View metadata, citation and similar papers at core.ac.uk
brought to you by CORE
provided by Ulster University's Research Portal

Connectivity Mapping (ssCMap) To Predict A20 Inducing Drugs Anti-inflammatory Action In Cystic Fibrosis

B Malcomson[1]*, H Wilson[1]*, E Veglia[2], G Thillaiyampalan[3], R Barsden[1], S Donegan[1], A El Banna[1], JS Elborn[1], M Ennis[1], C Kelly[4], SD Zhang[3,4] and BC Schock[1$].

[1]Centre for Infection and Immunity, Queen's University of Belfast, Belfast, BT9 7AE, UK,
[2]Dipartimento di Scienza e Tecnologia del Farmaco, Università di Torino, Via P. Giuria 9, 10125, Turin, Italy,
[3]Centre Cancer Research and Cell Biology, Queen's University of Belfast, Belfast, BT9 7AE, UK,
[4]Northern Ireland Centre for Stratified Medicine, University of Ulster, Altnagelvin Hospital Campus, Derry/Londonderry BT47 6SB, UK.

*Both junior authors contributed equally to this work

[$]Corresponding author contact details:

Address: Centre for Infection and Immunity, Queen's University of Belfast, Medical Biology Building, 97 Lisburn Road, Belfast, UK, BT9 7BL

Phone: +44 (0) 28 9097 2258

Fax: +44 (0) 28 90 972671

Email: b.schock@qub.ac.uk

Key Words: A20 protein; NF-kappaB; Connectivity Mapping, Drug repositioning, Airway epithelial cells; Inflammation.

Running title: Connectivity mapping to predict A20 inducing drugs

Significance Statement

This study reports that publically available gene array expression data together with statistically significant connections' map (sscMap) can successfully predict already licensed drugs to modify genes of interest. We applied this bioinformatics approach to the NF-κB regulator A20 (*TNFAIP3*), which is reduced in Cystic Fibrosis (CF) airway cells. sscMap predicted drugs that should or are predicted to induce A20 and normalise the inflammatory response in CF airways. Using airway epithelial cells we show that ikarugamycin and quercetin have anti-inflammatory effects mediated by induction of A20 in CF and non-CF airway epithelial cells. Using siRNA techniques we confirm that the anti-inflammatory effect of ikarugamycin and quercetin are mainly due to A20 induction as the anti-inflammatory effect is lacking in bronchial epithelial cells with A20 knockdown.

The effect of fluvastatin, which was predicted not to modify A20, was also confirmed. We have identified a process whereby already licensed drugs can be successfully repositioned for chronic inflammatory airway diseases.

Abstract

Cystic Fibrosis (CF) lung disease is characterised by a chronic and exaggerated inflammation in the airways. Despite recent developments to therapeutically overcome the underlying functional defect in CFTR (cystic fibrosis transmembrane conductance regulator), there is still an unmet need to also normalise the inflammatory response. The prolonged and heightened inflammatory response in CF is in part mediated by a lack of intrinsic downregulation of the pro-inflammatory NF-kB pathway. We have previously identified reduced expression of the NF-κB down-regulator A20 in CF as a key target to normalise the inflammatory response. Here we have used publically available gene array expression data together with sscMap (statistically significant connections' map) to successfully predict drugs already licensed for the use in humans to induce A20 mRNA and protein expression and thereby reduce inflammation. The effect of the predicted drugs on A20 and NF-kB(p65) expression (mRNA) as well as pro-inflammatory cytokine release (IL-8) in the presence and absence of bacterial LPS was shown in bronchial epithelial cells lines (16HBE14o-, CFBE41o-) and in primary nasal epithelial cells (PNECs) from patients with CF (Phe508del homozygous) and non-CF controls. Additionally, the specificity of the drug action on A20 was confirmed using cell lines with *TNFAIP3* (A20) knockdown (siRNA). We also show that the A20 inducing effect of ikarugamycin and quercetin is lower in CF derived airway epithelial cells than in non-CF cells.

\body

Introduction

The response to pathogens, recognised by pattern recognition receptors including Toll-like receptors (TLRs), triggers an acute innate immune response that is mediated by transcription factors such as nuclear factor-kappa-light-chain enhancer of B cells (NF-κB). NF-κB activation promotes the transcription of inflammatory mediators in a tightly regulated process. However, in individuals with underlying chronic inflammatory diseases, this regulation is compromised, leading to constitutive NF-κB activation and persistent inflammation (1-3)

The development of new first-in-class medicines is costly (approximately $1.2 billion for a single FDA-approved drug) and takes between 10 and 15 years (4, 5). Many newly developed drugs perform well in the preclinical testing, but fail when tested in humans (6). Thus alternative approaches using predictive models to identify new drugs are needed. Gene expression connectivity mapping (www.broadinstitute.org/cmap/) is an advanced bioinformatics technique to establish the connections among biological states via gene expression profiles/signatures. One major application of connectivity mapping is to identify potential small molecules able to inhibit a disease state or regulate the expression of a small number of genes (7-9). We used an advanced version of connectivity mapping, sscMap (statistically significant connections' map) (10), which has been successfully applied to phenotypic targeting and predicting effective drugs in cancer (10). However, this has not yet been applied to chronic inflammatory diseases.

Cystic Fibrosis (CF) is a chronic multi-organ inflammatory disease, caused by mutations in the CFTR gene (Cystic Fibrosis Transmembrane Conductance regulator) expressed on apical epithelial surfaces. It is the most common lethal genetic disease in Caucasian populations. Lung disease is the primary cause of morbidity and mortality in CF, resulting from dehydration of epithelial surfaces and reduced mucociliary clearance as a consequence of the ionic imbalance created by CFTR mutation. This leads to a cycle of infection and inflammation associated with a

progressive reduction in lung function and eventual respiratory failure. A common feature of CF is the heightened, chronic inflammatory response to *Pseudomonas aeruginosa* (*P. aeruginosa*), driven by constitutive NF-κB activation in airway and peripheral blood cells (2, 3, 11). Primary nasal epithelial cells (PNECs) from patients with the common F508del/F508del mutation and a milder genotype (R117H/F508del), show a significant increase in NF-κB(p65) which correlates with disease severity (12).

A20 (TNFAIP3) is a central negative regulator of NF-κB activation following stimulation of TLRs and/or TNF-receptor and regulates different signalling pathways such as NF-kB and interferon regulatory factor (IRF) signalling (13). A20 modifies classical immune cells (14, 15) as well as epithelial cells (12), endothelial cells (16), embryonic fibroblasts (17), osteoclasts (18) and pancreatic beta-cells (19) and diverse roles for A20 in innate immunity, apoptosis, autophagy and antigen processing (13, 15, 16, 20). Within the innate inflammatory immune response A20 regulates NF-κB signalling at the level of TRAF6 in mouse embryonic fibroblasts (MEFs) and osteoclasts (17, 18). In cultured human airway epithelial cells, A20 is rapidly induced by viral or bacterial compounds (21) and is essential for termination of the TLR4 signal (22). PNECs stimulated with *P. aeruginosa* LPS show a transient increase in A20, but CF PNECs display lower A20 expression basally and after LPS stimulation (12, 23).

Therefore, A20 induction should have anti-inflammatory effects within the tightly regulated NF-kB signalling pathway as shown by the induction of A20 through gibberellin (GA_3) in airway epithelial cells. GA_3 induced A20, reduced IL-8 secretion, stabilised cytosolic IkBα and reduced NF-κB (p65) activation (24). Here we set out to identify additional compounds able to induce A20. Thus, we performed a compound search using gene expression connectivity mapping to identify existing drugs that could induce A20 expression.

Results

1. ***Connectivity mapping (sscMap):***

The selection of gene array data and creation of the gene signature

Data sets that passed the selection criteria contained human primary nasal epithelial cells (PNECs) and the human bronchial epithelial cell lines CFBE41o-, Calu-3 and IB3-1 analysed basally and after exposure to *P. aeruginosa* LPS (***Table 1***). In total 76 samples from 4 different published gene array data sets were used. Linear expression correlation and GO enrichment analysis for NF-κB pathway genes identified the closest correlates to A20. ***Table 2*** shows the top 7 genes that subsequently served as the input to the connectivity mapping process.

