

Epilithic diatoms (Bacillariophyta) from cloud forest and alpine streams in Bolivia, South America II: A preliminary report on the diatoms from Sorata, Department of La Paz

Eduardo A. Morales^{1,2,3} Morgan L. Vis⁴, Erika Fernández⁵, J. Patrick Kociolek⁶

¹ Herbario Criptogámico, Departamento de Ciencias Exactas e Ingenierías Universidad Católica Boliviana, ² Patrick Center for Environmental Research, The Academy of Natural Sciences of Philadelphia, ³ Unidad de Limnología y Recursos Acuáticos Renovables, Universidad Mayor de San Simón, ⁴ Department of Environmental and Plant Biology, ⁵ Centro de Biodiversidad y Genética, Universidad Mayor de San Simón, ⁶ Diatom Collection, California Academy of Sciences.

e-mail: morales@acnatsci.org, vis-chia@ohio.edu,
erika_fer2003@yahoo.es, pkociolek@calacademy.org.

Abstract

A total of seven epilithic samples were collected from streams along an altitudinal gradient within the Bolivian Cloud Forest region near the capital La Paz. The diatom flora is diverse and no clear altitudinal trends were found. The flora is composed of cosmopolitan species and taxa restricted in distribution to the Andes with the genera *Gomphonema*, *Nitzschia*, *Encyonopsis*, and *Encyonema* containing the greatest number of species and individuals with higher frequencies in samples. Diatoms from this region are also represented by several alkaliphilous and alkalibiont taxa reflecting the high pH of the streams. General characteristics of the sampled sites as well as a list of taxa and figures of representative taxa are presented herein.

Key words: Diatoms, Bacillariophyta, Taxonomy, Sorata, Bolivia.

Resumen

Un total de siete muestras de *epilithon* fueron colectadas en ríos dentro de un gradiente altitudinal en el bosque nublado de Bolivia, cerca de la capital La Paz. La flora diatomológica es diversa y no se encontraron patrones altitudinales claros. Tal flora está compuesta por especies cosmopolitas y taxa restringidos a los Andes con los géneros *Gomphonema*, *Nitzschia*, *Encyonopsis* y *Encyonema* agrupando al mayor número de especies e individuos con mayores frecuencias. La flora está también representada por varios taxa alcalifilos y alcalibiontes, reflejando los altos valores de pH de los ríos. Las

características de los sitios muestreados, así como una lista de taxa y figuras representativas se presentan en este trabajo.

Palabras clave: Diatomeas, Bacillariophyta, Taxonomía, Sorata, Bolivia.

1 Introduction

Diatoms are photosynthetic, diploid, eukaryotic organisms placed in the Kingdom Chromista, along with brown algae, chrysophytes and other *c*-chlorophyll bearing microbionts, and slime molds [2]. Diatom communities have been extensively used in the assessment of past and present ecological conditions in the aquatic habitats in which they live [29]. Their indicative utility resides in that they are ubiquitous, their taxonomically diagnostic, hard siliceous cell walls are well preserved in sediments and many species form characteristic assemblages under different trophic or diversely contaminated conditions [12] [25] [26].

Bolivian diatoms have been poorly studied and a review of the literature (in preparation) shows that most research has been restricted to the Altiplano. Only a fraction of this literature is taxonomic in nature, which hinders the potential use of diatoms for bioindication. Only a few articles have focused on the diatoms from the Bolivian Cloud Forest [21] [28], which are insufficient to cover this extensive ecosystem.

The South American Cloud Forest (or Yungas) is one of the most biologically diverse ecosystems in South America [5] [23] located between 13°-17°S and 69°-63°W and running along the Eastern Cordillera of the Andes in Bolivia and Peru, between the high Puna or Altiplano and the Amazonian lowlands. In Bolivia, the Yungas has a wide altitudinal range (200-3500 m), and a wide range in temperature (annual means: 7°-30°C) and humidity (800-7000 mm per year). The geology of the Yungas is also varied, composed of four different geological formations [23], resulting in a mixture of landscapes with great influence on the vegetation and fauna of the region. Navarro & Maldonado [23] state that the Yungas is dominated by riverine systems with few lakes. First order streams with fast flowing waters of varied chemical composition predominate at higher altitudes while there is a predominance of second and third order streams with increased load of particulates at lower elevations.

Biologically, the diversity of plants and certain groups of animals such as birds, mammals and amphibians in the Yungas is the best documented [3] [22] [24] and shows high endemism in these groups e.g., Birds [1], Ferns [6] [7]. Within the algae, Thérézien [31] reported 243 soft algae taxa from a few samples collected in a small region of the Yungas, 8 of them being taxa new to science. McClintic *et al.* [15] studied 13 samples collected from different sites and reported more than 40 taxa of soft-bodied algae, of which 17 were new records for Bolivia. A study of the diatoms present in 6 of these 13 samples showed that about a quarter of the species are not found in any bibliographical reference for South America or similar regions of the world [21].

The current paper presents a taxonomic assessment of the periphytic diatom flora in seven streams located in Sorata, Province of Larecaja, Department of La Paz and it constitutes a preliminary contribution before the initiation of a larger sampling project throughout the Yungas region.

