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## **ARS Forum Review**

## Inflammation, lipid (per)oxidation and redox regulation

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## Running Head - Lipid (per)oxidation in inflammation and ageing

## Abstract

*Significance:* Inflammation increases during the ageing process. It is linked to mitochondrial dysfunction and increased ROS production. Mitochondrial macromolecules are critical targets of oxidative damage; they contribute to respiratory uncoupling with increased ROS production, redox stress, and a cycle of senescence, cytokine production and impaired oxidative phosphorylation. Targeting the formation or accumulation of oxidised biomolecules, particularly oxidised lipids, in immune cells and mitochondria could be beneficial for age-related inflammation and comorbidities.

**Recent Advances:** Inflammation is central to age-related decline in health and exhibits a complex relationship with mitochondrial redox state and metabolic function. Improvements in mass spectrometric methods have led to the identification of families of oxidised phospholipids, cholesterols and fatty acids that increase during inflammation and which modulate Nrf2, PPARy, AP1 and NFkB redox sensitive transcription factor activity. The kinetic and spatial resolution of the modified lipidome has profound and sometimes opposing effects on inflammation, promoting initiation at high concentration and resolution at low concentration of oxidised phospholipid.

*Future Directions:* There is an emerging opportunity to prevent or delay age-related inflammation and vascular co-morbidity through a resolving (oxy)lipidome that is dependent on improving mitochondrial quality control and restoring redox homeostasis.



Innovation: (Per)oxidised lipids are central mediators in the initiation and resolution of inflammation. Generalised targeting of lipid peroxidation with antioxidants such as tocopherols as a primary <text><text><text> prevention in age-related, inflammatory vascular disease has not proven successful. A more nuanced view is emerging where ROS/NO production is desirable during early phases of inflammation and for resolution. Indeed, the cell-targeted amplification of ROS in neutrophils may be required during inflammation to promote resolution. Phytochemicals that promote resolving oxylipid mediators and improve mitochondrial quality control merit further investigation as inhibitors of underlying sterile inflammation and to mitigate age-related vascular disease.

Key words: anti-inflammatory, oxidised phospholipids, oxysterols, metabolism, eicosanoids, reactive oxygen species

## 1. Introduction

Inflammation is an important physiological process that ensures homeostasis and survival. But as we age, inflammation is less well-controlled and may come at a cost. Usually, physiological inflammation is triggered by non-self materials that activate surveillance systems (complement and resident immune cells). The chemotactic mediators produced by activated complement and resident cells recruit nicotinamide adenine dinucleotide phosphate oxidase (NADPH) oxidase (NOX)-dependent reactive oxygen species (ROS)-producing monocytes, macrophages and neutrophils to kill the invaders. This is followed by lymphocyte recruitment and finally the switch-off of inflammation, in a process known as resolution. Figure 1 illustrates the key effector molecules and different cells involved during the phases of physiological inflammation. Central to the control of physiological inflammation is the metabolic inter-dependence of activation and resolution, and their redox sensitivities (72). Mitochondria are central to inflammation with associated co-morbid diseases.

This review considers the mechanistic relationship between modified lipids, produced enzymatically and by free radical reactions during inflammation, and mitochondrial metabolism, the inflammasome and the resolution phase of inflammation. We consider the impact of ageing and age-related vascular diseases on lipid oxidation and mechanisms by which the phytochemical classes, flavonoids and oxocarotenoids, may mitigate inflammation by preserving mitochondrial function and enhancing nitric oxide (\*NO) availability.

## 2. Physiological inflammation and redox regulation

The initial lines of defence towards invading pathogens are non-specific circulating proteins of the complement cascade combined with tissue resident cells of the innate immune system such as macrophages and dendritic cells. The complement cascade proteins will release chemotactic fragments following contact with unexpected damage-associated molecular pattern molecules (DAMPs) on pathogens (155). A well-known DAMP is bacterial lipopolysaccharide, LPS, that is used widely in studies of inflammation. LPS binds to Toll-like receptor (TLR)4 triggering endocytosis and NOX2 activation with associated production of superoxide anion radicals by neutrophils and macrophages (90). The tissue-resident macrophages are present in low numbers throughout the body, ready to perform a random surveillance role and initiate physiological inflammation. To achieve rapid deployment, innate immune cell activity is tightly coupled to differential use of substrates for energy and the metabolic phenotype (72). While inflammatory macrophages are glycolytic, rapidly producing ATP, alternatively activated anti-inflammatory macrophages are polarised toward mitochondrial biogenesis, oxidative phosphorylation and fatty acid oxidation (167). The high degree of metabolic plasticity of these cells enables them to mount a rapid response to infection and damage (159) and then switch to resolution.

Intertwined metabolic pathways play a major role in immune function and inflammation. To enable switching between different substrate sources according to inflammatory environmental triggers, glucose, amino acid and fatty acid metabolism are coordinated by the activity of redox sensitive transcription factors, nuclear factor erythroid 2-related factor 2 (Nrf2), peroxisome proliferator-activated receptor gamma (PPARy) and hypoxia-inducible factor 1-alpha (HIF1 $\alpha$ ) (72,159,191). It is lipid peroxidation products that are electrophilic activators of these transcription factors.

Oxidative damage to lipids, proteins and DNA is frequently observed during inflammation. Extracellular oxidised lipids are ligands for TLR4, either directly activating receptor or acting as a competitor for LPS and inhibiting receptor activation. Mitochondrial DNA is also recognised as an endogenous DAMP by the intracellular receptor, TLR9 (129). DAMPs also activate local resident innate macrophages, which possess a series of pattern recognition receptors (PRRs) including TLR4 and nucleotide-binding oligomerization domain (NOD)-like receptors (NLR) that are highly conserved (46), to produce chemotactic cytokines and activate the nucleotide-binding domain, leucine-rich-containing family, pyrin domain-containing-3 (NLRP3) inflammasome (78). In addition to being activated by oxidised lipids, the NLRP3 inflammasome can also be activated by endogenous metabolic DAMPs including many different native lipids; fatty acids, ceramides and free cholesterol (210,226,231).

The cells that respond most rapidly to infection and the emerging chemotactic gradients are neutrophils which phagocytose extracellular pathogens, killing them via NADPH oxidase-dependent ROS production within the phagosome (61). To increase their efficiency in pathogen capture, neutrophils also produce neutrophil extracellular traps (NETs) from mitochondrial DNA. NADPH oxidase -dependent ROS production and formation of NETS are also essential for the resolution of necrosis-induced inflammation (18) (Figure 2). Neutrophil nets, together with proteins within the activated complement and the clotting cascades immobilise pathogens and provide a physical barrier, preventing infection from spreading around the body.

While NADPH oxidase activation is essential for adequate NET production (61), ROS production causes non-specific bystander oxidative damage. Neutrophils themselves will die rapidly due to low antioxidant enzyme levels within 24 hours of recruitment; they are not equipped to survive under conditions of oxidative stress and this is exacerbated with age (92).

Monocytes and lymphocytes appear later than neutrophils and within 24 hours. The monocytes display a range of phenotypes and functions that are dependent on the local environment; for example, if concentrations of DAMPS and pro-inflammatory cytokines are high, monocytes differentiate to macrophages and adopt an M1-type of pro-inflammatory highly migratory phenotype to target infection to distant sites. This phenotype favours further NADPH oxidase activation with superoxide anion radical production, expression and activity of nitric oxide synthases (NOS) with associated nitic oxide production (although uncoupled NOS which is more common during ageing increases peroxynitrite ONOO<sup>-</sup> production) as reactive nitrogen species (RNS) and the production of inflammatory mediators. M1 type macrophages depend on glycolysis to meet the metabolic requirement for rapid energy production (Figure 3). The pentose phosphate pathway is also upregulated under oxidative stress and the presence of lipid peroxides in M1 macrophages via the Nrf2 pathway (128). This increases availability of NADPH as an essential reducing agent to restore the antioxidant glutathione and redoxin cycles. In an Nrf2 dependent process, the antioxidant enzymes mitochondrial superoxide dismutase-2 (SOD2), glutathione reductase, thioredoxin reductase and peroxiredoxin 1 but not catalase are upregulated in macrophages, enhancing their chance of survival during a high ROS environment (151). Neutrophils do not have this extent of adaptation and antioxidant activity. Instead, after killing pathogens, neutrophils die, newly recruited and resident macrophages sense the presence of apoptotic neutrophils via surface receptors such as CD36 and intercellular adhesion molecule 3, and then clear them in a non-inflammatory phagocytic process

(203). Local tissue macrophages may also "cloak" the damaged site to prevent further neutrophil recruitment (207).

Resolution of inflammation was once thought to be a passive process that simply 'burns out' with time, allowing for tissues to return to homeostasis. However, resolution of inflammation is now recognised as an active programme that involves sequential synthesis of lipid mediators of inflammation (described in Section 4)(178,180). To meet the demands for degradation of invaginated material and remodelling the extracellular environment to promote healing, macrophages adopt mitochondrial-dependent oxidative phosphorylation and secrete anti-inflammatory lipids formed by an enzyme-catalysed oxidation reaction from the n-3 fatty acids and transforming growth factor beta (TGF $\beta$ ), enabling the resolution of normal physiological inflammation (72). The enzyme-catalysed oxidation of n-3 fatty acids leads to the formation of protectins, maresins and resolvins (D- and Eseries) which normally resolve acute inflammation and prevent chronic inflammation from developing. They affect macrophage differentiation (explained in more detail in section 4 and illustrated in Figure 5). Interestingly, administration of resolvin D3 was also able to reduce joint inflammation in an arthritis model, suggesting that resolvins may also reverse and resolve chronic phase of inflammation. D- and E-series resolving effects are not limited to the innate immune system; they may also target recruited T and B cells at an inflammatory site. In parallel, the recruited lymphocytes (T cells normally) will recognise ROS-regulated, enzymatically-trimmed foreign molecules that have been produced from a pathogen (141). Processed antigens are carried on the surface of dendritic antigen presenting cells within a specialised protein, within the major histocompatibility class, that enables the antigen to be recognised as foreign by T cells. T cells in turn are activated, switch to glycolysis and promote the adaptive immune system to produce foreign-antigen specific antibodies on future contact with antigen and enable a specific targeted immune response to be elicited (204).

Dysregulation at each phase of inflammation may lead to a process that has been termed sterile inflammation which becomes more common with age when there is no pathogenic stimulus; non-resolution of inflammation is common during age co-morbidities.

## 3. Sterile inflammation and ageing

During ageing, the carefully orchestrated acute inflammatory response that enables effective recognition of tissue damage and removal of pathogens is less well regulated. A process described as "sterile" inflammation is frequently seen and is manifested in specific organs and tissue e.g. in the vasculature during atherosclerosis. Atherosclerosis is a maladaptive phenotype that promotes immune cell recruitment activation and impairs resolution, can accelerate ageing and age-related comorbidities, and will be considered further.

In addition to the production of ROS by NADPH oxidase, a significant increase in mitochondrial ROS production has been observed during ageing possibly due to ineffective mitophagy (196). Some have reported that overproduction of mitochondrial ROS may increase inflammation e.g. in atherosclerosis (223). Mitochondrial ROS generated by myeloid cells within atherosclerotic plaques increase the concentration of the free-radical mediated cholesterol oxidation product, 7-ketocholesterol (7-KC), which is the most abundant oxysterol in low density lipoprotein (LDL), and enhance the formation of atherogenic neutrophil NETs (224). An exaggerated "sterile" inflammatory response is generated to cholesterol-rich lipoproteins that are retained in the arterial wall in the absence of any infection or tissue damage (186). Shirai et al have identified that circulating monocytes from atherosclerosis

patients are primed to produce more inflammatory cytokines via mitochondrial ROS-mediated oxidation of the glycolytic enzyme pyruvate kinase M2 (188). ROS play a role in regulating proinflammatory priming of monocytes in response to oxidised lipids through epigenetic and metabolic re-programming e.g. by protein acetylation (191). For effective priming and memory of an inflammatory event, the acetylation of macrophage histone proteins is rate limiting according to the availability of one metabolite, acetyl CoA (formed in abundance by during lipid metabolism), that is consumed by mitochondria. These observations highlight that lipids are essential sources of energy and are increasingly recognised as important regulators of inflammatory signalling.

Histone deacetylation is necessary to switch macrophage activity towards resolution of inflammation. The associated modulation of gene expression is closely regulated by the activity of sirtuin (SIRT) proteins and availability of the reducing agent nicotinamide adenine dinucleotide (NADH); this again provides another link between inflammation and redox state, as the availability of NADH is controlled by mitochondrial activity. Macrophage SIRT3 plays an essential anti-inflammatory role through regulating mitochondrial bioenergetics and redox homeostasis as well as controlling activation of the inflammatory protein complex, known as the inflammasome (97). The NLRP3 inflammasome has emerged as an immune sensor that causally links systemic inflammation to ageing (230). It is responsible for the proteolytic activation of the inflammasome is organised following activation at the mitochondria and this is achieved through its binding to cardiolipin, a highly abundant phospholipid lipid in mitochondrial membranes (50) (84).

An increase of ROS/RNS production by immune cells is observed with age due to mitochondrial uncoupling and sterile inflammation and causes lipid peroxidation. There are many classes of lipids that may be oxidised or nitrated. These include phospholipids and cardiolipin, sterols and fatty acids. Oxidised cardiolipins are found circulating at higher concentrations during ageing and are proinflammatory (220). It remains to be seen whether oxidised cardiolipin accumulates in mitochondria with age and whether this influences inflammasome activity. Mass spectrometry methods are now becoming sufficiently sensitive to measure oxidised lipids in subcellular organelles to address this question.

Over the following sections we review three principle classes of oxidised lipids; 1) oxidised phospholipids, such as those formed by free radical oxidation of 1-palmitoyl-2-arachidonyl-sn-glycero-3-phosphorylcholine (PAPC), that have been shown to modulate signalling induced by bacterial lipopeptide or (LPS, (51,52,117); 2) oxysterols produced by free radical reactions or enzymaticallyproduced cholesterol oxidation, which amongst other inflammatory effects, can regulate the activation of NLRP3 inflammasome by LPS (37); and 3) the products of 12/15 lipoxygenase (12/15 LOX); specialised pro-resolving mediators e.g. lipoxins and resolvins, acting on G protein coupled receptors (GPCRs). These lipid per/oxidation products are stable and can exert distant sites from their site of production. This raises the potential for oxidised lipids to exert distant effects of ROS on inflammation in a time and concentration dependent manner.

We have previously described increased levels of lipid peroxidation products in the age-related diseases rheumatoid arthritis, Alzheimer's disease and cardiovascular disease (38-40,71) and atherosclerosis is a common comorbidity in all conditions. Since mitochondria have an important role in proinflammatory signalling role during ageing (69) we consider in the following sections how a vicious cycle of lipid (per)oxidation may perpetuate mitochondrial dysfunction and inflammation.

## 3.1 Peroxidised phospholipids, nitrated fatty acids, inflammation and age-related vascular disease

A. Oxidised phospholipids (OxPLs)

Phospholipids have structural roles in cell membranes and while maintaining membrane integrity they may also function as second messengers (3,23). Phospholipids are substrates for synthesis of lipid mediators including phosphoinositides, diacylglycerides, platelet-activating factor, sphingosine-derived phospholipids, phosphatidic acids, and eicosanoids (135).

Phospholipid bound polyunsaturated fatty acids (PUFA) such as arachidonic acid (AA), docosahexaenoic acid (DHA) present in the second position of the glycerol backbone of phospholipids are also major targets for non-enzymatic oxidation by ROS/RNS to produce the primary oxidation products, peroxyl radicals and hydroperoxides (23). Subsequent molecular rearrangement, cyclisation or fragmentation and oxidation yield a diverse pool of oxidised phospholipids (OxPLs). This includes full-length or shortened carbon chains containing oxygen functional groups such as hydroxy-, keto-, epoxy, hydroperoxyl- and prostane groups as well as carboxylic and aldehydic terminal groups (23). In addition to being esterified within phospholipids, esterified oxidised PUFAs can also be detected as cholesterol esters and triglycerides. All exert distinct biological effects compared to their reduced parent molecules in part due to increased polarity but also due to their chemical reactivity (22). The source of oxidants predicts the likely subcellular targets e.g. mitochondrial DNA and cardiolipin are readily oxidised as mitochondria become more uncoupled, and the nature of ROS/RNS informs the chemistry of oxidation and the products that are formed.

Several OxPL species may result from one individual precursor (48). For example, PAPC (the major phospholipid) oxidation results in oxygenated full-length products e.g. epoxyisoprostane (PEIPC) and epoxycyclopentenone (PECPC) as well as fragmented or truncated products. Truncated PAPC products include 1-palmitoyl-2-(5-oxovaleroyl)-sn-glycero-phosphatidylcholine [POVPC] and 1-palmitoyl-2-glutaroyl-sn-glycero-phosphatidylcholine [PGPC] and have carbonyl groups that readily form adducts with other biomolecules especially proteins (23,153). OxPLs can bind covalently to lysine, histidine, arginine and cysteines via Schiff bases or as Michael adducts as well as to the amine groups of other phospholipids. Improvements in MS methodologies have led to the identification of membrane and cytosolic targets of proteins in cultured mouse macrophages that may be modified by OxPLs; these included proteins involved with apoptosis, stress response, lipid metabolism and transport (194).

Using model membranes, OxPLs with an aldehyde or a carboxyl group at their truncated sn-2-chain end e.g. POVPC, PGPC were shown to be released rapidly, to be inflammatory and to increase local membrane permeability whereas oxygenated full-length products PEIPC and PECPC were released more slowly and stabilised membranes (80). This led the authors to suggest that early truncated oxidation products may promote vascular leakiness associated with inflammation whereas the release of full length oxygenated products, perhaps as ROS levels reduce during later phases of inflammation, may support resolution. In support of this hypothesis, the phospholipase catalysed release of 5,6epoxyisoprostane that is present in PEIPC has been shown to mimic the effects of pro-resolving prostanoids, through Michael addition to essential Kelch Like ECH Associated Protein 1 (KEAP-1) thiols, facilitating Nrf2 activation. Together these studies illustrate how OxPL are involved in induction and resolution of inflammation (24,234). OxPLs present on oxidised lipoproteins, senescent and apoptotic cells are pattern associate molecular patterns (PAMPs) that are recognised and removed by soluble and cell-associated PRR, including scavenger receptors such as CD36, natural (germ line-encoded) antibodies, vascular endothelial growth factor receptors on endothelial cells and C-reactive protein (23). Downstream signalling pathways in endothelial and mononuclear cells include receptor tyrosine kinase and MAP kinases, that signal through intermediates to promote inflammation by regulating Activator Protein 1 (AP1), NFκB, PPARy and Nrf2 pathways leading to the increased expression of cytokines and chemokines including TNFα, IL-1β, IL-6, IL-8, MCP-1, MIP2, the lipid oxidation enzyme cyclooxygenase (COX)-2, and the homing receptors CCR1, CCR2 and CCR5 (22). In endothelial cells, a high concentration of oxidised PAPC was shown to disrupt barrier function due to increases in intracellular ROS and downstream activation of Src (193) that catalysed the phosphorylation of the adherens junction protein vascular endothelial cadherin (VE-cadherin) at Tyr-731 and Tyr-658. This was not observed in endothelial cells treated with low OxPAPC concentrations, which conversely showed potent barrier protective effects (193) (19). Additionally, OxPCs with different chain lengths also show opposite effects on barrier function where short chain OxPCs are reported to have disruptive barrier function whilst long-chain OxPC induce protective barrier function (19,60,117).

