nrf2 in chronic kidney disease, Am J Physiol Renal Physiol 317(5) (2019) F1122-F1131.

[77] J.A. Leopold, Vascular calcification: Mechanisms of vascular smooth muscle cell calcification, Trends Cardiovasc Med 25(4) (2015) 267-74.

[78] P. Aghagolzadeh, R. Radpour, M. Bachtler, H. van Goor, E.R. Smith, A. Lister, A.
Odermatt, M. Feelisch, A. Pasch, Hydrogen sulfide attenuates calcification of vascular smooth muscle cells via KEAP1/NRF2/NQO1 activation, Atherosclerosis 265 (2017) 78-86.
[79] F. Ganster, M. Burban, M. de la Bourdonnaye, L. Fizanne, O. Douay, L. Loufrani, A.

Mercat, P. Calès, P. Radermacher, D. Henrion, P. Asfar, F. Meziani, Effects of hydrogen sulfide

on hemodynamics, inflammatory response and oxidative stress during resuscitated hemorrhagic shock in rats, Crit Care 14(5) (2010) R165.

[80] E. Hinoi, S. Fujimori, L. Wang, H. Hojo, K. Uno, Y. Yoneda, Nrf2 negatively regulates osteoblast differentiation via interfering with Runx2-dependent transcriptional activation, J Biol Chem 281(26) (2006) 18015-24.

[81] P. Zhang, Y. Li, Y. Du, G. Li, L. Wang, F. Zhou, Resveratrol Ameliorated Vascular Calcification by Regulating Sirt-1 and Nrf2, Transplant Proc 48(10) (2016) 3378-3386.
[82] K.D. Yoo, S. Kang, Y. Choi, S.H. Yang, N.J. Heo, H.J. Chin, K.H. Oh, K.W. Joo, Y.S. Kim, H. Lee, Sex, Age, and the Association of Serum Phosphorus With All-Cause Mortality in Adults With Normal Kidney Function, Am J Kidney Dis 67(1) (2016) 79-88.

[83] R.N. Foley, A.J. Collins, C.A. Herzog, A. Ishani, P.A. Kalra, Serum phosphorus levels associate with coronary atherosclerosis in young adults, J Am Soc Nephrol 20(2) (2009) 397-404.

[84] C. Ginsberg, A.J.H.M. Houben, R. Malhotra, T.T.J.M. Berendschot, P.C. Dagnelie, J.P. Kooman, C.A. Webers, C.D.A. Stehouwer, J.H. Ix, Serum Phosphate and Microvascular Function in a Population-Based Cohort, Clin J Am Soc Nephrol 14(11) (2019) 1626-1633.
[85] R. Wei, M. Enaka, Y. Muragaki, Activation of KEAP1/NRF2/P62 signaling alleviates high phosphate-induced calcification of vascular smooth muscle cells by suppressing reactive oxygen species production, Sci Rep 9(1) (2019) 10366.

[86] C.M. Ha, S. Park, Y.K. Choi, J.Y. Jeong, C.J. Oh, K.H. Bae, S.J. Lee, J.H. Kim, K.G. Park, d.Y. Jun, I.K. Lee, Activation of Nrf2 by dimethyl fumarate improves vascular calcification, Vascul Pharmacol 63(1) (2014) 29-36.

[87] L. Yao, J. Wang, B.Y. Tian, T.H. Xu, Z.T. Sheng, Activation of the Nrf2-ARE Signaling Pathway Prevents Hyperphosphatemia-Induced Vascular Calcification by Inducing Autophagy in Renal Vascular Smooth Muscle Cells, J Cell Biochem 118(12) (2017) 4708-4715.

[88] W. Zhang, Y. Li, H. Ding, Y. Du, L. Wang, Hydrogen peroxide prevents vascular calcification induced ROS production by regulating Nrf-2 pathway, Ren Fail 38(7) (2016) 1099-106.

[89] J.L. Kirkland, T. Tchkonia, Cellular Senescence: A Translational Perspective, EBioMedicine 21 (2017) 21-28.

[90] I. Sturmlechner, M. Durik, C.J. Sieben, D.J. Baker, J.M. van Deursen, Cellular senescence in renal ageing and disease, Nat Rev Nephrol 13(2) (2017) 77-89.

