Oxidative stress, telomeres and cellular senescence: What non-drug interventions might break the link?

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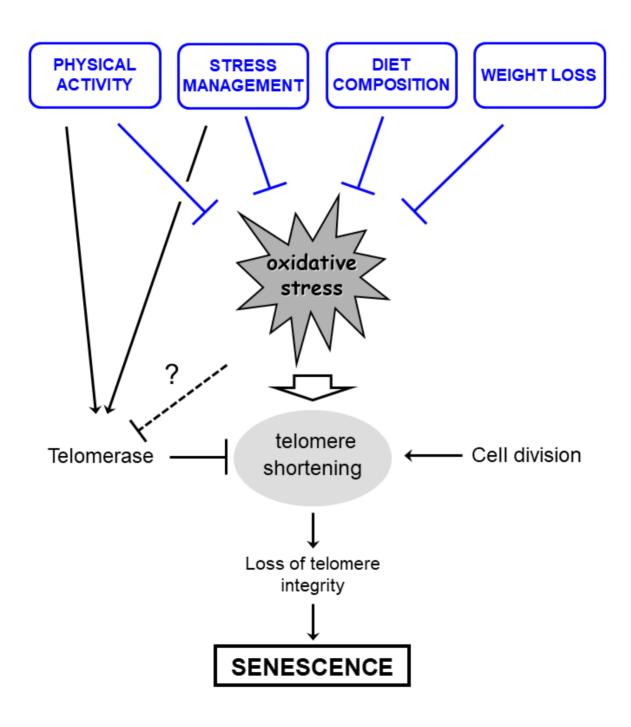
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Graphical abstract



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Abstract

Telomeres are higher order structures that cap and protect chromosome ends. Telomeric DNA naturally shortens during somatic cell division and as a result of oxidative stress. Excessive shortening disrupts the integrity of the telomere, causing cellular senescence, one of the hallmarks of organismal ageing. The accumulation of senescent cells with ageing contributes to the loss of tissue homeostasis and the development of age-related pathologies. Hence, counteracting telomere shortening may be one relevant approach to develop strategies for healthier ageing. In this review I present the case for the existence of a link between oxidative stress, accelerated telomere shortening and cellular senescence. I also examine findings from human observational studies exploring associations between telomere length and oxidative stress-related parameters. Finally, I discuss results from randomised control trials testing the impact of non-pharmacological lifestyle interventions on the maintenance of telomere length, considering the potential mechanisms that might be involved.

Keywords

Ageing, diet, lifestyle, oxidative stress, physical activity, psychological stress, senescence, telomere, weight loss

Introduction

Telomeres are higher-order structures made of a tandemly repeated guanine-rich DNA sequence (TTAGGG in vertebrates) and a specialized protein complex called shelterin, that cap and stabilize the physical ends of chromosomes, protecting them from degradation and end-to-end fusions [1, 2]. Synthesis and maintenance of telomeric DNA requires in addition to DNA polymerase, a specialized reverse transcriptase called telomerase [3, 4]. In humans telomerase is tightly regulated by differentiation and proliferation signals. It is expressed at high levels in embryonic tissues and adult stem cell compartments, but is hardly detectable, in most other human adult somatic cell types [5, 6]. In addition, telomerase activity may be upregulated in some cells under stimulatory conditions, e.g. in endothelial cells by growth factors [6, 7] or in lymphocytes by activation signals [8].

Due to the inability of conventional DNA polymerases to replicate the 5'-end of the lagging strand, when telomerase is repressed DNA synthesis during cell division results in a gradual loss of telomeric DNA [9]. In addition, because of its high guanine content, telomeric DNA is intrinsically susceptible to oxidative modifications and single strand break formation [10]. Furthermore, since at the telomere oxidative lesions are less efficiently removed, at the time of replication persistent lesions may also lead to the truncation of the telomeric DNA backbone (reviewed in [11]). Accordingly, work with cultured cells has shown that oxidative stress accelerates telomere shortening [12, 13].

Erosion of telomeric DNA eventually compromises the functional integrity of the whole telomere structure, triggering a DNA damage response, which in turn may result in the onset of cellular senescence [14]. Cellular senescence is a distinctive stress-response phenomenon that enforces an essentially irreversible cell-cycle arrest. In addition, it gives rise to a secretory phenotype endowed with pro-inflammatory, growth promoting and tissue remodelling properties [15]. This response may be beneficial in early life and adulthood, for example as an anti-neoplastic mechanism [16], or to promote wound healing [17]. In contrast, in later life, due to the age-related impairment of clearance systems, an accumulation of senescent cells equipped with a pro-inflammatory baggage is thought to contribute to the loss of tissue homeostasis and the development of age-related pathologies (reviewed in [18].

In human cells telomeric DNA is heterogeneous in length, measuring between ~4 to ~15 kb. A decrease in leukocyte telomere length (LTL), which generally correlates with telomere length in other body tissues [19, 20], is a feature of normal ageing [21]. In addition, greater telomere shortening in leukocytes or peripheral blood mononuclear cells (PBMC) is associated with an increased risk of age-related diseases [4, 22, 23] and mortality [4, 24, 25]. Hence, counteracting telomere shortening may be relevant to reduce the accumulation of senescent cells and their pathological consequences.

Telomere length is influenced by biological [26-29], environmental [30], psychosocial [31-35] and life-style factors [36-40]. These include, gene variants of telomerase and telomere protective proteins, epigenetic signatures, sex, paternal age at birth, ethnicity, body size, biological stress responsivity, levels of inflammation, tobacco smoking, physical activity, diet and nutrition, as well as psychosocial stress, education and pollution levels. Some of these factors are also known to affect the oxidative burden of the organism, raising the possibility that they influence telomere shortening in this way. In this review I summarize evidence linking oxidative stress to telomere shortening *in vivo* and highlight areas of controversy. Furthermore, given that the harmful biological impacts of some of the above factors could be mitigated by changing life style behaviours, I also discuss non-drug interventions that may have the potential to reduce telomere shortening via oxidative stress-suppressing mechanisms.

The relationships between oxidative stress and telomere length in humans

Heightened levels of reactive oxygen species (ROS) can cause macromolecular damage and thus contribute to the development of ageing phenotypes [41-43]. This notion, together with early research in cultured cells showing that biological mediators of disease which induce oxidative stress, also shorten telomeres and lead to premature senescence (reviewed in [7]), prompted a search for associations *in vivo* between telomere length and markers of oxidative damage or antioxidant defences. One of the first indications that linked oxidative stress with shorter telomeres in humans came from a small study investigating the relationship of these parameters with perceived psychological stress in healthy pre-menopausal women [31]. An inverse correlation between LTL and urine levels of the isoprostane 8-epi-PGF_{2α}, a marker of lipid peroxidation, was subsequently described in a larger sample

of Caucasian men [44]. However, further studies carried out in different population settings revealed a mixed picture (Table 1). Thus, associations between telomere length and selected markers of oxidative damage or antioxidant defences were found in type II diabetes or Parkinson's disease but not in healthy individuals [45-47]. Associations with plasma antioxidant micronutrients [48] and with oxidative stress-related gene polymorphisms [49] have also been documented. On the other hand, a handful of studies reported no associations. Thus, research investigating risks for breast cancer found no significant correlations between telomere length and urinary levels of 8-hydroxy-2-deoxyguanosine (8-OHdG) or 15-F₂-isoprostanes, either in the patient group or in the healthy controls [50]. Similarly, no correlations between LTL and diverse oxidative stress-related blood markers were found in Spanish patients with cardiovascular disease [51] or in mostly healthy Chinese individuals [52], except for a positive association with superoxide dismutase in the latter.

Importantly, in a recent study which examined interactions between redox state markers and telomere length in blood versus vascular tissue of patients with coronary artery disease, tissue-specific inverse associations between superoxide production and telomere length were described [53]. Furthermore, these relationships were influenced by the additive effects of two functional single nucleotide polymorphisms in the CYBA gene, which encodes one of the NADPH oxidase subunits, thus, suggesting causality. In addition, the study showed no associations between telomere length of blood cells and either plasma superoxide dismutase activity or total plasma anti-oxidant capacity, or between vascular telomere length and vascular superoxide dismutase activity, suggesting that antioxidant defences played no role in the regulation of telomere length.

Thus, taken together the available evidence only lends partial support to the concept that oxidative stress contributes to telomere shortening *in vivo*. Several factors could contribute to the discrepancies found between studies, including the ethnical and clinical characteristics of the populations studied, the type of tissue sampled and the markers of oxidative stress tested. Of note however, in those studies that reported associations with telomere length, only a minority showed these to be with antioxidant defences.