Elevated concentrations of fragmented OxPLs exert proinflammatory effects in monocytes and promote foam cell formation but, in contrast to LPS, do not activate granulocytes. Similar to the effects of OxPL on endothelial cells, it has been suggested that proinflammatory effects of OxPLs on monocyte/macrophages may be observed more frequently in sites of local to lipid accumulation sites (136). In contrast, at distant sites and with lower concentrations, OxPL may be anti-inflammatory, possibly by competing with PAMPs for TLR4 binding. Consistent with this, it has also been shown that when mouse macrophages are treated with OxPLs, they undergo differentiation resulting in a phenotype that is less phagocytic and with higher Nrf2 activation. However, these authors also fractionated the OxPLs and observed that truncated species of OxPAPC induced reprogramming of macrophage metabolism (increased *Glut1* expression, the major glucose transporter) to support antioxidant gene expression (e.g. haemoxygenase 1, *Ho1* thioredoxin reductase, *Txnrd1*; and *Gclm*, glutamate cysteine ligase), while the full-length OxPAPC treated macrophages showed increased expression of cytokines and chemokines e.g. *Il-16*, *Il-6*, and *Cxcl1* genes encoding interleukin-1β, interleukin-6 and the chemokine (C-X-C motif) ligand 1 (174,175).

The increased production of IL-1  $\beta$  by OxPAPC-treated monocyte macrophages and endothelial cells indicates activation of the inflammasome by OxPAPC. Again pleiotropic effects of OxPAPC on inflammasome activation are described; it has been shown by some authors that OxPAPC binds directly to caspase-4 and caspase-11, competes with LPS binding, and consequently inhibits LPSinduced pyroptosis, IL-1 $\beta$  release and septic shock (32), whereas others have shown that POVPC injection resulted in the production of caspase-1, IL-1 $\beta$ , and IL-18 in wild-type, but not in NLRP3deficient, mice. Furthermore, POVPC-induced inflammasome activation was dependent on an increase in mitochondrial ROS; ROS were increased following POVPC-mediated increases in intracellular Ca<sup>2+</sup> signaling and mitochondrial destabilisation (229), after transient POVPC stimulation of transient receptor potential channels (7). Collectively, these studies illustrate that OxPL are effectors of bioenergetic switch, mitochondrial ROS production and altered redox state, potentially at distant sites, and their local concentrations may influence whether a pro- or anti-inflammatory phenotype predominates.

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## B. Isoprostanes

Prostaglandin  $D_2$  and prostaglandin  $E_2$  GPCR receptors are important targets for prostanoid-like products formed from OxPAPC and PEIPC, but not POVPC (70). Common stable products of nonenzymatic free radical attack on phospholipids are isoprostane-like structures and chain-shortened products containing carboxylic acid groups, carbonyl and hydroxyl groups. These include arachidonic acid-derived F2 $\alpha$ -isoprostanes and other isoprostanes from  $\alpha$ -linolenic, eicosapentaenoic and docosahexaenoic such as F1-phytoprostanes, F3-isoprostanes and F4-neuroprostanes respectively (212). F2 $\alpha$ -isoprostanes regulate vasoactive, mitogenic and inflammatory properties but also may have inhibitory action via cAMP signalling from the thromboxane A2 receptor (TBXA2R) on platelets (89). Formation of  $F2\alpha$ -isoprostanes is significantly increased in age-associated diseases such as obesity, diabetes and atherosclerosis (16,195). F2 $\alpha$ -isoprostanes may also compete with receptors to inhibit drug action – it has been suggested that aspirin-insensitivity in cardiovascular diseases may be due to TBXA2 prostanoid receptor activation by F2 $\alpha$ -isoprostanes (16). The rearrangement of F2 $\alpha$ endoperoxide intermediates results in the formation of  $D_2/E_2$ -isoprostanes which can undergo further rearrangements generating the cyclopentenone A/J-isoprostanes (77). In contrast to the inflammatory effects of isoprostanes, the cyclopentenone deoxy-A2/J2-isoprostane-phosphocholines 15d-PGJ2 and 15d-PGJ2-PC, formed from 1-palmitoyl-2-arachidonoyl-sn-glycero-3-phosphocholine, prevent foam cell formation, induce anti-inflammatory and anti-oxidant responses in macrophages through targeting redox sensitive thiols within the NF-κB, PPARy and Nrf2 pathways (77,175). Some suggest that it is Nrf2 that is largely responsible for the anti-inflammatory actions of oxidised lipids (59).

Table 1 summarises the biological effects of OxPL and isoprostanes on inflammation and atherosclerosis, illustrating both pro- and anti-inflammatory effects (110).

## C. Nitrated fatty acids

RNS derived from 'NO, including ONOO-, nitrogen dioxide ('NO<sub>2</sub>) and nitrous oxide can also oxidise and nitrate unsaturated fatty acids via the homolytic addition of ('NO2) to a double bond, to form nitrated lipids (21,132). In addition, 'NO terminates peroxyl radical-induced chain propagation reactions of lipid peroxidation at a faster rate than tocopherol. Nitration specific products from arachidonic, linoleic and oleic acids include nitroalkenes, nitro-nitrile esters, nitrohydroxy species and β-nitroalkyl radicals. In addition, hydrogen abstraction may also occur, producing similar fragmented peroxidation products to oxygen radical species attack (21,108). Nitrated fatty acids were first described in 2003 and are considered to be protective in the vasculature; they are the subject of several reviews (91,206). They have potent electrophilic activity, forming adducts with proteins though Michael addition reactions (8,170). Their formation has been reported in the plasma membrane, mitochondria, lipoproteins and triglycerides and they may be transported to distant sites within lipoproteins (54). Free nitro-fatty acids are found at very low concentrations in the circulation, probably released by phospholipases (162). They can target extracellular receptors through nucleophilic attack and may be taken up in the esterified form by scavenger receptors. Intracellular nitrated fatty acids contribute to reversible and exchangeable nitrated fatty acid-thiol adducts occur under biological conditions (162) and are effective inhibitors of NF-kB; alkylation of macrophage p65 by nitroalkene fatty acids inhibits NF-kB DNA binding activity and represses downstream inflammatory target gene expression (36). Nitrolipids are also potent agonists of PPARy and Nrf2, binding to thiol on

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KEAP1 and releasing Nrf2 which directs antioxidant and anti-inflammatory and metabolic gene expression in monocytes/macrophages and endothelial cells. There are intrinsically different potencies of regioisomers of nitro fatty acids (8). In contrast to ROS-inducing activity of OxPLs, nitrolipids have been reported to inhibit activation of the NADPH oxidase via nucleophilic attack on thiol residues in p47 Phox (109), possibly involving the formation of a covalent Michael adduct between NO<sub>2</sub>-AA and critical nucleophilic residues (*e.g.* histidines or cysteines) preventing the formation of the active complex. Within mitochondria, mild uncoupling by nitrolipids has been described which in turn promoted adaptation during ischemia and reperfusion, a sterile inflammatory response involving production of ROS (126,127), and cardioprotection.

To date there are no reports describing any change of nitrolipids with age, however, it is known that endothelial NOS efficiency decreases with age due to uncoupling (44), potentially reducing the rate of anti-inflammatory nitro-lipid formation and increasing ONOO<sup>-</sup>-mediated lipid peroxidation. Furthermore, our previous study has shown that the median concentration of plasma linoleic acid is reduced by 50% in healthy adults over 50 years, reducing availability of a major substrate for nitration by •NO (145). The in situ generation of nitrolipids within mitochondria highlights their significant role for coordinating the regulation of metabolism and inflammation, driving an anti-inflammatory phenotype (213), and efforts to increase eNOS coupling may contribute to anti-inflammatory activity via nitrolipid formation to mitigate risk and severity of inflammageing and atherosclerosis.

#### 3.2 Oxysterols, inflammation and age-related vascular disease

ROS oxidise the unsaturated rings of cholesterol yielding several oxysterols that differ in the position of hydroxyl group addition. In addition to free radical mediated oxidation, some oxysterols are formed by specific enzymatic oxidation reactions to produce short-lived intermediates that are involved in cholesterol excretion pathways; they may also be taken up in the diet (Figure 4). Compared to highly abundant cholesterol that circulates in blood at millimolar levels, oxysterols are reported to found in 1,000-10,000 times lower concentrations around micromolar to submicromolar levels in human plasma (40). Regardless of the source or the concentration, oxysterols are taken up by a range of receptors (LDL receptors, EBI1; Epstein-Barr-virus-induced G-protein coupled receptor 2, also known as GPR183) , scavenger receptors, G-protein coupled receptor 17 GPR17, C-X-C Motif Chemokine Receptor 2 CXCR2, glucocorticoid receptor, purinergic P2X7 receptor, Smoothened, Frizzled Class Receptor SMO, Glutamate Ionotropic Receptor NMDA Type NMDA, NPC Intracellular Cholesterol Transporter 1 NPC1; see Figure 4) and exert a range of biological effects. At high concentrations, oxysterols are cytotoxic and pro-atherogenic compared to cholesterol, especially in vascular cells.

Similar to other oxidised lipids, addition of a hydroxyl, epoxide, hydroperoxyl, carboxyl or ketone moieties to the steroid nucleus or to the side chain, increases the polarity of oxysterols (20,73). To date, the subcellular distribution of oxysterols has not been characterised, although increased concentrations are predicted to occur in mitochondria during ageing, which may influence cellular metabolism. In membranes, a change in behaviour in micro environments by oxysterols has been shown to affect membrane fluidity (55), cell signalling and metabolism (27,68). Oxysterols are stable species that are found within lipoproteins or free in serum; they can diffuse within LDL to distant sites,

may cross the blood brain barrier (42,111,200) and are taken up by discrete receptors to influence cell cholesterol and mitochondrial metabolism (Figure 4).

Enzymatically produced oxysterols have been studied in relation to health and vascular diseases. The cytochrome P450 family gene, *CYP27A1*, encodes a mitochondrial enzyme which catalyses the formation of 27-hydroxycholesterol (27-OHC) and modulates the acidic biosynthetic pathway for bile acids. 27-OHC is the most prevalent enzymatically produced oxysterol in human circulation ranging from 150- 730 nM but increases more than two fold in atherosclerotic lesions (25). As competitors for binding estrogen receptor  $\alpha$ , atherogenic properties of 27-OHC have been attributed to the attenuation of estrogen-related atheroprotection (208). Additionally, 27-OHC induces adverse effects in the brain by passing through the blood-brain-barrier and interrupts local cholesterol homeostasis (120).

We have shown previously that 27-OHC alters the distribution of membrane microdomains in neuronlike cells to increase the activity of beta secretase to produce amyloid beta (41) in vitro. In neurones, cholesterol is converted into 24S-hydroxycholesterol (24-OHC). Excess 24-OHC crosses the bloodbrain-barrier to reach the blood and is metabolised by the liver, converted into biliary acids and eliminated into the bile. Both 24-OHC and 27-OHC increase proportionally to the number of e4 alleles in individuals with cognitive decline and a positive correlation has been reported for APOE genotype and 24-OHC levels in cerebrospinal fluid from patients with Alzheimer's disease and mild cognitive impairment (143). Another prospective ageing study reported that lower cholesterol present in ApoE epsilon 4 carriers was related to a higher rate of decline of information processing speed and a higher ratio of 27-OHC to cholesterol was related to a lower level of general performance and memory functioning. The authors concluded that lower total cholesterol and high oxysterol levels may be considered as a frailty marker, predictive of lower cognitive functioning in the elderly (209).

Both 27-OHC and 25-hydroxycholesterol (25-OHC) regulate immune cell function. 25-OHC is synthesised by the enzyme cholesterol 25-hydroxylase. Initial studies by Park and Scott revealed that 25-hydroxylase gene expression is low in resting macrophages under standard cell culture conditions but is rapidly induced by two orders of magnitude when cells are activated with TLR ligands (146). Other oxysterols,  $7\alpha$ -OHC and  $7\alpha$ , 25-dihydroxycholesterol ( $7\alpha$ , 25-OHC) are reported as a high-affinity ligands for the inflammatory receptor EBI2/ GPR183 that is induced in B cells upon viral infection (149).

In addition to a direct antiviral role, 25-OHC also regulates transcription of inflammatory genes. 25-OHC amplified the TLR-induced gene expression through a positive feedback response mediated, at least in part, via the transcription factor, AP1 (67). The induction of pro-inflammatory cytokines by 25-OHC is also dependent on increased NF $\kappa$ B activation, most likely following the activation of p38 MAPK and JNK (185). Umetani et al reported the activation of endothelial and macrophage NF $\kappa$ B pathway by 27-OHC (208). Peritoneal macrophages from 27-OHC treated mice upregulated mRNA level of TNF- $\alpha$ , IL-1 $\beta$  and IL-6 by 3 to 6 fold and promoted monocyte-endothelial cell adhesion through estrogen receptor (ER)- $\alpha$  driven pathway (208). Several closely related sterol regulatory transcription factors are regulated by oxysterols including LXR, RXR, ROR $\alpha$ , ROR $\beta$ , ROR $\gamma$ , ER $\alpha$ . Some oxysterols are able to induce metabolic reprogramming upon uptake; they are involved in polarising M1 to M2 macrophages through the LXR transcription factor (76). In this process, the oxysterol activated LXR pathway drives macrophages to recognise dead cells, promotes cholesterol efflux and down regulates inflammatory

gene expression, further illustrating the relationship between oxidised lipids, metabolism and inflammation.

Anti-inflammatory properties of oxysterols have also been reported; Vurusaner et al have shown that low micromolar concentrations of 27-OHC can activate the redox sensitive transcription factor, Nrf2 and subsequently the target genes, HO-1 and NQO-1 in monocyte-like cell line, U937 (217). Early and transient generation of ROS levels by 27-OHC enhanced MEK-ERK/PI3K-Akt phosphorylation, which in turn reduced the subsequent ROS production suggesting the ability of oxysterols as pro-survival inducers at low concentration (217,218). In the same cell line, 27-OHC activated an autophagic response by expressing upregulated microtubule-associated protein 1A/1B-light chain 3 (LC3) II/LC3 I ratio and Beclin 1 levels in a MEK-ERK/PI3K-Akt dependent manner (219).

To study the short term effect of free radical mediated cholesterol oxidation products acquired from the diet, Vine et al analysed non-enzymatically produced  $7\beta$ -hydroxycholesterol ( $7\beta$ -OHC), 7- KC and  $5\alpha$ ,  $6\alpha$ -epoxycholesterol after two weeks of feeding rabbits with chow supplemented with oxidised cholesterol (containing 6% oxysterols) (214). The authors did not observe any changes to  $7\beta$ -OHC levels but five times higher levels of  $5\alpha$ ,  $6\alpha$ -epoxycholesterol and double the levels of 7-KC were found in triglyceride-rich lipoproteins from oxidised cholesterol-fed animals compared to the purified cholesterol-fed animals. The oxidised cholesterol-fed animals also had a 64% increase in total aortic cholesterol. This is consistent with the observation that  $7\alpha$ -OHC,  $7\beta$ -OHC and 7-KC are found in relatively high concentration in foam cells and fatty streaks (26). Foam cells are formed after oxidised lipid loading into recruited macrophages; and it is estimated that oxysterols comprise up to 50% of total sterol content of OxLDL-loaded cells (26). The underpinning mechanism of plaque formation is likely to be via 7-KC enhanced leukocyte-endothelial interactions (199). In an attempt to prevent foam cell formation, macrophage lysosomal lipase hydrolyses the ingested cholesteryl esters. In concert, 25-OHC is synthesised, probably in mitochondria where it maintains membrane integrity, and prevents mitochondrial ROS production and NLRP3 inflammasome activation. Lysosomal lipase activity is also required for LXR-mediated activation of cholesterol efflux (Figure 4) (211); free intracellular cholesterol is transported to the mitochondria via steroidogenic acute regulatory protein where CYP27H catalyses oxysterol synthesis. 27-OHC is a regulatory oxysterol that not only activates LXR and mediates lipid efflux, but also modulates PPARy activity and suppresses IL-6, TNF $\alpha$  and IL-1 $\beta$ production by macrophages (133). This evidence supports an important role for oxysterols in regulation of inflammation via the mitochondrion.

In summary there is accumulating evidence for a role of oxysterols in regulating inflammation. The biological effects may be pro- or anti-inflammatory according to oxysterol concentration. However, whether they undergo covalent interactions or act as partial agonists/antagonists remains to be determined.

#### 3.3 Eicosanoids, inflammation and age-related vascular disease

The eicosanoids are another group of oxidised lipids that integrate with metabolism, regulating both the initiation and resolution of inflammation (**Figure 5**)(178,180). The immune system sentinels, neutrophils and monocytes/macrophages, require a directional signal both to migrate to the site of tissue injury, during the acute inflammation phase, and back out, removing cellular debris and clearing out invading pathogens without tissue injury. The cardinal 'signals' come in the form of

chemoattractant bioactive lipids that are produced by the coordinated action of receptors, phospholipases and oxygenases. Thus, they are considered to play key roles driving both proinflammatory and pro-resolving responses(179). While they are also electrophilic in nature and are formed through enzyme-controlled free radical reactions, the biological effects are not mediated through covalent modification of targets but rather as conventional ligands for receptors and transcription factors.

Pro-inflammatory lipid mediators are produced and released by the host cells with an aim to create chemotactic gradient that will support transendothelial migration of professional phagocytes to the site of injury. They include eicosanoids, namely leukotrienes (Lts)(147,166), prostaglandins (PGs)(154) and thromboxanes (TXs)(202), Figure 6. These lipid mediators are synthesised from membrane-derived AA within minutes of an acute challenge. Phospholipase A2 (PLA2) hydrolyses phospholipids at the sn2 position to generate fatty acid substrates for oxidation by cyclooxygenases (COX) and lipoxygenases (LOX).

Production of **leukotrienes** from free AA is initiated by the enzyme 5-lipoxygenase (5-LOX). The most potent of all, leukotriene  $B_4$  (Lt $B_4$ ) is produced mainly by neutrophils and pro-inflammatory macrophages. By ligating to G-protein coupled receptor BLT1/2, Lt $B_4$  supports further recruitment and activation of circulating leukocytes to the site of cellular injury(198).

Metabolism of AA by cyclooxygenase (COX) and thromboxane synthase leads to the formation of **thromboxanes**. Thromboxane A<sub>2</sub> (TXA<sub>2</sub>), mainly produced by monocytes and macrophages, exerts its pro-inflammatory actions through binding to the thromboxane receptors TP that belong to the family of G-protein coupled receptors(201). Thromboxane is best known for being a potent platelet aggregant and vasoconstrictor(130).

