Bone 131 (2020) 115160

Contents lists available at ScienceDirect

Bone

journal homepage: www.elsevier.com/locate/bone

Full Length Article

Lipidomic architecture shared by subclinical markers of osteoporosis and atherosclerosis: The Cardiovascular Risk in Young Finns Study

Binisha H. Mishra^{a,b,c,*}, Pashupati P. Mishra^{a,b,c}, Nina Mononen^{a,b,c}, Mika Hilvo^d, Harri Sievänen^e, Markus Juonala^{f,g,h}, Marika Laaksonenⁱ, Nina Hutri-Kähönen^j, Jorma Viikari^{f,g}, Mika Kähönen^{b,k}, Olli T. Raitakari^{h,l,m}, Reijo Laaksonen^{a,b,d}, Terho Lehtimäki^{a,b,c}

^a Department of Clinical Chemistry, Faculty of Medicine and Health Technology, Tampere University, Tampere, Finland

^b Finnish Cardiovascular Research Centre, Faculty of Medicine and Health Technology, Tampere University, Tampere, Finland

^c Department of Clinical Chemistry, Fimlab Laboratories, Tampere, Finland

^d Zora Biosciences Oy, Espoo, Finland

^e UKK Institute, Tampere, Finland

^f Department of Medicine, University of Turku, Turku, Finland

⁸ Division of Medicine, Turku University Hospital, Turku, Finland

^h Research centre of Applied and Preventive Cardiovascular Medicine, University of Turku, Turku, Finland

ⁱ Fazer Lab Research, Oy Karl Fazer Ab, Helsinki, Finland

^j Department of Paediatrics, Tampere University Hospital, Faculty of Medicine and Health Technology, Tampere University, Tampere, Finland

^k Department of Clinical Physiology, Tampere University Hospital, Tampere Finland.

¹Department of Clinical Physiology and Nuclear Medicine, Turku University Hospital, Turku, Finland

^m Centre for Population Health Research, University of Turku and Turku University Hospital, Turku Finland

ARTICLE INFO

Keywords: Osteoporosis Atherosclerosis Comorbidity Lipidomics Weighted co-expression network analysis

ABSTRACT

Background: Studies have shown that osteoporosis and atherosclerosis are comorbid conditions sharing common risk factors and pathophysiological mechanisms. Understanding these is crucial in order to develop shared methods for risk stratification, prevention, diagnosis and treatment. The aim of this study was to apply a system-level bioinformatics approach to lipidome-wide data in order to pinpoint the lipidomic architecture jointly associated with surrogate markers of these complex comorbid diseases.

Subjects and methods: The study was based on the Cardiovascular Risk in Young Finns Study cohort from the 2007 follow-up (n = 1494, aged 30–45 years, women: 57%). Liquid chromatography-tandem mass spectrometry (LC-MS/MS) was used to analyse the serum lipidome, involving 437 molecular lipid species. The subclinical osteoporotic markers included indices of bone mineral density and content, measured using peripheral quantitative computer tomography from the distal and shaft sites of both the tibia and the radius. The subclinical atherosclerotic markers included carotid and bulbus intima media thickness measured with high-resolution ultrasound. Weighted co-expression network analysis was performed to identify networks of densely interconnected lipid species (i.e. lipid modules) associated with subclinical markers of both osteoporosis and atherosclerosis. The levels of lipid species (lipid profiles) of each of the lipid modules were summarized by the first principal component termed as module eigenlipid. Then, Pearson's correlation (r) was calculated between the module eigenlipids and the markers. Lipid modules that were significantly and jointly correlated with subclinical markers of both osteoporosis and atherosclerosis were considered to be related to the comorbidities. The hypothesis that the eigenlipids and profiles of the constituent lipid species in the modules have joint effects on the markers was tested with multivariate analysis of variance (MANOVA).

Results: Among twelve studied molecular lipid modules, we identified one module with 105 lipid species significantly and jointly associated with both subclinical markers of both osteoporosis (r = 0.24, p-value = 2×10^{-20}) and atherosclerosis (r = 0.16, p-value = 2×10^{-10}). The majority of the lipid species in this module belonged to the glycerolipid (n = 60), glycerophospholipid (n = 13) and sphingolipid (n = 29) classes. The module was also enriched with ceramides (n = 20), confirming their significance in cardiovascular outcomes and suggesting their joint role in the comorbidities. The top three of the 37 statistically significant (adjusted p-value < 0.05) lipid species jointly associated with subclinical markers of both osteoporosis and atherosclerosis within the module were all triacylglycerols (TAGs) – TAG(18:0/18:0/18:1) with an adjusted p-

* Corresponding author at: Department of Clinical Chemistry, Fimlab Laboratories and Finnish Cardiovascular Research Centre Tampere, Finland *E-mail address:* binisha.hamalmishra@tuni.fi (B.H. Mishra).

https://doi.org/10.1016/j.bone.2019.115160

Received 17 June 2019; Received in revised form 2 November 2019; Accepted 18 November 2019 Available online 21 November 2019 8756-3282/ © 2019 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/BY-NC-ND/4.0/).







Bone



value of 8.6 \times 10⁻⁸, TAG(18:0/18:1/18:1) with an adjusted p-value of 3.7 \times 10⁻⁶, and TAG(16:0/18:0/18:1) with an adjusted p-value of 8.5 \times 10⁻⁶.

Conclusion: This study identified a novel lipid module associated with both surrogate markers of both subclinical osteoporosis and subclinical atherosclerosis. Alterations in the metabolism of the identified lipid module and, more specifically, the TAG related molecular lipids within the module may provide potential new biomarkers for testing the comorbidities, opening avenues for the emergence of dual-purpose prevention measures.

1. Introduction

Cardiovascular diseases and osteoporosis are both widely prevalent disorders, inducing serious morbidities, bone fractures and death [1-4]. Evidence indicates that there is a similar pathophysiological mechanism underlying both diseases [5]. Several association studies have linked bone measures with atherosclerosis-related measures, such as echogenic calcified atherosclerotic plaques, pulse wave velocity and coronary artery calcification [3,6-8]. Using human atherosclerotic plaque transcriptomics and confocal microscopic analysis, we have shown that advanced atherosclerotic lesions express a variety of markers related with osteoclastogenesis, osteoblastogenesis and calcification and that they involve osteoclast-like cells [9]. Furthermore, genetic polymorphism of apolipoprotein E, a key regulator of serum lipid levels [10] and atherosclerosis [11], has also been shown to be associated with bone structural traits [12]. Various studies have also revealed a positive biological effect of statin, a cholesterol-lowering drug used for the prevention of cardiovascular diseases, to be effective on bone density [13,14]. However, there are also studies that reveal no significant association between the bone and vascular markers [15–17]. Although, they share the same biomarkers and risk factors - for example, oestrogen deficiency, vitamin D abnormalities, dyslipidaemia, smoking, physical inactivity, intake of dietary calcium, dietary saturated fat, oxidative stress and genetic factors [18-21] - the nature and the mechanism involved remains elusive.

