

**Biomarkers of Inflammation and Lung recovery in ECMO patients with Persistent
Pulmonary Hypertension of the Newborn (PPHN): A feasibility study**

Paolo Pais¹, MD, Simon Robinson², MD, Gavin Majitha-Beet³, BSc, Attilio Lotto^{1,4}, MD FRCS
(CTh), Tracy Kumar¹, BSc, Claire Westrope², MD, Nikol Sullo^{1,7}, PhD, Bryony Eagle
Hemming¹, BSc, Lathishia Joel-David¹, BSc, Maria JnTala³, MSc, Claudio Corazzari^{1,5}, MD,
Lorenzo Grazioli^{2,6}, MD, Dawn Smallwood^{1,8}, PhD, Gavin J Murphy¹, MD FRCS (CTh),
Florence Y Lai¹, MPhil, and Marcin J Woźniak¹, PhD.

¹Leicester NIHR Biomedical Research Unit and Department of Cardiovascular Sciences,
University of Leicester, Glenfield Hospital, Leicester LE3 9QP, UK.

²Paediatric Intensive Care, University Hospitals Leicester NHS Trust, Glenfield Hospital,
Leicester LE3 9QP, UK

³Clinical Perfusion, University Hospitals Leicester NHS Trust, Glenfield Hospital, Leicester
LE3 9QP, UK.

⁴Department of Congenital Cardiac Surgery, Alder Hey Children's Hospital, Eaton Road,
Liverpool L12 2AP, UK

⁵Cardiac Surgery Unit, Insubria University, Varese, Italy.

⁶ASST Papa Giovanni XXIII, 24127 Bergamo, Italy

⁷University of Nottingham, Royal Derby Hospital, Derby, DE22 3DT

⁸ School of Allied Health Sciences, De Montfort University, Leicester LE1 9BH

Address for correspondence: Marcin J Woźniak, Leicester NIHR Biomedical Research
Unit and Department of cardiovascular Sciences, University of Leicester, Glenfield Hospital,
Leicester LE3 9QP, Email mw299@leicester.ac.uk. Telephone +44116 2583028

Financial support: This work was supported by the Heart Link Charity, Leicester NIHR
Biomedical Research Centre, British Heart Foundation RG/13/6/29947

Keywords: Extracorporeal membrane oxygenation, Hemolysis, Inflammation, Persistent
Fetal Circulation Syndrome, Cytokines.

ClinicalTrials.gov Identifier: NCT02940327

Copyright form disclosure: Drs. Kumar, Hemming, Joel-David, Smallwood, Murphy, and Wozniak received support for article research from the British Heart Foundation. Drs. Hemming, Joel-David, and Wozniak's institutions received funding from HeartLink, British Heart Foundation grant no RG/13/6/29947, and Leicester NIHR Biomedical Research Centre. Dr. Smallwood received funding from HeartLink. Dr. Lai disclosed work for hire. The remaining authors have disclosed that they do not have any potential conflicts of interest.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Abstract

Objective: Extra-corporeal membrane oxygenation (ECMO) is a treatment for Persistent Pulmonary Hypertension of the Newborn (PPHN) with high mortality. Hypothesis: the ECMO circuit results in inflammatory responses that mitigate against successful weaning.

Design: Single centre prospective observational feasibility study.

Setting: Paediatric Intensive Care Unit

Patients: Twenty-four neonates requiring ECMO support for PPHN.

Interventions: None

Measurements and Main Results: The reference outcome was death or more than 7 days of ECMO support. Other outcomes included serial measures of plasma-free haemoglobin and markers of its metabolism, leucocyte, platelet and endothelial activation, and biomarkers of inflammation. Of 24 participants recruited between February 2016 and June 2017, 10 died or required prolonged ECMO support. These patients were sicker at baseline with higher levels of plasma-free haemoglobin within 12 hours of cannulation (geometric mean ratio 1.92, 95% confidence intervals 1.00-3.67, $p=0.050$) but not thereafter, versus those requiring <7 days ECMO. Serum haptoglobin concentrations were significantly elevated in both groups. Patients who died or required prolonged ECMO support demonstrated elevated levels of platelet-leucocyte aggregation, but decreased concentrations of mediators of the inflammatory response: interleukin-8, C-reactive protein and Tumour Necrosis Factor α .

Conclusion: Clinical status at baseline and not levels of plasma-free haemoglobin or the systemic inflammatory response may determine the requirement for prolonged ECMO support in neonates.

Introduction

1
2
3 Persistent Pulmonary Hypertension of the Newborn (PPHN) complicates 1–2 per 1000 live
4 births, primarily affecting full term and late preterm babies.¹ Some neonates with PPHN and
5 severe respiratory failure require Extracorporeal Membrane Oxygenation (ECMO) support.
6
7 This reduces mortality rates from >80% to 10-20%.² However, ECMO remains an invasive
8 form of life support. Some 74% of neonates with PPHN develop complications while on
9 ECMO with cardiovascular and renal complications amongst the most common.³ The ECMO
10 circuit causes haemolysis and systemic inflammation along with leukocyte and platelet
11 activation that can lead to organ injury and prolonged cannulation times.⁴⁻⁷ Severe
12 haemolysis and rise in plasma-free haemoglobin (PFH) have been reported in over 10% of
13 neonatal ECMO cases.⁸⁻¹¹ PFH is highly reactive and can oxidize multiple species including
14 proteins and membrane lipids causing inflammation, endothelial injury and organ
15 dysfunction.¹²⁻¹⁷ Elevated PFH may also contribute to persistent pulmonary hypertension.¹⁸⁻²¹
16
17 These observations led us to hypothesise that damage to red blood cells by the exposure to
18 the ECMO circuit results in inflammatory responses that mitigate against successful weaning
19 from ECMO. As a first step to testing this hypothesis, we evaluated the feasibility of
20 measuring PFH and its metabolism, cellular activation processes, and inflammation, in
21 neonates on ECMO support.
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Materials and Methods

Study Population

The Markers of Inflammation and Lung recovery in ECMO patients for Persistent Pulmonary Hypertension of the Newborn (PPHN): MI-ECMO study, was a prospective, single-centre observational feasibility study. The study was reviewed by the 'North East – York' Research Ethics Committee on 11th December 2015, and granted a favourable ethical opinion on 23rd December 2015. Neonates (<30 days of age) having a diagnosis of PPHN and requiring ECMO support were eligible for inclusion. We excluded neonates with PPHN caused by a congenital heart pathology or those requiring ECMO for a congenital heart disease. We recruited consecutive patients referred to the Glenfield Hospital (a regional ECMO centre in the UK) according to the prevalent ECMO referral system who met the inclusion/exclusion criteria. Emergency assents were obtained from patients' parents/legal guardians within 12 hours of cannulation and full consents were obtained within 24 hours. Study withdrawal occurred at the parent/guardian's request or if the patient was found not to meet all the inclusion criteria. In the event of withdrawal, we requested that all patient's data and tissues collected until that time were retained for analysis. The study had ethical approval (REC reference 15/NE/0398). The trial protocol was registered (NCT02940327) and is attached as a digital supplement. The corresponding author (MJW) attests to the validity of the data, its analysis, and interpretations, on behalf of all the co-authors.