The **prostaglandin** profile in tissues depends on the activity of COX-1 and COX-2, enzymes with both cyclooxygenase and peroxidase activity(189). During homeostasis COX-1 is the dominant catalyst in formation of prostaglandins. Upon receiving an inflammatory stimulus, the expression of COX-2 becomes upregulated, leading to the synthesis of pro-inflammatory prostaglandin  $E_2$  (PGE<sub>2</sub>), prostaglandin  $D_2$  (PGD<sub>2</sub>) and prostaglandin  $F_{2\alpha}$  (PGF<sub>2\alpha</sub>)(202). Hence, the prostaglandin profile is dependent on both the levels of expression of enzymes responsible for their synthesis and the cell activation state. Indeed, TXA<sub>2</sub> is produce by the resting macrophages, while the synthesis of PGE<sub>2</sub> is favoured upon cellular activation.

The actions of  $PGF_{2\alpha}$ , are mediated by its individual receptors FP,  $PGD_2$  and  $PGE_2$  activate multiple receptors that are differentially expressed in immune cells, thereby triggering immune responses(202). In the context of this review, the most important cellular response mediated by  $PGE_2$  and  $PGD_2$  is the lipid mediator "class switch". While both known for their pro-inflammatory activities(58),  $PGE_2$  and  $PGD_2$  each promote a switch in the upregulation of key enzymes to promote synthesis of dual-acting specialised pro-resolving lipid mediators of inflammation (SPM)(100).

The dual-acting nature of SPM is reflected in their ability to selectively stop further infiltration of neutrophils to the site of acute challenge while supporting pro-resolving futures, such as recruitment and activation of monocytes without pro-inflammatory stimuli (non-phlogistic stimulation), phagocytosis of apoptotic cells and microorganisms, exit of phagocytes via lymphatics and upregulation of antimicrobial agents(177,178).

**Lipoxins**, namely lipoxin  $A_4$  (LxA<sub>4</sub>) and lipoxin  $B_4$  (LxB<sub>4</sub>), are the first SPM to be recognised for their dual-acting nature(176). They are derived from AAin a form of trihydroxy derivatives via two different routes. For example, in mucosal tissue, lipoxins are released due to the interactions between leukocytes and epithelial cells. Here, synthesis of lipoxins is initiated in epithelial cells by the 15lipoxygenase (15-LOX), leading to the formation of 15-hydroxy-eicosatetraenoic acid (15-HETE) intermediate. This substrate is provided to neutrophils for the final synthesis of lipoxins by 5-LOX. In blood vessels lipoxin synthesis is ensured by an interplay between leukocytes and platelets. Leukocyte-derived 5-LOX metabolises AA to form epoxide intermediate LtA<sub>4</sub>. Studies have shown that more than 50% of LtA<sub>4</sub> is further metabolised by platelet 12-lipoxygenase (12-LOX) to produce lipoxins(47,56,160). In macrophages, a unique two-step activation mechanism ensures controlled production of lipoxins; first TLR4 activation results in accumulation of an esterified form of the COX-2-derived lipoxin precursor 15-HETE which is stored within membrane phospholipids. Subsequent activation of P2X7 by extracellular ATP leads to phospholipase A2 activation, hydrolysis of 15-HETE from membranes and its conversion to lipoxins by 5-LOX, linking inflammasome activation to resolution (134). LxA<sub>4</sub> binds to lipoxin A4 receptor (ALX) on leukocytes thereby triggering cell-specific responses. In neutrophils, this interaction will stop neutrophil chemotaxis, adherence(57,101,144) and transmigration into the inflamed sites (86, 184), while in monocytes it stimulates chemotaxis (112) and non-phlogistic responses (66,123). LxA<sub>4</sub>-ALX interaction in T cells blocks secretion of TNF $\alpha$  providing the link between the innate and adaptive immune systems (94).

**E-series resolvins**, namely resolvin E1 (ResE1) and resolvin E2 (ResE2), are derived from 18-hydroxyeicosapentaenoic acid (18-HEPE) via two pathways. EPA can be converted into its hydroxy derivative in the presence of aspirin or by the cytochrome P450 enzymes, which can be transformed by 5-LOX in leukocytes (mainly neutrophils) to ResE1 and ResE2. ResE1 exerts its dual-action nature by interacting with at least two G protein coupled receptors. Interaction with BLT1 on neutrophils prevents transendothelial migration(10,181) while interaction with chemokine-like receptor 1 on monocytes supports non-phlogistic phagocytosis of apoptotic leukocytes(173). In peripheral blood leukocytes, ResE1 prevents the decrease in mitochondrial respiration, membrane potential, and the imbalance of mitochondrial fission and fusion that is induced by TNF $\alpha$ ; since mitochondrial fission alone can induced proinflammatory cytokines, a novel anti-inflammatory mechanism of ResE1 may be through maintenance of mitochondrial integrity (79).

Docosahexaenoic acid (DHA) is a PUFA that is highly enriched in brain, retina and synapses(17,165). Hence, **D-series resolvins** (ResD1-D4) derived from DHA via metabolism by 15-LOX/5-LOX(183) are of a great importance as 'guards' of neural tissues(114). It is important to note that synthesis of D-series resolvins can be also triggered with aspirin. In microglia, both ResD1 and its aspirin analogue indirectly block synthesis of pro-inflammatory interleukin-1β, which is rapidly released following brain injury(83). They also protect macrophages from efferocytosis-induced death via ResD1 mediated activation of cAMP-PKA signaling, which in turn inhibits p47 phox phosphorylation, suppressing NOX activation and limiting further oxidative damage (99). In the liver during ischemia reperfusion, ResD1 regulates mitophagy, mitochondrial biogenesis and mitochondrial fission via thioredoxin 2, and reduces mitochondrial oxidative stress (88). In another elegant study in macrophages, the binding of ResD2 to the GPR18 receptor was shown not only to inhibit the priming but also to expedite the deactivation of the NLRP3 inflammasome during the resolution process, probably by preventing apoptosis-associated speck-like protein oligomerisation at the mitochondria and inflammasome assembly (107).

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**Protectins** represent another class of bioactive lipid mediators produced from DHA via lipoxygenase pathway, with protectin D1 (PD1) being the most potent(12). They are produced by neutrophils, macrophages, T cells, glial cells and retinal pigment epithelium(45). Although they it is known that they can bind leukocytes and retinal pigment epithelium cells, the exact binding partner is yet to be determined(115). PD1 upregulates CCR5 expression in neutrophils(9) and stimulates non-phlogistic uptake of apoptotic leukocytes by macrophages(173). **Maresins** represent the third class of SPM derived from DHA via metabolism by 12-LOX/5-LOX. The exact receptors for maresins remain unknown. However there is an evidence that maresins limit neutrophil transendothelial migration(182), promote uptake of apoptotic leukocytes by macrophages(102), and shorten the resolution phase(197).

During ageing, inflammageing is associated with lower SPM concentration (11). Lower maresin, ResD and ResE concentrations during ageing associate with poor cardiovascular outcomes (75). Lower SPM concentration effects on heart function relate to a combination of altered metabolism and increased dietary intake of n-6 fatty acids that correlate with inflammation and oxidative stress (87). In support, delivery of resolvins in nanoparticles can reverse a decline in cardiac function in animal models by promoting efferocytosis and phenotype switching [145]. In a balloon model of vascular injury, both ResD1 and D2 inhibit vascular smooth muscle activation, monocyte recruitment and ROS production (124) and ResD2 promotes revascularisation post-ischemia via GPR18 receptor dependent mechanisms (233). In the brain, histological post-mortem brain studies from people with cognitive impairment but not healthy people have also shown increased levels of the cyclooxygenase, COX-2, greater oxidative damage and more oxidised lipid deposition (28,62,164). Furthermore, the eicosanoid precursors DHA and AA, are significantly dysregulated in the brains of patients with varying degrees of Alzheimer pathology (2,190). Therapeutic activation the resolution pathways has great potential for the treatment of inflammatory diseases. Could appropriate nutrition be a link between innate and adaptive immune system? Studies with n-3 PUFAs supplementation for inflammatory resolution and neuroinflammation have not shown consistent outcomes (1,85,164). It is known that older adults lack delta-6-desaturase (D6D) activity when compared to younger adults (145) which is required for conversion of fatty acids to n-3 PUFAs. In the absence of D6D, AA may predominate as a proinflammatory substrate for COX-2. Furthermore, before tissue-specific treatments become available, it is important to address whether dietary supplementation with PUFA or SPM intermediates can be beneficial for the local production of SPM, or if their supplementation in excess may have deleterious effects due to the increased risk for auto-oxidation. Instead, novel approaches that address mitochondrial quality may prevent the activation of the inflammasome (31,98), generation of high concentrations of oxidised lipids and enable macrophage differentiation to a resolving M2 phenotype and yielding anti-inflammatory outcomes.

This review is focussed on involvement of oxylipins in inflammation and its resolution with relation to age-related vascular disease. The vasculature itself is exposed to a range of oxygen tension, but the variance is less than in other tissues. It is worthy of note that some studies that have explored oxylipins under physiological conditions of extreme oxygen tension, for example in the lung (higher oxygen tension) or exercising muscle (hypoxia). The free-radical dependent formation of oxidised phospholipids and fatty acids is increased during hyperoxia and during reperfusion post-ischemia, however, their biological effects in lung and muscle are similar to those reported in the vasculature with both pro- and anti-inflammatory effects being described (119,161,232). Nevertheless, there is a

consistent view that the enzyme catalysed eicosanoids play an important role in the regulation of inflammation and may be targeted to improve muscle regeneration (30,65,233).

## 3.4 Analytical challenges of measuring products of lipid oxidation

The foregoing discussion has highlighted that oxidised lipids generated from PUFA, cholesterol and phospholipids through enzymatic or free radical mediated reactions are structurally similar, with similar chemical and physical properties. These properties make it difficult to simultaneously separate, and accurately identify and quantify oxidised lipids in biological samples (103,104). Since the regulation of different lipid oxidation species may shift the balance between inflammatory and anti-inflammatory status, and because these processes are fine-tuned, accurate and precise quantitation are important. It is also noteworthy that most of the oxidised lipids are unstable compounds that can degrade or further oxidise during sample collection, handling and storage. Therefore, special care and attention is needed starting from the point of experimental design. These challenges have been reviewed by others (152,168), which emphasise the need of standardised approaches to sample preparation lipid extraction and storing lipid oxidation products. For example, removal of highly abundant parent lipids (cholesterol, phospholipids) and concentration of oxidised species are required to avoid interference and to optimise detection limits. Techniques such as sample evaporation, liquid-liquid extraction and solid phase extraction have been reported in literature. Each of these techniques has their limitations on sample recovery and extraction efficiencies (104). Advances in analytical techniques, availability of high sensitive instruments and column chemistries are playing a major role in overcoming these challenges (43). However, still the complex nature of lipid species and their interaction with solid phase matrix in different strengths may pose challenges to extract all species with a same efficiency (116).

In recent years, much effort has been invested to accurately quantify lipid oxidation products from minimum sample volumes (4,5,192). The development of ultra-high performance liquid chromatography methods with short runtimes using targeted and sensitive quantification using quadrapole ion trap mass spectrometers has been successfully applied to different types of biological samples (6,163,221) and is likely to revolutionise our understanding of the spatial and temporal distribution of these modified lipids.

## 4. Flavonoids mitigating lipid oxidation, inflammation and age-associated vascular disease

Two classes of phytochemicals that are less-well studied but have positive age-related health outcomes are the oxocarotenoids and flavonoids. Oxocarotenoids are metabolised by mitochondria, promote membrane integrity and vascular risk factors. On the other hand, in the PREDIMED study the increase in circulating flavonoids were associated with preventing negative cardiovascular outcomes in older adults following dietary supplementation with olive oil and with or without nuts, and for type diabetes, catechins, proanthocyanins and hydroxybenzoic acid afforded the greater protection (205). The next two sections focus on the mechanisms and effects of these phytochemicals on oxidised lipids; despite being under-investigated in clinical studies, they show promise to exert anti-inflammatory effects and protect against age-related vascular disease.

Diet is a major determinant of healthy aging and the flavonoid phytonutrients may extend healthy lifespan and prevent age-related vascular mortality (53,227). This hypothesis is based on human epidemiological and clinical intervention studies and lifespan studies in model species. The mechanisms are not fully understood and include modulation of nitric oxide-dependent arterial function (106,171) regulation of NADPH oxidase, (158) immune and inflammatory function modulation via Nrf2 activity modulation of the inflammasome, (49) CNS/neuronal cell function

modulation (13). Pre-clinical studies in mice have shown that flavonoids may attenuate inflammation through inhibition of the inflammasome and promoting mitochondrial biogenesis is also enhanced in many tissues (125,225). Mitochondrial biogenesis increases the efficiency of mitochondrial metabolism and promotes differentiation to anti-inflammatory phenotype.

Flavonoids are the biggest group of polyphenolic secondary plant metabolites found in human diets (216). Major classes of flavonoids are flavan-3-ol monomers, theaflavins, proanthocyanidins, anthocyanidins, flavones, flavonols, and flavanones - they differ in their abundance in foods, absorption, metabolism, distribution, and excretion (156). An important reason for the lack of clarity of molecular targets and mechanisms is the likely importance of considering the absorption, distribution, metabolism, and excretion (ADME) of flavonoids when investigating their mechanisms; this was previously underappreciated (138). Most flavonoids undergo significant metabolism during absorption and therefore, the circulating metabolites significantly differ from the flavonoids in food. For instance, flavanols in cocoa consist of monomers, mainly (-)-epicatechin (20%) and procyanidins (80%) which are oligomers with a degree of polymerisation of up to 10 monomers. However, recent research has shown that (-)-epicatechin is methylated, sulphated and glucuronidated during absorption in the small intestine while procyanidins are not absorbed as such and are broken down by the gut microflora, reaching the circulation much later e.g. as valerolactones (139). While studies in the past have shown that many polyphenols including (-)-epicatechin and procyanidins can mediate a number of effects in vitro including inhibition of NADPH oxidase with the half maximal inhibitory concentration (IC50) values in the micromolar range, these are seldomly reached in vivo limiting the understanding of how flavanols mediate health benefits in humans in vivo (169). More recent work with structurally related (-)-epicatechin metabolites at physiologically relevant concentrations in human endothelial cells using systems biology-based network analysis suggests that epicatechin metabolites trigger complex nutri(epi)genomic changes that subsequently modulate endothelial cell adhesion function and permeability. This study also identified key molecular and cellular targets of epicatechin associated with their vascular protective effects in vivo (121). Flavanol metabolites reduce monocyte adhesion to endothelial cells through modulation of expression of genes via p38-MAPK and p65-NF-kB pathways (33). Whether there is any effect on concentration of (per)oxidised lipids is unknown. Comparison of the 253 differentially expressed genes with the 4787 TNFα-modulated genes revealed an overlap of 66 genes. Interestingly, among these 66 common genes, 44 presented opposing gene expression profiles, suggesting that epicatechin metabolites could partially counteract the genomic effect induced by  $TNF\alpha$  in endothelial cells. Another interesting class of flavonoids are anthocyanins which give blueberries their blue colour. While the parent compounds are not likely to be absorbed in relevant amounts, there are many smaller phenolic degradation products that reach micromolar plasma concentrations and may lead to significant health effects (157). Ex vivo experiments showed that anthocyanins and their gut metabolites can affect adenosine diphosphateinduced platelet activation and their aggregation with monocytes and neutrophils (96) and attenuate monocyte adhesion and transendothelial migration through nutrigenomic mechanisms regulating endothelial cell permeability (95).

Human flavonoid intake varies considerably, and epidemiological data rely on food frequency questionnaires. Based on food consumption data from the European Food Safety Authority and the FLAVIOLA Food Composition Database, it was estimated that in Europe the mean daily intake is 430 mg/d and the median id 160 mg/d with tea and fruit being the main source (216). Data from

anthropological research (82) and observational studies (81) suggested that flavonoids can reduce the risk of ageing-related pathologies such as cardiovascular disease although results from more recent studies were less clear (118,122,215) and the main beneficial effect emerges when comparing very low intake with low intake. Overall, a systematic review suggests that the dietary intakes of six classes of flavonoids, namely flavonols, anthocyanidins, proanthocyanidins, flavones, flavanones and flavan-3-ols, significantly decrease the risk of CVD (222). Unfortunately, randomised controlled or prospective trials studying human morbidity or mortality as an endpoint for flavonoid supplementation are scarce with the exception of the ongoing COSMOS trial (35).

Putative mechanisms that may underlie flavonoid benefit as identified in worms include energyrestriction-like effects, inhibition of insulin-like-growth factor signalling, induction of antioxidant defence mechanisms, lowered inflammation, hormesis, and antimicrobial properties. Lifespan studies show that flavonoids can promote longevity in a range of species (142). In most studies, worms and flies experienced lifespan extension when supplemented with flavonoids either as extracts or single compounds. Studies with mutant worms and flies give hints as to which gene products may be regulated by flavonoids and consequently enhance longevity. Although flavonoids can work as radical scavengers because of their chemical structure; their in vivo antioxidative action is more likely caused by them stabilising and up-regulating the antioxidant transcription factor Nrf2. Interestingly, in flies, cocoa powder was tested in SOD-deficient mutants and it prolonged life-span in flies that did not have the cytosolic SOD1 but was detrimental for flies without the mitochondrial SOD2. These findings suggest that when oxidative stress levels are very high, flavonoids could further increase oxidative damage (14). The metabolome of flavonoids differs widely between mice and men making comparative conclusions difficult (137), and studies in mice are not discussed further here. Readers are directed to a few studies in mice (140,157,172).

Taken together, while flavonoids are promising phytonutrients that may extend a healthy lifespan and limit vascular co-morbidities, this remains to be demonstrated clinically. The underlying mechanisms of individual flavonoids and their circulating putatively bioactive metabolites and their interactions with inflammatory pathways remain to be elucidated taking their ADME into account.

#### 5. Carotenoids, inflammation and age-related dementia and vascular disease

PUFA are highly susceptible to peroxidation. Carotenoids can quench singlet oxygen and scavenge lipid peroxides [9–11]. They are defined into two groups according to their polarity: xanthophylls (polar carotenoids such as astaxanthin,  $\beta$ -cryptoxanthin, lutein, and zeaxanthin) and carotenes (nonpolar carotenoids such as  $\alpha$ -carotene,  $\beta$ -carotene, and lycopene). In contrast to carotenes which are metabolised by BCO1 and retained in the cytoplasm, the enzyme BCO2 is associated with the inner mitochondrial membrane and metabolises xanthophylls (which accumulates in the inner mitochondrial membrane) into long-chain apo-carotenoids. BCO2 functions as a key regulatory enzyme that prevents toxicity caused by carotenoid accumulation and loss of BCO2 function is associated with the development of mitochondrial oxidative stress and metabolic diseases [16]. The uptake, distribution and metabolism of carotenoids have been attributed to the activities of different protein transporters (SR-BI, CD36, NPC1L1), digestion enzymes (PNLIP, CES), cleavage enzymes (BCO1/2), intracellular transporters (FABP2) and receptors (LPL, APOC/E, LDLR). It is predicted that carotenoids are widely bioavailable [37]. At a molecular level zeaxanthin not only attenuates lipid oxidation by scavenging mitochondrial ROS but also promotes mitochondrial biogenesis (105). The

oxocarotenoids are effective modulators of the physical properties of both natural and model membranes, increasing their rigidity and thermostability (93). Oxo-carotenoids stabilise membranes to a greater extent than  $\beta$ -carotene; they are incorporated into bilayers and span the membrane with their lipophilic core, being further anchored in the aqueous area with –OH groups, thus functioning like a molecular rivet (74,93).

