- 1 Commercially important species associated with horse mussel (*Modiolus*
- 2 *modiolus*) biogenic reefs: a habitat for biodiversity conservation and
- 3 fisheries benefits
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13 <u>Highlights</u>

- Communities of conspicuous megafauna were assessed on *Modiolus modiolus* reefs,
 sand and cobble habitats.
- Tissue samples from reef fauna were subject to stable isotope analysis to investigate
 trophic structure.
- *M. modiolus* reefs support a higher abundance of commercially important shellfish,
- including *Aequipecten opercularis*, *Buccinum undatum* and *Maja brachydactyla*, than
 cobble and sand habitats.
- Evidence of the commercial value of *M. modiolus* reefs in the UK.

22 <u>Abstract</u>

23 Horse mussel beds (*Modiolus modiolus*) are protected biodiversity hotspots in Marine

24 Protected Areas of the NE Atlantic. In this study, horse mussel beds, cobble habitats and

sandy habitats were assessed using underwater visual census and drop-down video techniques

26 in three UK regions. Megafauna were enumerated, differences in community composition

- 27 and individual species abundances were analysed. Samples of conspicuous megafauna were
- also collected from horse mussel beds in Orkney for stable isotope analysis.
- 29 Communities of conspicuous megafauna were different between horse mussel habitats and
- 30 other habitats throughout their range. Three commercially important species: whelks
- 31 (Buccinum undatum), queen scallops (Aequipecten opercularis) and spider crabs (Maja

brachydactyla) were significantly more abundant (by as much as 20 times) on horse mussel
beds than elsewhere. Isotopic analysis provided insights into their trophic relationship with
the horse mussel bed. Protection of *M. modiolus* habitat can achieve biodiversity conservation
objectives whilst benefiting fisheries also.

36

37 <u>Introduction</u>

38 Globally, fish stocks are under threat (Jackson et al., 2001) with the proportion of overexploited fish stocks at 32% in 2008 (FAO, 2010), leading to calls to halt this trend and 39 restore them (Worm et al., 2009). One method of improving fisheries management has been a 40 shift to 'Ecosystem-Based Management' (EBM) that involves a move from traditional single 41 species based management to a greater understanding of the interactions between habitats and 42 43 commercially important species. In the NE Atlantic, habitat-forming species such as maerl (a coralline red algae) create a physically complex and biodiverse seabed, which provides a 44 nursery ground for commercially important fish and shellfish (Kamenos et al., 2004a, 2004b). 45 Modiolus modiolus (horse mussel) reefs are structurally complex habitats, characterised by 46 high species diversity (Hirst et al., 2012; Rees et al., 2008; Sanderson et al., 2008). The 47 societal benefits of horse mussel reefs for fishermen has been demonstrated (Kent et al 2016); 48 yet the utilisation of horse mussel reefs as a resource for benthic consumers has not been 49 50 studied.

51

In the United States, the Sustainable Fisheries Act (SFA) now requires fisheries managers to
identify the 'Essential Fish Habitat' (EFH) for commercially important fish species (Fluharty,
2000), i.e. "those waters and substrate necessary to fish spawning, feeding or growth to
maturity" (NOAA, 1966). This concept is not confined to describing finfish habitat

associations; it has also been used for invertebrates such as conch (Glazer and Kidney, 2004),

57 octopus (Garofalo et al., 2011) and even bottlenose dolphin (Ingram and Rogan, 2002).

58

The EBM approach extends the focus of managing resources beyond the target species and includes impacts to non-target species and benthic habitats, supporting ecological processes that are required to sustain harvestable resources (Hughes et al., 2005). With an increasing human population comes an increased demand for ecosystem goods and services, i.e. "the benefits natural ecosystems provide to human society" (de Groot et al., 2002). In the marine environment, there is increasing interest in the relationship between biodiversity and
ecosystem services given the wide scale loss of biodiversity in marine ecosystems (Worm et
al., 2006). Marine Protected Areas (MPAs) are seen as an important management tool to
conserve species and habitats (Tundi Agardy, 1994) and, over time, well managed MPAs can
support ecosystem services to provide ecological and societal benefits (Fox et al., 2012).

70 Those implementing the Habitats Directive across Europe have rarely evaluated the ecosystem services or the commercial value of the biodiversity that they seek to maintain 71 72 (Maes et al., 2012). Marine biodiversity conservation is lagging behind the terrestrial equivalent, and increasing uses and demands on the marine environment have led to the 73 development of policy frameworks that integrate human activities as part of the system 74 (Atkins et al., 2011). Historically, biodiversity management and fisheries management 75 76 have been developed separately due to constraints imposed on sectors and fundamentally 77 different aspirations. Thus, the motivation to identify EFH is not provided through fisheries legislation in Europe or spatial management of marine habitats either. However, 78 recognition of habitat associations and appropriate management of EFH has the potential 79 80 to provide benefits for both fisheries and nature conservation.

81

Identifying a habitat as an EFH is not a simple task, especially for highly mobile species 82 83 that use a range of habitats throughout their life cycle. With limited resources, 84 conserving or restoring every habitat used by a fish is unrealistic, therefore identifying 85 habitats that are used during sensitive life stages are often prioritised (Levin and Stunz, 2005). Relative fish abundance measurements provide an initial insight into which areas 86 87 are important for fish and shellfish. However, to recognise a habitat as an EFH requires a greater understanding of the functional role of the habitat, i.e. whether it is being used 88 89 for feeding or reproduction, etc.

