



Astroglial atrophy in Alzheimer's disease

DOI:

[10.1007/s00424-019-02310-2](https://doi.org/10.1007/s00424-019-02310-2)

Document Version

Accepted author manuscript

[Link to publication record in Manchester Research Explorer](#)

Citation for published version (APA):

Verkh ratsky, A., Rodrigues, J. J., Pivoriunas, A., Zorec, R., & Semyanov, A. (2019). Astroglial atrophy in Alzheimer's disease. *Pflugers Archiv European Journal of Physiology*, 471(10), 1247-1261. <https://doi.org/10.1007/s00424-019-02310-2>

Published in:

Pflugers Archiv European Journal of Physiology

Citing this paper

Please note that where the full-text provided on Manchester Research Explorer is the Author Accepted Manuscript or Proof version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version.

General rights

Copyright and moral rights for the publications made accessible in the Research Explorer are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Takedown policy

If you believe that this document breaches copyright please refer to the University of Manchester's Takedown Procedures [<http://man.ac.uk/04Y6Bo>] or contact uml.scholarlycommunications@manchester.ac.uk providing relevant details, so we can investigate your claim.



Astroglial atrophy in Alzheimer's disease

Alexei Verkhratsky^{1,2}, Jose Julio Rodrigues^{3,4}, Augustas Pivoriunas⁵, Robert Zorec^{6,7} &
Alexey Semyanov^{8,9}

¹Faculty of Biology, Medicine and Health, The University of Manchester, Manchester, M13 9PT, UK;

²Achucarro Center for Neuroscience, IKERBASQUE, Basque Foundation for Science, 48011 Bilbao, Spain;

³BioCruces Health Research Institute, IKERBASQUE, Basque Foundation for Science 48011 Bilbao, Spain;

⁴Department of Neuroscience, The University of the Basque Country UPV/EHU, Plaza de Cruces 12, 48903, Barakaldo, Bizkaia, Spain;

⁵Department of Stem Cell Biology, State Research Institute Centre for Innovative Medicine, Santariškių street 5, LT-08406 Vilnius, Lithuania;

⁶University of Ljubljana, Medical Faculty, Institute of Pathophysiology, Laboratory of Neuroendocrinology and Molecular Cell Physiology, Zaloska cesta 4; SI-1000, Ljubljana, Slovenia;

⁷Celica, BIOMEDICAL, Technology Park 24, 1000 Ljubljana, Slovenia;

⁸Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Russian Academy of Sciences, Miklukho-Maklaya street 16/10, Moscow, 117997, Russia;

⁹Sechenov First Moscow State Medical University, Moscow, Russia

Send all correspondence to:

Alexei Verkhratsky

The University of Manchester,

Oxford Road,

Manchester, M13 9PT, UK,

Telephone +44 (0)161-2757324,

e-mail: Alexej.Verkhatsky@manchester.ac.uk

Abstract

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
Astrocytes, a class of morphologically and functionally diverse primary homeostatic neuroglia, are key keepers of neural tissue homeostasis and fundamental contributors to brain defence in pathological contexts. Failure of astroglial support and defence facilitate the evolution of neurological diseases, which often results in aberrant synaptic transmission, neurodegeneration, and death of neurones. In Alzheimer's disease (AD) astrocytes undergo complex and multifaceted metamorphoses ranging from atrophy with loss of function to reactive astrogliosis with hypertrophy. Astroglial asthenia underlies reduced homeostatic support and neuroprotection that may account for impaired synaptic transmission and neuronal demise. Reactive astrogliosis which mainly develops in astrocytes associated with senile plaque is prominent at the early to moderate stages of AD manifested by mild cognitive impairment; down-regulation of astrogliosis (reflecting astroglial paralysis) is associated with late stages of the disease characterised by severe dementia. Cell-specific therapies aimed at boosting astroglial supportive and defensive capabilities and preventing astroglial paralysis may offer new directions in preventing, arresting or even curing AD-linked neurodegeneration.

23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
Keywords: Astrocytes; Alzheimer's disease; Astroglial atrophy; Astrogliosis; Neurological diseases; Neurodegeneration

The epidemic of neurodegenerative diseases

The “epidemic” of neurodegenerative diseases prophesied by Robert Katzman in 1976 [63] spreads through the aging world population with little prospective for therapeutic containment. Despite remarkable progress in understanding the biochemistry and genetics of neurodegenerative processes the genesis and evolution of the majority of sporadic cases remain obscure, whereas pharmacological options remain symptomatic [122]. The ultimate outcome of neurodegeneration is neural cell death, brain atrophy and loss of brain function. A direct link between the decrease in the size (i.e., atrophy) of the brain tissue and decrease in cognitive capabilities (i.e., dementia) was suggested by Thomas de Willis at the end of 17th century [156]. Aberrant processing of proteins lies at the core of neurodegeneration; compromised synthesis/degradation or clearance of proteins results in accumulation of intra- or extracellular “toxic” proteins [55,118]. Despite the multitude of specific pathological pathways idiosyncratic for certain disease (e.g. β -amyloid accumulation and abnormal tau phosphorylation in Alzheimer’s disease (AD), α -synuclein accumulation in Parkinson’s disease or expression of mutant glial fibrillary acidic protein in Alexander disease) all neurodegenerative processes share a common pathological phenotype - they all trigger cell death and destroy connectivity in the neural networks.

Extracellular depositions of β -amyloid and intracellular accumulation of misphosphorylated tau protein (both processes are, most likely, interrelated with indications for tau pathology being driven by β -amyloid accumulation) are common histological denominators of the AD brains. Occurrence of these lesions, however, varies and there is no obvious correlation between their densities and the severity of dementia. The concept that tissue depositions of pathological material are causative for neurodegeneration was proposed by Oskar Fischer in 1907 [39,40]. The specific role for β -amyloid in the AD (the amyloid cascade hypothesis), remains, however, disputed [61,87,23]. Amyloid plaques occur in several neurological diseases; they were initially discovered by Paul Blocq and Gheorghe Marinescu in post-mortem brains from elderly patients with chronic epilepsy [11]; amyloid depositions populate posttraumatic nervous tissues, tissues infected with prions and brains affected by Parkinson’s or Huntington’s disease. Similarly, tau pathology is characteristic of fronto-temporal dementia and prion infection. In recent years the new concept of tau astrogliopathy had emerged, after the discovery of multiple pathological phenotypes of astrocytes infested with tau and related to specific forms of age-associated dementia [71]. Even acute sleep deprivation for a single night causes accumulation of β -amyloid, which is seemingly unrelated to any predisposition to AD [125].

It is, however, almost beyond dispute that the gross histopathological signs of AD became apparent at the late stages of the disease. The AD begins with prolonged (10 - 15 years) asymptomatic phase, when the overall cognitive function remains (almost) intact, although pathological changes begin to accumulate. It is most probable that from the very beginning AD-pathology affects synaptic transmission. There is a close correlation between synaptic alterations and cognitive impairments in AD patients, and these synaptic alterations are often considered to occur at the very early (pre-plaque) stages of the disease [31,135,55,89]. Nervous tissue, affected by AD is characterised by compromised synaptic connectivity and neuronal hyperexcitability, which are indicative of dyshomeostasis of ions and neurotransmitters [64,84,42,19]. The brain unwiring in AD is also manifested in white matter damage, which is

1 observed from the early stages and correlates with cognitive deficit [14]. Finally, the
2 AD alters metabolic homeostasis of the nervous system; region-specific
3 hypometabolism underlies AD-specific diagnostic phenotype used for FDG-PET
4 diagnostics [59]. All these features indicate that AD (similarly to other
5 neurodegenerative diseases) is a chronic homeostatic failure of the brain tissue,
6 which, naturally, has to be associated with the failure of the homeostatic neuroglia.
7

8 **Astrocytes provide homeostatic support and neuroprotection**

9

10 The human brain evolved for over ~500 million years from the diffuse nervous
11 system that appeared in the most primitive multicellular organisms. Evolution of
12 nervous system progressed through an increase in the complexity of nervous tissue
13 with a parallel increase in heterogeneity and specialisation of neural cells. Emergence
14 of the central nervous system (CNS) with intricate synaptic web required
15 sophisticated homeostatic support and thus much specialisation has occurred among
16 neural cells, which were fundamentally divided into neurones, representing the
17 executive arm and neuroglia, representing the housekeeping branch [111,143]. This
18 division of responsibilities reflected the perfection of fast neuronal signalling (action
19 potentials and synaptic transmission) that requires much energy and high level of
20 protein expression with an inevitable loss of neuronal abilities to control tissue
21 homeostasis. This task was transferred to neuroglia, which indeed is responsible for
22 each and every homeostatic cascade operating in the nervous system. Conceptually,
23 the term neuroglia covers cells responsible for homeostasis and defence of neural
24 tissue. Neuroglia include several types of heterogeneous (both morphologically and
25 functionally) cells of neural (astroglia, oligodendroglia, radial glia, NG2 cells,
26 peripheral glia and enteric glia) and non-neural (microglia) origin (for overview and
27 references see [67,139,48,144,147,66,129,69]).
28
29
30
31
32
33

34 The homeostasis of the CNS is mainly controlled by astrocytes, which are arguably
35 the most diverse glial cells in the brain and in the spinal cord (Fig. 1). Astrocytes are
36 defined as a class of neural cells, which sustain homeostasis and provide for
37 neuroprotection and defence of the CNS tissue [144]. Astroglial cells account
38 (depending on the brain region) for 20 - 40% of all neuroglial cells [140]. Astrocytes
39 demonstrate remarkable adaptive plasticity that defines the functional maintenance of
40 the CNS in development and ageing. Astrocytes maintain homeostasis of the CNS at
41 all levels of organisation, from molecular to organ. Astroglia contribute to ionostasis
42 of the CNS tissue by regulating fluxes of major ions; astrocytes control the turnover
43 of major neurotransmitters through dedicated systems responsible for their uptake and
44 for release of neurotransmitter precursors. Protoplasmic astrocytes divide (through the
45 process known as tiling) the grey matter into relatively independent “neurovascular
46 units”; the territorial domain of a single astrocyte (which overlaps in adult healthy
47 brain with neighbouring astrocytes only at the level of very distal processes)
48 integrates all neuronal elements and establishes direct link (through perivascular
49 process and the endfoot) with capillaries [57,56,20,94]. The morphology of astrocytic
50 processes defines their function. The processes can be classified into (i) astrocytic
51 branches of several orders that contain organelles including endoplasmic reticulum
52 Ca^{2+} stores, (ii) organelle-free perisynaptic leaflets that form astroglial cradle and (iii)
53 endfeet tiling the blood vessels [68,101,144]. Astrocytic branches are responsible for
54 amplification and propagation of Ca^{2+} signals within an astrocyte and possibly beyond
55 [123]. Astrocytic perisynaptic leaflets are rich in glutamate transporters as well as
56
57
58
59
60
61
62
63
64
65

1 other homeostatic transporters and selectively approach dendritic spines, providing
2 less coverage of presynaptic boutons and little coverage of dendritic shaft [44].
3 Endfeet intimately interact with cells of blood vessels supporting the blood-brain-
4 barrier [130]. Using perivascular processes and endfeet astrocytes adjust local blood
5 flow to the level of neuronal activity [90,165,131,38]. Astrocytes also act as an energy
6 source (being the main if not the only producer of glycogen in the CNS) by providing
7 neurones with lactate in an activity-dependent manner [104,80]. Furthermore,
8 astrocytes synthesize glutamate *de novo* and supply neurones with glutamine, which is
9 the main precursor for glutamate and GABA [54]. Astroglial leaflets cover central
10 synapses by a synaptic cradle and control neuronal excitability and synaptic
11 transmission in many ways from assisting synaptogenesis and synaptic maintenance
12 to regulating extracellular concentration of ions and major neurotransmitters (such as
13 glutamate, GABA monoamines and adenosine) [33,12,142,95,91]. Astrocytes are
14 central elements for cellular homeostasis of the CNS being responsible for embryonic
15 neurogenesis (which relies on radial glia) and for neurogenesis in the adult CNS (that
16 occurs from radial astrocytes of the neurogenic niches, [36,13]). Finally, astrocytes
17 are fundamental to brain defence through evolutionary conserved and complex
18 programmes controlling reactive astrogliosis launched in response to
19 polyaeathiological insults [103,128,151,102].
20
21
22
23

24 **Principles and classification of astroglipathology**

25
26
27 Neurological disorders are, in essence, the failures of homeostasis, and neuroglia,
28 being the homeostatic neural cells, are central to all types of neuropathologies.
29 Whatever the aetiology, neurological diseases are, to a great extent, pathologies of
30 neuroglia, the compromised function of which determines the survival or death of
31 neurones thus defining the progression and the outcome of neurological diseases. The
32 central role for neuroglia in neuropathology begun to be appreciated only recently
33 when numerous experimental finding questioned the neuronocentric views
34 [103,151,3,150,30,146,17,163,45,16,137,113].
35
36

37
38 The pathological potential of neuroglia was contemplated already by Rudolf Virchow
39 who prophesied that “This very interstitial tissue of the brain and spinal marrow (*i.e.*
40 *neuroglia - authors*) is one of the most frequent seats of morbid change” [153]. A
41 fundamental contribution of neuroglia to neuropathology was also considered by
42 Alois Alzheimer, Franz Nissl, Santiago Ramon-y-Cajal, Rio-Hortega and William
43 Lloyd Andriezen, the latter reflecting that astrocytes “exhibit a morbid hypertrophy in
44 pathological conditions” [2]. It is now the fact universally acknowledged that many
45 neurological diseases are associated with the astrocytic hypertrophy and activation of
46 microglia often referred to as reactive gliosis. For many years the reactive gliosis was
47 considered as a general non-specific pathological reaction often regarded as a basis
48 for neuroinflammation that is chiefly involved in exacerbating neuronal damage. This
49 oversimplification is, however, very much detached from reality. The main function
50 of neuroglia is the preservation of the nervous tissue, and as such multiple molecular
51 cascades expressed in glia are genuinely neuroprotective. Astrocytes, for example, are
52 primarily responsible for homeostasis of ions and neurotransmitters thus fencing
53 against excitotoxic damage. Oligodendrocytes maintain axonal survival and the death
54 of the former spells the imminent death on the latter. Microglia release multiple
55 trophic factors ensuring neuronal survival. The reactive changes instigated by a brain
56 lesion, represented by astrogliosis and microglial activation are therefore genuinely
57
58
59
60
61
62
63
64
65