Prediction of drugs to induce A20 in airway epithelial cells

This study sought small molecular compounds that may enhance A20 expression and as a confirming negative control, those compounds that may inhibit A20 expression. ***Table 3*** summarizes the top candidate drugs identified. The column entitled 'significance' shows the significance of drugs based on p values and the column 'z-score' shows the correlation of the drugs with the input gene signature. Positive z-scores indicate a positive correlation i.e. the input genes are induced when treated with the particular drug. The significant drugs with the highest positive z-scores along with a negative control were selected for laboratory validation. In addition to p values and z scores, stability of the connections was measured by altering the gene signature and the significance of the connections are given under the column 'perturbation stability'. Drugs with perturbation stability 1 represent strong connections which remain significant with 'perturbation' gene signatures. From these predictions two A20 inducing drugs (ikarugamycin and

quercetin) as well as one non-A20 inducing drug (fluvastatin) were chosen for further investigation.

2. Gene expression of gene signature

Expression of the genes identified as the A20/NF-kB gene signature in CF epithelial cells were analysed by qRT-PCR in 16HBE14o- and CFBE41o- cultured in the presence or absence of LPS for 0-24h (***Figure 1***).

Basal expression: CFBE41o- show significantly lower mRNA expression for A20, ATF3, Rab5c and ICAM1 compared to 16HBE14o- (all $p<0.05$, $n=5$). Expression of DENNDA4 and PSNE1 was also lower in CFBE41o- but this did not reach significance.

LPS induced expression: In 16HBE14o-, A20 mRNA is rapidly induced with expression peaking 1h after LPS exposure ($p<0.001$ compared to medium, $n=5$), while CFBE41o- show significantly lower (at 1h $p<0.001$ vs. 16HBE14o-, $n=5$) and delayed (maximal induction at 4-8 h, $p<0.01$ and 0.001 vs. medium, $n=5$) induction upon LPS stimulation. After LPS, ATF3 and ICAM1 expression was significantly lower in CFBE41o- compared to 16HBE14o- ($p<0.01$, $n=5$). Pom121 and PSNE1 expression increased in CFBE41o- compared to medium (8h, $p<0.05$ and $p<0.01$) and in CFBE41o- compared to 16HBE14o- (8h, $p<0.05$ and $p<0.001$). 16HBE14o- showed a significant reduction in DENNDA4 and Rab5c expression compared to medium (1h, $p<0.05$ and 4h, $p<0.05$, respectively), but there was no significant change in the expression of these genes in CFBE41o-.

3. Effect of A20 inducing drugs on cell lines

LDH release in drug exposed 16HBE14o- and CFBE41o-: LDH release was measured after exposure to the drugs alone (0.01–1000 µM) and with LPS stimulation (***supplement Figure 1S***). Quercetin did not cause any LDH release. Overall fluvastatin was almost without effect on LDH;

the exceptions were a slight but statistically significant increase at 10 µM alone (CFBE41o-) and in the presence of LPS (in CFBE41o- and 16HBE14o-). Ikarugamycin (1, 100 µM) caused a significantly higher LDH release in both cell types. In LPS stimulated cells, 0.1 µM ikarugamycin and higher concentrations showed a higher LDH release compared to LPS alone, but this did not reach statistical significance (***supplement Figure 1S***).

LPS stimulated IL-8 release in drug pre-treated 16HBE14o- and CFBE41o-: To assess the anti-inflammatory potential of the selected drugs, cells were pre-treated with the drug for 1h, stimulated with LPS and the IL-8 release measured and the relative IC_{50} calculated (***supplement S3***). In 16HBE14o-, all drugs reduced IL-8 release by at least 50% with an IC_{50} of 15.6 µM for ikarugamycin, 0.09 µM for quercetin and 0.11 µM for fluvastatin. In CFBE41o-, only quercitin (IC_{50} 0.03 µM) and fluvastatin (IC_{50} 0.001 µM) pre-treatment were able to reduce release by 50%. In contrast, pre-treatment of CFBE41o- with 1 µM ikarugamycin caused a significant increase in IL-8 release compared to LPS alone (LPS 269.9 ± 47.9 pg/ml vs. 590.7 ± 82.6 pg/ml, $p<0.05$, n=5). Therefore, a meaningful calculation of the relative IC_{50} for IL-8 release in ikarugamycin treated CFBE41o- cells was not possible.

A20 mRNA induction in drug treated 16HBE14o- and CFBE41o-: To elucidate if ikarugamycin and quercetin facilitate their anti-inflammatory action though the induction of A20 as predicted, A20 mRNA was determined by qRT-PCR. Fluvastatin was included as a negative control. Using the LDH and IL-8 release data two drug concentrations were selected for further investigations (***supplement S3***). In 16HBE14o-, LPS stimulation caused a significant induction of A20 1h after stimulation (***Figure 2***). Ikarugamycin (0.01 µM) alone did not cause a significant induction of A20, but additional LPS stimulation caused a significant A20 induction at 24h ($p<0.05$ vs LPS 24h). 1 µM ikarugamycin significantly induced A20 at 4h on its own, but the higher A20

expression in the presence of LPS (1-24h) did not reach statistical significance. Quercetin pre-treatment did not induce significant levels of A20 mRNA at 0.1 μM, alone or in the presence of LPS. However, at 100 μM quercetin A20 mRNA was significantly induced alone at 1h and 4h ($p<0.05$ vs. medium 1h and 4h) and in the presence of LPS at 4h ($p<0.05$ vs. LPS 4h). Fluvastatin alone did not induce A20 mRNA at any time or concentration (***Figure 2***), in the presence of LPS fluvastatin pre-treatment caused a significant reduction in A20 mRNA at both concentrations tested ($p<0.001$ for 0.1 μM+LPS 1h vs. LPS 1h; $p<0.05$ for 1 μM+LPS 1h vs. LPS 1h). In CFBE41o-, LPS significantly induced A20 at 4h ($p<0.05$), but induction levels were lower than in 16HBE14o- (***Figure 2***). Ikarugamycin (1 μM) induced significant levels of A20 at 4h and 24h (both $p<0.05$), this was further increased in the presence of LPS at 4h ($p<0.05$ vs LPS 4h) and 24h ($p<0.01$ vs LPS 24h). Quercetin treatment alone significantly induced A20 at 4h at both concentrations ($p<0.05$ 0.1 μM vs. medium 4h and $p<0.01$ 100 μM vs. medium 4h). In the presence of LPS, only 100 μM quercetin caused a significant induction of A20 at 24h ($p<0.05$). Fluvastatin alone did not induce A20 mRNA at any time or concentration (***Figure 2***), and in the presence of LPS fluvastatin caused a significant reduction in A20 mRNA at both concentrations tested ($p<0.05$ for 0.1 μM+LPS 4h vs. LPS 4h; $p<0.05$ for 1 μM+LPS 4h vs. LPS 4h).

Effect of selected components on NF-kB (p65) mRNA in 16HBE14o- and CFBE41o-: Next we investigated if A20 induction altered NF-κB(p65) mRNA levels (***Figure 2***). LPS stimulation caused a significant induction of p65 1h after stimulation in 16HBE14o- ($p<0.01$ – $p<0.001$ vs. medium). Ikarugamycin (0.01 μM) was without significant effect on p65 mRNA. At 1 μM, ikarugamycin alone induced p65 at 24h ($p<0.01$ vs. medium 24h) and in the presence of LPS at 1h and 4h (both $p<0.05$ vs. LPS). Quercetin alone showed no effect on p65 mRNA levels, but when stimulated with LPS both concentrations of quercetin (0.1, 100 μM) significantly reduced p65 mRNA levels at 1h ($p<0.01$ vs LPS 1h). Similar to quercetin, fluvastatin alone showed no effect

on p65 mRNA levels, but after LPS stimulation fluvastatin (0.1, 100 μM) significantly reduced p65 mRNA levels at 1h ($p<0.001$ and $p<0.05$ vs LPS 1h). In CFBE41o- LPS significantly induced p65 at 1h, 4h and 24h (p<0.05-0.001 vs medium) and overall CFBE41o- exhibited higher expression levels of p65 at 4h and 24h than 16HBE14o- (***Figure 2***). Ikarugamycin was without significant effect on p65 mRNA expression at any concentration or time point, although overall expression levels appear higher at 1 μM, when stimulated with LPS (***Figure 2***). In CFBE41o-, quercetin (0.1 μM) did not affect p65 mRNA levels. At 100 μM, quercetin significantly induced p65 ($p<0.05$: quercetin alone 100 μM 24h vs. medium 24h and quercetin 100 μM + LPS 24h vs. LPS 24h). Fluvastatin caused a significant reduction in p65 mRNA at both concentrations and all time points after LPS stimulation ($p<0.05$ and $p<0.01$ for 0.1 μM+LPS vs. LPS; $p<0.05$ and $p<0.01$ for 1 μM+LPS vs. LPS) (***Figure 2***).

