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Review Article

Ethnobotanical and pharmacological importance of *Taxus wallichiana* Zucc.

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Abstract

Taxus wallichiana Zucc. or the Himalayan Yew is a gymnosperm growing along the Himalayan region of Indian and adjoining countries. The plant is extensively used by local people for treatment of various diseases such as fever, headache, diarrhea, fractures, problems of nervous system etc. It also finds usage in Unani system of medicine. The plant is rich in various bioorganic compounds natural products such as hydrocarbons, terpene alcohols, terpenoids (including taxoids), organic acids etc. The plant has been explored for anti-inflammatory, analgesic, antipyretic, anticonvulsant, immunomodulatory, hepatoprotective and anticancer activity with satisfactory outcome. The pharmacological activity of the plant is largely due to the presence of large number of terpenoids. The bioactive constituents present in the plant interacts with a large number of biochemical pathways involved in inflammatory processes, cell division cycles and inhibits a number of enzymes to bring about its protective action against various diseases. In this review, an attempt have been made to highlight the beneficial properties of *Taxus wallichiana* in various levels of usage starting from its fundamental ethnobotanical use to pharmacological use involving both *in-vitro* and *in-vivo* studies. Insights into the molecular mechanisms of action of the active constituents in bringing about the beneficial activity have also been illustrated. The plant can very well become a source of medicine for better management of a large number of diseases including cancer.

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Introduction

The Himalayan region has been a rich source of medicinal plants for millions of population inhabiting in around the mountain ranges. The Himalayan region is rich in floral diversity and plants are extensively used by local people for their daily needs ranging from thatching and shelter, fuel, fodder, house hold items, medicines (1, 2). The Indian Himalayan region presently houses

8000 species of vascular plants of which 1748 are known to possess medicinal properties (3). Himalayan mountains are home of 232 families of angiosperms representing 2302 genera and 10452 species while the gymnosperms are represented by 8 families, 20 genera and 51 species (4). *Taxus wallichiana* is one such gymnosperm that grows in the Himalayan region. It is a small to medium sized evergreen tree with a height of 10 m to 28 m (5). The plant is widely distributed in Asia and its

occurrence spans from Afghanistan in the west to Philippines in the east and widely distributed in Himalayan regions of India and adjoining countries (6).

T. wallichiana is traditionally used by local people of Indian subcontinent for the cure of a number of ailments. In India, tincture prepared from aerial part of the plant is traditionally used for treatment of several diseases of central nervous system such as hysteria, grittiness, biliousness, epilepsy and nervousness. The plant also forms one of the components of the popular unani drug 'Zarnab' which is known to possess sedative and aphrodisiac properties (7). *T. wallichiana* is also used indigenously by people of Nepal for curing respiratory problems, bronchitis and cancer (8). The leaves of *T. wallichiana* is also used to prepare herbal tea for cure of epilepsy and indigestion (9). *T. wallichiana* is also reported to have immunomodulatory, anti-bacterial, anti-fungal, analgesic, anti-pyretic and anti-convulsant activities (10). Thus, based on the available reports, an attempt has been made to review the entire domain of beneficial properties of *T. wallichiana* with special emphasis on its chemical constituents, ethnobotanical uses and pharmaceutical applications.

Methodology

Extensive literature survey have been done using internet with PubMed, google scholar forming the search platform. Relevant research papers and review articles were selected in framing the article. The entire review article has been divided into an introductory phase which briefly describes geographical distribution, morphological features and taxonomic classification of *Taxus wallichiana*. Introductory segment is followed by detailed account of ethnobotanical uses of the plant. The article then extends into further insights into the chemical constituents of the plant and pharmacological uses and culminates with the discussion and conclusive remarks on the mode of action of the active principles in bringing about the pharmacological activities. The chemical structures have been drawn using ACD/Chem sketch Freeware 2018.2.5.

Results

Geographical distribution

Taxus wallichiana grows throughout the Himalayan region. In India it is widely distributed in the temperate zone of Himalayan mountains between altitudes of 1800m to 3300m above mean sea level (11). It is a slow growing understory species occurring in forests of Indian Himalayas and grows with *Abies pindrow*, *Quercus semecarpifolia*, *Q. floribunda*, *Q. leucotrichophora*, *Betula utilis*, *Acer caesium*, *Pinus wallichiana*,

Rhododendron arboreum and *Betula alnoides* (12). In the eastern Himalayas, the species grows in moist temperate zones at an altitude between 1600 to 2700 meters in West Kameng, Tawang, Lohit and Dibang Valley districts of Arunachal Pradesh (13). The plant also grows in hilly regions of Manipur and Meghalaya in north-eastern India (14). In Nepal, the plant is found in Rasuwa, Kavre, Kaski, Gorkha, Dolakha, Sindhupalanchok, Lamjung, Myagdi, Sakhawasabha, Taplejung and Katmandu districts (15). It is also found in the Kanchenjunga landscape, a region of eastern Nepal (parts of Taplejung, Panchthar, Ilam and Jhapa districts), Sikkim and North Bengal (Darjeeling and Jalpaiguri, and recently formed Alipurduar and Kalimpong districts) in India, and western Bhutan (portions of Haa, Chukha, Samtse, Dagana and Paro districts) (16). In Pakistan, the plant is present in Murree hills, Hazara, Swat, Dir, Chitral as undergrowth of other conifers and broad leaved deciduous plants (17). It is also reported from Gilgit-baltistan and also in SWAT valley on the foot hills of Hindukhush range in Pakistan (18, 19).

Description of the plant

Taxus wallichiana is a dioecious tree species (20). The stems are fluted with spreading branches. The barks are thin, reddish brown and scale like (21). Leaves dark grey in colour, glossy green above, paler beneath, linear, 2-3.8 × 0.3 cm in length, coriaceous, flattened, arranged in two vertical opposite rows. Cones are axillary and sessile. Male cones are solitary, axillary, sub-globose, bracts empty, with ten stamens. Female cones are solitary with few imbricate scales surrounding an erect ovule. Ovules are surrounded at base by membranous cup shaped disc. Fruit have bright red disc (Fig 1), succulent, enlarged, 7-8 mm in length. Seeds are olive-green in colour. Seeds dispersed by birds and animals. Growth of the trees are extremely low with 12-14 annual rings per 2.5 cm radius and girth increment 0.4 to 1.3 cm per year (22).

Taxonomic hierarchy (IUCN Taxonomic hierarchy)

Kingdom: Plantae

Division: Tracheophyta

Class: Coniferopsida

Order: Coniferales

Family: Taxaceae

Genus: *Taxus*

Species: *wallichiana* (23)

Synonym: *Taxus baccata* L. ssp. *Wallichiana* Zucc. Pilg.



Fig 1. Picture of *Taxus wallichiana* Zucc. showing ripe female fructification (24)

Population and Conservation status

Populations of *Taxus wallichiana* were existent in Himalayan regions since 3Ma BP (25). The plant is traditionally and extensively used by the locals of Himalayan region largely for primary healthcare purpose (26, 27). This has resulted in unregulated and unscientific harvesting of the plant ultimately leading to the decline and fragmentation of populations (28). The plant is thus marked as 'Endangered' in IUCN redlist with a continuous decrease in population (29). The plant is also included in CITES and negative list of exports of the government of India (30).

Ethnobotanical uses

Taxus wallichiana has been a plant of immense ethnobotanical use amongst the local people dwelling in the Himalayan region. They largely use *T. wallichiana* to cure various diseases. The various ethnobotanical use of *T. wallichiana* across various regions of Himalaya has been compiled in Table 1.

Table.1 Ethnobotanical use of *Taxus wallichiana* Zucc. across various Himalayan regions

#	Region	Vernacular Name	Ethnobotanical uses	Ref.
1.	Asi Ganga sub basin, Uttarakhand, India	Thuner	Bark and seed extract with warm water is given orally for treatment of internal wound.	(31)
2.	Urgam valley, Chamoli Garhwal, Uttarakhand, India	Thuner	Bark extract is used as tea for treatment of high blood pressure.	(32)
3.	Nanda Devi Biosphere Reserve, Uttarakhand, India	Thuner	Bark used as a substitute of tea. The powdered bark is used for the treatment of cold.	(33)
4.	Kedarnath wildlife sanctuary, Garhwal Himalayas, Uttarakhand, India		Bark and bark paste used for the treatment of fractured bones, headache, breast piles	(34)
5.	Niti Valley, Uttarakhand, India	Thuner	Dry powder of bark with salt and ghee is mixed with water to make tea and used for treatment of high blood pressure and cancer. Paste of bark with egg yolk is used as plaster for treatment of fracture.	(35)
6.	Jakholi Block,	Thuner	Juice of leaves are used for the	(36)
	Rudraprayag district, Uttarakhand, India		treatment of boils, cuts and wounds	
7.	Mornaula Reserve forest, Kumaon, Uttarakhand, India	Thuner	Bark, oil and leaves are used for treatment of cancer. Bark is also used as fuel.	(37)
8.	Shimla Hills, Himachal Pradesh, India	Thuno, Barmi	Tincture from young shoots are used for treatment of headache, giddiness, feeble and falling pulse, diarrhoea and severe biliousness. Leaves are antispasmodic and used for treatment of nervousness, hysteria, epilepsy and stones.	(38)
9.	Manali wildlife sanctuary, Himachal Pradesh	Rakhal	Barks and leaves are used for treatment of cancer, swelling and as contraceptive.	(39)
10.	Pabbar Valley, Himachal Pradesh	Thuna	Tea prepared from needle and bark are used for treatment of congestion and cough.	(40)
11.	Mandi and Hamirpur district, Himachal Pradesh	Rakhala/ Talispatra	Tea prepared from barks and leaves are used to treat asthma. Bark is used for the treatment of cancer.	(41)
12.	Churah subdivision, District Chamba, Himachal Pradesh, India	Nadgaun/ Brahmi	Bark is used as flavouring agent.	(42)
13.	Shimla water catchment sanctuary, Himachal Pradesh, India	Rakhal	Leaves used to cure cancer. Bark used for preparation of tea.	(43)
14.	Kathua, Jammu & Kashmir, India	Barmi	Decoction of leaves are used to cure asthma, bronchitis, cough, indigestion and epilepsy.	(44, 45)
15.	Rajouri, Jammu & Kashmir, India	Barmi	Decoction of leaves are used to cure asthma, bronchitis and cough.	(46)
16.	Ganderbal, Kashmir, India		Tea prepared from boiling bark in water is used for cure of asthma, giddiness, arthritis, tumour growths.	(47)
17.	Bangus valley, Kashmir, India	Postul	Tea made from bark is used to cure sickness in winter.	(48)
18.	Bandipora, Jammu and Kashmir, India	Postul /Brammi	Bark extract is made into a tea and is used for curing of asthma, headache, giddiness, tumour growths.	(49)
19.	Galliyat, NWFP, Pakistan	Bermi	Decoction of stem is used for treatment of tuberculosis.	(50)
20.	Neelam valley, Muzaffarabad, Pakistan occupied Kashmir	Birmi	Tea from leaves are used to cure asthma and high fever.	(51)
21.	Leepa valley, Muzaffarabad, Pakistan occupied Kashmir	Birmi	Leaf and bark extract is used to treat tumours. Decoction of leaf with honey is used for the treatment of hey fever, flatulation, epilepsy and asthma.	(52)
22.	Kel, Pakistan occupied Kashmir		Decoction of bark is used for treatment of cancer.	(53)
23.	Shogran valley, Pakistan		Plant used for the treatment of cancer, cardiac disorders, head ache, renal disorders and digestive disorders. The plant is antispasmodic, purgative and antirheumatic.	(54)
24.	Manaslu, Sagarmatha and Kanchenjunga region, Nepal		Used for treatment of cancer and jaundice.	(55)