2 Materials and Methods

2.1 Study area

Collection of samples took place in the vicinity of Sorata, about 100 km from the capital La Paz. Seven streams were sampled from an altitudinal range of ca. 2 600-4 060 m. Coordinates and physico-chemical characteristics of each sampling point are presented in Table 1 (samples 27-33) in McClintic *et al.* [15]. Four of the streams fell within the altitudinal range given for the Yungas by Navarro & Maldonado [23], while the remaining three correspond to the elevation usually given for the Puna. Despite the altitude difference there are no clear patterns in the physical and chemical characteristics of the streams, except for specific conductance which is lower in the higher streams [15].

Table 1. List of taxa found in the 7 samples collected from Sorata (Bolivia) streams. Each column contains the integration of data collected from two preparations from each sample. Stream numbers correspond to those presented in Table 1 in McClintic [15].

Taxon/Stream number	27	28	29	30	31	32	33
<i>Achnanthes minutissima</i> var. <i>jackii</i> (Rabenhorst) Lange-Bertalot et Ruppel			+	+	+	+	
<i>Achnantheidium exiguum</i> var. <i>heterovalvum</i> (Krasske) Czarnecki		+					
<i>Achnantheidium minutissimum</i> (Kützing) Czarnecki	+	+	+	+	+	+	
<i>Achnantheidium modestiforme</i> (Lange-Bertalot) Van de Vijver			+	+			
<i>Achnantheidium rivulare</i> Potapova et Ponader				+	+		
<i>Adlafia bryophila</i> (Petersen) Lange-Bertalot			+				
<i>Adlafia minuscula</i> (Grunow) Lange-Bertalot			+	+	+		
<i>Adlafia</i> sp. 2 SORATA		+					
<i>Adlafia suchlandtii</i> (Hustedt) Lange-Bertalot			+	+			
<i>Amphora carvajaliana</i> Patrick			+				
<i>Amphora montana</i> Krasske	+				+	+	
<i>Amphora pediculus</i> (Kützing) Grunow						+	
<i>Aulacoseira ambigua</i> (Grunow) Simonsen						+	

Taxon/Stream number	27	28	29	30	31	32	33
<i>Aulacoseira granulata</i> (Ehrenberg) Simonsen						+	
<i>Brachysira brebissonii</i> Ross				+			
<i>Brachysira lebmannaiae</i> Lange-Bertalot et Moser				+			
<i>Brachysira neoexilis</i> Lange-Bertalot			+	+			
<i>Brachysira serians</i> (Brébisson) Round et Mann				+			
<i>Caloneis bendeyi</i> Lange-Bertalot							+
<i>Caloneis molaris</i> (Grunow) Krammer						+	
<i>Caloneis</i> sp. 1 SORATA			+	+			
<i>Caloneis</i> sp. 2 SORATA						+	
<i>Cavinula protractolapidoso</i> Metzeltin, Lange-Bertalot et Garcia-Rodriguez			+				
<i>Cavinula pseudoscutiformis</i> (Hustedt) Mann et Stickle			+	+			
<i>Chamaepinnularia evanida</i> (Hustedt) Lange-Bertalot							+
<i>Chamaepinnularia soebrensis</i> var. <i>bassiaca</i> (Krasske) Lange-Bertalot			+	+			
<i>Chamaepinnularia soebrensis</i> var. <i>musciola</i> (Petersen) Lange-Bertalot et Krammer	+		+				
<i>Cocconeis placentula</i> var. <i>englypta</i> (Ehrenberg) Grunow	+	+	+				
<i>Cocconeis placentula</i> var. <i>lineata</i> (Ehrenberg) Van Heurck	+	+	+	+	+	+	
<i>Craticula</i> cf. <i>molestiformis</i> (Hustedt) Mayama							+
<i>Craticula halophiloides</i> (Hustedt) Lange-Bertalot							+
<i>Cymbella excisa</i> Kützing					+		
<i>Cymbella peraspera</i> Krammer			+				
<i>Cymbella tumida</i> (Brébisson ex Kützing) Van Heurck	+	+			+		
<i>Cymbopleura inca</i> (Lange-Bertalot, Krammer et Rumrich) Krammer				+			
<i>Diatoma hyemalis</i> (Roth) Heiberg			+	+			
<i>Diatoma mesodon</i> Kützing			+	+			
<i>Diatoma moniliformis</i> Kützing						+	
<i>Diploneis kablii</i> Lange-Bertalot et Rumrich			+	+			+
<i>Encyonema brevicapitatum</i> Krammer	+						
<i>Encyonema</i> cf. <i>brevicapitatum</i> Krammer	+		+	+	+		

