



PRIFYSGOL
BANGOR
UNIVERSITY

Dietary nitrate supplementation effect on dynamic cerebral autoregulation in normoxia and acute hypoxia

Horiuchi, Masahiro; Rossetti, Gabriella; Oliver, Sam

Journal of Cerebral Blood Flow and Metabolism

DOI:

[10.1177%2F0271678X20910053](https://doi.org/10.1177%2F0271678X20910053)

E-pub ahead of print: 09/03/2020

Peer reviewed version

[Cyswllt i'r cyhoeddiad / Link to publication](#)

Dyfyniad o'r fersiwn a gyhoeddwyd / Citation for published version (APA):

Horiuchi, M., Rossetti, G., & Oliver, S. (2020). Dietary nitrate supplementation effect on dynamic cerebral autoregulation in normoxia and acute hypoxia. *Journal of Cerebral Blood Flow and Metabolism*. <https://doi.org/10.1177%2F0271678X20910053>

Hawliau Cyffredinol / General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

This article was accepted in its current form to Journal of Cerebral Blood Flow & Metabolism on 31st Jan 2020. DOI:

Title: Dietary nitrate supplementation effect on dynamic cerebral autoregulation in normoxia and acute hypoxia

Running title: Dietary nitrate and cerebral autoregulation

Masahiro Horiuchi¹, Gabriella M.K. Rossetti², Samuel J. Oliver²

1. Division of Human Environmental Science, Mt. Fuji Research Institute
2. Extremes Research Group, College of Human Sciences, Bangor University

Corresponding author: Masahiro Horiuchi, Ph.D

Address; Kami-yoshida 5597-1, Fuji-yoshida city, Yamanashi, 4030005, Japan

e-mail; mhuriuchi@mfri.pref.yamanashi.jp

TEL; +81-555-72-6198

FAX; +81-555-72-6216

Abstract

We tested the hypothesis that increasing the nitric oxide (NO) bioavailability by dietary nitrate would recover the hypoxic-induced reduction in dynamic CA. Twelve healthy males (age 21 ± 2 years) completed four days of dietary supplementation with a placebo or inorganic nitrate drink (140-ml beetroot juice) followed by 60-min of normoxia or hypoxia (fraction of inspired oxygen [FiO_2] = 13%). Duplex ultrasonography was used to perform volumetric change-based assessment of dynamic CA in the internal carotid artery (ICA). Dynamic CA was assessed by rate of regulation (RoR) of vascular conductance using the thigh-cuff method. Four days of beetroot supplementation increased circulating nitrate by 208 [171,245] μ M (mean difference [95% confidence interval]) compared with placebo. Dynamic CA was lower in hypoxia than normoxia (RoR Δ -0.085 [-0.116, -0.054]). Compared with placebo, nitrate did not alter dynamic CA in normoxia (RoR Δ -0.022 [-0.060, 0.016]) or hypoxia (RoR Δ 0.017 [-0.019, 0.053]). Further, nitrate did not affect ICA vessel diameter, blood velocity or flow in either normoxia or hypoxia. Increased bioavailability of NO through dietary nitrate supplementation did not recover the hypoxia-induced reduction in dynamic CA. This suggests the mechanism of hypoxia-induced reduction in dynamic CA does not relate to the availability of NO.

Key words: *nitric oxide, acute hypotension, cerebral blood flow, vasodilation, vascular conductance*

INTRODUCTION

Cerebral autoregulation (CA) is essential to protect the brain against ischaemic injury, capillary damage, and edema^{1,2}. Dynamic CA is the maintenance of cerebral blood flow (CBF) at constant levels despite acute changes in cerebral perfusion pressure or in systemic blood pressure³. The physiological mechanisms responsible for the control of dynamic cerebral autoregulation are thought to involve a combination of nitric oxide (NO), neurogenic (sympathetic and cholinergic) and myogenic (stretching vascular smooth muscle) factors¹. While dynamic CA normally ensures the preservation of CBF, including to transient hypertension^{4,5}, studies demonstrate that dynamic CA is reduced during transient hypotension in normoxic and hypoxic conditions (i.e. acute hypoxia and at high altitude)⁴⁻¹⁴. The impairment in dynamic CA is characterized by a blunted increase in cerebrovascular conductance (CVC) in response to systemic hypotension, preventing normalization of CBF¹³. The mechanism by which hypoxia reduces dynamic CA is unclear but may relate to nitric oxide (NO) availability¹⁵.

NO is a potent vasodilator and plays a vital role in the regulation of CVC, and consequently CBF¹⁶. Previous research in humans, breathing normal ambient air, has shown dynamic CA to be substantially reduced when the enzyme NO synthase is inhibited¹⁵. In oxygen-depleted environments, the L-arginine pathway, which is helped by the enzyme NO-synthase, has a decreased ability to generate NO¹⁷. NO can also be

generated by the serial reduction of nitrate (NO_3^-) to nitrite (NO_2^-) and then NO, providing an alternative pathway for NO production in hypoxia^{18,19}. Dietary nitrate supplementation has been shown to be effective to increase the NO bioavailability²⁰, and improve cerebral regulation and CBF in normoxia^{21,22}. Specifically, dietary nitrate supplementation has previously been shown to enhance cerebral perfusion²¹ and increase middle cerebral artery blood flow velocity via a reduction in cerebrovascular resistance²². In addition, dietary nitrate has been shown to reverse the hypoxia-induced impairment in peripheral NO-dependent endothelial function²³. Combined, this suggests dietary nitrate may be effective to reverse the hypoxia-induced impairment in dynamic CA.

The aim of this study was to determine the role of NO availability in the hypoxia-induced reduction of dynamic CA. We hypothesized nitrate supplementation would reverse the hypoxia-induced reduction in dynamic CA, which would provide evidence that NO plays a central role in the mechanism of hypoxia-induced reduction in dynamic CA.