Outcome measures

Outcomes of interest included markers of haemoglobin and iron metabolism (plasma free haemoglobin, bilirubin, haptoglobin, ferritin, transferrin, total and catalytic iron), markers of platelet, leukocyte and endothelial cell activation (please see Research procedures below) and markers of the systemic inflammatory and acute phase response (interleukin (IL-) 1 β , IL-6, IL-8, IL-18, Tumour Necrosis Factor α (TNF α), Monocyte chemoattractant protein 1 (MCP1), C-reactive protein (CRP), fibrinogen) were measured in arterial blood at four time points after the initiation of ECMO - at 12, 24, 48 and 72 hours and at weaning 24 hours after

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

decannulation or immediately prior to termination of ECMO support in case of death or treatment withdrawal.

Research Procedures

PFH levels were estimated by measuring absorbance at 415, 450 and 700 nm of citrated plasma samples using EnSpire spectrophotometer (PerkinElmer, Waltham, USA) and calculated as described in Fairbanks et al.²² Total iron was measured in citrated plasma samples using Iron Assay Kit (Abcam, Cambridge, UK). Catalytic iron was estimated using bleomycin method, as described in Burkit et al.²³ Platelet and leukocyte activation, and leukocyte-platelet interactions were measured using flow cytometry (CyAn ADP, Beckman Coulter, Pasadena, USA) in citrated blood with specific fluorescently-labelled antibodies: PAC-1 (against activated GPIIb/IIIa, BD Biosciences, Abingdon, Oxford, UK), CD41 (Affymetrix, Santa Clara, USA) and CD62P (Abcam, Cambridge, UK) for platelets; CD64, CD163, CD11b (Affymetrix) for leukocytes; and CD14, CD16, CD41 (Affymetrix) for leukocyte-platelet interactions. Cytokines (IL-1 β , 6, 8, 18, TNFa) and soluble ICAM1, as a marker of endothelial activation, were measured in citrated plasma using MAGPIX platform (Luminex Corporation, Austin, USA) and magnetic Luminex assays (BioTechne, Abingdon, UK). Troponin (Enzo Biochem, Farmingdale, USA), Haptoglobin (Abcam), ferritin and transferrin (ThermoFisher Scientific, Waltham, USA), were measured in plasma using ELISA DS2 Dynex platform (Worthing, UK). Levels of bilirubin, alkaline phosphatase, pH, pO₂, pCO₂, SaO₂ and lactate were collected from routine hospital diagnostic procedures.

Clinical procedures

Pre-ECMO care and ECMO indication: Eligible patients received standard care during the pre-ECMO period, according to the different respiratory failure aetiology and protocols from referral hospitals. The data on the respiratory protocols at the referring hospitals were not collected.

The ECMO circuit and cannulation: The ECMO circuit consisted of vascular access cannulas (Biomedicus, Medtronic, Dublin, Ireland), polyvinyl chloride tubing for blood extracorporeal circulation (Leicester Neonatal circuit, Chalice Medical, Worksop, UK), a pump console (2nd generation Centrimag, Thoratec Corporation, Pleassanton, USA), a centrifugal pump (Centrimag, Thoratec Corporation) and head (PediVAS Blood pump., Thoratec Corporation) and an oxygenator (Paragon infant PMP oxygenator, Chalice Medical). Cannulation was always performed using an open surgical approach. VA (veno-arterial) or VV (veno-venous) cannulation was chosen based on the clinical status of the patient and the availability of the correct material by the attending clinician. VA ECMO involved surgical cannulation of the right common carotid artery and internal jugular vein, with the tip of the venous cannula 10fg Biomedicus (Medtronic, Minneapolis, USA) advanced into the right atrium and the arterial catheter positioned at the junction of the right common carotid artery and aortic arch. VV ECMO involved insertion of a 13F (OriGen Biomedical, Austin, USA) arterial / venous cannula advanced to the right atrium via the right internal jugular vein.

ECMO management: Established Glenfield ECMO centre protocols were used. In VA-ECMO flows were typically maintained at about $100\text{-}120\text{ mL kg}^{-1}\text{ min}^{-1}$ to achieve normal saturation ($\text{SvO}_2 > 65\%$), with a controlled pCO_2 to achieve a normal pH. The target of arterial blood oxygen saturation during VV ECMO was $>85\%$ with a venous oxygen saturation $> 60\%$ and low lactates with targeted flow rates of $80\text{ mL kg}^{-1}\text{ min}^{-1}$. Sweep gas was usually maintained at a FiO_2 100% and regulated on the pCO_2 basis; the usual starting ECMO flow:sweep gas ratio was 1:1.

Ventilator Management: During ECMO, patients were commonly ventilated in a “lung rest” setting defined as a respiratory rate of 10 breaths minute^{-1} , PEEP (positive end expiratory pressure) of $10\text{ cmH}_2\text{O}$ and pressure over PEEP $10\text{ cmH}_2\text{O}$ with FiO_2 40% for VA ECMO and 30% for VV ECMO. These parameters were changed according to clinical status.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Haemodynamic support: During VA-ECMO patients received haemodynamic support in addition to the respiratory support, and the usual inotropes requirement was reduced or null. During VV-ECMO inotropic or vasoactive support was used to maintain an adequate perfusion and cardiac output defined by general practice targets: Mean Arterial Blood Pressure (MABP) > 40 mmHg, lactate < 2 mmol L⁻¹, central venous saturation > 60% with evidence of end organ perfusion e.g. urine output.

Anaesthesia induction: The anaesthetic technique used a standard unit protocol that while titrated to effect, is typically Ketamine 2 mg kg⁻¹ for sedation and Atracurium 1 mg kg⁻¹ as a neuromuscular blocking agent. Underlying sedation was modified according to the requirements of the patient and deviations from the protocol were recorded. Details of concomitant medications and therapy including generic drug name, dose, route, frequency, duration and indication were recorded.

Fluid Management: Target urine output was > 0.5 mL kg⁻¹ h⁻¹. This was maintained using fluid boluses or diuretics at the discretion of the attending clinician. In some circumstances the clinician can choose CRRT (continuous renal replacement therapy) according the local protocols and fluid balance.

Transfusion: The standard unit protocol is transfusion with 10 mL kg⁻¹ of allogenic red cells if the haemoglobin < 13g dL⁻¹ and SvO₂ < 65% or in the presence of bleeding. Non red cell components were administered according to standard unit protocols, with the indication, volume and timing of their administration recorded.

Weaning: Clinician and patient clinical status guided ECMO weaning. VA ECMO was typically with “retrograde flow”, in this case the flow in the cannula is reversed inside the system using the ECMO as a brake and not as an engine. With this kind of procedure, the observational weaning time is prolonged with less clotting risk for the circuit. In VV ECMO, the support blood flow is maintained but the sweep gas is unplugged, the patient is fully ventilated to assess the lung function. Failure to wean the patient results in restarting ECMO support or treatment withdrawal.