90

Stable isotope analysis is becoming a popular method for studying food webs (Schaal et al., 2012). ¹⁵N becomes enriched during trophic transfers, with a mean increase (or "trophic enrichment factor", TEF) of 3.4‰ (Minagawa and Wada, 1984; Post, 2002) equivalent to approximately one trophic level, though there are variations, with TEF being larger for herbivorous fish (Mill et al., 2007). The δ^{13} C from the tissue of an animal is more conservative with trophic transfer, instead reflecting the source(s) of 97 carbon in the food chain. Analysis of δ^{15} N and δ^{13} C together can be used to determine 98 the feeding relationships on a biogenic habitat (Grall et al., 2006), bentho-pelagic 99 coupling (McIntyre et al., 2006) and habitat associations (Yeager and Layman, 2011).

100

UK biogenic reefs have been recognised as biodiversity hotspots, e.g. *M. modiolus* reefs; 101 102 (Mair et al., 2000; Moore et al., 2013; Rees, 2009) and mapped for conservation 103 purposes, e.g. Lindenbaum et al. (2008). However, our understanding of habitat 104 associations and the trophic structure of many biogenic reef communities is limited. Underwater Visual Censuses (UVCs) can be used to measure relative megafauna 105 abundance (Kamenos et al., 2004b) and the benefit of using this technique is that it is 106 non-destructive and independent of fishing gear bias. Remotely Operated Vehicles 107 (ROVs) and towed videos have also been used to record the abundance of mobile species 108 associated with coral reefs (Söffker et al., 2011) and gorgonians (De Clippele et al., 109 2015). 110

111

As structurally complex seabed features, *M. modiolus* reefs are expected to provide a 'habitat provision' ecosystem service that has been shown to be the case for other biogenic structures (De Clippele et al., 2015; Kamenos et al., 2004b; Margiotta et al., 2016). However, *M. modiolus* reefs are relatively inaccessible and their ecological function is understudied. The aim of this study is to identify the key megafaunal species (defined as animals greater than 2 cm maximum length) associated with three *M. modiolus* reefs in the UK and to investigate the trophic feeding niche structure of a *M. modiolus* reef megafaunal community.

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120 <u>Methods</u>

121 Megafauna abundance

Underwater Visual Censuses (UVCs) were carried out using SCUBA at twenty-three sites in 122 Shetland and Orkney (Figure 1 B and C) between 15 and 25 m below chart datum. It was 123 impractical to survey at multiple sites at Pen Llŷn (Figure 2 D) due to strong currents and 124 short slack water periods, therefore a Drop Down Video (DDV) camera system was used. 125 Sites in Orkney and Shetland were surveyed in May 2013 and September 2012 respectively. 126 Sites off Pen Llŷn were surveyed between May and June of 2008 and 2010. In all cases, 127 conspicuous megafauna, including fish and commercially important invertebrates such as 128 shellfish, were quantified. 129

- 131 UVC sites were selected at random using extant video footage and preliminary dive data to
- stratify sampling to 'horse mussel reef' (8 sites), 'sand' (8 sites) or 'cobble' (7 sites) habitats.
- Horse mussel reef sites had a density of > 20 M. *modiolus* m⁻². All observers were trained in
- species identification during a pilot study in May 2012 in Orkney and all surveys covered a
- 60 m^2 area delineated by a transect tape deployed from a shot-line. Habitats were verified as
- 136 cobble, sand or *M. modiolus* reef using photoquadrats and granulometry samples.
- 137

The DDV camera system used was a Sony Model DCR-TRV950 camcorder fitted into a 138 139 tubular aluminium housing with two HID video lamps and lasers (10cm apart) were used to quantify the seabed area covered. The video surface unit included a labelling system that 140 overlaid GPS position and depth information onto the surface image for recording and 141 viewing. DDV camera tows were approximately 100 m in length, but actual distances were 142 calculated from the GPS start and end points and a section of 4-8 minutes was edited from the 143 video clip for analysis. Video footage that was too fast (>30 m per minute) or with less than 1 144 m visibility was disregarded. Eight *M. modiolus* reef sites were sampled off Pen Llŷn as well 145 as 8 cobble sites and 8 sand sites. 146

147

148 Food web analysis

149 Thirty-seven samples were collected for stable isotope analysis from eleven species across

150 three *M. modiolus* reef sites in Orkney (see Supplementary Table S1 for sampling locations)

151 from the 1st-5th December 2013. *Aequipecten opercularis* (n=6), *Leocarcinus depurator*

152 (n=5), Asterias rubens (n=1) and Buccinum undatum (n=5) were collected using SCUBA by

searching the reef from a central location or on a drift dive. *M. modiolus* (n=14) and

polychaetes (n=3) were collected using a 'clump sample' technique where a clump of 3-5 M.

155 *modiolus* and associated fauna are placed into a bucket along with the underlying sediment

156 (Mair et al., 2000). *Gadus morhua* (n=1) and *Taurulus bubalis* (n=3) were collected from

157 sites adjacent to the reefs using baited creels.

158

159 White muscle was taken from the fish samples and adductor from the bivalves, while the foot

160 muscle was dissected from the *B. undatum* samples. Gill tissue was taken from the

161 crustaceans and tube feet from A. rubens. Samples were freeze-dried and ground to a fine

162 powder using a pestle and mortar. Approximately 1 mg of tissue from each sample was

loaded into a 4 x 6 mm tin capsule and combusted in continuous flow isotope ratio mass

spectrometer (CF-IRMS). Results are expressed as parts per thousand (‰) deviations from
international standards, in delta (δ) notation.

166

167 *Data Analysis*

168 Megafauna counts from the Underwater Visual Censuses (UVCs) and those from the Drop

169 Down Video (DDV) transects were analysed separately due to the different methodology

used. For the multivariate analysis, all data were log transformed to down-weigh very

abundant species (e.g. *Echinus esculentus* abundance varied by three orders of magnitude).