Non-pharmacological interventions

A substantial number of observational studies have described associations between telomere length and potentially modifiable lifestyle behaviours, although conflicting results have also been reported (reviewed in [54-60]). These observations led the way for interventional studies aimed at ascertaining whether non-pharmacological approaches focusing on life-style changes could influence telomere length. The first clear indication that such interventions might have an effect came from a five-year follow-up study carried out in men at low risk of prostate cancer [61]. At the end of the study the group that engaged in a comprehensive life-style change programme, which included diet, physical activity, stress management and social support, showed a significant increase in PBMC telomere length compared to a decrease in the control group [61]. Using a randomised control trial (RCT) design, other studies have evaluated mainly four distinct intervention modalities, namely diet composition, physical activity, weight loss and psychological stress management. Table 2 summarizes examples of such RCTs lasting for at least eight weeks. The studies were generally small and heterogeneous with respect to the age and health characteristics of the participants, as well as the duration of the interventions. The majority lasted for less than a year [62-72], while four trials were conducted for a year or more [73-77]. In all of them telomere length was determined in either leukocytes or PBMC before and after the intervention. However, although the type of interventions described are known to impinge on oxidative-stress responses [78-82], only a few related telomere length measurements to changes in oxidative stressrelated parameters [62, 64, 65]. A brief description of these studies is summarized below.

Dietary interventions

Some of the health benefits of fish-rich diets have been attributed to a high content of long chain omega-3 (n-3) polyunsaturated fatty acids (PUFAs) and the capacity of these compounds to decrease oxidative stress [83, 84]. In this respect, small trials have attempted to ascertain whether n-3 PUFA supplementation also affected telomere length. In one trial where participants received two different doses of n-3 PUFAs or a placebo for four months, changes in plasma F₂-isoprostanes were also recorded. In this study increases in telomere length in the PUFA supplementation groups were observed, compared to a decrease in the placebo group, although the

differences between groups were not significant [62]. However, a secondary analysis revealed that the increases in telomere length were associated with decreasing plasma n-6:n-3 PUFA ratios. n-3 PUFA supplementation was also shown to significantly reduce F₂-isoprostanes, but an attempt to correlate this measure with telomere length was apparently not carried out in this case [62]. A second trial with participants suffering from mild cognitive impairment evaluated a six month intervention with supplements rich in the n-3 PUFAs eicosapentanoic acid (EPA) or docosahexanoic acid (DHA) against the n-6 PUFA linolenic acid (LA) as the control [63]. In this case, telomere length decreased more in the LA group than in the DHA or EPA groups, but the differences were not statistically significant. Nevertheless, it was found that in the DHA group an increase in red blood cell DHA levels correlated with reduced telomere shortening [63]. Thus, while the above two studies do not seem to have sufficient statistical power to reach to a firm conclusion, taken together they suggest that telomere shortening may be attenuated by n-3 PUFA supplementation. Consistent with these findings, in an opportunistic two year trial conducted in cognitively healthy elders, a diet rich in walnuts, which is a source of n-3 PUFAs, showed a trend to preserve telomere length when compared to a control diet [73].

The Mediterranean diet (MD) is rich in sources of bioactive compounds that are thought to play a role in protecting the organism against inflammation and other sources of oxidative stress [60]. This protective function may be achieved in part by activating detoxifying stress responses, which *in vitro* are known to ameliorate telomere shortening and senescence [85]. *In vivo*, the effect of the MD on telomere length has been examined in a relatively large RCT, which had been originally designed to investigate the effects of this diet on the primary prevention of cardiovascular disease. In this study participants who were given the MD supplemented with olive oil showed no differences in telomere erosion over a five year period when compared to a control group given a low fat diet; unexpectedly, accelerated telomere shortening was observed in participants on the MD supplemented with nuts [75]. Importantly however, the study found that greater adherence to the MD reduced telomere shortening in individuals carrying the Ala variant of the PPARy2 gene, but not in those carrying the Pro variant [74]. Thus, although the study concluded that overall there was no beneficial effect of the MD

intervention on slowing telomere shortening in comparison to a low fat diet, it also highlighted the potential relevance of genetic factors when considering the effects of diets on telomere length. Unfortunately, although the MD is known to increase antioxidant defences [86] and reduce oxidative damage [87] in this high cardiovascular risk population, no comparisons between changes in oxidative stress markers and changes in telomere length were reported in this case.

Exercise and physical activity

Physical exercise increases ROS generation acutely. This in turn triggers an antioxidant defence response, which with regular exercise practice becomes protective [88]. This protection against oxidative stress is one mechanism by which physical activity might reduce telomere attrition. To test this concept, the effect of exercise training or increase in physical activity on telomere length was investigated in small RCTs carried out in a variety of settings (Table 2). Some of the studies also examined the effect of the intervention on enzymes involved in redox homeostasis [64, 65] or on telomerase activity [67, 89]. One intervention carried out in obese middle age women showed that although six month of aerobic exercise training resulted in an increase in glutathione peroxidase activity, it did not cause significant changes in telomere length [64]. In another study which evaluated the effect of a twelve week low frequency explosive-type resistance training in elderly people, telomere length increased in the intervention group and decreased in the control group, the difference in telomere length between trained and untrained participants being significant [65]. In this study, significant reductions of mitochondrial superoxide dismutase 2 and thioredoxin reductase 1 in PBMC, as well as in serum myeloperoxidase, were detected in the intervention group but not in controls [65]. In a third study in which overweight older adults received physical activity of low/moderate intensity on prescription for six months there were no significant differences in telomere length between the control and the intervention group. However, in the exercise group a reduction in the sitting time was strongly correlated with telomere lengthening, whereas no such correlation was observed in the control group [66]. More recently, in one RCT conducted with family caregivers, aerobic exercise training was shown to increase telomere length after six months, without causing changes in telomerase activity [67]. In contrast, in another RCT carried out

with healthy sedentary postmenopausal women, twelve month of aerobic exercise showed no effect on telomere length [76]. Finally, a recent RCT in healthy sedentary participants compared the effects of three exercise training modalities on telomerase activity and telomere length in PBMCs [89]. Notably, this study found that aerobic endurance training or high intensity interval training for six month increased both telomerase activity and telomere length; in contrast, no such changes were induced by resistance training. Thus, this study suggest that in otherwise sedentary healthy people, specific training protocols may have differential effects on telomere length.

Weight loss

Calorie restriction causing weight loss without malnutrition has been shown to improve telomere length maintenance [90, 91] as well as to reduce oxidative stress and senescence [90] in lean mice. Conversely, mice made obese by feeding a high fat diet show increased levels of oxidative DNA damage and a reduction in telomere length, compared to lean animals fed a normal chow [92]. In agreement with findings in mice, obesity is one of the factors more consistently associated with telomere shortness in human observational studies [29, 56]. However, whether low calorie diets aimed at reducing weight, ameliorate telomere attrition also in humans, remains to be demonstrated. Indeed, in a RCT that examined the effects of calorie restriction on predictors of longevity and disease risk factors in healthy non-obese individuals, telomere length was not measured [93]. Furthermore, in obese or overweight participants, low calorie diets on their own or in combination with physical activity have not reduced telomere shortening significantly [69, 77]. Thus, in a relatively large study in older women carried out over a twelve-month period, there were no significant changes in LTL in any intervention group, whether weight loss had been achieved by dietary restriction, aerobic exercise or a combination of both, compared to controls. In this study, no association between the magnitude of weight loss and telomere length was found either [77]. Similarly, in another smaller trial carried out in breast cancer survivors, which combined exercise with diet, while an increase in telomere length was detected in the intervention group at six months compared to a decrease in the control group, the difference between groups was not significant [69].

Stress management

A variety of studies has reported associations between telomere length, telomerase activity and psychosocial stress (reviewed in [94]) but the mechanisms underlying these relationships remain to be fully elucidated. Chronic psychological stress has been linked to higher levels of oxidative DNA damage and decrease in anti-oxidant defences [54], possibly triggered in part by higher levels of the stress-related hormone cortisol [95]. In support of this possibility, a longitudinal study in healthy late middle age people has recently suggested that heightened cortisol responsivity may mediate the relationship between psychological stress and telomere shortening [28]. It should be noted however, that the effect of cortisol on telomere length could be mediated also by an inhibition of telomerase activity [96] and not necessarily only by oxidative stress. In addition, other pathways could be involved in this relationship, including a psychological stress-induced increase in pro-inflammatory cytokines [97, 98], which are also known be associated to greater telomere attrition [62].