Taxon/Stream number	27	28	29	30	31	32	33
<i>Encyonema jemtlandicum</i> var. <i>venezolanum</i> Krammer			+				
<i>Encyonema lange-bertalotii</i> Krammer				+	+		
<i>Encyonema minutiforme</i> Krammer			+				
<i>Encyonema minutum</i> (Hilse) Mann			+	+	+	+	+
<i>Encyonema neogracile</i> Krammer			+				
<i>Encyonema schneideri</i> Krammer			+				
<i>Encyonema silesiacum</i> (Bleisch) Mann			+	+	+	+	
<i>Encyonema</i> sp. 1 COROICO				+			
<i>Encyonema</i> sp. 1 SORATA			+				
<i>Encyonema subminutum</i> Krammer et Lange-Bertalot				+			
<i>Encyonopsis</i> cf. <i>cesatii</i> (Rabenhorst) Krammer			+				
<i>Encyonopsis</i> cf. <i>krammeri</i> Reichardt			+				
<i>Encyonopsis</i> cf. <i>krammerioides</i> Lange-Bertalot et Rumrich				+			
<i>Encyonopsis</i> cf. <i>minuta</i> Krammer et Reichardt			+				
<i>Encyonopsis</i> cf. <i>subminuta</i> Krammer et Reichardt			+				
<i>Encyonopsis microcephala</i> (Grunow) Krammer	+						
<i>Encyonopsis</i> sp. 1 SORATA			+				
<i>Eolimna</i> cf. <i>minima</i> (Grunow) Lange-Bertalot						+	
<i>Eolimna minima</i> (Grunow) Lange-Bertalot				+			
<i>Eolimna</i> sp. 1 SORATA			+				
<i>Eolimna subminuscule</i> (Manguin) Moser, Lange-Bertalot et Metzeltin	+					+	
<i>Epithemia adnata</i> (Kützing) Brébisson	+			+			
<i>Eucocconeis flexella</i> (Kützing) Brun			+				
<i>Eucocconeis quadratarea</i> (Østrup) Lange-Bertalot				+		+	
<i>Eunotia boreoalpina</i> Lange-Bertalot et Nörpel-Schempp				+			
<i>Eunotia muscicola</i> var. <i>tridentula</i> Nörpel et Lange-Bertalot			+	+			
<i>Eunotia paludosa</i> Grunow				+			
<i>Eunotia tecta</i> Krasske							+
<i>Eunotia tenella</i> (Grunow) Hustedt				+			

Taxon/Stream number	27	28	29	30	31	32	33
<i>Fragilaria capucina</i> var. <i>gracilis</i> (Østrup) Hustedt			+	+	+		
<i>Fragilaria</i> sp. 1 SORATA			+				
<i>Fragilaria</i> sp. 2 SORATA							+
<i>Fragilaria vaucheriae</i> (Kützing) Petersen	+	+		+	+	+	
<i>Frankophila similioides</i> Lange-Bertalot et Rumrich			+	+			
<i>Frustulia crassinervia</i> (Brébisson) Lange-Bertalot et Krammer				+			+
<i>Frustulia</i> sp. 1 SORATA				+			
<i>Frustulia vulgaris</i> (Thwaites) DeToni			+	+			+
<i>Geissleria ignota</i> (Krasske) Lange-Bertalot et Metzeltin		+					
<i>Gomphonema</i> cf. <i>angustum</i> (Kützing) Rabenhorst		+					
<i>Gomphonema</i> cf. <i>lagenula</i> Kützing			+	+			+
<i>Gomphonema</i> cf. <i>micropus</i> Kützing				+			
<i>Gomphonema</i> cf. <i>parvulum</i> (Kützing) Kützing	+	+					
<i>Gomphonema</i> cf. <i>punae</i> Lange-Bertalot et Rumrich			+	+	+		
<i>Gomphonema coronatum</i> Ehrenberg			+	+			
<i>Gomphonema exilissimum</i> (Grunow) Lange-Bertalot et Reichardt							+
<i>Gomphonema lagenula</i> Kützing	+						
<i>Gomphonema micropus</i> Kützing			+				
<i>Gomphonema olivaceoides</i> Hustedt							+
<i>Gomphonema parvulus</i> (Lange-Bertalot et Reichardt) Lange-Bertalot et Reichardt	+						
<i>Gomphonema parvulum</i> (Kützing) Kützing							+
<i>Gomphonema pumilum</i> var. <i>elegans</i> fo. <i>biseriatum</i> Morales et Vis	+	+	+		+	+	
<i>Gomphonema pumilum</i> var. <i>elegans</i> Reichardt et Lange-Bertalot	+	+					
<i>Gomphonema pumilum</i> var. <i>rigidum</i> fo. <i>biseriatum</i> Morales et Vis	+		+		+	+	
<i>Gomphonema punae</i> Lange-Bertalot et Rumrich				+	+		
<i>Gomphonema</i> sp. 1 SORATA			+				
<i>Gomphonema</i> sp. 2 SORATA				+			
<i>Gomphonema</i> sp. 3 SORATA				+			
<i>Gomphonema</i> sp. 4 SORATA							+