Several intervention studies have investigated whether carotenoids improve vascular outcomes. Despite the lack of benefit of astaxanthin on vascular risk factors in renal transplant patients (34), a systematic review and meta-analysis has confirmed that increasing tomato and lycopene intake has positive effects on blood lipids, blood pressure and endothelial function (29). Smaller studies have also identified positive effects of lutein in early atherosclerosis (228,235). Previously, we have shown that a decrease of circulating carotenoids and tocopherols was associated in correlated with increased protein and lipid oxidation (148) in dementia and vascular disease patients that was correlated with the degree of cognitive impairment (39). In the MARK-Age study, lower levels of  $\beta$ -cryptoxanthin and zeaxanthin were found among 2220 randomly recruited age-stratified persons, in those who were physically, cognitively or psychologically frail [34]. Notably, carotenoid concentrations were inversely associated with the risk of being cognitively frail after adjusting for confounders (131).

Vascular disease is a common co-morbidity with dementia(15). Evidence from cross-sectional and observational studies has grown for an association between elevated serum cholesterol in mid-life and later development of dementia(187). Furthermore, chronic low-grade inflammation relates to age-related cognitive impairment (150). The epigenetic oxidative redox shift theory of ageing proposes that sedentary behavior associated with age triggers an oxidised redox shift and impaired mitochondrial function inducing an epigenetic vicious circle that promotes inflammation (64).

Conversely, optimal nutrition may negatively influence dementia risk through epigenetic mechanisms. The "LEARn" (latent early life-associated regulation) model suggests that environmental stress – likely and often oxidative stress and/or nutritional imbalance - marks a gene which later during life and in the presence of a secondary trigger, e.g. inflammation, it will be expressed aberrantly causing overt pathology (113). Preliminary studies on the role of vascular- and lifestyle-related preventive strategies show that vascular risk control and lifestyle improvement are indeed able to slow down the progression of cognitive impairment (63). Plasma levels of several lipophilic antioxidant micronutrients are significantly associated, independently of fruit and vegetable intake, with validated, accurate measures of both cognitive and physical performance in persons with mild cognitive impairment (MCI). This represents a further step in the field of nutritional cognitive neuroscience.

Taken together, while carotenoids are promising phytonutrients that may limit vascular comorbidities, this remains to be demonstrated clinically. The underlying mechanisms of individual oxocarotenoids and their interactions with mitochondria and inflammatory pathways remain to determined. In light of the increasing attention towards the nutritional cognitive neuroscience of carotenoids [43, 44], the use of computerised measures of cognitive performance might be further implemented in future studies investigating their effects on cognitive and physical impairment.

## **Overall conclusion**

<text> Inflammation associates with ROS/RNS production and with changes to the (oxy)lipidome. (Per)oxidised lipids are central mediators in the initiation and resolution of inflammation. Generalised targeting with antioxidants such as tocopherols as a primary prevention in age-related, inflammatory vascular disease has not proven successful. A more nuanced view is emerging where ROS/NO production is desirable during early phases of inflammation and for resolution. Indeed, the celltargeted amplification of ROS in neutrophils may be required in some circumstances (Reshetnikov et al. 2018) to promote resolution. Phytochemicals that promote resolving oxylipid mediators and improve mitochondrial quality control merit further investigation as inhibitors of underlying sterile inflammation and to mitigate age-related vascular disease.

# Table 1

Model	Target	Outcome	OxPL	Reference	Year
HEK, vascular	TRPC5 calcium	Increase in calcium	PGPC	(7)	2010
smooth	channel	permeability	PCPC		
muscle cells					
Pulmonary	Cytoskeletal	Reduced permeability	PEIPC	(19,193)	2012,
endothelial	remodelling, Src	with low			2013
cells	Kinase mediated	concentration and			
	VE-cadherin	full length oxPL.	PGPC		
	phosphorylation	High concentration	POVPC		
		and fragmented oxPL			
		increase permeability			
Myeloid cells	Chemokine and	Decreased	OxPAPC with	(24)	2015
	cytokine	inflammatory	ероху-		
	production via	response,	cyclopentenone		
	Nrf2	Reduced sepsis			
Macrophages	Non-canonical	Decrease in IL-1b and	OxPAPC	(32)	2018
not DCs	inflammasome	decrease n sepsis			
	pathway, TLR4				
Endothelial	Cell adherens	Fragment disrupts	LysoPC	(80)	2013
cells	junction	barrier			
		Complex mix	OxPAPC		
		enhances barrier			
Macrophages,	TLR2, TLR4, MD2	Decrease in LPS-	Ox PAPC,	(51)	2008
smooth		mediated	POVPC, PGPC		
muscle		inflammatory			
		signalling			
Bone-marrow	Mitochondrial	Increased IL-1b	POVPC	(229)	2017
derived	ROS,	production due to 🗸			
macrophages	inflammasome	mito-ROS and	D		
		calcium influx			
		activating NLRP3			
		_			
EK = hamster e	mbryonic kidney cell	S			

#### **Figure Legends**

Figure 1. Key mediators and cells involved in physiological and sterile inflammation responses

Figure 2. Innate immune recruitment to inflammatory sites; oxidised phospholipids contribute to the chemotactic gradient for recruitment of inflammatory cells. Their local production in the mitochondria results in impaired mitochondrial activity and failure to remove damaged mitochondria exacerbates ROS production. MtDNA and OxPL enhance inflammasome activation and pro-inflammatory cytokine release.

Figure 3. Monocyte and macrophage phenotype and metabolism

Figure 4: Cellular distribution of oxysterols. Cellular cholesterol is maintained by regulatory mechanisms that control synthesis, uptake, metabolism and efflux. Cells uptake cholesterol and oxysterols packed lipoproteins aided by many membrane receptors, such as LDL receptor (LDLR) family, EBI2 , G-protein coupled receptor, GPR17, C-X-C motif chemokine receptor 2 (CXCR2) and scavenger receptors. Inside the cell, endosomes containing cholesterol/oxysterol are transported to Endoplasmic reticulum (ER) mitochondria or peroxisomes. Within ER, sterols are further metabolised with help of many protein complexes including, oxysterol binding protein (OSBP), OSBP-related proteins (ORPs), the cellular nucleic acid binding protein, the sterol regulatory element binding protein (SREBP), insulin induced gene protein (INSIG) and Niemann-Pick protein C1 (NPC1). These metabolic activities within ER and golgi transfer signals to nucleus either to up- or down regulate endogenous cholesterol synthesis via 3-hydroxy-3-methylglutaryl CoA reductase (HMGR) pathway. Cellular cholesterol and oxysterol levels are also sensed a family of nuclear receptors family; Liver X receptors (LXR), Retinoid X receptor (RXR), Retinoic acid receptor (RAR)related orphan proteins (ROR $\alpha$ , ROR $\beta$  and ROR $\gamma$ ) and Estrogen receptor  $\alpha$  (ER $\alpha$ ). Activation of these transcription factors upregulate multiple genes involved in cellular cholesterol homeostasis, including ATP-binding cassette transporter A1 (ABCA1), ATP binding cassette transporter G1 (ABCG1), and APOE. Mitochondrial translocator protein (TSPO) has high affinity for cholesterol and uptake cholesterol from ER or from lipid droplets. Steroidogenic acute regulatory protein (StAR) transport cholesterol from the outer to inner membrane in mitochondria. Peroxisome proteins ATPbinding cassette transporter D1 (ABCD1) and acyl-CoA oxidase 1 (ACOX1) are also involve in transport of cholesterol and oxysterols.

Figure 5. Lipid-derived mediators of inflammation are key players that drive both pro-inflammatory and anti-inflammatory and pro-resolving responses. Infiltration of neutrophils from blood vessels into the site of injury, and later pro-inflammatory macrophages (M1 macrophages), is supported by leukotriene  $B_4$  and prostaglandin  $F_{2\alpha}$ . Resolution phase is initiated by the synthesis of prostaglandins that induce "class switch" from pro-inflammatory (LtB<sub>4</sub>, PGF<sub>2α</sub>) to the specialised pro-resolving mediators (SPM, such as lipoxins, resolvins, protectins and maresins). Their dual acting nature stops neutrophils from further infiltrating to the sites of injury, while supporting chemotaxis of pro-repair macrophages (M2 macrophages), phagocytosis of apoptotic material, and their exit via lymphatics.

Figure 6. Pathways of formation and molecular structures of eicosanoids

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## References

- 1. Abdelhamid AS, Brown TJ, Brainard JS, Biswas P, Thorpe GC, Moore HJ, Deane KHO, AlAbdulghafoor FK, Summerbell CD, Worthington HV, et al. Omega-3 fatty acids for the primary and secondary prevention of cardiovascular disease. *Cochrane Database of Systematic Reviews*, 2018.
- Abdullah L, Evans JE, Emmerich T, Crynen G, Shackleton B, Keegan AP, Luis C, Tai L, LaDu MJ, Mullan M, Crawford F, Bachmeier C. APOE ε4 specific imbalance of arachidonic acid and docosahexaenoic acid in serum phospholipids identifies individuals with preclinical Mild Cognitive Impairment/Alzheimer's Disease. *Aging* 9: 964-985, 2017.
- 3. Ademowo OS, Dias HKI, Burton DGA, Griffiths HR. Lipid (per) oxidation in mitochondria: an emerging target in the ageing process? *Biogerontology* 18: 859-879, 2017.
- 4. Ademowo OS, Dias HKI, Milic I, Devitt A, Moran R, Mulcahy R, Howard AN, Nolan JM, Griffiths HR. Phospholipid oxidation and carotenoid supplementation in Alzheimer's disease patients. *Free Radic Biol Med* 108: 77-85, 2017.
- 5. Ademowo OS, Dias HKI, Milic I, Devitt A, Moran R, Mulcahy R, Howard AN, Nolan JM, Griffiths HR. Phospholipid oxidation and carotenoid supplementation in Alzheimer's disease patients. *Free Radical Biology and Medicine* 108: 77-85, 2017.
- 6. Ademowo OS, Sharma P, Cockwell P, Reis A, Chapple IL, Griffiths HR, Dias IHK. Distribution of plasma oxidised phosphatidylcholines in chronic kidney disease and periodontitis as a comorbidity. *Free Radical Biology and Medicine*, 2019.
- Al-Shawaf E, Naylor J, Taylor H, Riches K, Milligan CJ, O'Regan D, Porter KE, Li J, Beech DJ. Short-term stimulation of calcium-permeable transient receptor potential canonical 5containing channels by oxidized phospholipids. *Arterioscler Thromb Vasc Biol* 30: 1453-9, 2010.
- 8. Alexander RL, Wright MW, Gorczynski MJ, Smitherman PK, Akiyama TE, Wood HB, Berger JP, King SB, Morrow CS. Differential potencies of naturally occurring regioisomers of nitrolinoleic acid in PPARgamma activation. *Biochemistry* 48: 492-8, 2009.
- 9. Ariel A, Fredman G, Sun YP, Kantarci A, Van Dyke TE, Luster AD, Serhan CN. Apoptotic neutrophils and T cells sequester chemokines during immune response resolution through modulation of CCR5 expression. *Nat Immunol* 7: 1209-16, 2006.
  - 10. Arita M, Ohira T, Sun YP, Elangovan S, Chiang N, Serhan CN. Resolvin E1 selectively interacts with leukotriene B4 receptor BLT1 and ChemR23 to regulate inflammation. *J Immunol* 178: 3912-7, 2007.
  - 11. Arnardottir HH, Dalli J, Colas RA, Shinohara M, Serhan CN. Aging delays resolution of acute inflammation in mice: reprogramming the host response with novel nano-proresolving medicines. *Journal of immunology (Baltimore, Md. : 1950)* 193: 4235-4244, 2014.
- 12. Aursnes M, Tungen JE, Vik A, Colas R, Cheng CY, Dalli J, Serhan CN, Hansen TV. Total synthesis of the lipid mediator PD1n-3 DPA: configurational assignments and anti-inflammatory and pro-resolving actions. *J Nat Prod* 77: 910-6, 2014.
- 13. Azam S, Jakaria M, Kim IS, Kim J, Haque ME, Choi DK. Regulation of Toll-Like Receptor (TLR) Signaling Pathway by Polyphenols in the Treatment of Age-Linked Neurodegenerative Diseases: Focus on TLR4 Signaling. *Front Immunol* 10: 1000, 2019.
- 14. Bahadorani S, Hilliker AJ. Cocoa confers life span extension in Drosophila melanogaster. *Nutr Res* 28: 377-82, 2008.
- 15. Ballard C, Gauthier S, Corbett A, Brayne C, Aarsland D, Jones E. Alzheimer's disease. *The Lancet* 377: 1019-1031.
- 16. Bauer J, Ripperger A, Frantz S, Ergun S, Schwedhelm E, Benndorf RA. Pathophysiology of isoprostanes in the cardiovascular system: implications of isoprostane-mediated thromboxane A2 receptor activation. *Br J Pharmacol* 171: 3115-31, 2014.

- Bazan NG, Birkle DL, Reddy TS. Docosahexaenoic acid (22:6, n-3) is metabolized to lipoxygenase reaction products in the retina. *Biochem Biophys Res Commun* 125: 741-7, 1984.
  - Biermann MH, Podolska MJ, Knopf J, Reinwald C, Weidner D, Maueroder C, Hahn J,
    Kienhofer D, Barras A, Boukherroub R, Szunerits S, Bilyy R, Hoffmann M, Zhao Y, Schett G,
    Herrmann M, Munoz LE. Oxidative Burst-Dependent NETosis Is Implicated in the Resolution of Necrosis-Associated Sterile Inflammation. *Front Immunol* 7: 557, 2016.
  - 19. Birukova AA, Starosta V, Tian X, Higginbotham K, Koroniak L, Berliner JA, Birukov KG. Fragmented oxidation products define barrier disruptive endothelial cell response to OxPAPC. *Transl Res* 161: 495-504, 2013.
  - 20. Björkhem I. Do oxysterols control cholesterol homeostasis? *Journal of Clinical Investigation* 110: 725-730, 2002.
  - 21. Bochkov V, Gesslbauer B, Mauerhofer C, Philippova M, Erne P, Oskolkova OV. Pleiotropic effects of oxidized phospholipids. *Free Radical Biology and Medicine* 111: 6-24, 2017.
  - 22. Bochkov V, Gesslbauer B, Mauerhofer C, Philippova M, Erne P, Oskolkova OV. Pleiotropic effects of oxidized phospholipids. *Free Radic Biol Med* 111: 6-24, 2017.
  - 23. Bochkov VN, Oskolkova OV, Birukov KG, Levonen AL, Binder CJ, Stockl J. Generation and biological activities of oxidized phospholipids. *Antioxid Redox Signal* 12: 1009-59, 2010.
  - 24. Bretscher P, Egger J, Shamshiev A, Trotzmuller M, Kofeler H, Carreira EM, Kopf M, Freigang S. Phospholipid oxidation generates potent anti-inflammatory lipid mediators that mimic structurally related pro-resolving eicosanoids by activating Nrf2. *EMBO Mol Med* 7: 593-607, 2015.
  - 25. Brown AJ, Jessup W. Oxysterols and atherosclerosis. *Atherosclerosis* 142: 1-28, 1999.
  - 26. Brown AJ, Mander EL, Gelissen IC, Kritharides L, Dean RT, Jessup W. Cholesterol and oxysterol metabolism and subcellular distribution in macrophage foam cells. Accumulation of oxidized esters in lysosomes. *J Lipid Res* 41: 226-37, 2000.
  - 27. Brown MS, Goldstein JL. The SREBP Pathway: Regulation of Cholesterol Metabolism by Proteolysis of a Membrane-Bound Transcription Factor. *Cell* 89: 331-340, 1997.
  - 28. Butterfield DA, Reed T, Perluigi M, De Marco C, Coccia R, Cini C, Sultana R. Elevated proteinbound levels of the lipid peroxidation product, 4-hydroxy-2-nonenal, in brain from persons with mild cognitive impairment. *Neuroscience Letters* 397: 170-173, 2006.
  - 29. Cheng HM, Koutsidis G, Lodge JK, Ashor A, Siervo M, Lara J. Tomato and lycopene supplementation and cardiovascular risk factors: A systematic review and meta-analysis. *Atherosclerosis* 257: 100-108, 2017.
  - 30. Chiu CY, Smyl C, Dogan I, Rothe M, Weylandt KH. Quantitative Profiling of Hydroxy Lipid Metabolites in Mouse Organs Reveals Distinct Lipidomic Profiles and Modifications Due to Elevated n-3 Fatty Acid Levels. *Biology (Basel)* 6, 2017.
  - 31. Chiu HW, Li LH, Hsieh CY, Rao YK, Chen FH, Chen A, Ka SM, Hua KF. Glucosamine inhibits IL-1beta expression by preserving mitochondrial integrity and disrupting assembly of the NLRP3 inflammasome. *Sci Rep* 9: 5603, 2019.
  - 32. Chu LH, Indramohan M, Ratsimandresy RA, Gangopadhyay A, Morris EP, Monack DM, Dorfleutner A, Stehlik C. The oxidized phospholipid oxPAPC protects from septic shock by targeting the non-canonical inflammasome in macrophages. *Nat Commun* 9: 996, 2018.
  - Claude S, Boby C, Rodriguez-Mateos A, Spencer JP, Gerard N, Morand C, Milenkovic D.
    Flavanol metabolites reduce monocyte adhesion to endothelial cells through modulation of expression of genes via p38-MAPK and p65-Nf-kB pathways. *Mol Nutr Food Res* 58: 1016-27, 2014.
  - 34. Coombes JS, Sharman JE, Fassett RG. Astaxanthin has no effect on arterial stiffness, oxidative stress, or inflammation in renal transplant recipients: a randomized controlled trial (the XANTHIN trial). *Am J Clin Nutr* 103: 283-9, 2016.