Statistical analysis

1
2
3 Continuous data are presented as the mean (standard deviation, SD) or median
4
5 (interquartile range, IQR) as appropriate. Categorical data are presented with frequency and
6
7 percentage. Duration on ECMO was recorded in terms of hours and converted to days by
8
9 dividing by 24. Patients were classified as prolonged stay if they stayed on ECMO for at
10
11 least 7 days or if they did not survive to discharge (patients who died cannulated or after
12
13 weaning). Patients not experiencing a prolonged stay were surviving patients who stayed on
14
15 ECMO for less than 7 days. Using the linear mixed effects model with patients as random
16
17 effect, we compared the trajectory of outcome measures between those experiencing a
18
19 prolonged stay on ECMO and those not. Separate models were built for each of the outcome
20
21 measures. All models included the main effects of group (prolonged stay or not) and time (5
22
23 time points) together with the group—time interaction term. A significant interaction term
24
25 indicates that the two groups exhibit different trajectories of the outcome or the effect of
26
27 groups depends on the time points. Models with a significant group effect but insignificant
28
29 interaction indicates the effect of groups are similar over the time points. Interaction or group
30
31 effects were considered significant if $p\text{-value} < 0.05$. For laboratory measurements and
32
33 ECMO and blood gas parameters where data prior to ECMO or at start of ECMO were
34
35 measured, the models would be adjusted for these baseline values. Outcome data analysed
36
37 on a logarithmic scale would be transformed back to the original scale after the analysis and
38
39 results reported as geometric mean. A p-value of <0.05 was considered statistical
40
41 significance. No adjustment for p-values has been made for multiple comparisons as this
42
43 was a feasibility study. All analyses were performed with SAS version 9.4 (SAS Institute Inc.,
44
45 Cary, NC, USA).
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Results

Study cohort

Between February 2016 and June 2017, we screened 52 neonates referred to the Glenfield Hospital for ECMO support, of which 24 with PPHN were eligible. Informed full consent was obtained from parents in every case. **Figure 1** and **Supplemental Table 1** show participant flow and completeness of data for individual endpoints. The reference outcome (7+ days on ECMO) occurred in 10 (41.7%) participants, two of which died while cannulated and 1 who died 6.3 days after weaning. A further 14 neonates with PPHN (58.3%) stayed on ECMO for <7 days and survived to discharge (<7 days). There was no difference between the two groups with respect to age (median 2 days), gestational age (median 40 weeks), male:female ratio, or birth weight (median 3.2kg) (**Table 1**). The aetiology of PPHN differed significantly between the two groups; congenital diaphragmatic hernia accounted for 8/10 (80%) in the 7+ days group and meconium aspiration syndrome (MAS) accounted for 12/14 (85%) of the <7 days group (**Table 1**). This was reflected by differences in ventilation strategies at baseline; 10/10 (100%) in the 7+ days group underwent HFOV whereas 7/14 (50%) of the <7 days group underwent SIMV (**Table 1**).

Baseline Clinical Status

A greater proportion of participants in the 7+ days group were haemodynamically unstable prior to cannulation; n=7/10 (70%) versus n=5/14 (36%, **Table 2**). Arterial pCO₂ and serum lactate concentrations were higher in the 7+ days group, and arterial SaO₂ was lower. None of these changes reached conventional statistical significance (P<0.05). Baseline values for PIM2 (mortality risk) or PMODS (organ dysfunction) scores at ICU admission were not significantly different between the two groups. At baseline participants in the <7 days group were less acidotic; mean PH 7.3, SD 0.08 versus 7.3, SD 0.12, $p_{Baseline}=0.018$, but had significantly higher levels of CRP (median 39 mg L⁻¹, IQR 11-18 versus 6 mg L⁻¹, IQR 5-15, $p_{Baseline}=0.019$) and alkaline phosphatase (median 198 iu L⁻¹, IQR 113 – 613, $p_{Baseline}=0.03$ vs 107 iu L⁻¹, IQR 66 – 131) relative to the 7+ days group.

ECMO Course

There was no difference between the groups with respect to use of VA (n=21) versus VV ECMO (n=3), choice of prime (blood/crystalloid), pump flows, or sweep (**Tables 1 & 2**). Participants in the <7 days group spent a median of 80 hours (IQR 59-91), and patients in the 7+ days group spent a median of 292 (186-360) hours on ECMO. Mean arterial blood pressure was similar between the groups for the duration of ECMO support (**Supplemental Table**). SaO₂ and PaO₂ were higher (all participants received FiO₂ 1.0, **Supplemental Figure 1A – 2B, Supplemental Table**) and PaCO₂ values were lower in the <7 days group (**Supplemental Figure 1C, Supplemental Table**). There was no difference between the groups for haemoglobin levels and haematocrit, both decreased after cannulation and remained unchanged until discharge (**Supplemental Table**). Serum lactate concentration was higher (not significantly) in the 7+ days group prior to cannulation. The difference was not sustained during ECMO and the concentrations gradually dropped during the course of ECMO ($p_{Time} < 0.001$). The 7+ days group experienced a greater number of adverse events (**Table 3**); median 4 (IQR 3-5) versus median 2 (IQR 0-2), $p=0.001$, received a larger volume of allogenic red cells, and clotting of the ECMO circuit (6 (60%) versus 6 (25%), $p=0.002$, **Table 3**). Patients in both groups had their PaO₂/FiO₂ ratio improved post weaning to 208 (SD 86) for 7+ days group and 212 (SD 69) for <7 days group and the difference between the two groups was not statistically significant ($p=0.944$; **Supplemental Table**).

Inflammation

Haemoglobin metabolism There was a significant interaction between group and time for PFH concentrations ($p_{group*time}=0.02$). Neonates requiring >7 days ECMO had higher levels at 12 hours post cannulation ($p_{12hr}=0.05$) but not thereafter (**Supplemental Figure 1D, Supplemental Table**). The >7 day group also had higher levels of serum bilirubin at 48 hours ($p_{48hr}=0.03$, **Supplemental Figure 1E, Supplemental Table**). There was no significant difference between groups with respect to plasma haptoglobin (**Supplemental Figure 1F, Supplemental Table**) although levels were high in both groups and gradually

1 increased during the ECMO course ($p_{Time}<0.001$). There was no difference between the
2 groups for catalytic iron, ferritin, transferrin and total iron concentrations (**Supplemental**
3 **Table**).

4
5
6
7 **Platelet, leucocyte and endothelial activation.** Participants in the 7+ days group showed
8 higher levels of platelet-granulocyte and platelet-monocyte aggregates (CD16/CD41 and
9 CD14/CD41, respectively; $p_{group}=0.001$, **Supplemental Figure 1G – H, Supplemental**
10 **Table**), peaking at 24 hr post cannulation in the 7+ days group ($p<0.001$) but not thereafter.
11 For levels of platelet-monocyte aggregates there was also a significant interaction between
12 group and time ($p_{group*time}=0.028$, **Supplemental Table**). Monocyte activation was also higher
13 in the +7 days group ($p_{group}=0.018$), with the greatest difference at 48 hours ($p_{48hr} = 0.02$;
14 **Supplemental Figure 1I, Supplemental Table**). There were no differences between groups
15 for platelets activation measured with P-selectin/CD62P antibodies, CD11b (total leukocyte
16 activation) and ICAM-1 (canonical endothelial cell activation, **Supplemental Table**).

17
18
19
20
21
22 **Inflammation biomarkers.** There was a significant interaction between groups and time for
23 the pro-inflammatory cytokines IL-8 ($p_{group*time}=0.003$, **Figure 2A, Supplemental Table**) and
24 MCP-1 ($p_{group*time}=0.013$, **Figure 2B, Supplemental Table**) as well as the acute phase
25 response biomarkers CRP ($p_{group*time}<0.001$, **Figure 2C and Supplemental Table**) and IL-1b
26 ($p_{group*time}<0.001$, **Figure 2D, Supplemental Table**). For all these markers, participants in the
27 <7 days group had significantly higher plasma concentrations of systemic inflammatory
28 response and acute phase response biomarkers during the first 12 to 48 hours post
29 cannulation versus those in the 7+ days group. TNF α and IL-1 β levels were higher in the <7
30 days ($p_{group}=0.018$ and 0.013, respectively) with the test for interaction between group and
31 time insignificant for TNF α (**Figure 2E, Supplemental Table**). There was no difference
32 between the groups for IL-6, IL-8 and MCP-1, and Fibrinogen (**Supplemental Table**).