Taxon/Stream number	27	28	29	30	31	32	33
<i>Gomphonema</i> sp. 5 SORATA						+	
<i>Gomphonema subclavatum</i> (Grunow) Grunow			+	+	+		+
<i>Hannea arcus</i> (Ehrenberg) Patrick	+	+	+	+	+	+	
<i>Hantzschia abundans</i> Lange-Bertalot	+		+				
<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	+					+	
<i>Kobayasiella</i> cf. <i>parasubtilissima</i> (Kobayasi et Nagumo) Lange-Bertalot							+
<i>Mayamaea atomus</i> (Kützing) Lange-Bertalot		+					
<i>Mayamaea atomus</i> var. <i>permitis</i> (Hustedt) Lange-Bertalot					+	+	
<i>Mayamaea</i> cf. <i>atomus</i> var. <i>alcimonica</i> (Reichardt) Reichardt					+	+	
<i>Melosira varians</i> Agardh	+	+				+	
<i>Microcostatus</i> sp. 1 SORATA			+				
<i>Navicula angusta</i> Grunow			+				
<i>Navicula capitatoradiata</i> Germain	+				+		
<i>Navicula caterva</i> Hohn et Hellermann	+					+	
<i>Navicula</i> cf. <i>lundii</i> Reichardt			+	+		+	
<i>Navicula</i> cf. <i>veneta</i> Kützing						+	
<i>Navicula cryptocephala</i> Kützing							
<i>Navicula cryptotenella</i> Lange-Bertalot	+	+				+	
<i>Navicula gregaria</i> Donkin	+	+	+	+	+	+	
<i>Navicula lanceolata</i> (Agardh) Kützing						+	
<i>Navicula libonensis</i> Schoeman						+	
<i>Navicula longicephala</i> var. <i>vilaplani</i> Lange-Bertalot et Sabater	+						
<i>Navicula rehardtiana</i> Lange-Bertalot						+	
<i>Navicula rhynchocephala</i> Kützing				+			+
<i>Navicula rostellata</i> Kützing		+					
<i>Navicula schroeteri</i> var. <i>escambia</i> Patrick	+	+				+	
<i>Navicula</i> sp. 1 SORATA			+				
<i>Navicula</i> sp. 2 SORATA			+				
<i>Navicula</i> sp. 3 SORATA			+				

Taxon/Stream number	27	28	29	30	31	32	33
<i>Navicula</i> sp. 4 SORATA							+
<i>Navicula submuralis</i> Hustedt		+					
<i>Navicula symmetrica</i> Patrick	+					+	
<i>Navicula tripunctata</i> (Müller) Bory	+	+	+	+	+	+	
<i>Navicula trivialis</i> Lange-Bertalot						+	
<i>Neidium longiceps</i> (Gregory) Ross				+			
<i>Nitzschia acidoclinata</i> Lange-Bertalot			+				
<i>Nitzschia archibaldii</i> Lange-Bertalot	+	+		+	+	+	+
<i>Nitzschia biacrula</i> Hohn et Hellerman						+	
<i>Nitzschia boliviana</i> Morales et Vis	+	+	+	+		+	
<i>Nitzschia</i> cf. <i>agnita</i> Hustedt				+			
<i>Nitzschia</i> cf. <i>inconspicua</i> Grunow				+			
<i>Nitzschia</i> cf. <i>perminuta</i> (Grunow) Peragallo				+	+		
<i>Nitzschia dissipata</i> (Kützing) Rabenhorst	+	+	+	+	+	+	
<i>Nitzschia fonticola</i> var. <i>pelagica</i> Hustedt	+						
<i>Nitzschia hantzschiana</i> Rabenhorst	+	+					+
<i>Nitzschia heufleriana</i> Grunow		+					
<i>Nitzschia intermedia</i> Hantzsch	+						
<i>Nitzschia linearis</i> (Agardh) Smith			+				
<i>Nitzschia linearis</i> var. <i>subtilis</i> (Grunow) Hustedt						+	
<i>Nitzschia oberheimiana</i> Rumrich et Lange-Bertalot			+				
<i>Nitzschia palea</i> (Kützing) Smith				+	+	+	
<i>Nitzschia palea</i> var. <i>debilis</i> (Kützing) Grunow			+	+		+	
<i>Nitzschia recta</i> Hantzsch		+		+			
<i>Nitzschia sinuata</i> var. <i>delognei</i> (Grunow) Lange-Bertalot	+						
<i>Nitzschia soratensis</i> Morales et Vis	+	+			+	+	
<i>Nitzschia</i> sp. 1 SORATA			+	+	+		
<i>Nitzschia</i> sp. 2 SORATA			+	+		+	
<i>Nitzschia</i> sp. 3 SORATA			+			+	