2		
3	35.	COSMOS. 2015. COcoa Supplement and Multivitamin Outcomes Study, NCT02422745
4		https://clinicaltrials.gov/ct2/show/NCT02422745.
5	36.	Cui T, Schopfer FJ, Zhang J, Chen K, Ichikawa T, Baker PR, Batthyany C, Chacko BK, Feng X,
6		Patel RP, Agarwal A, Freeman BA, Chen YE. Nitrated fatty acids: Endogenous anti-
7 8		inflammatory signaling mediators. J Biol Chem 281: 35686-98, 2006.
8 9	37.	Dang EV, McDonald JG, Russell DW, Cyster JG. Oxysterol Restraint of Cholesterol Synthesis
10	57.	Prevents AIM2 Inflammasome Activation. <i>Cell</i> 171: 1057-1071.e11, 2017.
11	38.	Dias HK, Brown CL, Polidori MC, Lip GY, Griffiths HR. LDL-lipids from patients with
12	50.	hypercholesterolaemia and Alzheimer's disease are inflammatory to microvascular
13		
14	20	endothelial cells: mitigation by statin intervention. <i>Clin Sci (Lond)</i> 129: 1195-206, 2015.
15	39.	Dias IH, Polidori MC, Li L, Weber D, Stahl W, Nelles G, Grune T, Griffiths HR. Plasma levels of
16		HDL and carotenoids are lower in dementia patients with vascular comorbidities. J
17		Alzheimers Dis 40: 399-408, 2014.
18	40.	Dias IHK, Milic I, Lip GYH, Devitt A, Polidori MC, Griffiths HR. Simvastatin reduces circulating
19		oxysterol levels in men with hypercholesterolaemia. <i>Redox Biol</i> 16: 139-145, 2018.
20	41.	Dias IHK, Mistry J, Fell S, Reis A, Spickett CM, Polidori MC, Lip GYH, Griffiths HR. Oxidized LDL
21		lipids increase $\beta$ -amyloid production by SH-SY5Y cells through glutathione depletion and
22		lipid raft formation. Free Radical Biology and Medicine 75: 48-59, 2014.
23	42.	Dias Irundika HK, Polidori Maria C, Griffiths Helen R. Hypercholesterolaemia-induced
24		oxidative stress at the blood-brain barrier. Biochemical Society Transactions 42: 1001-1005,
25 26		2014.
20	43.	Dias IHK, Wilson SR, Roberg-Larsen H. Chromatography of oxysterols. <i>Biochimie</i> 153: 3-12,
28		2018.
29	44.	Donato AJ, Morgan RG, Walker AE, Lesniewski LA. Cellular and molecular biology of aging
30		endothelial cells. J Mol Cell Cardiol 89: 122-35, 2015.
31	45.	Doyle R, Sadlier DM, Godson C. Pro-resolving lipid mediators: Agents of anti-ageing? Semin
32	45.	Immunol 40: 36-48, 2018.
33	16	
34	46.	Dunston CR, Griffiths HR. The effect of ageing on macrophage Toll-like receptor-mediated
35	47	responses in the fight against pathogens. <i>Clin Exp Immunol</i> 161: 407-16, 2010.
36	47.	Edenius C, Haeggstrom J, Lindgren JA. Transcellular conversion of endogenous arachidonic
37		acid to lipoxins in mixed human platelet-granulocyte suspensions. Biochem Biophys Res
38		Commun 157: 801-7, 1988.
39	48.	Egea J, Fabregat I, Frapart YM, Ghezzi P, Gorlach A, Kietzmann T, Kubaichuk K, Knaus UG,
40		Lopez MG, Olaso-Gonzalez G, Petry A, Schulz R, Vina J, Winyard P, Abbas K, Ademowo OS,
41 42		Afonso CB, Andreadou I, Antelmann H, Antunes F, Aslan M, Bachschmid MM, Barbosa RM,
42 43		Belousov V, Berndt C, Bernlohr D, Bertran E, Bindoli A, Bottari SP, Brito PM, Carrara G, Casas
43 44		AI, Chatzi A, Chondrogianni N, Conrad M, Cooke MS, Costa JG, Cuadrado A, My-Chan Dang P,
45		De Smet B, Debelec-Butuner B, Dias IHK, Dunn JD, Edson AJ, El Assar M, El-Benna J,
46		Ferdinandy P, Fernandes AS, Fladmark KE, Forstermann U, Giniatullin R, Giricz Z, Gorbe A,
47		Griffiths H, Hampl V, Hanf A, Herget J, Hernansanz-Agustin P, Hillion M, Huang J, Ilikay S,
48		Jansen-Durr P, Jaquet V, Joles JA, Kalyanaraman B, Kaminskyy D, Karbaschi M, Kleanthous M,
49		Klotz LO, Korac B, Korkmaz KS, Koziel R, Kracun D, Krause KH, Kren V, Krieg T, Laranjinha J,
50		Lazou A, Li H, Martinez-Ruiz A, Matsui R, McBean GJ, Meredith SP, Messens J, Miguel V,
51		Mikhed Y, Milisav I, Milkovic L, Miranda-Vizuete A, Mojovic M, Monsalve M, Mouthuy PA,
52		Mikhed F, Milisav F, Miković E, Milanda-Vizdete A, Mojović M, Molsave M, Modulidy PA, Mulvey J, Munzel T, Muzykantov V, Nguyen ITN, Oelze M, Oliveira NG, Palmeira CM,
53		
54		Papaevgeniou N, Pavicevic A, Pedre B, Peyrot F, Phylactides M, Pircalabioru GG, Pitt AR,
55		Poulsen HE, Prieto I, Rigobello MP, Robledinos-Anton N, Rodriguez-Manas L, Rolo AP,
56		Rousset F, Ruskovska T, Saraiva N, Sasson S, Schroder K, Semen K, Seredenina T,
57		Shakirzyanova A, Smith GL, Soldati T, Sousa BC, Spickett CM, Stancic A, Stasia MJ,
58		Steinbrenner H, Stepanic V, Steven S, Tokatlidis K, Tuncay E, Turan B, Ursini F, Vacek J,
59		Vajnerova O, Valentova K, Van Breusegem F, Varisli L, Veal EA, Yalcin AS, Yelisyeyeva O,
60		

Zarkovic N, Zatloukalova M, Zielonka J, Touyz RM, Papapetropoulos A, Grune T, Lamas S, Schmidt H, Di Lisa F, Daiber A. European contribution to the study of ROS: A summary of the findings and prospects for the future from the COST action BM1203 (EU-ROS). *Redox Biol* 13: 94-162, 2017.

- 49. Ellinger S, Stehle P. Impact of Cocoa Consumption on Inflammation Processes-A Critical Review of Randomized Controlled Trials. *Nutrients* 8, 2016.
- 50. Elliott EI, Miller AN, Banoth B, Iyer SS, Stotland A, Weiss JP, Gottlieb RA, Sutterwala FS, Cassel SL. Cutting Edge: Mitochondrial Assembly of the NLRP3 Inflammasome Complex Is Initiated at Priming. *J Immunol* 200: 3047-3052, 2018.
- 51. Erridge C, Kennedy S, Spickett CM, Webb DJ. Oxidized phospholipid inhibition of toll-like receptor (TLR) signaling is restricted to TLR2 and TLR4: roles for CD14, LPS-binding protein, and MD2 as targets for specificity of inhibition. *J Biol Chem* 283: 24748-59, 2008.
- 52. Erridge C, Webb DJ, Spickett CM. Toll-like receptor 4 signalling is neither sufficient nor required for oxidised phospholipid mediated induction of interleukin-8 expression. *Atherosclerosis* 193: 77-85, 2007.
- 53. Estruch R, Ros E, Salas-Salvado J, Covas MI, Corella D, Aros F, Gomez-Gracia E, Ruiz-Gutierrez V, Fiol M, Lapetra J, Lamuela-Raventos RM, Serra-Majem L, Pinto X, Basora J, Munoz MA, Sorli JV, Martinez JA, Martinez-Gonzalez MA. Primary prevention of cardiovascular disease with a Mediterranean diet. *N Engl J Med* 368: 1279-90, 2013.
- 54. Fazzari M, Vitturi DA, Woodcock SR, Salvatore SR, Freeman BA, Schopfer FJ. Electrophilic fatty acid nitroalkenes are systemically transported and distributed upon esterification to complex lipids. *J Lipid Res* 60: 388-399, 2019.
- 55. Filomenko R, Fourgeux C, Bretillon L, Gambert-Nicot S. Oxysterols: Influence on plasma membrane rafts microdomains and development of ocular diseases. *Steroids* 99: 259-65, 2015.
- 56. Fiore S, Serhan CN. Formation of lipoxins and leukotrienes during receptor-mediated interactions of human platelets and recombinant human granulocyte/macrophage colony-stimulating factor-primed neutrophils. *J Exp Med* 172: 1451-7, 1990.
- 57. Fiore S, Serhan CN. Lipoxin A4 receptor activation is distinct from that of the formyl peptide receptor in myeloid cells: inhibition of CD11/18 expression by lipoxin A4-lipoxin A4 receptor interaction. *Biochemistry* 34: 16678-86, 1995.
- 58. Flower RJ. Prostaglandins, bioassay and inflammation. *Br J Pharmacol* 147 Suppl 1: S182-92, 2006.
- 59. Friedli O, Freigang S. Cyclopentenone-containing oxidized phospholipids and their isoprostanes as pro-resolving mediators of inflammation. *Biochimica et Biophysica Acta* (*BBA*) *Molecular and Cell Biology of Lipids* 1862: 382-392, 2017.
- 60. Fu P, Birukov KG. Oxidized phospholipids in control of inflammation and endothelial barrier. *Transl Res* 153: 166-76, 2009.
- 61. Fuchs TA, Abed U, Goosmann C, Hurwitz R, Schulze I, Wahn V, Weinrauch Y, Brinkmann V, Zychlinsky A. Novel cell death program leads to neutrophil extracellular traps. *J Cell Biol* 176: 231-41, 2007.
- 62. Fujimi K, Noda K, Sasaki K, Wakisaka Y, Tanizaki Y, Iida M, Kiyohara Y, Kanba S, Iwaki T. Altered Expression of COX-2 in Subdivisions of the Hippocampus during Aging and in Alzheimer's Disease: The Hisayama Study. *Dementia and Geriatric Cognitive Disorders* 23: 423-431, 2007.
- 63. Gerger P, Pai RK, Stuckenschneider T, Falkenreck J, Weigert H, Stahl W, Weber B, Nelles G, Spazzafumo L, Schneider S, Polidori MC. Associations of Lipophilic Micronutrients with Physical and Cognitive Fitness in Persons with Mild Cognitive Impairment. *Nutrients* 11: 902, 2019.

2		
3	C A	Check D. LeV(suit KD. Demest AL. Drewer CL. A. Deversible Farth Ovidiand Deday State That
4	64.	Ghosh D, LeVault KR, Barnett AJ, Brewer GJ. A Reversible Early Oxidized Redox State That
5		Precedes Macromolecular ROS Damage in Aging Nontransgenic and 3xTg-AD Mouse
6		Neurons. The Journal of Neuroscience 32: 5821-5832, 2012.
7	65.	Giannakis N, Sansbury BE, Patsalos A, Hays TT, Riley CO, Han X, Spite M, Nagy L. Dynamic
8		changes to lipid mediators support transitions among macrophage subtypes during muscle
9		regeneration. Nat Immunol 20: 626-636, 2019.
10	66.	Godson C, Mitchell S, Harvey K, Petasis NA, Hogg N, Brady HR. Cutting edge: lipoxins rapidly
11		stimulate nonphlogistic phagocytosis of apoptotic neutrophils by monocyte-derived
12		macrophages. J Immunol 164: 1663-7, 2000.
13	67.	Gold ES, Diercks AH, Podolsky I, Podyminogin RL, Askovich PS, Treuting PM, Aderem A. 25-
14		Hydroxycholesterol acts as an amplifier of inflammatory signaling. Proceedings of the
15		National Academy of Sciences 111: 10666-10671, 2014.
16	68.	Goldstein JL, Brown MS. Regulation of the mevalonate pathway. <i>Nature</i> 343: 425-430, 1990.
17		
18	69.	Green DR, Galluzzi L, Kroemer G. Mitochondria and the Autophagy–Inflammation–Cell Death
19 20		Axis in Organismal Aging. Science 333: 1109, 2011.
20	70.	Greig FH, Kennedy S, Spickett CM. Physiological effects of oxidized phospholipids and their
21 22		cellular signaling mechanisms in inflammation. Free Radic Biol Med 52: 266-80, 2012.
22 23	71.	Griffiths HR, Aldred S, Dale C, Nakano E, Kitas GD, Grant MG, Nugent D, Taiwo FA, Li L,
23		Powers HJ. Homocysteine from endothelial cells promotes LDL nitration and scavenger
25		receptor uptake. Free Radic Biol Med 40: 488-500, 2006.
26	72.	Griffiths HR, Gao D, Pararasa C. Redox regulation in metabolic programming and
27		inflammation. <i>Redox Biology</i> 12: 50-57, 2017.
28	73.	Griffiths WJ, Wang Y. Oxysterol research: a brief review. <i>Biochemical Society Transactions</i> :
29		BST20180135, 2019.
30	74.	Grudzinski W, Nierzwicki L, Welc R, Reszczynska E, Luchowski R, Czub J, Gruszecki WI.
31	,	Localization and Orientation of Xanthophylls in a Lipid Bilayer. <i>Scientific reports</i> 7: 9619-
32		9619, 2017.
33	75	
34	75.	Halade GV, Kain V, Black LM, Prabhu SD, Ingle KA. Aging dysregulates D- and E-series
35		resolvins to modulate cardiosplenic and cardiorenal network following myocardial infarction.
36		Aging 8: 2611-2634, 2016.
37	76.	Han S, Zhuang H, Shumyak S, Wu J, Xie C, Li H, Yang L-J, Reeves WH. Liver X Receptor Agonist
38		Therapy Prevents Diffuse Alveolar Hemorrhage in Murine Lupus by Repolarizing
39		Macrophages. Frontiers in Immunology 9, 2018.
40	77.	Hardy KD, Cox BE, Milne GL, Yin H, Roberts LJ, 2nd. Nonenzymatic free radical-catalyzed
41		generation of 15-deoxy-Delta(12,14)-prostaglandin J(2)-like compounds (deoxy-J(2)-
42		isoprostanes) in vivo. J Lipid Res 52: 113-24, 2011.
43 44	78.	He Y, Hara H, Nunez G. Mechanism and Regulation of NLRP3 Inflammasome Activation.
44 45		Trends Biochem Sci 41: 1012-1021, 2016.
45	79.	Hecker M, Sommer N, Foch S, Hecker A, Hackstein H, Witzenrath M, Weissmann N, Seeger
47		W, Mayer K. Resolvin E1 and its precursor 18R-HEPE restore mitochondrial function in
48		inflammation. <i>Biochim Biophys Acta Mol Cell Biol Lipids</i> 1863: 1016-1028, 2018.
49	80.	Heffern CT, Pocivavsek L, Birukova AA, Moldobaeva N, Bochkov VN, Lee KY, Birukov KG.
50	80.	Thermodynamic and kinetic investigations of the release of oxidized phospholipids from lipid
51		
52	04	membranes and its effect on vascular integrity. <i>Chem Phys Lipids</i> 175-176: 9-19, 2013.
53	81.	Hertog MG, Feskens EJ, Hollman PC, Katan MB, Kromhout D. Dietary antioxidant flavonoids
54		and risk of coronary heart disease: the Zutphen Elderly Study. <i>Lancet</i> 342: 1007-1011, 1993.
55	82.	Hollenberg NK, Martinez G, McCullough M, Meinking T, Passan D, Preston M, Rivera A,
56		Taplin D, Vicaria-Clement M. Aging, acculturation, salt intake, and hypertension in the Kuna
57		of Panama. Hypertension 29: 171-176, 1997.
58		
59		
60		

83.	Hong S, Gronert K, Devchand PR, Moussignac RL, Serhan CN. Novel docosatrienes and 17S- resolvins generated from docosahexaenoic acid in murine brain, human blood, and glial cells. Autacoids in anti-inflammation. <i>J Biol Chem</i> 278: 14677-87, 2003.
84.	Horvath SE, Daum G. Lipids of mitochondria. <i>Prog Lipid Res</i> 52: 590-614, 2013.
85.	Jackson JC, Mozaffarian D, Graves AJ, Brown NJ, Marchioli R, Kiehl AL, Ely EW. Fish Oil Supplementation Does Not Affect Cognitive Outcomes in Cardiac Surgery Patients in the Omega-3 Fatty Acids for Prevention of Post-Operative Atrial Fibrillation (OPERA) Trial. <i>The</i> <i>Journal of nutrition</i> 148: 472-479, 2018.
86.	Jozsef L, Zouki C, Petasis NA, Serhan CN, Filep JG. Lipoxin A4 and aspirin-triggered 15-epi- lipoxin A4 inhibit peroxynitrite formation, NF-kappa B and AP-1 activation, and IL-8 gene expression in human leukocytes. <i>Proc Natl Acad Sci U S A</i> 99: 13266-71, 2002.
87.	Kain V, Ingle KA, Kachman M, Baum H, Shanmugam G, Rajasekaran NS, Young ME, Halade GV. Excess ω-6 fatty acids influx in aging drives metabolic dysregulation, electrocardiographic alterations, and low-grade chronic inflammation. <i>American journal of physiology</i> . <i>Heart and circulatory physiology</i> 314: H160-H169, 2018.
88.	Kang JW, Choi HS, Lee SM. Resolvin D1 attenuates liver ischaemia/reperfusion injury through modulating thioredoxin 2-mediated mitochondrial quality control. <i>Br J Pharmacol</i> 175: 2441-2453, 2018.
89.	Khasawneh FT, Huang JS, Mir F, Srinivasan S, Tiruppathi C, Le Breton GC. Characterization of isoprostane signaling: evidence for a unique coordination profile of 8-iso-PGF(2alpha) with the thromboxane A(2) receptor, and activation of a separate cAMP-dependent inhibitory pathway in human platelets. <i>Biochem Pharmacol</i> 75: 2301-15, 2008.
90.	Kim SY, Jeong JM, Kim SJ, Seo W, Kim MH, Choi WM, Yoo W, Lee JH, Shim YR, Yi HS, Lee YS, Eun HS, Lee BS, Chun K, Kang SJ, Kim SC, Gao B, Kunos G, Kim HM, Jeong WI. Pro- inflammatory hepatic macrophages generate ROS through NADPH oxidase 2 via endocytosis of monomeric TLR4-MD2 complex. <i>Nat Commun</i> 8: 2247, 2017.
91.	Koenitzer JR, Freeman BA. Redox signaling in inflammation: interactions of endogenous electrophiles and mitochondria in cardiovascular disease. <i>Ann N Y Acad Sci</i> 1203: 45-52, 2010.
92.	Kolaczkowska E. The older the faster: aged neutrophils in inflammation. <i>Blood</i> 128: 2280-2282, 2016.
93.	Kostecka-Gugała A, Latowski D, Strzałka K. Thermotropic phase behaviour of α- dipalmitoylphosphatidylcholine multibilayers is influenced to various extents by carotenoids containing different structural features- evidence from differential scanning calorimetry. <i>Biochimica et Biophysica Acta (BBA) - Biomembranes</i> 1609: 193-202, 2003.
94.	Kowal-Bielecka O, Kowal K, Distler O, Gay S. Mechanisms of Disease: leukotrienes and lipoxins in scleroderma lung diseaseinsights and potential therapeutic implications. <i>Nat Clin Pract Rheumatol</i> 3: 43-51, 2007.
95.	Krga I, Tamaian R, Mercier S, Boby C, Monfoulet LE, Glibetic M, Morand C, Milenkovic D. Anthocyanins and their gut metabolites attenuate monocyte adhesion and transendothelial migration through nutrigenomic mechanisms regulating endothelial cell permeability. <i>Free</i> <i>Radic Biol Med</i> 124: 364-379, 2018.
96.	Krga I, Vidovic N, Milenkovic D, Konic-Ristic A, Stojanovic F, Morand C, Glibetic M. Effects of anthocyanins and their gut metabolites on adenosine diphosphate-induced platelet activation and their aggregation with monocytes and neutrophils. <i>Arch Biochem Biophys</i> 645: 34-41, 2018.
97.	Kurundkar D, Kurundkar AR, Bone NB, Becker EJ, Jr., Liu W, Chacko B, Darley-Usmar V, Zmijewski JW, Thannickal VJ. SIRT3 diminishes inflammation and mitigates endotoxin- induced acute lung injury. <i>JCI Insight</i> 4, 2019.

