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Pure Isolates and Preparative HPLC Fractions or Crude Extract of *Inula viscosa*: Effect on β -hematin Inhibition *in vitro*

Abstract

The antimalarial activity of wild *Inula viscosa* (*I. viscosa*) plant leaves was investigated. The efficacy of the whole crude extract versus preparative HPLC fractions versus pure isolates were compared by measuring their effect on β -hematin inhibition *in-vitro*. The preparative HPLC experiments consisted of reversed phase preparative column (22.2mm x 250mm, 10 μ m) and linear gradient of water, acetonitrile as the mobile phase. Injection volume was 1000 μ l and the wavelengths range were from 200-450 nm using photodiode array detector (PDA). While fractions (IV, V and VI) showed antimalarial potential in comparison to chloroquine positive control, the rest of the fractions did not show any significant inhibition to the β -hematin formation. The antimalarial results showed that whole crude extract of the plant works better than the preparative fractions or the pure isolates presumably due to synergistic effect. The chemical identity of some of the pure isolates was explored using UHPLC-ESI-MS. Moreover, *I. viscosa* extract powder stayed stable over several years, while many other products such as *Artemisia annua* extract or Artemisinin Combined Therapy (ACT) drugs rapidly lost their efficiency under tropical storage conditions.

Keywords: *Inula viscosa*; Malaria; Hemozoin; β -hematin; Preparative HPLC; LC-MS

Research Article

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Introduction

Malaria is still one of the largest medical threats to humans worldwide. Despite the global efforts to control it, many areas around the world remain malaria endemic, mainly confined to poor tropics and subtropics with huge global economic burdens [1]. In 2015, the World Health Organization (WHO) estimated about half of the world population is at risk of this disease, malaria deaths are so frequent that statistically speaking every two minutes a child dies from it [2]. Malaria parasites (*Plasmodium*) are introduced into human hosts by feeding female anopheles mosquitoes. There are five species that infect humans worldwide of these, *Plasmodium falciparum* is the deadliest [3]. During the asexual part of the parasite life cycle (intra-erythrocytic stage), the *Plasmodium* feeds on the hemoglobin of the hosts RBC's [4]. This stage is responsible for the clinical symptoms of the disease and the end product of this consumption is the liberation of highly activated Ferriprotoporphyrin (IX) that can kill the parasites through cellular oxidative stress [5-6]. Unfortunately, the parasite has a solution to this problem by dimerizing this ferric heme into an insoluble more stable form called hemozoin or malaria pigment [7-9]. Purified hemozoin was found to be identical to the synthetic analogue, β -hematin, an insoluble form of ferric protoporphyrin IX (FP) [10], produced in the presence of acetic acid [11,12] which can be used as tool to test new potential antimalarials [13,14]. Due to indiscriminate use of drugs, an alarming increase in drug resistance emerges [15], which has become a global concern. Consequently, researchers are increasingly turning their attention

to folk medicine and exploring natural resources for new bioactive compounds, looking for new leads to develop better drugs [16]. According to the WHO, about 80% of the world's population relies on traditional medicine [17].

In recent years, resistance to artemisinin, the core compound in WHO recommended combination treatments for malaria [18] has been detected in south East Asia [19], necessitating the urge for finding a new drug against this dreadful disease. *Inula* is a large genus containing about 120 species that belongs to the family Asteraceae. *I. viscosa*, also known as *Dittrichia viscosa* [20] is a predominant Mediterranean herbaceous perennial plant that is traditionally used in Palestinian folk medicine with a large number of medical applications [21,22] such as a diuretic, antifungal [23] topical anti-inflammatory and hemostatic activity [24]. Several other studies have shown antimicrobial activity [25], antioxidant [26], and anti-inflammatory [27], and hepatoprotective activities [28]. *I. viscosa* contains a large number of pharmacologically active compounds. Previous studies isolated and identified various chemical compounds in *Inula* species including flavonoids [29,30], terpenes [31], sesquiterpenoids [32-33] and sesquiterpene Lactones; [34]. We have investigated the traditional Palestinian *I. viscosa* plant crude extract in the past and found it to exhibit potential anti-malarial activity *in vitro* [35]. The current work however, focuses on the preparative HPLC fractionation of the alcoholic extract of *I. viscosa* and their *in vitro* effect on β -hematin inhibition as a parameter to antimalarial activity. Moreover, the full scan UHPLC-ESI-MS was conducted

to reveal the structure of some isolated compounds probably responsible for the antimalarial activities.