Attenuating the physiological components of the stress response is a reasonable strategy to reduce the effects of psychological stress on telomere length and cellular ageing. In this context, a growing interest in the relationship between telomere length and psychological stress management has emerged over the last few years [99]. Yoga and meditation are two modalities that might reduce levels of systemic inflammation, stress-hormones and oxidative stress. These practices have also been linked with increases in telomerase activity and/or the amelioration of telomere maintenance in some observational and interventional studies [82, 99]. Two studies in particular examined the effect of yoga on telomere-related parameters and redox markers [100, 101]. The first one was a case-control study, which found that experienced yoga practitioners had significantly longer leukocyte telomeres, a higher total anti-oxidant status, and lower levels of malondialdehyde and homocysteine in serum, compared to healthy controls. In addition, in this study a strong correlation between telomere length and each of the redox markers was reported, suggesting that habitual yoga practice ameliorates telomere shortening by reducing systemic oxidative stress levels [100]. The second study was a prospective single arm exploratory trial lasting for three months in which 94 healthy participants were subjected to a yoga and meditation-based lifestyle intervention [101]. In this study, levels of 8-OHdG and ROS were significantly lower after the intervention compared to baseline, while the total anti-oxidant capacity and telomerase activity were

significantly higher. In contrast, although telomere length was shown to increase, the difference was not significant [101]. Notably, and consistent with this finding a handful of small RCTs lasting between two and nine months has shown no effect of meditation practices on telomere length (Table 2). Taken together, these studies suggest that stress management techniques on their own are unlikely to demonstrate significant effects on telomere length if applied for relatively short periods of time (e.g. for less than two years).

Conclusions, challenges and future perspectives

Mounting evidence indicates that life-style changes aimed at promoting health and wellbeing do so by reducing the pro-inflammatory and oxidative burden of the organism. In line with this notion, as described in this review, over the last 10 years an increasing number of RCT have tested whether non-drug interventions known for their oxidative stress-reducing potential, could also help to maintain telomere length over time. However, although mechanistically plausible, the picture that emerges from those studies is at present equivocal (Figure 1). Aside from limitations in the precision and heterogeneity of methods to measure telomere length and oxidative stress, including in some cases, the shortcoming inherent to the use of a single biomarker to measure the latter, inconsistencies between findings may be due to a variety of additional factors. These include the demographic, genetic and health characteristics of the groups studied, as well as the duration and intensity of the interventions. Furthermore, some studies could have been statistically underpowered to detect significant changes in telomere length. Finally, concerning the duration of the interventions, it should be emphasised that the majority lasted for less than a year and it is unclear whether telomere length can indeed change with interventions of such length [102, 103].

In considering the biological mechanisms that are recruited by non-pharmacological treatments, one should contemplate the role of telomerase (Figure 1). The weight of evidence suggests that meditation can reactivate telomerase in leukocytes with relatively short interventions [99]. Similarly, mouse and human studies indicate that exercise can also induce an increase in telomerase activity [68, 104]. Notably, in one prospective human study the exercise-induced increase in telomerase activity was shown to precede an increase in telomere length [104]. However, telomerase activity

and telomere length do not show always a positive correlation [61, 99, 105]; thus, whether such re-activation can affect telomere shortening in different scenarios is unclear. Telomerase activity is under multiple controls, some of which are also influenced by redox mechanisms [106]. Furthermore, oxidative stress supresses telomerase activity *in vitro* [13], although it is not known whether this is relevant *in vivo*. Thus, in evaluating the link between oxidative stress and telomere length *in vivo*, cross-talk between telomerase activity and the redox status of the organism and/or the relevant cell type should also be considered.

For practical reasons in human studies telomere length has been mostly measured in leukocytes or PBMC. In this respect, it should be considered that the expansion of leukocyte subsets that occurs during inflammation and immunological responses may also cause accelerated telomere shortening, irrespective of the levels of oxidative stress to which these cells are subjected. In fact, measurement of telomere length in blood should be regarded as a measure of the senescent status of circulating immune cells. Thus, to take account of these potential confounding effects future interventional studies should also look at tissues other than blood, for example the buccal epithelium.

Taken together the current evidence suggest that longer, larger (adequately powered) and better-designed RCT are required in order to confirm whether lifestyle changes are effective in reducing telomere-related cellular senescence. Larger interventions would also allow for stratification according to genetic variants and thus establish if genetic factors modulate the effectiveness of some of these interventions. These studies should consider where possible, taking samples from tissues other than blood, and include several reliable measures of oxidative stress, which are relevant to the cells and tissues from which telomere length measurements are taken. Finally, we should not neglect the notion that social and public health measures, aimed at increasing the educational attainment of the population or reduce pollution levels, are *bona fide* non-pharmacological interventions with a much wider remit, which might also contribute to extend health span by reducing telomere-related cellular senescence.

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References

- [1] Blackburn, E. H.; Greider, C. W.; Szostak, J. W. Telomeres and telomerase: the path from maize, Tetrahymena and yeast to human cancer and aging. *Nat Med* **12**:1133-1138; 2006.
- [2] Palm, W.; de Lange, T. How shelterin protects mammalian telomeres. *Annu Rev Genet* **42:**301-334; 2008.
- [3] Lingner, J.; Hughes, T. R.; Shevchenko, A.; Mann, M.; Lundblad, V.; Cech, T. R. Reverse transcriptase motifs in the catalytic subunit of telomerase. *Science* **276**:561-567; 1997.
- [4] Blackburn, E. H.; Epel, E. S.; Lin, J. Human telomere biology: A contributory and interactive factor in aging, disease risks, and protection. *Science* **350**:1193-1198; 2015.
- [5] Forsyth, N. R.; Wright, W. E.; Shay, J. W. Telomerase and differentiation in multicellular organisms: turn it off, turn it on, and turn it off again. *Differentiation* **69**:188-197; 2002.
- [6] Kurz, D. J.; Hong, Y.; Trivier, E.; Huang, H. L.; Decary, S.; Zang, G. H.; Luscher, T. F.; Erusalimsky, J. D. Fibroblast growth factor-2, but not vascular endothelial growth factor, upregulates telomerase activity in human endothelial cells. *Arterioscler. Thromb. Vasc. Biol* 23:748-754; 2003.
- [7] Erusalimsky, J. D. Vascular endothelial senescence: from mechanisms to pathophysiology. *J Appl. Physiol* **106**:326-332; 2009.
- [8] Weng, N. P.; Palmer, L. D.; Levine, B. L.; Lane, H. C.; June, C. H.; Hodes, R. J. Tales of tails: regulation of telomere length and telomerase activity during lymphocyte development, differentiation, activation, and aging. *Immunol Rev* 160:43-54; 1997.
- [9] Olovnikov, A. M. Telomeres, telomerase, and aging: origin of the theory. *Exp. Gerontol* **31**:443-448; 1996.
- [10] von Zglinicki, T.; Pilger, R.; Sitte, N. Accumulation of single-strand breaks is the major cause of telomere shortening in human fibroblasts. *Free Radic. Biol. Med* **28**:64-74; 2000.
- [11] Ahmed, W.; Lingner, J. Impact of oxidative stress on telomere biology. *Differentiation* 99:21-27; 2018.
- [12] von Zglinicki, T. Oxidative stress shortens telomeres. *Trends Biochem. Sci* 27:339-344; 2002.
- [13] Kurz, D. J.; Decary, S.; Hong, Y.; Trivier, E.; Akhmedov, A.; Erusalimsky, J. D. Chronic oxidative stress compromises telomere integrity and accelerates the onset of senescence in human endothelial cells. *J. Cell Sci* **117**:2417-2426; 2004.
- [14] d'Adda di Fagagna, F.; Reaper, P. M.; Clay-Farrace, L.; Fiegler, H.; Carr, P.; von Zglinicki, T.; Saretzki, G.; Carter, N. P.; Jackson, S. P. A DNA damage checkpoint response in telomereinitiated senescence. *Nature* **426**:194-198; 2003.
- [15] Tchkonia, T.; Zhu, Y.; van Deursen, J.; Campisi, J.; Kirkland, J. L. Cellular senescence and the senescent secretory phenotype: therapeutic opportunities. *J Clin Invest* **123**:966-972; 2013.
- [16] Campisi, J. Cellular senescence as a tumor-suppressor mechanism. *Trends Cell Biol* 11:S27-S31; 2001.
- [17] Demaria, M.; Ohtani, N.; Youssef, S. A.; Rodier, F.; Toussaint, W.; Mitchell, J. R.; Laberge, R. M.; Vijg, J.; Van Steeg, H.; Dolle, M. E.; Hoeijmakers, J. H.; de Bruin, A.; Hara, E.; Campisi, J. An essential role for senescent cells in optimal wound healing through secretion of PDGF-AA. *Developmental cell* **31**:722-733; 2014.
- [18] Childs, B. G.; Gluscevic, M.; Baker, D. J.; Laberge, R. M.; Marquess, D.; Dananberg, J.; van Deursen, J. M. Senescent cells: an emerging target for diseases of ageing. *Nat Rev Drug Discov* 16:718-735; 2017.
- [19] Daniali, L.; Benetos, A.; Susser, E.; Kark, J. D.; Labat, C.; Kimura, M.; Desai, K.; Granick, M.; Aviv, A. Telomeres shorten at equivalent rates in somatic tissues of adults. *Nat Commun* 4:1597; 2013.