Table 2. Major constituents of different plant parts of *Taxus wallichiana* Zucc

Plant Parts	Nature of Compounds	Name of Compound	Ref.	
Essential oil from leaves	Alkane	n-Eicosane; Docosane ; n-Pentacosane.	(67)	
	Alkene	Santolinatriene.		
	Alcohol	Geraniol; Globulol; Eugenol; Myrtenol; (E)-Verbenol; n-Hexenol; (Z)-3-Hexenol; (E)-2-Hexenol; n-Hep-tan-2-ol; 1-Hepten-3-ol; (E)-2-Octen-1-ol; (Z)-2-Octen-1-ol; (E)-2-Nonenol; 1-Octanol.		
	Aldehyde	Benzaldehyde; Anisaldehyde; n-Heptanal; n-Octanal; (E)-2-Octenal; n-Nonanal; (E)-2-Nonenal; Dodecanal.		
	Organic acids	Benzoic acid; Hexanoic acid.		
	Organic acid esters	(Z)-3-Hexenyl formate; Octyl formate; (Z)-3-Hexenyl acetate; (E)-2-Hexenyl acetate; (E)-3-Heptenyl acetate; Benzyl acetate; Anisyl acetate;n-Octyl acetate; Sabinyl acetate;(E)-2-Hexenyl-n-hexanoate;Isopropyl-n-octanoate; (Z)-3-Hexenyl benzoate; Methyl benzoate; Geranyl-n-heptanoate; Geranyl benzoate;n-Amyl anisoate; Geranyl tiglate; Methyl salicylate.		
	Terpenes	α -Pinene; β -Pinene; Camphor; β -Caryophyllene; Caryophyllene oxide; (Z)- β -Ocimene; (Z)-Sabinene hydrate; (Z)-Pinene hydrate.		
Leaves		Taxol; 10-deacetylbaccatin III; baccatin IV; 1-hydroxybaccatin I; 2'-deacetoxydecinnamoyltaxinine J; 2'-deacetoxytaxinine J;2-acetoxybrevifoliol	(68)	
		Brevifoliol;2-acetoxybrevifoliol	(68, 69)	
	Taxoids (terpenoids)		5 α O-(3'-dimethylamino-3'-phenylpropionyl) taxinine M;7-O-acetyltaxine A; 2 α -acetoxy-2' β -deacetylaustrospicatine.	(70)
			14- β -hydroxy- 10- deacetylbaccatin III;2-Debenzoyl-14 β -benzoyloxy-10-deacetylbaccatin III; 14 β -Hydroxy-1 0-deacetylbaccatin V.	(71)
			19-debenzoyl-19-acetyltaxinineM; 13-deacetyltaxuspine A; 10-debenzoyl-2a-acetoxybrevifoliol,	(72)
			2-deacetoxytaxinine B	(73)
			Wallifoliol; cephalomannine; 1-O-deacetylbaccatin 11	(69)
Barks	Ketone	4-(4'-hydroxyphenyl)-2-butanone	(56)	
	Alcohol	4-(4'-hydroxyphenyl)-2-butanol; 9-hydroxy-4,7-megastigmadiene-3-one-3-oxo- α -ionol		
	Taxoids (Terpenoids)	Taxayuntin E ; Taxayuntin G; Taxayuntin J; Taxinine A;2-Deacetoxy taxinine B; 2-Deacetyl-5-decinnamoyl taxinine E;Taxinine J ; 2-Deacetoxy taxinine J ;5-Decinnamoyl taxinine J; Taxinine M; 19-Debenzoyl-19-acetyl taxinine M; Taxchin A;Taxchin B; 1-Hydroxy-5-deacetyl baccatin I; Baccatin III; Baccatin IV; 10,13-Deacetyl-abeobaccatin IV; 1-Dehydroxy baccatin VI;13-Deacetyl baccatin VI;9-O-Benzoyl-9,10-dideacetyl (15-1)-abeo baccatin VI;9-Benzoyl-9-deacetyl11(15-1)abeo baccatin VI Taxayunnanine C; 2'-Deacetyl austrotaxine, 2-Acetoxy-2',7-dideacetoxy-1-hydroxy austrospicatine; 2-Acetoxy-2'-deacetoxy austrospicatine; 2'-Deacetoxy austrospicatine; 7-Xylosyl-10-deacetyl taxol D; 7-Xylosyl-10-deacetyl taxol;10-Deacetyl taxol; 7-Xylosyl taxol; 7-Xylosyl-10-deacetyl taxol C; N-Methyl taxol C ;10-Deacetyl epi-taxol; 7-Xylosyl taxol C; Epi-taxol; 2-Benzoyloxy-7,9,10, 13-tetraacetoxy-4(20); 11 -taxadiene,2,5,9-Trihydroxy-10, 13-diacetoxy-4(20), 11-taxadiene; 5-Hydroxy-9,10-dia-cetoxy-13-oxo-4(20), 11-taxadiene; 2-Hydroxy-5,10,14-triacetoxy 4(20), 11-taxadiene; 5,7,9,10,13-Pentaacetoxy 4(20), 11-taxadiene;7,9,10,13-Tetraacetoxy-5-(3'-acetylamino3'-phenyl)propionyloxy-4(20), 11-taxadiene;Hydroxy-triacetoxy-5(3'-dimethylamino-3'phenyl)propionyloxy4(20), 11-taxadiene;Taxusin;Yunnanxane.	(74)	
Heart wood		Taxusin ; 7-xylosyl-10deacetyltaxol C.	(75)	
	Taxoids	13-acetyl-13-decinnamoyltaxchinin.	(76)	
Roots	Taxoids	Baccatin III; Baccatin IV; Taxusin; 1 β -hydroxybaccatin I; Penta acetoxy taxadiene 7; 7-xyloxyl-10-deacetyl-taxol.	(65)	
	Lignans	(-) Seco-isolariciresinol; Taxiresinol; Isotaxiresinol.	(66)	
	Lignans	α -Conidendrin; Formosanol; Methyl- α -Conidendral; α -Intermedianol;1, 4-methano-2-benzoxepin-10-methanol, 1, 3, 4, 5-tetrahydro-7-hydroxy-5-(4-hydroxy-3methoxyphenyl)-8-methoxy.		
	Sesquiterpene lactone	Cinnamolide.		
Seeds	Triterpenoid	Ursolic acid.	(77)	
	Cyanogenic glucoside	Amygdalin.		
	Phytosterol	β -Sitosterol.		

Chemical constituents

Taxus wallichiana has been a plant of extensive study due to its diverse ethnobotanical uses. The plant also exhibits a wide range of

pharmacological properties and detailed investigations about the chemical constituents of the plant have been undertaken by a number of researchers. Essential oil is a major ingredient of

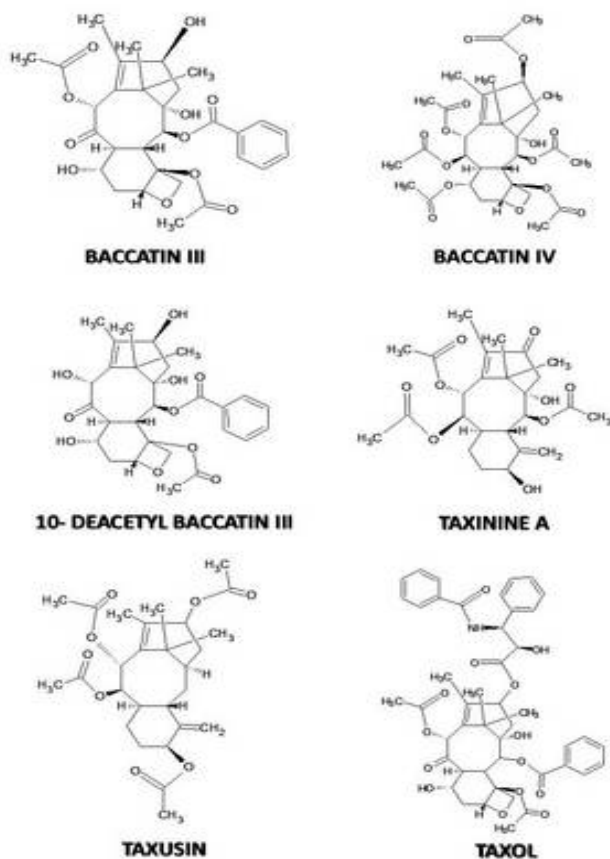


Fig 3. Chemical structures of selected Taxoids present in *Taxus wallichiana* Zucc.

