

# Rational design of a material for rapid colorimetric Fe<sup>2+</sup> detection

Nadia O. Laschuk,<sup>†</sup> Iraklii I. Ebralidze,<sup>†</sup> Simone Quaranta,<sup>†</sup> Stephen Kerr,<sup>†</sup> Jacquelyn G. Egan,<sup>†</sup> Scott Gillis,<sup>†</sup> Franco Gaspari,<sup>†</sup> Alessandro Latini,<sup>‡</sup> and Olena V. Zenkina<sup>\*†</sup>.

<sup>†</sup> Faculty of Science, University of Ontario Institute of Technology, 2000 Simcoe Street North, Oshawa, ON, Canada, L1H 7K4.

<sup>‡</sup> Dipartimento di Chimica, Università degli Studi di Roma "La Sapienza", Piazzale A. Moro 5, Rome 00185, Italy.

**ABSTRACT:** We report on the rational design of a novel TiO<sub>2</sub>-based screen printed material suitable for sensitive and selective detection of iron ions in water. This includes the synthesis and characterization of large mesoporous TiO<sub>2</sub> nanostructures, screen-printing of titania thick films on glass surfaces and their functionalization with 2,2':6',2''-terpyridin-4'-ylphosphonic acid (terpy). The ultra-high affinity between iron ions and TiO<sub>2</sub>-anchored terpy receptor makes this system potentially applicable for analysis of the iron content in environmental, food, biological, and biomedical systems by a readily quantifiable colour change. Rapid (<30 s) colour change of the material from white to magenta permits easy detection of as low as 0.3 ppm of Fe<sup>2+</sup> by the naked eye. Colour change intensity depends on the nature of the nanoparticles, the overall TiO<sub>2</sub> film thickness, and the Fe<sup>2+</sup> concentration. The material was characterized using profilometry, diffuse reflectance UV-Vis spectroscopy, and X-ray photoelectron spectroscopy (XPS) before and after treatment with aqueous solutions of Fe<sup>2+</sup>. The designed material shows colour reversibility upon treatment with EDTA solutions, which allows for multiple re-uses of the same film with no effect on sensitivity.

**KEYWORDS:** TiO<sub>2</sub>, screen printed films, terpy, iron detection.

## HIGHLIGHTS:

Depositing of terpy-based ligand, **L**, on TiO<sub>2</sub> support results in hybrid material able to selectively react with Fe<sup>2+</sup>.

Impregnation of iron (II) into material leads to distinct colour change from white to magenta.

A novel material based on screen-printed mesoporous TiO<sub>2</sub> beads functionalized by **L** was designed and characterized.

The material is able to quantify 0.3 to 5 ppm of Fe<sup>2+</sup> in aqueous solutions.

## 1. Introduction

Inexpensive fast detection and quantification of metal cations in aqueous solutions is in high demand for a variety of environmental, industrial, forensic, biological and health applications. Especially interesting are systems that allow colorimetric detection to be performed by non-trained personal in a field condition by the naked eye [1, 2]. Thus, the development of materials for such purpose is crucial [3]. Incorporation of sensing receptors on solid supports is a very promising strategy for the development of this type of material. The most common approach is the integration of the functional motif of interest, i.e. a sensing receptor into a polymeric matrix or formation of

self-assembled monolayers (SAMs) containing a sensing receptor on the surface by utilization of an appropriate linker. Various derivatives of 2,2':6',2''-terpyridine (terpy) have been developed and implemented as such sensing receptors as summarized in seminal works by Constable [4] and Hancock [5]. Terpy-based molecules were utilized to identify various metal cations [5-7] and cation-anions pairs [8, 9]. To adjust polarity and therefore solubility, as well as to enhance sensing characteristics, terpy motifs have been combined into a variety of organic, organometallic and supramolecular architectures. These structures were applied for metal ion sensing [10-12], counter anion sensing [8, 13, 14], sensing of bio-molecules [15, 16], and for bio-imaging [17]. Incorporation of terpy derivatives into polymeric matrices often results in the formation of functional polymeric materials that can be employed as effective sensors [18]. For example, solutions of polyiminofluorene based polymer decorated with terpy groups were utilized as a highly sensitive and specific fluorimetric zinc sensor in organic solvent media [10]. Very recently, Garcia et al. reported the incorporation of terpy motif into water soluble acrylic polymers for colorimetric detection of  $\text{Fe}^{3+}$ ,  $\text{Co}^{3+}$ ,  $\text{Sn}^{2+}$ , and  $\text{Cu}^{2+}$  in solution and on solid film [19]. In addition, they reported on the design of polymeric materials able to detect the sub-micromolar amounts of  $\text{Fe}^{3+}$  and  $\text{Co}^{3+}$  and micromolar amounts of  $\text{Cu}^{2+}$  and  $\text{Sn}^{2+}$ . Remarkably, varying the amount of terpy incorporated into the polymer allows for the effective tuning of the selectivity of the resulting polymeric material towards ion sensing. Moreover, a terpy-functionalized poly(ionic liquid) has been utilized for coordination and selective recognition of europium ions that further can be employed in the preparation of novel luminescent materials [20, 21].

Besides the successful integration of terpy into soft materials and polymers, incorporation of this functional component into solid support is an important step for the creation of materials for molecular electronics and photonics [22], electrochromic materials [23], and solar cells. Terpy-motifs have been anchored onto a large number of solid supports resulting in functional materials with enhanced stability and excellent light absorption properties [18, 23-29]. Nishihara et al. have achieved long range electron transport for molecular electric wires by assembling bis(terpyridine) metal complexes into molecular wires on gold surfaces. Successively, they have also used the same molecular motif to create electrochromic responsive nanostructures on the water-organic interface [30, 31]. Terpy-based macroligands were used to prepare yellow-green emitting  $\text{Zn}^{2+}$  metallo-polymer complexes with advanced photophysical properties for OLED applications [32]. Finally, the recent work of Li and Han shows that immobilization of non-planar terpy motifs on graphene-based porous materials through an azide-alkyne "click" reaction enhances porosity and carbon dioxide gas sorption capacity [33].

It is well documented that the terpy-core motif allows visual differentiation of iron cations due to a rapid color change from colorless to deep magenta and this effect has been widely employed for the determination of  $\text{Fe}^{2+}$ . Remarkably, after more than 80 years since the discovery of the first  $\text{Fe}^{2+}$  terpy complex [34], the use of terpy-derivatives for sensing of  $\text{Fe}^{2+}$  has mostly been performed in organic solvents or their mixtures with  $\text{H}_2\text{O}$  [11, 34-36]. To the best of our knowledge, a limited amount of truly water-operational terpy-based metal sensing systems have been reported [19]. In this paper we report the design of a novel material based on terpy-functionalized screen-printed  $\text{TiO}_2$  films, able to detect  $\text{Fe}^{2+}$  in water.

## 2. Experimental

### 2.1. Materials

Titanium (IV) isopropoxide (TIP) (Vertec, 97+%) and Ethylenediaminetetraacetic acid disodium salt dihydrate, 99+%, has been purchased from Alfa Aesar. Ethanol (absolute,  $\geq 99.8\%$ ), anhydrous terpineol, 5-15 mPa·s ethyl cellulose (48.0-49.5% w/w ethoxyl basis), 30-70 mPa·s ethyl cellulose (48.0-49.5% w/w ethoxyl basis), titanium(IV) oxide, anatase nanopowder, <25 nm particle size were purchased from Sigma Aldrich. Aeroxide VP P90 fumed titanium dioxide was kindly donated by Evonik. 2,2':6',2''-terpyridin-4'-ylphosphonic acid was synthesized according previously published procedures [25, 37-39].

### 2.2. Methods

#### 2.2.1. The morphology of SAM-anchored surfaces

STM images were recorded on Nanosurf NaoSTM using Nanosurf Nao 3.4.0 software. All STM images were obtained in the constant current mode by applying a tunneling current  $I_t$  of 70 to 250 pA and a sample bias  $V_t$  of 200 to 800 mV. The STM tips were mechanically cut from Pt/Ir wire (80/20, diameter 0.25 mm, Nanoscience). The calibration of the piezoelectric positioners was verified by atomic resolution imaging of graphite.