Discussion

1
2
3 In this single centre observational feasibility study we observed that neonates with PPHN
4 who required prolonged ECMO support or who died (7+ days group) were more likely to
5 have: 1. Presentation with congenital diaphragmatic hernia. 2. To be sicker at baseline with
6 more severe hypoxia and hypercapnoea during ECMO. 3. A higher number of adverse
7 events. 4. Higher PFH concentrations at 12 hours post cannulation but not thereafter and
8 higher bilirubin concentrations at 48 hours without group differences in other markers of
9 haemoglobin and iron metabolism. 5. Increased platelet (platelet-leukocyte aggregates²⁴)
10 and monocyte activation but no differences in canonical endothelial activation. 6. In an
11 apparent paradox, lower plasma concentrations of biomarkers of the systemic inflammatory
12 response (IL-8, MCP-1) and acute phase response (CRP, TNF α , IL-1 β) within 48 hours of
13 cannulation. These findings did not support our primary hypothesis that the release of PFH
14 by the ECMO circuit drives inflammatory responses that can prolong ECMO support.
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29

30 The study has several strengths. It is the first study to our knowledge that has systematically
31 phenotyped cellular and platelet activation, cytokine responses, and haemoglobin
32 metabolism in a cohort of neonates with PPHN requiring ECMO. The study results are
33 counter to our initial hypotheses but these are supported by the rigorousness of our
34 analytical methods, the high level of completeness of data, and the consistency of our
35 results. A limitation of the study is that we were not able to measure our biomarkers prior to
36 ECMO cannulation. This is because many neonates are placed on ECMO at their referring
37 centre by a retrieval team and prior to assent or consent by legal guardians for sample
38 collection. Stored samples collected at the time of cannulation were not suitable for the
39 inflammation assays used in the study and a window of 12 hours post cannulation was
40 therefore selected for the collection of baseline bloods for these assays. It is possible that
41 the changes measured at this time will have altered dramatically from the pre-cannulation
42 values, although this potential source of bias will have been present in both patient groups.
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58

59 The observational nature of the study means that we cannot demonstrate a causal
60
61
62
63
64
65

1 relationship between the processes that we have measured and clinical outcomes. The
2 small sample size is also a limitation. We cannot conclude that these findings would be
3 replicated in a larger cohort. Nonetheless, our findings were remarkably consistent and
4 highly statistically significant across measures of linked processes, indicating adequate
5 power to detect differences in markers of inflammation even in this limited sample.
6
7
8
9

10
11 The results have clinical application. First, outcomes in participants of the Mi-ECMO study
12 were significantly associated with patient status before cannulation. Earlier use of ECMO
13 support may be indicated in patients with congenital diaphragmatic hernia who were sicker
14 at presentation relative to other groups. Second, patients with higher levels of systemic
15 inflammation and acute phase inflammation were associated with shorter duration of
16 ECMO support. This contrasts findings in adults requiring ECMO support^{25,26} where
17 increased levels of pro-inflammatory cytokines²⁷ are predictive of mortality, but is in
18 agreement with the p-MIVAKI study where very different inflammatory responses were
19 observed in neonates versus older children undergoing cardiac surgery with CPB.²⁸ This
20 may reflect different pathologies at baseline; MAS is primarily an inflammatory condition,
21 whereas CDH is associated with lung hypoplasia and impaired chest wall dynamics, rather
22 than a pathological response to ECMO.⁷ Alternatively, the results may represent the failure
23 of the sicker patients in the 7+ days group to generate an inflammatory response. Third, the
24 cytokine levels observed in the <7 days group were relatively low and were not associated
25 with significant differences in downstream effectors of tissue inflammation IL-6 and IL-18, or
26 measures of myocardial or renal injury between groups. These changes may reflect an
27 inflammatory response that promotes repair rather than a dysregulated response that results
28 in organ injury.²⁹ Finally, the lower levels of inflammatory biomarkers are apparently
29 contradictory to the higher PFH measured in the 7+ days group with levels comparable to
30 those (0.3 mg mL⁻¹) associated with organ injury in other paediatric ECMO cohorts.³⁰ In
31 mitigation, the difference between the groups was transient. In addition, the levels of
32 haptoglobin in both groups were high, and comparable to levels observed in neonates with
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1 sepsis or other inflammatory insults.³¹ Neonates exposed to inflammatory stress undergo a
2 'haptoglobin switch' characterised by significant increases (20-30 fold) in protein expression
3 as part of the acute phase response.³² These changes may precondition neonates against
4 plasma free haem mediated tissue damage. A greater transfusion volume was observed in
5 the 7+ day group. The importance of this finding is unclear; the absolute difference in the
6 volume of red cells transfused was low and this may simply represent the effects of a
7 prolonged stay in a critical care environment on circulating red cell mass in the 7+ day
8 group. There was also no detectable difference in markers of haemolysis after 48 hours
9 suggestive of consumption by the circuit. A final observation is the increased circulating
10 levels of platelet-leukocyte aggregates, a sensitive measure of platelet activation ²⁴ observed
11 in the 7+ days group. PFH is known to activate platelets leading to adverse clinical
12 outcomes,^{10,33,34} however whether these processes are linked in the current study is
13 unproven.

14 **The study design dichotomising the analysis into two groups may have reduced the number**
15 **of significantly associating biomarkers. That provides scope for future post hoc analyses.**
16 **Also, in a larger study, it would be interesting to analyse the outcomes of patients with CDH**
17 **and those with adverse events during ECMO.**

18 **Conclusions** Neonates requiring ECMO support for PPHN attributable to congenital
19 diaphragmatic hernia, with higher levels of acidosis, PFH, platelet and leucocyte
20 aggregation, and lower levels of systemic and acute phase inflammation are likely to require
21 prolonged ECMO support. These findings highlight a knowledge gap with respect to the
22 mechanisms underlying failure to wean in this vulnerable population.