Effect of predicted drugs on A20 and p65 protein expression: We then determined the effect of the drugs on cytosolic A20 and p65 protein by Western Blotting using the same selected concentrations than before. Ikarugamycin (0.01 μM) induced A20 protein in both 16HBE14o- and CFBE41o-, with less A20 protein induction at 1 μM. Ikarugamycin also induced cytosolic p65 in both cell types (***Figure 3a***). Quercetin treatment caused a strong induction of A20 protein at both concentrations (0. 1, 100 μM) in 16HBE14o- and to a lower degree in CFBE41o-. Quercetin (100 μM) reduced cytosolic p65 in 16HBE14o- and in CFBE41o- (***Figure 3b***). Fluvastatin did not induce A20 protein at either concentration (0.1, 1 μM) in both 16HBE14o- and CFBE41o- cells. Fluvastatin pretreatment reduced cytosolic p65 protein in 16HBE14o- cells though this was only apparent at the higher concentration in CFBE41o- (***Figure 3c***).

Specificity of the drug effect on A20 mRNA expression using A20 siRNA: To confirm that the effect of the selected drugs is facilitated through A20 induction, we used siRNA to knock down

A20 expression in 16HBE14o- cells as previously described (30). Cells were pre-treated with quercetin or ikarugamycin prior to LPS and IL-8 determined. Results (***Figure 4***) showed that in 16HBE14o- LPS significantly induced IL-8 ($p<0.05$ compared to untreated control), but when A20 is knocked down IL-8 increased further (although not significantly different from LPS alone). When cells are pre-treated with quercetin (100 µM) or ikarugamycin (1 µM), the LPS induced IL-8 release was significantly reduced ($p<0.05$). However, when A20 is knocked down IL-8 levels were not different from LPS control (***Figure 4***).

4. Effect of A20 inducing drugs on PNECs

Effect on IL-8 release: LPS significantly induced IL-8 release from PNECs from non-CF and CF patients (non-CF: $p<0.01$, CF: $p<0.05$, Wilcoxon paired test, ***Figure 5***). IL-8 release from CF PNECs was significantly higher than from non-CF PNECs (600.6±62.8 pg/ml vs. 315.8±36.1 pg/ml, $p<0.01$, Mann-Whitney test). In non-CF PNECs (***Figure 5a***), pre-treatment with ikarugamycin at 0.01 µM, but not at 1 µM significantly reduced LPS induced IL-8 release ($p<0.05$). In quercetin and fluvastatin treated non-CF PNECs only the higher concentrations tested (quercetin: 100 µM and fluvastatin: 1 µM significantly reduced IL-8 release ($p<0.01$ and $p<0.05$, respectively, ***Figure 5a***). PNECs from patients with CF showed similar results with a significant IL-8 reduction at the lower concentration of ikarugamycin (0.01 µM, $p<0.01$) and the higher concentration of quercetin (100 µM, $p<0.05$). In CF PNECs fluvastatin treatment significantly reduced IL-8 release at both concentrations tested (0.1 µM, $p<0.05$; 1 µM, $p<0.01$, ***Figure 5b***).

A20 induction in PNECs: In PNECs from non-CF control subjects (***Figure 6***) LPS stimulation resulted in a rapid and significant upregulation of A20 mRNA within 1h and a peak expression at 4h ($p<0.05$, LPS vs. medium at 1h and 4h). Ikarugamycin alone at 0.01 µM increased A20

mRNA which reached significance at 24h ($p<0.01$) and in the presence of LPS this increase was significantly higher at 1h and 24h than LPS alone ($p<0.05$ and $p<0.01$ vs. LPS). Ikarugamycin (1 μM) had no effect on A20 mRNA induction, either alone or in the presence of LPS. Quercetin (0.1 μM) caused a significant induction of A20 compared to medium control at 1h, 4h and 24h (all $p<0.01$) with expression levels similar to those induced by LPS (***Figure 6***) and this was maintained the presence of LPS, with a peak A20 induction at 4h. The higher concentration of quercetin (100 μM) significantly induced A20 mRNA at 1h ($p<0.05$) and subsequent stimulation with LPS resulted in significantly increased A20 mRNA levels at 4h ($p<0.05$ vs LPS 4h). Fluvastatin treatment did not induce A20 mRNA expression, but 1μM reduced A20 mRNA at 4h LPS (***Figure 6***).

In CF PNECs LPS induced A20 mRNA expression was lower than in non-CF PNECs, but in CF PNECs LPS induced significant levels of A20 mRNA 4h after LPS ($p<0.05$ vs. medium 4h). Ikarugamycin at 0.01 μM (alone and in the presence of LPS) had no effect on A20 mRNA levels (***Figure 6***) but 1 μM ikarugamycin significantly induced A20 mRNA at 4 and 24h alone ($p<0.05$ vs medium control) and in the presence of LPS ($p<0.05$ and $p<0.001$ vs LPS). Similarly, quercetin treatment with the lower concentration (0.01 μM) alone and in the presence of LPS had no effect on A20 mRNA levels (***Figure 6***), while 100 μM quercetin significantly induced A20 mRNA alone (4h, $p<0.05$ vs medium control) and additionally above LPS induction (1h and 24h, both $p<0.05$ vs LPS). Similar to non-CF PNECs, fluvastatin treatment of CF PNECs had no significant effect on A20 mRNA expression levels.

NF-κB (p65) induction in PNECs: PNECs from non-CF control subjects respond to LPS exposure with a significant increase in NF-κB (p65) at 1h and 4h ($p<0.001$ vs medium control). Thereafter, p65 mRNA expression returns to its corresponding medium control value (***Figure 6***).

PNECs from patients with CF however show the expected high levels of p65 throughout the 24h studied ($p<0.01$ at 1h, $p<0.05$ at 4h and 24h vs medium control) (***Figure 6***).

In non-CF PNECs, ikarugamycin treatment alone did not change p65 mRNA levels (vs medium control), but in the presence of LPS p65 mRNA was significantly reduced ($p<0.05$ at 1h, $p<0.01$ at 4h vs LPS). Overall the higher concentration of ikarugamycin induced p65 mRNA levels with a significant increase at 4h ($p<0.05$ vs medium control). However, in the presence of LPS, p65 levels remained not significantly different from those after LPS exposure at 1h and 24h, but were significantly lower compared to LPS alone at 4h ($p<0.05$) (***Figure 6***). The lower concentration of 0.01 μM quercetin alone did not modify basal p65 mRNA. After subsequent LPS challenge p65 mRNA significantly decreased at 1h ($p<0.01$ vs LPS), but then increased in a similar manner to LPS alone. However, 100 μM quercetin alone significantly reduced p65 induction at 1h and 4h ($p<0.05$ and p0.01 vs. medium control). In the presence of LPS this reduction of p65 mRNA reached statistical significance at 4h and 24h (both $p<0.05$ vs LPS) (***Figure 6***). Fluvastatin (0.1 μM) induced p65 at 4h ($p<0.05$ vs. medium 4h), but p65 levels remain significantly lower when LPS is added ($p<0.05$, vs LPS at 1h and 4h). The higher concentration of fluvastatin (1 μM) did not change p65 levels alone, but after addition of LPS, p65 mRNA was initially reduced (1h $p<0.05$ vs LPS) but then induced similarly to LPS alone (***Figure 6***).

When PNECs from patients with CF were pre-treated with ikarugamycin, p65 levels dropped significantly at 0.01 μM ikarugamycin in the absence or presence of LPS ($p<0.05$-0.01). At the higher concentration of 1 μM p65 levels appeared lower, but this only reached statistical significance at 1h ($p<0.05$ vs LPS) (***Figure 6***). Pre-treatment of CF PNECs with quercetin did not affect p65 levels alone at either concentration tested, but at 0.1 μM quercetin in the presence of LPS significantly reduced p65 at 24h ($p<0.05$ vs LPS). Treatment with 100 μM quercetin significantly reduced p65 mRNA in the presence of LPS at all time points (all $p<0.05$ vs LPS) (***Figure 6***). Fluvastatin alone had no significant effect on p65 mRNA levels, but significantly

reduced LPS induced p65 at 1h, 4h and 24h ($p<0.05$ vs. medium 1h, 4h or 24h), while the higher concentration of fluvastatin (1 μM) showed no significant effect on basal or LPS induced p65 mRNA (***Figure 6***).