172 Using Bray-Curtis similarity, Analysis of Similarities (ANOSIM) (Clarke and Gorley, 2006)

was used to test for differences in megafaunal communities between *M. modiolus* reef, sandand cobble habitats.

175

A SIMPER analysis was used to examine the key species driving the differences in 176 community composition (Clarke and Gorley, 2006). Counts of the three species contributing 177 the most to the difference in megafauna communities at the reef, cobble and sand sites were 178 then analysed using a Kruskal-Wallis test with 'habitat type' as the independent variable. 179 Pairwise Kruskal-Wallis tests were then performed using a chi-squared probability 180 distribution with a Bonferroni correction for multiple comparisons. The stable isotope 181 samples were checked to ensure protein had been analysed: samples with a Carbon/Nitrogen 182 ratio of greater than four were removed as this implies a high lipid content or carbonate in the 183 184 sample. Samples were averaged for each species where possible and standard deviations calculated. All univariate analyses were carried out in R version 2.9.1 (R Core Development 185 186 Team, 2011).

187

188 Results

189 Megafauna community analysis

Fifteen megafauna species were recorded from 23 UVC samples in Orkney and Shetland. An ANOSIM shows that the megafaunal community was significantly different amongst habitats (r = 0.45, p = <0.001). Pairwise tests showed that the megafauna counted at the *M. modiolus* reefs were significantly different to those living on the sand habitats (r = 0.45, p=0.003) and the cobble habitats (r = 0.38, p = 0.002). The SIMPER analysis showed that the highest within-group similarity was in the *M. modiolus* reef group (72.68%). The sand habitats were

the least similar group, with an average similarity of 19.59%. The high counts of flatfish and

- dragonets at sand sites contributed 70% of the similarity within the sandy sites. *M. modiolus*
- reef sites were most dissimilar to the sand sites (90.41% dissimilarity) and 50% of this
- 199 dissimilarity was attributable to higher densities of *E. esculentus*, *A. opercularis* and *B.*
- 200 *undatum* on the reefs sites (SIMPER; Figure 3a, b and c).
- 201
- 202 *M. modiolus* reef sites had over five times more *A. opercularis* than the cobble sites (x^2)
- =11.011, p=0.001) and more than 20 times as many as the sand sites ($x^2 = 12.427$, p=<0.001). B. *undatum* were almost twenty times more abundant on reef sites compared to sand sites (x^2
- 205 = 7.136, p=0.008). However, the counts at the reef sites were not significantly different to the
- cobble sites at the 0.05 significance level (x^2 = 3.84, p=0.050). The most abundant species
- found on the *M. modiolus* reefs was *E. esculentus* (0.46 m⁻²), which was significantly greater
- than the cobble sites (x^2 =9.442, p=0.002) and the sand sites (x^2 =12.308, p=<0.001).
- 209
- 210 On the UVC transects, gobies (*Pomatoschistus pictus* and *Pomatoschistus microps*) were
- abundant in all habitats, but especially the sand sites (a maximum of 24 *P. pictus* counted on
- one transect). Juvenile flatfish (*Limanda limanda*) were abundant at two of the sand sites and
- 213 dragonets (*Callionymus spp*) were present at three out of eight sand sites (16 counted in
- total). The fish species associated with the *M. modiolus* sites were almost entirely different to
- those associated with the sand sites (with the exception of *Callionymus* sp. and
- 216 *Pomatoschistus pictus*, which occurred in both habitats). The cling-fish *Diplecogaster*
- 217 *bimaculata* occurred at the *M. modiolus* sites but not at the off-reef sites.
- 218
- A total of 22 species were identified from the DDV analysis at 23 sites off the Pen Llŷn. The 219 220 megafaunal community associated with the M. modiolus reef was significantly different to the sand and cobble habitats (ANOSIM, r=0.26, p=<0.001). The top three species highlighted 221 222 in the SIMPER analysis, contributing to the difference in community structure between M. modiolus reef sites and off-reef sites, were B. undatum, A. rubens and Maja brachydactyla 223 (previously known as Maja squinado: Sotelo et al., 2007). The cumulative contribution of 224 these three species in the difference between habitats was 44.15% between reef and sand 225 habitats, and 44.72% between reef and cobble habitats. Other key species causing differences 226 in the species assemblages were curled octopus (Eledone cirrhosa) and butterfish (Pholis 227 gunnelus), which were only seen at M. modiolus reef sites. 228
- 229

230 The DDV transects showed that A. rubens was the most abundant conspicuous megafaunal species and there were significantly more observed on the reef than the sand ($x^2 = 9.26$, 231 p = < 0.001) and cobble habitats ($x^2 = 11.618$, p = 0.002). B. undatum were rarely seen on the 232 DDV surveys compared to the UVCs (0.01 m⁻² compared to 0.11 m⁻² respectively). From the 233 DDV analysis, no *B. undatum* were observed on cobble sites, although they were 234 occasionally seen on the reef (0.02 m^{-2}) and sand sites (0.01 m^{-2}) . Ballan wrasse (Labrus 235 bergylta) occurred on the cobble sites but not on the sand sites. Sea scorpions (Taurulus 236 bubalis) occurred at the reef sites and the cobble sites but not on the sandy sites, whereas 237 238 dogfish (Scyliorhinus canicula) occurred on the reef sites and the sand sites but not the cobble sites. 239

240

241 *Foodweb analysis*

The *M. modiolus*, *A. opercularis*, and *B. undatum* samples from Orkney showed consistent
stable isotope signatures (Figure 4). Even *M. modiolus* samples from different reefs (up to

244 2km apart) showed very similar δ^{13} C and δ^{15} N values. For the megafauna tissue samples

analysed, δ^{15} N increases in the following order: A. opercularis; M. modiolus; B. undatum and

246 *Liocarcinus depurator; T. bubalis* and *G morhua*. The average δ^{15} N for *M. modiolus* is

247 9.21‰ compared with 11.56‰ for *B. undatum*. The highest δ^{15} N values were from the fish

samples, showing that these animals are at the top of the food chain in this study, and

possibly feeding on crustaceans and bivalves on the reef. The isotope ratios from *M. modiolus*

and *A. opercularis* samples formed two distinct groups with the δ^{15} N of *M. modiolus*

approximately 2‰ higher than *A. opercularis* (Figure 4).