METHODS

Participants

All procedures were approved by the ethical committee of Mt. Fuji Research Institute and were performed in accordance with the guidelines of the Declaration of Helsinki (ECMFRI-01-2014). After a detailed explanation of all study procedures, including the possible risks and benefits of participation, each participant gave his written consent. Twelve healthy male participants with a mean age of 21 ± 2 years, 174 ± 5 cm, and body mass 69 ± 8 kg (mean \pm standard deviation [SD]) were enrolled. They were free from any cardiovascular or cerebrovascular diseases, and were not taking any medications. Participants engaged in regular recreational sports (1-2 h per day, 2-4 days per week). None of the participants was exposed to an altitude higher than 1500 m within 6 months before the study. In addition, participants were asked to abstain from caffeine for 12 h and from strenuous exercise and alcohol for at least 24 h before the study. Participants were familiarized with measurement techniques (i.e., thigh-cuff testing for dynamic CA and measurement of blood flow in the internal carotid artery [ICA]). All studies were performed in an environmental chamber (TBR-4, 5SA2GX, Tabai Espec Co, Ltd., Tokyo, Japan) with set at an ambient temperature of 24°C and at relative humidity of 40%.

Study design

The study followed a double-blinded placebo-controlled crossover design. All participants performed two trials, each assessing dynamic CA in both normoxia and

hypoxia after four days of (1) nitrate-rich beetroot juice (nitrate) and (2) nitrate-depleted beetroot juice (placebo) dietary supplementation. These two trials were performed with at least a 2-week washout period and in random order. An overview of the study protocol is shown in Figure 1.

Supplementation

For the three days before the trials, participants consumed 140 ml per day of NO_3^- -rich beetroot juice (nitrate; Beet It, James White Drinks, Ltd., Ipswich, UK) or 140 mL per day of NO_3^- -depleted beetroot concentrate (placebo; James White Drinks, Ltd., Ipswich, UK). Both participants and researchers were blinded to the drink contents until the completion of the study. Participants were also provided a list of foods rich in NO_3^- , and instructed to avoid the consumption of these foods and to otherwise maintain their normal dietary intake for the duration of the study. In addition, they were asked to abstain from the use of antibacterial mouthwash for the duration of the study, as it eliminates the oral bacteria that reduce NO_3^- to NO_2^- ²⁴.

Study procedures

On each study day (nitrate or placebo), participants consumed their final dose of nitrate or placebo on arrival to the laboratory, about 1.5 h before the first dynamic CA test. After a 30 min supine and 45 min semi-recumbent rest on a manually inclined bed

(A4524-0096L, Paramount Bed Co., Ltd., Tokyo, Japan), a venous blood sample (10 mL) was collected from the antecubital vein into a vacutainer and immediately centrifuged at 1000 g for 15 min at 4°C (Kubota 5200, Kubota Co., Ltd., Tokyo, Japan) to separate serum from whole blood. The serum samples were frozen at -80°C for further analysis of NO₃⁻ content by SRL Co., Ltd. (Tokyo, Japan). Briefly, after deproteinization, serum NO₃⁻ was measured by using high-performance liquid chromatography²⁵. After attachment of all devices, normoxic baseline values were measured for 5 min; then, dynamic CA assessment in normoxia was performed. Thereafter, hypoxic gas (FiO₂ 0.13) was supplied via a commercial tent (about 5000 L) with a hypoxic gas generating system (Hypoxico Everest Summit 2: Will Co., Ltd., Tokyo, Japan). Hypoxic baseline values were measured during the last 5 min of the 60-min resting hypoxic exposure. Next, dynamic CA assessment in hypoxia was performed. Inspired oxygen concentration was verified before and after each experiment using a metabolic cart (AE-310s; Minato Medical Science, Osaka, Japan).

Dynamic CA assessment

Dynamic CA was assessed with participants sat on a comfortable chair in a semi-recumbent position. After 2 min of baseline data in normoxia were recorded, bilateral thigh-cuffs (width 22 cm) were inflated to a pressure of 220 mmHg for 3 min using a custom cuff inflator. To evaluate dynamic CA, the cuff was then rapidly deflated,

causing a transient drop in arterial blood pressure, and measurements were continued for an additional minute. Participants were instructed to remain relaxed and were not given feedback regarding the elapsed time during cuff occlusion. This protocol was repeated two or three times at 5-min intervals to allow for recovery to the resting value²⁶. If the first or second trial reduction in mean arterial pressure (MAP) was < 15 mmHg²⁷, a third trial was performed, and the average value of two trials was used¹³.

Measurements

Right ICA measurements were performed 1.0–1.5 cm distal to the carotid bifurcation with a Doppler ultrasound set at 10.0 MHz and a linear transducer (Logic-e; GE Healthcare, Tokyo, Japan). For baseline ICA flow measurement, the ICA blood flow was averaged over 2 min during the last 5 min of the initial 30-min resting exposure. For the dynamic ICA blood flow measurements, blood flow was continuously measured during the thigh cuff test. To calculate the average ICA blood flow, we analyzed the mean vessel diameter (D_{mean}) and flow velocity as described in a previous study²⁸. Briefly, after obtaining a clear image of the vessel using the brightness mode, the mean vessel diameter was calculated as: mean diameter = (systolic diameter \times 1/3) + (diastolic diameter \times 2/3). The time-averaged mean flow velocity obtained using the pulse wave mode was defined as the mean blood flow velocity (V_{mean} ; in centimeters per second). Blood flow was calculated by multiplying the cross-sectional area \times 60 (in