- [20] Wilson, W. R.; Herbert, K. E.; Mistry, Y.; Stevens, S. E.; Patel, H. R.; Hastings, R. A.; Thompson, M. M.; Williams, B. Blood leucocyte telomere DNA content predicts vascular telomere DNA content in humans with and without vascular disease. *Eur. Heart J* 29:2689-2694; 2008.
- [21] Aubert, G.; Lansdorp, P. M. Telomeres and aging. *Physiol Rev* 88:557-579; 2008.
- [22] Haycock, P. C.; Heydon, E. E.; Kaptoge, S.; Butterworth, A. S.; Thompson, A.; Willeit, P. Leucocyte telomere length and risk of cardiovascular disease: systematic review and metaanalysis. *BMJ* 349:g4227; 2014.
- [23] Forero, D. A.; Gonzalez-Giraldo, Y.; Lopez-Quintero, C.; Castro-Vega, L. J.; Barreto, G. E.; Perry, G. Meta-analysis of Telomere Length in Alzheimer's Disease. *J Gerontol A Biol Sci Med Sci* 71:1069-1073; 2016.
- [24] Cawthon, R. M.; Smith, K. R.; O'Brien, E.; Sivatchenko, A.; Kerber, R. A. Association between telomere length in blood and mortality in people aged 60 years or older. *Lancet* 361:393-395; 2003.
- [25] Wang, Q.; Zhan, Y.; Pedersen, N. L.; Fang, F.; Hagg, S. Telomere Length and All-Cause Mortality: A Meta-analysis. Ageing Res Rev 48:11-20; 2018.
- [26] Hunt, S. C.; Chen, W.; Gardner, J. P.; Kimura, M.; Srinivasan, S. R.; Eckfeldt, J. H.; Berenson, G. S.; Aviv, A. Leukocyte telomeres are longer in African Americans than in whites: the National Heart, Lung, and Blood Institute Family Heart Study and the Bogalusa Heart Study. *Aging Cell* 7:451-458; 2008.
- [27] Codd, V.; Nelson, C. P.; Albrecht, E.; Mangino, M.; Deelen, J.; Buxton, J. L.; Hottenga, J. J.; Fischer, K.; Esko, T.; Surakka, I.; Broer, L.; Nyholt, D. R.; Mateo Leach, I.; Salo, P.; Hagg, S.; Matthews, M. K.; Palmen, J.; Norata, G. D.; O'Reilly, P. F.; Saleheen, D.; Amin, N.; Balmforth, A. J.; Beekman, M.; de Boer, R. A.; Bohringer, S.; Braund, P. S.; Burton, P. R.; de Craen, A. J.; Denniff, M.; Dong, Y.; Douroudis, K.; Dubinina, E.; Eriksson, J. G.; Garlaschelli, K.; Guo, D.; Hartikainen, A. L.; Henders, A. K.; Houwing-Duistermaat, J. J.; Kananen, L.; Karssen, L. C.; Kettunen, J.; Klopp, N.; Lagou, V.; van Leeuwen, E. M.; Madden, P. A.; Magi, R.; Magnusson, P. K.; Mannisto, S.; McCarthy, M. I.; Medland, S. E.; Mihailov, E.; Montgomery, G. W.; Oostra, B. A.; Palotie, A.; Peters, A.; Pollard, H.; Pouta, A.; Prokopenko, I.; Ripatti, S.; Salomaa, V.; Suchiman, H. E.; Valdes, A. M.; Verweij, N.; Vinuela, A.; Wang, X.; Wichmann, H. E.; Widen, E.; Willemsen, G.; Wright, M. J.; Xia, K.; Xiao, X.; van Veldhuisen, D. J.; Catapano, A. L.; Tobin, M. D.; Hall, A. S.; Blakemore, A. I.; van Gilst, W. H.; Zhu, H.; Erdmann, J.; Reilly, M. P.; Kathiresan, S.; Schunkert, H.; Talmud, P. J.; Pedersen, N. L.; Perola, M.; Ouwehand, W.; Kaprio, J.; Martin, N. G.; van Duijn, C. M.; Hovatta, I.; Gieger, C.; Metspalu, A.; Boomsma, D. I.; Jarvelin, M. R.; Slagboom, P. E.; Thompson, J. R.; Spector, T. D.; van der Harst, P.; Samani, N. J. Identification of seven loci affecting mean telomere length and their association with disease. Nat Genet **45**:422-427, 427e421-422; 2013.
- [28] Steptoe, A.; Hamer, M.; Lin, J.; Blackburn, E. H.; Erusalimsky, J. D. The Longitudinal Relationship Between Cortisol Responses to Mental Stress and Leukocyte Telomere Attrition. *The Journal of clinical endocrinology and metabolism* **102**:962-969; 2017.
- [29] Gielen, M.; Hageman, G. J.; Antoniou, E. E.; Nordfjall, K.; Mangino, M.; Balasubramanyam, M.; de Meyer, T.; Hendricks, A. E.; Giltay, E. J.; Hunt, S. C.; Nettleton, J. A.; Salpea, K. D.; Diaz, V. A.; Farzaneh-Far, R.; Atzmon, G.; Harris, S. E.; Hou, L.; Gilley, D.; Hovatta, I.; Kark, J. D.; Nassar, H.; Kurz, D. J.; Mather, K. A.; Willeit, P.; Zheng, Y. L.; Pavanello, S.; Demerath, E. W.; Rode, L.; Bunout, D.; Steptoe, A.; Boardman, L.; Marti, A.; Needham, B.; Zheng, W.; Ramsey-Goldman, R.; Pellatt, A. J.; Kaprio, J.; Hofmann, J. N.; Gieger, C.; Paolisso, G.; Hjelmborg, J. B. H.; Mirabello, L.; Seeman, T.; Wong, J.; van der Harst, P.; Broer, L.; Kronenberg, F.; Kollerits, B.; Strandberg, T.; Eisenberg, D. T. A.; Duggan, C.; Verhoeven, J. E.; Schaakxs, R.; Zannolli, R.; Dos Reis, R. M. R.; Charchar, F. J.; Tomaszewski, M.; Mons, U.; Demuth, I.; Iglesias Molli, A. E.; Cheng, G.; Krasnienkov, D.; D'Antono, B.; Kasielski, M.; McDonnell, B. J.; Ebstein, R. P.; Sundquist, K.; Pare, G.; Chong, M.; Zeegers, M. P.; group, T. Body mass index is negatively

associated with telomere length: a collaborative cross-sectional meta-analysis of 87 observational studies. *Am J Clin Nutr* **108**:453-475; 2018.