Taxon/Stream number	27	28	29	30	31	32	33
<i>Nitzschia</i> sp. 4 SORATA				+		+	
<i>Nitzschia</i> sp. 5 SORATA							+
<i>Nitzschia tropica</i> Hustedt		+					+
<i>Pinnularia borealis</i> Ehrenberg			+	+			
<i>Pinnularia divergens</i> var. <i>sublinearis</i> Cleve			+				
<i>Pinnularia martinii</i> Krasske			+	+			
<i>Pinnularia obscura</i> Krasske				+			
<i>Pinnularia viridis</i> (Nitzsch) Ehrenberg				+			
<i>Planobididium</i> cf. <i>biporum</i> (Hohn et Hellermann) Lange-Bertalot						+	
<i>Planobididium frequentissimum</i> (Lange-Bertalot) Lange-Bertalot	+	+		+	+	+	
<i>Planobididium lanceolatum</i> (Brébisson ex Kützing) Lange-Bertalot	+	+	+	+	+	+	
<i>Pleurosira laevis</i> (Ehrenberg) Compère			+				
<i>Psammothidium grischunum</i> (Wuthrich) Bukhtiyarova et Round			+				
<i>Psammothidium subatomoides</i> (Hustedt) Bukhtiyarova et Round				+			
<i>Pseudostaurosira laucensis</i> var. <i>vulpina</i> (Lange-Bertalot et Rumrich) Morales comb. nov.			+				
<i>Reimeria sinuata</i> (Gregory) Kociolek et Stoermer	+	+	+	+	+	+	
<i>Rhoicosphenia abbreviata</i> (Agardh) Lange-Bertalot	+	+	+	+	+	+	
<i>Sellaphora</i> cf. <i>seminulum</i> (Grunow) Mann	+	+					
<i>Sellaphora seminulum</i> (Grunow) Mann							+
<i>Simonsenia delognei</i> (Grunow) Lange-Bertalot							+
<i>Stauroneis</i> sp. 1 SORATA							+
<i>Staurosirella</i> cf. <i>pinnata</i> (Ehrenberg) Williams et Round			+				
<i>Staurosirella leptostauron</i> (Ehrenberg) Williams et Round			+	+			
<i>Stephanodiscus</i> cf. <i>alpinus</i> Hustedt							+
<i>Stephanodiscus</i> cf. <i>minutulus</i> (Kützing) Cleve et Möller							+
<i>Surirella angusta</i> Kützing		+					+
<i>Surirella brebissonii</i> var. <i>kuetzingii</i> Krammer et Lange-Bertalo	+						
<i>Surirella minuta</i> Brébisson							+

Taxon/Stream number	27	28	29	30	31	32	33
<i>Tabellaria flocculosa</i> (Roth) Kützing			+	+			
<i>Tabellaria ventricosa</i> Kützing				+			
<i>Ulnaria</i> cf. <i>ulna</i> (Nitzsch) Compère		+		+			
<i>Ulnaria contracta</i> (Østrup) Morales et Vis	+	+			+	+	
<i>Ulnaria</i> sp. 1 SORATA			+	+			
Total number of taxa	45	37	81	78	37	61	21

2.2 Methods

At each site, five rocks were randomly selected and scraped with a stiff toothbrush. Scraped material from the rocks was rinsed with stream water, collected as a composite sample, and preserved with 2.5% CaCO₃-buffered glutaraldehyde. Data for the first two samples (27 and 28) was presented in [21] and are pooled here in order to provide a more complete list of diatoms.

For light microscopy (LM) analysis, a 10 ml periphyton subsample was extracted and cleaned using 30% H₂O₂ and concentrated HNO₃ [30]. Cleaned samples were suspended in glass-distilled water and air-dried. Dried samples were mounted on glass slides using NAPHRAX®. A Nikon Microphot-FXA microscope equipped with DIC was used to identify and measure diatoms at a magnification of 1 250X. A Spot Insight color digital camera was used to capture images of selected specimens. Identification of specimens present in two preparations from each sample was performed to the lowest taxonomic level possible using pertinent literature [14] [16] [17] [18] [27].

For scanning electron microscopy (SEM) studies, aliquots of clean material were air dried onto 15 X 15 cm pieces of aluminum foil. Smaller pieces were trimmed and mounted on aluminum stubs with double-sided tape. The stubs were coated with gold-palladium using a Polaron Sputter Coater for ca. 1 min at 1.8 kV. A Leo-Zeiss 982-DSM electron microscope was used for SEM analyses. LM and SEM digital images were directly captured into a computer file and plates were assembled using Adobe Photoshop v. 7.0.

3 Results and Discussion

A list of all taxa encountered is presented in Table 1. In total, 190 infrageneric taxa were identified. Of these, 53 entities could not be determined based on the literature for the region or from other parts of the world [8] [9] [10] [11]. Many of these entities probably represent taxa new to science, but none were sufficiently abundant to permit detailed studies. The majority of the undetermined species belonged to *Gomphonema* (20%), *Nitzschia* (16%), *Encyonopsis* (9%), and *Encyonema* (6%). These four genera were also the most common in samples. *Navicula* taxa accounted for 9% of the undetermined

species, due to the inclusion of small unknowns whose features were not clear in LM and may be placed in *Adlafia*, *Eolimna* and *Mayamaea* after thorough SEM analysis.

The streams were clear, small (width 0.85-8.3 m), and shallow (max. depth 12-40 cm). The smaller streams located at higher altitudes had a periphytic flora, which is characteristic of montane regions worldwide with cosmopolitan taxa, such as *Encyonema minutum* (Hilse) Mann, *Hannaea arcus* (Ehrenberg) Patrick and *Tabellaria flocculosa* (Roth) Kützing. Stream water specific conductance was generally low (20-210 $\mu\text{S}/\text{cm}$), with higher values potentially representing more human activity. In streams located at lower elevations, cosmopolitan taxa characteristic of more elevated trophic conditions were present (e.g., *Encyonema silesiacum* (Bleisch) Mann, *Melosira varians* Agardh, *Reimeria sinuata* (Gregory) Kociolek et Stoermer, *Rhoicosphenia abbreviata* (Agardh) Lange-Bertalot). The stream water pH was alkaline (7.4-8.6) with no clear altitudinal trend. Alkaliphilous (occurring at pH > 7 [32]) taxa were present: *Achnanthebidium exiguum* var. *heterovalvum* (Krasske) Czarnecki, *Frustulia vulgaris* (Thwaites) De Toni, and *Planothidium lanceolatum* (Brébisson ex Kützing) Lange-Bertalot. In addition, several alkalibionts (living exclusively at pH > 7) were collected such as *Epithemia adnata* (Kützing) Brébisson, *Pleurosira laevis* (Ehrenberg) Compère, *Stephanodiscus* cf. *minutulus* (Kützing) Cleve et Möller. A few taxa, reported only from the Andes (possible endemics), were present in the Sorata flora as follows: *Encyonema jemtlandicum* var. *venezolanum* Krammer, *Gomphonema punae* Lange-Bertalot et Rumrich and *Staurosira laucensis* var. *vulpina* Lange-Bertalot et Rumrich, a species herein transferred to *Pseudostaurosira* (see later).

