

Dr Ian Johnston

Microfluidics & Microengineering Research Group University of Hertfordshire

Sensors in Food and Agriculture 2018 Norwich, UK. 19.07.2018









- Large end-user led consortium. 16 Partners, end user focused, 11 SMEs, 3 academic institutions.
- Aim is to develop reliable and cost-effective risk management tools for early detection of harmful algal blooms (HAB), chemical contaminants, bacteria, viruses and toxins.

 Early warning system for aquaculture enterprises, providing improved food security and reducing economic loss.

- Near real-time environmental monitoring.
- Autonomous operation and data distribution.



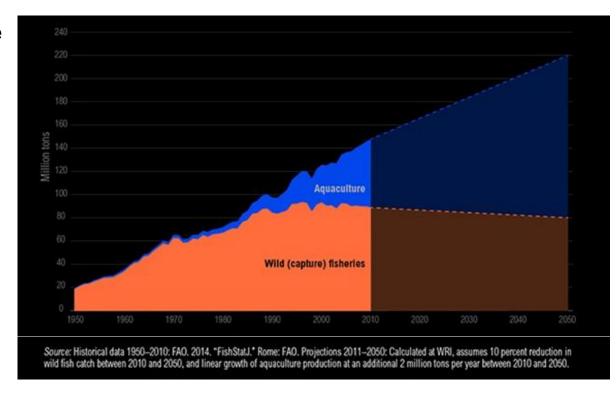








- Global fisheries and aquaculture production totalled 158 million tonnes in 2012.
- Combined this accounts for ~17% of global protein consumption.
- Aquaculture improves diet, especially in poor areas where essential nutrients are often scarce.





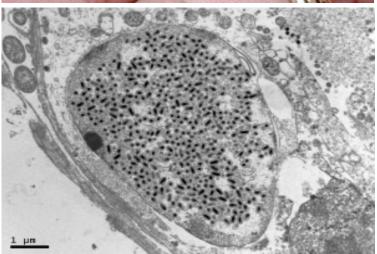






- Infectious microbial diseases limit yield.
- Global cost of these diseases exceed \$6bn pa.
- In specific sectors disease losses may exceed 40% of global capacity.
- Disease is the major restricting factor for expansion of the aquaculture industry to 2050.
- White spot in shrimp. Devastation since 1990's, accounting for at least \$1bn in losses every year since its emergence.













- In-situ early detection of Harmful Algae Blooms (HAB), chemical contaminants, viruses and toxins for aquaculture and environmental monitoring.
- Integration of 3 independent modular near realtime sensing systems.
- 3 independent sensing technologies and sensing bioassay systems.
- Potential for future integrated orthogonal detection.
- Integrated sample preprocessing.
- Integrated power supply and control systems.



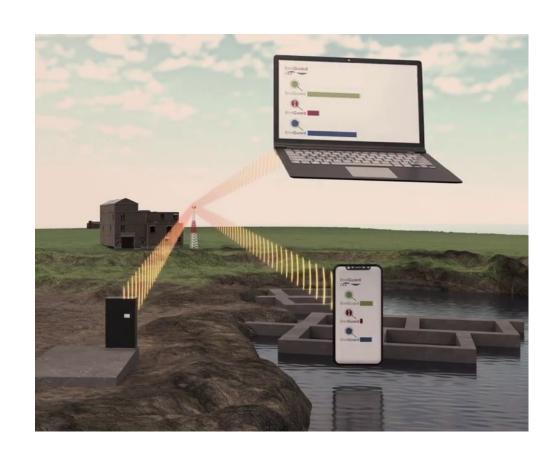








- Sensing systems integrated within a single EnviGuard Port platform.
- On-board data analysis.
- Cloud-based communication for system control and data transmission via a web interface.







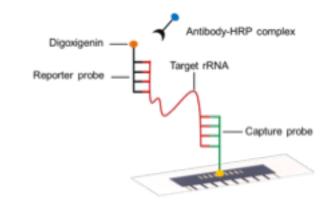






Algal Detection Unit

- A quantitative nucleic acid-based detection sensor for toxic algae in EU waters.
- Molecular detection and quantification uses a Sandwich Hybridisation Assay.
- The biosensor is calibrated for target species:
 - Protoceratium reticulatum
 - Dinophysis sp.
 - Pseudonitzschia sp.