Materials and Methods

Dimethyl sulfoxide (DMSO) of 99.5% purity, chloroquine diphosphate salt, sodium acetate of 99% purity and hemin chloride were procured from Sigma-Aldrich, Israel. Glacial acetic acid was purchased from Fluka. Ethanol (EtOH) solvents were purchased from Merck (Germany). Fresh wild leaves of Palestinian *I. viscosa* were collected towards the end of October 2015 from different areas around Jerusalem far from agricultural lands. Samples were air dried in the shade for 10 days. Grinded and kept in a fridge.

Preparation of plant extract

The crude extract was obtained by macerating (1:10) wt/vol of dried plant leaves in 35% ethanol, left for 24 hours at room temperature, then filtered using MN 615.Ø110 mm filter paper. The resultant extract was evaporated using (IKA WEREK RV06-ML) rotary evaporator at 70°C under reduced pressure. Then, it was lyophilized using Labconco freeze drier (USA), until constant weight was achieved. The final dried extract was stored in amber bottle and kept in the fridge until fractionated by preparative HPLC.

In vitro semi-quantitative test for screening of anti-malarial activity

As per the protocol described by Deharo *et al.* [36], a mixture containing 50 μ l of 0.5mg/ml haemin chloride (freshly dissolved in DMSO), 100 μ l of 0.5 M sodium acetate buffer (pH 4.4) and 50 μ l of the tested anti-malarial drug solution or control, was incubated in a normal non-sterile 96-well flat bottom plate at 37°C for 18-24 hours. It is imperative that the solutions are being added to the plate in this order. The plate was then centrifuged for 10 minutes at 4000 rpm. The supernatant was removed and the pH of reaction was measured. The final pH of the mixture should be between 5.0 and 5.2. The wells were washed with 200 μ l DMSO per well to remove free haemin chloride. The plate was centrifuged again, discharging the supernatant afterwards. The β -hematin remaining was then dissolved in 200 μ l of 0.1 M NaOH to form alkaline hematin that could be measured spectrophotometrically. Finally, the absorbance was determined at 405nm using an ELISA reader (Stat Fax-2100). Ultrapure water was used as negative control, whereas chloroquine dissolved in ultrapure water was used as positive control.

Chromatographic Analysis

Analytical HPLC, preparative HPLC and UHPLC-MS instrumentations

Analytical samples were analyzed using HPLC of Waters Alliance (e2695 separations module), which is equipped with 2998 Photo diode Array (PDA). Data acquisition and control were carried out using Empower 3 chromatography data software (Waters, Germany). The Preparative High Pressure Liquid Chromatography (Prep-HPLC) system consisted of 3535 quaternary gradient module equipped with 996 PDA detectors.

The UHPLC was performed under reverse phase conditions using a TSQ Quantum Access MAX (Thermo Scientific, San Jose, CA, USA) which includes a Dionex Pump with a degasser module, an Accela PDA detector and an Accela Autosampler. The chromatographic separations were performed on a Kinetex™ column (C8, 2.6 μ m particle size, 100 Å pore size, 100 x 2.1mm) from Phenomenex, USA. A UHPLC Security Guard cartridge (C8, for 2.1mm ID column) was from Phenomenex, USA. The injection volume was 10 μ L; the oven temperature was maintained at 35°C.

Chromatographic conditions

Crude *I. viscosa* from 35% ethanol was run on reversed phase ODS column of Waters (XBridge, 4.6 ID x 150mm, 5 μ m) with a guard column of XBridge ODS (20mm x 4.6mm ID, 5 μ m). The mobile phase consisted of a binary solvent mixture of 0.5% acetic acid solution (A) and acetonitrile (B) in linear gradient mode. A 100% of solvent A was initially used and then descended to 70% A in 40 minutes, then to 40% A in 20 minutes, finally to 10% A in 2 minutes and remained there for 6 minutes and then back to the initial conditions in 2 minutes. The HPLC system was equilibrated for 5 minutes with the initial acidic water mobile phase (100% A) before injecting the next sample. All the samples were filtered with a 0.45 μ m PTFE filter. The PDA wavelengths range was from 210-500 nm. The flow rate was 1 ml/min. Injection volume was 10 μ l and the column temperature was at room temperature. The preparative HPLC experiments were run on ODS column (Agilent PrepHT C18, 22.2 x 250 mm, 10 μ m). The gradient program started from 95% water, 5% acetonitrile to 100 % acetonitrile in 17 minutes and back to the starting mobile phase composition in 4 minutes. The flow rate was 12 ml/minutes, the injection volume was 1000 μ l, the column temperature was set at room temperature and the PDA wavelengths ranged from 200-450nm. The UHPLC-ESI-MS chromatographic separation was achieved using a linear gradient program at a constant flow rate of 0.4 mL/min over a total run time of 70 min. The mobile phase gradient program was the same as the HPLC except for the use of 0.1% formic acid instead of acetic acid. Samples were detected by a TSQ Quantum Access Max mass spectrometer in positive and negative ion mode using Electron Spray ionization (ESI) and full scan acquisition. Air was produced (SF 2 FF compressor, Atlas Copco, Belgium). Purified nitrogen was used as source and exhaust gases.