252

253 Discussion

Through *in situ* observations, this study provides evidence that *M. modiolus* reefs across the 254 255 UK support a high abundance of megafaunal species compared to other habitats, including the commercially valuable shellfish, A. opercularis, B. undatum and Maja brachydactyla. 256 These findings are in keeping with other biodiversity hotspots in the UK, such as maerl beds 257 that provide an important habitat for A. opercularis, A. rubens and E. esculentus (Kamenos et 258 al., 2004b). Structurally complex marine habitats such as oyster beds and seagrass beds also 259 provide a disproportionately important refuge for other species (Bertelli and Unsworth, 2014; 260 Shervette and Gelwick, 2008). However this is the first time that *M. modiolus* reefs have been 261

shown to provide a comparable functional role to these other complex habitats.

263

The stable isotope data also show the range of feeding mechanisms used within the reef 264 community and the different food sources on the reef available for mobile species. This also 265 gives an insight into the ecological function of the *M. modiolus* reefs and for example, shows 266 that B. undatum and L. depurator are approximately one tropic level higher than M. modiolus. 267 When considered in combination with the high abundance of *B. undatum* relative to other 268 'off-reef' habitats, this suggests that *B. undatum* may be feeding on *M. modiolus*. Although 269 270 normally considered a scavenger, predation on bivalves by B. undatum has been observed in 271 a laboratory setting (Scolding et al., 2007). Habitat heterogeneity is often important for mobile species during different life stages (Kimirei et al., 2011) and M. modiolus reef habitats 272 provide a range of physical conditions from fine sediment to large shell gravel and epifauna 273 (Nic Aonghua et al., 2001). 274

275

The two survey methods used in this study (UVCs and DDV) together provide a 276 comprehensive method for measuring conspicuous megafauna abundance; a wide range of 277 species were sampled and the area of seabed sampled was easily defined. However, it is 278 279 likely that cryptic fish species are under-represented in these more complex reef habitats, as 280 indicated by Bozec et al. (2011), and the use of different sampling techniques may be required to fully grasp the different fish species associated with *M. modiolus*. Similarly, the 281 282 high counts of B. undatum on M. modiolus reefs in Orkney and Shetland were not mirrored in the DDV transects off Pen Llŷn probably because this is a slow moving semi-cryptic species, 283 284 which is difficult to identify from DDV footage. Divers on the UVC transects were able to 285 look around objects and check if shellfish were live or dead, which was not possible on the 286 DDV transects. In a parallel study, comparatively high catch rates of *B. undatum* on *M.* modiolus reefs off Pen Llŷn using baited traps (Kent et al., In Review) corroborates the view 287 288 that DDV is not the best technique for recording this species.

289

Spider crabs (*Maja brachydactyla*) were highly abundant on the Pen Llŷn *M. modiolus* reef compared to the off-reef sites and this species is harvested throughout Europe, especially in the English Channel and the Mediterranean (Fahy, 2001). Within the UK, *M. brachydactyla* is largely distributed on the south coast of England and Wales; hence the lack of observations on the Orkney and Shetland reef sites, although its distribution may extend further north with rising seawater temperatures (Goodwin et al., 2013). *M. modiolus* reefs often form a mosaic with other biogenic habitat (e.g. maerl in Shetland; Hirst et al., 2013) and it is likely that mobile species benefit from this variety (Buhl-Mortensen et al., 2010). Further investigations

into the diet of the key mobile species identified in this study through stomach content

- analysis and a more comprehensive stable isotope study would develop understanding of the
- 300 predator-prey interactions occurring within and between biogenic habitats.
- 301

The Underwater Visual Censuses showed very high densities of *A. operculum* on the *M. modiolus* reefs. Elsewhere, juvenile *A. opercularis* has been shown to attach to hard
substrates and use rugose structures such as maerl, which provide a refuge from predators
(Kamenos et al., 2004c). It is likely that *A. opercularis* utilise *M. modiolus* shell crevices and
associated epibiota (Figure 2) to avoid predation and increase post-recruitment survival.
Juvenile *A. opercularis* are able to swim, providing the option to move to a more favourable
environment once settled, if a different habitat is required.

309

The present study shows that the two filter feeding species *M. modiolus* and *A. opercularis* fill different trophic niches, which is consistent with Yakovis et al. (2012) who found that horse mussels and ascidians growing together have distinct diets and there is no overlap in the origin of their food. Increasing evidence for differential particle selection in bivalves (Kiorboe and Mohlenberg, 1981; Shumway et al., 1985; Ward and Shumway, 2004) provides a possible explanation for how the filter feeding *M. modiolus* and *A. opercularis* can co-occur in high densities together as adults in a reef environment.