[91] J.P. Coppé, P.Y. Desprez, A. Krtolica, J. Campisi, The senescence-associated secretory phenotype: the dark side of tumor suppression, Annu Rev Pathol 5 (2010) 99-118.

[92] P. Stenvinkel, K. Luttropp, D. McGuinness, A. Witasp, A.R. Qureshi, A. Wernerson, L. Nordfors, M. Schalling, J. Ripsweden, L. Wennberg, M. Söderberg, P. Bárány, H. Olauson, P.G. Shiels, CDKN2A/p16INK4(a) expression is associated with vascular progeria in chronic kidney disease, Aging (Albany NY) 9(2) (2017) 494-507.

[93] L. Zhou, H. Zhang, K.J.A. Davies, H.J. Forman, Aging-related decline in the induction of Nrf2-regulated antioxidant genes in human bronchial epithelial cells, Redox Biol 14 (2018) 35-40.

[94] Z. Ungvari, L. Bailey-Downs, D. Sosnowska, T. Gautam, P. Koncz, G. Losonczy, P. Ballabh, R. de Cabo, W.E. Sonntag, A. Csiszar, Vascular oxidative stress in aging: a homeostatic failure due to dysregulation of NRF2-mediated antioxidant response, Am J Physiol Heart Circ Physiol 301(2) (2011) H363-72.

[95] S.S. Gounder, S. Kannan, D. Devadoss, C.J. Miller, K.J. Whitehead, K.S. Whitehead, S.J. Odelberg, M.A. Firpo, R. Paine, J.R. Hoidal, E.D. Abel, N.S. Rajasekaran, Impaired transcriptional activity of Nrf2 in age-related myocardial oxidative stress is reversible by moderate exercise training, PLoS One 7(9) (2012) e45697.

[96] S.M. Kuosmanen, V. Sihvola, E. Kansanen, M.U. Kaikkonen, A.L. Levonen, MicroRNAs mediate the senescence-associated decline of NRF2 in endothelial cells, Redox Biol 18 (2018) 77-83.

[97] G.A. Fulop, T. Kiss, S. Tarantini, P. Balasubramanian, A. Yabluchanskiy, E. Farkas, F. Bari, Z. Ungvari, A. Csiszar, Nrf2 deficiency in aged mice exacerbates cellular senescence promoting cerebrovascular inflammation, Geroscience 40(5-6) (2018) 513-521.

[98] T. Zhou, M. Zhang, L. Zhao, A. Li, X. Qin, Activation of Nrf2 contributes to the protective effect of Exendin-4 against angiotensin II-induced vascular smooth muscle cell senescence, Am J Physiol Cell Physiol 311(4) (2016) C572-C582.

[99] S. Hobson, S. Arefin, K. Kublickiene, P.G. Shiels, P. Stenvinkel, Senescent Cells in Early Vascular Ageing and Bone Disease of Chronic Kidney Disease-A Novel Target for Treatment, Toxins (Basel) 11(2) (2019).

[100] D. Kloska, A. Kopacz, A. Piechota-Polanczyk, W.N. Nowak, J. Dulak, A. Jozkowicz, A. Grochot-Przeczek, Nrf2 in aging - Focus on the cardiovascular system, Vascul Pharmacol 112 (2019) 42-53.

[101] P. Hiebert, M.S. Wietecha, M. Cangkrama, E. Haertel, E. Mavrogonatou, M. Stumpe, H. Steenbock, S. Grossi, H.D. Beer, P. Angel, J. Brinckmann, D. Kletsas, J. Dengjel, S. Werner, Nrf2-Mediated Fibroblast Reprogramming Drives Cellular Senescence by Targeting the Matrisome, Dev Cell 46(2) (2018) 145-161.e10.

[102] P. Stenvinkel, J. Painer, M. Kuro-O, M. Lanaspa, W. Arnold, T. Ruf, P.G. Shiels, R.J. Johnson, Novel treatment strategies for chronic kidney disease: insights from the animal kingdom, Nat Rev Nephrol 14(4) (2018) 265-284.

[103] S. Ruiz, P.E. Pergola, R.A. Zager, N.D. Vaziri, Targeting the transcription factor Nrf2 to ameliorate oxidative stress and inflammation in chronic kidney disease, Kidney Int 83(6) (2013) 1029-41.