Lipidomics offers a tool to investigate the systemic lipid profiles produced in the body's cells, tissues and organs, as well as their interactions with other molecular and cellular components [22]. An altered lipidome has been shown to be associated with several clinical conditions [23,24]. Understanding these alterations can provide useful insight into the development process of the diseases. A study investigating the shared underlining mechanism of atherosclerosis and osteoporosis comorbidity by utilizing lipidomics data is lacking. Therefore, the objective of the present study was to perform a system-level analysis of lipidomics data to identify networks of lipid species associated jointly with subclinical markers of both osteoporosis and atherosclerosis.

The lipidomics data in this study involved 437 molecular lipids generated with liquid chromatography-tandem mass spectrometry (LC-MS/MS) technique from the serum of 1494 participants. Traditional individual molecule-wise statistical methods are limited in their ability to provide a holistic system-level picture. We, therefore, performed a signed weighted lipid co-expression network analysis to identify networks of lipid species (modules) jointly associated with subclinical markers of both osteoporosis and atherosclerosis [25].

2. Material and methods

2.1. Study subjects

The Cardiovascular Risk in Young Finns Study (YFS) is a prospective multi-centre follow-up study investigating cardiovascular risk factors from childhood to adulthood [26]. The study was initiated in 1980 with 3596 children and adolescents aged 3–18 years. The participants were randomly selected from the areas of five university hospitals in Finland (Turku, Tampere, Helsinki, Kuopio and Oulu) and have been followed for nearly 40 years. The present study is based on 1494 participants aged 30–45 from the 2007 follow-up, with four atherosclerotic and six

osteoporotic markers, as summarized in Table 2.

2.2. Measurement of surrogate markers of subclinical atherosclerosis

Carotid and bulbus intima-media thickness (IMT) were used as surrogate markers of subclinical atherosclerosis. An ultrasound imaging device with a high-resolution system (Sequoia 512, Acuson) including 13.0 MHz linear array transducers was used for IMT measurement by trained sonographers following a standardized protocol. The image was focused on the posterior (far) wall, and images were recorded from the angle showing the greatest distance between the lumen-intima interface and the media-adventitia interface. A scan including the beginning of the carotid bifurcation and the common carotid artery was recorded and stored in digital format on optical discs for subsequent off-line analysis. All scans were analysed by one reader blinded to the participants' details. The best-quality end-diastolic frame was selected. Several measurements of the common carotid far wall were taken approximately 10 mm proximally to derive the maximal carotid IMT. To assess the reproducibility of the IMT measurements, we re-examined 60 participants 3 months after the initial visit (2.5% random sample). The between-visit coefficient of variation of IMT measurements was 6.4%. To assess the reproducibility of the IMT image analysis, 113 scans were re-analysed by a second observer, and the coefficient of variation was 5.2%. The mean and maximum carotid and bulbus IMT was used in the study.

2.3. Measurement of surrogate markers of subclinical osteoporosis

Two trained researchers in each study centre performed the peripheral quantitative computed tomography (pQCT) bone measurements from both the distal and the diaphysis sites of the radius and tibia. The same pQCT device was used in all five centres (XCT 2000R, Stratec, Medizintechnik, Pforzheim, Germany). The tomographic slices were taken from the shaft (a cortical-rich bone site) and the distal part (a trabecular-rich bone site) of the weight-bearing tibia (30% and 5% from the distal endplate of the tibia, respectively) and of the nonweightbearing radius (30% and 4% from the distal endplate of the radius, respectively) according to our standard procedures [27]. For the shaft regions, the analysed bone traits were total area (ToA, mm²), cortical area (CoA, mm²), and cortical density (CoD, mg/cm³). For the distal parts of the radius and tibia, the measured bone traits were ToA (mm²), CoA (mm²) and trabecular density (TrD, mg/cm³). The range of in vivo precision of the used pQCT-measured traits ranged from 0.5% (CoD of the radial shaft) to 4.4% (CoA of the distal radius). Mineral content was calculated as 0.2 \times (area/100) \times density. The measured indices are demonstrated in Table 2.

2.4. Health and life style data

The physical activity index was calculated as metabolic equivalents (METs) by combining information on the frequency, intensity and duration of physical activity including leisure-time physical activity and commuting to the workplace (MET h/wk). One MET corresponds to the energy consumption of one kilocalorie per kilogram of weight per hour at rest [28]. Alcohol consumption was measured by asking participants to report their alcohol consumption during the previous week. One unit is equivalent to 14 g of alcohol [29].

2.5. Lipidome-wide analysis

Lipidome quantification for the stored serum samples was performed at Zora Biosciences Oy (Espoo, Finland). Lipid extraction was based on a previously described method [30]. In brief, 10 µl of 10 mM 2.6-di-tert-butyl-4-methylphenol (BHT) in methanol was added to 10 ul of the sample, followed by 20 µl of internal standards (Avanti Polar Lipids Inc., Alabaster, AL) and 300 µl of chloroform:methanol (2:1, v:v) (Sigma-Aldrich GmbH, Steinheim, Germany). The samples were mixed and sonicated in a water bath for 10 min, followed by a 40-min incubation and centrifugation (15 min at 5700 \times g). The upper phase was transferred and evaporated under nitrogen. Extracted lipids were resuspended in 100 µl of water-saturated butanol and sonicated in a water bath for 5 min. Then, 100 µl of methanol was added to the samples before the extracts were centrifuged for 5 min at 3500 \times g, and finally the supernatants were transferred to the analysis plate for mass spectrometric (MS) analysis. The MS analyses have also been described in detail previously [31]. The analyses were performed on a hybrid triple quadrupole/linear ion trap mass spectrometer (QTRAP 5500, AB Sciex, Concord, Canada) equipped with ultra-high-performance liquid chromatography (UHPLC) (Nexera-X2, Shimadzu, Kyoto, Japan).



Chromatographic separation of the lipidomic screening platform was performed on an Acquity BEH C18, 2.1 \times 50 mm id. 1.7 μm column (Waters Corporation, Milford, MA, USA). The data were collected using a scheduled multiple reaction monitoring algorithm and processed using Analyst and MultiQuant 3.0 software (AB Sciex). The heights of the peaks obtained from the MS analysis were normalized with the internal standard of the lipid classes.