- [30] Zota, A. R.; Needham, B. L.; Blackburn, E. H.; Lin, J.; Park, S. K.; Rehkopf, D. H.; Epel, E. S. Associations of cadmium and lead exposure with leukocyte telomere length: findings from National Health and Nutrition Examination Survey, 1999-2002. *Am J Epidemiol* 181:127-136; 2015.
- [31] Epel, E. S.; Blackburn, E. H.; Lin, J.; Dhabhar, F. S.; Adler, N. E.; Morrow, J. D.; Cawthon, R. M. Accelerated telomere shortening in response to life stress. *Proc. Natl. Acad. Sci. U. S. A* 101:17312-17315; 2004.
- [32] Steptoe, A.; Hamer, M.; Butcher, L.; Lin, J.; Brydon, L.; Kivimaki, M.; Marmot, M.; Blackburn, E.; Erusalimsky, J. D. Educational attainment but not measures of current socioeconomic circumstances are associated with leukocyte telomere length in healthy older men and women. *Brain Behav Immun* 25:1292-1298; 2011.
- Brydon, L.; Lin, J.; Butcher, L.; Hamer, M.; Erusalimsky, J. D.; Blackburn, E. H.; Steptoe, A.
 Hostility and cellular aging in men from the Whitehall II cohort. *Biol Psychiatry* **71**:767-773; 2012.
- [34] Adler, N.; Pantell, M. S.; O'Donovan, A.; Blackburn, E.; Cawthon, R.; Koster, A.; Opresko, P.; Newman, A.; Harris, T. B.; Epel, E. Educational attainment and late life telomere length in the Health, Aging and Body Composition Study. *Brain Behav Immun* 27:15-21; 2013.
- [35] Needham, B. L.; Adler, N.; Gregorich, S.; Rehkopf, D.; Lin, J.; Blackburn, E. H.; Epel, E. S. Socioeconomic status, health behavior, and leukocyte telomere length in the National Health and Nutrition Examination Survey, 1999-2002. Soc Sci Med 85:1-8; 2013.
- [36] Valdes, A. M.; Andrew, T.; Gardner, J. P.; Kimura, M.; Oelsner, E.; Cherkas, L. F.; Aviv, A.; Spector, T. D. Obesity, cigarette smoking, and telomere length in women. *Lancet* 366:662-664; 2005.
- [37] Cassidy, A.; De Vivo, I.; Liu, Y.; Han, J.; Prescott, J.; Hunter, D. J.; Rimm, E. B. Associations between diet, lifestyle factors, and telomere length in women. *Am J Clin Nutr* **91**:1273-1280; 2010.
- [38] Sun, Q.; Shi, L.; Prescott, J.; Chiuve, S. E.; Hu, F. B.; De Vivo, I.; Stampfer, M. J.; Franks, P. W.; Manson, J. E.; Rexrode, K. M. Healthy lifestyle and leukocyte telomere length in U.S. women. *PLoS One* **7**:e38374; 2012.
- [39] Crous-Bou, M.; Fung, T. T.; Prescott, J.; Julin, B.; Du, M.; Sun, Q.; Rexrode, K. M.; Hu, F. B.; De Vivo, I. Mediterranean diet and telomere length in Nurses' Health Study: population based cohort study. *BMJ* 349:g6674; 2014.
- [40] Latifovic, L.; Peacock, S. D.; Massey, T. E.; King, W. D. The Influence of Alcohol Consumption, Cigarette Smoking, and Physical Activity on Leukocyte Telomere Length. *Cancer Epidemiol Biomarkers Prev* 25:374-380; 2016.
- [41] Finkel, T.; Holbrook, N. J. Oxidants, oxidative stress and the biology of ageing. *Nature* **408**:239-247; 2000.
- [42] Harrison, D.; Griendling, K. K.; Landmesser, U.; Hornig, B.; Drexler, H. Role of oxidative stress in atherosclerosis. *Am. J. Cardiol* **91:**7A-11A; 2003.
- [43] El Assar, M.; Angulo, J.; Carnicero, J. A.; Walter, S.; Garcia-Garcia, F. J.; Lopez-Hernandez, E.; Sanchez-Puelles, J. M.; Rodriguez-Manas, L. Frailty Is Associated With Lower Expression of Genes Involved in Cellular Response to Stress: Results From the Toledo Study for Healthy Aging. J Am Med Dir Assoc 18:734 e731-734 e737; 2017.
- [44] Demissie, S.; Levy, D.; Benjamin, E. J.; Cupples, L. A.; Gardner, J. P.; Herbert, A.; Kimura, M.; Larson, M. G.; Meigs, J. B.; Keaney, J. F.; Aviv, A. Insulin resistance, oxidative stress, hypertension, and leukocyte telomere length in men from the Framingham Heart Study. *Aging Cell* 5:325-330; 2006.
- [45] Sampson, M. J.; Winterbone, M. S.; Hughes, J. C.; Dozio, N.; Hughes, D. A. Monocyte telomere shortening and oxidative DNA damage in type 2 diabetes. *Diabetes Care* **29**:283-289; 2006.

- [46] Salpea, K. D.; Talmud, P. J.; Cooper, J. A.; Maubaret, C. G.; Stephens, J. W.; Abelak, K.; Humphries, S. E. Association of telomere length with type 2 diabetes, oxidative stress and UCP2 gene variation. *Atherosclerosis* **209**:42-50; 2010.
- [47] Watfa, G.; Dragonas, C.; Brosche, T.; Dittrich, R.; Sieber, C. C.; Alecu, C.; Benetos, A.; Nzietchueng, R. Study of telomere length and different markers of oxidative stress in patients with Parkinson's disease. J Nutr Health Aging 15:277-281; 2011.
- [48] Sen, A.; Marsche, G.; Freudenberger, P.; Schallert, M.; Toeglhofer, A. M.; Nagl, C.; Schmidt, R.; Launer, L. J.; Schmidt, H. Association between higher plasma lutein, zeaxanthin, and vitamin C concentrations and longer telomere length: results of the Austrian Stroke Prevention Study. J Am Geriatr Soc 62:222-229; 2014.
- [49] Starr, J. M.; Shiels, P. G.; Harris, S. E.; Pattie, A.; Pearce, M. S.; Relton, C. L.; Deary, I. J.
 Oxidative stress, telomere length and biomarkers of physical aging in a cohort aged 79 years from the 1932 Scottish Mental Survey. *Mech Ageing Dev* 129:745-751; 2008.
- [50] Shen, J.; Gammon, M. D.; Terry, M. B.; Wang, Q.; Bradshaw, P.; Teitelbaum, S. L.; Neugut, A. I.; Santella, R. M. Telomere length, oxidative damage, antioxidants and breast cancer risk. *Int J Cancer* **124**:1637-1643; 2009.
- [51] Corina, A.; Rangel-Zuniga, O. A.; Jimenez-Lucena, R.; Alcala-Diaz, J. F.; Quintana-Navarro, G.; Yubero-Serrano, E. M.; Lopez-Moreno, J.; Delgado-Lista, J.; Tinahones, F.; Ordovas, J. M.; Lopez-Miranda, J.; Perez-Martinez, P. Low Intake of Vitamin E Accelerates Cellular Aging in Patients With Established Cardiovascular Disease: The CORDIOPREV Study. J Gerontol A Biol Sci Med Sci 74:770-777; 2019.
- [52] Zhou, M.; Zhu, L.; Cui, X.; Feng, L.; Zhao, X.; He, S.; Ping, F.; Li, W.; Li, Y. Influence of diet on leukocyte telomere length, markers of inflammation and oxidative stress in individuals with varied glucose tolerance: a Chinese population study. *Nutr J* 15:39; 2016.
- [53] Margaritis, M.; Sanna, F.; Lazaros, G.; Akoumianakis, I.; Patel, S.; Antonopoulos, A. S.; Duke, C.; Herdman, L.; Psarros, C.; Oikonomou, E. K.; Shirodaria, C.; Petrou, M.; Sayeed, R.; Krasopoulos, G.; Lee, R.; Tousoulis, D.; Channon, K. M.; Antoniades, C. Predictive value of telomere length on outcome following acute myocardial infarction: evidence for contrasting effects of vascular vs. blood oxidative stress. *Eur Heart J* 38:3094-3104; 2017.
- [54] Lin, J.; Epel, E.; Blackburn, E. Telomeres and lifestyle factors: roles in cellular aging. *Mutat Res* 730:85-89; 2012.
- [55] Mundstock, E.; Zatti, H.; Louzada, F. M.; Oliveira, S. G.; Guma, F. T.; Paris, M. M.; Rueda, A. B.; Machado, D. G.; Stein, R. T.; Jones, M. H.; Sarria, E. E.; Barbe-Tuana, F. M.; Mattiello, R. Effects of physical activity in telomere length: Systematic review and meta-analysis. *Ageing Res Rev* 22:72-80; 2015.
- [56] Mundstock, E.; Sarria, E. E.; Zatti, H.; Mattos Louzada, F.; Kich Grun, L.; Herbert Jones, M.;
 Guma, F. T.; Mazzola In Memoriam, J.; Epifanio, M.; Stein, R. T.; Barbe-Tuana, F. M.; Mattiello,
 R. Effect of obesity on telomere length: Systematic review and meta-analysis. *Obesity (Silver Spring)* 23:2165-2174; 2015.
- [57] Mathur, M. B.; Epel, E.; Kind, S.; Desai, M.; Parks, C. G.; Sandler, D. P.; Khazeni, N. Perceived stress and telomere length: A systematic review, meta-analysis, and methodologic considerations for advancing the field. *Brain Behav Immun* 54:158-169; 2016.
- [58] Freitas-Simoes, T. M.; Ros, E.; Sala-Vila, A. Nutrients, foods, dietary patterns and telomere length: Update of epidemiological studies and randomized trials. *Metabolism* 65:406-415; 2016.
- [59] Arsenis, N. C.; You, T.; Ogawa, E. F.; Tinsley, G. M.; Zuo, L. Physical activity and telomere length: Impact of aging and potential mechanisms of action. *Oncotarget* **8**:45008-45019; 2017.
- [60] Davinelli, S.; Trichopoulou, A.; Corbi, G.; De Vivo, I.; Scapagnini, G. The potential nutrigeroprotective role of Mediterranean diet and its functional components on telomere length dynamics. *Ageing Res Rev* **49:**1-10; 2019.