[104] D. Li, G. Shi, J. Wang, D. Zhang, Y. Pan, H. Dou, Y. Hou, Baicalein ameliorates pristaneinduced lupus nephritis via activating Nrf2/HO-1 in myeloid-derived suppressor cells, Arthritis Res Ther 21(1) (2019) 105.

[105] F.C. Barreto, D.V. Barreto, Z.A. Massy, T.B. Drücke, Strategies for Phosphate Control in Patients With CKD, Kidney Int Rep 4(8) (2019) 1043-1056.

[106] C. Viegas, N. Araújo, C. Marreiros, D. Simes, The interplay between mineral metabolism, vascular calcification and inflammation in Chronic Kidney Disease (CKD): challenging old concepts with new facts, Aging (Albany NY) 11(12) (2019) 4274-4299.

[107] Y.L. Yan, B. Qiu, J. Wang, S.B. Deng, L. Wu, X.D. Jing, J.L. Du, Y.J. Liu, Q. She, Highintensity statin therapy in patients with chronic kidney disease: a systematic review and metaanalysis, BMJ Open 5(5) (2015) e006886.

[108] T.M. Huang, V.C. Wu, Y.F. Lin, J.J. Wang, C.C. Shiao, L. Chen, S.J. Chueh, E. Chueh, S.Y. Yang, T.S. Lai, S.L. Lin, T.S. Chu, K.D. Wu, N.T.U.H.S.G.o.A.R.F. (NSARF), Effects of Statin Use in Advanced Chronic Kidney Disease Patients, J Clin Med 7(9) (2018).
[109] M.R. Hager, A.D. Narla, L.R. Tannock, Dyslipidemia in patients with chronic kidney disease, Rev Endocr Metab Disord 18(1) (2017) 29-40.

[110] I. Capelli, G. Cianciolo, L. Gasperoni, A. Galassi, P. Ciceri, M. Cozzolino, Nutritional vitamin D in CKD: Should we measure? Should we treat?, Clin Chim Acta 501 (2020) 186-197.
[111] G.J. Elder, J. Center, The role of calcium and non calcium-based phosphate binders in chronic kidney disease, Nephrology (Carlton) 22 Suppl 2 (2017) 42-46.

[112] M. Xu, T. Pirtskhalava, J.N. Farr, B.M. Weigand, A.K. Palmer, M.M. Weivoda, C.L. Inman, M.B. Ogrodnik, C.M. Hachfeld, D.G. Fraser, J.L. Onken, K.O. Johnson, G.C. Verzosa, L.G.P. Langhi, M. Weigl, N. Giorgadze, N.K. LeBrasseur, J.D. Miller, D. Jurk, R.J. Singh, D.B. Allison, K. Ejima, G.B. Hubbard, Y. Ikeno, H. Cubro, V.D. Garovic, X. Hou, S.J. Weroha, P.D. Robbins, L.J. Niedernhofer, S. Khosla, T. Tchkonia, J.L. Kirkland, Senolytics improve physical function and increase lifespan in old age, Nat Med 24(8) (2018) 1246-1256.

[113] C.M. Roos, B. Zhang, A.K. Palmer, M.B. Ogrodnik, T. Pirtskhalava, N.M. Thalji, M. Hagler, D. Jurk, L.A. Smith, G. Casaclang-Verzosa, Y. Zhu, M.J. Schafer, T. Tchkonia, J.L. Kirkland, J.D. Miller, Chronic senolytic treatment alleviates established vasomotor dysfunction in aged or atherosclerotic mice, Aging Cell 15(5) (2016) 973-7.

[114] L.J. Hickson, L.G.P. Langhi Prata, S.A. Bobart, T.K. Evans, N. Giorgadze, S.K. Hashmi, S.M. Herrmann, M.D. Jensen, Q. Jia, K.L. Jordan, T.A. Kellogg, S. Khosla, D.M. Koerber, A.B. Lagnado, D.K. Lawson, N.K. LeBrasseur, L.O. Lerman, K.M. McDonald, T.J. McKenzie, J.F. Passos, R.J. Pignolo, T. Pirtskhalava, I.M. Saadiq, K.K. Schaefer, S.C. Textor, S.G. Victorelli, T.L. Volkman, A. Xue, M.A. Wentworth, E.O. Wissler Gerdes, Y. Zhu, T. Tchkonia, J.L. Kirkland, Senolytics decrease senescent cells in humans: Preliminary report from a clinical trial of Dasatinib plus Quercetin in individuals with diabetic kidney disease, EBioMedicine 47 (2019) 446-456.

