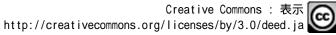




Complete Genome Sequence of an Anaerobic Benzene-Degrading Bacterium, Azoarcus sp. Strain DN11

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ABSTRACT Here, we present the complete genome sequence of *Azoarcus* sp. strain DN11, a denitrifying bacterium capable of anaerobic benzene degradation. The DN11 genome is 4,956,835 bp long with a G+C content of 66.3%. Genome analysis suggested the possibility that DN11 utilizes three proposed pathways for anaerobic benzene degradation, namely, methylation, hydroxylation, and carboxylation pathways.

A *zoarcus* sp. strain DN11 (NBRC 108892) is a denitrifying, facultative anaerobic bacterium isolated from gasoline-contaminated groundwater in Kumamoto, Japan (1). DN11 can anaerobically degrade benzene and other aromatic hydrocarbons, such as toluene and *m*-xylene, under denitrifying conditions (2). These biodegradative capabilities of DN11 demonstrate its potential for bioremediation of benzene-contaminated sites under oxygen-limited conditions where benzene is persistent (3).

Three anaerobic benzene-degradation pathways have been proposed based on the metabolites identified in benzene-degrading cultures, namely, methylation to toluene, hydroxylation to phenol, and carboxylation to benzoate (4). Toluene, phenol, and benzoate are further metabolized to a common intermediate, benzoyl-coenzyme A (CoA) (4). However, enzymes involved in the three pathways have not yet been fully elucidated. Here, we report the complete genome sequence of DN11 to better understand its biochemical pathway of anaerobic benzene degradation which might involve novel genes.

DN11 was grown in diluted casitone glycerol yeast autolysate (dCGY) medium (5), and its DNA was extracted and purified using a Wizard genomic DNA purification kit (Promega). TruSeq DNA PCR-free and Nextera mate-pair library preparation kits (Illumina) were used to generate a 1,300-bp paired-end (PE) PCR-free library and a 580-bp mate-pair (MP) library with an initial fragment size of 7,100 bp. Libraries were loaded into a 600-cycle V3 chemistry sequencing cartridge, and 300 bp of each library end was sequenced with a MiSeq instrument (Illumina). All kits were used following the manufacturer's instructions. Newbler 2.8 (Roche) was used for genome assembly (default parameters, except for overlapMinMatchIdentity of 98 and allContigThresh of 0), with 1 million PE (260 Mb) and 0.67 million MP (120 Mb) reads that were quality trimmed using ShortReadManager 0.995 (score_threshold, 30; length_threshold, 21; and 21-mers occurring more than twice regarded as valid) (6). *In silico* gap closing of the obtained scaffolds (67 in total) was done manually with AceFileViewer 1.5 and GenoFinisher 2.1 (default settings) (6) using the MP data, resulting in the complete resolution of

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Received 18 December 2018 Accepted 17 February 2019 Published 14 March 2019 repeat-induced gaps and unambiguous assembly of a single replicon. This sequence was checked for errors derived from the gap closing step with FinishChecker (an accessory of GenoFinisher) (7). Gene prediction and annotation were done using the Prokaryotic Genome Annotation Pipeline (PGAP) (8) and Microbial Genome Annotation Pipeline (MiGAP) (9), and annotated sequence files were first compared using Genome-Matcher 2.203 (10) to manually correct start/stop codon inconsistencies and then merged into a single annotated sequence file.

The chromosome of strain DN11 is 4,956,835 bp long ($77 \times$ coverage), with a G+C content of 66.3%. Also, 4,593 coding DNA sequences (CDSs), 4 copies of the rRNA operon, and 57 tRNA genes were identified. A BLAST search of the DN11 genome revealed two *bssABCD* gene clusters involved in the anaerobic conversion of toluene to benzylsuccinate, with amino acid identities ranging from 49% to 71% compared with another anaerobic benzene-degrading bacterium, *Geobacter metallireducens* (11, 12). The presence of a *ppsABC* gene cluster for anaerobic conversion of phenol to phenylphosphate was identified, with 39% to 77% amino acid identities. Lastly, 3 *bamY* genes for anaerobic conversion of benzoate to benzoyl-CoA had 44%, 49%, and 50% amino acid identities. They indicated the possibility that to degrade benzene under anaerobic conditions, DN11 may use all three pathways, namely, methylation, carboxylation, and hydroxylation pathways.