milliliters per minute). Throughout the measurement, care was taken to ensure that the probe position was stable, the insonation angle did not vary ($< 60^\circ$ in all cases) and the sample volume was positioned in the center of the vessel and adjusted to cover the width of the vessel diameter. Heart rate (HR) was monitored using three-lead ECG. Pulmonary ventilation (\dot{V}_E) and partial pressure of end-tidal carbon dioxide output (P_{ETCO_2}) were measured by a breath-by-breath gas analyzer (AE-310S; Minato Medical Science, Osaka, Japan). Standard gases (O_2 15.23%, CO_2 4.999%, and N_2 balance) and room air were used to calibrate the gas analyzer before each test. Beat-by-beat MAP was measured using finger photoplethysmography from the middle or index finger of the left hand (MUB-101; Medi-sens, Saiatama, Japan). Peripheral arterial oxygen saturation (SpO_2) was monitored by finger pulse oximetry (PULFIS WB-100, Japan Precision Instruments Inc., Gunma, Japan).

Data analysis

We calculated the rate of regulation (RoR) as an indicator of dynamic CA^{29} . For calculation of RoR, baseline values for MAP, ICA blood flow and cerebrovascular conductance (ICA/MAP) were defined as the mean during the 4 s immediately before the thigh-cuff release. The RoR was calculated from the slope of the regression line between ICA conductance and the time from 1.0 to 4.0 s after thigh-cuff deflation and normalized to the degree of cuff-release-induced hypotension. The $RoR = (\Delta ICA$

conductance/ Δt)/ Δ MAP, where Δ ICA conductance/ Δt is the slope of the linear regression between ICA conductance and time (t), and Δ MAP is the magnitude of the blood pressure step, which was calculated by subtracting the baseline MAP from the MAP averaged during the interval from 1.0 to 4.0 s. In a previous study, the data from 1.0 to 3.5 s after cuff deflation were used to calculate dynamic CA²⁹; however, we extended this time to 4.0 s after cuff deflation to ensure beat-by-beat blood flow data for at least three cardiac cycles^{13,28}. Trials in which the reduction in MAP was < 15 mmHg were excluded from analysis, and the average of two trials was used. The coefficient of variation of this measure is 13%.

Statistics

Values are expressed as means \pm SD unless otherwise stated. Two-way repeated-measures ANOVAs (oxygen \times supplement) were used for comparisons of cardiorespiratory, hemodynamic and dynamic CA variables across conditions. P values < 0.05 were considered to indicate statistical significance. The magnitude of effect of nitrate supplementation on the hypoxia-induced decline in dynamic CA variables was determined by ANCOVA comparison of nitrate and placebo hypoxia trials (with normoxia placebo as the covariate), and interpreted in relation to *a priori* meaningful differences³⁰. A sample size estimation for the primary analysis indicated that 10 participants were needed to produce an 80% chance of obtaining statistical significance

at the 0.05 level for a two-tailed design, based on a minimum important difference of 0.076 RoR, a standard deviation of the difference of 0.05, and an estimated correlation of 0.4¹¹. Statistical analyses were performed using commercial software packages (Sigma Stat 3.5, Hulinks, Chicago, IL, USA; SPSS V25, IBM Corp, Armonk; NY).

RESULTS

Serum nitrate

Four days inorganic nitrate supplementation increased serum NO₃⁻ concentrations compared to placebo (mean difference [95% confidence interval]: Δ208 [171, 245] μM; $P < 0.001$).

Cardiorespiratory and hemodynamic responses at rest

Effect of hypoxia. Compared to normoxia, acute hypoxia decreased SpO₂, and increased HR, ICA diameter, ICA blood flow, and CVC (all $P < 0.05$; Table 1). Further, acute hypoxia tended to increase \dot{V}_E ($P = 0.06$) and decrease P_{ET}CO₂ ($P = 0.09$). Hypoxia did not alter MAP or ICA blood flow velocity at rest ($P > 0.05$; Table 1).

Effect of nitrate. Dietary nitrate supplementation did not affect any of the resting cardiovascular or cerebrovascular responses in either normoxia or hypoxia (all $P > 0.05$; Table 1).

Dynamic cerebral autoregulation

Effect of hypoxia. Thigh-cuff release elicited similar MAP reductions in hypoxia and normoxia ($P = 0.7$; Table 2). In contrast, the ICA blood flow nadir in response to thigh-cuff was greater in hypoxia than normoxia ($P < 0.001$; Table 2). Dynamic CA assessment (1-4 s after deflation) indicated that hypoxia did not alter the Δ MAP, but did decrease the slope of CVC ($P < 0.05$; Table 2). Consequently, RoR was decreased in hypoxia compared to normoxia (Δ -0.085 [-0.116, -0.054]; $P < 0.05$; Figure 3).

Effect of nitrate. Thigh-cuff release elicited similar reductions in MAP and ICA flow after nitrate and placebo in normoxia and hypoxia (Table 2; interactions $P > 0.9$).

During dynamic CA assessment (1-4s after deflation), nitrate had no effect on any dynamic CA variable in normoxia or hypoxia when compared to the placebo (Table 2, Figures 2 and 3). Specifically, compared to placebo, nitrate supplementation did not alter RoR in normoxia (Δ -0.022 [-0.060, 0.016]; $P = 0.2$) or hypoxia (Δ 0.017 [-0.019, 0.053]; $P = 0.3$). Further, compared to placebo, nitrate supplementation did not effect the hypoxia-induced decline in dynamic CA variables (Figure 2): MAP (Δ -0.06 [-3.62, 3.50]; $P = 0.9$), ICA flow (Δ -1 [-26, 23]; $P = 0.9$), CVC (Δ -0.003 [-0.009, 0.002]; $P = 0.2$), or RoR (Δ 0.020 [-0.010, 0.051]; $P = 0.2$).