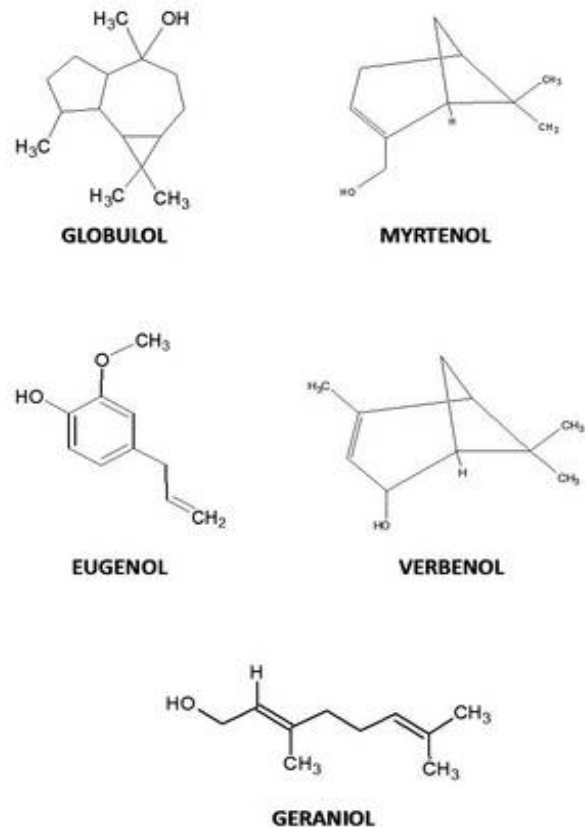


Fig 4. Chemical Structure of selected alcohols present in *Taxus wallichiana* Zucc.

leaves obtained through hydrodistillation process. The major constituents of essential oil are long chain hydrocarbons, terpene alcohols such as geraniol, globulol, eugenol, myrtenol and (E)-verbenol. In addition to it, the essential oil also contains fatty alcohol such with varying chain length, aldehydes, organic acid esters and terpenes (56). These terpenes and terpene alcohols are largely responsible for the characteristic fragrance of essential oil (57). In addition to essential oils, all parts of *T. wallichiana* contains the most unique group of compound by the name taxoids or taxanes. These compounds are largely responsible for pharmacological activities of *Taxus* (58, 59). The taxoids are fundamentally diterpenoids and contains 6/8/6-membered ring skeleton known as taxane skeleton which is chemically a pentamethyl [9.3.1.0] 3, 8 tricyclopentadecane skeleton (60, 61).

The most important and pharmacologically important representative of the taxoids present in *T. wallichiana* is Taxol. The biogenesis of taxol involves the condensation of the three isoprenyl diphosphate (IPP) units with dimethylallyl diphosphate (DMAPP) both of which are produced either through mevalonic (MVA) pathway in the cytosol or via the methyl erythritol phosphate (MEP) pathway in plastids. The first determining step in the biogenetic pathway is cyclization of geranyl geranyl diphosphate (GGPP) to taxadiene followed by eight cytochrome P450-mediated oxygenations, three CoA-dependent acyl/aroyl

transfers, an oxidation at C9, and oxetane (D-ring) formation finally resulting in formation of intermediate baccatin III (62, 63) (Fig. 2). The final stage of taxol biosynthesis involves the assembly of C13 side chain appended to the Baccatin molecule (64). Analysis of bark, root and other parts of the plants revealed the presence of taxol, baccatins and other intermediary molecules and taxol analogues (Table 2). Apart from taxoids, *T. wallichiana* also contains lignans such as (-) seco-isolariciresinol, taxiresinol, isotaxiresinol, formasonol, sesquiterpene lactone, triterpenoid cyanogenic glucoside and phytosterols (65, 66). Chemical structures of selected compounds present in *T. wallichiana* is depicted in Figs. 2, 3 & 4 respectively. Most of these compounds have been reported to have pharmacological importance and have been discussed in this article. The various compounds isolated from different parts of *T. wallichiana* are listed in Table 2.

Pharmacological activities of *Taxus wallichiana* Zucc.

Anti-inflammatory activity

Inflammation is associated with onset of a number of diseases including asthma (78), rheumatoid arthritis (79), atherosclerosis (80), chronic venous insufficiency (81), diabetes (82), cancer (83) etc. Herbal remedies have proven to be an effective way to reduce inflammatory processes (84). In the

same line *Taxus wallichiana* have proven its efficacy to combat inflammatory processes. Taxusabietane A, isolated from *T. wallichiana* was reported to possess anti-inflammatory potential.

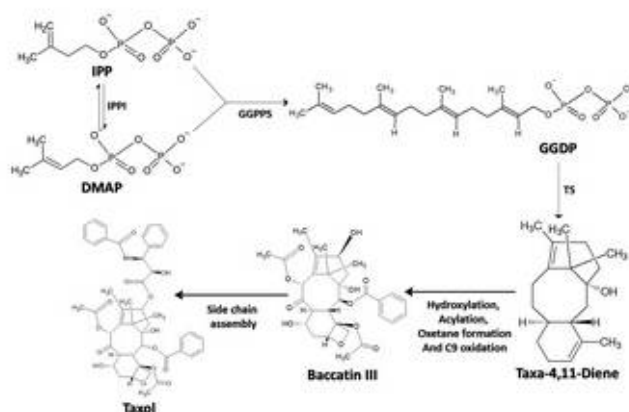


Fig 2. Schematic representation of major steps associated with biosynthesis of taxol

IPP: isopentenylidiphosphate

DMAPP: dimethylallyldiphosphate

GGDP: Geranylgeranyldiphosphate

IPPI: isopentenylidiphosphate isomerase

GGPPS: geranylgeranyldiphosphate synthase

TS: taxadiene synthase

The results obtained from a study indicated that taxusabietane A showed lipoxygenase inhibitory activity with an IC₅₀ value of 57±0.31 μM which was significant in relation to standard compounds baicalein and tenidap sodium with an IC₅₀ value of 22.1±0.03 and 41.6±0.02 μM respectively. In addition to it, taxusabietane A at 5mg/kg and 10mg/kg dose also inhibited carrageenan induced oedema in adult wistar rats though in this case standard compound indomethacin at same dose exhibited comparatively better result (85). Another study reported that taxoids namely tasumatrol B, 1, 13-diacetyl-10-deacetyl baccatin III (10-DAD) and 4-deacetylbaccatin III (4-DAB), isolated from *T. wallichiana* is significantly effective in bringing down inflammations induced by carrageenan and cotton pellet in rats. At 20mg/kg and 40mg/kg of dose, the tested compounds showed significant inhibition of paw oedema in carrageenan induced inflammation model out of which tasumatrol B was observed to be most effective. In addition to it, cotton pellet test revealed that all the three test compounds showed inhibition against inflammation and granuloma accumulation (86). Two more abietane diterpenoids namely taxusabietane C and taxamairin F have also reported to show lipoxygenase inhibitory effect with 69 ± 0.31 and 73 ± 0.14 μM, respectively (87).

Analgesic, antipyretic and anticonvulsant activity

The analgesic activity of tasumatrol B, a taxane isolated from the bark of *Taxus wallichiana* have been reported, It has been observed that

tasumatrol B administered at 40mg/kg in wistar rats significantly decreased the writhing count in acetic acid writhing experiment (86). The analgesic activity of *T. wallichiana* was observed by the formalin test in rats. It was observed that upon treatment with 50, 100 and 200 mg/kg of the extract of *T. wallichiana* to rats previously injected with 0.05 ml of formalin in the plantar surface of hind paw, the pain score severity showed a concentration dependent decline in both early and late phases of analgesia. In addition to it, the antipyretic activity was investigated by injecting 15% aqueous solution of yeast at 10 ml/kg dose to induce pyrexia. This was followed by intraperitoneal injection of 50, 100 and 200 mg/kg of extract. It was observed that all the dose of plant extract showed significant inhibition of pyrexia induced by yeast and the antipyretic effect of 200 mg/kg of extract is comparable to 20 mg/kg of paracetamol between 30 and 60 minute of treatment. The anticonvulsant activity was also tested observing the effect of plant extracts on pentylenetetrazole induced seizures in rats. It was observed that pretreatment of 50, 100 and 200 mg/kg of plant extract significantly reduced mioclonus seizures. Additionally, 100 and 200 mg/kg of extracts significantly delayed the onset of first clonus seizures while 200 mg/kg of extract delayed the tonous seizures. Extracts of all doses protected the rats from tonic-clonic seizures. Diazepam was used as a control and showed remarkable anticonvulsant activity at 7.5 mg/kg. There was also a decrease in percentage of mortality in animals upon pretreatment with extracts (88).

Hepatoprotective activity

The protective action of methanolic extract of *Taxus wallichiana* against carbon tetrachloride induced hepatotoxicity in rats was reported from a recent study. It was observed that CCl₄ treatment resulted in significant elevation of liver enzyme markers namely aspartate transaminase (AST), alanine transaminase (ALT) and lactate dehydrogenase (LDH). Treatment of animals with 1 ml of 100 and 300 mg/kg/body weight resulted in decrease in the levels of liver marker enzymes histopathological investigation of liver of rats treated with plant extracts revealed an almost normal hepatic architecture with less infiltration of fat and absence of necrosis all of which were prominent in rats treated with carbon tetrachloride (89).