1 survivalistic. These glial responses are complex and multistaged and are, as a rule,
 2 neuroprotective [103]. Being driven to an extreme by the severity of the insult, glial
 3 cells can assume the role of natural killer and contribute to the neural cell death, and
 4 yet even neurotoxicity of glia has, at its core, neuroprotective significance. Indeed,
 5 astroglial scar limits the area of damage and astrocytes and microglia surrounding the
 6 lesioned area through the releasing of neurotoxic factors rapidly exterminate
 7 pathologically affected cells thus contributing to the final clean-up (by phagocytic
 8 microglia and astrocytes [86,154]) and ultimate protection of the undamaged neural
 9 circuits lying outside of the damaged area. Finally, neuroglia is primarily responsible
 10 for the recovery of homeostasis in post-lesioned nervous tissue through promoting
 11 vascularization, reforming the blood-brain barrier, stimulating synaptogenesis and
 12 accomplishing re-myelination [128,151,1,18]. Conceptually, disruption of glial
 13 protection is fatal for the nervous system. All in all, neurones cannot function
 14 correctly or survive in the absence of glia, whereas glial cells can survive and operate
 15 in the presence of dead or dying neurones.
 16
 17
 18

19 Pathological metamorphoses of astrocytes are complex, disease- and disease-stage
 20 specific and may change substantially in the course of pathological evolution.
 21 Conceptually, astrogliopathological phenotypes (see Glossary) are classified into
 22 three major groups: (i) reactive astrogliosis (ii) astrodegeneration with astroglial
 23 atrophy and loss of function and (iii) pathological remodelling [103,151]. The latter
 24 two groups of the non-reactive pathological transformation of astrocytes can be
 25 summararily identified as astrocytopathies to distinguish from reactive astrogliosis [37].
 26 Astrocytes in the CNS are organised in the forms of syncytia confined to specific
 27 anatomical structures [46,47]. Astroglial syncytia are formed by intercellular gap
 28 junctions, which are composed from connexons (astrocytes express connexins 43, 30
 29 and 26; with predominant presence of Cx43) permeable for ions and small (<1000
 30 Da) molecules. Gap junctions participate in regulation of astrocyte electric properties
 31 [79], redistribution of K^+ (“spatial K^+ buffering” - [155]), diffusional transport of
 32 glucose [116] and propagation of Ca^{2+} waves [123,148]. Pathological changes in
 33 astrocytes frequently affect gap junctional connectivity within astroglial networks;
 34 which may define various aspects of neuropathological progression, for example, in
 35 epilepsy [9,106] or neurodegeneration [162].
 36
 37
 38
 39
 40

41 *****

42 **Glossary of astrogliopathology:**

43 **Reactive astrogliosis, reactive astrocytes:**

44 There is no universally agreed definition of the reactive astrogliosis [34]. We define
 45 reactive astrogliosis as an evolutionary conserved defensive response of astrocytes to
 46 pathological lesions caused by endo- or exogeneous agents. Reactive astrogliosis
 47 leads to substantial changes in gene expression, resulting in a remodelling of
 48 astroglial morphology, biochemistry, and function, thus producing reactive astrocytes
 49 with multiple phenotypes. These reactive phenotypes are disease- and context- (for
 50 example ageing) specific, while reactive astrocytes could be either neuroprotective or
 51 neurotoxic, again depending on the pathology and severity of lesion. Morphology of
 52 reactive astrocytes is characterised by hypertrophy of somata and primary processes
 53 and significant up-regulation of intermediate filament proteins GFAP and vimentin
 54 (see [34,102,103,151] for further details).
 55
 56
 57
 58
 59
 60
 61
 62
 63
 64
 65

Astroglial atrophy:

Defined as decrease in surface area and volume of astroglial morphological profiles; astroglial atrophy is manifested in specific diminution of peripheral and perisynaptic processes, which decrease synaptic coverage and synaptic homeostatic support. Astroglial atrophy has been reported in stress, depression, Alzheimer's disease and epilepsy [106,164,152,93].

Loss of function:

Decrease in expression or activity of astroglial homeostatic molecules or pathways (for example decrease in glutamate transporters activity), which reduce glial homeostatic support this instigating or exacerbating neuropathology.

Astroglial pathological remodelling:

Development of specific astroglial phenotype, which drives neuropathology. Examples are expression of mutant sporadically mutated GFAP in Alexander disease, which affect development of white matter and causes severe leicomalacia [82] or changes in glutamate/ammonium handling in hepatic encephalopathy, which affects neurotransmission and neuronal excitability thus causing psychotic symptoms and ultimately brain oedema [146].

Reactive astrogliosis is the most characterised response of astrocytes to pathological lesions [102]. Reactive astrogliosis can be defined as an evolutionary conserved defensive reprogramming of astroglia aimed at: (i) increased neuroprotection and trophic support of nervous tissue; (ii) isolation of the lesioned area; (iii) reconstruction of the damaged blood-brain barrier; and (iv) providing for post-lesion regeneration of brain circuits [102,103,128]. Astroglial reactivity is heterogeneous and disease-specific; reactive phenotypes may demonstrate both neuroprotective and neurotoxic features [76], although generally suppression of astroglial response exacerbates neuropathology [103].

Pathological remodelling of astrocytes reflects the emergence of abnormal astroglial phenotypes which either cause or drive neuropathological changes. Examples of pathological remodelling of astrocytes include for example leukodystrophies, such as Alexander disease, megalencephalic leukoencephalopathy with subcortical cysts or vanishing white matter syndrome. In all these pathologies pathologically modified astrocytes trigger white matter lesions [74]. Pathological remodelling of astrocytes was suggested as a contributing factor to mesial temporal lobe epilepsy. In this condition astrocytes acquire aberrant morphology, decrease gap junctional coupling and reduce K^+ buffering capability [9]. Yet another example of pathological remodelling of astrocytes is observed following infection with *Toxoplasma gondii*. Infiltration of astrocytes with *T. gondii* results in abnormal elevation of synthesis and release kynurenic acid that, through inhibition of NMDA and acetylcholine receptors, affects neurotransmission which may be linked to an increased risk of schizophrenia [120]. In AD astrocytes start to express GAD67 that also affects the balance of excitation and inhibition in the neuronal network [43,157].

1 Astrodegeneration appears as a decrease in astroglial density, is often accompanied by
2 morphological atrophy, and invariably is associated with a loss of function, i.e. in
3 decreased homeostatic, supportive and neuroprotective capabilities, which all
4 constitute astroglial asthenia. Astroglial atrophy and asthenia are observed in a wide
5 range of neurological disorders including neuropsychiatric diseases, addictive
6 disorders, epilepsy and neurodegeneration. Reduction of astroglial numbers, astroglial
7 morphological atrophy with decreased expression of GFAP as well as deficient
8 glutamate uptake are detected in major neuropsychiatric diseases such as bipolar
9 disease, major depression and schizophrenia [28,29,83,92,108-110,149,119].
10 Deficient astroglial support leads to abnormal neurotransmission and oxidative or
11 excitotoxic stress which contribute to psychotic symptomatology. Similar
12 morphological atrophy of astrocytes associated with decreased expression of
13 glutamate transporters and deficient glutamate uptake were identified in the nucleus
14 accumbens of cocaine-addicted rats [121]. Significant atrophy of astrocytes which
15 may underlie aberrant K⁺ buffering and glutamate homeostasis has been detected in
16 experimental epilepsy [106]. Finally, as will be discussed below, astroglial atrophy is
17 often seen in various neurodegenerative disorders.

21 Astrocytic atrophy may have a complex nature and affect astrocytic branches, leaflets
22 and endfeet differently. Since these astrocytic processes have distinct functions, the
23 effect on the neuronal network will be different. Atrophy of branches arguably affects
24 astroglial Ca²⁺ signalling possibly inducing further morphological changes [132].
25 Perisynaptic leaflets are highly plastic and their changes accompany (or even drive)
26 synaptic plasticity [105]. Hence, changes in perisynaptic leaflets can potentially
27 contribute to pathological processes. Finally, atrophy of endfeet can potentially
28 damage the neuron-glia-vascular unit, and affect the blood-brain barrier thus
29 promoting neurodegeneration [126,133,78].

34 **Morphological atrophy and functional impairment of astrocytes in** 35 **neurodegenerative disorders**

37 Degeneration of astroglia resulting in astroglial atrophy or death has been recently
38 documented for several classes of neurological diseases associated with disruption in
39 neural connectivity including excitotoxic neurodegeneration (Wernicke
40 encephalopathy) chronic neurodegeneration (amyotrophic lateral sclerosis and
41 Alzheimer's disease, thalamic dementia, fronto-temporal dementia) and psychiatric
42 disorders such as schizophrenia, bipolar disorder and major depression, which all
43 affect cognition. These dystrophic changes in astroglia often precede neurological
44 symptoms or are key pathogenic factors.

48 In acute excitotoxic neurodegeneration of Wernicke type massive neuronal death
49 results from functional astroglial degeneration manifested by down-regulation of
50 expression of astroglial glutamate transporters. This compromises the ability of
51 astrocytes to remove the excess of glutamate and to regulate glutamatergic
52 transmission, which in turn results in severe excitotoxicity that underlies rapid
53 development of severe dementia with prominent psychotic components [51,52].

57 In amyotrophic lateral sclerosis (ALS) astroglial degeneration precedes the
58 development of neuronal death and clinical symptoms in transgenic mice model of the
59 disease, in which cells express ALS-associated mutant human gene for superoxide
60
61
62
63
64
65

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

dismutase 1 (hSOD1^{G93A}). At the later stage of the disease neuronal death triggers reactive astrogliosis and yet atrophic astrocytes remain in the tissue [114,115,160,4]. Astrocytes selectively expressing SOD1 gene acquire vulnerability to glutamate excitotoxicity that may underlie their early degeneration. These degenerated astrocytes lose their ability to effectively control glutamate homeostasis through down-regulation of glutamate transporters expression that further exacerbates excitotoxicity and contributes to neuronal death [115]. Critically, specific silencing of SOD1 gene in astrocytes delays the progression of ALS symptoms in the mouse model [159].

Dystrophic changes in astrocytes have been observed in several types of neurodegenerative pathologies including fronto-temporal dementia and Pick's disease, some studies mentioned the direct correlation between the degree of glial atrophy and the severity of dementia [15,65]. In Huntington disease functional astrodegeneration is manifested by a decrease in expression of glutamate transporters and hence in exacerbated glutamate excitotoxicity [35]; similarly, astrocytes demonstrate deficits in K⁺ buffering [136]. In addition astrocytes in HD reduce production and release of glutathione and ascorbic acid that act as major scavengers of reactive oxygen species (ROS) in CNS; this further reduces astroglial neuroprotection.

Astroglial atrophy in Alzheimer's disease

Astroglial changes in AD are highly heterogeneous in different brain regions and are represented by both astrogliosis with astroglial hypertrophy and astroglial atrophy (Table 1 and [3,141,145]). When analysing longitudinal changes in expression of GFAP, a classic marker for astrogliosis, in a triple transgenic mouse model of AD a decrease in the morphological presence of GFAP profiles and an overall decrease in GFAP expression was observed in early pre-plaque stages of the disease (Fig. 3 and [73,97,98,161]). This decrease in GFAP-positive astroglial profiles appear very early (at ~ 1 month of age) in the entorhinal cortex, somewhat later (~ 3 months of age) in the prefrontal cortex and even later (~ 9 - 12 months) in the hippocampus. Morphological atrophy of astrocytes was also confirmed when analysing profiles labelled with antibodies against glutamine synthetase and protein s100b [98,161]; this staining reveals full extent of the cells (in contrast to GFAP which stains only primary processes). Emergence of senile plaques triggers astrogliosis, which, however, similarly differs between brain regions. In hippocampus β -amyloid depositions and β -amyloid plaques are surrounded by hypertrophic astrocytes [97], whereas little, if any, signs of astrogliosis were found in entorhinal and prefrontal cortices [73,161]. The GFAP-hypertrophic astrocytes in hippocampus were associated with β -amyloid depositions/plaques, while distantly to the plaques GFAP profiles remain atrophic [97]. Similar astroglial atrophy has been characterised in other models of AD including PDAPP-J20 transgenic mice, 5xTG-AD mice and Swiss 3 mouse AD model (Table 1, [58,107,32,8]). Astroglial atrophy was found not only in experimental model animals but in post-mortem tissues of patients with advanced (Braak V-VI) stages of the disease (Rodriguez and Verkhatsky, personal observations). Similarly, morphological atrophy was observed in astrocytes derived from induced pluripotent stem cells obtained from patients with both familial and sporadic forms of AD (Fig. 4; [60]). Enriched environment as well as physical activity prevents astroglial atrophy in an AD mouse model and ameliorates the symptoms, confirming the causal link between astrocyte remodelling and dementia [7,112].

1 It has to be noted, however, that animal models of AD only partially reproduce the
2 human disease. For example, while 3xTG-AD mouse develops both plaque and tangle
3 pathologies accompanied by some cognitive deficits, there is no massive neuronal loss
4 which is a hallmark of AD progression in patients [10]. In this regard, new stem cell
5 technologies that enable the generation of astrocytes from the disease-specific
6 inducible pluripotent stem cells (iPSCs) [117], or by direct reprogramming of patient-
7 specific fibroblasts [21] present a great promise. Although the field is still in its
8 infancy, current technologies allow generation of relatively pure populations of cells
9 exhibiting molecular and functional properties (electrophysiological signatures,
10 generation of spontaneous calcium signals, glutamate uptake, support of synapse
11 formation, etc.) similar to those of adult human astrocytes. The disease-specific
12 astrocytes also recapitulate some important pathological aspects of AD. Thus,
13 astrocytes derived from the familial AD and sporadic AD patients exhibited a less
14 complex morphological appearance, decreased heterogeneity, overall atrophic profiles
15 and abnormal localisation of key functional astroglial markers resembling changes
16 occurring during glial paralysis at the early stages of the disease [60]. Astrocytes
17 derived from familial AD patients also demonstrated increased β -amyloid production,
18 deregulated Ca^{2+} homeostasis, altered cytokine release, increased ROS, decreased
19 lactate production and compromised neuroprotection and neuronal support [85,96].
20 Astrocytes differentiated from isogenic APOE4 astrocytes were impaired in their
21 ability to clear extracellular β -amyloid and displayed aberrant cholesterol
22 accumulation [77]. A new human AD “triculture” model using neurones, astrocytes,
23 and microglia in a 3D microfluidic platform has been successfully used to study
24 neuroinflammatory responses [100]. These studies demonstrate the substantial
25 potential of iPSC-based human astrocyte models to reveal the cellular mechanisms of
26 AD. Undoubtedly, these technologies will improve our understanding of the
27 molecular mechanisms controlling astroglial response and also help to develop
28 astrocyte-specific therapies against AD.