317

Overall, the high abundance of A. opercularis, B. undatum and M. brachydactyla on reefs in 318 combination with the tropic structure of the former two (Figure 5), provides strong evidence 319 320 that *M. modiolus* reefs are 'Essential Fish Habitats' for these species. This highlights the value of horse mussel reefs in providing disproportionately more ecosystem services and 321 322 corresponding benefits to society than other habitats. However, M. modiolus reefs along with other shellfish habitats have declined over the past 100 years (Thurstan et al., 2013) and are 323 vulnerable to physical impacts (Cook et al., 2013). Strangford Lough is a Marine Protected 324 Area that was once home to the UK's largest *M. modiolus* reefs (> 10km²), and the *A*. 325 326 opercularis trawl fishery boomed there in the 1980s (Strain et al., 2012) causing long-term declines in *M. modiolus* and associated megafauna, with little sign of recovery of this long-327 lived reef forming species (Elsäßer et al., 2013; Fariñas-Franco and Roberts, 2014). 328 Ironically, therefore, the fisheries benefits that *M. modiolus* reefs can deliver to society can 329 330 lead to their demise. Indeed, given that many coastal habitats provide important functional

roles for commercially important fish (e.g. nursery, spawning and feeding grounds), it is
unsurprising that destruction results in the consequential decline in associated fish landings
(see Seitz et al., 2014).

334

In many countries, including the UK, biodiversity conservation and fisheries management 335 have historically been the responsibility of separate government departments or agencies. 336 However, marine conservation management has progressed from imposed, command-and-337 control 'fortress conservation' approaches to systems that increasingly combine both 'top-338 339 down' and 'bottom-up' governance (Jones, 2014; Kelleher, 1993). Indeed, these more balanced approaches are reflected globally, where the Convention on Biological Diversity 340 Aichi 6 arguably brings biodiversity concerns closer to fisheries management and UN 341 Sustainable Development Goals than ever before. Understanding the ecological interaction 342 between habitats and mobile species is required for the identification of EFH in contemporary 343 sustainable fisheries management. Here we present evidence to illustrate that this concept can 344 345 be usefully implemented in the context of Marine Spatial Planning to prioritise the most 346 valuable areas of the seabed (Crowder and Norse, 2008). Thus, the protection of fragile habitats of biodiversity importance can intersect with sustainable fisheries to achieve 347 348 ecosystem-based management.

349

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358 <u>References</u>

Atkins, J.P., Burdon, D., Elliott, M., Gregory, A.J., 2011. Management of the marine
environment: integrating ecosystem services and societal benefits with the DPSIR
framework in a systems approach. Mar. Pollut. Bull. 62, 215–226.

- Bertelli, C.M., Unsworth, R.K.F., 2014. Protecting the hand that feeds us: Seagrass (*Zostera marina*) serves as commercial juvenile fish habitat. Mar. Pollut. Bull. 83, 425–429.
- Bozec, Y.-M., Kulbicki, M., Laloë, F., Mou-Tham, G., Gascuel, D., 2011. Factors affecting

| 365 | the detection distances of reef fish: implications for visual counts. Mar. Biol. 158, 969. | | |
|--------------------------|---|--|--|
| 366 367 368 | Buhl-Mortensen, L., Vanreusel, A., Gooday, A.J., Levin, L.A., Priede, I.G., Buhl-Mortensen, P., Gheerardyn, H., King, N.J., Raes, M., 2010. Biological structures as a source of habitat heterogeneity and biodiversity on the deep ocean margins. Mar. Ecol. 31, 21–50. | | |
| 369 | Clarke, K.R., Gorley, R.N., 2006. PRIMER v6: User Manual/Tutorial. Prim. Plymouth, UK. | | |
| 370 371 372 373 | Cook, R., Fariñas-Franco, J.M., Gell, F.R., Holt, R.H.F., Holt, T., Lindenbaum, C., Porter, J.S., Seed, R., Skates, L.R., Stringell, T.B., Sanderson, W.G., 2013. The Substantial First Impact of Bottom Fishing on Rare Biodiversity Hotspots: A Dilemma for Evidence-Based Conservation. PLoS One 8, e69904. doi:10.1371/journal.pone.0069904 | | |
| 374 375 | Crowder, L., Norse, E., 2008. Essential ecological insights for marine ecosystem-based management and marine spatial planning. Mar. Policy 32, 772–778. | | |
| 376 377 | De Clippele, L.H., Buhl-Mortensen, P., Buhl-Mortensen, L., 2015. Fauna associated with cold water gorgonians and sea pens. Cont. Shelf Res. 105, 67–78. | | |
| 378 379 380 | de Groot, R.S., Wilson, M.A., Boumans, R.M.J., 2002. A typology for the classification, description and valuation of ecosystem functions, goods and services. Ecol. Econ. 41, 393–408. doi:http://dx.doi.org/10.1016/S0921-8009(02)00089-7 | | |
| 381 382 383 384 | Elsäßer, B., Fariñas-Franco, J.M., Wilson, C.D., Kregting, L., Roberts, D., 2013. Identifying optimal sites for natural recovery and restoration of impacted biogenic habitats in a special area of conservation using hydrodynamic and habitat suitability modelling. J. Sea Res. 77, 11–21. doi:http://dx.doi.org/10.1016/j.seares.2012.12.006 | | |
| 385 386 | Fahy, E., 2001. The Magharees spider crab <i>Maja squinado</i> fishery in 2000. Mar. Fish. Serv. Div. Mar. Institute, Abbotstown, Castleknock, Dublin 15. | | |
| 387 388 | FAO, 2010. The state of the World Fisheries and Aquaculture 2010. Food Agric. Organ. Rome, 197. | | |
| 389 390 | Fariñas-Franco, J.M., Roberts, D., 2014. Early faunal successional patterns in artificial reefs used for restoration of impacted biogenic habitats. Hydrobiologia 75. | | |
| 391 392 | Fluharty, D., 2000. Habitat protection, ecological issues, and implementation of the Sustainable Fisheries Act. Ecol. Appl. 10, 325–337. | | |
| 393 394 395 | Fox, H.E., Mascia, M.B., Basurto, X., Costa, A., Glew, L., Heinemann, D., Karrer, L.B., Lester, S.E., Lombana, A. V, Pomeroy, R.S., 2012. Reexamining the science of marine protected areas: linking knowledge to action. Conserv. Lett. 5, 1–10. | | |
| 396 397 398 399 | Garofalo, G., Fortibuoni, T., Gristina, M., Sinopoli, M., Fiorentino, F., 2011. Persistence and co-occurrence of demersal nurseries in the Strait of Sicily (central Mediterranean): Implications for fishery management. J. Sea Res. 66, 29–38. doi:DOI 10.1016/j.seares.2011.04.008 | | |
| 400 401 402 | Glazer, R.A., Kidney, J.A., 2004. Habitat associations of adult queen conch (<i>Strombus gigas</i> L.) in an unfished Florida Keys back reef: applications to essential fish habitat. Bull. Mar. Sci. 75, 205–224. | | |
| 403 | Goodwin, C.E., Strain, E.M.A., Edwards, H., Bennet, S.C., Breen, J., Picton, B.E., 2013. | | |