2.6. Biostatistical analysis

The lipid profiles were log_e transformed to correct for skewness. We used signed weighted co-expression network analysis implemented in R statistical software [25] to identify groups of densely interconnected lipid species, hereafter referred to as lipid modules. The analysis pipeline is illustrated in Fig. 1. Pearson's correlation coefficients (r) were calculated for all pairwise comparisons of lipid species across all participants. The correlation matrix was transformed to an adjacency matrix by raising it to the power of 5, chosen based on scale-free topology criteria (Fig. S1). The power transformation reduces noise by supressing low correlations and emphasizing stronger correlations between lipid species. The power term is chosen in a manner that leads to

Fig. 1. Weighted co-expression network analysis pipeline. 1) Lipidomics data generated with liquid chromatography-tandem mass spectrometry technique (LC-MS/MS). 2) Levels of lipid species. 3) Correlation matrix based on the pairwise correlation (Pearson) of the lipid species. 4) Hierarchical clustering of the dissimilarity matrix generated from the correlation matrix. 5) Identification of lipid species modules based on clustering. 6) Summarization of the levels of constituent lipid species in the modules by calculating their first principal component called as eigenlipid. 7) Correlation between eigenlipids (as representative of modules) and markers of subclinical osteoporosis and atherosclerosis. 8) Examination of the correlation between module membership and the lipid significance of lipid species in the selected modules as a quality check of the modules.

a scale-free network topology because most of the biological networks are expected to be approximately scale-free. The resulting adjacency matrix was used to generate a Topological Overlap Matrix (TOM). The TOM is a pairwise similarity matrix of the lipid species that considers topological similarity among lipid species. For example, a high TOM implies that a pair of lipid species shares several neighbour lipid species with similar levels. The TOM was transformed into a dissimilarity matrix. Average linkage hierarchical clustering of the dissimilarity matrix was performed to generate a hierarchical clustering tree of lipid species. Lipid modules that are weighted networks of lipid species were identified with a dynamic tree-cutting algorithm. The lipid profiles in each module were summarized by the module eigenlipid (ME), which is defined as the first principal component of the modules' lipid profiles. Association analysis was performed by calculating Pearson's correlation coefficients (r) between the modules and the studied markers. Multivariate analyses of variance (MANOVA) were conducted for significant modules and their constituent member lipid species in order to test the hypothesis that the eigenlipid and profiles of the constituent lipid species in the modules have joint effects on markers of both subclinical osteoporosis and subclinical atherosclerosis. All the multivariate analyses were adjusted for age and sex. All statistical analyses and data processing were performed using the statistical package R version 3.4.3 [32].

3. Results

3.1. Study population characteristics

The characteristics of the study population are shown in Table 1. The disease incidences are based on self-reports [27]. The measured markers of subclinical osteoporosis and atherosclerosis are shown in Table 2.

3.2. Association between surrogate markers of subclinical osteoporosis and atherosclerosis

The surrogate markers of subclinical osteoporosis had a weak but significant (p-value < 0.01) positive correlation with those of subclinical atherosclerosis (Fig. 2).

3.3. Identification of lipid modules

An adjacency matrix was generated from the correlation matrix of the molecular lipid species using a soft-thresholding power of five with the *WGCNA* R package. The threshold was chosen based on a network topology analysis. The network resembled a scale-free graph, with $r^2 > 0.80$, when the correlation matrix was raised to the power of five (Fig. S1). The hierarchical clustering of the TOM dissimilarity matrix defined 12 modules, containing 6–105 highly correlated lipid species (Fig. S2). The lipid modules were named according to colour for downstream analysis, as shown in Fig. 3.

3.4. Module trait relationships and identification of the most significant modules

Pearson's correlation between the module eigenlipids (MEs) and the studied markers was calculated. Three modules (turquoise, pink and yellow) were found to be significantly associated with several of both the osteoporotic and the atherosclerotic markers (Fig. 3). The turquoise module was significantly associated with the carotid-IMT-related variables *imtav* (r = 0.16, p-value = 2×10^{-10}) and *imtmax* (r = 0.16, p-value = 1×10^{-9}). The same module was also significantly associated with all of the pQCT bone measurements, the closest association being with *DTToMC* (r = 0.24, p-value = 2×10^{-20}). The pink and yellow modules were significantly associated with both bulbus IMT variables *bbav* (pink: r = 0.10, p-value = 7×10^{-5} , yellow: r = 0.11, p-

value = 1×10^{-5}) and *bbmax* (pink: r = 0.10, p-value = 8×10^{-5} , yellow: r = 0.12, p-value = 7×10^{-6}). The same modules were also significantly associated with five of the six pQCT-based subclinical osteoporosis indices. The exact lipid content of the most significant turquoise module is listed in Table S1 and explained under Section 3.6.

3.5. Lipid significance (LS) and module membership (MM)

LS is defined as the correlation between the module's member lipids and the study marker. MM is defined as the correlation between the eigenlipid and the other member lipids. An ideal module is the one where LS and MM are highly correlated suggesting that the lipids that are highly correlated with the biological marker of interest are also the important member of the analysed module [25]. Among the three significant modules (Fig. 4), the joint turquoise module has a highly significant correlation between LS and MM with respect to both subclinical atherosclerotic (*imtav*; r = 0.66, p-value = 1.9×10^{-14}) and subclinical osteoporotic (*DTToMC*; r = 0.64, p-value = 2×10^{-13}) markers (Fig. 3). The yellow module has a highly significant correlation between LS and MM only with respect to the subclinical atherosclerotic marker (*bbmax*; r = 0.49, p-value = .00013), whereas the pink module has no significant correlation between LS and MM with respect to any of the studied markers (data not shown).

3.6. Lipid species distribution in the joint turquoise module for subclinical osteoporotic and atherosclerosis markers

There were 105 lipid species in the joint turquoise module for subclinical markers of osteoporosis and atherosclerosis. The majority of the lipid species belonged to the classes of glycerolipid, glyceropho-spholipid and sphingolipid (Fig. 5A). The glycerolipid class included 19 diacylglycerol and 41 triacylglycerol (TAG) lipid species (Fig. 5B). The glycerophospholipid class had seven phosphatidylcholine lipid species, and the sphingolipid class was enriched with 20 ceramide species (Fig. 5B).

Table 1

Population characteristics of the Cardiovascular Risk in Young Finns Study cohort. Data are expressed as mean \pm SD or percentages.

	Men	Women
Number of subjects	646 (43%)	848 (57%)
Age, years	38 ± 5	38 ± 5
Body mass index, kg/m ²	26.5 ± 3.9	25.1 ± 4.7
Total cholesterol (mmol/l)	5.2 ± 0.9	4.9 ± 0.8
LDL cholesterol (mmol/l)	3.3 ± 0.8	3.0 ± 0.7
HDL cholesterol (mmol/l)	1.2 ± 0.3	1.5 ± 0.3
Triglycerides (mmol/l)	1.6 ± 0.9	1.2 ± 0.6
Serum glucose (mmol/l)	5.5 ± 0.6	5.2 ± 0.7
Insulin (IU/l)	9.9 ± 26.3	8.3 ± 8.6
C-reactive protein (mg/l)	1.6 ± 4.7	2.0 ± 3.5
Systolic blood pressure (mmHg)	125.2 13.1	116 ± 13.4
Diastolic blood pressure (mmHg)	78.3 ± 10.9	72.8 ± 10.7
Alcohol consumption, units/day	1.4 ± 1.9	0.6 ± 0.7
Physical activity index (MET h/wk)	20.4 ± 22.2	19.4 ± 20.1
Daily smoking, %	129/641 (20%)	121/843 (14%)
Daily calcium intake (mg)	1371 ± 602	1190 ± 483
Daily vitamin D intake (µg)	8.4 ± 4.5	7.3 ± 3.5
Family risk factor for Coronary Heart	107/646 (16.6%)	140/847 (16.5%)
Disease (%)		
Participants with osteoporosis (%)	3/641 (0.5%)	8/845 (1%)
Participants with epilepsy (%)	5/624 (0.8%)	7/835 (0.8%)
Participants with Crohn's disease (%)	5/625 (0.8%)	9/836 (1.1%)
Participants with Anorexia (%)	0	8/836 (1%)
Usage of corticosteroids at least once a	13/624 (2.1%)	54/837 (6.5%)
month (%)		

Table 2

Surrogate markers of both subclinical osteoporosis and subclinical atherosclerosis with their descriptive statistics among the study participants, expressed as mean \pm SD.