#### 2.2.2. Surface coverage determination

UV-vis spectroscopy was used to determine the molecular density of monolayer films deposited on 1 mm quartz substrates as previously reported [40]. The instrument used for these experiments was a dual-beam Varian Cary® 50 UV-Vis spectrophotometer, with a modified sample holder for thin film measurements. Assuming uniform distribution of FeL<sub>2</sub> molecules on the surface and expecting the extinction coefficient in solution and in a monolayer is the same, we can relate the absorbance spectra with a calculated surface concentration, rewriting the Beer–Lambert law as follows:

$$\Gamma_{monolayer} [molecules \cdot cm^{-2}] = \frac{A \cdot 10^{-3} [cm^{-3}] \cdot N_{Av} [molecules \cdot mol^{-1}]}{2 \cdot \epsilon [M^{-1} cm^{-1}]}, \quad (1)$$

Here  $\Gamma_{monolayer}$  is surface concentration,  $A$  is monolayer absorbance,  $N_{Av}$  is Avogadro's number, and  $\epsilon$  is molar extinction coefficient. The factor of 2 in denominator was implemented since quartz slides were functionalized from both sides.

The peak at 560 nm characteristic to FeL<sub>2</sub> gives a molar extinction coefficient in solution of 11970 M<sup>-1</sup> cm<sup>-1</sup>. The monolayer absorbance for this FeL<sub>2</sub> sample was 0.0029.

$$\Gamma_{monolayer} = \frac{0.0029 \cdot 10^{-3} [cm^{-3}] \cdot 6.02 \cdot 10^{23} [molecules \cdot mol^{-1}]}{2 \cdot 11970 [M^{-1} cm^{-1}]} = 7.29 \cdot 10^{13} [molecules \cdot cm^{-2}] \approx$$

0.7 [molecules · nm<sup>-2</sup>] This corresponds to area of 1/0.7=1.43 nm<sup>2</sup> per molecule. That is close to 1.21 nm<sup>2</sup> per molecule determined from the crystal structure [41].

The peak at 313 nm was chosen to estimate surface coverage of the ligand. This peak of L gives a molar extinction coefficient in solution of 4910 M<sup>-1</sup> cm<sup>-1</sup>. The monolayer absorbance for this L

sample was 0.002. Analogously, surface concentration of the **L** is 1.23 [molecules·nm<sup>-2</sup>] that corresponds to 1/1.23=0.81 nm<sup>2</sup> per molecule.

### **2.2.3. Anchoring of L onto TiO<sub>2</sub> Nanoparticles: Preparation of L-TiO<sub>2</sub> NPs**

Immobilization of **L** was accomplished by stirring of the suspension of TiO<sub>2</sub> anatase nanoparticles (200 mg) with the solution of **L** (40 mg, 0.127 mmol) in DI water (40 mL). The mixture was stirred for 24 hours at room temperature. After, the suspension was separated by centrifugation and the liquid part was removed. The solid residue was thoroughly washed with DI water (3 x 50 mL), centrifuged and decanted, washed with ethanol (3 x 50 mL), and dried in vacuum for 24 hours.

### **2.2.4. Reaction of L-TiO<sub>2</sub> NP with Fe<sup>2+</sup> solution.**

Suspension of L-TiO<sub>2</sub> NPs was reacted with 40 mL of 45 ppm solution of Fe<sup>2+</sup> in DI water. The mixture was stirred for 24 hours at room temperature. TiO<sub>2</sub> nanoparticles change colour from white to deep pink within 30 sec. Further stirring in the solution of Fe<sup>2+</sup> in DI water had no influence on the NP colour. After, the mixture was separated by centrifugation. Deep magenta solid residue was thoroughly washed with DI water (6 x 50 mL), and ethanol (3 x 50 mL), centrifuged and decanted. Then solid precipitate was dried in vacuum for 24 hours.

### **2.2.5. Characterization of bare and L-TiO<sub>2</sub> anatase NPs**

Bare and L-TiO<sub>2</sub> anatase NPs were characterized using Hitachi H7000 Transmission Electron Microscope operating under iTEM 5.2 software.

### **2.2.6. Syntheses and deposit of titanium dioxide scaffolds**

Mesoporous titanium dioxide beads (TiO<sub>2</sub> NPs) have been obtained by hydrothermal synthesis according to the procedure reported in Chen et al. [42] and Latini et al. [43]. Briefly, titanium tetraisopropoxide hydrolysis has been conducted in hydroethanolic medium by using hexadecylamine and KCl as templating agent and ionic strength buffer, respectively. Autoclaving and annealing conditions have been previously reported [43]. Screen printing paste comprised of titanium dioxide mesoporous beads (scaffold material), ethyl cellulose (binder and thickener) and  $\alpha$ -terpineol (solvent) has been prepared according to Ito et al. [44] and screen printed on clean soda lime glass or FTO-covered glass. Scaffold's film thickness was adjusted by one-by-one printing of several layers. Furthermore, commercially available high surface area P90 titanium dioxide was implemented into the paste as a scaffold material and screen printed for comparison.

### **2.2.7. Mesoporous TiO<sub>2</sub> NPs characterization**

~~XRD~~ Powder X-ray diffraction analysis of mesoporous TiO<sub>2</sub> beads has been carried out by using a Panalytical X'Pert PRO MPD diffractometer equipped with Cu K $\alpha$  source ( $\lambda = 0.154184$  nm) and a X'Celerator 1D RTMS silicon strip detector. The angular resolution (in  $2\theta$ ) is 0.001°. Beam divergence, peaks asymmetry, and beam width have been adjusted by using a 0.04 rad soller slit, a 1° divergence slit and a 20 mm mask on the incident beam path. Low background level and beam

collimation have been assured by using a 6.6 mm anti-scatter slit and a 0.04 rad collimator on the diffracted beam path. Rietveld refinement analysis has been performed through MAUD software [45]. ~~SEM~~ Scanning electron microscope images of the samples have been collected by using a Zeiss Auriga FE-SEM microscope. ~~Surface area and porosimetry~~. Specific surface area has been determined by using an ASAP 2020 Accelerated Surface Area and Porosimetry Analyzer (Micrometrics Instrument Corporation, U.S.) through the BET (Branauer-Emmett-Teller) procedure. Pore size distribution has been in the mesoporous range has been determined by the BJH (Barret-Joyner-Halenda) method by the desorption branch of the isotherm.

### 2.2.8. Anchoring of L onto TiO<sub>2</sub> screen-printed films

Screen-printed films of TiO<sub>2</sub> on glass were immersed into solution of L (20 mg, 0.064 mmol) in 100 mL of DI water. After 30 min films were copiously washed with DI water and isopropanol (4 x 4 mL) and dried by stream of air or N<sub>2</sub>. The same procedure was applied after each regeneration cycle.