36 **Reactive astrogliosis in AD: does it always signal neuroinflammation?**

37
38
39 Reactive astrogliosis is another hallmark of AD; increased expression of GFAP,
40 vimentin or S100B protein as well as astroglial hypertrophy has been observed in
41 post-mortem tissues from AD patients [6,49,81,88] as well as in brain samples of AD
42 animal models [97,145,62,99]. In AD brains reactive astrocytes are mainly associated
43 with senile plaques [72,97,99]. The idea that chronic neuroinflammation is directly
44 responsible for the progression of idiopathic AD from the very early stages, and may
45 even have an etiological significance, become quite popular in recent years [53]. It is
46 also generally assumed that this chronic neuroinflammation results from activation of
47 microglia and reactive astrogliosis. This statement however, needs clarification, as
48 indeed reactive gliosis is far from being a straight pathological reaction ultimately
49 resulting in damage to the brain tissue. The reactions of neuroglia to brain lesion
50 (reflected by both reactive astrogliosis and activation of microglia) essentially
51 represent a defensive response aimed at counteracting pathology and remodelling
52 post-lesioned circuitry. Neither astrogliosis, nor microglial activation develops in all-
53 or-none fashion; to the contrary they represent a continuum of phenotypic
54 remodelling, fundamentally associated with neuroprotection. There are many stages
55 and degrees in astrogliosis [103,127], and some of these are fully reversible. Limited
56 brain lesions trigger astroglial hypertrophy and biochemical remodelling without
57
58
59
60
61
62
63
64
65

1 affecting microdomain organisation (anisomorphic astrogliosis); and only severe
2 lesions disrupt astroglial territorial maps, arrange astrocytes in palisades and trigger
3 scar formation [127,128]. Even the scar formation is in essence defensive and
4 survivalistic reaction aimed at isolation of lesioned area from the healthy tissue.
5 Likewise, activation of microglia is a multistage process with a multitude of activated
6 phenotypes, many of which have a neuroprotective role [50,66]. In AD, both reactive
7 astrogliosis and activation of microglia are directly associated with plaque formation;
8 activated glial cells become elements of plaques. Importantly, activated astrocytes
9 surrounding the plaques retain their domain organisation and do not express severe
10 astroglial features; similarly, microglial cells do not turn in all-devouring
11 macrophages, but retain intermediate activated phenotype. Suppression of astroglial
12 reactivity, contrary to the inflammatory hypothesis, exacerbates β -amyloid load and
13 reduces neuroprotection [72].
14
15

16 **Astroglial atrophy contributes to synaptic dysfunction and cognitive deficits**

17
18
19 Atrophic changes in astrocytes, characterised in several AD animal models as well as
20 in stem-cells derived astrocytes appear as shrinkage of astroglial territories, with a
21 decrease in astroglial coverage of synaptic contacts and other neuronal structures with
22 the ultimate decline in astroglial homeostatic support. Astroglial atrophy and loss of
23 function may contribute to early cognitive deficits through dwindling synaptic support
24 and synaptic malfunction. Decreased astroglial synaptic coverage may also result in
25 neurotransmitter spillover with subsequent hyperexcitability of neuronal networks
26 often observed in neurodegeneration. In addition enhanced glutamate spillover may
27 increase the recruitment of extrasynaptic NR2B subunit-containing NMDA receptors
28 associated with long-term depression, LTD [5]. Thus decreased astroglial synaptic
29 coverage with a consequent reduction in astroglial glutamate uptake may impair upon
30 synaptic plasticity shifting it towards depression, which can in turn negatively affect
31 memory [138,27].
32
33
34
35

36
37 Notably, K^+ clearance and glutamate uptake by astrocytes are tightly linked [75].
38 During synaptic transmission most of K^+ entering the synaptic cleft is released
39 through postsynaptic AMPA and NMDA receptors [24]. Accumulation of K^+ in the
40 synaptic cleft depolarises presynaptic terminal causing activity-dependent facilitation
41 of glutamate release [124,26]. Reduction of glutamate uptake can enhance recruitment
42 of postsynaptic AMPA/NMDA receptors and causes further K^+ release. This
43 instigates positive feedback, which can potentially lead to an uncontrolled increase in
44 excitability of the synaptic network, excitotoxicity and neuronal death. In physiological
45 conditions, these processes are tightly controlled by astroglial glutamate uptake and
46 K^+ clearance. However, when glutamate uptake is impaired (due to the reduction of
47 transporter expression or withdrawal of perisynaptic leaflets) the resulting vicious
48 circle may contribute to neurodegeneration.
49
50
51

52
53 Furthermore limited astroglial support may instigate early extinction of synapses
54 [166]. Dysfunctional synaptic transmission and loss of synapses are indeed the very
55 first morphological changes in AD, which mount years before the occurrence of
56 specific clinical presentation [134,25,70]. Astroglial asthenia may also suppress
57 regenerative synaptogenesis; while the loss of astroglial transporters may contribute to
58 excitotoxicity through impaired glutamate and K^+ buffering [166,152].
59
60
61
62
63
64
65

Failure of astroglial reactivity paves the way to dementia?

As has been alluded above, astrogliosis represents a powerful defensive program, which arguably contains various pathological processes including β -amyloid pathology and AD. Indeed there is compelling evidence demonstrating the neuroprotective potential of reactive astrocytes in AD. In the Tg2576 mice model (that bears APPSwe mutation) astrogliosis became prominent rather early and this correlates with the relatively slow development of AD. Furthermore, senile plaques in these animals resemble human β -amyloid deposits being represented by fleecy, granular, cored and diffused amyloid plaques [158]. In 3xTg-AD animals reactive astrocytes are positioned around senile plaques and close to perivascular β -amyloid deposits [97,98]. Conversely, in entorhinal and prefrontal cortices, the emergence of extracellular β -amyloid deposits does not initiate astrogliosis [73,161] which may reflect a failure of astroglial neuroprotection. This coincides with (and arguably underlines) high vulnerability of both regions to AD pathology; suggesting that paralysis of astroglial defence exacerbates AD-like pathology [141].

This notion also has been indirectly confirmed by the *in vivo* brain imaging of reactive astroglia in AD patients. Astrogliosis was assessed by positron-emission tomography detection of ^{11}C -deuterium-L-deprenyl (^{11}C -DED); deprenyl is a specific inhibitor of monoaminoxidase-B (MAO-B) localised predominantly in astrocytes. An increase in ^{11}C -DED signal hence is considered to reflect astroglial hypertrophy [41]. When using a multi-tracer PET detecting ^{11}C -PIB (marker of fibrillar β -amyloid), ^{18}F -FDG (marker of cerebral glucose metabolism) and ^{11}C -DED (marker of astrogliosis) the highest binding of ^{11}C -DED (which reflects prominent astrogliosis) was observed in patients with mild cognitive impairment (MCI) and high levels of fibrillar amyloid plaques in the brain (PIB+) reflecting prodromal AD [22]. The decrease in astroglial reactivity parallels the switch from MCI to full blown AD with senile dementia again demonstrating the neuroprotective role of astroglial remodelling [141].

Conclusions: the need for new astrocentric therapies?

In recent decades astrocytes emerged as a fundamental elements in pathophysiology of numerous neurological and neuropsychiatric diseases. Pathomorphological examination of astroglia has become a standard in describing the histology of the diseased brain. Pathophysiology of astroglia is a complex and multifactorial combination of degenerative and reactive remodelling, which can support neuroprotection or project neurotoxicity. Failure of astrocytes to support homeostasis of neural tissue and to protect this tissue against insults is arguably critical for determining the pathological evolution and ultimately neurological deficits. In the context of Alzheimer's disease astrocytes undergo atrophy with loss of function which may stipulate impairments of synaptic connectivity as well as contribute to neuronal death due to deficient neuroprotective support. At the same time, reactive astrocytes surround senile plaques in the AD brains; while initial stages of the disease are characterised by prominent astrogliosis. Exhaustion of astroglial defensive capacities and down-regulation of astrogliosis coincides with (and again may be instrumental for) the switch from mild cognitive impairment (characteristic for early to moderate AD stages) to senile dementia (which reflects late stages of the disease). Cell-specific therapies aimed at boosting astroglial supportive and defensive

capabilities and preventing astroglial paralysis may offer new directions in preventing, arresting or even curing AD-linked neurodegeneration.

Acknowledgments

AV and AS were supported by Volkswagen Stiftung research grant A115105; RZ by Slovenian Research Agency Research core funding #P3 310 and projects # J3 6790, # J3 9266 and # J3 7605. AS was supported by and COMFI grant 17-00-00412 (K) from RFBR; AV and AP were supported by the Global Grant measure (No. 09.3.3-LMT-K-712-01-0082).

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

References

1. Anderson MA, Burda JE, Ren Y, Ao Y, O'Shea TM, Kawaguchi R, Coppola G, Khakh BS, Deming TJ, Sofroniew MV (2016) Astrocyte scar formation aids central nervous system axon regeneration. *Nature* 532:195-200. doi:10.1038/nature17623
2. Andriezen WL (1893) The neuroglia elements of the brain. *Br Med J* 2:227-230
3. Arranz AM, De Strooper B (2019) The role of astroglia in Alzheimer's disease: pathophysiology and clinical implications. *Lancet Neurol* 18:406-414. doi:10.1016/S1474-4422(18)30490-3
4. Barbeito L (2018) Astrocyte-based cell therapy: new hope for amyotrophic lateral sclerosis patients? *Stem Cell Res Ther* 9:241. doi:10.1186/s13287-018-1006-y
5. Bartlett TE, Bannister NJ, Collett VJ, Dargan SL, Massey PV, Bortolotto ZA, Fitzjohn SM, Bashir ZI, Collingridge GL, Lodge D (2007) Differential roles of NR2A and NR2B-containing NMDA receptors in LTP and LTD in the CA1 region of two-week old rat hippocampus. *Neuropharmacology* 52:60-70. doi:10.1016/j.neuropharm.2006.07.013
6. Beach TG, McGeer EG (1988) Lamina-specific arrangement of astrocytic gliosis and senile plaques in Alzheimer's disease visual cortex. *Brain Res* 463:357-361
7. Beauquis J, Pavia P, Pomilio C, Vinuesa A, Podlutskaya N, Galvan V, Saravia F (2013) Environmental enrichment prevents astroglial pathological changes in the hippocampus of APP transgenic mice, model of Alzheimer's disease. *Exp Neurol* 239:28-37. doi:10.1016/j.expneurol.2012.09.009
8. Beauquis J, Vinuesa A, Pomilio C, Pavia P, Galvan V, Saravia F (2014) Neuronal and glial alterations, increased anxiety, and cognitive impairment before hippocampal amyloid deposition in PDAPP mice, model of Alzheimer's disease. *Hippocampus* 24:257-269. doi:10.1002/hipo.22219
9. Bedner P, Dupper A, Huttmann K, Muller J, Herde MK, Dublin P, Deshpande T, Schramm J, Haussler U, Haas CA, Henneberger C, Theis M, Steinhauser C (2015) Astrocyte uncoupling as a cause of human temporal lobe epilepsy. *Brain* 138:1208-1222. doi:10.1093/brain/awv067
10. Bilkei-Gorzo A (2014) Genetic mouse models of brain ageing and Alzheimer's disease. *Pharmacol Ther* 142:244-257. doi:10.1016/j.pharmthera.2013.12.009
11. Blocq P, Marinesco G (1892) Sur les lésions et la pathogenie de l'épilepsie dite essentielle. *La Semaine Medicale* 12:445-446
12. Boison D, Chen JF, Fredholm BB (2010) Adenosine signaling and function in glial cells. *Cell Death Differ* 17:1071-1082. doi:10.1038/cdd.2009.131
13. Bonzano S, Crisci I, Podlesny-Drabiniok A, Rolando C, Krezel W, Studer M, De Marchis S (2018) Neuron-Astroglia Cell Fate Decision in the Adult Mouse Hippocampal Neurogenic Niche Is Cell-Intrinsically Controlled by COUP-TFI In Vivo. *Cell Rep* 24:329-341. doi:10.1016/j.celrep.2018.06.044
14. Bozzali M, Padovani A, Caltagirone C, Borroni B (2011) Regional grey matter loss and brain disconnection across Alzheimer disease evolution. *Curr Med Chem* 18:2452-2458. doi:BSP/CMC/E-Pub/2011/175
15. Broe M, Kril J, Halliday GM (2004) Astrocytic degeneration relates to the severity of disease in frontotemporal dementia. *Brain* 127:2214-2220
16. Burda JE, Bernstein AM, Sofroniew MV (2016) Astrocyte roles in traumatic brain injury. *Exp Neurol* 275 Pt 3:305-315. doi:10.1016/j.expneurol.2015.03.020