In general, the streams at higher altitudes (streams 29 and 30) harbored more diverse floras (81 and 78 taxa, respectively) and lower streams (No. 27, 28 and 31) contained fewer taxa (range 37-45 taxa) (Table 1). Stream 33 is located at higher elevation, but contained fewer taxa (21). Water temperature (16°C) and pH (8.4) were higher in stream 33 than in streams of similar altitude, while the current velocity (6.1 cm/sec) and conductivity (20 $\mu\text{S}/\text{cm}$) were lower. These conditions might favor the development of other groups of algae since McClintic *et al.* [15] reported high numbers of soft-bodied macro- and microalgae in this stream. In contrast, stream 32 was lower in elevation, but contained high diatom diversity (61 taxa). This stream was the shallowest (12 cm) of all streams sampled and had the fastest current velocity (52.6 cm/sec). Apart from these two factors, the stream appears similar in conditions to the other lower altitude streams and, thus, it is difficult to pinpoint the reason for its high diversity. For this same stream, McClintic *et al.* [15] report only three species of macroalgae.

Taxonomic section

The taxa listed in Table 1 represent the finest level of taxonomic resolution we can provide to date. Several of these are represented in Figures 1-68. Of interest is the presence of taxa whose specific epithets cannot yet be determined, and may represent species new to science. The identity and distribution of these potentially new taxa, in addition to a broader perspective on the diatoms from lotic systems throughout the Bolivian Yungas region, will be the focus of future collaborative research, funded in part by the California Academy of Sciences.