### 2.2.9. Characterization of screen-printed films

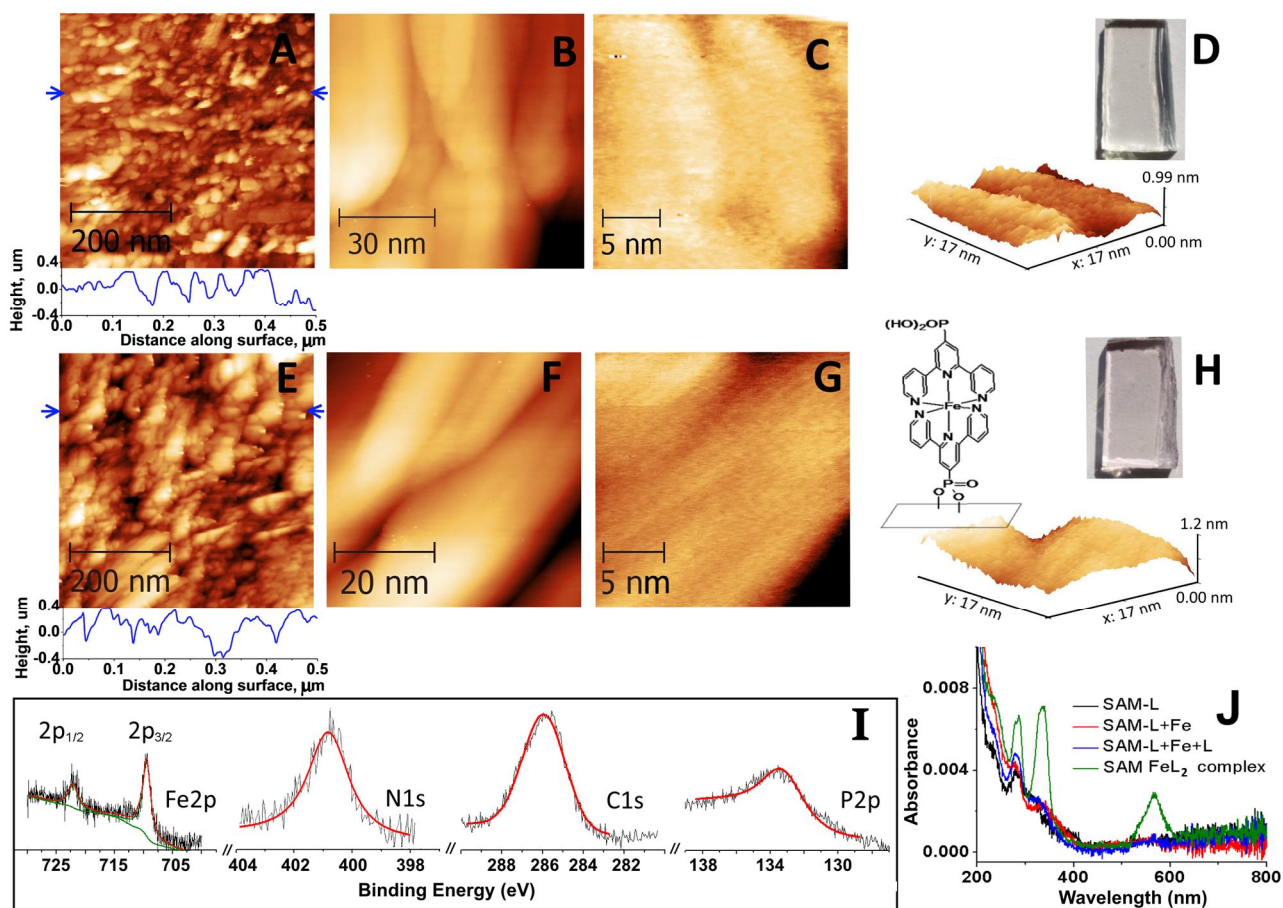
~~Profilometry~~ Tencor Alpha-Step-200 profilometer was equipped with a standard stylus of 12.5 micron radius. Sample leveling was automatically computed after each scan and digitized using GetData Graph Digitizer 2.24 software. Typical scan lengths were 10,000 um. ~~UV-Vis Diffuse Reflectance Spectroscopy~~. Diffuse reflectance measurements have been performed on the bare, *N*-aromatic ligand sensitized and Iron adsorbed films. The spectra have been collected in the range 280-1200 nm in transmittance mode using a double-beam Lambda 750S spectrophotometer (Perkin-Elmer, U.S.) equipped with a 60 mm integrating sphere. ~~X-Ray Photoelectron Spectroscopy~~ XPS measurements were performed using a Thermo Scientific K-Alpha surface analysis system equipped with a monochromated Al K $\alpha$  X-ray source (1486.7 eV). Ultra high vacuum (UHV) conditions ( $10^{-9}$  to  $10^{-10}$  Torr) were maintained during the experiment. The instrument is fitted with a flood source for an effective charge compensation, The Ti<sup>4+</sup> 2p<sub>3/2</sub> peak of TiO<sub>2</sub> at 458.6 eV was used as an internal reference peak for calibrating the binding energy. A Smart fit algorithm was used for background subtraction and a Powell peak-fitting algorithm for data analysis. The elemental ratio was determined using Wagner atomic sensitivity factors [46].

## 3. Results and discussion

~~It is well documented that the terpy core motif allows visual differentiation of iron cations due to a rapid color change from colorless to deep magenta and this effect has been widely employed for the determination of Fe<sup>2+</sup>. Remarkably, after more than 80 years since the discovery of the first Fe<sup>2+</sup> terpy complex [34], the use of terpy derivatives for sensing of Fe<sup>2+</sup> has mostly been performed in organic solvents or their mixtures with H<sub>2</sub>O [11, 34-36]. To the best of our knowledge, a limited amount of truly water operational terpy based metal sensing systems have been reported [19].~~ Recently, we utilized 2,2':6',2''-terpyridin-4'-ylphosphonic acid (L), which is able to interact with the number of metal ions via its *N*-aromatic terpy moiety, for the selective detection of Fe<sup>2+</sup>, Fe<sup>3+</sup>, Ru<sup>3+</sup> and Zn<sup>2+</sup> in aqueous media [41]. Reliable and fast quantification of 50 ppb to 1 ppm levels of each target cation was demonstrated in solution. In this paper ~~the present communication~~ we report the embedding of terpy-based water soluble receptors onto hydrophilic glass, fluorine-doped tin oxide

(FTO) coated glass, TiO<sub>2</sub> nanoparticles, and screen-printed porous surfaces, and explore the performance of these novel materials as Fe<sup>2+</sup> sensors. The phosphonate functional group not only makes the ligand water soluble, but allows for the formation of robust self-assembled monolayers on a variety of surfaces. Thus phosphonates have been utilized for the surface modification of a wide range of oxide substrates [47] such as titania [48, 49], alumina [50], iron oxides [51], hafnium oxide [52], indium-tin oxide (ITO) [53], and silica [54]. Phosphonate-anchored SAMs are easily prepared in air, and form well-ordered strongly bound films on oxide surfaces with molecular densities close to those found in single crystals [55]. Taking this into account, we prepared the complex FeL<sub>2</sub> and deposited it onto FTO/glass and quartz surfaces (Fig. 1). Scanning tunneling microscopy (STM) analysis shows that the roughness of the FeL<sub>2</sub> functionalized FTO/glass surface (Fig 1E-H) is very similar of that of unfunctionalized FTO/glass (Fig 1 A-D), which suggest the formation of a monolayer. The embedding of the complex on the glass, quartz or FTO/glass surfaces can be easily observed by the naked eye. On the contrary, ligand anchoring of L on quartz surfaces does not result in any visible colour. However, this process could be monitored by UV-vis spectroscopy which confirms the formation of a monolayer of L on the surface (Fig. 1J). Moreover, insertion of quartz slides with the pre-deposited L ligand in 0.1 mM solution of Fe<sup>2+</sup> results in insignificant changes in UV-vis spectra of the quartz substrates. Further immersion of these slides into a 0.1 mM solution of the ligand does not change the UV-vis spectra of surface-bonded layer at 560 nm, which is characteristic of a metal-to ligand charge transfer (MLCT) transition for a FeL<sub>2</sub> type complex [56], suggesting that Fe<sup>2+</sup> did not incorporate into the ligand anchored to the surface. In contrast, preparation of the FeL<sub>2</sub> complex in solution followed by its deposition on quartz slides results in a significant increase of intensities at 287 nm and 332 nm, as well as the appearance of a new peak at 569 nm (see Fig. 1J). The red shift of the on-surface FeL<sub>2</sub> monolayer with respect to FeL<sub>2</sub> in aqueous solution (278, 321 and 560 nm, respectively) is the consequence of the change in the microenvironment of FeL<sub>2</sub> in these two different media [57].

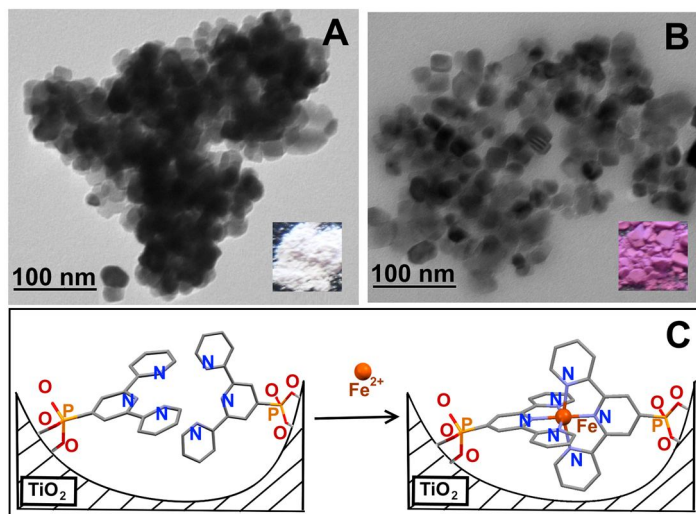
Further characterization of the FeL<sub>2</sub> complex deposited on glass surface by XPS (Fig. 1I) verifies the presence of iron, nitrogen, carbon and phosphorous on the surface within ratios close to the molecular stoichiometry. The binding energy of any XPS peak depends on the oxidation state of an element and on its binding site on the surface. Thus the binding energy of the Fe 2p<sub>3/2</sub> peak is located at 709.6 eV; the binding energy of Fe 2p<sub>1/2</sub> can be seen at 722.1 eV; both peaks are characteristic for Fe<sup>2+</sup>. The N 1s peak at 400.9 eV is evidently associated with aromatic nitrogen that provides the electron density to form the complex. The C 1s peak is centered at 286.0 eV, which is in a good agreement with aromatic carbon. An asymmetric broad peak observed at 133.6 eV is due to the overlapping of the P 2p<sub>3/2</sub> and P 2p<sub>1/2</sub> components, which is characteristic for phosphates.