- 1 17. Burda JE, Sofroniew MV (2014) Reactive gliosis and the multicellular response to
2 CNS damage and disease. *Neuron* 81:229-248.
3 doi:10.1016/j.neuron.2013.12.034
- 4 18. Burda JE, Sofroniew MV (2017) Seducing astrocytes to the dark side. *Cell Res*
5 27:726-727. doi:10.1038/cr.2017.37
- 6 19. Busche MA, Grienberger C, Keskin AD, Song B, Neumann U, Staufenbiel M,
7 Forstl H, Konnerth A (2015) Decreased amyloid- β and increased neuronal
8 hyperactivity by immunotherapy in Alzheimer's models. *Nat Neurosci*
9 18:1725-1727. doi:10.1038/nn.4163
- 10 20. Bushong EA, Martone ME, Jones YZ, Ellisman MH (2002) Protoplasmic
11 astrocytes in CA1 stratum radiatum occupy separate anatomical domains. *J*
12 *Neurosci* 22:183-192. doi:22/1/183
- 13 21. Caiazzo M, Giannelli S, Valente P, Lignani G, Carissimo A, Sessa A, Colasante
14 G, Bartolomeo R, Massimino L, Ferroni S, Settembre C, Benfenati F, Broccoli
15 V (2015) Direct conversion of fibroblasts into functional astrocytes by defined
16 transcription factors. *Stem Cell Reports* 4:25-36.
17 doi:10.1016/j.stemcr.2014.12.002
- 18 22. Carter SF, Scholl M, Almkvist O, Wall A, Engler H, Langstrom B, Nordberg A
19 (2012) Evidence for astrocytosis in prodromal Alzheimer disease provided by
20 ^{11}C -deuterium-L-deprenyl: a multitracer PET paradigm combining ^{11}C -
21 Pittsburgh compound B and ^{18}F -FDG. *J Nucl Med* 53:37-46.
22 doi:10.2967/jnumed.110.087031
- 23 23. Castellani RJ, Smith MA (2011) Compounding artefacts with uncertainty, and an
24 amyloid cascade hypothesis that is 'too big to fail'. *J Pathol* 224:147-152.
25 doi:10.1002/path.2885
- 26 24. Cheung G, Sibille J, Zapata J, Rouach N (2015) Activity-Dependent Plasticity of
27 Astroglial Potassium and Glutamate Clearance. *Neural Plast* 2015:109106.
28 doi:10.1155/2015/109106
- 29 25. Coleman P, Federoff H, Kurlan R (2004) A focus on the synapse for
30 neuroprotection in Alzheimer disease and other dementias. *Neurology*
31 63:1155-1162
- 32 26. Contini D, Price SD, Art JJ (2017) Accumulation of K^+ in the synaptic cleft
33 modulates activity by influencing both vestibular hair cell and calyx afferent
34 in the turtle. *J Physiol* 595:777-803. doi:10.1113/JP273060
- 35 27. Cui Z, Feng R, Jacobs S, Duan Y, Wang H, Cao X, Tsien JZ (2013) Increased
36 NR2A:NR2B ratio compresses long-term depression range and constrains
37 long-term memory. *Sci Rep* 3:1036. doi:10.1038/srep01036
- 38 28. Czeh B, Di Benedetto B (2013) Antidepressants act directly on astrocytes:
39 evidences and functional consequences. *Eur Neuropsychopharmacol* 23:171-
40 185. doi:10.1016/j.euroneuro.2012.04.017
- 41 29. Czeh B, Nagy SA (2018) Clinical Findings Documenting Cellular and Molecular
42 Abnormalities of Glia in Depressive Disorders. *Front Mol Neurosci* 11:56.
43 doi:10.3389/fnmol.2018.00056
- 44 30. De Keyser J, Mostert JP, Koch MW (2008) Dysfunctional astrocytes as key
45 players in the pathogenesis of central nervous system disorders. *J Neurol Sci*
46 267:3-16. doi:10.1016/j.jns.2007.08.044
- 47 31. DeKosky ST, Scheff SW (1990) Synapse loss in frontal cortex biopsies in
48 Alzheimer's disease: correlation with cognitive severity. *Ann Neurol* 27:457-
49 464. doi:10.1002/ana.410270502

- 1 32. Diniz LP, Tortelli V, Matias I, Morgado J, Bergamo Araujo AP, Melo HM, Seixas
2 da Silva GS, Alves-Leon SV, de Souza JM, Ferreira ST, De Felice FG, Gomes
3 FCA (2017) Astrocyte Transforming Growth Factor β 1 Protects Synapses
4 against Abeta Oligomers in Alzheimer's Disease Model. *J Neurosci* 37:6797-
5 6809. doi:10.1523/JNEUROSCI.3351-16.2017
- 6 33. Eroglu C, Barres BA (2010) Regulation of synaptic connectivity by glia. *Nature*
7 468:223-231. doi:10.1038/nature09612
- 8 34. Escartin C, Guillemaud O, Carrillo-de Sauvage M-A (2019) Questions and (some)
9 answers on reactive astrocytes. *Glia* E-pub ahead of print.
10 doi:10.1002/glia.23687
- 11 35. Estrada-Sánchez AM, Rebec GV (2012) Corticostriatal dysfunction and glutamate
12 transporter 1 (GLT1) in Huntington's disease: Interactions between neurons
13 and astrocytes. *Basal Ganglia* 2:57-66
- 14 36. Falk S, Gotz M (2017) Glial control of neurogenesis. *Curr Opin Neurobiol*
15 47:188-195. doi:10.1016/j.conb.2017.10.025
- 16 37. Ferrer I (2017) Diversity of astroglial responses across human neurodegenerative
17 disorders and brain aging. *Brain Pathol* 27:645-674. doi:10.1111/bpa.12538
- 18 38. Filosa JA, Morrison HW, Iddings JA, Du W, Kim KJ (2016) Beyond
19 neurovascular coupling, role of astrocytes in the regulation of vascular tone.
20 *Neuroscience* 323:96-109. doi:10.1016/j.neuroscience.2015.03.064
- 21 39. Fischer O (1907) Miliare Nekrosen mit drusigen Wucherungen der Neurofibrillen,
22 eine regelmässige Veränderung der Hirnrinde bei seniler Demenz. *Monatsschr*
23 *Psychiatr Neurol* 22:361-372
- 24 40. Fischer O (1910) Die presbyofrene Demenz, deren anatomischen Grundlage und
25 klinische Abgrenzung. *Z Ges Neurol Psychiatr* 3:371-471
- 26 41. Fowler JS, Volkow ND, Wang GJ, Logan J, Pappas N, Shea C, MacGregor R
27 (1997) Age-related increases in brain monoamine oxidase B in living healthy
28 human subjects. *Neurobiol Aging* 18:431-435
- 29 42. Frazzini V, Guarnieri S, Bomba M, Navarra R, Morabito C, Mariggio MA, Sensi
30 SL (2016) Altered $K_v2.1$ functioning promotes increased excitability in
31 hippocampal neurons of an Alzheimer's disease mouse model. *Cell Death Dis*
32 7:e2100. doi:10.1038/cddis.2016.18
- 33 43. Garaschuk O, Verkhratsky A (2019) GABAergic astrocytes in Alzheimer's
34 disease. *Aging (Albany NY)* 11:1602-1604. doi:10.18632/aging.101870
- 35 44. Gavrilov N, Golyagina I, Brazhe A, Scimemi A, Turlapov V, Semyanov A (2018)
36 Astrocytic Coverage of Dendritic Spines, Dendritic Shafts, and Axonal
37 Boutons in Hippocampal Neuropil. *Front Cell Neurosci* 12:248.
38 doi:10.3389/fncel.2018.00248
- 39 45. Giaume C, Kirchhoff F, Matute C, Reichenbach A, Verkhratsky A (2007) Glia:
40 the fulcrum of brain diseases. *Cell Death Differ* 14:1324-1335.
41 doi:10.1038/sj.cdd.4402144
- 42 46. Giaume C, Koulakoff A, Roux L, Holcman D, Rouach N (2010) Astroglial
43 networks: a step further in neuroglial and gliovascular interactions. *Nat Rev*
44 *Neurosci* 11:87-99. doi:10.1038/nrn2757
- 45 47. Giaume C, Liu X (2012) From a glial syncytium to a more restricted and specific
46 glial networking. *J Physiol Paris* 106:34-39.
47 doi:10.1016/j.jphysparis.2011.09.001
- 48 48. Ginhoux F, Garel S (2018) The mysterious origins of microglia. *Nat Neurosci*
49 21:897-899. doi:10.1038/s41593-018-0176-3
- 50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

- 1 49. Griffin WS, Stanley LC, Ling C, White L, MacLeod V, Perrot LJ, White CL, 3rd,
2 Araoz C (1989) Brain interleukin 1 and S-100 immunoreactivity are elevated
3 in Down syndrome and Alzheimer disease. *Proc Natl Acad Sci U S A*
4 86:7611-7615
- 5 50. Hanisch UK, Kettenmann H (2007) Microglia: active sensor and versatile effector
6 cells in the normal and pathologic brain. *Nat Neurosci* 10:1387-1394.
7 doi:10.1038/nrn1997
- 8 51. Hazell AS (2009) Astrocytes are a major target in thiamine deficiency and
9 Wernicke's encephalopathy. *Neurochem Int* 55:129-135
- 10 52. Hazell AS, Sheedy D, Oanea R, Aghourian M, Sun S, Jung JY, Wang D, Wang C
11 (2009) Loss of astrocytic glutamate transporters in Wernicke encephalopathy.
12 *Glia* 58:148-156
- 13 53. Herrup K (2010) Reimagining Alzheimer's disease--an age-based hypothesis. *J*
14 *Neurosci* 30:16755-16762. doi:10.1523/JNEUROSCI.4521-10.2010
- 15 54. Hertz L, Zielke HR (2004) Astrocytic control of glutamatergic activity: astrocytes
16 as stars of the show. *Trends Neurosci* 27:735-743.
17 doi:10.1016/j.tins.2004.10.008
- 18 55. Huang Y, Mucke L (2012) Alzheimer mechanisms and therapeutic strategies. *Cell*
19 148:1204-1222. doi:10.1016/j.cell.2012.02.040
- 20 56. Iadecola C (2004) Neurovascular regulation in the normal brain and in
21 Alzheimer's disease. *Nat Rev Neurosci* 5:347-360. doi:10.1038/nrn1387
- 22 57. Iadecola C, Nedergaard M (2007) Glial regulation of the cerebral
23 microvasculature. *Nat Neurosci* 10:1369-1376. doi:10.1038/nrn2003
- 24 58. Iram T, Trudler D, Kain D, Kanner S, Galron R, Vassar R, Barzilai A, Blinder P,
25 Fishelson Z, Frenkel D (2016) Astrocytes from old Alzheimer's disease mice
26 are impaired in Aβ uptake and in neuroprotection. *Neurobiol Dis* 96:84-94.
27 doi:10.1016/j.nbd.2016.08.001
- 28 59. Johnson KA, Fox NC, Sperling RA, Klunk WE (2012) Brain imaging in
29 Alzheimer disease. *Cold Spring Harb Perspect Med* 2:a006213.
30 doi:10.1101/cshperspect.a006213
- 31 60. Jones VC, Atkinson-Dell R, Verkhatsky A, Mohamet L (2017) Aberrant iPSC-
32 derived human astrocytes in Alzheimer's disease. *Cell Death Dis* 8:e2696.
33 doi:10.1038/cddis.2017.89
- 34 61. Kametani F, Hasegawa M (2018) Reconsideration of Amyloid Hypothesis and
35 Tau Hypothesis in Alzheimer's Disease. *Front Neurosci* 12:25.
36 doi:10.3389/fnins.2018.00025
- 37 62. Kamphuis W, Kooijman L, Orre M, Stassen O, Pekny M, Hol EM (2015) GFAP
38 and vimentin deficiency alters gene expression in astrocytes and microglia in
39 wild-type mice and changes the transcriptional response of reactive glia in
40 mouse model for Alzheimer's disease. *Glia* 63:1036-1056.
41 doi:10.1002/glia.22800
- 42 63. Katzman R (1976) Editorial: The prevalence and malignancy of Alzheimer
43 disease. A major killer. *Arch Neurol* 33:217-218
- 44 64. Kazim SF, Chuang SC, Zhao W, Wong RK, Bianchi R, Iqbal K (2017) Early-
45 Onset Network Hyperexcitability in Presymptomatic Alzheimer's Disease
46 Transgenic Mice Is Suppressed by Passive Immunization with Anti-Human
47 APP/Aβ Antibody and by mGluR5 Blockade. *Front Aging Neurosci* 9:71.
48 doi:10.3389/fnagi.2017.00071
- 49
- 50
- 51
- 52
- 53
- 54
- 55
- 56
- 57
- 58
- 59
- 60
- 61
- 62
- 63
- 64
- 65

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
65. Kersaitis C, Halliday GM, Kril JJ (2004) Regional and cellular pathology in frontotemporal dementia: relationship to stage of disease in cases with and without Pick bodies. *Acta Neuropathol* 108:515-523
 66. Kettenmann H, Hanisch UK, Noda M, Verkhratsky A (2011) Physiology of microglia. *Physiol Rev* 91:461-553. doi:10.1152/physrev.00011.2010
 67. Kettenmann H, Ransom BR (eds) (2013) *Neuroglia*. Oxford University Press, Oxford
 68. Khakh BS, Sofroniew MV (2015) Diversity of astrocyte functions and phenotypes in neural circuits. *Nat Neurosci* 18:942-952. doi:10.1038/nn.4043
 69. Kimelberg HK, Nedergaard M (2010) Functions of astrocytes and their potential as therapeutic targets. *Neurotherapeutics* 7:338-353. doi:10.1016/j.nurt.2010.07.006
 70. Koffie RM, Hyman BT, Spires-Jones TL (2011) Alzheimer's disease: synapses gone cold. *Mol Neurodegener* 6:63. doi:10.1186/1750-1326-6-63
 71. Kovacs GG, Ferrer I, Grinberg LT, Alafuzoff I, Attems J, Budka H, Cairns NJ, Crary JF, Duyckaerts C, Ghetti B, Halliday GM, Ironside JW, Love S, Mackenzie IR, Munoz DG, Murray ME, Nelson PT, Takahashi H, Trojanowski JQ, Ansorge O, Arzberger T, Baborie A, Beach TG, Bieniek KF, Bigio EH, Bodi I, Dugger BN, Feany M, Gelpi E, Gentleman SM, Giaccone G, Hatanpaa KJ, Heale R, Hof PR, Hofer M, Hortobagyi T, Jellinger K, Jicha GA, Ince P, Kofler J, Kovari E, Kril JJ, Mann DM, Matej R, McKee AC, McLean C, Milenkovic I, Montine TJ, Murayama S, Lee EB, Rahimi J, Rodriguez RD, Rozemuller A, Schneider JA, Schultz C, Seeley W, Seilhean D, Smith C, Tagliavini F, Takao M, Thal DR, Toledo JB, Tolnay M, Troncoso JC, Vinters HV, Weis S, Wharton SB, White CL, 3rd, Wisniewski T, Woulfe JM, Yamada M, Dickson DW (2016) Aging-related tau astroglialopathy (ARTAG): harmonized evaluation strategy. *Acta Neuropathol* 131:87-102. doi:10.1007/s00401-015-1509-x
 72. Kraft AW, Hu X, Yoon H, Yan P, Xiao Q, Wang Y, Gil SC, Brown J, Wilhelmsson U, Restivo JL, Cirrito JR, Holtzman DM, Kim J, Pekny M, Lee JM (2013) Attenuating astrocyte activation accelerates plaque pathogenesis in APP/PS1 mice. *FASEB J* 27:187-198. doi:10.1096/fj.12-208660
 73. Kulijewicz-Nawrot M, Verkhratsky A, Chvatal A, Sykova E, Rodriguez JJ (2012) Astrocytic cytoskeletal atrophy in the medial prefrontal cortex of a triple transgenic mouse model of Alzheimer's disease. *J Anat* 221:252-262. doi:10.1111/j.1469-7580.2012.01536.x
 74. Lanciotti A, Brignone MS, Bertini E, Petrucci TC, Aloisi F, Ambrosini E (2013) Astrocytes: Emerging stars in leukodystrophy pathogenesis. *Transl Neurosci* 4. doi:10.2478/s13380-013-0118-1
 75. Lebedeva A, Plata A, Nosova O, Tyurikova O, Semyanov A (2018) Activity-dependent changes in transporter and potassium currents in hippocampal astrocytes. *Brain Res Bull* 136:37-43. doi:10.1016/j.brainresbull.2017.08.015
 76. Liddelow SA, Guttenplan KA, Clarke LE, Bennett FC, Bohlen CJ, Schirmer L, Bennett ML, Munch AE, Chung WS, Peterson TC, Wilton DK, Frouin A, Napier BA, Panicker N, Kumar M, Buckwalter MS, Rowitch DH, Dawson VL, Dawson TM, Stevens B, Barres BA (2017) Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 541:481-487. doi:10.1038/nature21029
 77. Lin YT, Seo J, Gao F, Feldman HM, Wen HL, Penney J, Cam HP, Gjoneska E, Raja WK, Cheng J, Rueda R, Kritskiy O, Abdurrob F, Peng Z, Milo B, Yu CJ,