Description (unit)	Abbreviations	Mean (\pm SD)
Subclinical atherosclerosis		
Carotid intima-media thickness (average, mm)	imtav	0.6 ± 0.1
Carotid intima-media thickness (maximum, mm)	imtmax	0.7 ± 0.2
Bulbus intima-media thickness (average, mm)	bbav	0.8 ± 0.1
Bulbus intima-media thickness (maximum, mm)	bbmax	0.8 ± 0.1
Subclinical osteoporosis		
Total mineral density of the distal radius's trabecular bone (mg/cm ³)	DRTrD	224.4 ± 36.1
Total mineral density of the distal tibia's trabecular bone (mg/cm ³)	DTTrD	$240.3~\pm~34.1$
Total mineral content of the distal radius (mg)	DRToMC	243.6 ± 64.2
Total mineral content in the radial shaft's cortical bone (mg)	RSCoMC	214.2 ± 44.9
Total mineral content in the distal tibia (mg)	DTToMC	602.1 ± 126.9
Total mineral content in the tibia shaft's cortical bone (mg)	TSCoMC	646.4 ± 110.5

3.7. Multivariate analysis of the turquoise module and its constituent lipid species with subclinical markers of osteoporosis and atherosclerosis

In multivariate analysis of variance (adjusted with age and sex), average carotid intima media thickness (*imtav*) for subclinical atherosclerosis and total mineral content in the distal tibia (*DTToMC*) for subclinical osteoporosis were chosen as outcomes because they obtained the maximum correlation and the minimum p-value in a module–trait relationship analysis (Fig. 3). There was a statistically significant joint association between the turquoise eigenlipid and the

markers of subclinical osteoporosis and atherosclerosis, F(2, 1489) = 12.50, p-value = 4.1×10^{-6} , Pillais' Trace = 0.01. The turquoise eigenlipid had a statistically significant positive association with both markers (p-value with *imtav*: 2.7×10^{-6} and p-value with *DTToMC*: 0.03) in separate regression analyses.

Multivariate analysis of all the member lipid species in the turquoise module with *imtav* and *DTToMC* as outcomes, identified 37 lipid species that were jointly associated with the markers, with a Bonferroni-adjusted p-value of < 0.05 (Table S1). The three most significant joint biomarkers of both osteoporosis and atherosclerosis were TAG (18:0/18:1), TAG (18:0/18:1/18:1) and TAG (16:0/18:0/18:1), with adjusted p-values of 8.6 × 10^{-8} , 3.7×10^{-6} , and 8.5×10^{-6} , respectively.

In separate regression analyses of each member lipid species and *imtav*, 36 out of the 37 lipid species were found to be positively associated, with a Bonferroni-adjusted p-value of < 0.05 (Table S2). Similarly, regression analyses of each lipid species with *DTToMC* were also performed. All the 37 lipid species that were found to be jointly associated with markers of subclinical osteoporosis and atherosclerosis were positively associated with *DTToMC*; 16 of these were nominally significant (p-value < 0.05), but none of the lipid species reached a Bonferroni-adjusted p-value of 0.05 (Table S3).

4. Discussion

To the best of our knowledge, this is the first lipidome-wide systemlevel association study investigating the joint lipid architecture of surrogate markers of both subclinical osteoporosis and subclinical atherosclerosis. We performed lipidomics analysis to identify modules of lipid species that are significantly and jointly associated with the markers' of both of the studied comorbidities. We identified a shared module that is significantly associated with subclinical markers of both osteoporosis (pQCT bone measurements) and atherosclerosis



Fig. 2. Pearson's correlation coefficients (r) between surrogate markers of subclinical osteoporosis and atherosclerosis. All correlations are statistically significant (p-value < 0.01). The abbreviations in this figure are explained in Table 2.



Module-trait relationships

Fig. 3. Module-surrogate marker relationships. The rows correspond to the different modules and their eigenlipids (ME). The columns correspond to the measured subclinical osteoporotic and atherosclerotic markers of the study. The values in the cells represent Pearson's correlation coefficients (r), with the associated p-values in parentheses. The modules are named according to colour and the correlation coefficients have a colour-coding shown in the colour legend (between -1 and +1) on the right side of the figure. The abbreviations for the subclinical osteoporotic and atherosclerotic markers in the column names are explained in Table 2. (For interpretation of the references to colour in this figure, the reader is referred to the web version of this article.)



module membership (MM) in the turquoise and the vellow modules. The left panel corresponds to subclinical atherosclerotic markers and the right panel to subclinical osteoporotic markers. Abbreviations: imtav, carotid intima media thickness (average); bbmax, bulbus intima media thickness (maximum); DTToMC, total mineral content in distal tibia: DRToMC, total mineral content of the distal radius.

(ultrasound carotid IMT).

Whether osteoporosis and atherosclerosis are independent conditions that only share common risk factors, such as aging, or also constitute comorbid conditions with a similar pathophysiological mechanism is an active field of research [19,33]. Several studies have shown an association between decreased bone mass density and increased carotid IMT in different study groups [34-36]. Other studies

have suggested an association between osteoporosis and cardiovascular mortality [37-40]. Similarly, one study suggested that defects in bone mineralization and arterial calcification have a similar pathogenesis [41].

In contrast to most of the published findings, we identified weak, but statistically significant positive correlations between surrogate markers of these two diseases. The positive correlations might be due to



Fig. 5. Distribution of lipids classes (A) and constituent lipids (B) in the joint turquoise module for subclinical markers of osteoporosis and atherosclerosis.

the relatively younger age of the study participants who have not yet developed the clinical manifestations of the diseases. Thus, the positive associations might arise from the shared biological mechanisms between bone and vascular tissue during their normal growth and development. Similar results have been published elsewhere [42]. We speculate that the dynamics of the lipid molecules that are associated with both bone-related and vascular markers change during adverse conditions leading to the comorbidity. Knowledge of the lipid molecules that are associated with the surrogate markers of both the diseases is crucial for identifying alterations in molecular dynamics that take place during the disease. This will not only confirm whether or not the diseases are comorbid but can also potentially improve the risk stratification, prevention, diagnosis and treatment of the diseases.