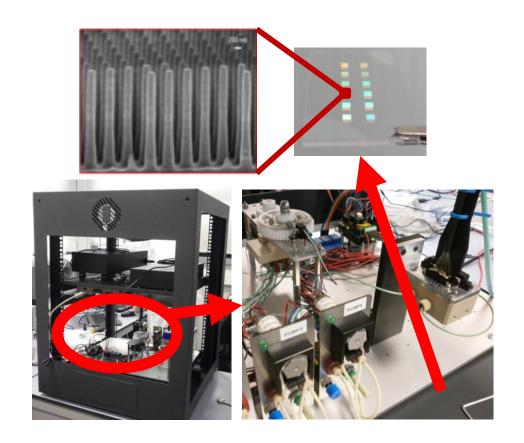


Chemical Detection Unit

- An optical nano-biosensing unit making use of antibodies for the detection of chemicals & toxins.
- Optical interrogation of biological antibody-based binding to arrays of nano-pillars.
- The biosensor is calibrated for target species:
 - PCBs
 - Okadaic Acid
 - Saxitoxin (ongoing)











- Application of UHs extensive expertise in the field of biological sample processing and system integration.
- Filter-based pathogen concentration, hydrodynamic separation, ultrasonic disruption.
- Numerous end-user driven prototype automated biological sample processing systems have been developed.
- Physical delivery of systems for further extended evaluation.













Pathogen Detection Unit

- UH have developed a prototype sample processing and analysis system for pathogen detection.
- Laser illumination and PMT-based optical fluorescence detection with a large stokes shift.
- Partners ttz Breherhaven have developed a sandwich assay employing both binding aptamer and antibody labelling stages, for target species:
 - > E.coli
 - Betanodavirus



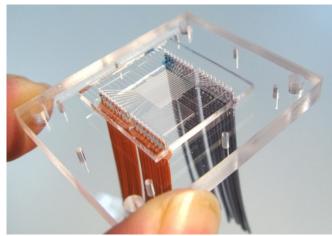


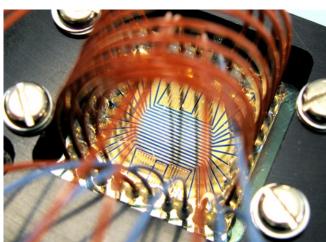






- UH have significant expertise in development of complex microfluidic devices and systems.
- For higher TRL application with real-world samples strategic decisions were made to make trade-offs between state of the art technology and in-field usability.
- Ease of use, reliability and performance.









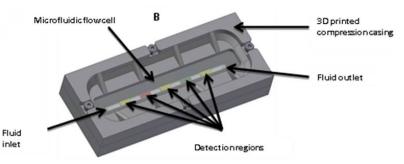




- Robust and fluidically simple microfluidic consumable design.
- Developed primarily for system development with mass manufacture capability.
- Simple easy to pattern the bioassay and manually assemble.
- Plug and play packaging for end user operation.









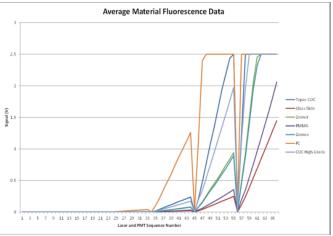






- Polymer material selection based on in system autofluorescence testing.
- Fluorophore selected with large stokes shift to minimize non specific background signal. Dynomics DY-521XL, 523 nm / 668 nm.
- PMMA was selected as the optical flow cell material due to ease of fabrication and low autofluorescence.













- Holistic approach to system design.
- In-group development of all core aspects of system design and manufacture.
- Electronics, software, custom fluid devices, packaging etc...
- Working with OEM manufacturers when there are specific requirements to use emerging commercial technologies.