Discussion

Airways infection and the subsequent inflammation are deleterious for patients suffering from CF. Current drugs targeting the mutated CFTR (potentiators/correctors) improve expression and function of CFTR on epithelial surfaces and patients showed improved lung function and reduced frequency of pulmonary exacerbations, hospitalization and use of intravenous antibiotics, bu augmented CFTR function failed to reduce inflammatory markers in sputum (e.g. IL-1,-6,-8) (25) and heterogeneous responses to the treatment have been reported (26), suggesting that CFTR correction/potentiation may not directly improve the underlying compromised immune response. The negative NF-κB regulator A20 *(TNFAIP3)* is reduced in CF airway epithelial cells, basally and after LPS stimulation (23) and is associated with markers of inflammation and decreased lung function (12). A20 silencing increased TRAF6 and NF-κB activity (18), and A20 over-expression had protective effects in airway inflammation in 'asthmatic mice' (27), suggesting that A20 augmentation normalises the inflammatory response in the airways.

In order to find agents to induce A20 in CF we employed sscMAP, which has been widely used in drug development uncovering potential new indications for existing drugs as well as predicting side effects (28). Using disease specific publicly available gene array data (GEO data sets), we used connectivity mapping to firstly identify the target gene (A20) related gene signature and to secondly predict already licensed drugs to induce A20 expression. We included a total of 76 gene array data from primary nasal epithelial cells and cell lines commonly used in CF research (***Table 1***). Gene array databases were first selected in August 2013, but a

recent (Jan 2016) search revealed no further significant published gene array data on CF (primary nasal) epithelial cells.

The applied linear regression model (Pearson's correlation coefficient) is an established robust method to identify the correlates of a known gene expression estimating the strength of a linear relationship between two random normally distributed variables (29). The application of the linear regression model with GO (gene ontology) selection revealed a gene signature of 6 genes additionally to the seed gene A20 (***Table 2***) and we confirmed that the expression of these genes is similarly reduced in CF epithelial cells, basally and after LPS stimulation (***Figure 1***). The identified A20 correlates were *ATF3*, a transcriptional repressor that binds to cAMP response elements (CRE); RAB5C, a small ubiquitously expressed GTPase; *DENND4A,* which encodes the *C-Myc* Promoter Binding Protein (MBP-1); *POM121*, a nuclear transmembrane protein and essential component of the nuclear pore complex; *ICAM1,* a cell surface glycoprotein typically expressed on endothelial and immune cells, especially during inflammation and *PSEN* 1 (Presenilin 1), a catalytic component of γ-secretase and a DREAM binding protein. Further descriptions of these genes and their involvements in inflammation can be found in the online appendix. These genes, as a combined gene signature, were then input into the sscMAP process comparing the gene expression of the gene signature with the gene expression in the reference database (www.broadinstitute.org), which was obtained from systematic microarray gene expression profiling.

SscMAP predicted a short list of drugs that should modify the expression profile of the gene signature genes, including A20. Those drugs included azacyclonol, ikarugamycin, quercetin and karakoline (***Table 3***). Azacyclonol is a drug used in psychotic individuals (30). We excluded azacyclonol, as its use requires special permission through relevant government authorities. Interestingly, the anti-histamine terfenadine is metabolised to azacyclonol and terfenadine (31). Karakoline is a highly toxic plant diterpenoid (32) and the pharmacological

effects of preparations of Aconitum roots are attributed to diterpenoid alkaloids (33). The anti-inflammatory activity of gibberellin (GA_3), also a plant-derived diterpenoid, is mediated through A20 induction (24). We therefore selected ikarugamycin and quercetin for further studies.

Ikarugamycin is a macrolide antibiotic with cytostatic effects against Gram-positive bacteria. We show that ikarugamycin exhibits anti-inflammatory properties in LPS stimulated airway cells. In 16HBE14o- ikarugamycin showed a dose-dependent reduction of LPS-induced IL-8 release (***supplement S4***), through induction of A20 and reduction of p65 (***Figure 2***). 16HBE14o- and CFBE41o- did not show reduced cell viability at concentrations lower than 1 μM, higher concentrations increased LDH release suggesting a cytotoxic effect (***supplement S2***). CFBE41o- appear more sensitive to ikarugamycin treatment (***supplement S4***), which made it not possible to calculate a meaningful relative IC_{50} value (***supplement S3***), although p65 protein expression was not increased (***Figure 2***). In HL-60 cells, ikarugamycin reduced cell viability and increased DNA fragmentation starting at 0.1 μM (IC_{50} of 0.22 μM), while MCF-7 cells and peripheral blood mononuclear cells showed higher resistance. Furthermore, ikarugamycin treatment of HL-60 cells caused a significant caspase activation, increase in intracellular calcium and p38 MAP kinase activation (34). However, investigating the pro-apoptotic mechanisms in bronchial epithelial cells was beyond the scope of this study. Nonetheless, our ikarugamycin data at near cytotoxic levels add valuable information: Firstly, sscMap correctly predicted that ikarugamycin would induce A20 mRNA but sscMap does not predict the physiological effect of the gene induction. CF cells overall show a limited ability to induce A20, however, our results show that – given the right stimulus – CF cells are indeed able to induce A20 mRNA and the high induction of A20 at near cytotoxic levels may be able to counteract the pro-apoptotic stimulation of ikarugamycin.

Quercetin, a flavonoid, is known for its anti-inflammatory effects. *In vivo* studies have shown antioxidant, anti-inflammatory, anti-tumour and even anti-infectious properties of quercetin, which are promoted through its effects on signalling pathways such as NF-κB (35). In lung epithelial cells, quercetin inhibited IL-1 and TNF-α induced IκBα degradation and NF-κB activity through modification of the MAPK pathway (AP-1) (36). SscMap correctly predicted that quercetin can induce A20 mRNA, adding a new mechanism for the anti-inflammatory effects of quercetin. It also significantly reduced LPS-induced IL-8 release in both cell types with a relative IC_{50} of 0.15 and 0.04 μM in 16HBE14o- and CFBE41o-, respectively. Quercetin at concentrations up to 1000 μM did not show any cytotoxicity, although in neuronal cell cultures quercetin higher than 100 μM was cytotoxic (37). Within the *in vivo* antioxidant network, quercetin has been described to be oxidised and to yield an ortho-quinone, which, in absence of reducing glutathione, can oxidise protein thiols impairing enzyme activities (38). We have not investigated the antioxidant status of our cell culture, but we took precautions to minimise oxidation when preparing our quercetin dilutions.

To further investigate the A20-dependent mechanism of the anti-inflammatory action of quercetin and ikarugamycin, we used A20 knock down in 16HBE14o-. As previously described for the A20 inducing anti-inflammatory compound gibberellin (24), we were able to confirm that the anti-inflammatory effect of the predicted drugs was indeed mainly mediated by the induction of A20.

We also tested ***fluvastatin***, which was predicted not to affect or reduce A20 gene expression (negative z-score). Although fluvastatin exerted anti-inflammatory effects (IL-8) in both cell lines, our data show that this was not mediated by the induction of A20 (mRNA), clearly confirming the sscMap prediction. In asthma, fluvastatin inhibits eosinophil adhesion to ICAM-1 (39) and fibroblast proliferation (40). Using similar concentrations we did not

observe any reduced proliferation. Fluvastatin at a concentration range similar to those we used, reduced basal and LPS-induced IL-8 release from LPS stimulated whole blood cells, with CF cells appearing more sensitive to fluvastatin than control cells (IC_{50}: 19.1 μM in non-CF cells, 4.6 μM in CF blood cells) (41). In isolated LPS-stimulated peripheral blood monocytes from patients with chronic kidney disease fluvastatin had a significant anti-inflammatory effect (IL-8, IL-6) at a concentration range of 0.0001–1 μM (42). Patients with heart transplants receiving 40 mg fluvastatin/day for 4 weeks showed a significant reduction in total cholesterol levels and a maximum blood fluvastatin concentration of 2.11 and 3.77 μM. These studies suggest that we have covered a physiologically relevant range of fluvastatin. However, fluvastatin metabolism may be affected by concomitant therapies, especially substances competing with cytochrome enzymes and in such cases fluvastatin levels may need to be monitored (43). Any reactions with other therapies (as they would appear in patients with CF) were not investigated in our manuscript as they would have been beyond the scope of the study.

Similar to ikarigamycin, fluvastatin has been described to have pro-apoptotic effects e.g. in human lymphoma cells, human smooth muscle cells and in rat neonatal cardiac myocytes or rat vascular smooth muscle cells (44, 45), mediated through activation of caspase-3, reactive oxygen species and activation of p38 MAPK (44, 55). However, statins, through their inhibition of 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA), but not through induction of A20, may still have therapeutic potential in airway and systemic inflammation in CF (41).