I. viscosa preparative sample preparation

700 mg of crude Palestinian *I. viscosa* was dissolved in 15 ml ethanol with 5 drops of acetic acid, and then the solution was directly filtered through PTFE 0.45 μ m membrane. One ml (46.6 mg) of the clear solution was directly injected to the preparative HPLC column and nine fractions were collected. Fractions collected time were as follows: I (4-6.5), II (9.3-11.2), III (11.2-13.7), IV (13.7-19), V (20.5-23), VI (23-23.8), VII (23.8-24.5), VIII (24.5-25.5), IX (25.5-26) minutes.

Results and Discussion

It is known from previous literature that the chemical constituents from the genus *Inula* contains more than 117 different compounds including volatile, semi-volatile and non-

volatiles polar compounds [28]. This includes broad range of monoterpenes, sesquiterpenes, diterpenes, flavonoids, and glycolipid. Depending on the nature of the extracting solvent used, one can roughly predict the nature of the isolated compounds. In the current effort, 35% ethanol was utilized as a macerating agent at room temperature to extract out the polar non-volatiles phenolic compounds. The gentle extraction method used was to deliberately avoid the degradation of any thermo labile active compounds and to imitate as much as we can the traditional use of natural products simple extractions to cure malaria. We have previously investigated the crude raw material powder from 35% ethanol extract of *I. viscosa* and measured its *in vitro* inhibitory effect on β -hematin formation. The crude alcoholic extract of *I. viscosa* has the capability to impede the formation of β -hematin *in-vitro*; with an efficiency of about 93% when compared to the standard chloroquine which gave 94% at comparable concentrations [35]. However, pure isolates or preparative HPLC fractions have never been explored yet. Surprisingly, *I. viscosa* crude ethanolic extract was found to be quite stable when left at room temperature for about four years by maintaining its *in vitro* efficiency (Figure 1). As in Figure 1, the absorption at 405nm of the same whole crude extract almost stayed the same over a period of more than three years. This is a very important finding because in comparison to the *Artemisia annua* extract, we noticed that the latter lost its antimalarial activity within few days [37]. Stability problems are known to exist for some isolated pure molecules such as artesunate and amodiaquine as well [38] especially under tropical climate hot and humid weather. This addresses a major concern in alternative therapy: in Southern countries the whole plant extract is usually used rather than isolated pure individual compounds and in the Western therapy isolated pure isolates are used exclusively. The stability of *I. viscosa* extracts may have another impact. The food industry is desperate for plant extracts which could have a stabilizing and anti-oxidant effect on their produces.

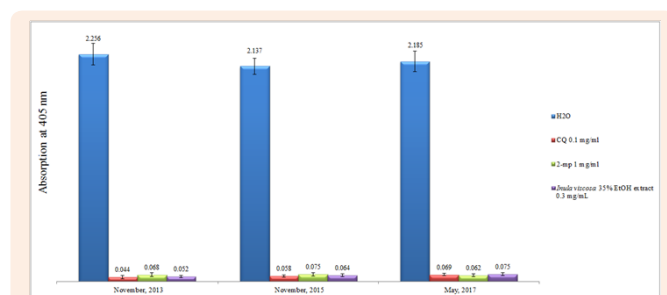


Figure 1: Column diagram representing the effect of time on the efficacy of potential anti-malarial drug 35% ethanol extract of *I. viscosa* leaves compared to the control water and positive controls chloroquine (CQ) 0.1 mg/mL, 2-mercaptopyrimidine (2-mp) 1mg/mL; showing the absorption values of dissolved β -hematin (alkaline hematin) at 405 nm using ELISA reader; according to E. Deharo semi-quantitative method; each result represents the average of 24 individual experiments.