[115] B. Hegner, T. Schaub, D. Janke, D. Zickler, C. Lange, M. Girndt, J. Jankowski, R. Schindler, D. Dragun, Targeting proinflammatory cytokines ameliorates calcifying phenotype conversion of vascular progenitors under uremic conditions in vitro, Sci Rep 8(1) (2018) 12087.
[116] O.H. Jeon, C. Kim, R.M. Laberge, M. Demaria, S. Rathod, A.P. Vasserot, J.W. Chung, D.H. Kim, Y. Poon, N. David, D.J. Baker, J.M. van Deursen, J. Campisi, J.H. Elisseeff, Local clearance of senescent cells attenuates the development of post-traumatic osteoarthritis and creates a pro-regenerative environment, Nat Med 23(6) (2017) 775-781.

[117] H. Zhang, W. Zheng, X. Feng, F. Yang, H. Qin, S. Wu, D.X. Hou, J. Chen, Nrf2⁻ARE Signaling Acts as Master Pathway for the Cellular Antioxidant Activity of Fisetin, Molecules 24(4) (2019).

[118] Y. Chien, C. Scuoppo, X. Wang, X. Fang, B. Balgley, J.E. Bolden, P. Premsrirut, W. Luo, A. Chicas, C.S. Lee, S.C. Kogan, S.W. Lowe, Control of the senescence-associated secretory phenotype by NF-κB promotes senescence and enhances chemosensitivity, Genes Dev 25(20) (2011) 2125-36.

[119] A. Freund, C.K. Patil, J. Campisi, p38MAPK is a novel DNA damage responseindependent regulator of the senescence-associated secretory phenotype, EMBO J 30(8) (2011) 1536-48.

[120] N. Herranz, S. Gallage, M. Mellone, T. Wuestefeld, S. Klotz, C.J. Hanley, S. Raguz, J.C. Acosta, A.J. Innes, A. Banito, A. Georgilis, A. Montoya, K. Wolter, G. Dharmalingam, P. Faull, T. Carroll, J.P. Martínez-Barbera, P. Cutillas, F. Reisinger, M. Heikenwalder, R.A. Miller, D.

Withers, L. Zender, G.J. Thomas, J. Gil, mTOR regulates MAPKAPK2 translation to control the senescence-associated secretory phenotype, Nat Cell Biol 17(9) (2015) 1205-17.

[121] R.M. Laberge, Y. Sun, A.V. Orjalo, C.K. Patil, A. Freund, L. Zhou, S.C. Curran, A.R. Davalos, K.A. Wilson-Edell, S. Liu, C. Limbad, M. Demaria, P. Li, G.B. Hubbard, Y. Ikeno, M.

Javors, P.Y. Desprez, C.C. Benz, P. Kapahi, P.S. Nelson, J. Campisi, MTOR regulates the protumorigenic senescence-associated secretory phenotype by promoting IL1A translation, Nat Cell Biol 17(8) (2015) 1049-61.

[122] D.S. Hong, R. Kurzrock, J.G. Supko, X. He, A. Naing, J. Wheler, D. Lawrence, J.P. Eder, C.J. Meyer, D.A. Ferguson, J. Mier, M. Konopleva, S. Konoplev, M. Andreeff, D. Kufe, H. Lazarus, G.I. Shapiro, B.J. Dezube, A phase I first-in-human trial of bardoxolone methyl in patients with advanced solid tumors and lymphomas, Clin Cancer Res 18(12) (2012) 3396-406. [123] S.M. Tan, A. Sharma, N. Stefanovic, D.Y. Yuen, T.C. Karagiannis, C. Meyer, K.W. Ward, M.E. Cooper, J.B. de Haan, Derivative of bardoxolone methyl, dh404, in an inverse dosedependent manner lessens diabetes-associated atherosclerosis and improves diabetic kidney disease, Diabetes 63(9) (2014) 3091-103.

[124] P.E. Pergola, M. Krauth, J.W. Huff, D.A. Ferguson, S. Ruiz, C.J. Meyer, D.G. Warnock, Effect of bardoxolone methyl on kidney function in patients with T2D and Stage 3b-4 CKD, Am J Nephrol 33(5) (2011) 469-76.