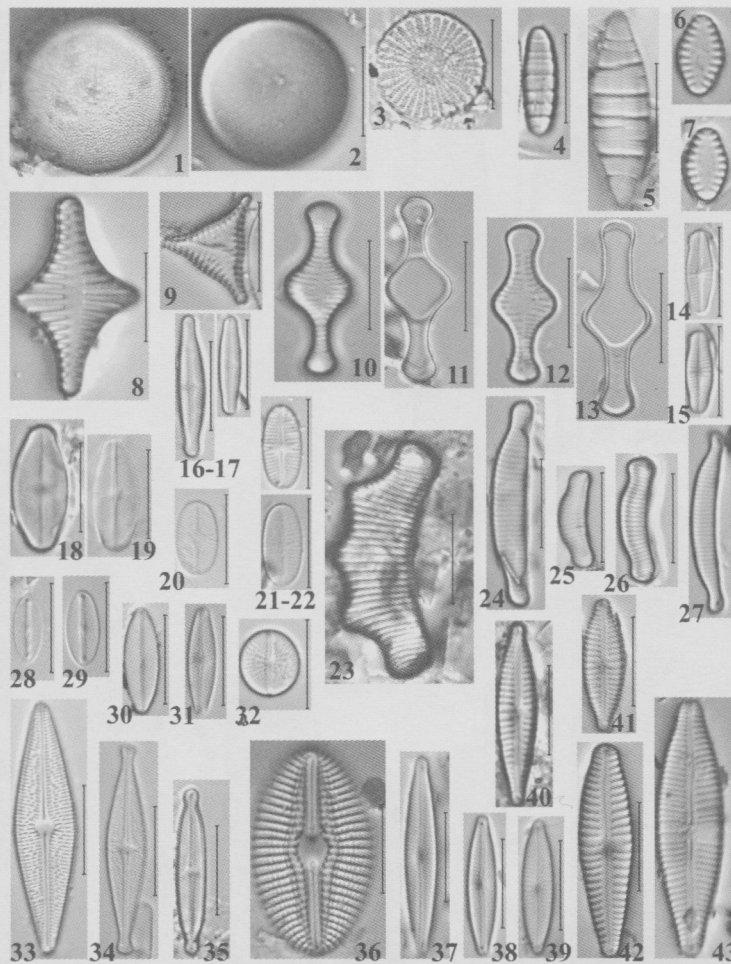
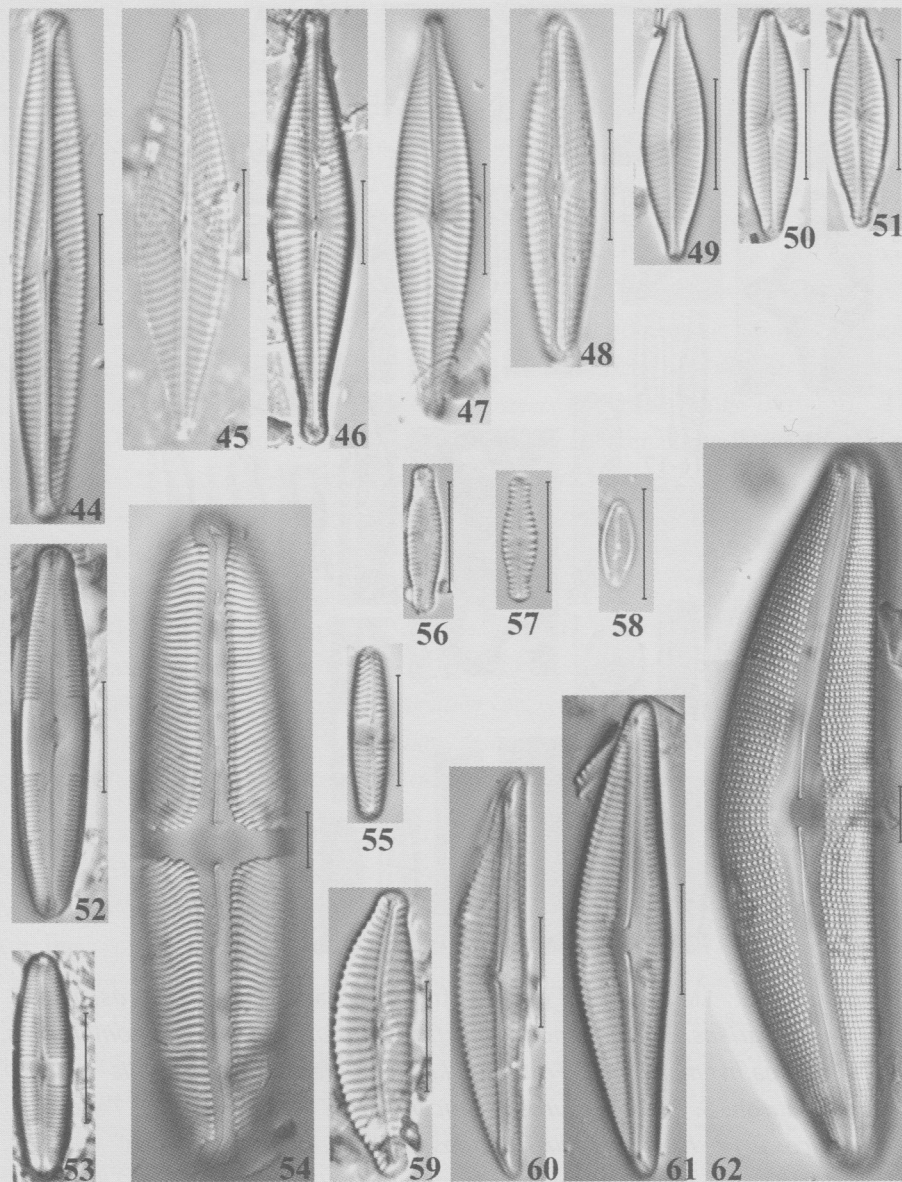
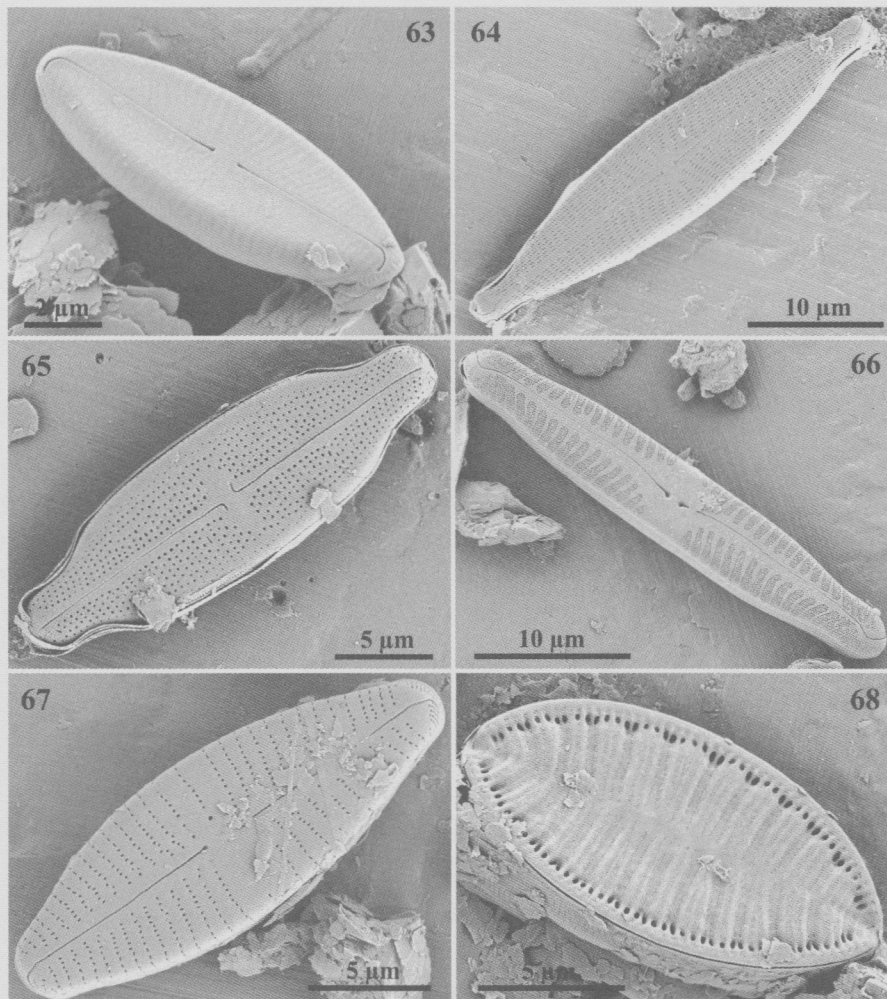