2		
3	98.	Lee HE, Yang G, Park YB, Kang HC, Cho YY, Lee HS, Lee JY. Epigallocatechin-3-Gallate Prevents
4		Acute Gout by Suppressing NLRP3 Inflammasome Activation and Mitochondrial DNA
5		Synthesis. <i>Molecules</i> 24, 2019.
6 7	99.	Lee HN, Surh YJ. Resolvin D1-mediated NOX2 inactivation rescues macrophages undertaking
8		efferocytosis from oxidative stress-induced apoptosis. <i>Biochem Pharmacol</i> 86: 759-69, 2013.
9	100.	Levy BD, Clish CB, Schmidt B, Gronert K, Serhan CN. Lipid mediator class switching during
10		acute inflammation: signals in resolution. Nat Immunol 2: 612-9, 2001.
11	101.	Levy BD, Fokin VV, Clark JM, Wakelam MJ, Petasis NA, Serhan CN. Polyisoprenyl phosphate
12		(PIPP) signaling regulates phospholipase D activity: a 'stop' signaling switch for aspirin-
13		triggered lipoxin A4. FASEB J 13: 903-11, 1999.
14 15	102.	Li Y, Dalli J, Chiang N, Baron RM, Quintana C, Serhan CN. Plasticity of leukocytic exudates in
15 16		resolving acute inflammation is regulated by MicroRNA and proresolving mediators.
17		Immunity 39: 885-98, 2013.
18	103.	Liakh I, Pakiet A, Sledzinski T, Mika A. Modern Methods of Sample Preparation for the
19		Analysis of Oxylipins in Biological Samples. <i>Molecules</i> 24: 1639, 2019.
20	104.	Liakh I, Pakiet A, Sledzinski T, Mika A. Modern Methods of Sample Preparation for the
21		Analysis of Oxylipins in Biological Samples Molecules 24: 1639, 2019.
22	105.	Liu M, Zheng M, Cai D, Xie J, Jin Z, Liu H, Liu J. Zeaxanthin promotes mitochondrial biogenesis
23		and adipocyte browning via AMPKalpha1 activation. <i>Food Funct</i> 10: 2221-2233, 2019.
24	106.	Loke WM, Hodgson JM, Proudfoot JM, McKinley AJ, Puddey IB, Croft KD. Pure dietary
25 26		flavonoids quercetin and (-)-epicatechin augment nitric oxide products and reduce
20		endothelin-1 acutely in healthy men. Am J Clin Nutr 88: 1018-25, 2008.
28	107.	Lopategi A, Flores-Costa R, Rius B, Lopez-Vicario C, Alcaraz-Quiles J, Titos E, Claria J. Frontline
29		Science: Specialized proresolving lipid mediators inhibit the priming and activation of the
30		macrophage NLRP3 inflammasome. <i>J Leukoc Biol</i> 105: 25-36, 2019.
31	108.	Losito I, Facchini L, Diomede S, Conte E, Megli FM, Cataldi TRI, Palmisano F. Hydrophilic
32		interaction liquid chromatography-electrospray ionization-tandem mass spectrometry of a
33		complex mixture of native and oxidized phospholipids. <i>J Chromatogr A</i> 1422: 194-205, 2015.
34 35	109.	Lu H, Sun J, Liang W, Zhang J, Rom O, Garcia-Barrio MT, Li S, Villacorta L, Schopfer FJ,
36		Freeman BA, Chen YE, Fan Y. Novel gene regulatory networks identified in response to nitro-
37		conjugated linoleic acid in human endothelial cells. <i>Physiol Genomics</i> 51: 224-233, 2019.
38	110.	Lu J, Guo S, Xue X, Chen Q, Ge J, Zhuo Y, Zhong H, Chen B, Zhao M, Han W, Suzuki T, Zhu M,
39		Xia L, Schneider C, Blackwell TS, Porter NA, Zheng L, Tsimikas S, Yin H. Identification of a
40		novel series of anti-inflammatory and anti-oxidative phospholipid oxidation products
41		containing the cyclopentenone moiety in vitro and in vivo: Implication in atherosclerosis. J
42		Biol Chem 292: 5378-5391, 2017.
43 44	111.	Lütjohann D, Björkhem I, Locatelli S, Dame C, Schmolling J, Bergmann Kv, Fahnenstich H.
44		Cholesterol dynamics in the foetal and neonatal brain as reflected by circulatory levels of
46		24S-hydroxycholesterol. Acta Paediatrica 90: 652-657, 2001.
47	112.	Maddox JF, Hachicha M, Takano T, Petasis NA, Fokin VV, Serhan CN. Lipoxin A4 stable
48		analogs are potent mimetics that stimulate human monocytes and THP-1 cells via a G-
49		protein-linked lipoxin A4 receptor. J Biol Chem 272: 6972-8, 1997.
50	113.	Maloney B, Sambamurti K, Zawia N, Lahiri DK. Applying epigenetics to Alzheimer's disease
51		via the latent early-life associated regulation (LEARn) model. <i>Curr Alzheimer Res</i> 9: 589-99,
52 53		2012.
53 54	114.	Marcheselli VL, Hong S, Lukiw WJ, Tian XH, Gronert K, Musto A, Hardy M, Gimenez JM,
55		Chiang N, Serhan CN, Bazan NG. Novel docosanoids inhibit brain ischemia-reperfusion-
56		mediated leukocyte infiltration and pro-inflammatory gene expression. J Biol Chem 278:
57		43807-17, 2003.
58	115.	Marcheselli VL, Mukherjee PK, Arita M, Hong S, Antony R, Sheets K, Winkler JW, Petasis NA,
59		Serhan CN, Bazan NG. Neuroprotectin D1/protectin D1 stereoselective and specific binding
60		

with human retinal pigment epithelial cells and neutrophils. *Prostaglandins Leukot Essent Fatty Acids* 82: 27-34, 2010.

- 116. Martín-Venegas R, Casillas R, Jáuregui O, Moreno JJ. Rapid simultaneous analysis of cyclooxygenase, lipoxygenase and cytochrome P-450 metabolites of arachidonic and linoleic acids using high performance liquid chromatography/mass spectrometry in tandem mode. *Journal of Pharmaceutical and Biomedical Analysis* 56: 976-982, 2011.
- 117. Mauerhofer C, Philippova M, Oskolkova OV, Bochkov VN. Hormetic and anti-inflammatory properties of oxidized phospholipids. *Mol Aspects Med* 49: 78-90, 2016.
- 118. McCullough ML, Peterson JJ, Patel R, Jacques PF, Shah R, Dwyer JT. Flavonoid intake and cardiovascular disease mortality in a prospective cohort of US adults. *Am J Clin Nutr* 95: 454-64, 2012.
- 119. Meliton AY, Meng F, Tian Y, Sarich N, Mutlu GM, Birukova AA, Birukov KG. Oxidized phospholipids protect against lung injury and endothelial barrier dysfunction caused by heat-inactivated Staphylococcus aureus. *Am J Physiol Lung Cell Mol Physiol* 308: L550-62, 2015.
- 120. Merino-Serrais P, Loera-Valencia R, Rodriguez-Rodriguez P, Parrado-Fernandez C, Ismail MA, Maioli S, Matute E, Jimenez-Mateos EM, Bjorkhem I, DeFelipe J, Cedazo-Minguez A. 27-Hydroxycholesterol Induces Aberrant Morphology and Synaptic Dysfunction in Hippocampal Neurons. Cereb Cortex 29: 429-446, 2019.
- 121. Milenkovic D, Berghe WV, Morand C, Claude S, van de Sandt A, Gorressen S, Monfoulet LE, Chirumamilla CS, Declerck K, Szic KSV, Lahtela-Kakkonen M, Gerhauser C, Merx MW, Kelm M. A systems biology network analysis of nutri(epi)genomic changes in endothelial cells exposed to epicatechin metabolites. *Sci Rep* 8: 15487, 2018.
- 122. Mink PJ, Scrafford CG, Barraj LM, Harnack L, Hong CP, Nettleton JA, Jacobs DR, Jr. Flavonoid intake and cardiovascular disease mortality: a prospective study in postmenopausal women. *Am J Clin Nutr* 85: 895-909, 2007.
- 123. Mitchell S, Thomas G, Harvey K, Cottell D, Reville K, Berlasconi G, Petasis NA, Erwig L, Rees AJ, Savill J, Brady HR, Godson C. Lipoxins, aspirin-triggered epi-lipoxins, lipoxin stable analogues, and the resolution of inflammation: stimulation of macrophage phagocytosis of apoptotic neutrophils in vivo. *J Am Soc Nephrol* 13: 2497-507, 2002.
- 124. Miyahara T, Runge S, Chatterjee A, Chen M, Mottola G, Fitzgerald JM, Serhan CN, Conte MS. D-series resolvin attenuates vascular smooth muscle cell activation and neointimal hyperplasia following vascular injury. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 27: 2220-2232, 2013.
- 125. Moreno-Ulloa A, Nogueira L, Rodriguez A, Barboza J, Hogan MC, Ceballos G, Villarreal F, Ramirez-Sanchez I. Recovery of Indicators of Mitochondrial Biogenesis, Oxidative Stress, and Aging With (-)-Epicatechin in Senile Mice. *J Gerontol A Biol Sci Med Sci* 70: 1370-8, 2015.
- 126. Nadtochiy SM, Baker PR, Freeman BA, Brookes PS. Mitochondrial nitroalkene formation and mild uncoupling in ischaemic preconditioning: implications for cardioprotection. *Cardiovasc Res* 82: 333-40, 2009.
- 127. Nadtochiy SM, Zhu QM, Urciuoli W, Rafikov R, Black SM, Brookes PS. Nitroalkenes confer acute cardioprotection via adenine nucleotide translocase 1. *J Biol Chem* 287: 3573-80, 2012.
- 128. Nagy C, Haschemi A. Time and Demand are Two Critical Dimensions of Immunometabolism: The Process of Macrophage Activation and the Pentose Phosphate Pathway. *Front Immunol* 6: 164, 2015.
- 129. Nakayama H, Otsu K. Mitochondrial DNA as an inflammatory mediator in cardiovascular diseases. *Biochem J* 475: 839-852, 2018.
- 130. Narumiya S, Sugimoto Y, Ushikubi F. Prostanoid receptors: structures, properties, and functions. *Physiol Rev* 79: 1193-226, 1999.

2		
3	131.	Nelson KJ, Klomsiri C, Codreanu SG, Soito L, Liebler DC, Rogers LC, Daniel LW, Poole LB. Use
4		of dimedone-based chemical probes for sulfenic acid detection methods to visualize and
5		identify labeled proteins. <i>Methods Enzymol</i> 473: 95-115, 2010.
6 7	132.	Ni Z, Milic I, Fedorova M. Identification of carbonylated lipids from different phospholipid
8		classes by shotgun and LC-MS lipidomics. Anal Bioanal Chem 407: 5161-73, 2015.
9	133.	Ning Y, Bai Q, Lu H, Li X, Pandak WM, Zhao F, Chen S, Ren S, Yin L. Overexpression of
10		mitochondrial cholesterol delivery protein, StAR, decreases intracellular lipids and
11		inflammatory factors secretion in macrophages. Atherosclerosis 204: 114-20, 2009.
12	134.	Norris PC, Gosselin D, Reichart D, Glass CK, Dennis EA. Phospholipase A2 regulates
13		eicosanoid class switching during inflammasome activation. Proc Natl Acad Sci U S A 111:
14		12746-51, 2014.
15 16	135.	O'Donnell VB, Murphy RC. New families of bioactive oxidized phospholipids generated by
10		immune cells: identification and signaling actions. <i>Blood</i> 120: 1985-92, 2012.
18	136.	Oskolkova OV, Afonyushkin T, Preinerstorfer B, Bicker W, von Schlieffen E, Hainzl E,
19		Demyanets S, Schabbauer G, Lindner W, Tselepis AD, Wojta J, Binder BR, Bochkov VN.
20		Oxidized phospholipids are more potent antagonists of lipopolysaccharide than inducers of
21		inflammation. J Immunol 185: 7706-12, 2010.
22	137.	Ottaviani JI, Borges G, Momma TY, Spencer JP, Keen CL, Crozier A, Schroeter H. The
23	-	metabolome of [2-(14)C](-)-epicatechin in humans: implications for the assessment of
24		efficacy, safety, and mechanisms of action of polyphenolic bioactives. Sci. Rep. 6: 29034,
25 26		2016.
20 27	138.	Ottaviani JI, Heiss C, Spencer JPE, Kelm M, Schroeter H. Recommending flavanols and
28		procyanidins for cardiovascular health: Revisited. Mol Aspects Med 61: 63-75, 2018.
29	139.	Ottaviani JI, Kwik-Uribe C, Keen CL, Schroeter H. Intake of dietary procyanidins does not
30		contribute to the pool of circulating flavanols in humans. Am J Clin Nutr 95: 851-8, 2012.
31	140.	Ottaviani JI, Momma TY, Heiss C, Kwik-Uribe C, Schroeter H, Keen CL. The stereochemical
32		configuration of flavanols influences the level and metabolism of flavanols in humans and
33		their biological activity in vivo. Free Radic Biol Med 50: 237-44, 2011.
34 25	141.	Paardekooper LM, Dingjan I, Linders PTA, Staal AHJ, Cristescu SM, Verberk W, van den
35 36		Bogaart G. Human Monocyte-Derived Dendritic Cells Produce Millimolar Concentrations of
37		ROS in Phagosomes Per Second. Front Immunol 10: 1216, 2019.
38	142.	Pallauf K, Duckstein N, Rimbach G. A literature review of flavonoids and lifespan in model
39		organisms. Proc Nutr Soc 76: 145-162, 2017.
40	143.	Papassotiropoulos A, Lütjohann D, Bagli M, Locatelli S, Jessen F, Buschfort R, Ptok U,
41		Björkhem I, von Bergmann K, Heun R. 24S-hydroxycholesterol in cerebrospinal fluid is
42		elevated in early stages of dementia. <i>Journal of Psychiatric Research</i> 36: 27-32, 2002.
43	144.	Papayianni A, Serhan CN, Brady HR. Lipoxin A4 and B4 inhibit leukotriene-stimulated
44 45		interactions of human neutrophils and endothelial cells. J Immunol 156: 2264-72, 1996.
43 46	145.	Pararasa C, Ikwuobe J, Shigdar S, Boukouvalas A, Nabney IT, Brown JE, Devitt A, Bailey CJ,
47		Bennett SJ, Griffiths HR. Age-associated changes in long-chain fatty acid profile during
48		healthy aging promote pro-inflammatory monocyte polarization via PPARy. Aging cell 15:
49		128-139, 2016.
50	146.	Park K, Scott AL. Cholesterol 25-hydroxylase production by dendritic cells and macrophages
51		is regulated by type I interferons. <i>Journal of leukocyte biology</i> 88: 1081-1087, 2010.
52	147.	Peters-Golden M, Canetti C, Mancuso P, Coffey MJ. Leukotrienes: Underappreciated
53		Mediators of Innate Immune Responses. <i>The Journal of Immunology</i> 174: 589, 2005.
54 55	148.	Polidori MC, Mattioli P, Aldred S, Cecchetti R, Stahl W, Griffiths H, Senin U, Sies H, Mecocci P.
55 56		Plasma antioxidant status, immunoglobulin g oxidation and lipid peroxidation in demented
57		patients: relevance to Alzheimer disease and vascular dementia. <i>Dement Geriatr Cogn Disord</i>
58		18: 265-70, 2004.
59		,, <b></b> ,,,,,
60		

1	
2	
3	
4	
5	
6 7	
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o 9	
9	
10 11 12 13 14 15 16 17	
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46	
47	
48	
49	
50	
51	
52	
53	
55 54	
56	
57	
58	
59	
60	

149.	Preuss I, Ludwig M-G, Baumgarten B, Bassilana F, Gessier F, Seuwen K, Sailer AW. Transcriptional regulation and functional characterization of the oxysterol/EBI2 system in primary human macrophages. <i>Biochemical and Biophysical Research Communications</i> 446:
	663-668, 2014.
150.	Ravaglia G, Forti P, Maioli F, Brunetti N, Martelli M, Servadei L, Bastagli L, Bianchin M, Mariani E. Serum C-Reactive Protein and Cognitive Function in Healthy Elderly Italian Community Dwellers. <i>The Journals of Gerontology Series A: Biological Sciences and Medical</i> <i>Sciences</i> 60: 1017-1021, 2005.
151.	Regdon Z, Robaszkiewicz A, Kovacs K, Rygielska Z, Hegedus C, Bodoor K, Szabo E, Virag L. LPS protects macrophages from AIF-independent parthanatos by downregulation of PARP1 expression, induction of SOD2 expression, and a metabolic shift to aerobic glycolysis. <i>Free Radic Biol Med</i> 131: 184-196, 2019.
152.	Reis A. Oxidative Phospholipidomics in health and disease: Achievements, challenges and hopes. <i>Free Radical Biology and Medicine</i> 111: 25-37, 2017.
153.	Reis A, Spickett CM. Chemistry of phospholipid oxidation. <i>Biochim Biophys Acta</i> 1818: 2374-87, 2012.
154.	Ricciotti E, FitzGerald GA. Prostaglandins and inflammation. <i>Arteriosclerosis, thrombosis, and vascular biology</i> 31: 986-1000, 2011.
155.	Ricklin D, Hajishengallis G, Yang K, Lambris JD. Complement: a key system for immune surveillance and homeostasis. <i>Nat Immunol</i> 11: 785-97, 2010.
156.	Rodriguez-Mateos A, Heiss C, Borges G, Crozier A. Berry (Poly)phenols and Cardiovascular Health. <i>Journal of Agricultural and Food Chemistry</i> 62: 3842-3851, 2014.
157.	Rodriguez-Mateos A, Istas G, Boschek L, Feliciano RP, Mills CE, Boby C, Gomez-Alonso S, Milenkovic D, Heiss C. Circulating anthocyanin metabolites mediate vascular benefits of blueberries: insights from randomized controlled trials, metabolomics, and nutrigenomics. <i>J Gerontol A Biol Sci Med Sci</i> , 2019.
158.	Rodriguez-Mateos A, Rendeiro C, Bergillos-Meca T, Tabatabaee S, George TW, Heiss C, Spencer JPE. Intake and time dependence of blueberry flavonoid-induced improvements in vascular function: a randomized, controlled, double-blind, crossover intervention study with mechanistic insights into biological activity. <i>American Journal of Clinical Nutrition</i> 98: 1179-1191, 2013.
159.	Rodriguez-Prados JC, Traves PG, Cuenca J, Rico D, Aragones J, Martin-Sanz P, Cascante M, Bosca L. Substrate fate in activated macrophages: a comparison between innate, classic, and alternative activation. <i>J Immunol</i> 185: 605-14, 2010.
160.	Romano M, Serhan CN. Lipoxin generation by permeabilized human platelets. <i>Biochemistry</i> 31: 8269-77, 1992.
161.	Romero F, Shah D, Duong M, Penn RB, Fessler MB, Madenspacher J, Stafstrom W, Kavuru M, Lu B, Kallen CB, Walsh K, Summer R. A pneumocyte-macrophage paracrine lipid axis drives the lung toward fibrosis. <i>Am J Respir Cell Mol Biol</i> 53: 74-86, 2015.
162.	Rudolph V, Schopfer FJ, Khoo NK, Rudolph TK, Cole MP, Woodcock SR, Bonacci G, Groeger AL, Golin-Bisello F, Chen CS, Baker PR, Freeman BA. Nitro-fatty acid metabolome: saturation, desaturation, beta-oxidation, and protein adduction. <i>J Biol Chem</i> 284: 1461-73, 2009.
163.	Rund KM, Ostermann AI, Kutzner L, Galano J-M, Oger C, Vigor C, Wecklein S, Seiwert N, Durand T, Schebb NH. Development of an LC-ESI(-)-MS/MS method for the simultaneous quantification of 35 isoprostanes and isofurans derived from the major n3- and n6-PUFAs. <i>Analytica Chimica Acta</i> 1037: 63-74, 2018.
164.	Ryan VH, Primiani CT, Rao JS, Ahn K, Rapoport SI, Blanchard H. Coordination of gene expression of arachidonic and docosahexaenoic acid cascade enzymes during human brain development and aging. <i>PloS one</i> 9: e100858-e100858, 2014.
165.	Salem N, Jr., Litman B, Kim HY, Gawrisch K. Mechanisms of action of docosahexaenoic acid in the nervous system. <i>Lipids</i> 36: 945-59, 2001.