Immunomodulatory activity

The immunomodulatory potential of *Taxus wallichiana* was investigated using human lymphocyte as the experimental system. In the experiment, cyclophosphamide was used to suppress the proliferation of lymphocytes and this condition was reversed by cotreatment of 1-hydroxy-2-deacetoxy-5-decinnamoyl-taxinine J (1

µg/ml) with concanavaline A (5 µg/ml), an immune stimulant (90).

Anticancer activity

Some studies have been undertaken to investigate the anticancer potential of compounds isolated from *Taxus wallichiana*. A study reported that 1-hydroxy-2-deacetoxy-5-decinnamoyl-taxinine J was cytotoxic to five cancer cell lines namely MCF7, WRL-68, KB, PA-1, Colo 320DM human cancer cell lines as determined by MTT and clonogenic assays (90). Another study have shown cytotoxicity of taxiresinol, a lignan isolated from heartwood of the plant against human liver, colon, ovarian and breast cancer cell lines (91). Taxawallin I, a new taxoid isolated from *T. wallichiana* has been reported to show toxicity against HepG2, A498, NCI-H226 and MDR 2780AD cancer cells (92). Another study reported isolation of four novel taxane derivatives namely N-debenzoyl-N-methyl-N-heptanoyl-taxol, N-debenzoyl-N-methyl-N-octanoyl-taxol, N-debenzoyl-N-methyl-N-(4-methylhexanoyl)-taxol, and N-debenzoyl-N-methyl-N-[(4Z)-1-oxo-4-tenenoyl]-taxol from ethanol extract of whole plant. It is also reported that the taxanes were inhibitory towards MCF-7, A549, and 3-AO cancer cell lines and had microtubule stabilizing properties (93).

The term taxane or taxoids refers to all the terpenoid compounds having molecular structure based on baccatin unit, either obtained naturally from *Taxus sp* or are semi synthetically or synthetically prepared exhibiting anticancer properties. Taxanes interacts with the microtubules involved in mitotic process. Taxanes stabilize the microtubules of cells which counteract their depolymerization. Thus, correct separation of two identical sets of chromosomes and their consequent transfer during cell division are inhibited resulting in blockage of cell mitosis ultimately leading to cell death (94). Taxanes promote microtubule polymerization and arrest mitosis through activation of spindle assembly check point and keeping a small number of unattached kinetochores to the microtubules. This delays mitotic metaphase progression and inhibits anaphase prompting complex (95). On a molecular level, binds to a pocket in β - tubulin that faces microtubule lumen and is near the lateral interface between protofilaments thereby affecting normal function and cellular processes (96). The binding of Paclitaxel to β - tubulin subunit results in stabilization of microtubules through induction of conformational changes of the M-loop of β - tubulin which result in more stable lateral interaction between adjacent protofilaments thereby changing microtubule dynamics and inducing mitotic block and triggering apoptosis of cancer cells (97, 98).

Moreover, taxanes also exhibit apoptotic action interacting with various proteins and enzymes which are involved in cell cycle, apoptosis and cell death. Increased reactive oxygen species (ROS) is one of the earliest events of apoptosis and

is brought about by taxanes (99). Taxanes also initiate decrease in mitochondrial membrane potential ($\Delta\Psi_m$) and induces opening of mitochondrial membrane permeability pore resulting in release of calcium and cytochrome c from mitochondria (100, 101). Caspases are the family of endoproteases that play an important role in cell inflammation and cell death (102). Taxanes also activates caspases thus initiating cell death and apoptosis (103). B-cell lymphoma (Bcl-2) is the key protein which regulates programmed cell death and apoptosis (104). They may be divided into two major groups namely (a) antiapoptotic protein (BCL-2, BCL-XL, MCL-1, BFL-1, BCL-W, and BCL2L10) and (b) proapoptotic proteins (BAK, BAX) (105). The apoptosis in cancer cells is further induced by phosphorylation and inactivation of antiapoptotic Bcl-2 by taxane (106). Along with Bcl-2, p53 also plays a major role in apoptotic process. It is an important tumour suppressor gene which regulates downstream expression of other genes involved in DNA repair, cell cycle arrest and apoptosis (107). Taxanes are reported to act as a p53 inducers thereby enhancing the apoptotic process (108). The p21 is an universal cyclin dependent kinase (CDK) inhibitor, controlled by p53 and physically interacts and inhibits cyclin-CDK2, cyclin-CDK1, cyclin-CDK4/6 complexes thus regulates progression of cell cycle during G1 and S phases (109). Taxanes results in increase of expression p21 through upregulation of p53 (110).

Toxicity

Toxicity of *Taxus sp* is known to humans since early civilization. Juice of the plant was applied to arrows for hunting and leaves were used for homicide and suicide (111). The main active compounds of the plant include a mixture of diterpenoid alkaloids namely taxine A and B, isotaxine B, taxol B and are responsible for toxicity causing toxicity resulting in the occurrence of symptoms like nausea, vomiting, diffuse abdominal pain, tachycardia (initially) and convulsions, followed by bradycardia and respiratory muscle paralysis (112). The time from ingesting lethal dose of *Taxus* toxin to death usually varies between 2-5 hours with symptoms occurring between 30 minutes to 1 hour after ingestion (113). Generally, the *Taxus* toxins acts by generation of a block in the distal portion of the conduction system of the heart resulting in fatal arrhythmia (114). It is further reported from a study that taxine-B inhibits calcium and sodium transport in myocardial cells and interferes with heart's conducting system thereby acting as a cardiac depressant (115).

Conservational approaches and production of bioactive compounds

The population of *Taxus wallichiana* have taken a toll due to its ever increasing demand among the localities accompanied by high rate of collection. To make the things even more grave, the plant exhibits slow growth and poor regeneration (116). This has resulted in number of conservational

approaches to save the population from extinction. Tissue culture method have been a versatile approach for mass propagation of a number of plants (117). The long dormancy period of the plant can overcome by culture of zygotic embryos which can develop into full grown seedlings in 10-12 weeks (118). A study reported that shoot elongation and root induction through shoot tip culture is feasible and may be applicable for propagation of the plant (119). Another study reported regeneration of *T. wallichiana* plant via shoot organogenesis from callus cultures derived from zygotic embryos (120). Apart from tissue culture techniques, stem cuttings treated with growth promoting substances have also been proved effective for propagation of the plant (121). Another recent study reported that treatment of shoot cuttings of the plant with indole acetic acid (IAA), indole butyric acid (IBA) and naphthoxy acetic acid (NAA) resulted in effective initiation of roots (122). Tissue culture techniques are not only used for mass propagation but also for the production of bioactive compounds. There have been reports that extracts from the cell cultures of the plant contains taxol, deacetyl baccatin III and baccatin III (123). Another study detected the presence of taxol (0.8499%) in the callus culture (124). Use of bioreactors for production of taxol and baccatin III from suspension cultures have also been reported from a study (125).

Discussion

The genus *Taxus* or Yew holds immense importance globally from pharmacological point of view. A lot of research work have been undertaken to explore the pharmacological importance of the genus. The most important amongst them is exploration of Pacific Yew or *Taxus brevifolia*. Exploration on the antineoplastic activity of the active constituent of *T. brevifolia* started way back in 1960 (126). It consequently underwent a series of modifications and culminated in commercial production as Paclitaxel by Bristol-Myers Squibb (BMS) in the year 1992 (127). Since then there have been no stopping and a number of variants and semisynthetic analogues such as docetaxel and cabazitaxel were developed all of which possess same anticancer activity. Moreover, studies showed that extracts of European yew, *Taxus baccata*, also have the potential to inhibit Caov-4 and HeLa cancer cell lines (128). It is also reported from a study that the bark extract of *T. baccata* also possess anti-inflammatory properties (129). There are also reports that the leaves of *T. baccata* also have bronchodilating properties and beneficial effect on asthma (130). Japanese Yew, *Taxus cuspidata* is also reported to possess inhibitory activity against MCF-7 (breast), PG and A549 (lung), PC-3M-1E8 (prostate), BGC-823 (gastric), WM451 (melanoma), Bel-7402 (hepatocellular), KB (oral squamous),

HeLa (cervical), and HL-60 (leukemic) cell lines (131).

Across the world there has been a growing demand of herbal medicines and 80% of the total population are relying on herbal remedies for primary healthcare (132). The figures are similar for Indian population of which 70% relies on plants as source of medicines (133). Now coming back to Himalayan Yew or *Taxus wallichiana*, it is reported that the plant finds extensive use by the people living in various location of Himalayas. The use of plant has been assimilated in their tradition and thus justifies its ethnobotanical and ethnomedicinal significance. Thus, there is a strong requirement for a reverse pharmacological process in which the traditional knowledge and usage pattern of Himalayan Yew can be better explored in the laboratory condition for validation of its pharmacological efficacy. In this regard, the plant has been investigated to some extent for a number of pharmacological activities which has yielded promising outcome. Thus, this species is at par with the other species of *Taxus*, many of which have been extensively investigated for their pharmacological actions. However most of the investigations have been made of cell lines or in animal systems. Experimentation using clinical trials using humans are further required for consequent development of drugs. Additionally, exploration of anticancer potential from *Taxus* sp has become a matter of priority since last 50 years and as mentioned earlier. Paclitaxel is a noble outcome of extensive research on *T. brevifolia*. In a similar manner, the Himalayan Yew can also be explored further for its anticancer potential leading to development and commercialization of drugs required for cancer treatment. In this aspect there are few reports of detection taxol in cell and callus culture of *T. wallichiana*. This avenue also needs further elaboration and investigation with a motive of increasing the concentration and yield of Taxol through modification of ambient culture condition. This would lead to a cost effective approach in the development of anticancer drugs from *T. wallichiana*.