[125] Y.J. Chen, L. Kong, Z.Z. Tang, Y.M. Zhang, Y. Liu, T.Y. Wang, Y.W. Liu, Hesperetin ameliorates diabetic nephropathy in rats by activating Nrf2/ARE/glyoxalase 1 pathway, Biomed Pharmacother 111 (2019) 1166-1175.

[126] M.C. Lu, J.A. Ji, Z.Y. Jiang, Q.D. You, The Keap1-Nrf2-ARE Pathway As a Potential Preventive and Therapeutic Target: An Update, Med Res Rev 36(5) (2016) 924-63.

[127] A. Cuadrado, A.I. Rojo, G. Wells, J.D. Hayes, S.P. Cousin, W.L. Rumsey, O.C. Attucks, S. Franklin, A.L. Levonen, T.W. Kensler, A.T. Dinkova-Kostova, Therapeutic targeting of the NRF2 and KEAP1 partnership in chronic diseases, Nat Rev Drug Discov 18(4) (2019) 295-317.
[128] N. Robledinos-Antón, R. Fernández-Ginés, G. Manda, A. Cuadrado, Activators and Inhibitors of NRF2: A Review of Their Potential for Clinical Development, Oxid Med Cell Longev 2019 (2019) 9372182.

[129] R. Gold, D.L. Arnold, A. Bar-Or, M. Hutchinson, L. Kappos, E. Havrdova, D.G. MacManus, T.A. Yousry, C. Pozzilli, K. Selmaj, M.T. Sweetser, R. Zhang, M. Yang, J. Potts, M. Novas, D.H. Miller, N.C. Kurukulasuriya, R.J. Fox, T.J. Phillips, Long-term effects of delayed-release dimethyl fumarate in multiple sclerosis: Interim analysis of ENDORSE, a randomized extension study, Mult Scler 23(2) (2017) 253-265.

[130] R.A. Linker, D.H. Lee, S. Ryan, A.M. van Dam, R. Conrad, P. Bista, W. Zeng, X. Hronowsky, A. Buko, S. Chollate, G. Ellrichmann, W. Brück, K. Dawson, S. Goelz, S. Wiese, R.H. Scannevin, M. Lukashev, R. Gold, Fumaric acid esters exert neuroprotective effects in neuroinflammation via activation of the Nrf2 antioxidant pathway, Brain 134(Pt 3) (2011) 678-92.

[131] M.P. Chin, G.L. Bakris, G.A. Block, G.M. Chertow, A. Goldsberry, L.A. Inker, H.J.L. Heerspink, M. O'Grady, P.E. Pergola, C. Wanner, D.G. Warnock, C.J. Meyer, Bardoxolone Methyl Improves Kidney Function in Patients with Chronic Kidney Disease Stage 4 and Type 2 Diabetes: Post-Hoc Analyses from Bardoxolone Methyl Evaluation in Patients with Chronic Kidney Disease and Type 2 Diabetes Study, Am J Nephrol 47(1) (2018) 40-47.

[132] A. Arlt, S. Sebens, S. Krebs, C. Geismann, M. Grossmann, M.L. Kruse, S. Schreiber, H. Schäfer, Inhibition of the Nrf2 transcription factor by the alkaloid trigonelline renders pancreatic cancer cells more susceptible to apoptosis through decreased proteasomal gene expression and proteasome activity, Oncogene 32(40) (2013) 4825-35.

[133] D. Ren, N.F. Villeneuve, T. Jiang, T. Wu, A. Lau, H.A. Toppin, D.D. Zhang, Brusatol enhances the efficacy of chemotherapy by inhibiting the Nrf2-mediated defense mechanism, Proc Natl Acad Sci U S A 108(4) (2011) 1433-8.

[134] A. Limonciel, P. Jennings, A review of the evidence that ochratoxin A is an Nrf2 inhibitor: implications for nephrotoxicity and renal carcinogenicity, Toxins (Basel) 6(1) (2014) 371-9.
[135] H. Zhang, K.J.A. Davies, H.J. Forman, Oxidative stress response and Nrf2 signaling in aging, Free Radic Biol Med 88(Pt B) (2015) 314-336.