166.

167.

168.

169.

170.

171.

172.

173.

174.

175.

176.

177.

178.

179.

180.

181.

2014.

Samuelsson B. Leukotrienes: mediators of immediate hypersensitivity reactions and

Sanin DE, Matsushita M, Klein Geltink RI, Grzes KM, van Teijlingen Bakker N, Corrado M,

Kabat AM, Buck MD, Qiu J, Lawless SJ, Cameron AM, Villa M, Baixauli F, Patterson AE, Hassler F, Curtis JD, O'Neill CM, O'Sullivan D, Wu D, Mittler G, Huang SC, Pearce EL, Pearce EJ. Mitochondrial Membrane Potential Regulates Nuclear Gene Expression in Macrophages

Schaich KM. CHAPTER 1 - Challenges in Elucidating Lipid Oxidation Mechanisms: When, Where, and How Do Products Arise? In: *Lipid Oxidation*. edited by Logan A, Nienaber U, Pan

Schewe T, Steffen Y, Sies H. How do dietary flavanols improve vascular function? A position

Schopfer FJ, Batthyany C, Baker PR, Bonacci G, Cole MP, Rudolph V, Groeger AL, Rudolph TK, Nadtochiy S, Brookes PS, Freeman BA. Detection and quantification of protein adduction by electrophilic fatty acids: mitochondrial generation of fatty acid nitroalkene derivatives. *Free* 

Schroeter H, Heiss C, Balzer J, Kleinbongard P, Keen CL, Hollenberg NK, Sies H, Kwik-Uribe C, Schmitz HH, Kelm M. (-)-Epicatechin mediates beneficial effects of flavanol-rich cocoa on

Schuler D, Sansone R, Freudenberger T, Rodriguez-Mateos A, Weber G, Momma TY, Goy C, Altschmied J, Haendeler J, Fischer JW, Kelm M, Heiss C. Measurement of endotheliumdependent vasodilation in mice--brief report. *Arterioscler Thromb Vasc Biol* 34: 2651-7,

Serbulea V, Upchurch CM, Ahern KW, Bories G, Voigt P, DeWeese DE, Meher AK, Harris TE,

Leitinger N. Macrophages sensing oxidized DAMPs reprogram their metabolism to support redox homeostasis and inflammation through a TLR2-Syk-ceramide dependent mechanism.

Serbulea V, Upchurch CM, Schappe MS, Voigt P, DeWeese DE, Desai BN, Meher AK, Leitinger N. Macrophage phenotype and bioenergetics are controlled by oxidized phospholipids

identified in lean and obese adipose tissue. Proc Natl Acad Sci U S A 115: E6254-e6263, 2018.

Serhan CN. Lipoxins and aspirin-triggered 15-epi-lipoxins are the first lipid mediators of endogenous anti-inflammation and resolution. *Prostaglandins Leukot Essent Fatty Acids* 73:

Serhan CN. Resolution phase of inflammation: novel endogenous anti-inflammatory and

Serhan CN, Brain SD, Buckley CD, Gilroy DW, Haslett C, O'Neill LAJ, Perretti M, Rossi AG,

Wallace JL. Resolution of inflammation: state of the art, definitions and terms. *FASEB journal* : official publication of the Federation of American Societies for Experimental Biology 21:

Serhan CN, Chiang N, Dalli J. The resolution code of acute inflammation: Novel pro-resolving

Serhan CN, Chiang N, Van Dyke TE. Resolving inflammation: dual anti-inflammatory and pro-

Serhan CN, Clish CB, Brannon J, Colgan SP, Chiang N, Gronert K. Novel functional sets of lipidderived mediators with antiinflammatory actions generated from omega-3 fatty acids via cyclooxygenase 2-nonsteroidal antiinflammatory drugs and transcellular processing. *J Exp* 

33

proresolving lipid mediators and pathways. Annu Rev Immunol 25: 101-37, 2007.

vascular function in humans. Proc. Natl. Acad. Sci. USA 103: 1024-1029, 2006.

Schwab JM, Chiang N, Arita M, Serhan CN. Resolvin E1 and protectin D1 activate

inflammation-resolution programmes. Nature 447: 869-74, 2007.

lipid mediators in resolution. Semin Immunol 27: 200-15, 2015.

resolution lipid mediators. Nat Rev Immunol 8: 349-61, 2008.

Exposed to Prostaglandin E2. Immunity 49: 1021-1033.e6, 2018.

paper. Archives of Biochemistry and Biophysics 476: 102-106, 2008.

inflammation. Science 220: 568-75, 1983.

X. AOCS Press; 2013. pp. 1-52.

Radic Biol Med 46: 1250-9, 2009.

Mol Metab 7: 23-34, 2018.

141-62, 2005.

325-332, 2007.

Med 192: 1197-204, 2000.

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2 3 4 5	
4	
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6 7 8	
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- Serhan CN, Dalli J, Colas RA, Winkler JW, Chiang N. Protectins and maresins: New pro-resolving families of mediators in acute inflammation and resolution bioactive metabolome.
  *Biochim Biophys Acta* 1851: 397-413, 2015.
- Serhan CN, Hong S, Gronert K, Colgan SP, Devchand PR, Mirick G, Moussignac RL. Resolvins:
  a family of bioactive products of omega-3 fatty acid transformation circuits initiated by aspirin treatment that counter proinflammation signals. J Exp Med 196: 1025-37, 2002.
- 184. Serhan CN, Maddox JF, Petasis NA, Akritopoulou-Zanze I, Papayianni A, Brady HR, Colgan SP, Madara JL. Design of lipoxin A4 stable analogs that block transmigration and adhesion of human neutrophils. *Biochemistry* 34: 14609-15, 1995.
- 185. Serra G, Deiana M, Spencer JPE, Corona G. Olive Oil Phenolics Prevent Oxysterol-Induced Proinflammatory Cytokine Secretion and Reactive Oxygen Species Production in Human Peripheral Blood Mononuclear Cells, Through Modulation of p38 and JNK Pathways. *Mol Nutr Food Res* 61, 2017.
- 186. Sheedy FJ, Grebe A, Rayner KJ, Kalantari P, Ramkhelawon B, Carpenter SB, Becker CE, Ediriweera HN, Mullick AE, Golenbock DT, Stuart LM, Latz E, Fitzgerald KA, Moore KJ. CD36 coordinates NLRP3 inflammasome activation by facilitating intracellular nucleation of soluble ligands into particulate ligands in sterile inflammation. *Nat Immunol* 14: 812-20, 2013.
- 187. Shepardson Ne SGMSDJ. Cholesterol level and statin use in alzheimer disease: I. review of epidemiological and preclinical studies. *Archives of Neurology* 68: 1239-1244, 2011.
- 188. Shirai T, Nazarewicz RR, Wallis BB, Yanes RE, Watanabe R, Hilhorst M, Tian L, Harrison DG, Giacomini JC, Assimes TL, Goronzy JJ, Weyand CM. The glycolytic enzyme PKM2 bridges metabolic and inflammatory dysfunction in coronary artery disease. *J Exp Med* 213: 337-54, 2016.
- 189. Smith WL, DeWitt DL, Garavito RM. Cyclooxygenases: structural, cellular, and molecular biology. *Annu Rev Biochem* 69: 145-82, 2000.
- 190. Snowden SG, Ebshiana AA, Hye A, An Y, Pletnikova O, O'Brien R, Troncoso J, Legido-Quigley C, Thambisetty M. Association between fatty acid metabolism in the brain and Alzheimer disease neuropathology and cognitive performance: A nontargeted metabolomic study. *PLoS Med* 14: e1002266, 2017.
- 191. Sohrabi Y, Lagache SMM, Schnack L, Godfrey R, Kahles F, Bruemmer D, Waltenberger J, Findeisen HM. mTOR-Dependent Oxidative Stress Regulates oxLDL-Induced Trained Innate Immunity in Human Monocytes. *Front Immunol* 9: 3155, 2018.
- 192. Solheim S, Hutchinson SA, Lundanes E, Wilson SR, Thorne JL, Roberg-Larsen H. Fast liquid chromatography-mass spectrometry reveals side chain oxysterol heterogeneity in breast cancer tumour samples. *The Journal of Steroid Biochemistry and Molecular Biology* 192: 105309, 2019.
- 193. Starosta V, Wu T, Zimman A, Pham D, Tian X, Oskolkova O, Bochkov V, Berliner JA, Birukova AA, Birukov KG. Differential regulation of endothelial cell permeability by high and low doses of oxidized 1-palmitoyl-2-arachidonyl-sn-glycero-3-phosphocholine. *Am J Respir Cell Mol Biol* 46: 331-41, 2012.
- 194. Stemmer U, Ramprecht C, Zenzmaier E, Stojcic B, Rechberger G, Kollroser M, Hermetter A. Uptake and protein targeting of fluorescent oxidized phospholipids in cultured RAW 264.7 macrophages. *Biochim Biophys Acta* 1821: 706-18, 2012.
- 195. Su G, Zhang T, Yang HX, Dai WL, Wang T, Tian L, Mi SH. Association of Isoprostanes-Related Oxidative Stress with Vulnerability of Culprit Lesions in Diabetic Patients with Acute Coronary Syndrome. *Int Heart J* 60: 271-279, 2019.
- 196. Sun N, Youle RJ, Finkel T. The Mitochondrial Basis of Aging. *Mol Cell* 61: 654-666, 2016.
  - 197. Sun Q, Wu Y, Zhao F, Wang J. Maresin 1 Ameliorates Lung Ischemia/Reperfusion Injury by Suppressing Oxidative Stress via Activation of the Nrf-2-Mediated HO-1 Signaling Pathway. Oxid Med Cell Longev 2017: 9634803, 2017.

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Tager AM, Luster AD. BLT1 and BLT2: the leukotriene B(4) receptors. Prostaglandins Leukot

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199.	<i>Essent Fatty Acids</i> 69: 123-34, 2003. Tani M, Kamata Y, Deushi M, Osaka M, Yoshida M. 7-Ketocholesterol enhances leukocy
155.	adhesion to endothelial cells via p38MAPK pathway. <i>PLoS One</i> 13: e0200499, 2018.
200.	Testa G, Staurenghi E, Zerbinati C, Gargiulo S, Iuliano L, Giaccone G, Fanto F, Poli G,
200.	Leonarduzzi G, Gamba P. Changes in brain oxysterols at different stages of Alzheimer's
	disease: Their involvement in neuroinflammation. <i>Redox Biol</i> 10: 24-33, 2016.
201.	Thomas DW, Rocha PN, Nataraj C, Robinson LA, Spurney RF, Koller BH, Coffman TM.
201.	
	Proinflammatory actions of thromboxane receptors to enhance cellular immune respo
202	<i>Immunol</i> 171: 6389-95, 2003. Tilley SL, Coffman TM, Koller BH. Mixed messages: modulation of inflammation and im
202.	
202	responses by prostaglandins and thromboxanes. <i>J Clin Invest</i> 108: 15-23, 2001.
203.	Torr EE, Gardner DH, Thomas L, Goodall DM, Bielemeier A, Willetts R, Griffiths HR, Mar
	LJ, Devitt A. Apoptotic cell-derived ICAM-3 promotes both macrophage chemoattractic
204	and tethering of apoptotic cells. <i>Cell Death Differ</i> 19: 671-9, 2012.
204.	Torrao RC, Bennett SJ, Brown JE, Griffiths HR. Does metabolic reprogramming underpir
	associated changes in T cell phenotype and function? Free Radic Biol Med 71: 26-35, 20
205.	Tresserra-Rimbau A, Castro-Barquero S, Vitelli-Storelli F, Becerra-Tomas N, Vázquez-Ru
	Díaz-López A, Corella D, Castañer O, Romaguera D, Vioque J, Alonso-Gómez ÁM, Wärnl
	Martínez JA, Serra-Majem L, Estruch R, Tinahones FJ, Lapetra J, Pintó X, Tur JA, López-
	Miranda J, García-Molina L, Delgado-Rodríguez M, Matía-Martín P, Daimiel L, Rubín-Ga
	M, Vidal J, Galdon A, Ros E, Basterra-Gortari FJ, Babio N, Sorlí JV, Hernáez Á, Konieczna
	Notario-Barandiaran L, Tojal-Sierra L, Pérez-López J, Abete I, Álvarez-Pérez J, Fernández
	García JC, Santos-Lozano JM, Galera-Cusí A, Julibert A, Ruiz-Canela M, Martinez-Lacruz
	Pérez-Vega K-A, Galmes-Panades AM, Pastor-Polo C, Moreno-Rodriguez A, Gea A, Fitó
	Lamuela-Raventós RM, Salas-Salvadó J. Associations between Dietary Polyphenols and
	2 Diabetes in a Cross-Sectional Analysis of the PREDIMED-Plus Trial: Role of Body Mass
	and Sex. Antioxidants 8: 537, 2019.
206.	Trostchansky A, Rubbo H. Anti-inflammatory signaling actions of electrophilic nitro-
	arachidonic acid in vascular cells and astrocytes. Arch Biochem Biophys 617: 155-161, 2
207.	Uderhardt S, Martins AJ, Tsang JS, Lammermann T, Germain RN. Resident Macrophage
	Cloak Tissue Microlesions to Prevent Neutrophil-Driven Inflammatory Damage. Cell 17
	541-555.e17, 2019.
208.	Umetani M, Ghosh P, Ishikawa T, Umetani J, Ahmed M, Mineo C, Shaul Philip W. The
	Cholesterol Metabolite 27-Hydroxycholesterol Promotes Atherosclerosis via
	Proinflammatory Processes Mediated by Estrogen Receptor Alpha. Cell Metabolism 20
	182, 2014.
209.	van den Kommer TN, Dik MG, Comijs HC, Fassbender K, Lütjohann D, Jonker C. Total
	cholesterol and oxysterols: Early markers for cognitive decline in elderly? Neurobiology
	Aging 30: 534-545, 2009.
210.	Vandanmagsar B, Youm YH, Ravussin A, Galgani JE, Stadler K, Mynatt RL, Ravussin E,
	Stephens JM, Dixit VD. The NLRP3 inflammasome instigates obesity-induced inflammat
	and insulin resistance. Nat Med 17: 179-88, 2011.
211.	Viaud M, Ivanov S, Vujic N, Duta-Mare M, Aira LE, Barouillet T, Garcia E, Orange F, Duga
	Hainault I, Stehlik C, Marchetti S, Boyer L, Guinamard R, Foufelle F, Bochem A, Hovingh
	Thorp EB, Gautier EL, Kratky D, Dasilva-Jardine P, Yvan-Charvet L. Lysosomal Cholester
	Hydrolysis Couples Efferocytosis to Anti-Inflammatory Oxysterol Production. Circ Res 1
	1369-1384, 2018.
212.	Vigor C, Bertrand-Michel J, Pinot E, Oger C, Vercauteren J, Le Faouder P, Galano JM, Le
	Durand T. Non-enzymatic lipid oxidation products in biological systems: assessment of

metabolites from polyunsaturated fatty acids. J Chromatogr B Analyt Technol Biomed Life Sci 964: 65-78, 2014.