Presently, over exploitation of *T. wallichiana* have resulted in its depletion. Isolation of population through habitat fragmentation leads to restriction of connectivity, resulting in low levels of gene flow between the population with subsequent lower genetic diversity and higher genetic differentiation in and among remaining populations (134). Moreover, smaller isolated populations also undergo frequent inbreeding which often express deleterious alleles leading to reduction in reproductive capacity and low offspring survival (135). Thus, studies on the genetic diversity of the plant is extremely relevant as an important avenue towards conservation as well an indicator of possible extinction. An important approach towards conservation of diminishing population of *T. wallichiana* is to

maintain an effective population size so that the level of genetic variation can be maintained. Thus, proper planning and management of this species in their natural habitat will be extremely fruitful in harvesting pharmacologically active principles from the plant. Techniques of molecular biology and biotechnology should also be involved in the overall process of conservation.

Conclusion

Taxus wallichiana or Himalayan Yew is an endangered gymnosperm that grows in the Himalayan region and is of immense Ethnobotanical importance amongst the people. They contain huge amount of phenolics and terpenoids which forms the backbone of their medicinal potential. The plant has proven its efficacy from pharmacological point of view and can be very well utilised as a cost effective source of medicine for management of diseases of the people of Indian subcontinent. The bioprospection of this plant thus becomes extremely relevant for the benefit of humans.

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Conflict of Interest

Author declares that he has no conflict of interest in the publication.

References

- Rana D, Bhatt A, Lal B. Ethnobotanical knowledge among the semi-pastoral Gujjar tribe in the high altitude (Adhwari's) of Churah subdivision, district Chamba, Western Himalaya. *J Ethnobiol Ethnomed.*2019;15(1):10. <https://doi.org/10.1186/s13002-019-0286-3>
- Pala NA, Sarkar BC, Shukla G, Chettri N, Deb S, Bhat JA, et al. Floristic composition and utilization of ethnomedicinal plant species in home gardens of the Eastern Himalaya. *J Ethnobiol Ethnomed.*2019;15(1):14. <https://doi.org/10.1186/s13002-019-0293-4>
- Joshi RK, Satyal P, Setzer WN. Himalayan Aromatic Medicinal Plants: A Review of their Ethnopharmacology, Volatile Phytochemistry, and Biological Activities. *Medicines (Basel).* 2016;3(1):6. <https://doi.org/10.3390/medicines3010006>
- Rana SK, Rawat GS. Database of Himalayan Plants Based on Published Floras during a Century. *Data.* 2017;2(4):36. <https://doi.org/10.3390/data2040036>.
- Juyal D, Thawani V, Thaledi S, Joshi M. Ethnomedical properties of *Taxus wallichiana* Zucc. (Himalayan yew). *J Tradit Complement Med.*2014;4(3):159-61. <https://doi.org/10.4103/2225-4110.136544>
- Hussain A, Qarshi IA, Nazir H, Ullah I, Rashid M, Shinwari ZK. In-vitro callogenesis and organogenesis in *Taxus wallichiana* Zucc. *The Himalayan Yew. Pak J Bot.*2013;45(5):1755-59.
- Sharma H, Garg M. Neuropharmacological activities of *Taxus wallichiana* bark in Swiss albino mice. *Indian J Pharmacol.*2015;47(3):299-303. <http://dx.doi.org/10.4103/0253-7613.157128>
- Gaire BP, Subedi L. Medicinal Plant Diversity and their Pharmacological aspects of Nepal Himalayas. *Phcog J.*2011;3(25):6-17. <https://doi.org/10.5530/pj.2011.25.2>
- Aboutabl ME. Antiepileptic drugs: progress and development. *Egypt Pharmaceut J.*2018;17(3):129-40.
- Rahman S, Salehin F, Uddin MJ, Zahid A. *Taxus Wallichiana* Zucc. (Himalayan Yew): insights on its antimicrobial and pharmacological activities. *OA Alternative Medicine.*2013;1(1): 3.
- Purohit A, Maikhuri RK, Rao KS, Nautiyal S. Impact of bark removal on survival of *Taxus baccata* L. (Himalayan yew) in Nanda Devi Biosphere Reserve, Garhwal Himalaya, India. *Curr Sci.*2001;81(5):586-90.
- Pant S, Samant SS. Population Ecology of the Endangered Himalayan Yew in Khokhan Wildlife Sanctuary of North Western Himalaya for Conservation Management. *J Mt Sci.* 2008;5:257-64. <https://doi.org/10.1007/s11629-008-0078-z>
- Nimasow G, Nimasow OD, Rawat JS, Tsering G, Litin T. Remote sensing and GIS-based suitability modeling of medicinal plant (*Taxus baccata* Linn.) in Tawang district, Arunachal Pradesh, India. *Curr Sci.*2016;110(2):219-27. <https://doi.org/10.18520/cs/v110/i2/219-227>
- Patel P, Patel K, Gandhi T. Evaluation of Effect of *Taxus baccata* Leaves Extract on Bronchoconstriction and Bronchial Hyperreactivity in Experimental Animals. *J Young Pharm.*2011;3(1):41-47. <https://doi.org/10.4103/0975-1483.76418>
- Poudel RC, Gao LM, Möller M, Baral SR, Uprety Y, Liu J, et al. Yews (*Taxus*) along the Hindu Kush-Himalayan region: exploring the ethnopharmacological relevance among communities of Mongol and Caucasian origins. *J Ethnopharmacol.*2013;147(1):190-203. <https://doi.org/10.1016/j.jep.2013.02.031>
- Kandel P, Chettri N, Chaudhary RP, Badola HK, Gaira KS, Wangchuk S, et al. Plant diversity of the Kangchenjunga Landscape, Eastern Himalayas. *Plant Divers.* 2019; 41(3):153-65. <https://doi.org/10.1016/j.pld.2019.04.006>
- Chaudhri II, Distribution of gymnosperms in West Pakistan. *Vegetatio.*1963;11(5-6):372-82.
- Ismail I, Sohail M, Gilani H, Ali A, Hussain K, Hussain K, et al. Forest inventory and analysis in Gilgit-Baltistan A contribution towards developing a forest inventory for all Pakistan. *International Journal of Climate Change Strategies and Management.*2018;10(4):616-31. <https://doi.org/10.1108/IJCCSM-05-2017-0100>
- Ali K. A comparative assessment of climate change effect on some of the important tree species of Hindu-Kush Himalayas, using predictive modelling techniques. *Int J Adv Res.*2015;3(5):1230-40.
- Yang JB, Li HT, Li DZ, Liu J, Gao LM. Isolation and Characterization of Microsatellite Markers in the Endangered Species *Taxus wallichiana* Using the FIASCO Method. *Hort Sci.*2009;44(7):2043-45. <https://doi.org/10.21273/HORTSCI.44.7.2043>
- Bhuju S, Gauchan DP. *Taxus wallichiana* (Zucc.), an Endangered Anti-Cancerous Plant: A Review. *Int J Res.*2018;5(21):10-21.