DISCUSSION

This study is the first to investigate the effects of dietary nitrate supplementation on dynamic CA in hypoxia. The major findings of this study are therefore that increased bioavailability of NO by dietary nitrate supplementation did not influence dynamic CA in acute hypoxia or recover the hypoxia-induced reduction in dynamic CA (Figure 3). In addition, we also show that dietary nitrate supplementation did not influence cerebrovascular responses, including dynamic CA, whilst participants breathed normal ambient air (normoxia). A particular strength of this study was the method used to determine dynamic CA, i.e. duplex Doppler ultrasound in conjunction with the traditional thigh-cuff test. In brief, we used duplex Doppler ultrasound as it enables the assessment of blood flow velocity and vessel diameter, which contrasts the easier-to-perform and more commonly used TCD method that only assesses blood flow velocity. Previous research has shown the thigh-cuff method, which imposes a large and abrupt change in blood pressure, is more sensitive to interventions, such as hypoxia, than transfer function analysis that examines spontaneous fluctuations in blood pressure²⁶. The importance of using these methods is emphasised when considering the only other study to have examined dietary nitrate supplementation on dynamic CA in normoxia reported inconclusive effects³¹ that may have resulted from using TCD in conjunction transfer function analysis, which is a less sensitive method to assess dynamic CA than used in the present study.

In our study, acute hypoxia reduced dynamic CA which is a consistent finding with many other studies⁶⁻¹⁴. Indeed, 10 of 12 participants had reduced dynamic CA with hypoxia irrespective of supplementation. However, in contrast with our hypothesis, and previous research that suggests NO may have an important role in regulating brain vascular tone³² and dynamic CA¹⁵, dietary nitrate supplementation did not influence any physiological determinant of dynamic CA (Figure 2), or dynamic CA (Figure 3), differently than the placebo in acute hypoxia. Indeed, the mean difference in RoR between nitrate and placebo and the 95% confidence intervals indicate any effect of nitrate is trivial in magnitude. This finding suggests NO bioavailability has limited influence on the hypoxia-induced reduction in dynamic CA observed in young healthy adults. Of note, our results do not discount the role of NO in CBF responses including dynamic CA in hypoxia. CBF regulation is a complex, multifaceted physiological process, of which the action of NO is a key component³³. The data from this study simply suggest that insufficient bioavailability of NO is not the primary cause of hypoxia-induced reduction in dynamic CA.

A possible alternative mechanism for the hypoxia-induced reduction in dynamic CA could be attributed to increased vasodilation at rest²⁹. Hypoxia is a potent vasodilator³⁴, and therefore cerebral blood vessels may already be close to maximally dilated in hypoxia, limiting capacity for further vasodilation to maintain CVC during dynamic CA. In our study hypoxia substantially increased ICA diameter at rest (Table

1), which may have limited the capacity for further dilation in dynamic CA response to the thigh cuff release. Regions with already elevated CBF at rest in hypoxia have been previously shown to minimally increase CBF with experimental manipulations known to cause vasodilation³⁵, supporting the idea that baseline diameter influences capacity for further dilation. Indeed, previous research supports that dynamic CA depends on basal vascular tone, and that the autoregulatory response in middle cerebral artery blood flow velocity is slower when cerebral vessels are already dilated²⁹. Similarly, lower cerebrovascular resistance (higher cerebrovascular conductance) resulted in higher transfer function gain, indicating that dynamic CA is reduced when cerebral vessels are dilated³⁶. Other factors, including prostaglandins³⁷⁻⁴⁰, and sympathetic nerve activity^{33,41}, have been shown to contribute the regulation of CBF and may also be responsible for the hypoxia-induced reduction in dynamic CA.

Limitations and perspectives

Our findings should be considered carefully within the context they were obtained. To assess dynamic CA we measured changes in ICA blood vessel diameter and blood velocity flow. Global CBF however consists of ~75% ICA flow and ~25% vertebral artery (VA) flow⁴² so, although technically challenging, a more complete assessment of CA would have included both ICA and VA. Further, as one previous magnetic resonance imaging study observed regional but not global CBF differences after nitrate

supplementation²¹, future studies examining the influence of dietary nitrate on regional extra- and intracranial cerebral blood flow and autoregulation are warranted. Dietary nitrate supplementation has been shown to be effective to recover peripheral vascular function at high altitude²³. In contrast, dietary nitrate did not affect cerebral vascular function during hypoxic exposure in the present study. This heterogenous response between vascular beds is consistent with a previous study where dietary nitrate increased muscle oxygenation in hypoxia, but did not affect cerebral oxygenation⁴³. In combination these findings provide evidence dietary nitrate and NO influence peripheral and cerebral vascular responses to hypoxia by different mechanisms. The limited effects of dietary nitrate observed on dynamic CA in this study may be a consequence of the young healthy population recruited. Indeed, as other studies in young healthy individuals we report no effect of dietary nitrate on resting physiological responses in normoxia or hypoxia^{23,44,45}. Whether dietary nitrate influences cerebrovascular responses and dynamic CA in persons with reduced vascular function, poor NO bioavailability, or clinical populations with cerebrovascular disease remains to be investigated. Future research should also resolve whether reductions in dynamic CA are related to an increase in adverse outcomes such as headache and Acute Mountain Sickness, which are common in hypoxic conditions^{7,9,13,46}. Resolving this is important as dynamic CA could be a useful screening-tool to identify those persons at risk of hypoxia-induced adverse events.

CONCLUSION

In conclusion, a four-day dietary inorganic nitrate supplementation did not affect resting cerebrovascular responses or dynamic CA in either normoxia or hypoxia (after 60-min exposure with 13% O₂) in healthy men. This suggests NO availability is not responsible for hypoxia-induced reduction in dynamic CA.