The majority of the lipid species in the most significant joint module belonged to the glycerolipid, glycerophospholipid and sphingolipid classes. Within glycerophospholipids, one of the phosphatidylcholine lipid species, namely lysophosphatidylcholine (LPC), is a pro-inflammatory lipid that is generated by various pathological activities and is a major component of oxidized low-density lipoprotein (LDL) [43]. Oxidized LDL is known to be a potential factor for the co-occurrence of vascular calcification with the loss of bone mass [44]. Studies have shown that oxidized LDL promotes atherosclerosis via a chemotactic and proliferative mechanism on monocytes by stimulating their adhesion into the endothelial cells and by initiating the formation of foam cells [45]. Oxidized-LDL has also been shown to proliferate and stimulate the migration of smooth muscle cells into the tunica media, which stimulates the production of collagen, thus contributing the fibrous lining in the atherosclerosis plaque [46]. Studies have also suggested that oxidized LDL inhibits osteoblastic differentiation and bone formation and promotes osteoblast cell death [47,48]. A recent study suggested that there is a causal effect of LDL cholesterol on bone mass density [49]. However, clinical findings related to oxidized LDL in the context of cardiovascular diseases have been controversial [50-52].

The identified joint module includes high-risk cardiovascular ceramides among 20 other ceramides, which confirms their previously shown association with cardiovascular outcomes [53,54] and suggests their potential role in subclinical osteoporosis as well. Ceramides are responsible for the activation of NF- κ B (nuclear factor kappa-lightchain-enhancer of activated B cells) that causes the apoptosis of bone cells [55]. A study has demonstrated an association between ceramides and trabecular bone density in mice [56]. In addition, sphingomyelins have been found to be decreased in the bone tissue of mice with osteoporosis [57]. Ceramides also promote lipoprotein infiltration into the vessel wall by acting as a key signalling molecule [58].

An earlier study has shown a significantly increased level of cholesterol ester in the arterial wall of atherosclerotic lesions [59]. An elevated level of triglycerides is known to be an important biomarker in the development of cardiovascular disease [60]. Lipoproteins that carry triglycerides in the blood stream accumulate in the artery wall intima and are taken up by macrophages to form foam cells that contribute to the build-up of plaque along the walls of artery [61]. The triglyceride metabolism in bone tissue has been shown to diminish in subjects with osteoporosis, when compared with the healthy controls [62]. Furthermore, a study with middle-aged women in Japan revealed that patients with hypertriglyceridemia had reduced bone resorption and were at risk of fractures [63]. Furthermore, among the 37 joint lipid species identified herein by multivariate analysis of variance as being significantly associated with both osteoporosis and atherosclerosis, the top three were triglycerides namely TAG(18:0/18:0/18:1), TAG(18:0/18:1/18:1) and TAG(16:0/18:0/18:1). A previous study has shown an association between TAG(18:0/18:1/18:1) and cardiovascular disease [64]. Furthermore, TAG(16:0/18:0/18:1) has been linked to a faster progression of type 2 diabetes [65] which is a risk factor for both cardiovascular disease [66] and bone fractures [67].

This study is limited to the subclinical phase of atherosclerosis and osteoporosis, as it is based on a relatively young cohort population with very few diagnosed cases of cardiovascular disease and osteoporosis. Therefore, further research on lipidome-wide associations with clinical comorbidities in a case–control setting is crucial. Furthermore, as all of the participants of this study are of Caucasian origin, studies with populations of different ethnicities are needed.

5. Summary and conclusion

Several earlier studies have shown that osteoporosis and atherosclerosis are comorbid conditions, emphasizing that these conditions should be investigated in detail to identify common risk factors and joint molecular mechanisms and to develop common methods for risk stratification, prevention, diagnosis and treatment. In the present study, we identified a lipidome module, with its specific molecular lipids, that was significantly associated with surrogate markers of the subclinical phase of both osteoporosis and atherosclerosis. Alteration in the metabolism of the identified lipid species might contribute to the comorbid conditions and yield new possibilities for their dual-based prevention methods.

Funding

The Cardiovascular Risk in Young Finns Study has been financially supported by the Academy of Finland (grants 322098, 286284, 134309 [Eye], 126925, 121584, 124282, 129378 [Salve], 117787 [Gendi], and 41071 [Skidi]); the Social Insurance Institution of Finland; Competitive State Research Financing of the Expert Responsibility area of Kuopio, Tampere and Turku University Hospitals (grant X51001); the Juho Vainio Foundation; the Paavo Nurmi Foundation; the Finnish Foundation for Cardiovascular Research; the Finnish Cultural Foundation; the Sigrid Jusélius Foundation; the Tampere Tuberculosis Foundation; the Emil Aaltonen Foundation; the Yrjö Jahnsson Foundation; the Signe and Ane Gyllenberg Foundation; as well as the EU Horizon 2020 programme (grant 755320 for TAXINOMISIS); and the European Research Council (grant 742927 for MULTIEPIGEN project); and the foundation Tays tukisäätiö.

Ethical approval

The study was approved by the ethical committee of the Hospital District of Southwest Finland on 20 June 2017 (ETMK:68/1801/2017), and all participants have given an informed written consent. Data protection will be handled according to current regulations.

Declaration of competing interest

None.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.bone.2019.115160.