- Effects of two decades of rising sea surface temperatures on sublittoral macrobenthic
 communities in Northern Ireland, UK. Mar. Environ. Res. 85, 34–44.
- Grall, J., Le Loc'h, F., Guyonnet, B., Riera, P., 2006. Community structure and food web
 based on stable isotopes (delta N-15 and delta C-13) analysis of a North Eastern Atlantic
 maerl bed. J. Exp. Mar. Bio. Ecol. 338, 1–15. doi:10.1016/j.jembe.2006.06.013
- Hirst, N.E., Clark, L., Sanderson, W.G., 2012. The distribution of selected MPA search
 features and Priority Marine Features off the NE coast of Scotland (JOUR), Scottish
 Natural Heritage Commissioned Report no. 500. Scottish Natural Heritage.
- Hirst, N.E., Kamphausen, L.M., Cook, R.L., Porter, J.S., Sanderson, W.G., 2013. Distribution
 and status of proposed protected features in the Fetlar to Haroldswick MPA proposal.
 Scottish Nat. Herit. Comm. Rep. no. 599.
- Hughes, T.P., Bellwood, D.R., Folke, C., Steneck, R.S., Wilson, J., 2005. New paradigms for
 supporting the resilience of marine ecosystems. Trends Ecol. Evol. 20, 380–386.
 doi:http://dx.doi.org/10.1016/j.tree.2005.03.022
- Ingram, S.N., Rogan, E., 2002. Identifying critical areas and habitat preferences of bottlenose
 dolphins Tursiops truncatus. Mar. Ecol. Prog. Ser. 244, 247–255.
- 420 Jackson, J.B.C., Kirby, M.X., Berger, W.H., Bjorndal, K.A., Botsford, L.W., Bourque, B.J., Bradbury, R.H., Cooke, R., Erlandson, J., Estes, J.A., Hughes, T.P., Kidwell, S., Lange, 421 C.B., Lenihan, H.S., Pandolfi, J.M., Peterson, C.H., Steneck, R.S., Tegner, M.J., 422 423 Warner, R.R., Jeremy, B.C.J., Kirby, M.X., Berger, W.H., Bjorndal, K.A., Botsford, L.W., Bourque, B.J., Bradbury, R.H., Cooke, R., Erlandson, J., Estes, J.A., Hughes, 424 T.P., Kidwell, S., Lange, C.B., Lenihan, H.S., Pandolfi, J.M., Peterson, C.H., Steneck, 425 426 R.S., Tegner, M.J., Warner, R.R., 2001. Historical Overfishing and the Recent Collapse of Coastal Ecosystems. Science (80-.). 293, 629. 427
- Jones, P.J.S., 2014. Governing marine protected areas: resilience through diversity.
 Routledge, Oxon, UK.
- Kamenos, N.A., Moore, P.G., Hall-Spencer, J.M., 2004a. Maerl grounds provide both refuge
 and high growth potential for juvenile queen scallops (*Aequipecten opercularis* L.). J.
 Exp. Mar. Bio. Ecol. 313, 241–254.
- Kamenos, N.A., Moore, P.G., Hall-Spencer, J.M., 2004b. Nursery-area function of maerl
 grounds for juvenile queen scallops *Aequipecten opercularis* and other invertebrates.
 Mar. Ecol. Prog. Ser. 274, 183–189.
- Kamenos, N.A., Moore, P.G., Hall-Spencer, J.M., 2004c. Attachment of the juvenile queen
 scallop (*Aequipecten opercularis* (L.)) to maerl in mesocosm conditions; juvenile habitat
 selection. J. Exp. Mar. Bio. Ecol. 306, 139–155.
 doi:http://dx.doi.org/10.1016/j.jembe.2003.10.013
- Kelleher, G., 1993. Guidelines for Marine Protected Areas. IUCN, Gland, Switzerland and
 Cambridge, UK.
- Kent, F.E.A., Gray, M., Last, K.S., Sanderson, W.G., 2016. Horse mussel reef ecosystem
 services: Evidence for a whelk nursery habitat supporting a shellfishery. Int. J.
 Biodivers. Sci. Ecosyst. Serv. Manag. 12, 172–180.