Overall, our study shows that connectivity mapping (sscMap) can predict A20 inducing drugs. Pre-treatment of cells with both ikarugamycin and quercetin reduces LPS-induced IL-8 secretion by induction of A20. In non-CF PNECs both drugs upregulated A20 and reduced IL-8 and p65 mRNA at lower concentrations than in cell lines. CF PNECs,

however, have a reduced and delayed A20 inducing response to LPS and to the tested drugs and a significant A20 induction appears at the higher concentration of the drugs tested, which might be near to the cytotoxic effect. A20 reduces apoptosis (46, 47) and mutational loss of A20 resulted in rapid apoptosis and inflammation in hematopoietic cells (48). We did not determine markers of apoptosis in our study, but the huge increase in A20 mRNA may indicate a possible counteraction to pro-apoptotic changes in response to ikarugamycin treatment. Of particular interest, this may indicate a higher susceptibility/sensitivity of CF cells to pro-apoptotic stimuli.

Our study has several limitations. Firstly, for the sscMAP process a huge number of gene array samples are required and although the database search gave a high number of initial results, upon detailed inspection several gene array studies could not be included. Connectivity mapping uses gene array data run on Affimetrix platforms and we selected those performed on these platforms. Several published gene array studies were performed in cell lines. However, the majority of samples selected were PNECs (n=40), but we also included data from cell lines. Furthermore, every published dataset has been performed using a specific experimental design with respect to treatments and time points. We selected experiments that used either no stimulation or exposure to *P.aeruginosa* LPS or to *P.aeruginosa* itself. A sample size of 50-100 individual samples is a statistically acceptable sample size to produce an unbiased result and we used 76 individual gene array samples.

Secondly, the reference database was generated using different cell lines: MCF7, HL60, PC3 and SKMEL5. Although all of human origin, none of these cell lines are airway derived. We therefore confirmed the effect of the predicted drugs in airway relevant and disease specific cell lines, determined an effective drug concentration in our disease model and confirmed their effect in primary cells. Additionally, factors such as the interaction between various signalling pathways

and the interplay between genes can affect the functions of the predicted and validated drugs when used in humans.

The aim of our study was to investigate the potential of sscMap to predict A20-inducing drugs from a list of drugs already licenced for the use in humans to make them available for drug repositioning. As a proof of concept we have focused on the LPS induced expression of A20, p65 and cytokines IL-6 and IL-8. However, in addition to its direct regulation of TLR-induced NF-κB activation, A20 is also involved in the negative regulation of the of NLRP3 inflammasome via TLR3/4-(TRIF)-RIPK3 (49) may also inhibit inflammation induced regulated necrosis (necroptosis) via RIPK3 (50), adding further levels of action and complexity to the anti-inflammatory action of A20. Therefore, future work analysing further NF-κB driven cytokines such as TNFα and IL-1β would also indicate if the predicted drugs are able to modify A20 action on the inflammasome.

Summary

To date there is still a need for alternative anti-inflammatory drugs for patients with CF as restoring CFTR function with potentiators and correctors does not directly affect the inherent innate immune defect. The exaggerated inflammatory response is in part due to the lack of the NF-κB regulator A20 and pharmacological induction of A20 is anti-inflammatory. We have shown here that sscMap is a potent tool to predict effective drugs that can modify A20 without totally inhibiting NF-κB. This is particularly important in the clinical setting as pharmacological suppression of inflammation may increase the incidence of infective exacerbations (51). In addition, A20 inducing drugs have to be carefully adjusted as in addition to the A20 induction e.g. ikarugamycin can be pro-apoptotic. A20 inhibits TNF-induced pro-apoptotic signalling by inhibiting both, the activation of caspase 8 and the activation of c-Jun (52). However, neither the drug induced pro-apoptotic mechanisms nor

the A20 induced anti-apoptotic mechanisms have been investigated in this study, but our observation of cytotoxicity despite high anti-apoptotic A20 mRNA levels may suggest an overriding mechanism. Therefore, while sscMap successfully predicts drugs to modify A20, the effect of the candidate drugs must to be confirmed in a suitable model system to optimise treatments.

Our study also suggests that pharmacological induction of A20 may be less efficient in CF airway cells, but given the appropriate stimulation, A20 induction is indeed possible. Tiruppathi *et al*. recently showed that A20 induction may be regulated not only via NF-κB, but also through the opposing effects of the repressor DREAM and transcription factor USF1 (53). Future work will investigate if modifying the balance between A20 repressor and transcription factor in CF can more efficiently re-establish A20 induction.

Materials and Methods

Selection of Gene Array data

A search of PubMed GEO data sets (http://www.ncbi.nlm.nih.gov/gds/) was performed in August 2013 using the search terms “Cystic Fibrosis”, “epithelial cells”, “airways” and “primary cells”. Data sets that passed the search criteria and were compatible to Affymetrix Human Genome U133A Array were selected.

Connectivity Mapping (sscMap)

Gene expression profiles were generated using Affimetrix Gene chip Microarray and the relative expression of treatment vs. control was sorted in descending order giving rise to ~22000 rank ordered genes and their expression.

Determination of the gene signature

A gene signature (a set of genes that behaves in the same way or uniquely under a biological state) was created. A20 (*TNFAIP3*) was used as the known seed gene and a linear regression model was applied to create the gene signature from all selected GEO data. A20 correlates were identified by calculating the *Pearson* correlation coefficient between the expression of A20 and other genes using the formula (where $\bar{x}$ and $\bar{y}$ are the sample means of the two arrays of values and r was calculated in Excel:

$$r = \frac{\sum(x-\bar{x})(y-\bar{y})}{\sqrt{\sum(x-\bar{x})^2 \sum(y-\bar{y})^2}}$$

The correlation coefficient r demonstrates the association between A20 and other genes, either in a positive or negative direction. The significance of the observed correlations was measured by calculating the corresponding p values and applying a stringent p value threshold 1/N where N is the number of genes analysed. The significant r values were

selected as correlates of A20. Gene ontology (GO) enrichment analysis (http://geneontology.org/) was then applied to further filter the A20 correlates identifying those related to NF-κB. The subsequent gene signature (including A20) was used as the input query to evaluate the connection between them and the reference profiles (GEO, Accession Number GSE5258) (8). Based on the principles of Lamb's connectivity mapping, we used a simpler and more robust method (9, 10), called sscMap (statistically significant connections' map), to determine the connections between the gene signature and the reference profile. The similarity between the gene signature and each gene expression reference profile was assessed via a connection score. Connection scores are a function of expression profile and the query gene signature, which is expected to reflect the underlying connection between them. The sscMap applies a robust and improved scoring system based on the following formulae:

Connection strength = $C(\mathbf{R}, \mathbf{s}) = \sum_{i=1}^{n} R(g_i)s(g_i),$

and:

Connection Score = $\frac{\text{Connection Strength}}{\text{Maximum possible connection strength}}$

(Where gi represents the i th gene in the signature, s(gi) is its signed rank in the signature, and R(gi) is this gene's signed rank in the reference profile). To calculate the p value, after calculating the connection strength between a gene signature and the reference profile a large number of random gene signatures are created and the same number of connection scores are calculated and the proportion of scores higher than the observed score in absolute values is the p value. In addition to controlling false positives, sscMap is extended to measure the stability of the connections discovered by gene signature perturbation. In order to implement this, one gene is left out from the gene signature to derive 'perturbation' gene signature and the changes in the significant connections were observed. The connections which stay stable over the changes were given the perturbation stability score, defined as the fraction of times a drug remained significant under the perturbation process (54).

Cell Culture

The bronchial epithelial cell lines 16HBE14o- (control) and CFBE41o- (CF, F508del/F508del), obtained from D. Gruenert, UCSF, USA were cultured as described (32). Primary nasal epithelial cells (PNECs) from CF patients (all F508del/F508del, n=5) and healthy volunteers (n=5, informed consent given, research ethics approval 07/NIR02/23) were cultured as previously described (55). Control participants did not have any acute airways disease at the time of sampling, or a history of any chronic airways inflammation.

Cell culture stimulations

Cells were exposed to the selected drugs (ikarugamycin, quercetin, fluvastatin (all Sigma-Aldrich, SML0188, Q0125, SML0038, 0.01–1000 µM) at 0-1000 µM for 1 h prior to LPS stimulation (*P. aeruginosa* LPS, Sigma-Aldrich, L9143, 10 µg/mL, up to 24h). Stock solutions of the drugs were kept at -20°C for up to 3 months. To minimise oxidation or degradation of the compounds, the working dilutions were freshly prepared.

Determination of LPS induced cytokine release (IL-6 and IL-8)

IL-6 and IL-8 in cell-free culture supernatants were measured by a commercially available ELISA (PeproTech EC Ltd.) according to the manufacturer’s instructions.