References

- [1] G.S. Berenson, S.R. Srinivasan, W. Bao, W.P. Newman, R.E. Tracy, W.A. Wattigney, Association between multiple cardiovascular risk factors and atherosclerosis in children and young adults. The Bogalusa Heart Study, N. Engl. J. Med. 338 (23) (1998) 1650–1656, https://doi.org/10.1056/NEJM199806043382302.
- [2] S.R. Cummings, L.J. Melton, Osteoporosis I: epidemiology and outcomes of osteoporotic fractures, Lancet (2002, May 18), https://doi.org/10.1016/S0140-6736(02) 08657-9 Elsevier Limited.
- [3] L. Jørgensen, O. Joakimsen, G.K.R. Berntsen, I. Heuch, B.K. Jacobsen, Low bone mineral density is related to echogenic carotid artery plaques: a population-based study, Am. J. Epidemiol. 160 (6) (2004) 549–556, https://doi.org/10.1093/aje/ kwh252.
- [4] E. Hernlund, A. Svedbom, M. Ivergård, J. Compston, C. Cooper, J. Stenmark, ... J.A. Kanis, Osteoporosis in the European Union: Medical management, epidemiology and economic burden: A report prepared in collaboration with the International Osteoporosis Foundation (IOF) and the European Federation of Pharmaceutical Industry Associations (EFPIA), Archives of Osteoporosis 8 (1–2) (2013), https://doi.org/10.1007/s11657-013-0136-1.
- [5] G.N. Farhat, J.A. Cauley, The link between osteoporosis and cardiovascular disease, Clin. Cases Miner. Bone Metab. 5 (1) (2008) 19–34.
- [6] N.L. Kim, H.M. Jang, S.K. Kim, K.D. Ko, I.C. Hwang, H.S. Suh, Association of arterial stiffness and osteoporosis in healthy men undergoing screening medical examination, J. Bone Metab. 21 (2) (2014) 133, https://doi.org/10.11005/jbm.2014.21.2. 133.
- [7] J.A. Hyder, M.A. Allison, N. Wong, A. Papa, T.F. Lang, C. Sirlin, ... M.H. Criqui, Association of coronary artery and aortic calcium with lumbar bone density: The MESA abdominal aortic calcium study, American Journal of Epidemiology 169 (2) (2009) 186–194, https://doi.org/10.1093/aje/kwn303.
- [8] N. Campos-Obando, M. Kavousi, J.E. Roeters van Lennep, F. Rivadeneira, A. Hofman, A.G. Uitterlinden, ... M.C. Zillikens, Bone health and coronary artery calcification: The Rotterdam Study, Atherosclerosis 241 (1) (2014) 278–283, https://doi.org/10.1016/j.atherosclerosis.2015.02.013.
- [9] N. Oksala, M. Levula, M. Pelto-Huikko, L. Kytömïki, J.T. Soini, J. Salenius, ... T. Lehtimäki, Carbonic anhydrases II and XII are up-regulated in osteoclast-like cells in advanced human atherosclerotic plaques—Tampere Vascular Study, Annals of Medicine 42 (5) (2010) 360–370, https://doi.org/10.3109/07853890.2010. 486408.
- [10] J.P. Karjalainen, N. Mononen, N. Hutri-Kähönen, M. Lehtimäki, M. Juonala, M. Ala-Korpela, ... T. Lehtimäki, The effect of apolipoprotein E polymorphism on serum metabolome – a population-based 10-year follow-up study, Scientific Reports 9 (1) (2019), https://doi.org/10.1038/s41598-018-36450-9.
- [11] E. Ilveskoski, M. Perola, T. Lehtimäki, P. Laippala, V. Savolainen, J. Pajarinen, ... P.J. Karhunen, Age-dependent association of apolipoprotein E genotype with coronary and aortic atherosclerosis in middle-aged men: An autopsy study, Circulation 100 (6) (1999) 608–613, https://doi.org/10.1161/01.CIR.100.6.608.
- [12] S. Tolonen, V. Mikkilä, M. Laaksonen, H. Sievänen, N. Mononen, J. Hernesniemi, ... T. Lehtimäki, Association of apolipoprotein E promoter polymorphisms with bone structural traits is modified by dietary saturated fat intake - The Cardiovascular Risk in Young Finns Study, Bone 48 (5) (2011) 1058–1065, https://doi.org/10.1016/j. bone.2011.01.013.
- [13] C.J. Edwards, D.J. Hart, T.D. Spector, Oral statins and increased bone-mineral density in postmenopausal women, Lancet 355 (9222) (2000) 2218–2219, https:// doi.org/10.1016/S0140-6736(00)02408-9.
- [14] K. Bleicher, R.G. Cumming, V. Naganathan, M.J. Seibel, P.N. Sambrook, F.M. Blyth, ... L.M. Waite, Lifestyle factors, medications, and disease influence bone mineral density in older men: Findings from the CHAMP study, Osteoporosis International 22 (9) (2011) 2421–2437, https://doi.org/10.1007/s00198-010-1478-9.
- [15] M.E. Mussolino, H.K. Armenian, Low bone mineral density, coronary heart disease,

and stroke mortality in men and women: the third National Health and Nutrition Examination Survey, Ann. Epidemiol. 17 (11) (2007) 841–846, https://doi.org/10.1016/j.annepidem.2007.06.005.