22. ENVIS Centre on Floral Diversity. Hosted by Botanical Survey of India, Kolkata, West Bengal. Sponsored by: Ministry of Environment, Forest & Climate Change, Govt of India. Available from: <http://www.bsienvs.nic.in/CITES/Taxus%20wallichiana.pdf>
23. India Biodiversity Portal. Available from: <https://indiabiodiversity.org/species/show/258690>
24. Viet Delta Corporation. Available from: <http://herbvietnam.com/>
25. Uniyal SK. Bark removal and population structure of *Taxus wallichiana* Zucc. in a temperate mixed conifer forest of western Himalaya. *Environ Monit Assess.* 2013;185(4):2921-28. <https://doi.org/10.1007/s10661-012-2760-4>
26. Malik K, Ahmad M, Zafar M, Ullah R, Mahmood HM, Parveen B, et al. An ethnobotanical study of medicinal plants used to treat skin diseases in northern Pakistan. *BMC Complement Altern Med.* 2019;19(1):210. <https://doi.org/10.1186/s12906-019-2605-6>
27. Rana PK, Kumar P, Singhal VK, Rana JC. Uses of local plant biodiversity among the tribal communities of Pangi Valley of District Chamba in cold desert Himalaya, India. *Scientific World Journal.* 2014; 2014:75389. <https://doi.org/10.1155/2014/753289>
28. Poudel RC, Möller M, Liu J, Gao LM, Baral SR, Li DZ. Low genetic diversity and high inbreeding of the endangered yews in Central Himalaya: implications for conservation of their highly fragmented populations. *Biodiversity distrib.* 2014;20:1270-84. <https://doi.org/10.1111/ddi.12237>
29. Thomas P, Farjon A. *Taxus wallichiana*. The IUCN Red List of Threatened Species 2011: e.T46171879A9730085. <http://dx.doi.org/10.2305/IUCN.UK.2011-2.RLTS.T46171879A9730085.en>. Available from: <https://www.iucnredlist.org/species/46171879/9730085>
30. ENVIS Centre on Floral Diversity Hosted by Botanical Survey of India, Kolkata, West Bengal. Available from: http://www.bsienvs.nic.in/Database/bsi_3949.aspx
31. Nand K, Naithani S. Ethnobotanical uses of wild medicinal plants by the local community in the Asi Ganga sub-basin, Western Himalaya. *J Complement Med Res.* 2018;9(1):34-46. <https://doi.org/10.5455/jcmr.20180507034822>
32. Singh A, Hart R, Chandra S, Nautiyal MC, Sayok AK. Traditional Herbal Knowledge among the Inhabitants: A Case Study in Urgan Valley of Chamoli Garhwal, Uttarakhand, India. *Evid Based Complement Alternat Med.* 2019;2019:5656925. <https://doi.org/10.1155/2019/5656925>.
33. Tiwari JK, Dangwal LR, Rana CS, Tiwari P, Ballabha R. Indigenous uses of plant species in Nanda Devi Biosphere Reserve, Uttarakhand, India. *Rep Opinion.* 2010;2(2):67-70.
34. Bhat JA, Kumar M, Bussmann RW. Ecological status and traditional knowledge of medicinal plants in Kedarnath Wildlife Sanctuary of Garhwal Himalaya, India. *J Ethnobiol Ethnomed.* 2013; 9:1. <https://doi.org/10.1186/1746-4269-9-1>
35. Phondani PC, Maikhuri RK, Rawat LS, Farooquee NA, Kala CP, Visvakarma SCR, et al. Ethnobotanical Uses of Plants among the Bhotiya Tribal Communities of Niti Valley in Central Himalaya, India. *Ethnobot Res Appl.* 2010;8:233-44.
36. Singh A, Nautiyal MC, Kunwar RM, Bussmann RW. Ethnomedicinal plants used by local inhabitants of Jakholi block, Rudraprayag district, western Himalaya, India. *J Ethnobiol Ethnomed.* 2017;13(1):49. <https://doi.org/10.1186/s13002-017-0178-3>
37. Pant S, Samant SS. Ethnobotanical observations in the Mornaula Reserve Forest of Kumaon, West Himalaya, India. *Ethnobot leaflets.* 2010;14:193- 217.
38. Singh KJ, Thakur AK. Medicinal Plants of the Shimla hills, Himachal Pradesh: A Survey. *Int J Herb Med.* 2014;2(2):118-127.
39. Rana MS, Samant SS. Diversity, indigenous uses and conservation status of medicinal plants in Manali wildlife sanctuary, North Western Himalaya. *Indian J Tradit Knowl.* 2011;10(3):439- 459.
40. Chauhan PP, Nigam A, Santvan VK. Ethnobotanical study of wild fruits in Pabbar Valley, District Shimla, Himachal Pradesh. *J Med Plants Stud.* 2016;4(2):216- 220.
41. Kumari P, Samant SS, Puri S. Diversity, distribution, indigenous uses and conservation of medicinal plants in central Himachal Pradesh, North Western Himalaya. *J Med Plants Stud.* 2018;6(5):45- 68.
42. Rana D, Bhatt A, Lal B. Ethnobotanical knowledge among the semi-pastoral Gujjar tribe in the high altitude (Adhwari's) of Churah subdivision, district Chamba, Western Himalaya. *J Ethnobiol Ethnomed.* 2019;15(1):10. <https://doi.org/10.1186/s13002-019-0286-3>.
43. Rana D, Masoodi HUR. Ethno-botanical survey for wild plants in fringe villages around Shimla Water Catchment Sanctuary, Himachal Pradesh, India. *J Appl Nat Sci.* 2014;6(2):720-24. <https://doi.org/10.31018/jans.v6i2.525>
44. Rao PK, Hasan SS, Bhellum BL, Manhas RK. Ethnomedicinal plants of Kathuadistrict, J&K, India. *J Ethnopharmacol.* 2015;171:12-27. <https://doi.org/10.1016/j.jep.2015.05.028>
45. Kumar R, Bhagat N. Ethnomedicinal plants of district Kathua (J & K). *Int J Med Arom Plants.* 2012;603-11.
46. Rashid A. Ethnomedicinal plants used in traditional phytotherapy of chest diseases by Gujjar-Bakerwal tribe of district Rajouri of Jammu & Kashmir state. *Int J Pharm Sci Res.* 2013;4(1):328-33.
47. Seikh MA, Chishti S, Chishti NTN. Medicinally important plants from Ganderbal, Kashmir, India- An ethnomedicinal survey. *Eur J Pharm Med Res.* 2016;3(4):176-83.
48. Ishtiyak P, Hussain SA. Traditional Use of Medicinal Plants among Tribal Communities of Bangus Valley, Kashmir Himalaya, India. *Ethno Med.* 2017;11(4):318-31. <https://doi.org/10.1080/09735070.2017.1335123>
49. Lone PA, Bhardwaj AK, Shah KW, Tabasum S. Ethnobotanical survey of threatened medicinal plants of Kashmir Himalaya, India. *J Med Plant Res.* 2014;8(47):1362-73.
50. Ahmed E, Arshad M, Ahmad M, Saeed M, Ishaque M. Ethnopharmacological Survey of Some Medicinally Important Plants of Galliyat Areas of NWFP, Pakistan. *Asian J Plant Sci.* 2004;3(4):410-15. <https://doi.org/10.3923/ajps.2004.410.415>
51. Ishtiaq M, Iqbal P, Hussain T. Ethnobotanical uses of gymnosperms of Neelum and Muzaffarabad of Kashmir. *Indian J Tradit Knowl.* 2013;12(3):404-10.
52. Ishtiaq M, Mumtaz AS, Hussain T, Ghani A. Medicinal plant diversity in the flora of Leepa Valley, Muzaffarabad (AJK), Pakistan. *Afr J Biotechnol.* 2012;11(13):3087-98 <http://dx.doi.org/10.5897/AJB11.2711>
53. Ahmad KS, Hamid A, Nawaz F, Hameed M, Ahmad F, Deng J, et al. Ethnopharmacological studies of indigenous plants in Kel village, Neelum Valley, Azad Kashmir, Pakistan. *J Ethnobiol Ethnomed.* 2017;13(1):68. <https://doi.org/10.1186/s13002-017-0196-1>
54. Ummara U, Bokhari TZ, Altaf A, Younis U, Dasti AA. Pharmacological Study of Shogran Valley Flora, Pakistan. *International Journal of Scientific & Engineering Research.* 2013;4(9):1419-27.
55. Gajurel JP, Srestha KK, Wreth S, Scheidegger C. *Taxus wallichiana* (Himalayan Yew) for the livelihood of local