**Figure 1.** Representative STM images: (A-D) bare FTO/glass and (E-H) SAM-FeL<sub>2</sub> on FTO/glass slides. Representative XPS data for SAM-FeL<sub>2</sub> on glass slides (I): Fe(2p), N(1s), C(1s), and P(2p). The black line shows the experimental data, while the colored lines are the overall fitted spectra. UV-vis spectra (J) of as-deposited SAM-L (black), followed by its step-by-step treatment with Fe<sup>2+</sup> aqueous solution (red) and solution of L (blue) show no significant change of the spectra suggesting no FeL<sub>2</sub> complex formation. In comparison, UV-vis spectrum of FeL<sub>2</sub> complex pre-formed in solution and then deposited as a SAM on quartz surface (green) show a distinguishable characteristic peak at 569 nm.

To estimate the packing density of FeL<sub>2</sub> molecules on glass surfaces, the two-dimensional surface concentration  $\Gamma_{\text{monolayer}}$  (molecules cm<sup>-2</sup>) was calculated based on UV-vis data [40]. For this FeL<sub>2</sub> solution molar extinction coefficient of  $\epsilon_{560\text{nm}} = 11\,970\text{ M}^{-1}\text{ cm}^{-1}$  and the absorbance observed for the monolayer (see Fig. 1J and experimental section) resulted in a  $\Gamma_{\text{monolayer}}$  of  $7 \cdot 10^{13}$  molecules/cm<sup>2</sup>. This corresponds to a surface area occupied by one molecule of FeL<sub>2</sub> equal to 1.43 nm<sup>2</sup>. The surface area estimated from the crystal structure of FeL<sub>2</sub> is close to 1.2 nm<sup>2</sup> per molecule. The surface concentration of L was found to be 1.23 molecules/nm<sup>2</sup> that corresponds to 0.81 nm<sup>2</sup> per molecule. This suggests that the ligand forms well-ordered and dense packed monolayers on relatively smooth surfaces and thus due to the intermolecular  $\pi$ - $\pi$  interactions the rotation of the terpy moieties that are anchored to the surface is limited. Reduced flexibility of the terpy moieties on the surface, in turn, restricts the participation of two neighboring surface-anchored ligands in complex formation.

We proposed that when a high specific surface area substrate is functionalized with the ligand, a certain fraction of the ligand molecules might be available for  $\text{FeL}_2$  complex formation. Indeed, modification of commercially available titanium (IV) oxide anatase nanopowder by **L** followed by treatment of the functionalized **L**- $\text{TiO}_2$  NPs with  $\text{Fe}^{2+}$  solutions results in significant colour change (Fig. 2, A&B insets). Particle size distribution of the nanopowder functionalized by **L** before (Fig. 2A) and after treatment with  $\text{Fe}^{2+}$  (Fig. 2B) remains very similar and close to 25 nm in diameter as declared for bare  $\text{TiO}_2$  nanopowder by the manufacturer.

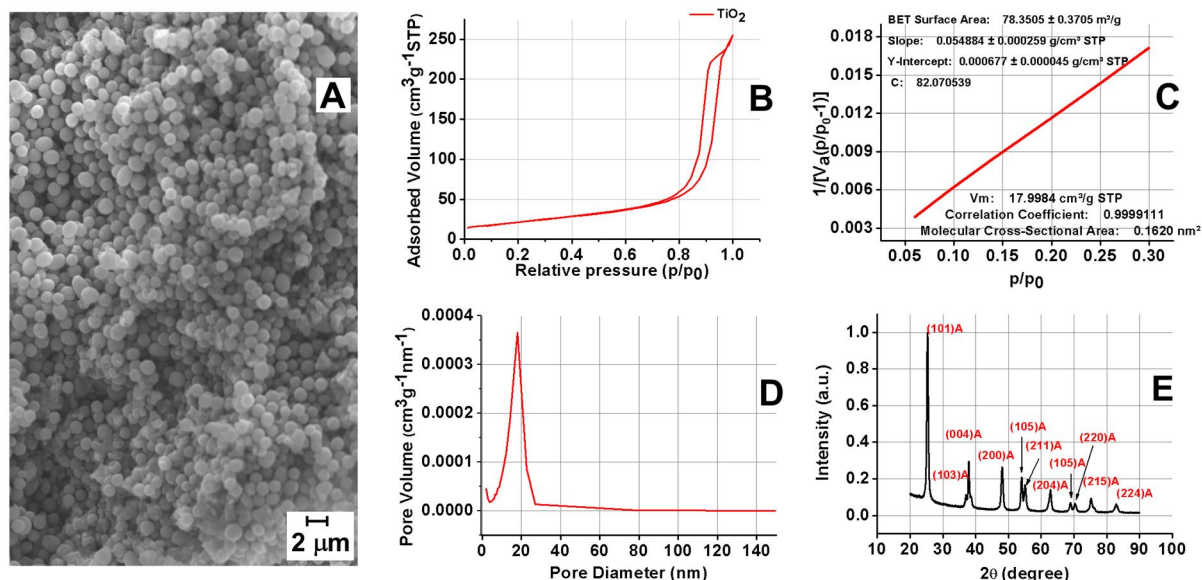


**Figure 2.** Representative TEM images of commercial  $\text{TiO}_2$ , anatase nanopowder, functionalized by **L**. **A:** before and **B:** after treatment with  $\text{Fe}^{2+}$  aqueous solutions. **C:** Proposed reaction mechanism of  $\text{Fe}^{2+}$  interactions with **L** molecules bound to  $\text{TiO}_2$  nanopowder surface.

To make this system more practical and applicable for easy and rapid  $\text{Fe}^{2+}$  detection, a paste comprised of high (70-110  $\text{m}^2/\text{g}$ ) specific surface area Evonik P-90 commercial  $\text{TiO}_2$  nanopowder (scaffold material), ethyl cellulose (binder and thickening agent) and  $\alpha$ -terpineol (solvent) has been prepared according to previously reported procedure [44]. The paste was screen printed on base piranha solution pre-treated soda lime glass as 1x1 cm squares, dried and thermally treated in air. One, three and six layers (2  $\mu\text{m}$  each) were screen printed one-by-one to increase the volume of the detecting cell and thus tune the detection range. When prepared, layered screen printed P-90- $\text{TiO}_2$  films were functionalized via immersion into a 0.65 mM aqueous solution of **L** for one hour followed by washing with isopropyl alcohol (3x3 mL) and dried by a stream of air. The resulting sensing strips were treated with 150  $\mu\text{L}$  of 45 ppm  $\text{Fe}^{2+}$  aqueous solution; all sensing strips turned deep magenta with comparable intensity. These results confirm that novel material can detect  $\text{Fe}^{2+}$  in an aqueous solution. However, we believe that P90 small particles size (typically 15 nm) favors sintering processes resulting in the formation of surfaces where diffusion through the layers is impeded. As such only a reduced amount of surface remains accessible for the functionalization by the molecular receptor **L**, which is active in metal-ion recognition. As a result, P90- $\text{TiO}_2$  based screen-printed materials do not allow reasonable control of sensitivity since no significant difference in colour intensity was observed between one and multiple (up to 6 layers) screen-printed films.