- 1 Elmsaouri S, Dey D, Ko T, Yankner BA, Tsai LH (2018) APOE4 Causes
2 Widespread Molecular and Cellular Alterations Associated with Alzheimer's
3 Disease Phenotypes in Human iPSC-Derived Brain Cell Types. *Neuron*
4 98:1141-1154 e1147. doi:10.1016/j.neuron.2018.05.008
- 5 78. Liu CY, Yang Y, Ju WN, Wang X, Zhang HL (2018) Emerging Roles of
6 Astrocytes in Neuro-Vascular Unit and the Tripartite Synapse With Emphasis
7 on Reactive Gliosis in the Context of Alzheimer's Disease. *Front Cell*
8 *Neurosci* 12:193. doi:10.3389/fncel.2018.00193
- 9 79. Ma B, Buckalew R, Du Y, Kiyoshi CM, Alford CC, Wang W, McTigue DM,
10 Enyeart JJ, Terman D, Zhou M (2016) Gap junction coupling confers
11 isopotentiality on astrocyte syncytium. *Glia* 64:214-226.
12 doi:10.1002/glia.22924
- 13 80. Magistretti PJ, Allaman I (2018) Lactate in the brain: from metabolic end-product
14 to signalling molecule. *Nat Rev Neurosci* 19:235-249.
15 doi:10.1038/nrn.2018.19
- 16 81. Meda L, Baron P, Scarlato G (2001) Glial activation in Alzheimer's disease: the
17 role of A β and its associated proteins. *Neurobiol Aging* 22:885-893
- 18 82. Messing A, Brenner M, Feany MB, Nedergaard M, Goldman JE (2012) Alexander
19 disease. *J Neurosci* 32:5017-5023. doi:10.1523/JNEUROSCI.5384-11.2012
- 20 83. Miguel-Hidalgo JJ, Waltzer R, Whittom AA, Austin MC, Rajkowska G,
21 Stockmeier CA (2010) Glial and glutamatergic markers in depression,
22 alcoholism, and their comorbidity. *J Affect Disord* 127:230-240.
23 doi:10.1016/j.jad.2010.06.003
- 24 84. Minkeviciene R, Rheims S, Dobszay MB, Zilberter M, Hartikainen J, Fulop L,
25 Penke B, Zilberter Y, Harkany T, Pitkanen A, Tanila H (2009) Amyloid beta-
26 induced neuronal hyperexcitability triggers progressive epilepsy. *J Neurosci*
27 29:3453-3462. doi:10.1523/JNEUROSCI.5215-08.2009
- 28 85. Mohamet L, Jones VC, Dayanithi G, Verkhatsky A (2018) Pathological human
29 astroglia in Alzheimer's disease: opening new horizons with stem cell
30 technology. *Future Neurology* 13:87 - 99. doi:10.2217/fnl-2017-0029
- 31 86. Morizawa YM, Hirayama Y, Ohno N, Shibata S, Shigetomi E, Sui Y, Nabekura J,
32 Sato K, Okajima F, Takebayashi H, Okano H, Koizumi S (2017) Reactive
33 astrocytes function as phagocytes after brain ischemia via ABCA1-mediated
34 pathway. *Nat Commun* 8:28. doi:10.1038/s41467-017-00037-1
- 35 87. Morris GP, Clark IA, Vissel B (2014) Inconsistencies and controversies
36 surrounding the amyloid hypothesis of Alzheimer's disease. *Acta Neuropathol*
37 *Commun* 2:135. doi:10.1186/s40478-014-0135-5
- 38 88. Mrak RE, Griffin WS (2005) Glia and their cytokines in progression of
39 neurodegeneration. *Neurobiol Aging* 26:349-354
- 40 89. Mucke L, Selkoe DJ (2012) Neurotoxicity of amyloid β -Protein: synaptic and
41 network dysfunction. *Cold Spring Harb Perspect Med* 2:a006338
- 42 90. Mulligan SJ, MacVicar BA (2004) Calcium transients in astrocyte endfeet cause
43 cerebrovascular constrictions. *Nature* 431:195-199. doi:10.1038/nature02827
- 44 91. Nagai J, Rajbhandari AK, Gangwani MR, Hachisuka A, Coppola G, Masmanidis
45 SC, Fanselow MS, Khakh BS (2019) Hyperactivity with Disrupted Attention
46 by Activation of an Astrocyte Synaptogenic Cue. *Cell* 177:1280-1292 e1220.
47 doi:10.1016/j.cell.2019.03.019
- 48 92. Nagy C, Torres-Platas SG, Mechawar N, Turecki G (2017) Repression of
49 Astrocytic Connexins in Cortical and Subcortical Brain Regions and
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

- 1 Prefrontal Enrichment of H3K9me3 in Depression and Suicide. *Int J*
2 *Neuropsychopharmacol* 20:50-57. doi:10.1093/ijnp/pyw071
- 3 93. Naskar S, Chattarji S (2019) Stress Elicits Contrasting Effects on the Structure
4 and Number of Astrocytes in the Amygdala versus Hippocampus. *eNeuro* 6:
5 e0338. doi:10.1523/ENEURO.0338-18.2019
- 6 94. Nedergaard M, Ransom B, Goldman SA (2003) New roles for astrocytes:
7 redefining the functional architecture of the brain. *Trends Neurosci* 26:523-
8 530
- 9 95. Nedergaard M, Verkhratsky A (2012) Artifact versus reality--how astrocytes
10 contribute to synaptic events. *Glia* 60:1013-1023. doi:10.1002/glia.22288
- 11 96. Oksanen M, Petersen AJ, Naumenko N, Puttonen K, Lehtonen S, Gubert Olive M,
12 Shakirzyanova A, Leskela S, Sarajarvi T, Viitanen M, Rinne JO, Hiltunen M,
13 Haapasalo A, Giniatullin R, Tavi P, Zhang SC, Kanninen KM, Hamalainen
14 RH, Koistinaho J (2017) PSEN1 Mutant iPSC-Derived Model Reveals Severe
15 Astrocyte Pathology in Alzheimer's Disease. *Stem Cell Reports* 9:1885-1897.
16 doi:10.1016/j.stemcr.2017.10.016
- 17 97. Olabarria M, Noristani HN, Verkhratsky A, Rodriguez JJ (2010) Concomitant
18 astroglial atrophy and astrogliosis in a triple transgenic animal model of
19 Alzheimer's disease. *Glia* 58:831-838. doi:10.1002/glia.20967
- 20 98. Olabarria M, Noristani HN, Verkhratsky A, Rodriguez JJ (2011) Age-dependent
21 decrease in glutamine synthetase expression in the hippocampal astroglia of
22 the triple transgenic Alzheimer's disease mouse model: mechanism for
23 deficient glutamatergic transmission? *Mol Neurodegener* 6:55.
24 doi:10.1186/1750-1326-6-55
- 25 99. Osborn LM, Kamphuis W, Wadman WJ, Hol EM (2016) Astrogliosis: An integral
26 player in the pathogenesis of Alzheimer's disease. *Prog Neurobiol* 144:121-
27 141. doi:10.1016/j.pneurobio.2016.01.001
- 28 100. Park J, Wetzel I, Marriott I, Dreau D, D'Avanzo C, Kim DY, Tanzi RE, Cho H
29 (2018) A 3D human triculture system modeling neurodegeneration and
30 neuroinflammation in Alzheimer's disease. *Nat Neurosci* 21:941-951.
31 doi:10.1038/s41593-018-0175-4
- 32 101. Patrushev I, Gavrilov N, Turlapov V, Semyanov A (2013) Subcellular location
33 of astrocytic calcium stores favors extrasynaptic neuron-astrocyte
34 communication. *Cell Calcium* 54:343-349. doi:10.1016/j.ceca.2013.08.003
- 35 102. Pekny M, Pekna M (2014) Astrocyte reactivity and reactive astrogliosis: costs
36 and benefits. *Physiol Rev* 94:1077-1098. doi:10.1152/physrev.00041.2013
- 37 103. Pekny M, Pekna M, Messing A, Steinhauser C, Lee JM, Parpura V, Hol EM,
38 Sofroniew MV, Verkhratsky A (2016) Astrocytes: a central element in
39 neurological diseases. *Acta Neuropathol* 131:323-345. doi:10.1007/s00401-
40 015-1513-1
- 41 104. Pellerin L, Magistretti PJ (2012) Sweet sixteen for ANLS. *J Cereb Blood Flow*
42 *Metab* 32:1152-1166. doi:10.1038/jcbfm.2011.149
- 43 105. Perez-Alvarez A, Navarrete M, Covelo A, Martin ED, Araque A (2014)
44 Structural and functional plasticity of astrocyte processes and dendritic spine
45 interactions. *J Neurosci* 34:12738-12744. doi:10.1523/JNEUROSCI.2401-
46 14.2014
- 47 106. Plata A, Lebedeva A, Denisov P, Nosova O, Postnikova TY, Pimashkin A,
48 Brazhe A, Zaitsev AV, Rusakov DA, Semyanov A (2018) Astrocytic Atrophy
49 Following Status Epilepticus Parallels Reduced Ca²⁺ Activity and Impaired
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

- Synaptic Plasticity in the Rat Hippocampus. *Front Mol Neurosci* 11:215. doi:10.3389/fnmol.2018.00215
107. Polis B, Srikanth KD, Elliott E, Gil-Henn H, Samson AO (2018) L-Norvaline Reverses Cognitive Decline and Synaptic Loss in a Murine Model of Alzheimer's Disease. *Neurotherapeutics* 15:1036-1054. doi:10.1007/s13311-018-0669-5
108. Rajkowska G, Legutko B, Moulana M, Syed M, Romero DG, Stockmeier CA, Miguel-Hidalgo JJ (2018) Astrocyte pathology in the ventral prefrontal white matter in depression. *J Psychiatr Res* 102:150-158. doi:10.1016/j.jpsychires.2018.04.005
109. Rajkowska G, Miguel-Hidalgo JJ (2007) Gliogenesis and glial pathology in depression. *CNS Neurol Disord Drug Targets* 6:219-233
110. Rajkowska G, Stockmeier CA (2013) Astrocyte pathology in major depressive disorder: insights from human postmortem brain tissue. *Curr Drug Targets* 14:1225-1236
111. Reichenbach A, Pannicke T (2008) Neuroscience. A new glance at glia. *Science* 322:693-694. doi:10.1126/science.1166197
112. Rodriguez JJ, Terzieva S, Olabarria M, Lanza RG, Verkhratsky A (2013) Enriched environment and physical activity reverse astroglial degeneration in the hippocampus of AD transgenic mice. *Cell Death Dis* 4:e678. doi:10.1038/cddis.2013.194
113. Rossi D (2015) Astrocyte physiopathology: At the crossroads of intercellular networking, inflammation and cell death. *Prog Neurobiol* 130:86-120. doi:10.1016/j.pneurobio.2015.04.003
114. Rossi D, Brambilla L, Valori CF, Roncoroni C, Crugnola A, Yokota T, Bredesen DE, Volterra A (2008) Focal degeneration of astrocytes in amyotrophic lateral sclerosis. *Cell Death Differ* 15:1691-1700
115. Rossi D, Volterra A (2009) Astrocytic dysfunction: Insights on the role in neurodegeneration. *Brain Res Bull* 80:224-232
116. Rouach N, Koulakoff A, Abudara V, Willecke K, Giaume C (2008) Astroglial metabolic networks sustain hippocampal synaptic transmission. *Science* 322:1551-1555. doi:10.1126/science.1164022
117. Rowland HA, Hooper NM, Kellett KAB (2018) Modelling Sporadic Alzheimer's Disease Using Induced Pluripotent Stem Cells. *Neurochem Res* 43:2179-2198. doi:10.1007/s11064-018-2663-z
118. Rubinsztein DC (2006) The roles of intracellular protein-degradation pathways in neurodegeneration. *Nature* 443:780-786. doi:10.1038/nature05291
119. Sanacora G, Banasr M (2013) From pathophysiology to novel antidepressant drugs: glial contributions to the pathology and treatment of mood disorders. *Biol Psychiatry* 73:1172-1179. doi:10.1016/j.biopsych.2013.03.032
120. Schwarcz R, Hunter CA (2007) *Toxoplasma gondii* and schizophrenia: linkage through astrocyte-derived kynurenic acid? *Schizophr Bull* 33:652-653. doi:10.1093/schbul/sbm030
121. Scofield MD, Li H, Siemsen BM, Healey KL, Tran PK, Woronoff N, Boger HA, Kalivas PW, Reissner KJ (2016) Cocaine self-administration and extinction leads to reduced glial fibrillary acidic protein expression and morphometric features of astrocytes in the nucleus accumbens core. *Biol Psychiatry* 80:207-215. doi:10.1016/j.biopsych.2015.12.022
122. Selkoe D, Mandelkow E, Holtzman D (2012) Deciphering Alzheimer disease. *Cold Spring Harb Perspect Med* 2:a011460. doi:10.1101/cshperspect.a011460