Acknowledgements

The authors are grateful to the staff of the Glenfield Paediatric Intensive Care Unit and ECMO teams for their assistance.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

References

1. Konduri GG, Kim UO: Advances in the diagnosis and management of persistent pulmonary hypertension of the newborn. *Pediatr Clin North Am* 2009; 56: 579-600, Table of Contents
2. Bahrami KR, Van Meurs KP: ECMO for neonatal respiratory failure. *Semin Perinatol* 2005; 29: 15-23
3. Lazar DA, Cass DL, Olutoye OO, et al.: The use of ECMO for persistent pulmonary hypertension of the newborn: a decade of experience. *J Surg Res* 2012; 177: 263-7
4. Fortenberry JD, Bhardwaj V, Niemer P, et al.: Neutrophil and cytokine activation with neonatal extracorporeal membrane oxygenation. *J Pediatr* 1996; 128: 670-8
5. Graulich J, Walzog B, Marcinkowski M, et al.: Leukocyte and endothelial activation in a laboratory model of extracorporeal membrane oxygenation (ECMO). *Pediatr Res* 2000; 48: 679-84
6. Mc IRB, Timpa JG, Kurundkar AR, et al.: Plasma concentrations of inflammatory cytokines rise rapidly during ECMO-related SIRS due to the release of preformed stores in the intestine. *Lab Invest* 2010; 90: 128-39
7. Raffaelli G, Ghirardello S, Passera S, et al.: Oxidative Stress and Neonatal Respiratory Extracorporeal Membrane Oxygenation. *Frontiers in Physiology* 2018; 9: 1739-1739
8. Lou S, MacLaren G, Best D, et al.: Hemolysis in pediatric patients receiving centrifugal-pump extracorporeal membrane oxygenation: prevalence, risk factors, and outcomes. *Crit Care Med* 2014; 42: 1213-20
9. Lubnow M, Philipp A, Foltan M, et al.: Technical complications during veno-venous extracorporeal membrane oxygenation and their relevance predicting a system-exchange--retrospective analysis of 265 cases. *PLoS One* 2014; 9: e112316

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
10. Omar HR, Mirsaeidi M, Socias S, et al.: Plasma Free Hemoglobin Is an Independent Predictor of Mortality among Patients on Extracorporeal Membrane Oxygenation Support. PLoS One 2015; 10: e0124034
11. Williams DC, Turi JL, Hornik CP, et al.: Circuit oxygenator contributes to extracorporeal membrane oxygenation-induced hemolysis. ASAIO J 2015; 61: 190-5
12. Haase M, Bellomo R, Haase-Fielitz A: Novel biomarkers, oxidative stress, and the role of labile iron toxicity in cardiopulmonary bypass-associated acute kidney injury. J Am Coll Cardiol 2010; 55: 2024-33
13. Hanssen SJ, van de Poll MC, Houben AJ, et al.: Hemolysis compromises nitric oxide-dependent vasodilatory responses in patients undergoing major cardiovascular surgery. Thorac Cardiovasc Surg 2012; 60: 255-61
14. Mamikonian LS, Mamo LB, Smith PB, et al.: Cardiopulmonary bypass is associated with hemolysis and acute kidney injury in neonates, infants, and children*. Pediatr Crit Care Med 2014; 15: e111-9
15. Rother RP, Bell L, Hillmen P, et al.: The clinical sequelae of intravascular hemolysis and extracellular plasma hemoglobin: a novel mechanism of human disease. JAMA 2005; 293: 1653-62
16. Schaer DJ, Vinchi F, Ingoglia G, et al.: Haptoglobin, hemopexin, and related defense pathways-basic science, clinical perspectives, and drug development. Front Physiol 2014; 5: 415
17. Vermeulen Windsant IC, de Wit NC, Sertorio JT, et al.: Hemolysis during cardiac surgery is associated with increased intravascular nitric oxide consumption and perioperative kidney and intestinal tissue damage. Front Physiol 2014; 5: 340
18. Brittain EL, Janz DR, Austin ED, et al.: Elevation of plasma cell-free hemoglobin in pulmonary arterial hypertension. Chest 2014; 146: 1478-1485
19. Buehler PW, Baek JH, Lisk C, et al.: Free hemoglobin induction of pulmonary vascular disease: evidence for an inflammatory mechanism. Am J Physiol Lung Cell Mol Physiol 2012; 303: L312-26

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
20. Irwin DC, Baek JH, Hassell K, et al.: Hemoglobin-induced lung vascular oxidation, inflammation, and remodeling contribute to the progression of hypoxic pulmonary hypertension and is attenuated in rats with repeated-dose haptoglobin administration. *Free Radic Biol Med* 2015; 82: 50-62
21. Valladolid C, Yee A, Cruz MA: von Willebrand Factor, Free Hemoglobin and Thrombosis in ECMO. *Frontiers in Medicine* 2018; 5: 228-228
22. Fairbanks VF, Ziesmer SC, O'Brien PC: Methods for measuring plasma hemoglobin in micromolar concentration compared. *Clin Chem* 1992; 38: 132-40
23. Burkitt MJ, Milne L, Raafat A: A simple, highly sensitive and improved method for the measurement of bleomycin-detectable iron: the 'catalytic iron index' and its value in the assessment of iron status in haemochromatosis. *Clin Sci (Lond)* 2001; 100: 239-47
24. Smout J, Dyker A, Cleanthis M, et al.: Platelet Function Following Acute Cerebral Ischemia. *Angiology* 2009; 60: 362-369
25. Chung JH, Yeo HJ, Kim D, et al.: Changes in the levels of beta-thromboglobulin and inflammatory mediators during extracorporeal membrane oxygenation support. *Int J Artif Organs* 2017; 40: 575-580
26. Mildner RJ, Taub N, Vyas JR, et al.: Cytokine imbalance in infants receiving extracorporeal membrane oxygenation for respiratory failure. *Biology of the neonate* 2005; 88: 321-7
27. Risnes I, Wagner K, Ueland T, et al.: Interleukin-6 may predict survival in extracorporeal membrane oxygenation treatment. *Perfusion* 2008; 23: 173-8
28. Sullo N, Mariani S, JnTala M, et al.: An Observational Cohort Feasibility Study to Identify Microvesicle and Micro-RNA Biomarkers of Acute Kidney Injury Following Pediatric Cardiac Surgery. *Pediatr Crit Care Med* 2018; 19: 816-830
29. Ortega SB, Pandiyan P, Windsor J, et al.: A Pilot Study Identifying Brain-Targeting Adaptive Immunity in Pediatric Extracorporeal Membrane Oxygenation Patients With Acquired Brain Injury. *Critical Care Medicine* 2019: 1-1

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
30. Okochi S, Cheung EW, Barton S, et al.: An Analysis of Risk Factors for Hemolysis in Children on Extracorporeal Membrane Oxygenation. *Pediatr Crit Care Med* 2018; 19: 1059-1066
 31. Mithal LB, Palac HL, Yogev R, et al.: Cord Blood Acute Phase Reactants Predict Early Onset Neonatal Sepsis in Preterm Infants. *PLoS One* 2017; 12: e0168677
 32. Buhimschi CS, Bhandari V, Dulay AT, et al.: Proteomics mapping of cord blood identifies haptoglobin "switch-on" pattern as biomarker of early-onset neonatal sepsis in preterm newborns. *PLoS One* 2011; 6: e26111
 33. Dalton HJ, Cashen K, Reeder RW, et al.: Hemolysis During Pediatric Extracorporeal Membrane Oxygenation: Associations With Circuitry, Complications, and Mortality. *Pediatr Crit Care Med* 2018; 19: 1067-1076
 34. Lehle K, Philipp A, Zeman F, et al.: Technical-Induced Hemolysis in Patients with Respiratory Failure Supported with Veno-Venous ECMO - Prevalence and Risk Factors. *PLoS One* 2015; 10: e0143527

Table and Figure legends

Table 1. Demographics and diagnosis. Data is expressed as n (%) for binary variables, and mean(SD) or median (Q1-Q3) where appropriate for continuous variables. # n=1 from 7+ days group missing; **ECMO**: Extra-Corporeal Membrane Oxygenation; **PPHN**: Persistent Pulmonary Hypertension of the Newborn; **RV** – right ventricular; **LV** – left ventricular; **SIMV** – synchronised intermittent mechanical ventilation; **PSV** - Pressure support ventilation; **HFOV** - High frequency oscillatory ventilation

Table 2. Baseline clinical characteristics. Data expressed as n(%) for binary variables, and mean(SD) or median (Q1-Q3) where appropriate for continuous variables. **PIM2**: Paediatric Index of Mortality v2; **PMODS**: Paediatric Multiple Organ Dysfunction Score (range from 0 to 20); **FiO₂** - Fraction of inspired oxygen; **PiO₂** – partial pressure of oxygen; **pCO₂** (partial pressure of carbon dioxide); **SaO₂** – oxygen saturation; **MAP** - mean arterial pressure; **ACT** - activated clotting time.