- [16] M.E. Mussolino, J.H. Madans, R.F. Gillum, Bone mineral density and stroke, Stroke 34 (5) (2003), https://doi.org/10.1161/01.STR.0000065826.23815.A5.
- [17] W.S. Browner, A.R. Pressman, M.C. Nevitt, J.A. Cauley, S.R. Cummings, S Cummings, ... B. Packer, Association between low bone density and stroke in elderly women: The study of osteoporotic fractures, Stroke 24 (7) (1993) 940–946, https://doi.org/10.1161/01.STR.24.7.940.
- [18] T.M. Doherty, L.A. Fitzpatrick, D. Inoue, J.H. Qiao, M.C. Fishbein, R.C. Detrano, ... T.B. Rajavashisth, Molecular, Endocrine, and Genetic Mechanisms of Arterial Calcification, Endocrine Reviews (2004, August), https://doi.org/10.1210/er.2003-0015.
- [19] P. Anagnostis, A. Karagiannis, A.I. Kakafika, K. Tziomalos, V.G. Athyros, D.P. Mikhailidis, Atherosclerosis and osteoporosis: age-dependent degenerative processes or related entities? Osteoporos. Int. (2009), https://doi.org/10.1007/ s00198-008-0648-5 shaffersh.
- [20] O.I. Stojanovic, M. Lazovic, M. Lazovic, M. Vuceljic, Association between atherosclerosis and osteoporosis, the role of vitamin D, Arch. Med. Sci. (2011), https:// doi.org/10.5114/aoms.2011.22066.
- [21] S.J. Chen, C.S. Lin, C.L. Lin, C.H. Kao, Osteoporosis is associated with high risk for coronary heart disease: a population-based cohort study, Medicine 94 (27) (2015) e1146, https://doi.org/10.1097/MD.00000000001146.
- [22] K. Yang, X. Han, Lipidomics: techniques, applications, and outcomes related to biomedical sciences, Trends in Biochemical Sciences, Elsevier Ltd., 2016, https:// doi.org/10.1016/j.tibs.2016.08.010.
- [23] C. Wadsack, T. Stojakovic, R. Birner-Gruenberger, V. Binder, A. Heinemann, G. Marsche, D. El-Gamal, Uremia alters HDL composition and function, J. Am. Soc. Nephrol. 22 (9) (2011) 1631–1641, https://doi.org/10.1681/asn.2010111144.
- [24] C. Xu, D. Zhou, Y. Luo, S. Guo, T. Wang, J. Liu, ... Z. Li, Tissue and serum lipidome shows altered lipid composition with diagnostic potential in mycosis fungoides, Oncotarget 8 (29) (2017), https://doi.org/10.18632/oncotarget.18228.
- [25] P. Langfelder, S. Horvath, WGCNA: an R package for weighted correlation network analysis, BMC Bioinf. 9 (2008) 559, https://doi.org/10.1186/1471-2105-9-559.
- [26] O.T. Raitakari, M. Juonala, T. Rönnemaa, L. Keltikangas-Järvinen, L. Räsänen, M. Pietikäinen, ... J.S.A. Viikari, Cohort profile: The cardiovascular risk in Young Finns Study, International Journal of Epidemiology 37 (6) (2008) 1220–1226, https://doi.org/10.1093/ije/dym225.
- [27] M.M.L. Laaksonen, H. Sievänen, S. Tolonen, V. Mikkilä, L. Räsänen, J. Viikari, ... O.T. Raitakari, Determinants of bone strength and fracture incidence in adult Finns: Cardiovascular Risk in Young Finns Study (the GENDI pQCT study), Archives of Osteoporosis 5 (1–2) (2010) 119–130, https://doi.org/10.1007/s11657-010-0043-7.
- [28] K.S. Pälve, K. Pahkala, C.G. Magnussen, T. Koivistoinen, M. Juonala, M. Kähönen, ... O.T. Raitakari, Association of physical activity in childhood and early adulthood with carotid artery elasticity 21 years later: The cardiovascular risk in Young Finns Study, Journal of the American Heart Association 3 (2) (2014), https://doi.org/10. 1161/JAHA.113.000594.
- [29] M. Juonala, J.S.A. Viikari, M. Kähönen, T. Laitinen, L. Taittonen, B.M. Loo, ... O.T. Raitakari, Alcohol consumption is directly associated with carotid intimamedia thickness in Finnish young adults. The Cardiovascular Risk in Young Finns Study, Atherosclerosis 204 (2) (2009), https://doi.org/10.1016/j.atherosclerosis. 2008.11.021.
- [30] G. Wong, C.K. Barlow, J.M. Weir, J.B.M. Jowett, D.J. Magliano, ... P.Meikle Zimmet, Inclusion of Plasma Lipid Species Improves Classification of Individuals at Risk of Type 2 Diabetes, PLoS ONE 8 (10) (2013), https://doi.org/10.1371/journal.pone. 0076577.
- [31] E.I. Braicu, S. Darb-Esfahani, W.D. Schmitt, K.M. Koistinen, L. Heiskanen, P. Pöhö, ... M. Hilvo, High-grade ovarian serous carcinoma patients exhibit profound alterations in lipid metabolism, Oncotarget 8 (61) (2017) 102912–102922, https:// doi.org/10.18632/oncotarget.22076.
- [32] R Core Team, R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing, Vienna, Austria, 2013 URL http://www.Rproject.org/.
- [33] D. Hamerman, Osteoporosis and atherosclerosis: biological linkages and the emergence of dual-purpose therapies, QJM (2005), https://doi.org/10.1093/ qjmed/hci077.
- [34] J.R. Shaffer, C.M. Kammerer, D.L. Rainwater, D.H. O'Leary, J.M. Bruder, R.L. Bauer, B.D. Mitchell, Decreased bone mineral density is correlated with increased subclinical atherosclerosis in older, but not younger, Mexican American women and men: the San Antonio family Osteoporosis Study, Calcif. Tissue Int. 81 (6) (2007) 430–441, https://doi.org/10.1007/s00223-007-9079-0.
- [35] J. Tamaki, M. Iki, Y. Hirano, Y. Sato, E. Kajita, S. Kagamimori, ... H. Yoneshima, Low bone mass is associated with carotid atherosclerosis in postmenopausal women: The Japanese Population-based Osteoporosis (JPOS) Cohort Study, Osteoporosis International 20 (1) (2009) 53–60, https://doi.org/10.1007/s00198-008-0633-z.
- [36] S.N. Kim, H.S. Lee, H.S. Nam, H.R. Lee, J.M. Kim, S.W. Han, ... J.H. Park, Carotid Intima-Media Thickness is Inversely Related to Bone Density in Female but not in Male Patients with Acute Stroke, Journal of Neuroimaging 26 (1) (2016) 83–88, https://doi.org/10.1111/jon.12284.
- [37] P. Von der Recke, M.A. Hansen, C. Hassager, The association between low bone mass at the menopause and cardiovascular mortality, Am. J. Med. 106 (3) (1999) 273–278, https://doi.org/10.1016/S0002-9343(99)00028-5.
- [38] P.A. Marcovitz, H.H. Tran, B.A. Franklin, W.W. O'Neill, M. Yerkey, J. Boura, ... C.Z. Dickinson, Usefulness of bone mineral density to predict significant coronary

artery disease, American Journal of Cardiology 96 (8) (2005) 1059–1063, https://doi.org/10.1016/j.amjcard.2005.06.034.

- [39] D.M. Kado, W.S. Browner, T. Blackwell, R. Gore, S.R. Cummings, Rate of bone loss is associated with mortality in older women: a prospective study, J. Bone Miner. Res. 15 (10) (2000) 1974–1980, https://doi.org/10.1359/jbmr.2000.15.10.1974.
- [40] W.S. Browner, D.G. Seeley, T.M. Vogt, S.R. Cummings, Non-trauma mortality in elderly women with low bone mineral density. Study of Osteoporotic Fractures Research Group, Lancet 338 (8763) (1991) 355–358 Retrieved from http://www. ncbi.nlm.nih.gov/pubmed/1677708.
- [41] T.M. Doherty, K. Asotra, L.A. Fitzpatrick, J.-H. Qiao, D.J. Wilkin, R.C. Detrano, ... T.B. Rajavashisth, Calcification in atherosclerosis: bone biology and chronic inflammation at the arterial crossroads, Proceedings of the National Academy of Sciences of the United States of America 100 (20) (2003) 11201, https://doi.org/ 10.1073/pnas.1932554100 6.
- [42] M. Frysz, K. Deere, D.A. Lawlor, L. Benfield, J.H. Tobias, C.L. Gregson, Bone mineral density is positively related to carotid intima-media thickness: findings from a population-based study in adolescents and premenopausal women, J. Bone Miner. Res. 31 (12) (2016) 2139–2148, https://doi.org/10.1002/jbmr.2903.
- [43] G. Schmitz, K. Ruebsaamen, Metabolism and atherogenic disease association of lysophosphatidylcholine, Atherosclerosis (2010), https://doi.org/10.1016/j. atherosclerosis.2009.05.029.
- [44] R. Wu, Y.H. Huang, L.S. Elinder, J. Frostegård, Lysophosphatidylcholine is involved in the antigenicity of oxidized LDL, Arterioscler. Thromb. Vasc. Biol. 18 (4) (1998) 626–630, https://doi.org/10.1161/01.ATV.18.4.626.
- [45] T. Matsumoto, T. Kobayashi, K. Kamata, Role of lysophosphatidylcholine (LPC) in atherosclerosis, Curr. Med. Chem. 14 (30) (2007) 3209–3220, https://doi.org/10. 2174/092986707782793899.
- [46] A. Stiko-Rahm, A. Hultgårdh-Nilsson, J. Regnström, A. Hamsten, J. Nilsson, Native and oxidized LDL enhances production of PDGF AA and the surface expression of PDGF receptors in cultured human smooth muscle cells, Arterioscler. Thromb. Vasc. Biol. 12 (9) (1992) 1099–1109, https://doi.org/10.1161/01.ATV.12.9.1099.
- [47] F. Parhami, A.D. Morrow, J. Balucan, N. Leitinger, A.D. Watson, Y. Tintut, ... L.L. Demer, Lipid oxidation products have opposite effects on calcifying vascular cell and bone cell differentiation: A possible explanation for the paradox of arterial calcification in osteoporotic patients, Arteriosclerosis, Thrombosis, and Vascular Biology 17 (4) (1997) 680–687, https://doi.org/10.1161/01.ATV.17.4.680.
- [48] M.R. Brodeur, L. Brissette, L. Falstrault, V. Luangrath, R. Moreau, Scavenger receptor of class B expressed by osteoblastic cells are implicated in the uptake of cholesteryl ester and estradiol from LDL and HDL3, J. Bone Miner. Res. 23 (3) (2008) 326–337, https://doi.org/10.1359/jbmr.071022.
- [49] X.L. Yang, Z.Z. Cui, H. Zhang, X.T. Wei, G.J. Feng, L. Liu, L. Zhang, Causal link between lipid profile and bone mineral density: a Mendelian randomization study, Bone (2019), https://doi.org/10.1016/j.bone.2019.05.037.
- [50] Y.K. Lee, D.H. Lee, J.K. Kim, M.J. Park, J.J. Yan, D.K. Song, ... J.W. Noh, Lysophosphatidylcholine, oxidized low-density lipoprotein and cardiovascular disease in Korean hemodialysis patients: Analysis at 5 years of follow-up, Journal of Korean Medical Science 28 (2) (2013) 268–273, https://doi.org/10.3346/jkms. 2013.28.2.268.
- [51] C. Fernandez, M. Sandin, J.L. Sampaio, P. Almgren, K. Narkiewicz, ... M.Melander Hoffmann, Plasma Lipid Composition and Risk of Developing Cardiovascular Disease, PLoS ONE 8 (8) (2013), https://doi.org/10.1371/journal. pone.0071846.
- [52] A. Ganna, S. Salihovic, J. Sundström, C.D. Broeckling, Å.K. Hedman, P.K.E. Magnusson, ... E. Ingelsson, Large-scale metabolomic Profiling Identifies Novel Biomarkers for Incident Coronary Heart Disease, PLoS Genetics 10 (12) (2014), https://doi.org/10.1371/journal.pgen.1004801.