Acknowledgements: The authors thank all participants for their time and effort. The authors also thank Mrs. Kirihara-Handa Yoko, and Hamada-Endo Junko for their technical assistance.

Funding: This study was partly supported by the funding of Japan Society for the Promotion of Science (# 26440268 to M.H.)

Author contributions: M. H. designed and performed the study. M.H., G. M. K. R., and S. J. O. analysed and interpreted results. M.H., G. M. K. R., and S. J. O. prepared tables and figures. M.H. drafted the first manuscript. M.H., G. M. K. R., and S. J. O. edited and revised the manuscript. All authors have approved the final version.

Conflict of interest: The authors declared no conflicts of the interest with respect to this study.

References

1. Tzeng YC, Ainslie PN. Blood pressure regulation IX: Cerebral autoregulation under blood pressure challenges. *Eur J Appl Physiol* 2014; 114: 545–559.
2. Paulson OB, Strandgaard S, Edvinsson L. Cerebral autoregulation. *Cerebrovasc Brain Metab Rev* 1990; 2: 161–192.
3. Lassen NA. Cerebral blood flow and oxygen consumption in man. *Physiol Rev* 1959; 39: 183–238.
4. Aaslid R, Blaha M, Svirid G, et al. Asymmetric dynamic cerebral autoregulatory response to cyclic stimuli. *Stroke* 2007; 38(5):1465-9.
5. Tzeng YC, Willie CK, Atkinson G, et al. Cerebrovascular regulation during transient hypotension and hypertension in humans. *Hypertension* 2010; 56: 268-273.
6. Iwasaki KI, Ogawa Y, Shibata S, et al. Acute exposure to normobaric mild hypoxia alters dynamic relationships between blood pressure and cerebral blood flow at very low frequency. *J Cereb Blood Flow Metab* 2007; 27: 776–784.
7. Bailey DM, Evans KA, James PE, et al. Altered free radical metabolism in acute mountain sickness: implications for dynamic cerebral autoregulation and blood-brain barrier function. *J Physiol* 2009; 587: 73–85.
8. Subudhi AW, Panerai RB, Roach RC. Acute hypoxia impairs dynamic cerebral autoregulation: results from two independent techniques. *J Appl Physiol* 2009;

- 107: 1165–1171.
9. Subudhi AW, Panerai RB, Roach RC. Effects of hypobaric hypoxia on cerebral autoregulation. *Stroke* 2010; 41: 641–646.
 10. Nishimura N, Iwasaki K, Ogawa Y, et al. Decreased steady-state cerebral blood flow velocity and altered dynamic cerebral autoregulation during 5-h sustained 15% O₂ hypoxia. *J Appl Physiol* 2010; 108: 1154–1161.
 11. Ogoh S, Nakahara H, Ainslie PN, et al. The effect of oxygen on dynamic cerebral autoregulation: critical role of hypocapnia. *J Appl Physiol* 2010; 108: 538–543.
 12. Ainslie PN, Lucas SJE, Fan J-L, et al. Influence of sympathoexcitation at high altitude on cerebrovascular function and ventilatory control in humans. *J Appl Physiol* 2012; 113: 1058–1067.
 13. Horiuchi M, Endo J, Dobashi S, et al. Effect of progressive normobaric hypoxia on dynamic cerebral autoregulation. *Exp Physiol* 2016; 82: 1496–1514.
 14. Subudhi AW, Fan JL, Evero O, et al. AltitudeOmics: Effect of ascent and acclimatization to 5260 m on regional cerebral oxygen delivery. *Exp Physiol* 2014; 99: 772–781.
 15. White RP, Vallance P, Markus HS. Effect of inhibition of nitric oxide synthase on dynamic cerebral autoregulation in humans. *Clin Sci* 2000; 99: 555–60.
 16. Toda N, Ayajiki K, Okamura T. Cerebral blood flow regulation by nitric oxide: recent advances. *Pharmacol Rev* 2009; 61: 62–97.

17. McQuillan LP, Leung GK, Marsden PA, et al. Hypoxia inhibits expression of eNOS via transcriptional and posttranscriptional mechanisms. *Am J Physiol Circ Physiol* 2017; 268: 1–1.
18. Duncan C, Dougall H, Johnston P, et al. Chemical generation of nitric oxide in the mouth from the enterosalivary circulation of dietary nitrate. *Nat Med* 1995; 1: 546–551.
19. Lundberg JO, Weitzberg E, Gladwin MT. The nitrate-nitrite-nitric oxide pathway in physiology and therapeutics. *Nat Rev Drug Discov* 2008; 7: 156–167.
20. Wylie LJ, Kelly J, Bailey SJ, et al. Beetroot juice and exercise: pharmacodynamic and dose-response relationships. *J Appl Physiol* 2013; 115: 325–336.
21. Presley TD, Morgan AR, Bechtold E, et al. Acute effect of a high nitrate diet on brain perfusion in older adults. *Nitric Oxide* 2011; 24: 34–42.
22. Bond V, Curry BH, Adams RG, et al. Effects of dietary nitrates on systemic and cerebrovascular hemodynamics. *Cardiol Res Pract* 2013: 435629, DOI: 10.1155/2013/435629.
23. Bakker E, Engan H, Patrician A, et al. Acute dietary nitrate supplementation improves arterial endothelial function at high altitude: a double-blinded randomized controlled cross over study. *Nitric Oxide* 2015; 50: 58–64.
24. Govoni M, Jansson EA, Weitzberg E, et al. The increase in plasma nitrite after a dietary nitrate load is markedly attenuated by an antibacterial mouthwash. *Nitric*