LDH Cytotoxicity assay

Lactate dehydrogenase (LDH) release into cell culture supernatants was determined using a LDH-Cytotoxicity Assay Kit (BioVision Ltd.) according to the manufacturer’s instructions.

Real time qPCR

*T*otal RNA was extracted (GenElute™, RTN350, Sigma-Aldrich) and quantified. Equal amounts of RNA (250 ng) were reverse transcribed into cDNA (High-Capacity cDNA Reverse Transcription Kit, Applied Biosystems™) and quantitative RT-PCR performed (LightCycler thermal cycler system, Roche). Expression of A20, p65 and β-actin were assessed using primer sequences previously described (23) and given in supplement table S1. Relative expression to β-actin was calculated as ΔΔCt. Jurkat cell cDNA acted as an internal calibrator for all experiments and was used to determine differences in basal gene expression.

Western Blotting

Cytosolic protein expression was determined by Western Blotting after extraction in RIPA buffer containing protease inhibitors (cOmplete™, Mini, Roche). Lysates were separated by SDS-PAGE and PVDF membranes incubated with 1 µg/ml primary antibody: A20 (ab74037, Abcam); p65 (C-20, Santa Cruz Biotechnology), washed, incubated with appropriate horseradish peroxidase-conjugated antibody and visualized on a BioRadChemi Doc XRS system (BioRad). Anti-GAPDH-HRP (ab-9484, Abcam) was used as a loading control.

Transfections

16HBE140- were seeded at $4x10^4$ cells/well and allowed to attach overnight. Custom FlexiTube siRNA (QIAGEN, UK) was designed against *TNFAIP3* and both cell lines transfected with 50 nM siRNA and Lipofectamine Transfection Reagent (Invitrogen, UK) over 72h. All experiments included mock transfection and scrambled controls. Gene silencing was assessed by qPCR as described above with knockdown of 74% ± 7.2; n=5) achieved.

Statistical analysis

All data are presented as the means ± SEM. Differences between groups were analysed using the Kruskal-Wallis non-parametric ANOVA with Dunn's post-test (*$p<0.05$, **$p<0.01$ and ***$p<0.001$). $ denotes a significant difference compared to medium control, while * denotes significant differences between groups (CF vs. non-CF, comparing the same time points or LPS vs. treatment + LPS). The logarithmic inhibitor concentration versus the relative IL-8 response achieving a 50% inhibition was calculated as the relative IC_{50}. GraphPad Prism (La Jolla, California) was used to plot graphs and to analyse the data.

Acknowledgements

The authors would like to express their thanks to all CF patients and volunteers that took part in this study. BM and HW were in part supported through a summer studentship obtained from the CF Trust UK. SDZ was supported by a BBSRC/MRC/EPSRC co-funded grant (BB/I009051/1). We thank Declan McGuigan and Fiona Manderson Koivula (University of Ulster) for technical assistance with the transfection experiments.

REFERENCES

1. Knorre A, Wagner M, Schaefer HE, Colledge WH, Pahl HL. (2002) DeltaF508-CFTR causes constitutive NF-kappaB activation through an ER-overload response in cystic fibrosis lungs. Biol Chem 383(2):271-82.
2. Cohen TS, Prince A. (2012) Cystic fibrosis: a mucosal immunodeficiency syndrome. Nat Med 18(4):509-19.
3. Blackwell TS, Stecenko AA, Christman JW. (2001) Dysregulated NF-kappaB activation in cystic fibrosis: evidence for a primary inflammatory disorder. Am J Physiol Lung Cell *Mol Physiol* 281(1):L69-70.
4. Paul SM, Mytelka DS, Dunwiddie CT, Persinger CC, Munos BH, Lindborg SR, Schacht AL. (2010) How to improve R&D productivity: the pharmaceutical industry's grand challenge. Nat Rev Drug Discov. 3:203-214.
5. DiMasi JA, Hansen RW & Grabowski HG. (2003) The price of innovation: new estimates of drug development costs. *Journal of health economics* 22:151-185.
6. Mak IWY, Evaniew N, Ghert M. (2014) Lost in translation: animal models and clinical trials in cancer treatment. Am J Transl Res 6(2):114-118.
7. Lamb J, Crawford ED, Peck D, Modell JW, Blat IC, Wrobel MJ, Lerner J, Brunet JP, Subramanian A, Ross KN, Reich M, Hieronymus H, Wei G, Armstrong SA, Haggarty SJ, Clemons PA, Wei R, Carr SA, Lander ES, Golub TR. (2006) The Connectivity Map: using gene-expression signatures to connect small molecules, genes, and disease. Science 29;313(5795):1929-35.
8. Zhang SD, Gant TW. (2008) A simple and robust method for connecting small-molecule drugs using gene-expression signatures. BMC Bioinformatics 9:258.

9. Zhang SD, Gant TW. (2009) sscMap: an extensible Java application for connecting small-molecule drugs using gene-expression signatures. BMC Bioinformatics 10:236.
10. Ramsey JM, Kettyle LM, Sharpe DJ, Mulgrew NM, Dickson GJ, Bijl JJ, Austin P, Mayotte N, Cellot S, Lappin TR, Zhang SD, Mills KI, Krosl J, Sauvageau G, Thompson A. (2013) Entinostat prevents leukemia maintenance in a collaborating oncogene-dependent model of cytogenetically normal acute myeloid leukemia. Stem Cells 31(7):1434-45.
11. Zaman MM, Gelrud A, Junaidi O, Regan MM, Warny M, Shea JC, Kelly C, O'Sullivan BP, Freedman SD. (2004) Interleukin 8 secretion from monocytes of subjects heterozygous for the deltaF508 cystic fibrosis transmembrane conductance regulator gene mutation is altered. Clin Diagn Lab Immunol 11(5):819–824.
12. Kelly C, Williams MT, Elborn JS, Ennis M, Schock BC. (2013) Expression of the inflammatory regulator A20 correlates with lung function in patients with cystic fibrosis. J Cyst Fibros 12(4):411-5.
13. Catrysse L, Vereecke L, Beyaert R, van Loo G. A20 in inflammation and autoimmunity. Trends Immunol. 2014 Jan;35(1):22-31.
14. Li M, Shi X, Qian T, Li J, Tian Z, Ni B, Hao F. A20 overexpression alleviates pristine-induced lupus nephritis by inhibiting the NF-κB and NLRP3 inflammasome activation in macrophages of mice. Int J Clin Exp Med. 2015 Oct 15;8(10):17430-40. eCollection 2015.
15. Matsuzawa Y, Oshima S, Takahara M, Maeyashiki C, Nemoto Y, Kobayashi M, Nibe Y, Nozaki K, Nagaishi T, Okamoto R, Tsuchiya K, Nakamura T, Ma A, Watanabe M.TNFAIP3 promotes survival of CD4 T cells by restricting MTOR and promoting autophagy. Autophagy. 2015;11(7):1052-62.

16. Gu G, Zhang Y, Guo L. Ubiquitin E3 ligase A20 is required in degradation of microbial superantigens in vascular endothelial cells. Cell Biochem Biophys. 2013 Jul;66(3):649-55.
17. Shembade N, Ma A, Harhaj EW. (2010) Inhibition of NF-kappaB signaling by A20 through disruption of ubiquitin enzyme complexes. Science. 327(5969):1135-9.
18. Mabilleau G, Chappard D, Sabokbar A. (2011) Role of the A20-TRAF6 axis in lipopolysaccharide-mediated osteoclastogenesis. J Biol Chem 286(5):3242-9.
19. Catrysse L, Fukaya M, Sze M, Meyerovich K, Beyaert R, Cardozo AK, van Loo G. A20 deficiency sensitizes pancreatic beta cells to cytokine-induced apoptosis in vitro but does not influence type 1 diabetes development in vivo. Cell Death Dis. 2015 Oct 15;6:e1918.
20. An YF, Li TL, Geng XR, Yang G, Zhao CQ, Yang PC. Ubiquitin E3 ligase A20 facilitates processing microbial product in nasal epithelial cells. J Biol Chem. 2012 Oct 12;287(42):35318-23.
21. Onose A, Hashimoto S, Hayashi S, Maruoka S, Kumasawa F, Mizumura K, Jibiki I, Matsumoto K, Gon Y, Kobayashi T, Takahashi N, Shibata Y, Abiko Y, Shibata T, Shimizu K, Horie T. An inhibitory effect of A20 on NF-kappaB activation in airway epithelium upon influenza virus infection. Eur J Pharmacol. 2006 Jul 17;541(3):198-204.
22. Gon Y, Asai Y, Hashimoto S, Mizumura K, Jibiki I, Machino T, Ra C, Horie T. (2004) A20 inhibits toll-like receptor 2- and 4-mediated interleukin-8 synthesis in airway epithelial cells. Am J Respir Cell Mol Biol 31(3):330-6.
23. Kelly C, Williams MT, Mitchell K, Elborn JS, Ennis M, Schock BC. (2013) Expression of the nuclear factor-κB inhibitor A20 is altered in the cystic fibrosis epithelium. Eur Respir J. 41(6):1315-23.
24. Reihill JA, Malcomson B, Bertelsen A, Cheung S, Czerwiec A, Barsden R, Elborn JS, Dürkop H, Hirsch B, Ennis M, Kelly C, Schock BC. (2016) Induction of the

inflammatory regulator A20 by gibberellic acid in airway epithelial cells. Br J Pharmacol Feb;173(4):778-89.