Figure 1–43: LM images of diatoms from Sorata. 1. *Orthoseira roseana*. 2. *Melosira varians*. 3. *Stephanodiscus cf. minutulus*. 4. *Diatoma moniliformis*. 5. *Diatoma hyemalis*. 6–7. *Frankophila similioides*. 8. *Staurosirella leptostauron*. 9. *Pseudostaurosira laucensis* var. *vulpina*. 10–11. *Tabellaria ventricosa*. 12–13. *Tabellaria flocculosa*. 14–15. *Achnantheidium modestiforme*. 16–17. *Achnantheidium minutissimum* var. *jackii*. 18–19. *Eucocconeis quadratarea*. 20. *Psammothidium subatomoides*. 21–22. *Psammothidium grischunum*. 23. *Eunotia tecta*. 24. *Eunotia boreoalpina*. 25–26. *Eunotia tenella*. 27. *Eunotia paludosa*. 28. *Mayamaea atomus* var. *permitis*. 29. *Mayamaea cf. atomus* var. *alcimonica*. 30. *Adlafia minuscula*. 31. *Adlafia suchlandtii*. 32. *Cavinula pseudoscutiformis*. 33. *Brachysira lehmanniae*. 34. *Brachysira neoexilis*. 35. *Kobayasiella cf. parasubtilissima*. 36. *Diploneis kahlii*. 37–38. *Encyonopsis cf. krammerioides*. 39. *Encyonopsis cf. krammeri*. 40. *Gomphonema exilissimum*. 41. *Gomphonema parvulum*. 42. *Gomphonema punae*. 43. *Gomphonema micropus*.



Figures 44–62. LM images of diatoms from Sorata. 44. *Navicula angusta*. 45. *Navicula trivialis*. 46. *Navicula rhynchocephala*. 47. *Navicula arkona*. 48. *Navicula libonensis*. 49. *Navicula cryptocephala*. 50. *Navicula* cf. *lundii*. 51. *Navicula reichardtiana*. 52. *Caloneis molaris*. 53. *Caloneis hendeyi*. 54. *Pinnularia divergens* var. *linearis*. 55. *Pinnularia obscura*. 56–57. *Chamaepinnularia soehrensii* var. *hassiacae*. 58. *Chamaepinnularia evanida*. 59. *Cymbella excisa*. 60. *Encyonema neogratile*. 61. *Encyonema schneideri*. 62. *Cymbella peraspera*.



Figures 63–68. SEM images of diatoms from Sorata. 63. *Adlafia minuscula*. 64. *Navicula arkona*. This taxon has been misidentified in Morales & Vis [21] as *Navicula capitatoradiata* (Figs 103, 104, and 122). 65. *Neidium longiceps*. 66. *Pinnularia obscura*. 67. *Gomphonema micropus*. 68. *Surirella angusta*.

Presence of one taxon in our samples requires a nomenclatural transfer. We present:

Pseudostaurosira laucensis* var. *vulpina (Lange-Bertalot et Rumrich) Morales comb. nov. (Fig. 9).

Basionym: *Staurosira laucensis* var. *vulpina* Lange-Bertalot et Rumrich in Rumrich *et al.*, 2000, Diatoms of the Andes from Venezuela to Patagonia/Tierra del Fuego, *Iconographia Diatomologica* 9, p. 223-224, Plate 10, Figs 1-11.

Morales & Vis [21] presented a new combination for *Staurosira laucensis* Lange-Bertalot et Rumrich within the genus *Pseudostaurosira* Williams et Round [33] based on

the resemblance of this taxon with other species included in the latter genus. The combination presented herein is a follow up to that work. Based on the discussion presented by Morales & Vis [21], several species recently described and transferred by Lange-Bertalot and collaborators within *Staurosira* Ehrenberg must be moved into other genera. This is because the concept of *Staurosira* handled by Lange-Bertalot is untenable and does not comply with extensive studies presented by several subsequent authors regarding the distinctiveness of genera erected and resurrected by Williams & Round [33] to better delimit the formerly widely circumscribed genus *Fragilaria* Lyngbye ([4] [13] [19] [20], among others).

4 Conclusion

The diatom flora from Sorata, Bolivia is diverse and comprises a variety of cosmopolitan and geographically restricted diatoms with many species adapted to alkaline habitats. This is in agreement with two previous studies showing high diversity for different localities within the Yungas [21] [28]. Many of the diatoms observed in collected samples could be new, but their description should be postponed until additional data are collected.

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