- Oxide* 2008; 19: 333–337.
25. Takahashi H, Nakanishi T, Nishimura M, et al. Measurements of serum levels of nitrate ions in men and women: implications of endothelium-derived relaxing factor in blood pressure regulation and atherosclerosis. *Journal Cardiovasc Pharmacol* 1992; 20: S214–S216.
 26. Subudhi AW, Grajzel K, Langolf RJ, et al. Cerebral autoregulation index at high altitude assessed by thigh-cuff and transfer function analysis techniques. *Exp Physiol* 2015; 100: 173–181.
 27. Tiecks FP, Lam AM, Aalsid R, et al. Comparison of static and dynamic cerebral autoregulation measurements. *Stroke* 1995; 26: 1014–1019.
 28. Sato K, Fisher JP, Seifert T, et al. Blood flow in internal carotid and vertebral arteries during orthostatic stress. *Exp Physiol* 2012; 97: 1272–1280.
 29. Aaslid R, Lindegaard KF, Sorteberg W, et al. Cerebral autoregulation dynamics in humans. *Stroke* 1989; 20: 45–52.
 30. Hopkins WG, Batterham AM. Error rates, decisive outcomes and publication bias with several inferential methods. *Sport Med* 2016; 46: 1563–1573.
 31. Fan J-L, O'Donnell T, Gray CL, et al. Dietary nitrate supplementation enhances cerebrovascular CO₂ reactivity in a sex-specific manner. *J Appl Physiol* 2019; 127: 760–769.
 32. Van Mil AHM, Spilt A, Van Buchem MA, et al. Nitric oxide mediates hypoxia-

- induced cerebral vasodilation in humans. *J Appl Physiol* 2002; 92: 962–966.
33. Ainslie PN, Ogoh S. Regulation of cerebral blood flow in mammals during chronic hypoxia: a matter of balance. *Exp Physiol* 2010; 95: 251–262.
 34. Ainslie PN, Subudhi AW. Cerebral blood flow at high altitude. *High Alt Med Biol* 2014; 15: 133–40.
 35. Lawley JS, Macdonald JH, Oliver SJ, et al. Unexpected reductions in regional cerebral perfusion during prolonged hypoxia. *J Physiol* 2017; 595: 935–947.
 36. Serrador JM, Sorond FA, Vyas M, et al. Cerebral pressure-flow relations in hypertensive elderly humans: transfer gain in different frequency domains. *J Appl Physiol* 2005; 98: 151–159.
 37. Fan JL, Burgess KR, Thomas KN, et al. Influence of indomethacin on the ventilatory and cerebrovascular responsiveness to hypoxia. *Eur J Appl Physiol* 2011; 111: 601–610.
 38. Harrell JW, Schrage WG. Cyclooxygenase-derived vasoconstriction restrains hypoxia-mediated cerebral vasodilation in young adults with metabolic syndrome. *Am J Physiol Circ Physiol* 2014; 306: H261–H269.
 39. Bowton DL, Haddon WS, Prough DS, et al. Theophylline effect on the cerebral blood flow response to hypoxemia. *Chest* 1988; 94: 371–375.
 40. Meno JR, Ngai AC, Richard Winn H. Changes in pial arteriolar diameter and CSF adenosine concentrations during hypoxia. *J Cereb Blood Flow Metab* 1993; 13:

214–220.

41. Curran-Everett DC, Meredith MP, Krasney JA. Acclimatization to hypoxia alters cerebral convective and diffusive O₂ delivery. *Respir Physiol* 1992; 88: 355–371.
42. Schöning M, Walter J, Scheel P. Estimation of cerebral blood flow through color duplex sonography of the carotid and vertebral arteries in healthy adults. *Stroke* 1994; 25: 17–22.
43. Masschelein E, Van Thienen R, Wang X, et al. Dietary nitrate improves muscle but not cerebral oxygenation status during exercise in hypoxia. *J Appl Physiol* 2012; 113: 736–745.
44. Lefferts WK, Hughes WE, White CN, et al. Effect of acute nitrate supplementation on neurovascular coupling and cognitive performance in hypoxia. *Appl Physiol Nutr Metab* 2016; 4: 133–141.
45. Rossetti GMK, Macdonald JH, Wylie LJ, et al. Dietary nitrate supplementation increases acute mountain sickness severity and sense of effort during hypoxic exercise. *J Appl Physiol* 2017; 123: 983–992.
46. Cochand NJ, Wild M, Brugniaux JV, et al., Sea-level assessment of dynamic cerebral autoregulation predicts susceptibility to acute mountain sickness at high altitude. *Stroke* 2011; 42(12):3628-30.

Figure Legends

Figure 1. The protocol of the present study. BL, baseline; dCA, dynamic cerebral autoregulation assessment; FiO₂, fraction of inspired oxygen.