25. Rowe SM, Heltshe SL, Gonska T, Donaldson SH, Borowitz D, Gelfond D, Sagel SD, Khan U, Mayer-Hamblett N, Van Dalfsen JM, Joseloff E, Ramsey BW; GOAL Investigators of the Cystic Fibrosis Foundation Therapeutics Development Network (2014) Clinical mechanism of the cystic fibrosis transmembrane conductance regulator potentiator ivacaftor in G551D-mediated cystic fibrosis. Am J Respir Crit Care Med 90:175–84.
26. Jones AM and Barry PJ. (2015) Lumacaftor/ivacaftor for patients homozygous for Phe508del-CFTR: should we curb our enthusiasm? Thorax 70:615-616.
27. Kang NI, Yoon HY, Lee YR, Won M, Chung MJ, Park JW, Hur GM, Lee HK, Park BH. (2009) A20 attenuates allergic airway inflammation in mice. J Immunol 183(2):1488-95.
28. Qu XA, Rajpal DK. (2012) Applications of Connectivity Map in drug discovery and development Drug Discov Today 17(23-24):1289-98.
29. Casson RJ & Farmer LDM. (2014) Understanding and checking the assumptions of linear regression: a primer for medical researchers. Clinical & Experimental Ophthalmology 42(6):590-596.
30. Odland TM. (1957) Azacyclonol (Frenquel) hydrochloride in the treatment of chronic schizophrenia. A double-blind, controlled study. *JAMA* 165(4):333-335.
31. Ling KH, Leeson GA, Burmaster SD, Hook RH, Reith MK, Cheng LK. (1995) Metabolism of terfenadine associated with CYP3A(4) activity in human hepatic microsomes. Drug Metab Dispos 23(6):631-6.
32. Díaz JG, Ruiz JG, Herz W. (2004) Alkaloids from Delphinium pentagynum. Phytochemistry 65(14):2123-7.

33. Ameri A. (1998) The effects of Aconitum alkaloids on the central nervous system. Prog Neurobiol. 56(2):211-35.

34. Popescu R, Heiss EH, Ferk F, Peschel A, Knasmueller S, Dirsch VM, Krupitza G, Kopp B. (2011) Ikarugamycin induces DNA damage, intracellular calcium increase, p38 MAP kinase activation and apoptosis in HL-60 human promyelocytic leukemia cells. Mutat Res 709-710:60-6.

35. Askari G, Ghiasvand R, Feizi A, Ghanadian SM, Karimian J. (2012) The effect of quercetin supplementation on selected markers of inflammation and oxidative stress. J Res Med Sci. 17(7):637-41.

36. Ying B, Yang T, Song X, Hu X, Fan H, Lu X, Chen L, Cheng D, Wang T, Liu D, Xu D, Wei Y, Wen F. (2009) Quercetin inhibits IL-1 beta-induced ICAM-1 expression in pulmonary epithelial cell line A549 through the MAPK pathways. Mol Biol Rep 36(7):1825-32.

37. Dajas F. (2012) Life or death neuroprotective and anticancer effects of quercetin. *J Ethnopharmacol* 143(2):283-96.

38. Boots AW, Kubben N, Haenen GR, Bast A. (2003) Oxidized quercetin reacts with thiols rather than with ascorbate: implication for quercetin supplementation. Biochem Biophys Res Commun. Aug 29;308(3):560-5.

39. Robinson AJ, Kashanin D, O'Dowd F, Fitzgerald K, Williams V, Walsh GM. (2009) Fluvastatin and lovastatin inhibit granulocyte macrophage-colony stimulating factor-stimulated human eosinophil adhesion to inter-cellular adhesion molecule-1 under flow conditions. Clin Exp Allergy 39(12):1866-74.

40. Folli C, Descalzi D, Bertolini S, Riccio AM, Scordamaglia F, Gamalero C, Barbieri M, Passalacqua G, Canonica GW. (2008) Effect of statins on fibroblasts from human nasal polyps and turbinates. Eur Ann Allergy Clin Immunol 40(3):84-9.

41. Jouneau S, Bonizec M, Belleguic C, Desrues B, Brinchault G, Galaine J, Gangneux JP, Martin-Chouly C. (2011) Anti-inflammatory effect of fluvastatin on IL-8

production induced by Pseudomonas aeruginosa and Aspergillus fumigatus antigens in cystic fibrosis. PLoS One 6(8):e22655.

42. Mantuano E, Santi S, Filippi C, Manca-Rizza G, Paoletti S, Consani C, Giovannini L, Tramonti G, Carpi A, Panichi V. Simvastatin and fluvastatin reduce interleukin-6 and interleukin-8 lipopolysaccharide (LPS) stimulated production by isolated human monocytes from chronic kidney disease patients. Biomed Pharmacother. 2007 Jul;61(6):360-5.
43. Park JW, Siekmeier R, Lattke P, Merz M, Mix C, Schüler S, Jaross W. Pharmacokinetics and pharmacodynamics of fluvastatin in heart transplant recipients taking cyclosporine A. J Cardiovasc Pharmacol Ther. 2001 Oct;6(4):351-61.
44. Qi XF, Zheng L, Lee KJ, Kim DH, Kim CS, Cai DQ, Wu Z, Qin JW, Yu YH, Kim SK. HMG-CoA reductase inhibitors induce apoptosis of lymphoma cells by promoting ROS generation and regulating Akt, Erk and p38 signals via suppression of mevalonate pathway. Cell Death Dis. 2013 Feb 28;4:e518.
45. Takahashi M, Ogata Y, Okazaki H, Takeuchi K, Kobayashi E, Ikeda U, Shimada K. Fluvastatin enhances apoptosis in cytokine-stimulated vascular smooth muscle cells. J Cardiovasc Pharmacol. 2002 Feb;39(2):310-7.
46. Gray ST, Arvelo MB, Hasenkamp W, Bach FH, Ferran C. (1999) A20 Inhibits Cytokine-Induced Apoptosis and Nuclear Factor κB–Dependent Gene Activation in Islets. JEM 190(8):1135-1146.
47. He KL, Ting AT. (2002) A20 Inhibits Tumor Necrosis Factor (TNF) Alpha-Induced Apoptosis by Disrupting Recruitment of TRADD and RIP to the TNF Receptor 1 Complex in Jurkat T Cells. Mol Cell Biol 22(17):6034-6045.
48. Nagamachi A, Nakata Y, Ueda T, Yamasaki N, Ebihara Y, Tsuji K, Honda Z, Takubo K, Suda T, Oda H, Inaba T, Honda H. (2014) Acquired deficiency of A20 results in rapid apoptosis, systemic inflammation, and abnormal hematopoietic stem cell function. PLoS One. 9(1):e87425.

49. Duong BH, Onizawa M, Oses-Prieto JA, Advincula R, Burlingame A, Malynn BA, Ma A. A20 restricts ubiquitination of pro-interleukin-1β protein complexes and suppresses NLRP3 inflammasome activity. Immunity. 2015 Jan 20;42(1):55-67.
50. Onizawa M, Oshima S, Schulze-Topphoff U, Oses-Prieto JA, Lu T, Tavares R, Prodhomme T, Duong B, Whang MI, Advincula R, Agelidis A, Barrera J, Wu H, Burlingame A, Malynn BA, Zamvil SS, Ma A. The ubiquitin-modifying enzyme A20 restricts ubiquitination of the kinase RIPK3 and protects cells from necroptosis. Nat Immunol. 2015 Jun;16(6):618-27.
51. Konstan MW, Döring G, Heltshe SL, Lands LC, Hilliard KA, Koker P, Bhattacharya S, Staab A, Hamilton A; Investigators and Coordinators of BI Trial 543.45. (2014) A randomized double blind, placebo controlled phase 2 trial of BIIL 284 BS (an LTB4 receptor antagonist) for the treatment of lung disease in children and adults with cystic fibrosis. J Cyst Fibros 13(2):148-55.
52. Lademann U, Kallunki T, Jäättelä M. A20 zinc finger protein inhibits TNF-induced apoptosis and stress response early in the signaling cascades and independently of binding to TRAF2 or 14-3-3 proteins. Cell Death Differ. 2001 Mar;8(3):265-72.
53. Tiruppathi C, Soni D, Wang DM, Xue J, Singh V, Thippegowda PB, Cheppudira BP, Mishra RK, Debroy A, Qian Z, Bachmaier K, Zhao YY, Christman JW, Vogel SM, Ma A, Malik AB. (2014) The transcription factor DREAM represses the deubiquitinase A20 and mediates inflammation. Nat Immunol 15(3):239-47.
54. McArt DG, Zhang SD. (2011) Identification of candidate small-molecule therapeutics to cancer by gene-signature perturbation in connectivity mapping. PLoS One 6(1):e16382.
55. de Courcey F, Zholos AV, Atherton-Watson H, Williams MT, Canning P, Danahay HL, Elborn JS, Ennis M. (2012) Development of primary human nasal epithelial cell

cultures for the study of cystic fibrosis pathophysiology. Am J Physiol Cell Physiol. 303(11):C1173-9.