- 1 123. Semyanov A (2019) Spatiotemporal pattern of calcium activity in astrocytic
2 network. *Cell Calcium* 78:15-25. doi:10.1016/j.ceca.2018.12.007
- 3 124. Shih PY, Savtchenko LP, Kamasawa N, Dembitskaya Y, McHugh TJ, Rusakov
4 DA, Shigemoto R, Semyanov A (2013) Retrograde synaptic signaling
5 mediated by K⁺ efflux through postsynaptic NMDA receptors. *Cell Rep*
6 5:941-951. doi:10.1016/j.celrep.2013.10.026
- 7 125. Shokri-Kojori E, Wang GJ, Wiers CE, Demiral SB, Guo M, Kim SW, Lindgren
8 E, Ramirez V, Zehra A, Freeman C, Miller G, Manza P, Srivastava T, De
9 Santi S, Tomasi D, Benveniste H, Volkow ND (2018) β -Amyloid
10 accumulation in the human brain after one night of sleep deprivation. *Proc*
11 *Natl Acad Sci U S A* 115:4483-4488. doi:10.1073/pnas.1721694115
- 12 126. Simon MJ, Wang MX, Murchison CF, Roesse NE, Boespflug EL, Woltjer RL,
13 Iliff JJ (2018) Transcriptional network analysis of human astrocytic endfoot
14 genes reveals region-specific associations with dementia status and tau
15 pathology. *Sci Rep* 8:12389. doi:10.1038/s41598-018-30779-x
- 16 127. Sofroniew MV (2009) Molecular dissection of reactive astrogliosis and glial scar
17 formation. *Trends Neurosci* 32:638-647. doi:10.1016/j.tins.2009.08.002
- 18 128. Sofroniew MV (2014) Astrogliosis. *Cold Spring Harb Perspect Biol* 7:a020420.
19 doi:10.1101/cshperspect.a020420
- 20 129. Sofroniew MV, Vinters HV (2010) Astrocytes: biology and pathology. *Acta*
21 *Neuropathol* 119:7-35. doi:10.1007/s00401-009-0619-8
- 22 130. Sweeney MD, Zhao Z, Montagne A, Nelson AR, Zlokovic BV (2019) Blood-
23 Brain Barrier: From Physiology to Disease and Back. *Physiol Rev* 99:21-78.
24 doi:10.1152/physrev.00050.2017
- 25 131. Takano T, Tian GF, Peng W, Lou N, Libionka W, Han X, Nedergaard M (2006)
26 Astrocyte-mediated control of cerebral blood flow. *Nat Neurosci* 9:260-267.
27 doi:10.1038/nn1623
- 28 132. Tanaka M, Shih PY, Gomi H, Yoshida T, Nakai J, Ando R, Furuichi T,
29 Mikoshiba K, Semyanov A, Itohara S (2013) Astrocytic Ca²⁺ signals are
30 required for the functional integrity of tripartite synapses. *Mol Brain* 6:6.
31 doi:10.1186/1756-6606-6-6
- 32 133. Tarantini S, Tran CHT, Gordon GR, Ungvari Z, Csiszar A (2017) Impaired
33 neurovascular coupling in aging and Alzheimer's disease: Contribution of
34 astrocyte dysfunction and endothelial impairment to cognitive decline. *Exp*
35 *Gerontol* 94:52-58. doi:10.1016/j.exger.2016.11.004
- 36 134. Terry RD (2000) Cell death or synaptic loss in Alzheimer disease. *J Neuropathol*
37 *Exp Neurol* 59:1118-1119
- 38 135. Terry RD, Masliah E, Salmon DP, Butters N, DeTeresa R, Hill R, Hansen LA,
39 Katzman R (1991) Physical basis of cognitive alterations in Alzheimer's
40 disease: synapse loss is the major correlate of cognitive impairment. *Ann*
41 *Neurol* 30:572-580. doi:10.1002/ana.410300410
- 42 136. Tong X, Ao Y, Faas GC, Nwaobi SE, Xu J, Haustein MD, Anderson MA, Mody
43 I, Olsen ML, Sofroniew MV, Khakh BS (2014) Astrocyte K_{ir}4.1 ion channel
44 deficits contribute to neuronal dysfunction in Huntington's disease model
45 mice. *Nat Neurosci* 17:694-703. doi:10.1038/nn.3691
- 46 137. Valori CF, Guidotti G, Brambilla L, Rossi D (2019) Astrocytes: Emerging
47 Therapeutic Targets in Neurological Disorders. *Trends Mol Med.*
48 doi:10.1016/j.molmed.2019.04.010
- 49 138. Valtcheva S, Venance L (2019) Control of Long-Term Plasticity by Glutamate
50 Transporters. *Front Synaptic Neurosci* 11:10. doi:10.3389/fnsyn.2019.00010
- 51
- 52
- 53
- 54
- 55
- 56
- 57
- 58
- 59
- 60
- 61
- 62
- 63
- 64
- 65

- 1 139. Verkhratsky A, Butt AM (2013) *Glial Physiology and Pathophysiology*. Wiley-
2 Blackwell, Chichester
- 3 140. Verkhratsky A, Butt AM (2018) The History of the Decline and Fall of the Glial
4 Numbers Legend. *Neuroglia* 1:188 - 192. doi:10.3390/neuroglia1010013
- 5 141. Verkhratsky A, Marutle A, Rodriguez-Arellano JJ, Nordberg A (2015) Glial
6 asthenia and functional paralysis: A new perspective on neurodegeneration
7 and Alzheimer's disease. *Neuroscientist* 21:552-568.
8 doi:10.1177/1073858414547132
- 9 142. Verkhratsky A, Nedergaard M (2014) Astroglial cradle in the life of the synapse.
10 *Philos Trans R Soc Lond B Biol Sci* 369:20130595.
11 doi:10.1098/rstb.2013.0595
- 12 143. Verkhratsky A, Nedergaard M (2016) The homeostatic astroglia emerges from
13 evolutionary specialization of neural cells. *Philos Trans R Soc Lond B Biol*
14 *Sci* 371. doi:10.1098/rstb.2015.0428
- 15 144. Verkhratsky A, Nedergaard M (2018) Physiology of Astroglia. *Physiol Rev*
16 98:239-389. doi:10.1152/physrev.00042.2016
- 17 145. Verkhratsky A, Olabarria M, Noristani HN, Yeh CY, Rodriguez JJ (2010)
18 Astrocytes in Alzheimer's disease. *Neurotherapeutics* 7:399-412.
19 doi:10.1016/j.nurt.2010.05.017
- 20 146. Verkhratsky A, Parpura V (2016) Astroglipathology in neurological,
21 neurodevelopmental and psychiatric disorders. *Neurobiol Dis* 85:254-261.
22 doi:10.1016/j.nbd.2015.03.025
- 23 147. Verkhratsky A, Parpura V, Rodriguez JJ (2011) Where the thoughts dwell: the
24 physiology of neuronal-glia "diffuse neural net". *Brain Res Rev* 66:133-151.
25 doi:10.1016/j.brainresrev.2010.05.002
- 26 148. Verkhratsky A, Rodriguez JJ, Parpura V (2012) Calcium signalling in astroglia.
27 *Mol Cell Endocrinol* 353:45-56. doi:10.1016/j.mce.2011.08.039
- 28 149. Verkhratsky A, Rodriguez JJ, Steardo L (2014) Astroglipathology: a central
29 element of neuropsychiatric diseases? *Neuroscientist* 20:576-588.
30 doi:10.1177/1073858413510208
- 31 150. Verkhratsky A, Sofroniew MV, Messing A, deLanerolle NC, Rempe D,
32 Rodriguez JJ, Nedergaard M (2012) Neurological diseases as primary
33 gliopathies: a reassessment of neurocentrism. *ASN Neuro* 4.
34 doi:10.1042/AN20120010
- 35 151. Verkhratsky A, Zorec R, Parpura V (2017) Stratification of astrocytes in healthy
36 and diseased brain. *Brain Pathol* 27:629-644. doi:10.1111/bpa.12537
- 37 152. Verkhratsky A, Zorec R, Rodriguez JJ, Parpura V (2016) Astroglia dynamics in
38 ageing and Alzheimer's disease. *Curr Opin Pharmacol* 26:74-79.
39 doi:10.1016/j.coph.2015.09.011
- 40 153. Virchow R (1858) *Die Cellularpathologie in ihrer Begründung auf*
41 *physiologische and pathologische Gewebelehre*. Zwanzig Vorlesungen
42 gehalten während der Monate Februar, März und April 1858 im
43 pathologischen Institut zu Berlin. First edition edn. August Hirschwald, Berlin
- 44 154. Wakida NM, Cruz GMS, Ro CC, Moncada EG, Khatibzadeh N, Flanagan LA,
45 Berns MW (2018) Phagocytic response of astrocytes to damaged neighboring
46 cells. *PLoS One* 13:e0196153. doi:10.1371/journal.pone.0196153
- 47 155. Wallraff A, Kohling R, Heinemann U, Theis M, Willecke K, Steinhauser C
48 (2006) The impact of astrocytic gap junctional coupling on potassium
49 buffering in the hippocampus. *J Neurosci* 26:5438-5447.
50 doi:10.1523/JNEUROSCI.0037-06.2006
- 51
- 52
- 53
- 54
- 55
- 56
- 57
- 58
- 59
- 60
- 61
- 62
- 63
- 64
- 65

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
156. Willis T (1672) *De Anima Brutorum quae Hominis Vitalis ac Sensitiva est. E Theatro Sheldoniano*; Ric.Davis Oxonii (Oxford)
 157. Wu Z, Guo Z, Gearing M, Chen G (2014) Tonic inhibition in dentate gyrus impairs long-term potentiation and memory in an Alzheimer's [corrected] disease model. *Nat Commun* 5:4159. doi:10.1038/ncomms5159
 158. Yamaguchi H, Sugihara S, Ogawa A, Saido TC, Ihara Y (1998) Diffuse plaques associated with astroglial amyloid beta protein, possibly showing a disappearing stage of senile plaques. *Acta Neuropathol* 95:217-222
 159. Yamanaka K, Chun SJ, Boillee S, Fujimori-Tonou N, Yamashita H, Gutmann DH, Takahashi R, Misawa H, Cleveland DW (2008) Astrocytes as determinants of disease progression in inherited amyotrophic lateral sclerosis. *Nat Neurosci* 11:251-253
 160. Yamanaka K, Komine O (2018) The multi-dimensional roles of astrocytes in ALS. *Neurosci Res* 126:31-38. doi:10.1016/j.neures.2017.09.011
 161. Yeh CY, Vadhvana B, Verkhratsky A, Rodriguez JJ (2012) Early astrocytic atrophy in the entorhinal cortex of a triple transgenic animal model of Alzheimer's disease. *ASN Neuro* 3:271-279. doi:10.1042/AN20110025
 162. Yi C, Koulakoff A, Giaume C (2017) Astroglial Connexins as a Therapeutic Target for Alzheimer's Disease. *Curr Pharm Des* 23:4958-4968. doi:10.2174/1381612823666171004151215
 163. Zeidan-Chulia F, Salmina AB, Malinovskaya NA, Noda M, Verkhratsky A, Moreira JC (2014) The glial perspective of autism spectrum disorders. *Neurosci Biobehav Rev* 38:160-172. doi:10.1016/j.neubiorev.2013.11.008
 164. Zhao Y, Lin Z, Chen L, Ouyang L, Gu L, Chen F, Zhang Q (2018) Hippocampal astrocyte atrophy in a mouse depression model induced by corticosterone is reversed by fluoxetine instead of benzodiazepine diazepam. *Prog Neuropsychopharmacol Biol Psychiatry* 83:99-109. doi:10.1016/j.pnpbp.2018.01.011
 165. Zonta M, Angulo MC, Gobbo S, Rosengarten B, Hossmann KA, Pozzan T, Carmignoto G (2003) Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation. *Nat Neurosci* 6:43-50. doi:10.1038/nn980
 166. Zorec R, Parpura V, Vardjan N, Verkhratsky A (2017) Astrocytic face of Alzheimer's disease. *Behav Brain Res* 322:250-257. doi:10.1016/j.bbr.2016.05.021

Figure legends

Figure 1. Diversity of astrocytes.

Figure 2. Classification of astroglipathology. Modified from [103].

Figure 3. Astroglia atrophy in hippocampal (a) and cortical (B) regions of the wild type (WT) and triple transgenic AD model mouse (3xTG-AD). The images show GFAP-positive profiles of astroglial cells.

Collated from [73,97,161].

Figure 4. Astrocytes derived from iPSCs isolated from familial AD patient carrying *PSEN1* M146L mutation and sporadic AD patient carrying *ApoE4*^{+/+} demonstrate significant atrophy when compared to those from healthy person.

(A) Morphological appearance (arborised, polarised and fibroblast-like) of astrocytes derived from iPSCs of healthy controls, familial and sporadic AD (FAD and SAD respectively). (B). Exemplar 3D IsoSurface renders constructed from serial confocal z-stacks display clear differences in cell size and overall morphology Scale bar = 10 μm . Quantification of cells using these renders by way of surface area (C), cell volume (D) and SA:Vol ratio (E) reveal significant differences in all aspects of cellular morphology between healthy and diseased astrocytes. Quantification of mean fluorescence intensity per immunoreactive cell reveals no significant difference in GFAP staining intensities between AD- and control astrocytes (F) but S100B, EAAT1 and GS intensities are reduced in both FAD and SAD cells (G, H and I, respectively). Asterisks on graph; *** $p < 0.001$, ** $p < 0.005$, * $p < 0.05$.

Reproduced from [60].