- people in some protected areas of Nepal. *J Nat Hist Mus.* 2014;28:1-8. <https://doi.org/10.3126/jnhm.v28i0.14162>
56. Rehman HU, Arfan M, Rahman AU, Choudhary MI, Khan AM. Chemical constituents of *Taxus wallichiana* Zucc. *Jour Chem Soc Pak.*2003; 25(4):337-40.
 57. Sowndhararajan K, Kim S. Influence of Fragrances on Human Psychophysiological Activity: With Special Reference to Human Electroencephalographic Response. *Sci Pharm.* 2016;84(4):724-51. <https://doi.org/10.3390/scipharm84040724>
 58. Hao J, Guo H, Shi X, Wang Y, Wan Q, Song YB, et al. Comparative proteomic analyses of two *Taxus* species (*Taxus × media* and *Taxus mairei*) reveals variations in the metabolisms associated with paclitaxel and other metabolites. *Plant Cell Physiol.*2017;58(11):1878-90. <https://doi.org/10.1093/pcp/pcx128>
 59. Barrales-Cureño HJ, Farrera RA, Reyes RC, Hernández FIY, García AE, Chávez SS. Taxol generalities: a systematic review. *Rev Med UV.*2016; 16(1):75-91.
 60. Shigemori H, Kobayashi J. Biological Activity and Chemistry of Taxoids from the Japanese Yew, *Taxus cuspidata*. *J Nat Prod.*2004;67:245-56. <https://doi.org/10.1021/np030346y>
 61. Baloglu E, Kingston DGI. The Taxane Diterpenoids. *J Nat Prod.*1999;62:1448-72. <https://doi.org/10.1021/np990176j>
 62. Howat S, Park B, Oh IS, Jin YW, Lee EK, Loake GJ. Paclitaxel: biosynthesis, production and future prospects. *N Biotechnol.*2014;31(3):242-45. <https://doi.org/10.1016/j.nbt.2014.02.010>
 63. Croteau R, Ketchum RE, Long RM, Kaspera R, Wildung MR. Taxol biosynthesis and molecular genetics. *Phytochem Rev.* 2006;5(1):75-97. <https://doi.org/10.1007/s11101-005-3748-2>
 64. Zhou T, Luo X, Yu C, Zhang C, Zhang L, Song YB, et al. Transcriptome analyses provide insights into the expression pattern and sequence similarity of several taxol biosynthesis-related genes in three *Taxus* species. *BMC Plant Biol.*2019;19(1):33. <https://doi.org/10.1186/s12870-019-1645-x>
 65. Chattopadhyay SK, Kulshrestha M, Tripathi V, Saha GC, Sharma RP, Mehta VK. Studies on Himalayan Yew *Taxus wallichiana*: Part IV- The taxoids and phenolic constituents of the roots of *Taxus wallichiana*. *Indian J Chem.*1999;701-04.
 66. Phu DH, Trong PNH, Hien TTT, Nhan NT. Chemical constituents from the roots of *Taxus wallichiana* Zucc. *J Sci Technol.*2013;51(5B):233-37.
 67. Khan M, Verma SC, Srivastava SK, Shawl AS, Syamsundar KV, Khanjua SPS, et al. Essential oil composition of *Taxus wallichiana* Zucc. from the Northern Himalayan region of India. *Flavour Fragr J.*2006;21:772-75. <https://doi.org/10.1002/ffj.1682>
 68. Bala S, Uniyal GC, Chattopadhyay SK, Tripathi V, Sashidhara KV, Kulshrestha M, et al. Analysis of taxol and major taxoids in Himalayan yew, *Taxus wallichiana*. *J Chromatogr A.*1999;858(2):239-44. [https://doi.org/10.1016/S0021-9673\(99\)00841-9](https://doi.org/10.1016/S0021-9673(99)00841-9)
 69. Vander Velde DG, Georg GI, Gollapudi SR, Jampani HB, Liang XZ, Mitscher LA, et al. Wallifoliol, a taxol congener with a novel carbon skeleton, from Himalayan *Taxus wallichiana*. *J Nat Prod.*1994;56:861-67. <https://doi.org/10.1021/np50108a032>
 70. Prasain JK, Stefanowicz P, Kiyota T, Habeichi F, Konishi Y. Taxines from the needles of *Taxus wallichiana*. *Phytochemistry.*2001;58(8):1167-70. [https://doi.org/10.1016/S0031-9422\(01\)00305-3](https://doi.org/10.1016/S0031-9422(01)00305-3)
 71. Appendino G, Ozen HC, Gariboldi O, Torregiani E, Gabetta B, Nizzola R, et al. New oxetane type taxanes from *Taxus wallichiana* Zucc. *J Chem Soc Perkin Trans I.*1993;14:1563-66. <https://doi.org/10.1039/P19930001563>
 72. Barboni L, Gariboldi P, Torregiani E, Appendino G, Varese M, Gabetta B, et al. Minor taxoids from *Taxus wallichiana*. *J Nat Prod.* 1995;58(6):934-39. <https://doi.org/10.1021/np50120a019>
 73. Srestha TB, Khatri Chetri SK, Banskota AH, Manandhar MD, Taylor WC. 2-Deacetoxytaxinine B: A New Taxane from *Taxus wallichiana*. *J Nat Prod.*1997;60:820-21. <https://doi.org/10.1021/np9606153>
 74. Madhusudan KP, Chattopadhyay SK, Tripathi VK, Sashidhara KV, Kukreja AK, Jain SP. LC-ESI-MS analysis of taxoids from the bark of *Taxus wallichiana*. *Biomed Chromatogr.* 2002;16:343-55. <https://doi.org/10.1002/bmc.163>
 75. Chattopadhyay SK, Kulshrestha M, Saha GC, Sharma RP, Jain SP, Kumar S. The Taxoid constituents of the heartwood of *Taxus wallichiana*. *Planta Med.*1996;62:482. <https://doi.org/10.1055/s-2006-957949>
 76. Joshi BS, Roy R, Chattopadhyay SK, Madhusudanan KP. An NMR and LC-MS based approach for Mixture Analysis involving Taxoid molecules from *Taxus wallichiana*. *J Mol Struct.* 2003;235-48. [https://doi.org/10.1016/S0022-2860\(02\)00576-8](https://doi.org/10.1016/S0022-2860(02)00576-8)
 77. Chattopadhyay SK, Tripathi V, Sashidhara KV, Mehta VK. Studies on Himalayan Yew *Taxus wallichiana*: Part IX- The chemical constituents of seeds of *Taxus wallichiana*. *Indian J Biochem.*2002;41B:225-27.
 78. Bush A. Pathophysiological Mechanisms of Asthma. *Front Pediatr.* 2019;7:68. <https://doi.org/10.3389/fped.2019.00068>
 79. Balkrishna A, Sakat SS, Joshi K, Paudel S, Joshi D, Joshi K, et al. Anti-Inflammatory and Anti-Arthritic Efficacies of an Indian Traditional Herbo-Mineral Medicine "Divya Amvadari Ras" in Collagen Antibody-Induced Arthritis (CAIA) Mouse Model Through Modulation of IL-6/IL-1 β /TNF- α /NF κ B Signaling. *Front Pharmacol.* 2019; 10:659. <https://doi.org/10.3389/fphar.2019.00659>
 80. Bäck M, Yurdagul A Jr, Tabas I, Öörni K, Kovanen PT. Inflammation and its resolution in atherosclerosis: mediators and therapeutic opportunities. *Nat Rev Cardiol.* 2019;16(7):389-406. <https://doi.org/10.1038/s41569-019-0169-2>
 81. Lichota A, Gwozdziński L, Gwozdziński K. Therapeutic potential of natural compounds in inflammation and chronic venous insufficiency. *Eur J Med Chem.* 2019;176:68-91. <https://doi.org/10.1016/j.ejmech.2019.04.075>
 82. Tsalamandris S, Antonopoulos AS, Oikonomou E, Papamikroulis GA, Vogiatzi G, Papaioannou S, et al. The Role of Inflammation in Diabetes: Current Concepts and Future Perspectives. *Eur Cardiol.*2019;14(1):50-59. <https://doi.org/10.15420/ecr.2018.33.1>.
 83. Abufaraj M, Tabung FK, Shariat SF, Moschini M, Devore E, Papantoniou K, et al. Association between Inflammatory Potential of Diet and Bladder Cancer Risk: Results of 3 United States Prospective Cohort Studies. *J Urol.* 2019;202(3):484-89. <https://doi.org/10.1097/JU.0000000000000279>
 84. Oguntibeju OO. Medicinal plants with anti-inflammatory activities from selected countries and regions of Africa. *J Inflamm Res.* 2018;11:307-17. <https://doi.org/10.2147/JIR.S167789>
 85. Khan I, Nisar M, Shah MR, Shah H, Gilani SN, Gul F, et al. Anti-inflammatory activities of *Taxus abietane* A isolated from *Taxus wallichiana* Zucc. *Fitoterapia.*2011;82(7):1003-07. <https://doi.org/10.1016/j.fitote.2011.06.003>