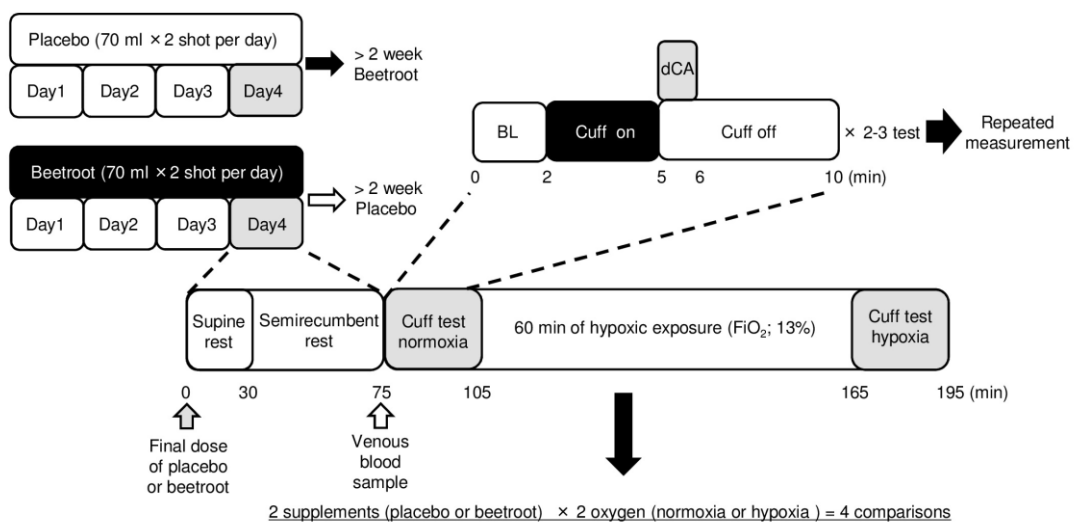


Figure 2. Comparisons in the components of dynamic cerebral autoregulation among the four conditions. The supplement contrast is defined by dashed (placebo) versus solid (nitrate) lines, the oxygen contrast is defined by black (normoxia) versus grey (hypoxia) lines. As such, the four conditions are represented as follows: placebo normoxia, dashed black line; nitrate normoxia, solid black line; placebo hypoxia, dashed grey line; nitrate hypoxia, solid grey line. All values are presented as changes from baseline to cuff release, normalised to baseline in each condition. Hypoxia (A) had no effect on mean arterial pressure (MAP), (B) decreased internal carotid artery (ICA) flow, and (C) decreased cerebrovascular conductance (CVC) following thigh cuff release ($*P < 0.05$). However, nitrate had no effect on (A) MAP, (B) ICA flow, or (C) CVC (all $P > 0.05$). Values are means \pm SD. $*P < 0.05$ between normoxia and hypoxia.

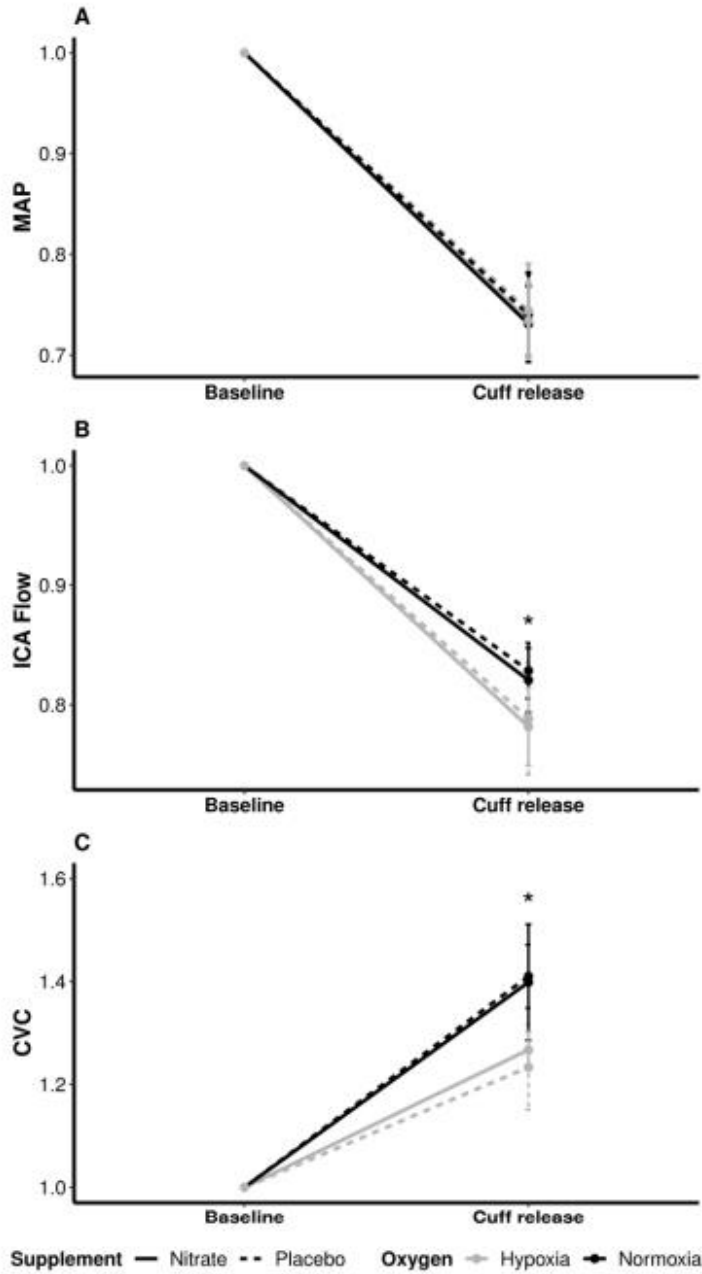


Figure 3. Dynamic cerebral autoregulation (rate of regulation: RoR) in normoxia and hypoxia after placebo and nitrate supplementation. * Significant difference between normoxia and hypoxia ($P > 0.05$). Values are means \pm standard deviation (SD).