Data availability. The complete genome sequence of *Azoarcus* sp. strain DN11 has been deposited at DDBJ/ENA/GenBank under the accession number CP021731 (software parameters and details of the use and rationale of bioinformatics tools for the completion of the genome sequence are available in the "comment" section). Raw sequencing data have been deposited in the SRA under the accession number PRJNA388763.

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REFERENCES

- Kasai Y, Takahata Y, Manefield M, Watanabe K. 2006. RNA-based stable isotope probing and isolation of anaerobic benzene-degrading bacteria from gasoline-contaminated groundwater. Appl Environ Microbiol 72: 3586–3592. https://doi.org/10.1128/AEM.72.5.3586-3592.2006.
- Kasai Y, Kodama Y, Takahata Y, Hoaki T, Watanabe K. 2007. Degradative capacities and bioaugmentation potential of an anaerobic benzenedegrading bacterium strain DN11. Environ Sci Technol 41:6222–6227. https://doi.org/10.1021/es062842p.
- Weelink SAB, van Eekert MHA, Stams AJM. 2010. Degradation of BTEX by anaerobic bacteria: physiology and application. Rev Environ Sci Biotechnol 9:359–385. https://doi.org/10.1007/s11157-010-9219-2.
- Vogt C, Kleinsteuber S, Richnow H-H. 2011. Anaerobic benzene degradation by bacteria. Microb Biotechnol 4:710–724. https://doi.org/10 .1111/j.1751-7915.2011.00260.x.
- Watanabe K, Teramoto M, Futamata H, Harayama S. 1998. Molecular detection, isolation, and physiological characterization of functionally dominant phenol-degrading bacteria in activated sludge. Appl Environ Microbiol 64:4396–4402.
- Ohtsubo Y, Maruyama F, Mitsui H, Nagata Y, Tsuda M. 2012. Complete genome sequence of *Acidovorax* sp. strain KKS102, a polychlorinatedbiphenyl degrader. J Bacteriol 194:6970–6971. https://doi.org/10.1128/ JB.01848-12.
- 7. Ohtsubo Y, Kishida K, Sato T, Tabata M, Kawasumi T, Ogura Y, Hayashi T,

Tsuda M, Nagata Y. 2014. Complete genome sequence of *Pseudomonas* sp. strain TKP, isolated from a γ -hexachlorocyclohexane-degrading mixed culture. Genome Announc 2:e01241-13. https://doi.org/10.1128/genomeA.01241-13.

- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi .org/10.1093/nar/gkw569.
- 9. Sugawara H, Ohyama A, Mori H, Kurokawa K. 2009. Microbial Genome Annotation Pipeline (MiGAP) for diverse users, abstr S-001, p 1–2. Abstr. 20th Int Conf Genome Informatics, Kanagawa, Japan.
- Ohtsubo Y, Ikeda-Ohtsubo W, Nagata Y, Tsuda M. 2008. GenomeMatcher: a graphical user interface for DNA sequence comparison. BMC Bioinformatics 9:376. https://doi.org/10.1186/1471-2105-9-376.
- Aklujkar M, Krushkal J, DiBartolo G, Lapidus A, Land ML, Lovley DR. 2009. The genome sequence of *Geobacter metallireducens*: features of metabolism, physiology and regulation common and dissimilar to *Geobacter sulfurreducens*. BMC Microbiol 9:109. https://doi.org/10.1186/1471-2180 -9-109.
- Zhang T, Tremblay P-L, Chaurasia AK, Smith JA, Bain TS, Lovley DR. 2013. Anaerobic benzene oxidation via phenol in *Geobacter metallireducens*. Appl Environ Microbiol 79:7800–7806. https://doi.org/10.1128/AEM .03134-13.