- Kimirei, I.A., Nagelkerken, I., Griffioen, B., Wagner, C., Mgaya, Y.D., 2011. Ontogenetic
 habitat use by mangrove/seagrass-associated coral reef fishes shows flexibility in time
 and space. Estuar. Coast. Shelf Sci. 92, 47–58.
- Kiorboe, T., Mohlenberg, F., 1981. Particle selection in suspension-feeding bivalves. Mar.
 Ecol. Prog. Ser.
- Levin, P.S., Stunz, G.W., 2005. Habitat triage for exploited fishes: Can we identify essential
 "Essential Fish Habitat?" Estuar. Coast. Shelf Sci. 64, 70–78.
 doi:http://dx.doi.org/10.1016/j.acss.2005.02.007
- 452 doi:http://dx.doi.org/10.1016/j.ecss.2005.02.007
- Lindenbaum, C., Bennell, J.D., Rees, E.I.S., McClean, D., Cook, W., Wheeler, A.J.,
 Sanderson, W.G., 2008. Small-scale variation within a *Modiolus modiolus* (Mollusca : Bivalvia) reef in the Irish Sea: I. Seabed mapping and reef morphology. J. Mar. Biol.
 Assoc. United Kingdom 88, 133–141.
- Maes, J., Paracchini, M.L., Zulian, G., Dunbar, M.B., Alkemade, R., 2012. Synergies and
 trade-offs between ecosystem service supply, biodiversity, and habitat conservation
 status in Europe. Biol. Conserv. 155, 1–12.
- Mair, J.M., Moore, C.G., Kingston, P.F., Harries, D.B., 2000. A review of the status, ecology
 and conservation of horse mussel *Modiouls modiolus* beds in Scotland. Scottish Nat.
 Herit. Comm. Rep. F99PA08, 89.
- Margiotta, A.M., Shervette, V.R., Hadley, N.H., Plante, C.J., Wilber, D.H., 2016. Speciesspecific responses of resident crabs to vertical habitat complexity on intertidal oyster
 reefs. J. Exp. Mar. Bio. Ecol. 477, 7–13.
- McIntyre, J.K., Beauchamp, D.A., Mazur, M.M., Overman, N.C., 2006. Ontogenetic trophic
 interactions and benthopelagic coupling in Lake Washington: Evidence from stable
 isotopes and diet analysis. Trans. Am. Fish. Soc. 135, 1312–1328. doi:10.1577/t05099.1
- Mill, A.C., Pinnegar, J.K., Polunin, N.V.C., 2007. Explaining isotope trophic-step
 fractionation: why herbivorous fish are different. Funct. Ecol. 21, 1137–1145.
- 472 Minagawa, M., Wada, E., 1984. Stepwise enrichment of 15 N along food chains: further
 473 evidence and the relation between δ 15 N and animal age. Geochim. Cosmochim. Acta
 474 48, 1135–1140.
- Moore, C.G., Harries, D.B., Cook, R.L., Hirst, N.E., Saunders, G.R., Kent, F.E.A., Trigg, C.,
 Lyndon, A.R., 2013. The distribution and condition of selected MPA search features
 within Lochs Alsh, Duich, Creran and Fyne. Scottish Nat. Herit. Comm. Rep. 566, 197.
- 478 Nic Aonghua, C., Wilson, J.G., Mackie, A.S.Y., 2001. Sediments. Wilson. J. G. al Benthic
 479 Biodivers. South. Irish Sea 2. South West Irish Sea Surv. Stud. Biodivers. Syst. from
 480 Natl. Museum Wales. BIOMOR Rep. 2 71–103.
- 481 NOAA, 1966. Magnuson-Stevens Fishery Management and Conservation Act amended
 482 through October 1996. NOAA Tech. Memo. NMFS-F/SPO.
- 483 Post, D.M., 2002. Using stable isotopes to estimate trophic position: models, methods, and
 484 assumptions. Ecology 83, 703–718.

- 485 R Core Development Team, 2011. R: A language and environment for statistical computing.
 486 R Found. Stat. Comput. Vienna, Austria.
- 487 Rees, E.I.S., 2009. Assessment of *Modiolus modiolus* beds in the OSPAR area (JOUR), Joint
 488 Nature Conservation Comittee.
- Rees, E.I.S., Sanderson, W.G., Mackie, A.S.Y., Holt, R.H.F., 2008. Small-scale variation
 within a *Modiolus modiolus* (Mollusca : Bivalvia) reef in the Irish Sea. III. Crevice,
 sediment infauna and epifauna from targeted cores. J. Mar. Biol. Assoc. United
 Kingdom 88, 151–156. doi:10.1017/S0025315408000052
- Sanderson, W.G., Holt, R.H.F., Kay, L., Ramsay, K., Perrins, J., McMath, A.J., Rees, E.I.S.,
 2008. Small-scale variation within a *Modiolus modiolus* (Mollusca : Bivalvia) reef in the
 Irish Sea. II. Epifauna recorded by divers and cameras. J. Mar. Biol. Assoc. United
 Kingdom 88, 143–149.
- 497 Schaal, G., Riera, P., Leroux, C., 2012. Food web structure within kelp holdfasts (Laminaria):
 498 a stable isotope study. Mar. Ecol. Evol. Perspect. 33, 370–376.
- Scolding, J.W.S., Richardson, C.A., Luckenbach, M.J., 2007. Predation of cockles
 (*Cerastoderma edule*) by the whelk (*Buccinum undatum*) under laboratory conditions.
- Seitz, R.D., Wennhage, H., Bergström, U., Lipcius, R.N., Ysebaert, T., 2014. Ecological
 value of coastal habitats for commercially and ecologically important species. ICES J.
 Mar. Sci. J. du Cons. 71, 648–665.
- Shervette, V.R., Gelwick, F., 2008. Relative nursery function of oyster, vegetated marsh
 edge, and nonvegetated bottom habitats for juvenile white shrimp *Litopenaeus setiferus*.
 Wetl. Ecol. Manag. 405.
- Shumway, S.E., Cucci, T.L., Newell, R.C., Yentsch, C.M., 1985. Particle selection, ingestion,
 and absorption in filter-feeding bivalves. J. Exp. Mar. Bio. Ecol. 91, 77–92.
 doi:10.1016/0022-0981(85)90222-9
- Söffker, M., Sloman, K.A., Hall-Spencer, J.M., 2011. In situ observations of fish associated
 with coral reefs off Ireland. Deep. Res. Part I 58, 818–825.
 doi:10.1016/j.dsr.2011.06.002
- Sotelo, G., Morán, P., Posada, D., 2007. Identification and characterization of microsatellite
 loci in the spiny spider crab *Maja brachydactyla*. Conserv. Genet. 8, 245.
- Strain, E.M.A., Allcock, A.L., Goodwin, C.E., Maggs, C.A., Picton, B.E., Roberts, D., 2012.
 The long-term impacts of fisheries on epifaunal assemblage function and structure, in a
 Special Area of Conservation. J. Sea Res. 67, 58–68.
- Thurstan, R.H., Hawkins, J.P., Raby, L., Roberts, C.M., 2013. Oyster (*Ostrea edulis*)
 extirpation and ecosystem transformation in the Firth of Forth, Scotland. J. Nat.
 Conserv. 21, 253–261.
- Tundi Agardy, M., 1994. Advances in marine conservation: the role of marine protected
 areas. Trends Ecol. Evol. 9, 267–270. doi:http://dx.doi.org/10.1016/01695347(94)90297-6
- 524 Ward, J.E., Shumway, S.E., 2004. Separating the grain from the chaff: particle selection in