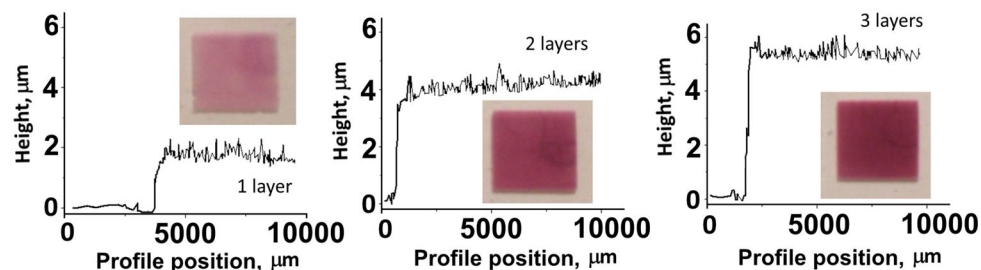
To validate our initial assumption regarding the role of nanoparticle dimensions and pore size, we synthesized TiO<sub>2</sub> nanoparticles (crystallites size estimated by XRD ~25 nm) arranged in large (1263±86 nm in diameter) mesoporous (Fig. 3A) hierarchical spheres (beads). The presence of mesopores (IUPAC classification 2-50 nm) is easily recognizable by the characteristic hysteresis in the N<sub>2</sub> adsorption-desorption graph (Fig. 3B). Specific surface area determined by the BET (Branauer-Emmett-Teller) method has been found to be 78.4±0.4 m<sup>2</sup>/g. Pore size distribution was determined by the BJH (Barret-Joyner-Halenda) method (Fig. 3D) and it has revealed a pore diameter distribution peaking at ca. 18 nm. Thus, although specific surface area of synthesized mesoporous TiO<sub>2</sub> nanoparticles is comparable to the P-90 one, the former are characterized by an almost monodisperse porosity stemming from spherical organization of randomly oriented nanocrystals. Moreover, the spherical shape of mesoporous TiO<sub>2</sub> beads minimize the surface energy tend to avoid surface shrinking during the annealing process. Further analysis of the synthesized mesoporous TiO<sub>2</sub> material by X-ray powder diffraction followed by Rietveld refinement analysis (MAUD software)[45] results in the following lattice parameters: a = 3.782±0.001 Å; c = 9.492±0.003 Å. The data are in good agreement with the reference data (a = 3.784479 Å, c = 9.5137 Å) for the tetragonal crystal structure of anatase (space group i41/amd).



**Figure 3.** **A:** Typical SEM image of synthesized mesoporous TiO<sub>2</sub> NPs. **B:** Adsorption-desorption isotherm for mesoporous TiO<sub>2</sub> NPs. **C:** BET linear region fit for TiO<sub>2</sub> mesoporous beads. **D:** Pore diameters distribution obtained by BJH method. **E:** Powder diffraction spectrum of mesoporous TiO<sub>2</sub> beads.

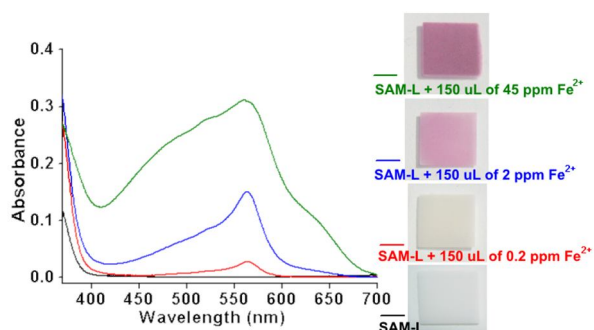
Synthesis of mesoporous beads followed by their layer-by-layer additive screen-printing and functionalization by L, results in Fe<sup>2+</sup> detecting films with a well-tunable thickness (Fig. 4). These mesoporous screen printed materials, in contrast to ones made from commercially available P90 TiO<sub>2</sub> NPs, benefit from better water diffusion and could be applied for quantitative Fe<sup>2+</sup> detection. Dropwise addition of 150 μL of a 45 ppm aqueous solution of Fe<sup>2+</sup> on 1, 2, and 3 layered screen printed films of mesoporous TiO<sub>2</sub> nanopowder results in a rapid colour change from white to 3

shades of purple starting from less intense for the 1 layered film to most intense for the 3 layered one (Fig. 4). We think both mesoporosity and particle size are important factors allowing for modulation of sensitivity and better quantitative analysis.



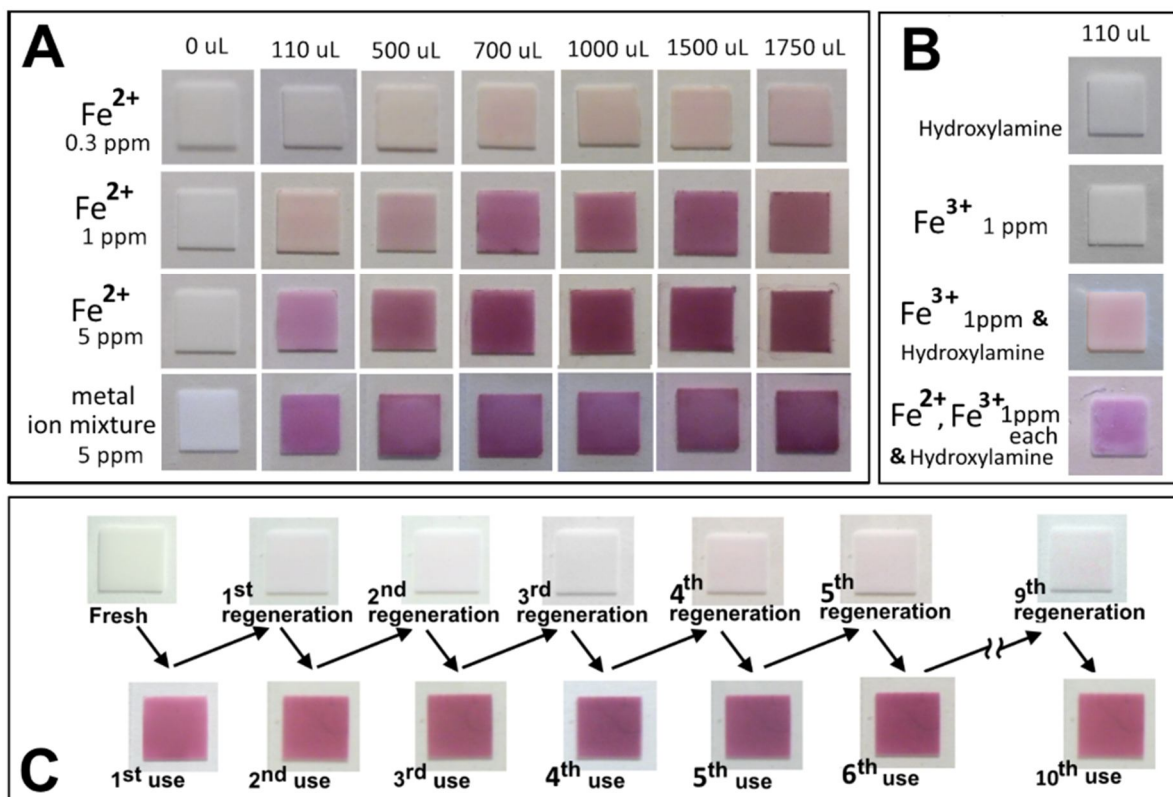
**Figure 4.** Typical profilometry curves of the mesoporous  $\text{TiO}_2$  nanopowder deposited and sintered on FTO glass substrates show thickness of each layer of 2  $\mu\text{m}$ . Inserts: the images of corresponding films after applying of 150  $\mu\text{L}$  of 45 ppm  $\text{Fe}^{2+}$  aqueous solution.

Three layered (6  $\mu\text{m}$  height) screen-printed films of mesoporous  $\text{TiO}_2$  functionalized by **L** (further – detecting films) were further studied to determine the detectable  $\text{Fe}^{2+}$  concentration range. Applying 150  $\mu\text{L}$  of 0.2 ppm, 2 ppm, and 45 ppm solutions of  $\text{Fe}^{2+}$  to the detecting films results in the appearance of a peak at 560 nm on the diffuse reflectance UV-Vis spectra (Fig. 5). Slight colour change takes place in the case of 0.2 ppm  $\text{Fe}^{2+}$  solution, but a naked eye distinguishable colour change was observed when 2 ppm and 45 ppm solutions have been used.



**Figure 5.** Diffuse reflectance UV-Vis spectra and optical photographs of mesoporous  $\text{TiO}_2$  screen printed film (3 screen printed layers) functionalized by **L** and after treatment with 150  $\mu\text{L}$  of  $\text{Fe}^{2+}$  aqueous solutions. The characteristic peak at 560 nm confirms the formation of the  $\text{FeL}_2$  complex.

It should be mentioned that according to the U.S. Environmental Protection Agency secondary drinking water standard and Guidelines for Canadian drinking water quality, maximum contaminant level of iron is 0.3 ppm; the United Nations Food and Agriculture Organization recommended level for irrigation waters is 5 ppm. On the other hand, iron in groundwater (in ferrous i.e.  $\text{Fe}^{2+}$  form) vary from 0.1 to 10 ppm [58]. Needless to say, the development of inexpensive, reusable and reliable materials which are able to rapidly detect iron in field conditions is in high demand. To insure our detecting films can be used as a material for  $\text{Fe}^{2+}$  determination, certain amounts of  $\text{Fe}^{2+}$  aqueous solutions were added to each film creating an appropriate coloring wheel (Fig. 6).