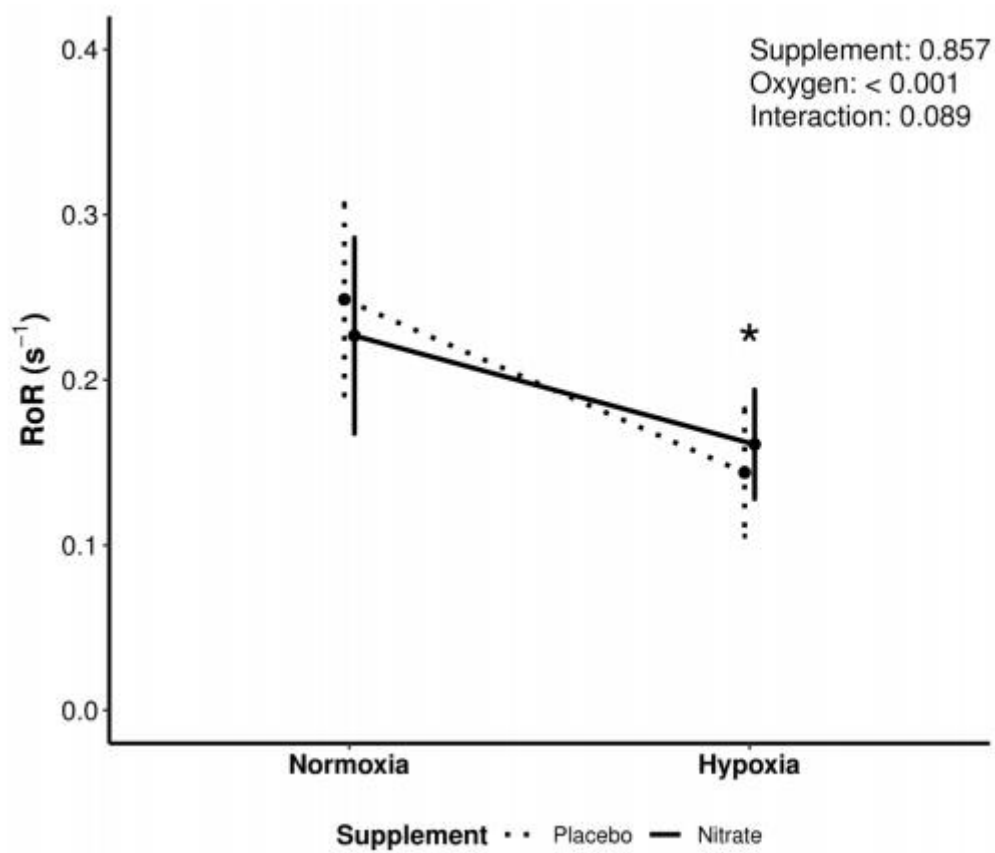


Table 1. Resting cardiorespiratory and hemodynamic responses in normoxia and hypoxia after placebo and nitrate supplementation.

	Placebo		Nitrate		Two-way ANOVA				
	Normoxia	Hypoxia	Normoxia	Hypoxia	Suppl.	Oxygen	Interaction		
<i>Cardiorespiratory</i>									
\dot{V}_E , l min ⁻¹	10.0 ± 1.4	10.6 ± 1.4	10.2 ± 0.9	10.6 ± 1.7	0.678	0.059	0.784		
P _{ET} CO ₂ , mmHg	37.8 ± 1.9	36.5 ± 1.9	37.7 ± 2.1	36.5 ± 1.7	0.852	0.094	0.903		
HR, bpm	63 ± 5	68 ± 5*	64 ± 5	69 ± 4*	0.137	<0.001	0.828		
MAP, mmHg	83 ± 5	85 ± 6	84 ± 5	85 ± 5	0.342	0.133	0.712		
SpO ₂ , %	98.2 ± 0.5	85.2 ± 4.8*	98.4 ± 0.8	85.9 ± 4.0*	0.160	<0.001	0.442		
<i>Internal carotid artery</i>									
Blood flow, ml min ⁻¹	327 ± 20	345 ± 26*	337 ± 21	356 ± 36*	0.274	0.022	0.880		
Blood velocity, cm s ⁻¹	29.0 ± 5.3	26.4 ± 2.5	27.9 ± 4.9	26.2 ± 3.5	0.555	0.149	0.655		
Diameter, cm	0.49 ± 0.05	0.53 ± 0.03*	0.51 ± 0.04	0.54 ± 0.04*	0.252	0.015	0.821		
CVC, ml min ⁻¹ mmHg ⁻¹	3.90 ± 0.38	4.05 ± 0.34*	4.07 ± 0.45	4.21 ± 0.52*	0.256	0.034	0.928		

Values are means ± standard deviation (SD). \dot{V}_E , pulmonary ventilation, P_{ET}CO₂, partial pressure of end-tidal carbon dioxide; HR, heart rate; bpm, beats per minute; MAP, mean arterial pressure; SpO₂, arterial oxygen saturation; CVC, cerebrovascular conductance; Suppl., supplementation. * indicates a statistical difference between normoxia and hypoxia within the same supplementation.

Table 2. Hemodynamic responses to the thigh-cuff test in normoxia and hypoxia after placebo and nitrate supplementation.

	Placebo		Nitrate		Two-way ANOVA		
	Normoxia	Hypoxia	Normoxia	Hypoxia	Suppl.	Oxygen	Interaction
<i>Thigh-cuff response</i>							
Max. Δ MAP (%)	-26.0 \pm 4.1	-25.6 \pm 4.6	-26.9 \pm 3.8	-26.6 \pm 3.8	0.525	0.683	0.913
Max. Δ ICA flow (%)	-17.2 \pm 2.3	-21.2 \pm 4.7*	-17.9 \pm 2.7	-21.8 \pm 3.3*	0.557	<0.001	0.987
<i>Dynamic CA variables (1-4 s after deflation)</i>							
Slope of CVC	0.041 \pm 0.006	0.023 \pm 0.008*	0.040 \pm 0.011	0.026 \pm 0.004*	0.646	<0.001	0.345
Δ MAP (%)	-17.0 \pm 3.0	-16.7 \pm 5.3	-17.9 \pm 3.5	-17.1 \pm 3.4	0.631	0.591	0.804

Values are mean \pm SD. MAP, mean arterial pressure; ICA, internal carotid artery; CVC, cerebrovascular conductance; Suppl., supplementation. * indicate significant difference between normoxia and hypoxia within the same supplementation.