- suspension- and deposit-feeding bivalves. J. Exp. Mar. Bio. Ecol. 300, 83–130.
 doi:10.1016/j.jembe.2004.03.002
- Worm, B., Barbier, E.B., Beaumont, N., Duffy, J.E., Folke, C., Halpern, B.S., Jackson,
 J.B.C., Lotze, H.K., Micheli, F., Palumbi, S.R., Sala, E., Selkoe, K.A., Stachowicz, J.J.,
 Watson, R., 2006. Impacts of biodiversity loss on ocean ecosystem services. Science
 (80-.). 314, 787–790. doi:10.1126/science.1132294
- Worm, B., Hilborn, R., Baum, J.K., Branch, T.A., Collie, J.S., Costello, C., Fogarty, M.J.,
 Fulton, E.A., Hutchings, J.A., Jennings, S., Jensen, O.P., Lotze, H.K., Mace, P.M.,
 McClanahan, T.R., Minto, C., Palumbi, S.R., Parma, A.M., Ricard, D., Rosenberg, A.A.,
 Watson, R., Zeller, D., 2009. Rebuilding global fisheries. Science (80-.). 325, 578–585.
- Yakovis, E.L., Artemieva, A. V, Fokin, M. V, Varfolomeeva, M.A., 2012. Intraspecific
 variation in stable isotope signatures indicates no small-scale feeding interference
 between a horse mussel and an ascidian. Mar. Ecol. Prog. Ser. 467, 113–120.
 doi:10.3354/meps09951
- Yeager, L.A., Layman, C.A., 2011. Energy flow to two abundant consumers in a subtropical
 oyster reef food web. Aquat. Ecol. 45, 267–277. doi:10.1007/s10452-011-9352-1
- 541
- 542
- 543 <u>Figures</u>





Figure 1. A) *M. modiolus* reefs study regions for megafauna abundance within the UK. B) Survey
locations in Shetland (UVC method); C) Orkney (UVC method and stable isotope samples collected);

and D) Pen Llŷn (Drop Down Video method). Positions are labelled with the number of sites where

548 markers overlap. Black circle with green cross = sand habitats (16 sites in total); solid square = M.

549 *modiolus* reefs (15 sites in total); red triangle = cobble habitats (15 sites in total).



551 Figure 2. A) UVC transect technique, B) Aequipecten opercularis, C) Pholis gunnelus, D) Buccinum

- 552 *undatum*, E) *Echinus esculentus* and *Asterias rubens* and F) *Pomatoschistus pictus*. Images A, B, D,
- and E: Richard Shucksmith, images C and F: George Stoyle.





Figure 3. Counts of the key megafauna species from the UVC transects a) b) and c) and the DDV
transects d), e) and f). Bars show the mean ± 1SE. Groups that share a capital letter are not
significantly different at the 0.017 probability threshold (Bonferroni correction on Kruskal Wallis
test).



Cobble & Sand

Reef & Sand

10.8482

12.3077

0.0009

0.0005

Video (DDV)

585 key species pairwise comparisons (Kruskal Wallis tests), Bonferroni correction; significance at p <

586 0.016.

587

| Key Species | Habitat comparison | \mathbf{X}^2 | P value |
|--------------------|--------------------|----------------|---------|
| Buccinum undatum | Reef & Cobble | 6.55 | 0.0100 |
| | Reef & Sand | 0.0269 | 0.8690 |
| | Cobble & Sand | 6.5502 | 0.0100 |
| Maja brachydactyla | Reef & Cobble | 2.6042 | 0.1066 |
| | Reef & Sand | 2.6042 | 0.1066 |
| | Cobble & Sand | 0 | 1.0000 |
| Asterias rubens | Reef & Cobble | 9.258 | 0.0023 |
| | Cobble & Sand | 0.0509 | 0.8214 |
| | Reef & Sand | 11.618 | 0.0007 |