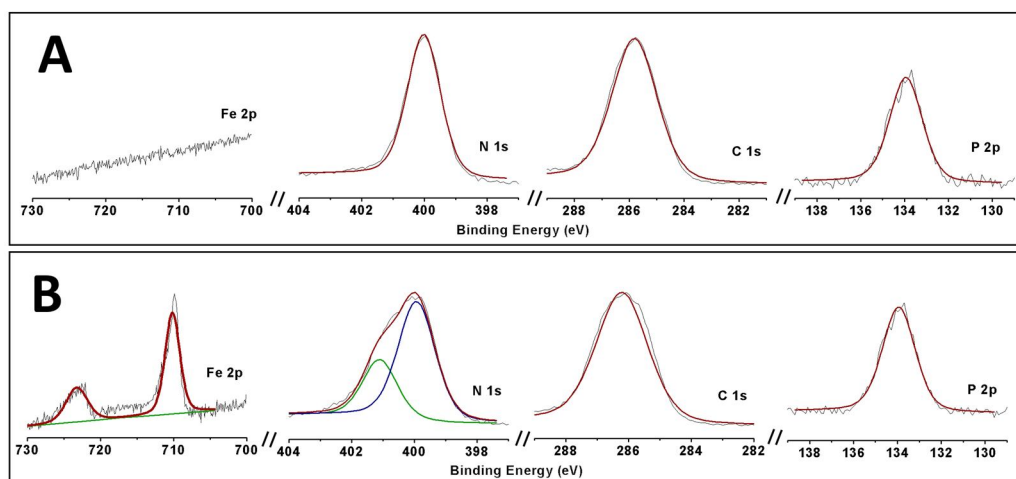
**Figure 6. A:** Colour wheel showing the colour change of detecting film after applying certain volumes of Fe<sup>2+</sup> aqueous solutions: 0.3 ppm (upper row), 1ppm (2<sup>nd</sup> line from the top), 5 ppm (3<sup>rd</sup> line from the top), and a mixture of metal ions (including Fe<sup>2+</sup>) 5 ppm each (bottom row) see main text for details. **B:** Water solutions of hydroxylamine do not change the colour of the detecting film, but reduce Fe<sup>3+</sup> to Fe<sup>2+</sup> making possible the detection of total iron in solution. **C:** Detecting films can be recovered by EDTA and reused, see main text for details.

Colour wheel results are consistent with our expectations: the colour intensity depends on the amount of Fe<sup>2+</sup> loaded. This permits the visual quantification of iron cations in aqueous solutions ranging from 0.3 ppm to 5 ppm. To investigate the detecting films' selectivity towards Fe<sup>2+</sup> in the presence of potentially interfering ions, an aqueous solution containing the following ions: Na<sup>+</sup>, Fe<sup>2+</sup>, Cr<sup>3+</sup>, Sn<sup>2+</sup>, Ca<sup>2+</sup>, Li<sup>+</sup>, Co<sup>2+</sup>, Mg<sup>2+</sup>, Cu<sup>2+</sup>, Cs<sup>+</sup>, Ni<sup>2+</sup>, Ba<sup>2+</sup>, K<sup>+</sup>, and Mn<sup>2+</sup> (5 ppm each) was prepared and portionwise applied to the detecting films. Fig. 6A clearly shows similar colour intensity of the detecting films after dripping certain volumes of 5 ppm solutions containing solely Fe<sup>2+</sup> and Fe<sup>2+</sup> in a mixture with thirteen other metal ions. This shows the material is highly selective to Fe<sup>2+</sup> determination.

In addition, we studied the ability of our screen-printed material to detect Fe<sup>3+</sup> ions in the aqueous solutions. Despite of the fact that aqueous solutions of Fe<sup>3+</sup> do not change colour of the detecting film, Fe<sup>3+</sup> can be detected in aqueous solutions if reduced to Fe<sup>2+</sup> by hydroxylamine [59] (Fig. 6B).

Notably, the detecting films can be reused by immersing for 48 hours into a 0.5 mM solution of ethylenediaminetetraacetic acid disodium salt (EDTA). Iron release from the films into the EDTA

solution can be monitored by the visual color change of the films. Figure 6C shows color films behavior for 10 successful cycles of resetting, no evidence of sensitivity degradation can be detected. Increasing the temperature of the EDTA solution to 80°C reduces the film recovery time to 24 h.



**Figure 7.** Comparative X-ray photoelectron spectra of the **L** anchored to mesoporous TiO<sub>2</sub> screen printed films **A**: before and **B**: after applying of 110 μL of 5 ppm Fe<sup>2+</sup> aqueous solution.

XPS analysis of detecting films before interaction with Fe<sup>2+</sup> confirms the presence of N, C, and P of **L** on the surface (Fig. 7A). The sharp (with full width at half-maximum (FWHM) of 1.9 eV) N 1s peak is centered at 400.0 eV. The C 1s peak is centered at 285.8 eV, as expected for aromatic carbon. An asymmetric broad peak observed at 133.8 eV is characteristic for P 2p. The N:C:P ratio of 2.5:17.4:1.0 is close to 3:15:1 of **L** molecular stoichiometry. When 110 μL of 5 ppm Fe<sup>2+</sup> aqueous solution was applied to the detecting film (Fig 7B) the appearance of two new peaks was observed. The peaks with the binding energies of 709.9 eV and 722.4 eV are characteristic for Fe 2p<sub>3/2</sub> and Fe 2p<sub>1/2</sub> of Fe<sup>2+</sup>, respectively. Interestingly, the P 2p peak position remains within the experimental error the same (133.8 eV) as before Fe<sup>2+</sup> solution application, the C 1s peak is insignificantly shifted to 286.1 eV, but N 1s peak undergoes a significant change and now can be deconvoluted using a Powell peak-fitting algorithm with full width at half-maximum (FWHM) of 1.3 eV yielding a peak centered at 400.0 eV and a new peak centered at 401.1 eV. This value is close to 400.9 eV, which was observed for the FeL<sub>2</sub> complex deposited on the FTO surface (Fig. 11). The P:Fe ratio is 1:0.01 suggesting that as expected, just a part of **L** molecules is able to form FeL<sub>2</sub> complex. On the other hand, assuming that the complex formation takes place not on the flat surface, but inside the pores, the iron content can be underestimated due to the signal attenuation.

#### 4. Conclusions

We observed that 2,2':6',2''-terpyridin-4'-ylphosphonic acid, **L**, forms dense SAMs on FTO/glass and quartz surfaces. Due to the close packing of the **L** molecules, these SAMs, once formed, cannot react with Fe<sup>2+</sup> to form the FeL<sub>2</sub> complex. On the other hand, in-solution, pre-formed the FeL<sub>2</sub> complex forms dense SAMs on glass surface which are observable by the naked eye as pink colouration of the glass. Depositing of SAMs of **L** on high surface area TiO<sub>2</sub> nanostructures results

in a situation when some of the **Ls** are able to form  $\text{FeL}_2$  complex. The formation of the  $\text{Fe}^{2+}$  complex in turn leads to significant colouration of the material. To improve detection abilities and adjust the  $\text{Fe}^{2+}$  detection limits, large mesoporous  $\text{TiO}_2$  beads have been synthesized and screen-printed on glass slides. Functionalization of these beads by **L** results in reusable detecting films able to quantify  $\text{Fe}^{2+}$  in aqueous solutions with the concentration range from 0.3 ppm to 5 ppm. The selectivity of  $\text{Fe}^{2+}$  detection in the presence of 13 other metal ions was shown. This material is expected to be useful for the design of more advanced efficient optical, chemical, and biological sensors. Such sensors would benefit from high accuracy, reliability, portability and cost effectiveness.

## AUTHOR INFORMATION

### Corresponding Author

E-mail address: [olena.zenkina@uoit.ca](mailto:olena.zenkina@uoit.ca) (Olena Zenkina)

## ACKNOWLEDGMENT

Research supported by the University of Ontario Institute of Technology start-up grant and NSERC Discovery grant.