Table 3. Clinical outcomes. Data is expressed as n(%) for binary outcomes, and mean(SD) or median (Q1-Q3) where appropriate for continuous outcomes. **RBC** – red blood cells; **FFP** – fresh frozen plasma; **SVT/AF** - Supraventricular tachycardia/Atrial fibrillation.

Figure 1. CONSORT Diagram showing patient flow through the study.

Figure 2. Inflammation markers Geometric mean and data points for **A** – IL-8, **B** – MCP-1, **C** – CRP, **D** – IL-1 β , **E** – TNF α) over time in patients who died or required prolonged stay (7+ days) vs those requiring less than 7 days (<7 days). p_{group} refers to the p-value for the effect of group (7+ vs <7 days). $p_{\text{group*time}}$ refers to the p-value for the interaction between group and time.

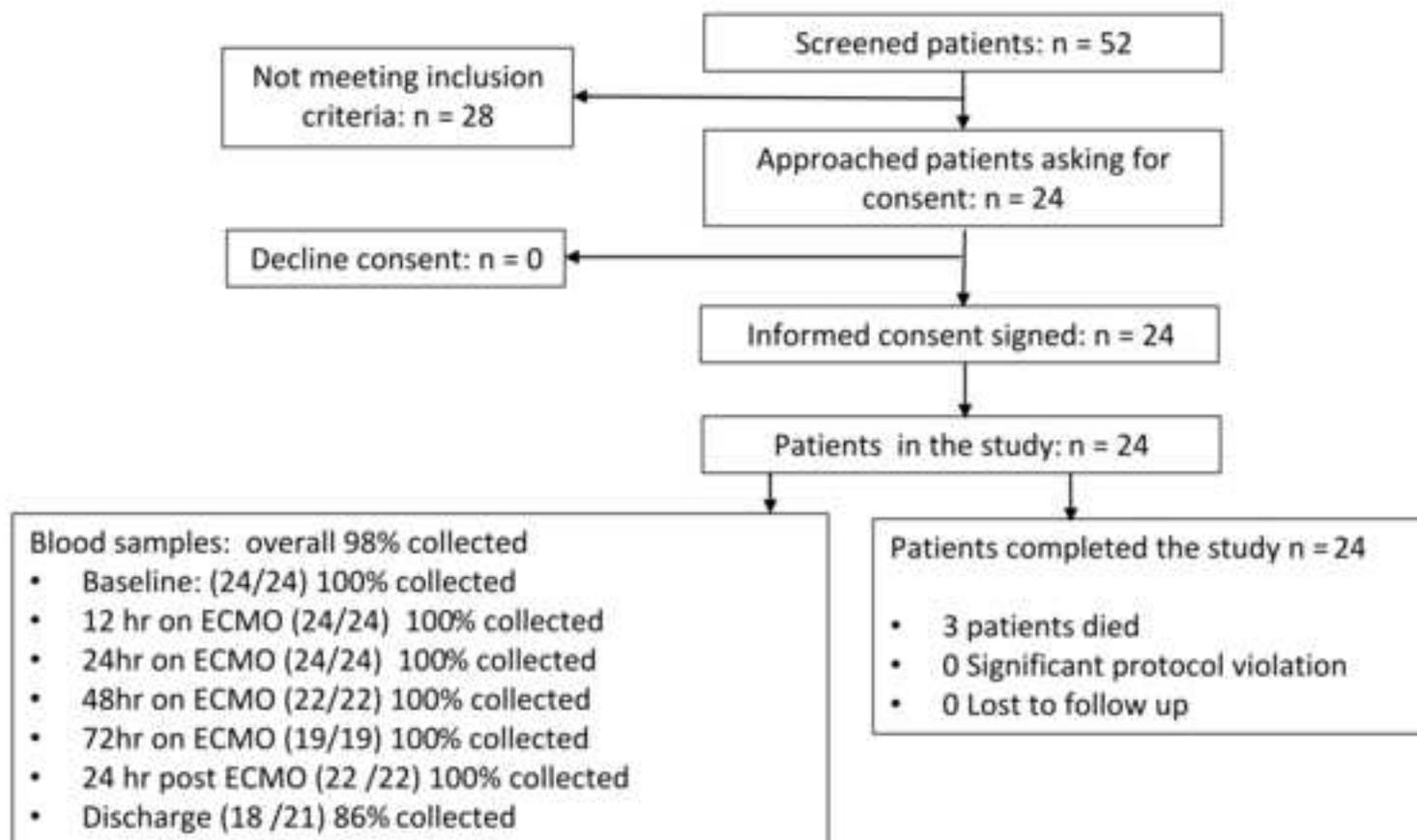
Supplemental Table: The table shows the mean (arithmetic or geometric for variables not requiring or requiring log transformation, respectively) and standard deviation (**SD**) for 7+days and <7 days groups. For **ECMO course** and **ECMO parameters** variables, **p-value**

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

column shows the p-values from the linear mixed models with effects of group, and time and interaction of group*time adjusted for baseline. For variables in **plasma free haemoglobin**, **cellular activation** and **Inflammation biomarkers**, **p-value** shows the p-values from the linear mixed models with effects of group, and time and interaction group*time; and **P_timept** shows the p-value for the effect of groups at each measured time point. **FValue** – F-statistics with corresponding degrees of freedom (**NumDF** and **DenDF**).

Supplemental Figure 1. Oxygenation, iron metabolism and cellular activation

Estimated mean and data points for **A** – SaO₂, **B** – PO₂, **C** – pCO₂, geometric mean and data points for **D** – Plasma Free Haemoglobin, **E** – Bilirubin, **F** – Haptoglobin, platelet (**A** – platelet-granulocyte aggregates CD16/CD41, **B** – platelet-monocyte aggregates) and monocyte activation biomarkers (**C** – CD14/CD41) over time in arterial blood of patients who died or required prolonged stay (7+ days) vs those requiring less than 7 days (<7 days). All participants received 100% FiO₂. p_{group} refers to the p-value for the effect of group (7+ vs <7 days). $p_{\text{group*time}}$ refers to the p-value for the interaction between group and time.