Table 1. Astrocytes in AD: Atrophy and loss of function

Model/Age/Preparation	Properties	Loss of function	Reference
5xFAD/12 - 14 months/ Primary culture		Reduced β -amyloid ₄₂ uptake; reduced neuroprotection reduced ability to promote neuronal growth	[58]
3xTG/ 4 Month/Fixed brain slices/Immunocytochemistry	Reduced morphological profiles and arborization	Treatment with L-norvaline rescued astroglial atrophy, increased dendritic spines densities and improved memory deficits	[107]
Swiss mice/ 3 month/Intracerebroventricular injection of 10 pM β -amyloid oligomers/immunocytochemistry	Morphological atrophy of astrocytes; ~23% reduction in a number of processes and ~40% reduction in surface area.	Decreased neuroprotection; increase in synaptic loss; both effects ameliorated by intracerebroventricular injection of 10 ng transforming growth factor- β 1	[32]
PDAPP-J20 mice/5 months/Immunocytochemistry	Decreased volume and complexity of hippocampal astrocytes		[8]
3xTG AD mice/1 to 24 months/Immunocytochemistry	Decreased volume, surface area and complexity of astrocytes in entorhinal and prefrontal cortices and in the hippocampus.	Loss of homoeostatic support	[73,97,98,161]

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Ref.: Ms. No. PAEJ-D-19-00142
Astroglial atrophy in Alzheimer's disease
Pflügers Archiv - European Journal of Physiology

Responses to reviewer's comments

Reviewer #1:

In their manuscript, the authors acknowledged the growing evidence that glial cells are not only affected by neurodegenerative processes, but are causative for neurodegeneration in diseases such as Alzheimer's. The authors are well-known experts in the field. The composition of the manuscript is reasonable and the selected literature well-balanced. However, there are many minor points, mainly addressing spelling, grammar and style, some of which are listed below. Therefore, I recommend a thorough check of the text.

Our reply: Thank you very much for constructive comments; we incorporated all your suggestions.

1) Many commas are missing.

Our reply: we proof read the text in particular from commas angle

2) P. 3, l. 18: Delete "n" in "Alzheimer's"

Our reply: Done

3) P. 3, l. 44: Delete one "single night" in "Even acute single night of sleep deprivation for a single night causes ..."

Our reply: Done

4) P. 5, l. 1: Change "reach" to "rich"

Our reply: Done

5) P. 6, l. 19: It is not clear what "disease- and disease-stage" means. The phrase appears to be incomplete.

Our reply: Thank you we meant "disease- and disease-stage specific"; the text is amended

6) P. 6, l. 20: Reconsider the sentence and the numbering of "Conceptually, astroglipathological phenotypes are classified into three major groups: (i) reactive astrogliosis (ii) astrodegeneration with astroglial atrophy and loss of function; (ii) pathological remodelling"

Our reply: Done

7) P. 8, l. 57: The parenthesis is not closed in "(Table 1, [53,92,29,7]."

Our reply: Done

8) P. 9, l. 4: Change "AD mice model" in "AD mouse models"

Our reply: Done

9) P. 9, l. 8: Delete "s" in "signals"

Our reply: Done

10) P. 9, l. 49: Change "stem cells derived astrocytes" in "stem cell-derived astrocytes"

Our reply: Done

11) P. 10, l. 50: Lower case "p" in "Positron-emission"

Our reply: Done

12) P. 11, l. 7: Change "In recent decade" either in "In the recent decade" or in "In recent decades"

Our reply: Done

13) P. 11, l. 9: Change "patomorphological" in "pathomorphological"

Our reply: Done

14) P. 11, l. 16: "Alzheimer" to "Alzheimer's"

Our reply: Done

15) Fig. 3, legend: Add "A" and "B" to the legend.

Our reply: Done

16) Fig. 4. Panel A is not correctly explained in the legend. "(A)" actually refers to panel B. Information referring to panel A is missing.

Our reply: Done

17) Fig. 4. "Scale bar = 10 um". there is no scale bar in the figure.

Our reply: it was actually - but too thin, and made it ticker.

Reviewer #2:

Verkhatsky and co-authors provide an up-to-date and timely review focusing on the changes in astrocytes in neurodegenerative (and Alzheimer's) disease, which will be very valuable to the broad readership of Pfluegers Archive. The historical views and broad introduction are informative and these days neglected by most authors. My comments to the abstract reflect my most important concerns; the authors should make an effort to better clarify the topic of astrocytopathy versus different from of astrogliosis that in are discussed throughout the text.

Our reply: Thank you very much for constructive comments; we incorporated your suggestions.

1. Abstract: several terms are used which are not clearly defined. Reactive gliosis is generally viewed as being accompanied by hypertrophy of astrocytes, the latter mostly referring to the increase in GFAP expression and increased thickness of primary processes. What then is meant by "hypotrophy"? The same is true for the terms "astrogliosis"/"astrogliotic response"/"astroglial hypotrophy / atrophy / paralysis". The abstract would greatly profit from a clear definition and separation of these terms. Alternatively, these terms should be omitted and more general summarizing statements be made in the abstract.

Our reply: We edited the abstract; obviously the word "hypotrophy" was a typo; astrogliosis is indeed characterised by hypertrophic changes.

As to terminology: (i) Astrogliosis;(ii) Astroglial atrophy and (iii) Pathological remodelling of astrocytes - this has been defined in several recent publications and are quite generally accepted (see consensus paper by Pekny M et al. (2016) Astrocytes: a central element in neurological diseases. *Acta Neuropathol* 131:323-345 doi:10.1007/s00401-015-1513-1; and recent essay by us Verkhatsky A, Zorec R, Parpura V (2017) Stratification of astrocytes in healthy and diseased brain. *Brain Pathol* 27:629-644 doi:10.1111/bpa.12537). To make this terminology easy for a reader we have now included a text box with a glossary of the terms.

2. There is a lot of evidence showing that astrocytes are not just "homeostatic" cells but actively contribute to brain communication.

Our reply: We fully agree that astrocytes communicate with other brain cells through numerous mechanisms; all their homeostatic cascades are "active" and do affect neurones and synapses. We have discussed these matters in depth in previous publications (e.g. *Physiol Rev* 2018).

3. Page 3, line 59: what is meant by "compromised synaptic physiology"; isn't hyperexcitability always "aberrant"?

Our reply: we agree and we amended the text accordingly

4. Page 4, lines 1-7: white matter (oligodendrocyte) damage is probably not primarily associated with failure in homeostatic neuroglia, which, in the context of the review here, relates to astrocytes?

Our reply: Oligodendrocytes are as homeostatic as astrocytes in white matter - they support axons in many ways including for example supplying them with lactate. In this context we simply mentioned that oligodendrodegeneration (which is rather prominent in AD) contributes to overall "unwiring" of the brain.

5. The term "homeostasis/homeostatic" is used quite often throughout the text. What exactly is meant by that? It is used as if "homeostasis" is the ultimate goal and ultimate condition to attain. Life as such is probably never in a "homeostatic" condition.

Our reply: this is of course very interesting and philosophical question; we as such are following the ideas of Claude Bernard, Walter Cannon and Joseph Barcroft. Failure in physiological homeostasis means disease and inability of the organism to maintain physiological homeostasis leads to extinction of biological entity; failure of homeostatic control of brain tissue signals its inability to perform - i.e. dementia in the broadest reading of it.

6. Page 5, line 28: There is no clear evidence that "neurological disorders are the failure of tissue homeostasis". This is a rather general statement and as such, debatable. What is meant by tissue homeostasis?

Our reply: This is how we define the disease: disease is failure of homeostatic systems which are unable anymore to maintain structural and functional stability. This definition is obviously open to debate, which we of course welcome. We agree however that the word "tissue" is inappropriate in this context and we amended the text accordingly.

7. Page 6, lines 16: Neurones can well survive without glia, this is the basis for primary neuronal cell cultures...

Our reply: well, purified cultures of central neurones (i.e. without astrocytes) do not thrive and form much less synapses; at the organism level ablation of glia is lethal for

neurones.

8. Pages 3-6: the text somewhat jumps back and forth, addressing properties of astrocytes and their arrangement in networks, as well as their physiological roles in healthy tissue and then mentioning changes in response to different pathologies. The review would greatly profit from a more organized separation of topics to discuss: e.g. first describe astrocytes and their function /organization in the healthy brain, and only then describe astrocytes under different pathological conditions.

Our reply: this is the matter of opinion: we feel that we did exactly what is proposed by the reviewer.

9. Page 6/7 and following: it is clearly debatable that reactive gliosis is a purely defensive programme. Reactive astrocytes reduce their "homeostatic" functions and have been reported to "de-differentiate", including a reduced expression of glutamate transporters, KIR-channels, NKA, gap junction coupling. Increased expression of GFAP should be mentioned, as well as changes in their morphology, retraction of processes This, however, is described only later (page 7), and referred to as "astrodegeneration". Again, it seems mandatory, that the different terms used ("reactive astrogliosis", "astrodegeneration", "astrotrophy",...) are better defined. Glial scar formation, involving reactive gliosis and proliferation of astrocytes must be separated from "hypertrophy" and "astrotrophy".

Our reply: we are somewhat confused by this logic. We of course agree that astroglial reactivity may produce neurotoxic phenotypes and we state that ("Being driven to an extreme by the severity of the insult, glial cells can assume the role of natural killer and contribute to the neural cell death, and yet even neurotoxicity of glia has, at its core, neuroprotective significance. "; our argument is rather simple - evolution never selects for negative mechanisms only for positive or neutral; hence astroglial neurotoxicity has at its core some beneficial effects - e.g. elimination of compromised neurones which may help faster resolution of pathology. The scar is obviously defensive, this seems rather obvious - a making a wall between healthy and damaged tissues.; of course if driven out of balance neurotoxic phenotypes facilitates cell death and overall demise of tissue which we clearly acknowledged. We do not use term "astrotrophy"? The terms "atrophy" and loss of function which we employ are of very general usage in medical literature. Now we have introduced a glossary to clearly define these.

10. Page 9, lines 38 ff: reactive astrocyte near lesions are characterized by aberrant calcium handling, which should be discussed.

Our reply: We fully agree with this; and yet in this paper we do not discuss much astroglial reactivity in AD; this has been covered by many excellent reviews; we specifically focus on astroglial atrophy, and hence this discussion is, in our opinion, outside of the scope.

11. Page 9, lines 54/55: the consequences of a retraction of astrocyte processes from synapses are not clear. Retraction of glial processes might also cause a widening of the ECS and a "dilution" of glutamate that escapes the synaptic cleft.

Our reply: In fact the main consequence of astroglial process retraction would be removal of glutamate transporters from the synapse. Indeed enhanced spillover obviously causes aberrant synaptic transmission - and weakens synaptic potentiation - this is exactly our logic.

12. Page 10, lines 8/9: reduction of glutamate uptake might be accompanied by desensitization of AMPA receptors.

Our reply: possibly yes however we are not aware about any data on this matter in the context of AD.

13. Page 10, line30: it is not clear from the text above, that astrogliosis is a "powerful defensive programme". The examples mentioned later show, at best, a correlation between astrocyte gliosis, amyloid deposition, and AD. At present, this seems rather like a "chicken and egg" problem, which should be acknowledged.

Our reply: we politely disagree; again this are all matters of opinion and writing style.

14. Fig. 1: the direct line between "specialized astrocytes" and "Human astrocytes" does not make sense.

Our reply: Thank you, we amended a figure accordingly

15. Fig. 2: should be revised and then could help in clarifying the different expressions used (see comments above). The term "astrogliopathy" somewhat bears the notion that astrocytes degenerate themselves. Reactive astrocytes are e.g. also observed during healthy ageing, is this pathological already? Again, please define what is mean by reactive astrogliosis and include all other terms used in the text.

Our reply: This figure shows classification of pathological changes which has been explained in detail in previous papers, which is of course fully acknowledged - this figure is take from Pekny et al., Acta Neuropathol, 2016. We do not call anywhere reactive astrocytes "pathological"; and yes astrocytes do degenerate by themselves in many neurological diseases; all this has been covered previously and we see no reason in repeating this here. We added the Glossary of terms to the text.

16. Fig. 3: apparently shows GFAP-labelling? Please state in the legend. GFAP does not delineate the entire morphology of the cells.

Our reply: Thank you; and we modified figure legend and the text accordingly.

17. Fig. 4: needs further explanation in the legend as well. These are cells derived from iPSCs?

Our reply: Yes and we edited the figure legend

18. General: the text is clearly written, but many typos and grammatical errors remain (e.g. page 3, lines 8/9, should read: "processes"; line 10: "A direct link..."; ...). Please proofread carefully.

Our reply: we read the paper and amended many of those small typos - thank you.



Change of authorship request form (pre-acceptance)

Please read the important information on page 4 before you begin

This form should be used by authors to request any change in authorship including changes in corresponding authors. Please fully complete all sections. Use black ink and block capitals and provide each author's full name with the given name first followed by the family name.

Please note: In author collaborations where there is formal agreement for representing the collaboration, it is sufficient for the representative or legal guarantor (usually the corresponding author) to complete and sign the Authorship Change Form on behalf of all authors.

Manuscript ID no. in case of unpublished manuscript: PAEJ-D-19-00142R1

Section 1: Please provide the current title of manuscript
 (For journals: Please provide the manuscript ID, title and/or DOI if available.)
 (For books: Please provide the title, ISBN and/or DOI if available.)

Manuscript ID no. in case of unpublished manuscript:
DOI in case of published manuscript:
ISBN (for books):

Title: Astroglial atrophy in Alzheimer's disease

Section 2: Please provide the previous authorship, in the order shown on the manuscript before the changes were introduced. Please indicate the corresponding author by adding (CA) behind the name.

	First name(s)	Family name	ORCID or SCOPUS id, if available
1 st author	Alexei	Verkhatsky CA	
2 nd author	Jose Julio	Rodriguez	
3 rd author	Robert	Zorec	
4 th author	Alexey	Semyanov	
5 th author			
6 th author			
7 th author			

Please use an additional sheet if there are more than 7 authors.

Change of authorship request form (pre-acceptance)

Section 3: Please provide a justification for change. Please use this section to explain your reasons for changing the authorship of your manuscript, e.g. what necessitated the change in authorship? Please refer to the (journal) policy pages for more information about authorship. Please explain why omitted authors were not originally included and/or why authors were removed on the submitted manuscript.

Section 4: Proposed new authorship. Please provide your new authorship list in the order you would like it to appear on the manuscript. Please indicate the corresponding author by adding (CA) behind the name. If the corresponding author has changed, please indicate the reason under section 3.

	First name(s)	Family name (this name will appear in full on the final publication and will be searchable in various abstract and indexing databases)
1 st author	Alexei	Verkhratsky CA
2 nd author	Jose Julio	Rodriguez
3 rd author	Augustas	Pivoriunas
4 th author	Robert	Zorec
5 th author	Alexey	Semyanov
6 th author		
7 th author		

Please use an additional sheet if there are more than 7 authors.