## REFERENCES

1. N. Kaur, S. Kumar, Colorimetric metal ion sensors, *Tetrahedron* 67 (48) (2011) 9233-9264.
2. G. Korotcenkov, B.K. Cho, Instability of metal oxide-based conductometric gas sensors and approaches to stability improvement (short survey), *Sens. Actuators, B* 156 (2) (2011) 527-538.
3. W.E. Duckworth, Sensor materials, *Mater. Des.* 9 (2) (1988) 72-77.
4. Constable E.C. *The Coordination Chemistry of 2,2':6',2''-Terpyridine -Terpyridine and Higher Oligopyridines*. Academic Press: 1986; Vol. 30, p 69-121.
5. J.M. Hamilton, M.J. Anhorn, K.A. Oscarson, J.H. Reibenspies, R.D. Hancock, Complexation of Metal Ions, Including Alkali-Earth and Lanthanide(III) Ions, in Aqueous Solution by the Ligand 2,2',6',2''-Terpyridyl, *Inorganic Chemistry* 50 (7) (2011) 2764-2770.
6. F. Barigelletti, L. Flamigni, G. Calogero, L. Hammarstrom, J.-P. Sauvage, J.-P. Collin, A functionalized ruthenium(II)-bis-terpyridine complex as a rod-like luminescent sensor of zinc(II), *Chem. Commun.* (21) (1998) 2333-2334.
7. R.D. Hancock, The pyridyl group in ligand design for selective metal ion complexation and sensing, *Chem. Soc. Rev.* 42 (4) (2013) 1500-1524.
8. C. Bhaumik, S. Das, D. Saha, S. Dutta, S. Baitalik, Synthesis, Characterization, Photophysical, and Anion-Binding Studies of Luminescent Heteroleptic Bis-Tridentate Ruthenium(II) Complexes Based on 2,6-Bis(Benzimidazole-2-yl)Pyridine and 4'-Substituted 2,2':6',2'' Terpyridine Derivatives, *Inorganic Chemistry* 49 (11) (2010) 5049-5062.
9. G. Baggi, M. Boiocchi, C. Ciarrocchi, L. Fabbrizzi, Enhancing the Anion Affinity of Urea-Based Receptors with a Ru(terpy) $^{2+}$  Chromophore, *Inorganic Chemistry* 52 (9) (2013) 5273-5283.
10. A.R. Rabindranath, A. Maier, M. Schäfer, B. Tiede, Luminescent and Ionochromic Polyiminofluorene with Conjugated Terpyridine Substituent Groups, *Macromol. Chem. Phys.* 210 (8) (2009) 659-668.
11. Y. Hong, S. Chen, C.W.T. Leung, J.W.Y. Lam, J. Liu, N.-W. Tseng, R.T.K. Kwok, Y. Yu, Z. Wang, B.Z. Tang, Fluorogenic Zn(II) and Chromogenic Fe(II) Sensors Based on Terpyridine-Substituted

- Tetraphenylethenes with Aggregation-Induced Emission Characteristics, *ACS Appl. Mater. Interfaces* 3 (9) (2011) 3411-3418.
12. P. Wang, T.-a. Okamura, H.-P. Zhou, W.-Y. Sun, Y.-P. Tian, Metal complex with terpyridine derivative ligand as highly selective colorimetric sensor for iron(III), *Chin. Chem. Lett.* 24 (1) (2013) 20-22.
  13. S.M. Brombosz, A.J. Zuccherro, R.L. Phillips, D. Vazquez, A. Wilson, U.H.F. Bunz, Terpyridine-Based Cruciform-Zn<sup>2+</sup> Complexes as Anion-Responsive Fluorophores, *Org. Lett.* 9 (22) (2007) 4519-4522.
  14. X. Peng, Y. Xu, S. Sun, Y. Wu, J. Fan, A ratiometric fluorescent sensor for phosphates: Zn<sup>2+</sup>-enhanced ICT and ligand competition, *Org. Biomol. Chem.* 5 (2) (2007) 226-228.
  15. M. Ali, S. Nasir, Q.H. Nguyen, J.K. Sahoo, M.N. Tahir, W. Tremel, W. Ensinger, Metal Ion Affinity-based Biomolecular Recognition and Conjugation inside Synthetic Polymer Nanopores Modified with Iron-Terpyridine Complexes, *J. Am. Chem. Soc.* 133 (43) (2011) 17307-17314.
  16. T.A. Ali, G.G. Mohamed, A.H. Farag, Electroanalytical studies on Fe(III) ion-selective sensors based on 2-methyl-6-(4-methylenecyclohex-2-en-1-yl)hept-2-en-4-one ionophore, *Int. J. Electrochem. Sci.* 10 (1) (2015) 564-578.
  17. Q. Zhang, X. Tian, Z. Hu, C. Brommesson, J. Wu, H. Zhou, S. Li, J. Yang, Z. Sun, Y. Tian, K. Uvdal, A series of Zn(II) terpyridine complexes with enhanced two-photon-excited fluorescence for in vitro and in vivo bioimaging, *J. Mater. Chem. B.* 3 (36) (2015) 7213-7221.
  18. A. Wild, A. Winter, F. Schlutter, U.S. Schubert, Advances in the field of pi-conjugated 2,2':6',2''-terpyridines, *Chem Soc Rev* 40 (3) (2011) 1459-1511.
  19. M. Trigo-López, A. Muñoz, S. Ibeas, F. Serna, F.C. García, J.M. García, Colorimetric detection and determination of Fe(III), Co(II), Cu(II) and Sn(II) in aqueous media by acrylic polymers with pendant terpyridine motifs, *Sens. Actuators, B* 226 (2016) 118-126.
  20. Q. Ru, Z. Xue, Y. Wang, Y. Liu, H. Li, Luminescent Materials of Europium(III) Coordinated by a Terpyridine-Functionalized Poly(Ionic Liquid), *Eur. J. Inorg. Chem.* 2014 (3) (2014) 469-474.
  21. D. Wang, H. Wang, H. Li, Novel luminescent soft materials of terpyridine-containing ionic liquids and europium(III), *ACS Appl Mater Interfaces* 5 (13) (2013) 6268-6275.
  22. M. Ichikawa, T. Yamamoto, H.-G. Jeon, K. Kase, S. Hayashi, M. Nagaoka, N. Yokoyama, Benzene substituted with bipyridine and terpyridine as electron-transporting materials for organic light-emitting devices, *J. Mater. Chem.* 22 (14) (2012) 6765.
  23. A. Et Taouil, J. Husson, L. Guyard, Synthesis and characterization of electrochromic [Ru(terpy)<sub>2</sub>selenophene]-based polymer film, *J. Electroanal. Chem.* 728 (2014) 81-85.
  24. A. Winter, U.S. Schubert, New Polyester-Based Terpyridine Macroligands and their Blue Iron(II) Complexes, *Macromol. Chem. Phys.* 208 (18) (2007) 1956-1964.
  25. C. Monnereau, J. Gomez, E. Blart, F. Odobel, S. Wallin, A. Fallberg, L. Hammarström, Photoinduced Electron Transfer in Platinum(II) Terpyridinyl Acetylide Complexes Connected to a Porphyrin Unit, *Inorganic Chemistry* 44 (13) (2005) 4806-4817.
  26. F. Camerel, R. Ziessel, B. Donnio, D. Guillon, Engineering of an iron-terpyridine complex with supramolecular gels and mesomorphic properties, *New J. Chem.* 30 (2) (2006) 135-139.
  27. H.A. Saadeh, E.A.A. Shairah, N. Charef, M.S. Mubarak, Synthesis and adsorption properties, toward some heavy metal ions, of a new polystyrene-based terpyridine polymer, *J. Appl. Polym. Sci.* 124 (4) (2012) 2717-2724.
  28. S. Köytepe, M.H. Demirel, A. Gültek, T. Seçkin, Metallo-supramolecular materials based on terpyridine-functionalized polyhedral silsesquioxane, *Polym. Int.* 63 (4) (2014) 778-787.
  29. A.O.T. Patrocínio, K.P.M. Frin, N.Y. Murakami Iha, Solid State Molecular Device Based on a Rhenium(I) Polypyridyl Complex Immobilized on TiO<sub>2</sub> Films, *Inorganic Chemistry* 52 (10) (2013) 5889-5896.
  30. R. Sakamoto, S. Katagiri, H. Maeda, H. Nishihara, Bis(terpyridine) metal complex wires: Excellent long-range electron transfer ability and controllable intrawire redox conduction on silicon electrode, *Coord. Chem. Rev.* 257 (9-10) (2013) 1493-1506.