Additional references (appendix):

56. Lai P-F, Cheng C-F, Lin H, Tseng T-L, Chen H-H, Chen S-H. (2013) ATF3 Protects against LPS-Induced Inflammation in Mice via Inhibiting HMGB1 Expression. Evidence-Based Complementary and Alternative Medicine. ID 716481, 14 pages.
57. Boespflug ND, Kumar S, McAlees JW, Phelan JD, Grimes HL, Hoebe K, Hai T, Filippi MD, Karp CL. (2014) ATF3 is a novel regulator of mouse neutrophil migration. Blood 123(13):2084-93.
58. Hai T. (2010) ATF3, a Hub of the Cellular Adaptive-Response Network, in the Pathogenesis of Diseases: Is Modulation of Inflammation a Unifying Component. Gene Expression 15 (1):1-11.
59. Zeigerer A, Gilleron J, Bogorad RL, Marsico G, Nonaka H, Seifert S, Epstein-Barash H, Kuchimanchi S, Peng CG, Ruda VM, Del Conte-Zerial P, Hengstler JG, Kalaidzidis Y, Koteliansky V, Zerial M. (2012) Rab5 is necessary for the biogenesis of the endolysosomal system in vivo. Nature 485: 465–470.
60. Kelly C, Canning P, Buchanan PJ, Williams MT, Brown V, Gruenert DC, Elborn JS, Ennis M, Schock BC. (2013) Toll-like receptor 4 is not targeted to the lysosome in cystic fibrosis airway epithelial cells. Am J Physiol Lung Cell Mol Physiol. Mar 1;304(5):L371-82.
61. Tvrdík D, Dundr P, Povýsil C, Prikryl P, Melcáková S, Planková M. (2007) Downregulation of myc promoter-binding protein 1 (MBP-1) in growth-arrested malignant B cells. Folia Biol (Praha) 53(6):207-15.

62. Sedoris KC, Thomas SD, Miller DM. (2007) C-myc promoter binding protein regulates the cellular response to an altered glucose concentration. Biochemistry 46(29):8659-68.

63. Funakoshi T, Maeshima K, Yahata K, Sugano S, Imamoto F, Imamoto N. (2007) Two distinct human POM121 genes: requirement for the formation of nuclear pore complexes. FEBS Lett 581: 4910-4916.

64. Depping R, Jelkmann W, Kosyna FK. (2015) Nuclear-cytoplasmatic shuttling of proteins in control of cellular oxygen sensing. J Mol Med (Berl) 93(6):599-608.

65. Tabary O, Corvol H, Boncoeur E, Chadelat K, Fitting C, Cavaillon JM, Clément A, Jacquot J. (2006) Adherence of airway neutrophils and inflammatory response are increased in CF airway epithelial cell-neutrophil interactions. Am J Physiol Lung Cell Mol Physiol 290(3):L588-96.

66. Greene CM, Carroll TP, Stephen G. J. Smith SGJ, Taggart CC, Devaney J, Griffin S, O'Neill SJ, McElvaney NG. (2005) TLR-Induced Inflammation in Cystic Fibrosis and Non-Cystic Fibrosis Airway Epithelial Cells. J Immunol 174: 1638 –1646.

67. Shen J, Bronson RT, Chen DF, Xia W, Selkoe DJ, Tonegawa S. (1997) Skeletal and CNS defects in presenilin-1-deficient mice. Cell 89:629–639.

68. Wong PC, Zheng H, Chen H, Becher MW, Sirinathsinghji DJS, Trumbauer ME, Chen HY, et al. (1997) Presenilin 1 is required for Notch1 and Dll1 expression in the paraxial mesoderm. Nature 387:288–292.

69. Herreman A, Hartmann D, Annaert W, Saftig P, Craessaerts K, Serneels L, Umans L, Schrijvers V, Checler F, Vanderstichele H, et al. (1999) Presenilin 2 deficiency causes a mild pulmonary phenotype and no changes in amyloid precursor protein processing but enhances the embryonic lethal phenotype of presenilin 1 deficiency. Proc Natl Acad Sci USA 96:11872–11877.

70. Jiang X, Zhang D, Shi J, Chen Y, Zhang P, Mei B. (2009) Increased inflammatory response both in brain and in periphery in presenilin 1 and presenilin 2 conditional double knock-out mice. J Alzheimers Dis 18(3):515-23.

Legends:

Figure 1: ***Gene expression profile of the gene signature genes associated with A20.*** 16HBE14o- (grey) and CFBE41o- (green) were stimulated (LPS, 10 µg/ml 0-24 h) and mRNA levels of A20, ATF3, Rab5c, DENNDA4, POM121, ICAM-1 and PSNE1 determined as described. $ significant difference compared to medium control, * significant differences between genotypes.

Figure 2: ***A20 and p65 mRNA expression of ikarugamycin, quercetin and fluvastatin treated cells.*** 16HBE14o- and CFBE41o- were pre-incubated with (a) ikarugamycin, (0.01, 1 µM), (b) quercetin (0.1, 100 µM) or (c) fluvastatin (0.1, 1 µM) stimulated (LPS, 10 µg/ml, 0-24 h), A20 mRNA determined (qRT-PCR) and expressed as A20/β-actin relative to the internal control.

Figure 3: ***Effect of ikarugamycin, quercetin and fluvastatin on A20 and p65 protein expression.*** 16HBE14o- and CFBE41o- were pre-incubated with (a) ikarugamycin, (0.01, 1 µM), (b) quercetin (0.1, 100 µM) or (c) fluvastatin (0.1, 1 µM) and stimulated (LPS, 10 µg/ml, 0-24 h). Cytosolic A20 and p65 protein was determined by Western Blotting: 1 = Ctr, 2 = LPS, 3 = drug at lower conc., 4 = drug at lower conc. + LPS, 5 = drug at higher conc., 6 = drug at higher conc., + LPS.

Figure 4: ***The anti-inflammatory effect of ikarugamycin and quercetin is mediated by A20 induction.*** 16HBE14o- with and without knock-down of A20 were pre-incubated with ikarugamycin, (1 µM) or quercetin (100 µM), stimulated and IL-8 analysed. Drug treatment caused a significant reduction in IL-8 ($p<0.05$) compared to LPS stimulation alone. A20 knock-down resulted in a lack of the anti-inflammatory effect of the drug tested.

Figure 5: ***Effect of ikarugamycin, quercetin and fluvastatin on IL-8 release from PNECs from (a) healthy controls and (b) patients with CF.*** The release of IL-8 (pg/ml) was determined using a commercially available IL-8 ELISA kit. Statistical analysis was performed using Wilcoxon paired ranked t-test.

Figure 6: ***Effect of ikarugamycin, quercetin and fluvastatin on A20 and p65 mRNA expression in PNECs from (a) healthy controls and (b) patients with CF.*** Cells were pre-incubated with ikarugamycin, quercetin or fluvastatin at the indicated concentrations (0.01 – 100 µM) and then stimulated (LPS, 10 µg/ml, 0- 24 h). A20 mRNA was determined by qRT-PCR and expressed as A20/β-actin relative to internal control.

Table 1: ***Selected GEO gene expression data*** sets selected for the connectivity mapping process.

Table 2*: Gene expression profile of NF-kB/A20 related genes in CF airway disease*. Genes behaving in a similar way than the target gene A20 were determined using linear correlation analyses of the selected gene expression data sets.

Table 3: ***Candidate compounds predicted to induce or reduce A20 expression***.