[136] A.I. Rojo, O.N. Medina-Campos, P. Rada, A. Zúñiga-Toalá, A. López-Gazcón, S. Espada, J. Pedraza-Chaverri, A. Cuadrado, Signaling pathways activated by the phytochemical nordihydroguaiaretic acid contribute to a Keap1-independent regulation of Nrf2 stability: Role of glycogen synthase kinase-3, Free Radic Biol Med 52(2) (2012) 473-87.

[137] O.C. Attucks, K.J. Jasmer, M. Hannink, J. Kassis, Z. Zhong, S. Gupta, S.F. Victory, M. Guzel, D.R. Polisetti, R. Andrews, A.M. Mjalli, M.J. Kostura, Induction of heme oxygenase I (HMOX1) by HPP-4382: a novel modulator of Bach1 activity, PLoS One 9(7) (2014) e101044.
[138] L. Fão, S.I. Mota, A.C. Rego, Shaping the Nrf2-ARE-related pathways in Alzheimer's and Parkinson's diseases, Ageing Res Rev 54 (2019) 100942.

[139] E. Panieri, L. Saso, Potential Applications of NRF2 Inhibitors in Cancer Therapy, Oxid Med Cell Longev 2019 (2019) 8592348.

[140] S. Dibbert, B. Clement, T. Skak-Nielsen, U. Mrowietz, M. Rostami-Yazdi, Detection of fumarate-glutathione adducts in the portal vein blood of rats: evidence for rapid dimethylfumarate metabolism, Arch Dermatol Res 305(5) (2013) 447-51.

[141] A. Cuadrado, G. Manda, A. Hassan, M.J. Alcaraz, C. Barbas, A. Daiber, P. Ghezzi, R. León, M.G. López, B. Oliva, M. Pajares, A.I. Rojo, N. Robledinos-Antón, A.M. Valverde, E. Guney, H.H.H.W. Schmidt, Transcription Factor NRF2 as a Therapeutic Target for Chronic Diseases: A Systems Medicine Approach, Pharmacol Rev 70(2) (2018) 348-383.

[142] J.I. Castillo-Quan, L. Li, K.J. Kinghorn, D.K. Ivanov, L.S. Tain, C. Slack, F. Kerr, T. Nespital, J. Thornton, J. Hardy, I. Bjedov, L. Partridge, Lithium Promotes Longevity through GSK3/NRF2-Dependent Hormesis, Cell Rep 15(3) (2016) 638-650.

[143] T. Fujiki, F. Ando, K. Murakami, K. Isobe, T. Mori, K. Susa, N. Nomura, E. Sohara, T. Rai, S. Uchida, Tolvaptan activates the Nrf2/HO-1 antioxidant pathway through PERK phosphorylation, Sci Rep 9(1) (2019) 9245.

[144] P. Basak, P. Sadhukhan, P. Sarkar, P.C. Sil, Perspectives of the Nrf-2 signaling pathway in cancer progression and therapy, Toxicol Rep 4 (2017) 306-318.

[145] M.J. Kerins, A. Ooi, A catalogue of somatic NRF2 gain-of-function mutations in cancer, Sci Rep 8(1) (2018) 12846.

[146] T. Gambichler, I. Rüddel, S. Hessam, F.G. Bechara, E. Stockfleth, L. Schmitz, Altered epigenetic pathways and cell cycle dysregulation in healthy appearing skin of patients with koebnerized squamous cell carcinomas following skin surgery, J Eur Acad Dermatol Venereol 32(9) (2018) 1485-1491.

[147] C. To, C.S. Ringelberg, D.B. Royce, C.R. Williams, R. Risingsong, M.B. Sporn, K.T. Liby, Dimethyl fumarate and the oleanane triterpenoids, CDDO-imidazolide and CDDO-methyl ester, both activate the Nrf2 pathway but have opposite effects in the A/J model of lung carcinogenesis, Carcinogenesis 36(7) (2015) 769-81.

[148] T. Fulop, A. Larbi, G. Dupuis, A. Le Page, E.H. Frost, A.A. Cohen, J.M. Witkowski, C. Franceschi, Immunosenescence and Inflamm-Aging As Two Sides of the Same Coin: Friends or Foes?, Front Immunol 8 (2017) 1960.

[149] J.Y. Chan, M. Kwong, Impaired expression of glutathione synthetic enzyme genes in mice with targeted deletion of the Nrf2 basic-leucine zipper protein, Biochim Biophys Acta 1517(1) (2000) 19-26.