The typical analytical HPLC-PDA chromatogram of the crude extract of *I. viscosa* (5mg/ml, 10 μ l injection) at 320 nm is shown in Figure 2. The preparative chromatograms of the 700mg crude extract (1000 μ l injection) at 320nm are shown in Figure 3. The

analytical and the preparative chromatograms showed peaks that share the same maximum of UV-Vis wavelengths at 324-328 nm and with a shoulder at 290-295nm. This observation indicates that the separated compounds share the same main functional groups, i.e., chromophores that are most likely belong to the flavonoids family. It is clearly seen that the major doublet peak in the preparative HPLC fraction (IV) that eluted at 16 minutes is the same peaks eluted at 27.1-27.6 minutes using the analytical mode. While preparative fraction III corresponds to the peak eluted at 14 minutes. Three fractions (IV, V and VI) out of nine (I-IX) from the 35% ethanolic extracts of *I. viscosa* leaves were shown to possess *in vitro* effect on β -hematin inhibition. It is worthwhile mentioning that the fractions VI-VIII are representing isolated pure compounds (Figure 3). Absorption results obtained at 405 nm were compared to positive controls of chloroquine (CQ) and amodiaquine (AD), at 0.1 mg/ml each, and negative controls of ultrapure H₂O and DMSO. According to the semi-quantitative method performed, the absorption is inversely proportional to fraction efficiency and to β -hematin content (Figure 4). Fraction IV that comprises two major compounds emerged together at retention interval of 13.7-19 minutes (Figure 3) showed an absorption value of 0.221 at a concentration of 0.3 mg/ml. The less active fraction V that eluted between 20.5-23 minutes showed an absorption value of 0.648. Fraction VI which is a pure individual compound eluted at 23-23.8 minutes (Figure 3) showed moderate activity (absorption of 0.338) in comparison to the positive controls (Figure 4). Although fractions VII and VIII are presumably pure isolates, nonetheless, they are inactive.

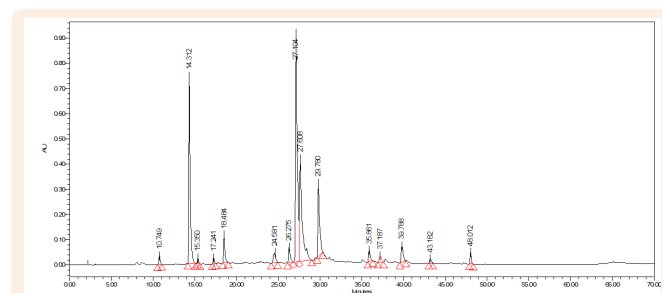


Figure 2: Typical HPLC-PDA chromatogram of all the peaks present in crude 35% ethanol extract of *I. viscosa* at 320 nm and their overlaid UV-Vis spectra between 210-500 nm.

In poor countries drinking herbal tea extract is affordable and cheap in comparison to pure drugs offered by western therapy. Although the whole crude extract contains individual antimalarial compounds or fractions at a much lower dose, the antimalarial activity is superior due to synergistic interaction between compounds present in the *I. viscosa* extract. Recent LC-PDA-ESI-MSMS investigation has characterized the presumed antioxidants and antifungal phenolic compounds in methanolic *I. viscosa* extract in the negative ESI mode [39]. The phenolic profile generated was dominated by hydroxycinnamic acid, flavonols and flavanones derivatives. The most prominent components were mono- and dicaffeoylquinic acids [39]. In order to explore the structural features of the phenolic compounds from the 35% ethanolic extract, a full scanned UHPLC-PDA-ESI-MS using the negative and positive electro spray ionization modes was

examined. Figure 5 showed the UHPLC chromatogram of the major components at 320 nm. As seen in Figure 5, the major peaks at 10.97, 27.38, 28.42 and 31.05 minutes are very similar to the major peak profiles of the analytical HPLC at 14.31, 27.16, 27.60 and 29.78 respectively (Figure 2). The major peak eluted at 10.97 minutes (in fraction III) gave a deprotonated pseudo-molecular ion $[M-H]^-$ at m/z of 407.17 Da with a base fragment ion at 375.27 Da. Other major peaks appeared at retention of 16.5 minutes showed molecular ion $[M-H]^-$ at m/z of 515.45 Da and a fragment at 353.28 Da presumably signifying to the caffeoylquinic acid (chlorogenic acid) skeleton which has a molecular weight of 354 Da (Figure 5). Other adjacent peaks at a retention time of 27.5 and 28.5 minutes showed a deprotonated peak molecular ion $[M-H]^-$ at 515.45 Da and 405.31 Da and the same chlorogenic acid backbone base peak at 353.30 Da. The deprotonated 515.45 Da ion may indicate the presence of more than one caffeoyl group attached to different quinic acid OH groups. The same observation was seen in Mahmoudi et al. [39]. Both 27.5 and 28.5 minutes peaks were eluted in fraction IV of the preparative HPLC fraction (Figure 3). The peak at 31 minutes exhibited the same deprotonated molecular ion at m/z 515.45 and 353.33 base peak indicative of another geometrical isomer of dicaffeoylquinic acid. It is evident that all the major peaks showed a fragment at m/z of 353.30 Da which indicates that the isomeric flavonoids are a shared caffeoylquinic acid compound, which is the ester of caffeic acid and (-)-quinic acid, is known for its antioxidant activity [40]. Fraction VI that eluted at 23-23.8 minutes using preparative HPLC presumably is indicative of the presence of sakuranetin compound since it showed a protonated molecular ion $[M+H]^+$ at m/z of 284.20 Da and a deprotonated molecular ion $[M-H]^-$ at m/z of 286.20 Da.