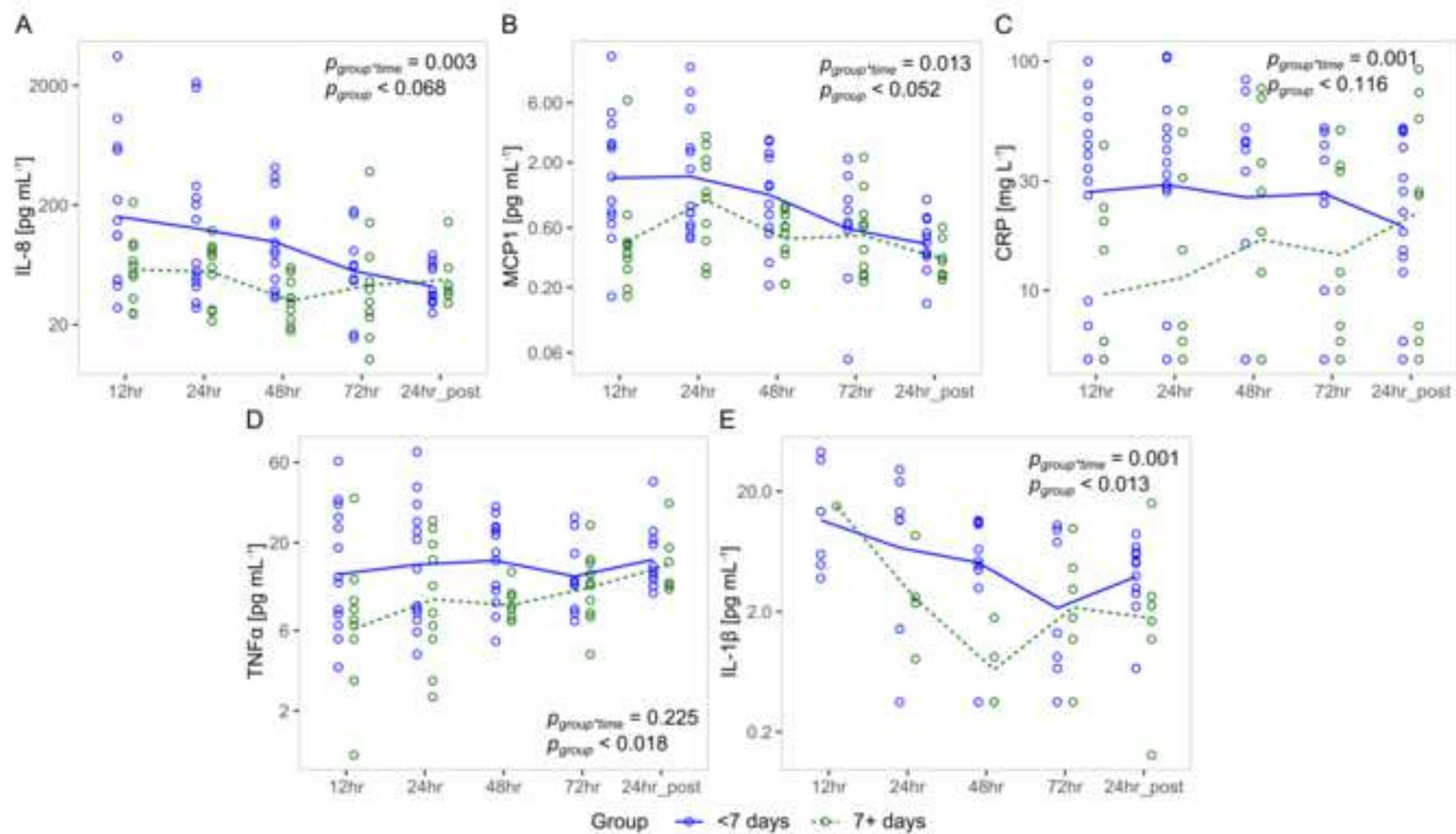


Table 1. Demographics and diagnosis. Data is expressed as n (%) for binary variables, and mean(SD) or median (Q1-Q3) where appropriate for continuous variables. # n=1 from 7+ days group missing; **ECMO**: Extra-Corporeal Membrane Oxygenation; **PPHN**: Persistent Pulmonary Hypertension of the Newborn; **RV** – right ventricular; **LV** – left ventricular; **SIMV** – synchronised intermittent mechanical ventilation; **PSV** - Pressure support ventilation; **HFOV** - High frequency oscillatory ventilation

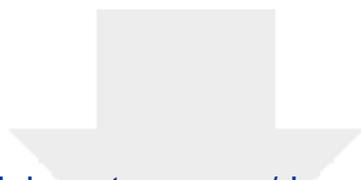
Parameter	overall (n = 24)	ECMO 7+ days (n = 10)	ECMO <7days (n = 14)	p- value
Age (day) on ECMO	2 (2 - 4)	2 (2 - 3)	2 (2 - 4)	0.890
Gestational age (week)	40.6 (2.6)	40.2 (3.3)	40.9 (2.1)	0.508
Sex - male	13 (54.2%)	5 (50.0%)	8 (57.1%)	0.999
Ethnicity - White British	24 (100%)	10 (100%)	14 (100%)	--
Weight at birth (kg)	3.2 (2.9 - 3.5)	3.1 (2.8 - 3.3)	3.2 (3.1 - 4.1)	0.102
Diagnosis				
Idiopathic PPHN	1 (4.2%)	0 (0.0%)	1 (7.1%)	0.999
Meconium Aspiration Syndrome	12 (50.0%)	0 (0.0%)	12 (85.7%)	<0.001
Congenital Diaphragmatic Hernia	9 (37.5%)	8 (80.0%)	1 (7.1%)	0.001
Pneumonia/ Respiratory Distress Syndrome	4 (16.7%)	1 (10.0%)	3 (21.4%)	0.615
other diagnosis	4 (16.7%)	1 (10.0%)	3 (21.4%)	0.615
Cardiac function				
LV - moderate/severe impairment	2 (8.3%)	1 (10.0%)	1 (7.1%)	0.999
RV - moderate/severe impairment	3 (12.5%)	1 (10.0%)	2 (14.3%)	0.999
Shunt#				
Left to right	8 (34.8%)	2 (22.2%)	6 (42.9%)	0.333
Right to left	6 (26.1%)	3 (33.3%)	3 (21.4%)	
Bidirectional	5 (21.7%)	1 (11.1%)	4 (28.6%)	
None	4 (17.4%)	3 (33.3%)	1 (7.1%)	
Mechanical ventilation	24 (100.0%)	10 (100.0%)	14 (100.0%)	--
Modality				
SIMV	7 (29.2%)	0 (0.0%)	7 (50.0%)	0.006
PSV	1 (4.2%)	0 (0.0%)	1 (7.1%)	
HFOV	16 (66.7%)	10 (100.0%)	6 (42.9%)	
Cannulation data				
Site – VV	3 (12.5%)	1 (10.0%)	2 (14.3%)	0.999
Blood priming	2 (8.3%)	2 (20.0%)	0 (0.0%)	0.163
Clear priming	14 (58.3%)	5 (50.0%)	9 (64.3%)	0.679

Table 2. Baseline clinical characteristics. Data expressed as n(%) for binary variables, and mean(SD) or median (Q1-Q3) where appropriate for continuous variables. **PIM2:** Paediatric Index of Mortality v2; **PMODS:** Paediatric Multiple Organ Dysfunction Score (range from 0 to 20); **FiO₂** - Fraction of inspired oxygen; **PiO₂** – partial pressure of oxygen; **pCO₂** (partial pressure of carbon dioxide); **SaO₂** – oxygen saturation; **MAP** - mean arterial pressure; **ACT** - activated clotting time.