- [61] Ornish, D.; Lin, J.; Chan, J. M.; Epel, E.; Kemp, C.; Weidner, G.; Marlin, R.; Frenda, S. J.; Magbanua, M. J. M.; Daubenmier, J.; Estay, I.; Hills, N. K.; Chainani-Wu, N.; Carroll, P. R.; Blackburn, E. H. Effect of comprehensive lifestyle changes on telomerase activity and telomere length in men with biopsy-proven low-risk prostate cancer: 5-year follow-up of a descriptive pilot study. *Lancet Oncol* 14:1112-1120; 2013.
- [62] Kiecolt-Glaser, J. K.; Epel, E. S.; Belury, M. A.; Andridge, R.; Lin, J.; Glaser, R.; Malarkey, W. B.; Hwang, B. S.; Blackburn, E. Omega-3 fatty acids, oxidative stress, and leukocyte telomere length: A randomized controlled trial. *Brain Behav Immun* 28:16-24; 2013.
- [63] O'Callaghan, N.; Parletta, N.; Milte, C. M.; Benassi-Evans, B.; Fenech, M.; Howe, P. R. Telomere shortening in elderly individuals with mild cognitive impairment may be attenuated with omega-3 fatty acid supplementation: a randomized controlled pilot study. *Nutrition* **30**:489-491; 2014.
- [64] Shin, Y. A.; Lee, J. H.; Song, W.; Jun, T. W. Exercise training improves the antioxidant enzyme activity with no changes of telomere length. *Mech Ageing Dev* **129:2**54-260; 2008.
- [65] Dimauro, I.; Scalabrin, M.; Fantini, C.; Grazioli, E.; Beltran Valls, M. R.; Mercatelli, N.; Parisi, A.; Sabatini, S.; Di Luigi, L.; Caporossi, D. Resistance training and redox homeostasis: Correlation with age-associated genomic changes. *Redox Biol* **10**:34-44; 2016.
- [66] Sjogren, P.; Fisher, R.; Kallings, L.; Svenson, U.; Roos, G.; Hellenius, M. L. Stand up for health-avoiding sedentary behaviour might lengthen your telomeres: secondary outcomes from a physical activity RCT in older people. *Br J Sports Med* **48**:1407-1409; 2014.
- [67] Puterman, E.; Weiss, J.; Lin, J.; Schilf, S.; Slusher, A. L.; Johansen, K. L.; Epel, E. S. Aerobic exercise lengthens telomeres and reduces stress in family caregivers: A randomized controlled trial - Curt Richter Award Paper 2018. *Psychoneuroendocrinology* **98**:245-252; 2018.
- [68] Werner, C.; Furster, T.; Widmann, T.; Poss, J.; Roggia, C.; Hanhoun, M.; Scharhag, J.; Buchner, N.; Meyer, T.; Kindermann, W.; Haendeler, J.; Bohm, M.; Laufs, U. Physical exercise prevents cellular senescence in circulating leukocytes and in the vessel wall. *Circulation* **120**:2438-2447; 2009.
- [69] Sanft, T.; Usiskin, I.; Harrigan, M.; Cartmel, B.; Lu, L.; Li, F. Y.; Zhou, Y.; Chagpar, A.; Ferrucci, L. M.; Pusztai, L.; Irwin, M. L. Randomized controlled trial of weight loss versus usual care on telomere length in women with breast cancer: the lifestyle, exercise, and nutrition (LEAN) study. *Breast Cancer Res Treat* **172**:105-112; 2018.
- [70] Wang, X.; Sundquist, K.; Hedelius, A.; Palmer, K.; Memon, A. A.; Sundquist, J. Leukocyte telomere length and depression, anxiety and stress and adjustment disorders in primary health care patients. *BMC Psychiatry* **17**:148; 2017.
- [71] Carlson, L. E.; Beattie, T. L.; Giese-Davis, J.; Faris, P.; Tamagawa, R.; Fick, L. J.; Degelman, E. S.; Speca, M. Mindfulness-based cancer recovery and supportive-expressive therapy maintain telomere length relative to controls in distressed breast cancer survivors. *Cancer* **121**:476-484; 2015.
- [72] Puhlmann, L. M. C.; Valk, S. L.; Engert, V.; Bernhardt, B. C.; Lin, J.; Epel, E. S.; Vrticka, P.; Singer, T. Association of Short-term Change in Leukocyte Telomere Length With Cortical Thickness and Outcomes of Mental Training Among Healthy Adults: A Randomized Clinical Trial. JAMA Netw Open 2:e199687; 2019.
- [73] Freitas-Simoes, T. M.; Cofan, M.; Blasco, M. A.; Soberon, N.; Foronda, M.; Serra-Mir, M.; Roth,
 I.; Valls-Pedret, C.; Domenech, M.; Ponferrada-Ariza, E.; Calvo, C.; Rajaram, S.; Sabate, J.; Ros,
 E.; Sala-Vila, A. Walnut Consumption for Two Years and Leukocyte Telomere Attrition in
 Mediterranean Elders: Results of a Randomized Controlled Trial. *Nutrients* 10; 2018.
- [74] Garcia-Calzon, S.; Martinez-Gonzalez, M. A.; Razquin, C.; Corella, D.; Salas-Salvado, J.; Martinez, J. A.; Zalba, G.; Marti, A. Pro12Ala polymorphism of the PPARgamma2 gene interacts with a mediterranean diet to prevent telomere shortening in the PREDIMED-NAVARRA randomized trial. *Circ Cardiovasc Genet* 8:91-99; 2015.

- [75] Garcia-Calzon, S.; Martinez-Gonzalez, M. A.; Razquin, C.; Aros, F.; Lapetra, J.; Martinez, J. A.; Zalba, G.; Marti, A. Mediterranean diet and telomere length in high cardiovascular risk subjects from the PREDIMED-NAVARRA study. *Clin Nutr* **35**:1399-1405; 2016.
- [76] Friedenreich, C. M.; Wang, Q.; Ting, N. S.; Brenner, D. R.; Conroy, S. M.; McIntyre, J. B.; Mickle, A.; Courneya, K. S.; Beattie, T. Effect of a 12-month exercise intervention on leukocyte telomere length: Results from the ALPHA Trial. *Cancer Epidemiol* 56:67-74; 2018.
- [77] Mason, C.; Risques, R. A.; Xiao, L.; Duggan, C. R.; Imayama, I.; Campbell, K. L.; Kong, A.; Foster-Schubert, K. E.; Wang, C. Y.; Alfano, C. M.; Blackburn, G. L.; Rabinovitch, P. S.; McTiernan, A. Independent and combined effects of dietary weight loss and exercise on leukocyte telomere length in postmenopausal women. *Obesity (Silver Spring)* **21**:E549-554; 2013.
- [78] Powers, S. K.; Radak, Z.; Ji, L. L. Exercise-induced oxidative stress: past, present and future. J Physiol 594:5081-5092; 2016.
- [79] Bjorklund, G.; Chirumbolo, S. Role of oxidative stress and antioxidants in daily nutrition and human health. *Nutrition* **33:**311-321; 2017.
- [80] Luo, H.; Chiang, H. H.; Louw, M.; Susanto, A.; Chen, D. Nutrient Sensing and the Oxidative Stress Response. *Trends Endocrinol Metab* **28**:449-460; 2017.
- [81] Himbert, C.; Thompson, H.; Ulrich, C. M. Effects of Intentional Weight Loss on Markers of Oxidative Stress, DNA Repair and Telomere Length - a Systematic Review. *Obes Facts* 10:648-665; 2017.
- [82] Rathore, M.; Abraham, J. Implication of Asana, Pranayama and Meditation on Telomere Stability. *Int J Yoga* **11**:186-193; 2018.
- [83] Nalsen, C.; Vessby, B.; Berglund, L.; Uusitupa, M.; Hermansen, K.; Riccardi, G.; Rivellese, A.; Storlien, L.; Erkkila, A.; Yla-Herttuala, S.; Tapsell, L.; Basu, S. Dietary (n-3) fatty acids reduce plasma F2-isoprostanes but not prostaglandin F2alpha in healthy humans. J Nutr 136:1222-1228; 2006.
- [84] Romieu, I.; Garcia-Esteban, R.; Sunyer, J.; Rios, C.; Alcaraz-Zubeldia, M.; Velasco, S. R.; Holguin,
 F. The effect of supplementation with omega-3 polyunsaturated fatty acids on markers of oxidative stress in elderly exposed to PM(2.5). *Environ Health Perspect* 116:1237-1242; 2008.
- [85] Romero, A.; San Hipolito-Luengo, A.; Villalobos, L. A.; Vallejo, S.; Valencia, I.; Michalska, P.; Pajuelo-Lozano, N.; Sanchez-Perez, I.; Leon, R.; Bartha, J. L.; Sanz, M. J.; Erusalimsky, J. D.; Sanchez-Ferrer, C. F.; Romacho, T.; Peiro, C. The angiotensin-(1-7)/Mas receptor axis protects from endothelial cell senescence via klotho and Nrf2 activation. *Aging Cell* 18:e12913; 2019.
- [86] Razquin, C.; Martinez, J. A.; Martinez-Gonzalez, M. A.; Mitjavila, M. T.; Estruch, R.; Marti, A. A 3 years follow-up of a Mediterranean diet rich in virgin olive oil is associated with high plasma antioxidant capacity and reduced body weight gain. *Eur J Clin Nutr* **63**:1387-1393; 2009.
- [87] Mitjavila, M. T.; Fandos, M.; Salas-Salvado, J.; Covas, M. I.; Borrego, S.; Estruch, R.; Lamuela-Raventos, R.; Corella, D.; Martinez-Gonzalez, M. A.; Sanchez, J. M.; Bullo, M.; Fito, M.; Tormos, C.; Cerda, C.; Casillas, R.; Moreno, J. J.; Iradi, A.; Zaragoza, C.; Chaves, J.; Saez, G. T. The Mediterranean diet improves the systemic lipid and DNA oxidative damage in metabolic syndrome individuals. A randomized, controlled, trial. *Clin Nutr* **32**:172-178; 2013.
- [88] Radak, Z.; Chung, H. Y.; Goto, S. Systemic adaptation to oxidative challenge induced by regular exercise. *Free radical biology & medicine* **44**:153-159; 2008.
- [89] Werner, C. M.; Hecksteden, A.; Morsch, A.; Zundler, J.; Wegmann, M.; Kratzsch, J.; Thiery, J.; Hohl, M.; Bittenbring, J. T.; Neumann, F.; Bohm, M.; Meyer, T.; Laufs, U. Differential effects of endurance, interval, and resistance training on telomerase activity and telomere length in a randomized, controlled study. *Eur Heart J* 40:34-46; 2019.
- [90] Wang, C.; Maddick, M.; Miwa, S.; Jurk, D.; Czapiewski, R.; Saretzki, G.; Langie, S. A.; Godschalk, R. W.; Cameron, K.; von Zglinicki, T. Adult-onset, short-term dietary restriction reduces cell senescence in mice. *Aging (Albany NY)* 2:555-566; 2010.