Change of authorship request form (pre-acceptance)

Section 5: Author contribution, Acknowledgement and Disclosures. Please use this section to provide a new disclosure statement and, if appropriate, acknowledge any contributors who have been removed as authors and ensure you state what contribution any new authors made (if applicable per the journal or book (series) policy). **Please ensure these are updated in your manuscript - after approval of the change(s) - as our production department will not transfer the information in this form to your manuscript.**


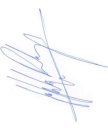
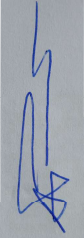


New acknowledgements:	N/A
New Disclosures (financial and non-financial interests, funding):	N/A
New Author Contributions statement (if applicable per the journal policy):	N/A

State 'Not applicable' if there are no new authors.

Change of authorship request form (pre-acceptance)

Section 6: Declaration of agreement. All authors, unchanged, new and removed must sign this declaration.

(NB: Please print the form, sign and return a scanned copy. Please note that signatures that have been inserted as an image file are acceptable as long as it is handwritten. Typed names in the signature box are unacceptable.) * Please delete as appropriate. Delete all of the bold if you were on the original authorship list and are remaining as an author.

	First name	Family name	I agree to the proposed new authorship shown in section 4 /and the addition/removal* of my name to the authorship list.	Signature	Affiliated institute	Date
1 st author	Alexei	Verkhatsky CA	I agree to the proposed new authorship shown in section 4 /and the addition/removal* of my name to the authorship list.		The University of Manchester	Aug 22 2019
2 nd author	Jose Julio	Rodriguez	I agree to the proposed new authorship shown in section 4 /and the addition/removal* of my name to the authorship list.		The University of the Basque Country	Aug 22 2019
3 rd author	Augustas	Pivoriunas	I agree to the proposed new authorship shown in section 4 /and the addition/removal* of my name to the authorship list.		State Research Institute Centre for Innovative Medicine	Aug 22 2019
4 th authors	Robert	Zorec	I agree to the proposed new authorship shown in section 4 /and the addition/removal* of my name to the authorship list.		University of Ljubljana	Aug 22 2019
5 th author	Alexey	Semyanov	I agree to the proposed new authorship shown in section 4 /and the addition/removal* of my name to the authorship list.		Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry	Aug 22 2019
6 th author			I agree to the proposed new authorship shown in section 4 /and the addition/removal* of my name to the authorship list.			
7 th author			I agree to the proposed new authorship shown in section 4 /and the addition/removal* of my name to the authorship list.			

Please use an additional sheet if there are more than 7 authors.

Important information. Please read.

- Please return this form, fully completed, to Springer Nature. We will consider the information you have provided to decide whether to approve the proposed change in authorship. We may choose to contact your institution for more information or undertake a further investigation, if appropriate, before making a final decision.
- By signing this declaration, all authors guarantee that the order of the authors are in accordance with their scientific contribution, if applicable as different conventions apply per discipline, and that only authors have been added who made a meaningful contribution to the work.
- Please note, we cannot investigate or mediate any authorship disputes. If you are unable to obtain agreement from all authors (including those who you wish to be removed) you must refer the matter to your institution(s) for investigation. Please inform us if you need to do this.
- If you are not able to return a fully completed form within **30 days** of the date that it was sent to the author requesting the change, we may have to withdraw your manuscript. We cannot publish manuscripts where authorship has not been agreed by all authors (including those who have been removed).
- Incomplete forms will be rejected.

Astroglia

(20 - 40% of all glial cells in human CNS)

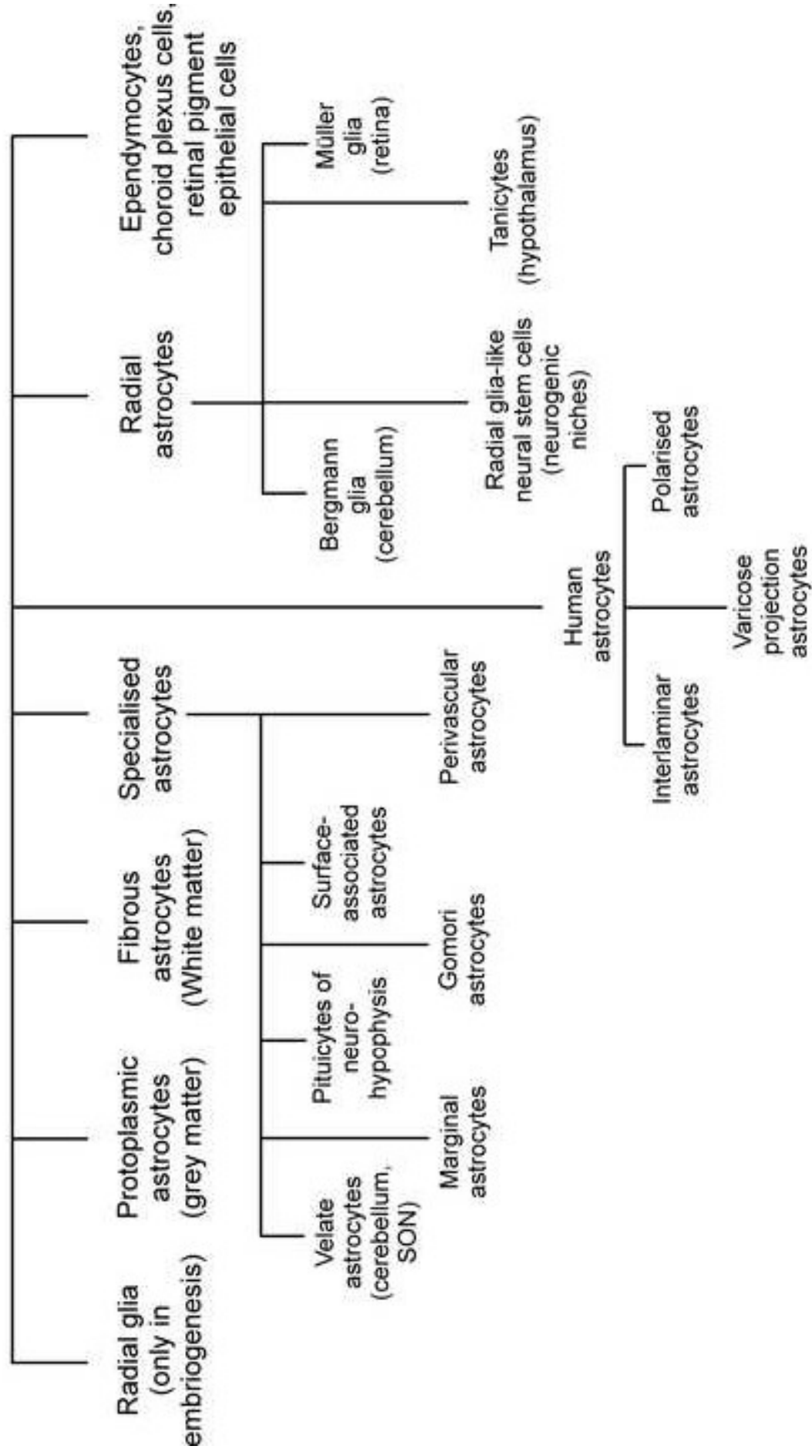


Fig. 1

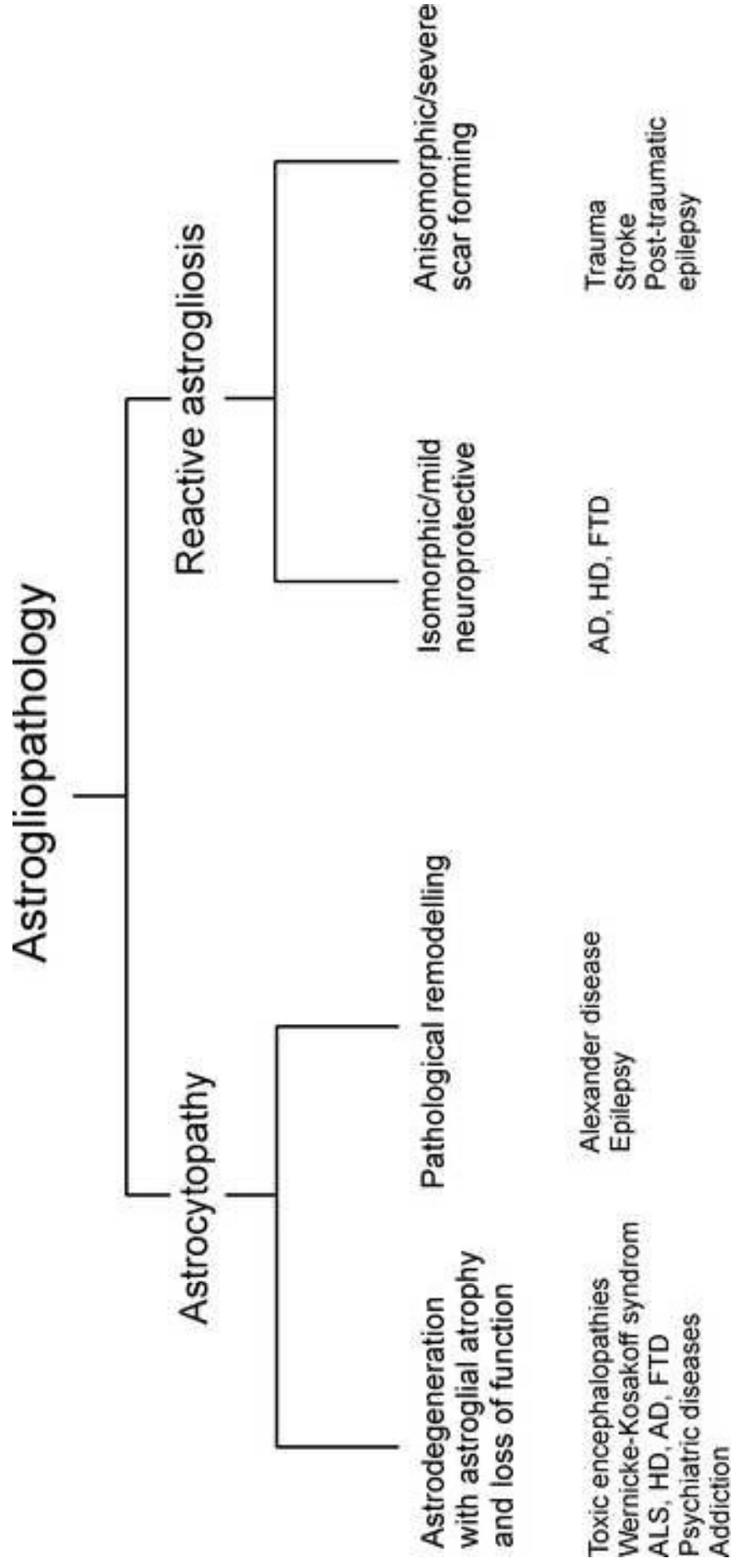


Fig. 2

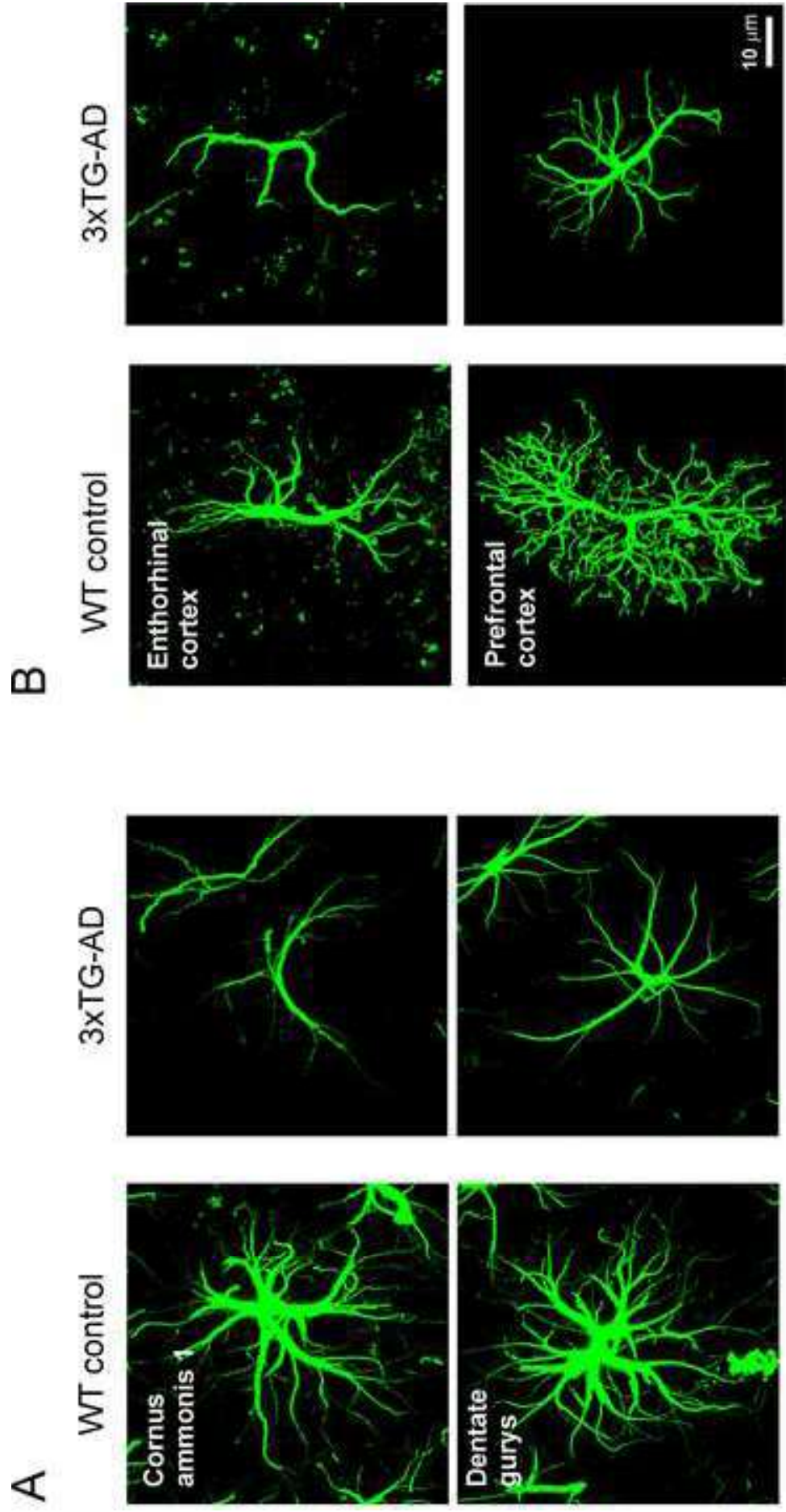


Fig. 3