[150] P. Stenvinkel, J. Painer, R.J. Johnson, B. Natterson-Horowitz, Biomimetics - Nature's roadmap to insights and solutions for burden of lifestyle diseases, J Intern Med (2019).

[151] M. Martucci, R. Ostan, F. Biondi, E. Bellavista, C. Fabbri, C. Bertarelli, S. Salvioli, M. Capri, C. Franceschi, A. Santoro, Mediterranean diet and inflammaging within the hormesis paradigm, Nutr Rev 75(6) (2017) 442-455.

[152] E.J. Calabrese, R. Blain, The occurrence of hormetic dose responses in the toxicological literature, the hormesis database: an overview, Toxicol Appl Pharmacol 202(3) (2005) 289-301.
[153] T.G. Son, S. Camandola, M.P. Mattson, Hormetic dietary phytochemicals, Neuromolecular Med 10(4) (2008) 236-46.

[154] J. Maher, M. Yamamoto, The rise of antioxidant signaling--the evolution and hormetic actions of Nrf2, Toxicol Appl Pharmacol 244(1) (2010) 4-15.

[155] J.S.D. Anjos, L.F.M.F. Cardozo, A.P. Black, G. Santos da Silva, D.C.M. Vargas Reis, R. Salarolli, J.C. Carraro-Eduardo, D. Mafra, Effects of Low Protein Diet on Nuclear Factor Erythroid 2-Related Factor 2 Gene Expression in Nondialysis Chronic Kidney Disease Patients, J Ren Nutr (2019).

[156] J.S. Anjos, L.F.M.F. Cardozo, M. Esgalhado, B. Lindholm, P. Stenvinkel, D. Fouque, D. Mafra, Could Low-Protein Diet Modulate Nrf2 Pathway in Chronic Kidney Disease?, J Ren Nutr 28(4) (2018) 229-234.

[157] M.B. Stockler-Pinto, C.O. Soulage, N.A. Borges, L.F.M.F. Cardozo, C.J. Dolenga, L.S. Nakao, R. Pecoits-Filho, D. Fouque, D. Mafra, From bench to the hemodialysis clinic: protein-bound uremic toxins modulate NF-κB/Nrf2 expression, Int Urol Nephrol 50(2) (2018) 347-354.
[158] M. Liu, M. Deng, Q. Luo, X. Dou, Z. Jia, High-Salt Loading Downregulates Nrf2 Expression in a Sodium-Dependent Manner in Renal Collecting Duct Cells, Front Physiol 10 (2019) 1565.

[159] Q. Wu, D. Zhang, N. Tao, Q.N. Zhu, T. Jin, J.S. Shi, J. Liu, Induction of Nrf2 and metallothionein as a common mechanism of hepatoprotective medicinal herbs, Am J Chin Med 42(1) (2014) 207-21.

[160] M. Esgalhado, P. Stenvinkel, D. Mafra, Nonpharmacologic Strategies to Modulate Nuclear Factor Erythroid 2-related Factor 2 Pathway in Chronic Kidney Disease, J Ren Nutr 27(4) (2017) 282-291.

[161] R. Singh, S. Chandrashekharappa, S.R. Bodduluri, B.V. Baby, B. Hegde, N.G. Kotla, A.A. Hiwale, T. Saiyed, P. Patel, M. Vijay-Kumar, M.G.I. Langille, G.M. Douglas, X. Cheng, E.C. Rouchka, S.J. Waigel, G.W. Dryden, H. Alatassi, H.G. Zhang, B. Haribabu, P.K. Vemula, V.R. Jala, Enhancement of the gut barrier integrity by a microbial metabolite through the Nrf2 pathway, Nat Commun 10(1) (2019) 89.

[162] A.B. Kunnumakkara, D. Bordoloi, G. Padmavathi, J. Monisha, N.K. Roy, S. Prasad, B.B. Aggarwal, Curcumin, the golden nutraceutical: multitargeting for multiple chronic diseases, Br J Pharmacol 174(11) (2017) 1325-1348.