- [91] Vera, E.; Bernardes de Jesus, B.; Foronda, M.; Flores, J. M.; Blasco, M. A. Telomerase reverse transcriptase synergizes with calorie restriction to increase health span and extend mouse longevity. *PLoS One* **8**:e53760; 2013.
- [92] Vergoni, B.; Cornejo, P. J.; Gilleron, J.; Djedaini, M.; Ceppo, F.; Jacquel, A.; Bouget, G.; Ginet, C.; Gonzalez, T.; Maillet, J.; Dhennin, V.; Verbanck, M.; Auberger, P.; Froguel, P.; Tanti, J. F.; Cormont, M. DNA Damage and the Activation of the p53 Pathway Mediate Alterations in Metabolic and Secretory Functions of Adipocytes. *Diabetes* 65:3062-3074; 2016.
- [93] Ravussin, E.; Redman, L. M.; Rochon, J.; Das, S. K.; Fontana, L.; Kraus, W. E.; Romashkan, S.; Williamson, D. A.; Meydani, S. N.; Villareal, D. T.; Smith, S. R.; Stein, R. I.; Scott, T. M.; Stewart, T. M.; Saltzman, E.; Klein, S.; Bhapkar, M.; Martin, C. K.; Gilhooly, C. H.; Holloszy, J. O.; Hadley, E. C.; Roberts, S. B. A 2-Year Randomized Controlled Trial of Human Caloric Restriction: Feasibility and Effects on Predictors of Health Span and Longevity. *J Gerontol A Biol Sci Med Sci* **70**:1097-1104; 2015.
- [94] Epel, E. S.; Prather, A. A. Stress, Telomeres, and Psychopathology: Toward a Deeper Understanding of a Triad of Early Aging. *Annu Rev Clin Psychol* **14**:371-397; 2018.
- [95] Aschbacher, K.; O'Donovan, A.; Wolkowitz, O. M.; Dhabhar, F. S.; Su, Y.; Epel, E. Good stress, bad stress and oxidative stress: insights from anticipatory cortisol reactivity. *Psychoneuroendocrinology* **38**:1698-1708; 2013.
- [96] Choi, J.; Fauce, S. R.; Effros, R. B. Reduced telomerase activity in human T lymphocytes exposed to cortisol. *Brain Behav Immun* **22**:600-605; 2008.
- [97] Steptoe, A.; Hamer, M.; Chida, Y. The effects of acute psychological stress on circulating inflammatory factors in humans: a review and meta-analysis. *Brain Behav Immun* 21:901-912; 2007.
- [98] Maes, M.; Song, C.; Lin, A.; De Jongh, R.; Van Gastel, A.; Kenis, G.; Bosmans, E.; De Meester, I.; Benoy, I.; Neels, H.; Demedts, P.; Janca, A.; Scharpe, S.; Smith, R. S. The effects of psychological stress on humans: increased production of pro-inflammatory cytokines and a Th1-like response in stress-induced anxiety. *Cytokine* 10:313-318; 1998.
- [99] Conklin, Q. A.; Crosswell, A. D.; Saron, C. D.; Epel, E. S. Meditation, stress processes, and telomere biology. *Curr Opin Psychol* **28**:92-101; 2018.
- [100] Krishna, B. H.; Keerthi, G. S.; Kumar, C. K.; Reddy, N. M. Association of leukocyte telomere length with oxidative stress in yoga practitioners. *J Clin Diagn Res* **9**:CC01-03; 2015.
- [101] Tolahunase, M.; Sagar, R.; Dada, R. Erratum to "Impact of Yoga and Meditation on Cellular Aging in Apparently Healthy Individuals: A Prospective, Open-Label Single-Arm Exploratory Study". Oxid Med Cell Longev 2017:2784153; 2017.
- [102] Steenstrup, T.; Hjelmborg, J. V.; Kark, J. D.; Christensen, K.; Aviv, A. The telomere lengthening conundrum--artifact or biology? *Nucleic Acids Res* **41:**e131; 2013.
- [103] Bateson, M.; Nettle, D. The telomere lengthening conundrum it could be biology. Aging Cell 16:312-319; 2017.
- [104] Melk, A.; Tegtbur, U.; Hilfiker-Kleiner, D.; Eberhard, J.; Saretzki, G.; Eulert, C.; Kerling, A.; Nelius, A. K.; Homme, M.; Strunk, D.; Berliner, D.; Rontgen, P.; Kuck, M.; Bauersachs, J.; Hilfiker, A.; Haverich, A.; Bara, C.; Stiesch, M. Improvement of biological age by physical activity. *Int J Cardiol* **176**:1187-1189; 2014.
- [105] Zalli, A.; Carvalho, L. A.; Lin, J.; Hamer, M.; Erusalimsky, J. D.; Blackburn, E. H.; Steptoe, A. Shorter telomeres with high telomerase activity are associated with raised allostatic load and impoverished psychosocial resources. *Proc Natl Acad Sci U S A* **111**:4519-4524; 2014.
- [106] Haendeler, J.; Hoffmann, J.; Diehl, J. F.; Vasa, M.; Spyridopoulos, I.; Zeiher, A. M.; Dimmeler, S. Antioxidants inhibit nuclear export of telomerase reverse transcriptase and delay replicative senescence of endothelial cells. *Circ. Res* 94:768-775; 2004.

Figure Legend

Figure 1: Targeting telomere shortening and senescence by life-style interventions that activate telomerase and/or reduce oxidative stress. The dashed line indicates a possible connection

Journal Prevention

Tissue	Population or disease	Sample size	Age*	Association with oxidative stress-related parameters **	Reference
WBC vs urine/serum	Healthy women caregivers vs controls	19 control 39 carers	38 (20-50)	Negative w/ F ₂ -isoprostane/VitE index (F(1,40)=3.49; <i>P</i> =0.035)	[31]
WBC vs urine	Framingham Heart Study Caucasian men	327	62.2 (40-89)	8-epi-PGF _{2α} (r=-0.16; <i>P</i> =0.005)	[44]
WBC	Caucasian men with T2D vs controls	28 control 21 T2D	61.5 (50-65)	8-oxoguanine (<i>r</i> =–0.55; <i>P</i> =0.018) in T2D only	[45]
WBC vs plasma	Caucasian T2D patients	561	68 (24–92)	TAC (r=0.12, <i>P</i> <0.01)	[46]
WBC vs plasma	Parkinson's disease (PD) vs controls	20 PD 15 control	>65	protein carbonyls (r=–0.544 <i>, P</i> =0.04) in PD only NOT w/SOD or GSH in either control, PD or overall	[47]
WBC vs plasma	Austrian Stroke Prevention Study	786	66 (46-85)	Antioxidant micronutrients: Lutein-zeaxanthin (β = 0.120, <i>P</i> =0.006), Vitamin C β =0.146, <i>P</i> =0.004	[48]
WBC	1921 Lothian Birth Cohort and Newcastle Thousand Family Study	190 LBC 318 NTF	79 50	Oxidative stress-related gene polymorphisms Methionine sulfoxide reductase A and NADH:Ubiquinone dehydrogenase A3 in both cohorts	[49]
WBC vs urine	Women from the Long Island Breast Cancer Study	1061 cases 1108 controls	(20-98)	NOT w/ 15- F_2 isoprostane or 8-OHdG	[50]
WBC vs plasma	Spanish CVD patients of CORDIOPREV study	290	(20-75)	NOT with lipid peroxides, GSH,GSSG, protein carbonyls, nitrotyrosine, NO or GPx activity	[51]
WBC vs serum	Chinese population	556	53.2 (11.7)	Positive w/ serum SOD (<i>P</i> =0.001) NOT w/ glutathione reductase or 8-OHdG	[52]
WBC, plasma saphenous vein (SV), mammary artery (IMA)	Stable CAD patients	128 WBC 32 IMA 24 SV	65.9 (9.6)	Tissue specific with O_2^{-1} production (IMA: r=-0.49, <i>P</i> =0.004; SV: r=-0.52, <i>P</i> =0.01). Weak w/ plasma MDA. NOT w/ plasma SOD or plasma TAC. Influence of <i>CYBA</i> gene (p22 ^{phox}) polymorphisms	[53]