31. R. Sakamoto, Y. Ohirabaru, R. Matsuoka, H. Maeda, S. Katagiri, H. Nishihara, Orthogonal bis(terpyridine)-Fe(II) metal complex oligomer wires on a tripodal scaffold: rapid electron transport, *Chem. Commun.* 49 (64) (2013) 7108-7110.
32. A. Winter, C. Friebe, M.D. Hager, U.S. Schubert, Advancing the Solid State Properties of Metallo-Supramolecular Materials: Poly( $\epsilon$ -caprolactone) Modified  $\pi$ -Conjugated Bis(terpyridine)s and their Zn(II) Based Metallo-Polymers, *Macromol. Rapid Commun.* 29 (20) (2008) 1679-1686.
33. D. Zhou, Q.-Y. Cheng, Y. Cui, T. Wang, X. Li, B.-H. Han, Graphene-terpyridine complex hybrid porous material for carbon dioxide adsorption, *Carbon* 66 (2014) 592-598.
34. G.T. Morgan, F.H. Burstall, 3. Dehydrogenation of pyridine by anhydrous ferric chloride, *J. Chem. Soc.* (0) (1932) 20-30.
35. K. Takada, R. Sakamoto, S.-T. Yi, S. Katagiri, T. Kambe, H. Nishihara, Electrochromic Bis(terpyridine)metal Complex Nanosheets, *J. Am. Chem. Soc.* 137 (14) (2015) 4681-4689.
36. E.C. Constable, *Modern Terpyridine Chemistry*. By Ulrich S. Schubert, Harald Hofmeier and George R. Newkome, *Angew. Chem. Int. Ed.* 46 (16) (2007) 2748-2749.
37. E.C. Constable, M.D. Ward, Synthesis and co-ordination behaviour of 6',6''-bis(2-pyridyl)-2,2':4,4'' : 2'',2'''-quaterpyridine; 'back-to-back' 2,2':6',2''-terpyridine, *Journal of the Chemical Society, Dalton Transactions* (4) (1990) 1405-1409.
38. P. Pechy, F.P. Rotzinger, M.K. Nazeeruddin, O. Kohle, S.M. Zakeeruddin, R. Humphry-Baker, M. Graetzel, Preparation of phosphonated polypyridyl ligands to anchor transition-metal complexes on oxide surfaces: application for the conversion of light to electricity with nanocrystalline TiO<sub>2</sub> films, *J. Chem. Soc., Chem. Commun.* (1) (1995) 65-66.
39. K.T. Potts, D. Konwar, Synthesis of 4'-vinyl-2,2':6',2''-terpyridine, *J. Org. Chem.* 56 (15) (1991) 4815-4816.
40. X. Guo, M. Myers, S. Xiao, M. Lefenfeld, R. Steiner, G.S. Tulevski, J. Tang, J. Baumert, F. Leibfarth, J.T. Yardley, M.L. Steigerwald, P. Kim, C. Nuckolls, Chemoresponsive monolayer transistors, *Proc. Natl. Acad. Sci. U S A.* 103 (31) (2006) 11452-11456.
41. N.O. Laschuk, I.I. Ebralidze, D. Spasyuk, O.V. Zenkina, Multi-readout logic gate for selective metal ions detection in parts per billion levels, Submitted (2016)
42. D. Chen, L. Cao, F. Huang, P. Imperia, Y.-B. Cheng, R.A. Caruso, Synthesis of Monodisperse Mesoporous Titania Beads with Controllable Diameter, High Surface Areas, and Variable Pore Diameters (14-23 nm), *J. Am. Chem. Soc.* 132 (12) (2010) 4438-4444.
43. A. Latini, C. Cavallo, F.K. Aldibaja, D. Gozzi, D. Carta, A. Corrias, L. Lazzarini, G. Salviati, Efficiency Improvement of DSSC Photoanode by Scandium Doping of Mesoporous Titania Beads, *J. Phys. Chem. C* 117 (48) (2013) 25276-25289.
44. S. Ito, P. Chen, P. Comte, M.K. Nazeeruddin, P. Liska, P. Péchy, M. Grätzel, Fabrication of screen-printing pastes from TiO<sub>2</sub> powders for dye-sensitised solar cells, *Prog. Photovoltaics: Res. Appl.* 15 (7) (2007) 603-612.
45. L. Lutterotti, Total pattern fitting for the combined size-strain-stress-texture determination in thin film diffraction, *Nuclear Inst. Methods in Phys. Res., B* 268 (3-4) (2010) 334-340.
46. C.D. Wagner, Sensitivity factors for XPS analysis of surface atoms, *J. Electron Spectrosc. Relat. Phenom.* 32 (2) (1983) 99-102.
47. G. Guerrero, J.G. Alauzun, M. Granier, D. Laurencin, P.H. Mutin, Phosphonate coupling molecules for the control of surface/interface properties and the synthesis of nanomaterials, *Dalton Trans* 42 (35) (2013) 12569-12585.
48. M. Becuwe, P. Rouge, C. Gervais, M. Courty, A. Dassonville-Klimpt, P. Sonnet, E. Baudrin, A new sensitive organic/inorganic hybrid material based on titanium oxide for the potentiometric detection of iron(III), *J. Colloid Interface Sci.* 388 (1) (2012) 130-136.
49. G.J. Ruiterkamp, M.A. Hempenius, H. Wormeester, G.J. Vancso, Surface functionalization of titanium dioxide nanoparticles with alkanephosphonic acids for transparent nanocomposites, *J. Nanopart. Res.* 13 (7) (2010) 2779-2790.

50. P. Thissen, M. Valtiner, G. Grundmeier, Stability of phosphonic acid self-assembled monolayers on amorphous and single-crystalline aluminum oxide surfaces in aqueous solution, *Langmuir* 26 (1) (2010) 156-164.
51. C. Tudisco, V. Oliveri, M. Cantarella, G. Vecchio, G.G. Condorelli, Cyclodextrin Anchoring on Magnetic Fe<sub>3</sub>O<sub>4</sub> Nanoparticles Modified with Phosphonic Linkers, *Eur. J. Inorg. Chem.* 2012 (32) (2012) 5323-5331.
52. H. Ma, O. Acton, D.O. Hutchins, N. Cernetic, A.K.Y. Jen, Multifunctional phosphonic acid self-assembled monolayers on metal oxides as dielectrics, interface modification layers and semiconductors for low-voltage high-performance organic field-effect transistors, *Phys. Chem. Chem. Phys.* 14 (41) (2012) 14110-14126.
53. P.J. Hotchkiss, S.C. Jones, S.A. Paniagua, A. Sharma, B. Kippelen, N.R. Armstrong, S.R. Marder, The Modification of Indium Tin Oxide with Phosphonic Acids: Mechanism of Binding, Tuning of Surface Properties, and Potential for Use in Organic Electronic Applications, *Accounts of Chemical Research* 45 (3) (2012) 337-346.
54. A. Vega, P. Thissen, Y.J. Chabal, Environment-controlled tethering by aggregation and growth of phosphonic acid monolayers on silicon oxide, *Langmuir* 28 (21) (2012) 8046-8051.
55. M. McDowell, I.G. Hill, J.E. McDermott, S.L. Bernasek, J. Schwartz, Improved organic thin-film transistor performance using novel self-assembled monolayers, *Appl. Phys. Lett.* 88 (7) (2006) 073505.
56. J.K. Klosterman, A. Linden, D.K. Frantz, J.S. Siegel, Manisyl-substituted polypyridine coordination compounds: Metallo-supramolecular networks of interdigitated double helices assembled via CH[three dots, centered][small pi] and [small pi]-[small pi] interactions, *Dalton Transactions* 39 (6) (2010) 1519-1531.
57. F. Guérin, Y. Tian, J.H. Fendler, Dihexadecyl Phosphate Monolayer Supported [Ru(bpy)<sub>3</sub>]<sup>2+</sup> Crystallites Investigated by Near-Field Scanning Optical Microscopy, *J. Phys. Chem. B* 103 (37) (1999) 7882-7888.
58. Standard Methods for the Examination of Water and Wastewater Standard Methods for the Examination of Water and Wastewater. 21 ed.; the American Public Health Association, the American Water Works Association and the Water Environment Federation: 2005. <http://www.standardmethods.org>.
59. R.L. Morris, Determination of iron in water in the presence of heavy metals, *Anal. Chem.* 24 (1952) 1376-1378.