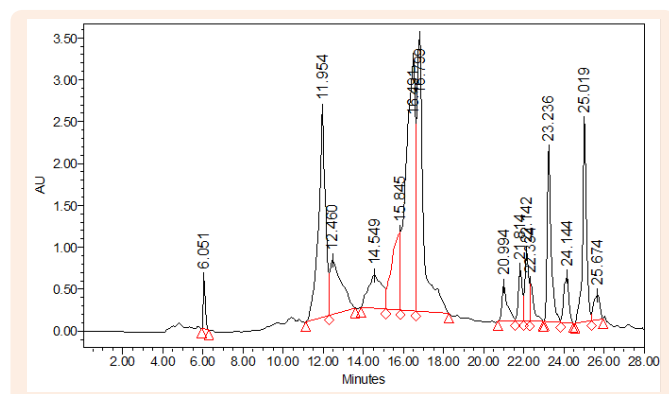


Figure 3: Typical preparative HPLC-PDA chromatogram of crude 35% ethanol extract of *I. viscosa*. 1 ml was injected at flow rate of 12ml/min at 320 nm using inch reversed phase column. Fractions collected time were as follows: I(4-6.5), II (9.3-11.2), III (11.2-13.7), IV (13.7-19), V (20.5-23), VI (23-23.8), VII (23.8-24.5), VIII (24.5-25.5), IX (25.5-26) minutes.

Conclusion

The emergence of resistant strains of *Plasmodium falciparum* to common antimalarial drugs, ranging from chloroquine to artemisinin combined therapy has been recognized. Development and search of novel and effective antimalarial agents to overcome chloroquine resistance have become an indispensable issue.

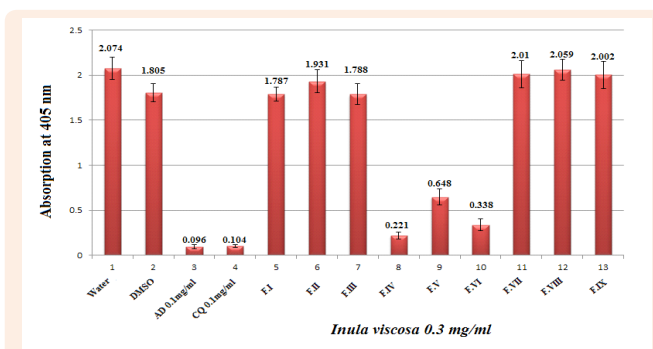


Figure 4: Column diagram representing the efficiency of nine fractions obtained from preparative HPLC separation of 35% ethanol extract *I. viscosa* leaf, dissolved in water-DMSO at concentration of 0.3mg/ml. The results are compared to negative and positive controls, showing the absorption values of dissolved β -hematin (alkaline hematin) at 405 nm using ELISA reader. Each result represents an average of 16 individual experiments.

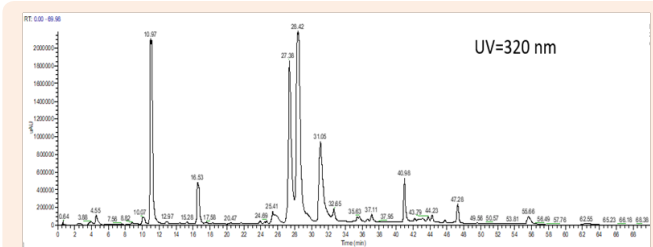


Figure 5: UHPLC-PDA-ESI-MS chromatogram of the *I. viscosa* 35% ethanol extract.

As unfold from the results of this study, three fractions (IV, V and VI) out of nine from the 35% ethanolic extracts of *I. viscosa* leaves were shown to possess *in vitro* effect on β -hematin inhibition in comparison to the positive controls. The crude whole extract versus the preparative HPLC fractions that includes pure compounds (VI, VII, VIII) were tested and the crude extract was found to be superior on β -hematin inhibition. This observation demonstrates that raw extracts sometimes exceed the efficacy of either the preparative fractions or isolated pure compounds due to synergistic effect.

Acknowledgement

None.

Conflict of Interest

None.

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