[163] A.S. Axelsson, E. Tubbs, B. Mecham, S. Chacko, H.A. Nenonen, Y. Tang, J.W. Fahey, J.M.J. Derry, C.B. Wollheim, N. Wierup, M.W. Haymond, S.H. Friend, H. Mulder, A.H. Rosengren, Sulforaphane reduces hepatic glucose production and improves glucose control in patients with type 2 diabetes, Sci Transl Med 9(394) (2017).

[164] N.D. Vaziri, J. Wong, M. Pahl, Y.M. Piceno, J. Yuan, T.Z. DeSantis, Z. Ni, T.H. Nguyen, G.L. Andersen, Chronic kidney disease alters intestinal microbial flora, Kidney Int 83(2) (2013) 308-15.

[165] D. Mafra, N. Borges, L. Alvarenga, M. Esgalhado, L. Cardozo, B. Lindholm, P. Stenvinkel, Dietary Components That May Influence the Disturbed Gut Microbiota in Chronic Kidney Disease, Nutrients 11(3) (2019).

[166] W.L. Lau, S.M. Liu, S. Pahlevan, J. Yuan, M. Khazaeli, Z. Ni, J.Y. Chan, N.D. Vaziri, Role of Nrf2 dysfunction in uremia-associated intestinal inflammation and epithelial barrier disruption, Dig Dis Sci 60(5) (2015) 1215-22.

[167] M. Metzger, W.L. Yuan, J.P. Haymann, M. Flamant, P. Houillier, E. Thervet, J.J. Boffa, F. Vrtovsnik, M. Froissart, L. Bankir, D. Fouque, B. Stengel, Association of a Low-Protein Diet With Slower Progression of CKD, Kidney Int Rep 3(1) (2018) 105-114.

[168] X. Chen, G. Wei, T. Jalili, J. Metos, A. Giri, M.E. Cho, R. Boucher, T. Greene, S. Beddhu, The Associations of Plant Protein Intake With All-Cause Mortality in CKD, Am J Kidney Dis 67(3) (2016) 423-30.

[169] M. Rossi, D.W. Johnson, H. Xu, J.J. Carrero, E. Pascoe, C. French, K.L. Campbell, Dietary protein-fiber ratio associates with circulating levels of indoxyl sulfate and p-cresyl sulfate in chronic kidney disease patients, Nutr Metab Cardiovasc Dis 25(9) (2015) 860-5.

[170] N. Vargas-Mendoza, Á. Morales-González, E.O. Madrigal-Santillán, E. Madrigal-Bujaidar, I. Álvarez-González, L.F. García-Melo, L. Anguiano-Robledo, T. Fregoso-Aguilar, J.A. Morales-Gonzalez, Antioxidant and Adaptative Response Mediated by Nrf2 during Physical Exercise, Antioxidants (Basel) 8(6) (2019).

[171] L. George, M.F. Lokhandwala, M. Asghar, Exercise activates redox-sensitive transcription factors and restores renal D1 receptor function in old rats, Am J Physiol Renal Physiol 297(5) (2009) F1174-80.

[172] C.C. Abreu, L.F. Cardozo, D. Mafra, Could physical exercises modulate Nrf2-Keap1 pathway in chronic kidney disease?, Med Hypotheses 84(1) (2015) 44-6.

[173] C.C. Abreu, L.F.M.F. Cardozo, M.B. Stockler-Pinto, M. Esgalhado, J.E. Barboza, R. Frauches, D. Mafra, Does resistance exercise performed during dialysis modulate Nrf2 and NFκB in patients with chronic kidney disease?, Life Sci 188 (2017) 192-197.

[174] A.J. Done, M.J. Gage, N.C. Nieto, T. Traustadóttir, Exercise-induced Nrf2-signaling is impaired in aging, Free Radic Biol Med 96 (2016) 130-8.

[175] M.J. Magbanua, E.L. Richman, E.V. Sosa, L.W. Jones, J. Simko, K. Shinohara, C.M. Haqq, P.R. Carroll, J.M. Chan, Physical activity and prostate gene expression in men with low-risk prostate cancer, Cancer Causes Control 25(4) (2014) 515-23.

[176] E. Barrón-Cabrera, O. Ramos-Lopez, K. González-Becerra, J.I. Riezu-Boj, F.I. Milagro, E. Martínez-López, J.A. Martínez, Epigenetic Modifications as Outcomes of Exercise Interventions Related to Specific Metabolic Alterations: A Systematic Review, Lifestyle Genom (2019) 1-20.