Parameter	n	Overall (n=24)	ECMO 7+ days (n = 10)	ECMO <7days (n = 14)	p-value
ICU admission					
PIM2 risk	24	0.39 (0.21 - 0.55)	0.37 (0.24 - 0.43)	0.43 (0.20 - 0.56)	0.623
Haemodynamic instability prior to ECMO		12 (50.0%)	7 (70.0%)	5 (35.7%)	0.214
PMODS PICU admission	23	8 (7 - 9)	9 (7 - 11)	7 (6 - 9)	0.122
Lactates (mmol L ⁻¹)	24	4.0 (3.4)	5.3 (4.6)	3.1 (2.0)	0.191
PaO ₂ /FiO ₂ ratio	24	49.8 (25.4)	46.6 (27.7)	52.1 (24.5)	0.618
Total bilirubin (umol L ⁻¹)	24	51 (29 - 70)	51 (34 - 59)	51 (17 - 70)	0.526
Fibrinogen (g L ⁻¹)	23	1.8 (1.4 - 2.9)	1.5 (1.1 - 2.0)	2.5 (1.7 - 2.9)	0.085
Urea (mmol L ⁻¹)	24	4.8 (1.6)	5.3 (1.4)	4.5 (1.6)	0.197
Start of ECMO parameters					
MAP(mmHg)	23	48 (44 - 61)	59 (40 - 75)	46 (45 - 50)	0.289
FiO ₂ on ECMO (%)	23	100 (100 - 100)	100 (100 - 100)	100 (100 - 100)	
ACT (sec)	17	304.3 (62.9)	320.3 (36.4)	293.1 (76.2)	0.398
Pump flow (ml min ⁻¹)	22	362.3 (83.5)	358.0 (59.6)	365.8 (101.9)	0.833
Sweep gas (ml min ⁻¹)	22	300 (200 - 300)	250 (200 - 300)	300 (225 - 310)	0.244
RMP (revolutions min ⁻¹)	22	2,786 (361)	2,840 (423)	2,742 (312)	0.537
Start of ECMO - arterial blood gas					
pH	24	7.3 (0.12)	7.2 (0.14)	7.3 (0.08)	0.018
PiO ₂ (kPa)	24	6.1 (1.2)	6.0 (1.3)	6.2 (1.2)	0.710
pCO ₂ (kPa)	24	6.4 (1.7)	7.2 (2.1)	5.8 (0.9)	0.078
SaO ₂ (%)	23	84.3 (7.4)	80.9 (5.1)	86.9 (8.0)	0.050
Glu (mmol L ⁻¹)	24	7.9 (3.2)	8.0 (2.8)	7.9 (3.5)	0.925
HCO ₃ (mmol L ⁻¹)	24	20.8 (3.8)	21.1 (4.4)	20.5 (3.4)	0.700
Biochemistry					
C-reactive protein (mg L ⁻¹)	24	16 (5 - 47)	6 (5 - 15)	39 (11 - 48)	0.019
Platelet count (x10 ⁹ L ⁻¹)	24	152.6 (83.1)	139.9 (99.1)	161.6 (72.1)	0.539
Creatinine (umol L ⁻¹)	24	63 (44 - 71)	53 (43 - 66)	66 (51 - 73)	0.358
Adjusted Calcium (mmol L ⁻¹)	23	2.24 (0.17)	2.19 (0.16)	2.27 (0.19)	0.326
Alkaline Phosphatase (iu L ⁻¹)	23	131 (75 - 207)	107 (66 - 131)	198 (113 - 613)	0.030
Haemoglobin (g L ⁻¹)	24	148.0 (37.5)	151.5 (33.0)	145.4 (41.5)	0.705
Haematocrit (L L ⁻¹)	21	0.44 (0.12)	0.45 (0.11)	0.43 (0.14)	0.725
International Normalised Ratio	23	1.8 (0.5)	1.7 (0.4)	1.8 (0.6)	0.661
Activated partial thromboplastin time	22	1.9 (1.2 - 6)	1.8 (1.5 - 6)	1.9 (1.2 - 5.6)	0.357

Table 3. Clinical outcomes. Data is expressed as n(%) for binary outcomes, and mean(SD) or median (Q1-Q3) where appropriate for continuous outcomes. **RBC** – red blood cells; **FFP** – fresh frozen plasma; **SVT/AF** - Supraventricular tachycardia/Atrial fibrillation.

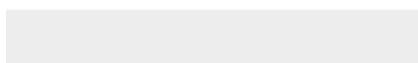
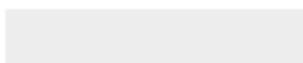
Clinical outcomes	overall (n = 24)	ECMO 7+ days (n = 10)	ECMO <7days (n = 14)	p- value
Transfusion				
RBC transfusion	24 (100.0%)	10 (100.0%)	14 (100.0%)	--
RBC transfusion volume	289.6 (241.0)	479.2 (268.2)	154.1 (74.6)	0.004
any non-RBC transfusion	20 (83.3%)	10 (100.0%)	10 (71.4%)	0.114
FFP transfusion	16 (66.7%)	7 (70.0%)	9 (64.3%)	0.999
Platelet transfusion	19 (79.2%)	10 (100.0%)	9 (64.3%)	0.053
Cryoprecipitate transfusion	13 (54.2%)	8 (80.0%)	5 (35.7%)	0.047
Complications				
SVT/AF requiring treatment	1 (4.2%)	1 (10.0%)	0 (0.0%)	0.417
ECMO circuit clotting	6 (25.0%)	6 (60.0%)	0 (0.0%)	0.002
Inotrope	20 (83.3%)	10 (100.0%)	10 (71.4%)	0.114
Vasopressor	17 (70.8%)	9 (90.0%)	8 (57.1%)	0.172
Tracheostomy	1 (4.2%)	1 (10.0%)	0 (0%)	0.417
Pneumothorax or effusion requiring drainage	5 (20.8%)	5 (50.0%)	0 (0%)	0.006
Acute kidney injury	3 (12.5%)	3 (30.0%)	0 (0%)	0.059
Peptic Ulcer / GI Bleed / Perforation	2 (8.3%)	2 (20.0%)	0 (0%)	0.163
Transient ischaemic attack	1 (4.2%)	1 (10.0%)	0 (0%)	0.417
Excess bleeding (400ml hr ⁻¹ for 1 hour or 200ml hr ⁻¹ for 4 hours))	1 (4.2%)	1 (10.0%)	0 (0%)	0.417
Wound requiring treatment	1 (4.2%)	1 (10.0%)	0 (0%)	0.417
Residual anatomical abnormalities requiring surgery	1 (4.2%)	1 (10.0%)	0 (0%)	0.417
Catheter related blood stream infection	1 (4.2%)	1 (10.0%)	0 (0%)	0.417
Any complications	20 (83.3%)	10 (100.0%)	10 (71.4%)	0.114
Number of complications	2 (1.5 - 4)	4 (3 - 5)	2 (0 - 2)	0.001
Outcome				
Chest open	1 (4.2%)	1 (10.0%)	0 (0.0%)	0.417
Survival to discharge	21 (87.5%)	7 (70.0%)	14 (100.0%)	0.059
Duration on ECMO (hours)	101 (76 - 242)	292 (186-360)	80 (59 - 91)	<0.001

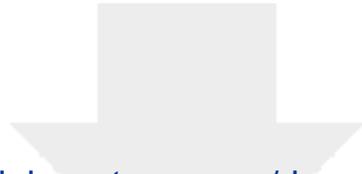


Click here to access/download

Supplemental Data File (.doc, .tif, pdf, etc.)

Supplemental Table.docx





Click here to access/download

Supplemental Data File (.doc, .tif, pdf, etc.)
fig2.tif