- 213. Villacorta L, Minarrieta L, Salvatore SR, Khoo NK, Rom O, Gao Z, Berman RC, Jobbagy S, Li L, Woodcock SR, Chen YE, Freeman BA, Ferreira AM, Schopfer FJ, Vitturi DA. In situ generation, metabolism and immunomodulatory signaling actions of nitro-conjugated linoleic acid in a murine model of inflammation. *Redox Biol* 15: 522-531, 2018.
- 214. Vine DF, Mamo JCL, Beilin LJ, Mori TA, Croft KD. Dietary oxysterols are incorporated in plasma triglyceriderich lipoproteins, increase their susceptibility to oxidation and increase aortic cholesterol concentration of rabbits. *Journal of Lipid Research* 39: 1995-2004, 1998.
- 215. Vogiatzoglou A, Mulligan AA, Bhaniani A, Lentjes MA, McTaggart A, Luben RN, Heiss C, Kelm M, Merx MW, Spencer JP, Schroeter H, Khaw KT, Kuhnle GG. Associations between flavan-3-ol intake and CVD risk in the Norfolk cohort of the European Prospective Investigation into Cancer (EPIC-Norfolk). Free Radic Biol Med 84: 1-10, 2015.
- 216. Vogiatzoglou A, Mulligan AA, Lentjes MA, Luben RN, Spencer JP, Schroeter H, Khaw KT, Kuhnle GG. Flavonoid intake in European adults (18 to 64 years). *PLoS One* 10: e0128132, 2015.
- 217. Vurusaner B, Gamba P, Gargiulo S, Testa G, Staurenghi E, Leonarduzzi G, Poli G, Basaga H. Nrf2 antioxidant defense is involved in survival signaling elicited by 27-hydroxycholesterol in human promonocytic cells. *Free Radical Biology and Medicine* 91: 93-104, 2016.
- 218. Vurusaner B, Gamba P, Testa G, Gargiulo S, Biasi F, Zerbinati C, Iuliano L, Leonarduzzi G, Basaga H, Poli G. Survival signaling elicited by 27-hydroxycholesterol through the combined modulation of cellular redox state and ERK/Akt phosphorylation. *Free Radical Biology and Medicine* 77: 376-385, 2014.
- 219. Vurusaner B, Gargiulo S, Testa G, Gamba P, Leonarduzzi G, Poli G, Basaga H. The role of autophagy in survival response induced by 27-hydroxycholesterol in human promonocytic cells. *Redox Biology* 17: 400-410, 2018.
- 220. Wan M, Hua X, Su J, Thiagarajan D, Frostegård AG, Haeggström JZ, Frostegård J. Oxidized but not native cardiolipin has pro-inflammatory effects, which are inhibited by Annexin A5. *Atherosclerosis* 235: 592-598, 2014.
- 221. Wang W, Qin S, Li L, Chen X, Wang Q, Wei J. An Optimized High Throughput Clean-Up Method Using Mixed-Mode SPE Plate for the Analysis of Free Arachidonic Acid in Plasma by LC-MS/MS. *International journal of analytical chemistry* 2015: 374819-374819, 2015.
- 222. Wang X, Ouyang YY, Liu J, Zhao G. Flavonoid intake and risk of CVD: a systematic review and meta-analysis of prospective cohort studies. *Br J Nutr* 111: 1-11, 2014.
- 223. Wang Y, Tabas I. Emerging roles of mitochondria ROS in atherosclerotic lesions: causation or association? *J Atheroscler Thromb* 21: 381-90, 2014.
- 224. Wang Y, Wang W, Wang N, Tall AR, Tabas I. Mitochondrial Oxidative Stress Promotes Atherosclerosis and Neutrophil Extracellular Traps in Aged Mice. *Arterioscler Thromb Vasc Biol* 37: e99-e107, 2017.
- 225. Watanabe Y, Nagai Y, Honda H, Okamoto N, Yamamoto S, Hamashima T, Ishii Y, Tanaka M, Suganami T, Sasahara M, Miyake K, Takatsu K. Isoliquiritigenin Attenuates Adipose Tissue Inflammation in vitro and Adipose Tissue Fibrosis through Inhibition of Innate Immune Responses in Mice. *Sci Rep* 6: 23097, 2016.
- 226. Wen H, Gris D, Lei Y, Jha S, Zhang L, Huang MT, Brickey WJ, Ting JP. Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signaling. *Nat Immunol* 12: 408-15, 2011.
- 227. Williamson G, Sies H, Heber D, Keen CL, Macdonald IA, ctis-Gorreta L, Momma TY, Ottaviani JI, Holt RR, Schroeter H, Heiss C. Functional foods for health promotion: state-of-the-science on dietary flavonoids Extended abstracts from the 12 Annual Conference on Functional Foods for Health Promotion, April 2009. *Nutrition Reviews* 67: 736-743, 2009.

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- Xu XR, Zou ZY, Xiao X, Huang YM, Wang X, Lin XM. Effects of lutein supplement on serum inflammatory cytokines, ApoE and lipid profiles in early atherosclerosis population. J Atheroscler Thromb 20: 170-7, 2013.
- Yeon SH, Yang G, Lee HE, Lee JY. Oxidized phosphatidylcholine induces the activation of NLRP3 inflammasome in macrophages. J Leukoc Biol 101: 205-215, 2017.
- Youm YH, Grant RW, McCabe LR, Albarado DC, Nguyen KY, Ravussin A, Pistell P, Newman S, Carter R, Lague A, Munzberg H, Rosen CJ, Ingram DK, Salbaum JM, Dixit VD. Canonical NIrp3 inflammasome links systemic low-grade inflammation to functional decline in aging. Cell Metab 18: 519-32, 2013.
- Youm YH, Kanneganti TD, Vandanmagsar B, Zhu X, Ravussin A, Adijiang A, Owen JS, Thomas MJ, Francis J, Parks JS, Dixit VD. The NIrp3 inflammasome promotes age-related thymic demise and immunosenescence. Cell Rep 1: 56-68, 2012.
- Zemski Berry KA, Murphy RC. Phospholipid Ozonation Products Activate the 5-Lipoxygenase Pathway in Macrophages. Chem Res Toxicol 29: 1355-64, 2016.
- Zhang MJ, Sansbury BE, Hellmann J, Baker JF, Guo L, Parmer CM, Prenner JC, Conklin DJ, Bhatnagar A, Creager MA, Spite M. Resolvin D2 Enhances Postischemic Revascularization While Resolving Inflammation. Circulation 134: 666-680, 2016.
- Zhong W, Springstead JR, Al-Mubarak R, Lee S, Li R, Emert B, Berliner JA, Jung ME. An epoxyisoprostane is a major regulator of endothelial cell function. J Med Chem 56: 8521-32, 2013.
- Zou ZY, Xu XR, Lin XM, Zhang HB, Xiao X, Ouyang L, Huang YM, Wang X, Liu YQ. Effects of lutein and lycopene on carotid intima-media thickness in Chinese subjects with subclinical atherosclerosis: a randomised, double-blind, placebo-controlled trial. Br J Nutr 111: 474-80, 2014.

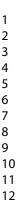
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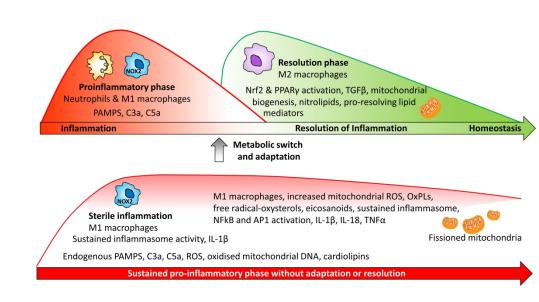
# List of Abbreviations

5-LOX	5-lipoxygenase
7α,25-ΟΗC	7α, 25-dihydroxycholesterol
7β-ΟΗC	7β-hydroxycholesterol
7-KC	7-ketocholesterol
12-LOX	12-lipoxygenase
12/15 LOX	12/15 lipoxygenase
15-HETE	15-hydroxy-eicosatetraenoic acid
15-LOX	15-lipoxygenase
18-HEPE	18-hydroxy-eicosapentacnoic acid
24-OHC	24S-hydroxycholesterol
25-OHC	25-hydroxycholesterol
27-OHC	27-hydroxycholesterol
AA	arachidonic acid
ABCD1	ATP-binding cassette transporter D1
ABCA1	ATP-binding cassette transporter A1
ABCG1	ATP binding cassette transporter G1
ACOX1	acyl-CoA oxidase 1
ADME	absorption, distribution, metabolism, a
	excretion
ALX	lipoxin A4 receptor
AP1	Activator Protein 1
COX	cyclooxygenases
CXCR2	C-X-C motif chemokine receptor 2
D6D	Delta 6 desaturase
DAMPS	damage-associated molecular patterns
DHA	Docosahexaenoic acid
EBI2	Epstein-Barr-virus-induced G-protein coupl
	receptor 2
ER	Endoplasmic reticulum
Gclm	glutamate cysteine ligase
GPCR	G protein coupled receptors
HIF1α	Hypoxia-inducible factor 1-alpha
HMGR	3-hydroxy-3-methylglutaryl CoA reductase
Ho-1	haemoxygenase 1
IC50	inhibitory concentration
INSIG	insulin induced gene protein
KEAP-1	Kelch Like ECH Associated Protein 1
LEARn	latent early life-associated regulation
LC3	light chain 3
LDL	low density lipoprotein
LDLR	LDL receptor
LOX	lipoxygenases
LPS	lipopolysaccharide
LtB <sub>4</sub>	leukotriene B <sub>4</sub>
Lts	leukotrienes
LxA <sub>4</sub>	lipoxin A <sub>4</sub>
LxB <sub>4</sub>	lipoxin B <sub>4</sub>
LXR	Liver X receptors

lyso-PC	lysophosphatidyl choline
MCI	mild cognitive impairment
NADH	Nicotinamide adenine dinucleotide
NADPH	nicotinamide adenine dinucleotide phospha oxidase
NET	neutrophil extracellular traps
NLR	NOD-like receptors
NLRP	nucleotide-binding domain, leucine-ric
	containing family, pyrin domain-containing
•NO	nitric oxide
•NO <sub>2</sub>	nitrogen dioxide
NOD	nucleotide-binding oligomerization domain
NOS	nitric oxide synthases
NOX	(NADPH) oxidase
NPC1	
	Niemann-Pick protein C1
Nrf2	nuclear factor erythroid 2-related factor 2
OSBP	oxysterol binding protein
ORP	OSBP-related proteins
OxPLs	Oxidised phospholipids
PAPC	1-palmitoyl-2-arachidonyl-sn-glycero-3-
	phosphorylcholine
PEIPC	epoxyisoprostane
PECPC	epoxycyclopentenone
PD1	protectin D1
PGs	prostaglandins
PGE <sub>2</sub>	prostaglandin E <sub>2</sub>
PGD <sub>2</sub>	prostaglandin D <sub>2</sub>
PGF <sub>2α</sub>	prostaglandin $F_{2\alpha}$
PGPC	1-palmitoyl-2-glutaroyl-sn-glycero- phosphatidylcholine
PLA2	Phospholipase A2
POVPC	1-palmitoyl-2-(5-oxovaleroyl)-sn-glycero-
	phosphatidylcholine
ΡΡΑRγ	peroxisome proliferator-activated recept
	gamma
PUFA	polyunsaturated fatty acids
PRR	pattern recognition receptors
ResE1	resolvin E1
ResE2	resolvin E2
RNS	reactive nitrogen species
RXR	Retinoid X receptor
RAR	Retinoic acid receptor
ROS	reactive oxygen species
SOD2	superoxide dismutase-2
SIRT	sirtuin
SPM	specialised pro-resolving lipid mediators inflammation
SREBP	sterol regulatory element binding protein
StAR	Steroidogenic acute regulatory protein
TBXA2R	thromboxane A2 receptor

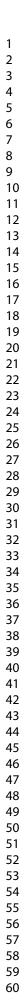
Antioxidants & Redox Signaling		Page 40 of 46
ΤGFβ	Transforming growth factor beta	
TLR	Toll-like receptor	
TSPO	Mitochondrial translocator protein	
TXs	thromboxanes	
TXA <sub>2</sub>	Thromboxane A <sub>2</sub>	
Txnrd1	thioredoxin reductase	
VE-cadherin	vascular endothelial cadherin	
e destroy all records after use for peer revie	ew. Mary Ann Liebert Inc., 140 Huguenot Street, New Rochelle, N	<b>40</b> Y 10801

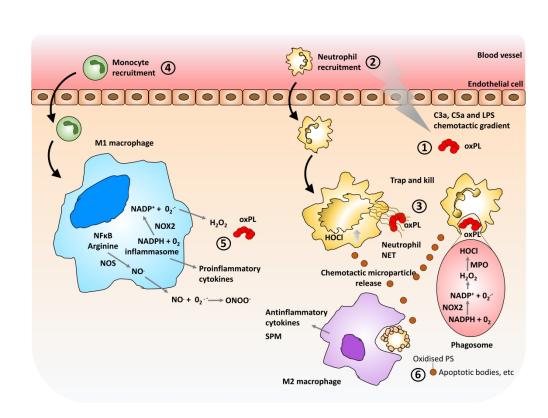




Key mediators and cells involved in physiological and sterile inflammation responses

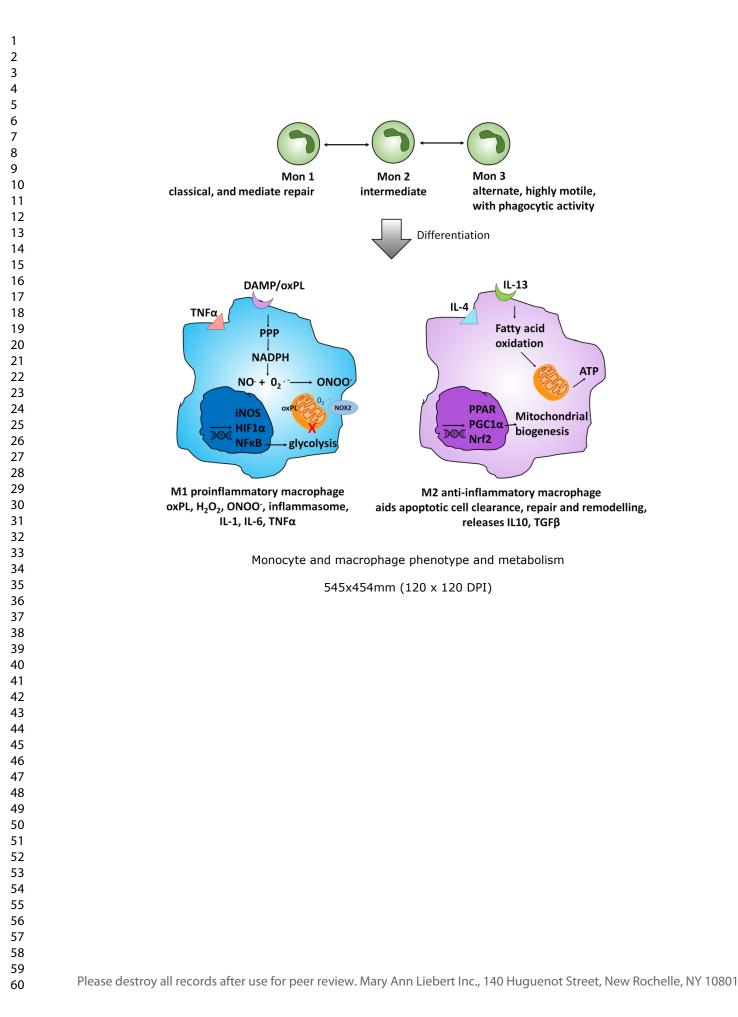
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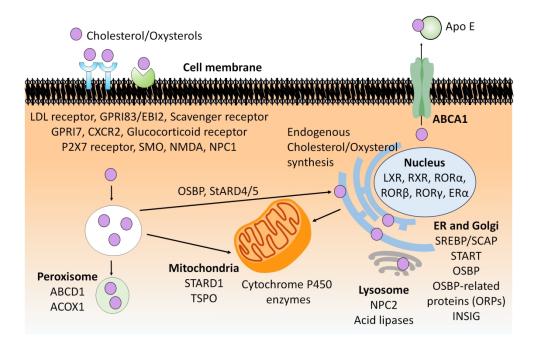




Innate immune recruitment to inflammatory sites; oxidised phospholipids contribute to the chemotactic gradient for recruitment of inflammatory cells. Their local production in the mitochondria results in impaired mitochondrial activity and failure to remove damaged mitochondria exacerbates ROS production. MtDNA and OxPL enhance inflammasome activation and pro-inflammatory cytokine release.

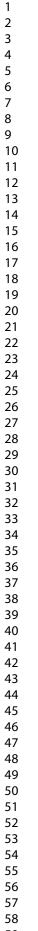
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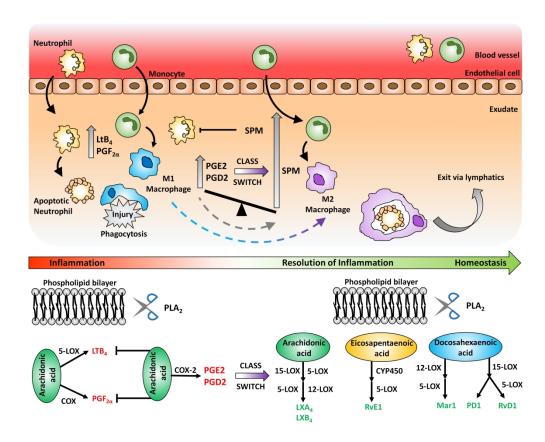




Cellular distribution of oxysterols. Cellular cholesterol is maintained by regulatory mechanisms that control synthesis, uptake, metabolism and efflux. Cells uptake cholesterol and oxysterols packed lipoproteins aided by many membrane receptors, such as LDL receptor (LDLR) family, EBI2 , G-protein coupled receptor, GPR17, C-X-C motif chemokine receptor 2 (CXCR2) and scavenger receptors. Inside the cell, endosomes containing cholesterol/oxysterol are transported to Endoplasmic reticulum (ER) mitochondria or peroxisomes. Within ER, sterols are further metabolised with help of many protein complexes including, oxysterol binding protein (OSBP), OSBP-related proteins (ORPs), the cellular nucleic acid binding protein, the sterol regulatory element binding protein (SREBP), insulin induced gene protein (INSIG) and Niemann-Pick protein C1 (NPC1). These metabolic activities within ER and golgi transfer signals to nucleus either to up- or down regulate endogenous cholesterol synthesis via 3-hydroxy-3-methylglutaryl CoA reductase (HMGR) pathway. Cellular cholesterol and oxysterol levels are also sensed a family of nuclear receptors family; Liver X receptors (LXR), Retinoid X receptor (RXR), Retinoic acid receptor (RAR)-related orphan proteins (RORa, RORβ and RORγ) and Estrogen receptor a (ERa). Activation of these transcription factors upregulate multiple genes involved in cellular cholesterol homeostasis, including ATP-binding cassette transporter A1 (ABCA1), ATP binding cassette transporter G1 (ABCG1), and APOE. Mitochondrial translocator protein (TSPO) has high affinity for cholesterol and uptake cholesterol from ER or from lipid droplets. Steroidogenic acute regulatory protein (StAR) transport cholesterol from the outer to inner membrane in mitochondria. Peroxisome proteins ATP-binding cassette transporter D1 (ABCD1) and acyl-CoA oxidase 1 (ACOX1) are also involve in transport of cholesterol and oxysterols.

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Lipid-derived mediators of inflammation are key players that drive both pro-inflammatory and antiinflammatory and pro-resolving responses. Infiltration of neutrophils from blood vessels into the site of injury, and later pro-inflammatory macrophages (M1 macrophages), is supported by leukotriene B4 and prostaglandin F2a. Resolution phase is initiated by the synthesis of prostaglandins that induce "class switch" from pro-inflammatory (LtB4, PGF2a) to the specialised pro-resolving mediators (SPM, such as lipoxins, resolvins, protectins and maresins). Their dual acting nature stops neutrophils from further infiltrating to the sites of injury, while supporting chemotaxis of pro-repair macrophages (M2 macrophages), phagocytosis of apoptotic material, and their exit via lymphatics.

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