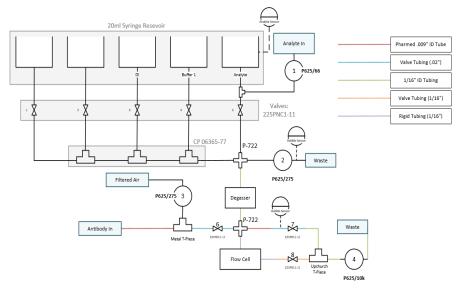


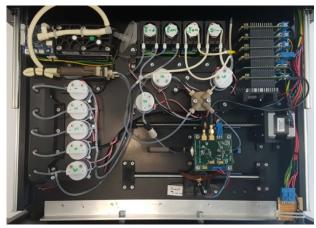






- System complexity.
- Multi analyte processes and analyte carry over.
- Sample manipulation for optimized delivery of target pathogens within collected samples.
- Sample specific requirements eg. outgassing of liquid samples.













- Redesign and repackaging for system integration.
- Designed to minimise interfacing and complexity for end-users.
- Disposable fluid reservoir consumables.
- Automated cleaning procedures.
- But with ease of access for required system maintenance.





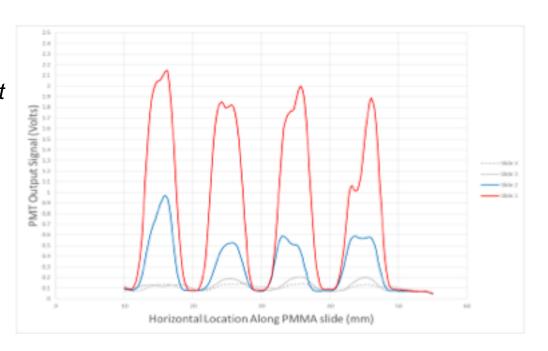








- Early example of PDU output signals along the horizontal length of the patterned PMMA slides.
- Slide1 (red) spotted regions represent exposure to 10⁷ killed E.coli cells per ml.
- Slide 2 (blue) spotted regions represent exposure to 10⁶ killed E.coli cells per ml.
- Slides 3 and 4 (grey) spotted regions are negative controls without E.coli cells, and with 10⁷ E.coli cells per ml without antibody.











- Simple to install and get operational.
- Remote operation and data transmission processes all validated.
- Includes custom designed components for:
 - Precision antibody introduction and dispensing.
 - > Bubble free operation.
 - > Fluid level monitoring.











- System testing and field trials are comencing on Orkney now at Orkney Shellfish Hatchery.
- EnviGuard Training Workshops are planned for:
 - August 1st Orkney Islands, UK.
 - November 7th Bremerhaven
 Germany (+ Final project conference).













<u>Acknowledgements</u>

D.McCluskey¹, R.Kaye¹, I. Munro¹, B. Suckow², I. Klarholz², P. Ciaurriz³, E. Teiletxea³, F. Fernández³, I. Cornago³, A. Baron⁴, C. Deffaud⁴, A. L. Hernandez⁵, R. Casquel⁵, M. Holgado⁵, T. T. Veenstra⁶, H. von Hörsten⁷, F. Dortu⁷, M. Maigler⁸, K. Metfies⁹, J. Hessel⁹, P. Sprong⁹, T. Hanken¹⁰

- 1 University of Hertfordshire, College Lane, Hatfield, Herts., UK.
- 2 ttz Bremerhaven, Am Lunedeich 12, 27572 Bremerhaven, Germany.
- 3 Cemitec, ADItech, Polígono Mocholí, Plaza Cein 4, 31110 Noain, Spain.
- 4 Biotem, Parc d'Activités Bièvre Dauphine, Alphonse Gourju 885, Apprieu, France.
- 5 Center for Biological Technology, Technical University of Madrid, 28223 Pozuelo, Spain.
- 6 LioniX BV, PO Box 456, 7500 AL Enschede, The Netherlands.
- 7 Multitel a.s.b.l., 2 Rue Pierre et Marie Curie, BE-7000 Mons, Belgium.
- 8 Bio Optical Detection S.L., Technical University of Madrid, 28223 Pozuelo, Spain.
- 9 Alfred Wegener Institute, Helmholtz Centre for Polar and Marine Research, Am Handelshafen 12, 27570 Bremerhaven, Germany.
- 10 iSiTEC GmbH, Bussestraße 27, 27570 Bremerhaven, Germany.









www.enviguard.net























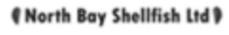












This project has received funding from the European Union's Seventh Framework Programme for research, technological development and demonstration under grant agreement no 614057