Table 1: Human observational studies exploring the association between telomere length and oxidative stress-related parameters

*Age is indicated as median or mean and/or (range or SD)

** Where data are available the strength of associations are described by correlations coefficients (r), ANCOVA F-statistics or standardized regression cofficients (β)

CAD, coronary artery disease; GSH, Glutathione; GSSG, oxidized glutathione; GPx, Glutathione peroxidase; MDA: malondyaldehyde; NO, nitric oxide; SOD, superoxide dismutase; TAC: total antioxidant capacity; T2D, type 2 diabetes; WBC, white blood cells

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Table 2: Randomised controlled trials of non-drug	interventions to modulate telomere length

Intervention	Population	Age [*]	Study Design	Main findings	Reference
n-3 PUFA supplements	Healthy overweight U.S. subjects	40-85	1.25 g/d (n=40) or 2.5 g/d (n=35) n-3 PUFA vs placebo (n=31); 4m	15% ↓urinary F-2 isoprostanes in PUFA groups $\leftrightarrow \Delta TL$ between groups; association between $\uparrow TL$ and ↓plasma n-6:n-3 PUFA ratio (<i>P</i> =0.02)	[62]
n-3 PUFA supplements	Australian subjects w/ MCI	>65	EPA (n=12) or DHA (n=12) vs LA (n=9); 6m	$\leftrightarrow \Delta TL$ between groups; correlation of RBC ΔDHA with ΔTL in DHA group (r=-0.67, <i>P</i> =0.02)	[63]
Walnut-rich diet	Cognitively healthy Spanish elders	63-79	Walnut rich diet (n=80) vs habitual diet (n=69); 2y	$\leftrightarrow \Delta TL$ between groups but tendency to reduced attrition in walnut diet group	[73]
Mediterranean diet	PREDIMED- NAVARRA high CVD risk Spanish subjects	55-80	MD +olive oil (n=211) or MD+ nuts (n=170) vs control low fat diet (n=140); 5y	Δ TL by high adherence to MD in PPARγ2 gene Ala carriers but not in Pro/Pro carriers (<i>P</i> < 0.001); \leftrightarrow ΔTL in MD + EV olive oil vs control, \uparrow -ΔTL in MD + nuts vs control (<i>P</i> =0.003)	[74, 75]
Aerobic exercise	Obese middle age women	47 (6)	supervised aerobic exercise (EX) (n=8) vs control (n=8); 6m	↔TL in control or EX group; ↔serum MDA or SOD, 个 GPX (<i>P</i> <0.05) in EX group	[64]
Resistance training	Elderly people	70-75	explosive moderate intensity resistance training (EMRT) (n=10) vs control (n=10); 12 w	 ↑ TL in EMRT group vs control group (P<0.05); ↓ MnSOD (P<0.01), ↓ TrxR1 (P<0.01), ↓ MPO (P<0.05) in EMRT group vs baseline 	[65]
Physical activity on prescription	Sedentary overweight older adults	68	individualised low/moderate intensity physical activity (n=24) vs control (n=25), 6m	\leftrightarrow TL exercise vs control; correlation of Δ TL with reduced sitting time in exercise group (r=-0.68, <i>P</i> =0.02)	[66]
Aerobic exercise	Family caregivers with high stress	61 (6)	highly supervised aerobic exercise (n=34) vs control (n=34); 6 m	\leftrightarrow TA in either group; \leftrightarrow Δ TL in control group $\land \Delta$ TL in exercise group (<i>P</i> =0.03)	[67]
Aerobic exercise	Healthy postmenopausal sedentary women	50-74	highly supervised aerobic exercise (n=99) vs usual inactivity (113); 1y	$\leftrightarrow \Delta TL$ between groups	[76]
Various exercise modalities	Healthy sedentary men and women	30-60	aerobic endurance (AET) (n=26), interval (IT) (n=29) or resistance training (RT) (n=34) vs control (n=35);	ΔTA (<i>P</i> <0.01) and ΔTL (<i>P</i> <0.05) in AET or IT ↔ΔTA and ↔ΔTL in RT or controls	[89]

			6m		
Weight loss	Overweight or obese U.S older women	50-75	dietary weight loss (n=118), aerobic exercise (n=117) or diet + exercise (n=117) vs control (n=87); 1y	$\leftrightarrow \Delta TL$ in any intervention vs control; no association between Δ weight loss and ΔTL	[77]
Weight loss	Overweight or obese breast cancer surviving women	58 (8)	diet + exercise (n=93) vs usual care control (n=58), 6m	↔ΔTL between groups	[69]
Mindfulness- based group therapy	Patients with depression, anxiety or stress and adjustment disorders	20-64	mindfulness based group therapy (n=88) vs treatment as usual control (n=89); 8w	↔ TL vs baseline or between groups	[70]
Mindfulness- based group therapy	Breast cancer survivors	55 (9)	8w of mindfulness-based cancer recovery (n=34), 12w of supportive expressive group therapy (n=36) vs control (n=18)	↔TL vs baseline ,but trend effect between combined intervention groups vs control group (F (1,84)=3.82, <i>P</i> =0.054)	[71]
Contemplative mental training	Healthy adults	20-55	mental training (n=222) vs control (n=76); 9 m	\leftrightarrow Δ TL between groups	[72]

*age is indicated as mean (SD) or range

Abbreviations: CVD, cardiovascular disease; DHA, docosahexanoic acid; EPA, eicosapentanoic acid; LA, linolenic acid; MDA, malondyaldehyde; MCI, mild cognitive impairment, PUFA, polyunsaturated fatty acids; SOD, superoxide dismutase; TL, telomere length; RBC, red blood cells;

 \leftrightarrow , no significant difference between groups, \uparrow , increase; \downarrow , decrease

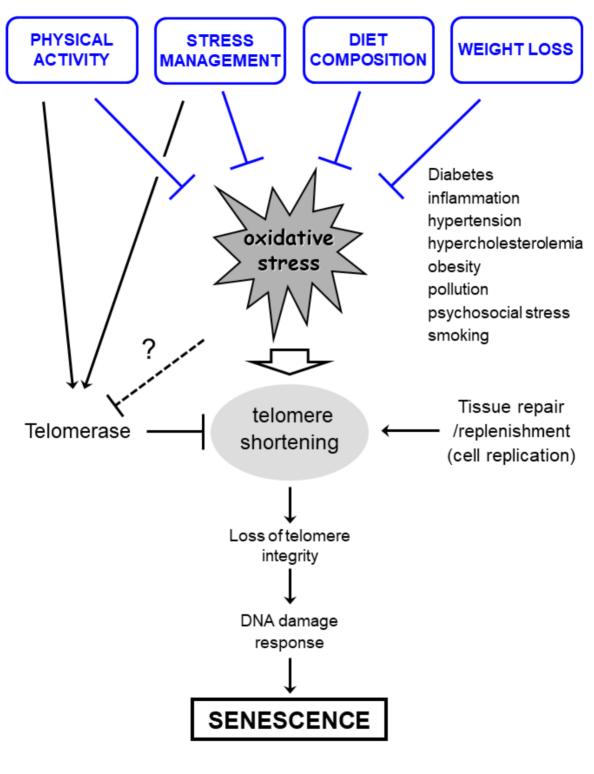


Figure 1

Highlights

- Oxidative stress accelerates telomere shortening resulting in cellular • senescence
- There is evidence linking oxidative stress to telomere shortening in humans •
- Non-drug interventions that curb oxidative stress may reduce telomere shortening