- [53] R. Laaksonen, K. Ekroos, M. Sysi-Aho, M. Hilvo, T. Vihervaara, D. Kauhanen, ... T.F. Lüscher, Plasma ceramides predict cardiovascular death in patients with stable coronary artery disease and acute coronary syndromes beyond LDL-cholesterol, European Heart Journal 37 (25) (2016) 1967–1976, https://doi.org/10.1093/ eurheartj/ehv148.
- [54] J.W. Meeusen, L.J. Donato, S.C. Bryant, L.M. Baudhuin, P.B. Berger, A.S. Jaffe, Plasma ceramides: a novel predictor of major adverse cardiovascular events after coronary angiography, Arterioscler. Thromb. Vasc. Biol. 38 (8) (2018) 1933–1939, https://doi.org/10.1161/ATVBAHA.118.311199.
- [55] I. Kitajima, Y. Soejima, I. Takasaki, H. Beppu, T. Tokioka, I. Maruyama, Ceramideinduced nuclear translocation of NF-κB is a potential mediator of the apoptotic response to TNF-α in murine clonal osteoblasts, Bone 19 (3) (1996) 263–270, https://doi.org/10.1016/8756-3282(96)00181-0.
- [56] Z. Saeed, C. Guilbault, J.B. De Sanctis, J. Henri, D. Marion, R. St-Arnaud, D. Radzioch, Fenretinide prevents the development of osteoporosis in Cftr-KO mice, J. Cyst. Fibros. 7 (3) (2008) 222–230, https://doi.org/10.1016/j.jcf.2007.09.001.
- [57] H. Zhao, X. Li, D. Zhang, H. Chen, Y. Chao, K. Wu, J. Su, Integrative bone metabolomics—lipidomics strategy for pathological mechanism of postmenopausal osteoporosis mouse model, Sci. Rep. 8 (1) (2018), https://doi.org/10.1038/s41598-018-34574-6.
- [58] W. Li, X. Yang, S. Xing, F. Bian, W. Yao, X. Bai, S. Jin, Endogenous ceramide contributes to the transcytosis of oxLDL across endothelial cells and promotes its subendothelial retention in vascular wall, Oxidative Med. Cell. Longev. 2014 (2014) 1–11, https://doi.org/10.1155/2014/823071.
- [59] E.B. Smith, The relationship between plasma and tissue lipids in human atherosclerosis, Advances in Lipid Research, 12 Elsevier, 1974, pp. 1–49, , https://doi. org/10.1016/B978-0-12-024912-1.50008-9.
- [60] R. Do, C.J. Willer, E.M. Schmidt, S. Sengupta, C. Gao, G.M. Peloso, ... S. Kathiresan, Common variants associated with plasma triglycerides and risk for coronary artery disease, Nature Genetics (2013, November), https://doi.org/10.1038/ng.2795.
- [61] K.M. Botham, E.H. Moore, C.D. Pascale, F. Bejta, The induction of macrophage foam cell formation by chylomicron remnants, Biochem. Soc. Trans. 35 (3) (2007) 454–458, https://doi.org/10.1042/bst0350454.
- [62] J. Dragojevič, J. Zupan, G. Haring, S. Herman, R. Komadina, J. Marc, Triglyceride metabolism in bone tissue is associated with osteoblast and osteoclast differentiation: a gene expression study, J. Bone Miner. Metab. 31 (5) (2013) 512–519, https://doi.org/10.1007/s00774-013-0445-x.
- [63] K. Ando, A. Tanaka, T. Tazaki, T. Yokoe, K. Okuda, T. Ohnishi, ... S. Inoue, Association between Casual Serum Triglyceride Levels and Bone resorption Activity in Japanese Middle-aged and Elderly Women, Showa Univ. J. Med. Sci. 28 (4) (2017) 349–357, https://doi.org/10.15369/sujms.28.349.
- [64] C. Stegemann, R. Pechlaner, P. Willeit, S.R. Langley, M. Mangino, U. Mayr, ... M. Mayr, Lipidomics profiling and risk of cardiovascular disease in the prospective population-based Bruneck study, Circulation 129 (18) (2014) 1821–1831, https:// doi.org/10.1161/CIRCULATIONAHA.113.002500.
- [65] M. Mamtani, H. Kulkarni, G. Wong, J.M. Weir, C.K. Barlow, T.D. Dyer, ... J.E. Curran, Lipidomic risk score independently and cost-effectively predicts risk of future type 2 diabetes: Results from diverse cohorts, Lipids in Health and Disease 15 (1) (2016), https://doi.org/10.1186/s12944-016-0234-3.
- [66] T.R. Einarson, A. Acs, C. Ludwig, U.H. Panton, Prevalence of cardiovascular disease in type 2 diabetes: a systematic literature review of scientific evidence from across the world in 2007–2017, Cardiovascular Diabetology, BioMed Central Ltd., 2018, June 8, https://doi.org/10.1186/s12933-018-0728-6.
- [67] P. Vestergaard, Discrepancies in bone mineral density and fracture risk in patients with type 1 and type 2 diabetes - a meta-analysis, Osteoporos. Int. 18 (4) (2007) 427–444, https://doi.org/10.1007/s00198-006-0253-4.