86. Qayum M, Nisar M, Shah MR, Adhikari A, Kaleem WA, Khan I, et al. Analgesic and antiinflammatory activities of toxoids from *Taxus wallichiana* Zucc. *Phytother Res.*2012;26(4):552-56. <https://doi.org/10.1002/ptr.3574>
87. Khan I, Nisar M, Zarrelli A, Fabio GD, Gul F, Gilani SN, et al. Molecular insights to explore abietane diterpenes as new LOX inhibitors. *Med Chem Res.*2013;22(12):5809-13. <https://doi.org/10.1007/s00044-013-0559-7>
88. Nisar M, Khan I, Simjee SU, Gilani AH, Obaidullah, Perveen H. Anticonvulsant, analgesic and antipyretic activities of *Taxus wallichiana* Zucc. *J Ethnopharmacol.*2008;116(3):490-94. <https://doi.org/10.1016/j.jep.2007.12.021>
89. Bhat MA, Ganie SA, Dar KB, Ali R, Hamid R. Invitro antioxidant potential and hepatoprotective activity of *Taxus wallichiana*. *Asian J Pharm Clin Res.*2018;11(8):237-43. <https://doi.org/10.22159/ajpcr.2018.v11i8.22345>
90. Chattopadhyay SK, Pal A, Maulik PR, Kaur T, Garg A, Khanuja SP. Taxoid from the needles of the Himalayan yew *Taxus wallichiana* with cytotoxic and immunomodulatory activities. *Bioorg Med Chem Lett.*2006;16(9):2446-49. <https://doi.org/10.1016/j.bmcl.2006.01.077>
91. Chattopadhyay SK, Kumar TR, Maulik PR, Srivastava S, Garg A, Sharon A, et al. Absolute configuration and anticancer activity of taxiresinol and related lignans of *Taxus wallichiana*. *Bioorg Med Chem.*2003;11(23):4945-48. <https://doi.org/10.1016/j.bmc.2003.09.010>
92. Khan I, Nisar M, Ahmad M, Shah H, Iqbal Z, Saeed M, et al. Molecular simulations of Taxawallin I inside classical taxol binding site of β -tubulin. *Fitoterapia.*2011;82(2):276-81. <https://doi.org/10.1016/j.fitote.2010.10.011>
93. Wang Y, Wang J, Wang H, Ye W. Novel taxane derivatives from *Taxus wallichiana* with high anticancer potency on tumor cells. *Chem Biol Drug Des.*2016;88(4):556-61. <https://doi.org/10.1111/cbdd.12782>
94. Zwawiak J, Zaprutko L. A brief history of taxol. *J Med Sci.*2014;1(83):47-52.
95. Weaver BA. How Taxol/paclitaxel kills cancer cells. *Mol Biol Cell.* 2014;25(18):2677-81. <https://doi.org/10.1091/mbc.e14-04-0916>
96. Kellogg EH, Hejab NMA, Howes S, Northcote P, Miller JH, Fernando Diaz J, et al. Insights into the distinct mechanisms of action of taxane and non-taxane microtubule stabilizers from cryo-EM structures. *J Mol Biol.*2017;429(5):633-46. <https://doi.org/10.1016/j.jmb.2017.01.001>
97. Mukhtar E, Adhami VM, Mukhtar H. Targeting microtubules by natural agents for cancer therapy. *Mol Cancer Ther.*2014;13(2):275-84. <https://doi.org/10.1158/1535-7163.MCT-13-0791>
98. Yanamadala G, Praveen srikumar P, Rushyendra GV, Gupta VRM, Srinivasarao S. Development and validation of a novel RP-HPLC method for the determination of cabazitaxel in bulk and formulations. *Indo Am J Pharm Res.* 2013; 3(10):8266-72.
99. Bidkar AP, Sanpui P, Ghosh SS. Efficient induction of apoptosis in cancer cells by paclitaxel-loaded selenium nanoparticles. *Nanomedicine (Lond).* 2017;12(21):2641-51. <https://doi.org/10.2217/nnm-2017-0189>
100. Brewer JR, Morrison G, Dolan ME, Fleming GF. Chemotherapy-induced peripheral neuropathy: Current status and progress. *Gynecol Oncol.* 2016;140(1):176-83. <https://doi.org/10.1016/j.ygyno.2015.11.011>
101. Sun J, Jiang L, Lin Y, Gerhard EM, Jiang X, Li L, et al. Enhanced anticancer efficacy of paclitaxel through multistage tumor-targeting liposomes modified with RGD and KLA peptides. *Int J Nanomedicine.* 2017;12:1517-37. <https://doi.org/10.2147/IJN.S122859>
102. McIlwain DR, Berger T, Mak TW. Caspase functions in cell death and disease. *Cold Spring Harb Perspect Biol.* 2015; 7(4):a026716. Erratum In: *Cold Spring Harb Perspect Biol.* 2015;5(4). pii: a008656. <https://doi.org/10.1101/cshperspect.a026716>
103. Jelínek M, Balušíková K, Schmiedlová M, Němcová-Fürstová V, Šrámek J, Stančíková J, et al. The role of individual caspases in cell death induction by taxanes in breast cancer cells. *Cancer Cell Int.*2015;15(1):8. <https://doi.org/10.1186/s12935-015-0155-7>
104. Yao Y, Marassi FM. Reconstitution and Characterization of BCL-2 Family Proteins in Lipid Bilayer Nanodiscs. *Methods Mol Biol.* 2019;1877:233-46. https://doi.org/10.1007/978-1-4939-8861-7_16
105. Tessoulin B, Papin A, Gomez-Bougie P, Bellanger C, Amiot M, Pellat-Deceunynck C, et al. BCL2-Family Dysregulation in B-Cell Malignancies: From Gene Expression Regulation to a Targeted Therapy Biomarker. *Front Oncol.*2019;8: 645. <https://doi.org/10.3389/fonc.2018.00645>
106. Correia C, Lee SH, Meng XW, Vincelette ND, Knorr KL, Ding H, et al. Emerging understanding of Bcl-2 biology: Implications for neoplastic progression and treatment. *Biochim Biophys Acta.*2015;1853(7):1658-71. <https://doi.org/10.1016/j.bbamcr.2015.03.012>
107. Hientz K, Mohr A, Bhakta-Guha D, Efferth T. The role of p53 in cancer drug resistance and targeted chemotherapy. *Oncotarget.* 2016;8(5):8921-46. <https://doi.org/10.18632/oncotarget.13475>
108. Simabuco FM, Morale MG, Pavan ICB, Morelli AP, Silva FR, Tamura RE. p53 and metabolism: from mechanism to therapeutics. *Oncotarget.* 2018;9(34):23780-823. <https://doi.org/10.18632/oncotarget.25267>
109. Georgakilas AG, Martin OA, Bonner WM. p21:A Two-Faced Genome Guardian. *Trends Mol Med.*2017;23(4):310-19. <https://doi.org/10.1016/j.molmed.2017.02.001>
110. Lv C, Qu H, Zhu W, Xu K, Xu A, Jia B, et al. Low-Dose Paclitaxel Inhibits Tumor Cell Growth by Regulating Glutaminolysis in Colorectal Carcinoma Cells. *Front Pharmacol.* 2017;8:244. <https://doi.org/10.3389/fphar.2017.00244>
111. Wehner F, Gawatz O. [Suicidal yew poisoning—from Caesar to today—or suicide instructions on the internet]. *Arch Kriminol.*2003;211(1-2):19-26.
112. Kobusiak-Prokopowicz M, Marciniak A, Ślusarczyk S, Ściborski K, Stachurska A, Mysiak A, et al. A suicide attempt by intoxication with *Taxus baccata* leaves and ultra-fast liquid chromatography-electrospray ionization-tandem mass spectrometry, analysis of patient serum and different plant samples: case report. *BMC Pharmacol Toxicol.* 2016;17(1):41. <https://doi.org/10.1186/s40360-016-0078-5>
113. Piskač O, Stříbrný J, Rakovcová H, Malý M. Cardiotoxicity of yew. *Cor et Vasa.* 2015;57(3):e234-e238. <https://doi.org/10.1016/j.crvasa.2014.11.003>
114. Perju-Dumbravă D, Morar S, Chiroban O, Lechintan E, Cioca A. Suicidal poisoning by ingestion of *Taxus Baccata* leaves. Case report and literature review. *Rom J Leg Med.* 2013;21:115-18. <https://doi.org/10.4323/rjlm.2013.115>
115. Willaert W, Claessens P, Vankelecom B, Vanderheyden M. Intoxication with *Taxus baccata*: cardiac arrhythmias following yew leaves ingestion. *Pacing Clin. Electrophysiol.* 2002;25(4 Pt 1):511-12. <https://doi.org/10.1046/j.1460-9592.2002.00511.x>
116. Poudel RC, Möller M, Gao LM, Ahrends A, Baral SR, Liu J, et al. Using morphological, molecular and climatic data to delimitate yews along the Hindu Kush-Himalaya and adjacent regions. *PLoS One.* 2012;7(10):e46873. <https://doi.org/10.1371/journal.pone.0046873>

117. Eibl R, Meier P, Stutz I, Schildberger D, Hühn T, Eibl D. Plant cell culture technology in the cosmetics and food industries: current state and future trends. *Appl Microbiol Biotechnol.* 2018;102(20):8661-75. <https://doi.org/10.1007/s00253-018-9279-8>
118. Dutta MM, Jha S. Embryo Culture of *Taxus wallichiana* Zucc. *J Plant Biotechnol.*2004;6(4):213-19.
119. Hussain A, Qarshi IA, Nazir H, Ullah I, Rashid M, Shinwari ZK. In-vitro callogenesis and organogenesis of *Taxus wallichiana*. *Pak J Bot.*2013;45(5):1755-59.
120. Datta MM, Majumder A, Jha S. Organogenesis and plant regeneration in *Taxus wallichiana* (Zucc.). *Plant Cell Rep.* 2006;25(1):11-18. <https://doi.org/10.1007/s00299-005-0027-z>
121. Aslam M, Raina PA, Rafiq RU, Siddiqi TO, Reshi ZA. Adventitious root formation in branch cuttings of *Taxus wallichiana* Zucc.(Himalayan yew): A clonal approach to conserve the scarce resource. *Curr Bot.*2017;8:127-35. <https://doi.org/10.19071/cb.2017.v8.3231>
122. Nazir N, Kamili AN, Shah D, Zargar MY. Adventitious Rooting in Shoot Cuttings of *Taxus wallichiana* Zucc., an Endangered Medicinally Important Conifer of Kashmir Himalaya. *Forest Res.* 2018;7:2. <https://doi.org/10.4172/2168-9776.1000221>
123. Veeresham C, Mamatha CH, Babu P, Srisilam K, Kokate CK. Production of Taxol and its Analogues from Cell Cultures of *Taxus wallichiana*. *Pharm Biol.*2003;41(6):426-30. <https://doi.org/10.1076/phbi.41.6.426.17822>
124. Das K, Dang R, Ghanshala N, Rajashekharan PE. In vitro establishment and maintenance of callus of *Taxus wallichiana* Zucc. for production of secondary metabolites. *Nat Prod Rad.*2008;7(2):150-53.
125. Navia-Osorio A, Garden H, Cusidó RM, Palazón J, Alfermann AW, Piñol MT. Production of paclitaxel and baccatin III in a 20-L airlift bioreactor by a cell suspension of *Taxus wallichiana*. *Planta Med.* 2002;68(4):336-40. <https://doi.org/10.1055/s-2002-26739>
126. Barbuti AM, Chen ZS. Paclitaxel Through the Ages of Anticancer Therapy: Exploring Its Role in Chemoresistance and Radiation Therapy. *Cancers (Basel).* 2015;7(4):2360-71. <https://doi.org/10.3390/cancers7040897>
127. Zhang D, Yang R, Wang S, Dong Z. Paclitaxel: new uses for an old drug. *Drug Des Devel Ther.*2014;8:279-84. <https://doi.org/10.2147/DDDT.S56801>
128. Kajani AA, Zarkesh-Esfahani SH, Bordbar AK, Khosropour AR, Razmjou A, Kardi M. Anticancer effects of silver nanoparticles encapsulated by *Taxus baccata* extracts. *J Mol Liq.* 2016;223:549-56. <https://doi.org/10.1016/j.molliq.2016.08.064>
129. Dutta S, Mariappan G, Sarkar D, Sarkar P. Assessment of Anti-inflammatory Activity of *Taxus Baccata* Linn. Bark Extract. *Anc Sci Life.*2010;29(3):19-21.
130. Patel P, Patel K, Gandhi T. Evaluation of Effect of *Taxus baccata* Leaves Extract on Bronchoconstriction and Bronchial Hyperreactivity in Experimental Animals. *J Young Pharm.* 2011;3(1):41-47. <https://doi.org/10.4103/0975-1483.76418>
131. Shang W, Qiao J, Gu C, Yin W, Du J, Wang W, et al. Anticancer activity of an extract from needles and twigs of *Taxus cuspidata* and its synergistic effect as a cocktail with 5-fluorouracil. *BMC Complement Altern Med.* 2011;11:123. <https://doi.org/10.1186/1472-6882-11-123>
132. Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Front Pharmacol.*2014;4:177. <https://doi.org/10.3389/fphar.2013.00177>
133. Datta T, Patra AK, Dastidar SG. Medicinal plants used by tribal population of Coochbehar district, West Bengal, India-an ethnobotanical survey. *Asian Pac J Trop Biomed.*2014;4(Suppl 1):S478-S482. <https://doi.org/10.12980/APJTB.4.2014C1122>
134. Xie DF, Li MJ, Tan JB, Price M, Xiao QY, Zhou SD, et al. Phylogeography and genetic effects of habitat fragmentation on endemic *Urophysa* (Ranunculaceae) in Yungui Plateau and adjacent regions. *PLoS One.* 2017;12(10):e0186378. <https://doi.org/10.1371/journal.pone.0186378>
135. Keller LF, Waller DM. Inbreeding effects in wild populations. *Trends in Ecology & Evolution.* 2002;17(5):230-41.

