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## MORPHOLOGICAL DIVERSITY OF *Meloidogyne* spp. FROM CARROT (*Daucus carota* subsp. *Sativus*) IN VIETNAM

THI DUYEN NGUYEN<sup>1,2</sup>, HUU TIEN NGUYEN<sup>1</sup>, NERIZA NOBLEZA<sup>3</sup>,  
THI MAI LINH LE<sup>1,2</sup> AND QUANG PHAP TRINH<sup>1,2\*</sup>

<sup>1</sup>Institute of Ecology and Biological Resource, Vietnam Academy of Sciences and Technology, 18 Hoang Quoc Viet, Cau Giay, 100000 Hanoi, Vietnam.

<sup>2</sup>Graduate University of Science and Technology, Vietnam Academy of Sciences and Technology, 18 Hoang Quoc Viet, Cau Giay, 100000 Hanoi, Vietnam.

<sup>3</sup>College of Agriculture, Mindanao State University, Main Campus, Marawi City, 9700 Lanao del Sur, Philippines.

### AUTHORS' CONTRIBUTIONS

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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### ABSTRACT

Carrot (*Daucus carota* subsp. *Sativus*) is known as one of the most widely cultivated and widely consumed vegetables in the world due to its nutritional and economic values. During a survey of nematodes parasites carrots from Vietnam, six populations of three species of root-knot nematodes, namely *M. incognita*, *M. arenaria*, and *M. graminicola* were found. The species specific primers were confirmed before morphological studies. By combining the morphology and morphometry of the females, males, and juveniles, this study provided useful references for classification of *Meloidogyne* on carrots in the future. Quantitative morphological studies reveal profound changes corresponding with the generation of morphological disparity at high taxonomic diversity. Especially, this study provided the first morphological and morphometric information of *M. graminicola*, that is known as aquatic root-knot nematodes, on carrots.

**Keywords:** Root-knot nematodes; *Meloidogyne*; carrot; Vietnam; morphological; new records.

### 1. INTRODUCTION

Currently, carrot (*Daucus carota* subsp. *Sativus*) is one of the most widely cultivated and widely consumed vegetables in the world, thanks to its nutritional and economic values [1,2]. Many reports indicated that carrots, which were infested by plant-parasitic nematodes (PPN), are often stunted, underdeveloped, or even the high density of PPN could trigger the death of seedlings.

Root-knot nematodes (*Meloidogyne* spp.) parasitize almost every species of higher plant and are the

economically most important plant-parasitic nematodes that caused billions of dollars losses each year in the world [3,4,5]. At present, more than 104 species of this genus have been described [5,6,7,8,9]. *Meloidogyne* species are characterized primarily on morphological features of females, males and second-stage juveniles. The traditional identification of root-knot nematodes generally relies not only on close observation of three important taxonomic criteria, including the perineal patterns of females, male head shape, and stylet morphology [10]. Detailed diagnostic characters differentiating *Meloidogyne* species have been given by authors such as Eisenback

\*Corresponding author: Email: [tqphap@gmail.com](mailto:tqphap@gmail.com);

*et al.*, Eisenback, Hirschmann, Jepson, Taylor and Eisenback and Triantaphyllou [11,12,13,14,15,16]. Problematically, most descriptions are only based on a limited number of morphological features but so far, hardly any information pertaining to morphological and morphometrical [17]. The taxonomical work of root-knot nematodes has been limited in Vietnam. The features of Vietnamese populations of root-knot nematodes is available, the intraspecific variability of *Meloidogyne* species are still lacking [18].

This study provides the morphological and morphometric characteristics of Vietnamese populations of *Meloidogyne* spp. on carrots, as well as their host and distribution around the world. These information are really necessary for the sustainable management and development of carrots.

## 2. MATERIALS AND METHODOLOGY

**Sampling:** Two hundred and forty samples were collected from 22 fields from 4 provinces (Lam Dong, Hai Duong, Ha Noi, Hung Yen) in Vietnam. Each sample consisted of 1kg soil from upper 30cm soil layer, and 1 tuber with roots that were sampled at equal intervals in a grid pattern (10×10m) at each field.

**Nematode extraction:** The vermiform nematodes were extracted by the modified Baermann tray method [19]. Swollen nematodes were extracted directly from the galls under a stereomicroscope, using a scalpel and forceps [20].

**Nematode identification:** The nematode populations were checked in Multiplex-PCR before morphological study following Kiewnick et al. [21]. Nematodes were fixed in TAF (91ml H<sub>2</sub>O; 7ml Formalin 40%; 2ml Triethanolamine) and transferred to anhydrous glycerin to make permanent slides following Seinhorst (1959) [22]. Perineal patterns of *Meloidogyne* spp. were cut, cleaned, and mounted in glycerin following Hartman & Sasser [10]. Microphotographs were taken from permanent slides using a Carl Zeiss Axio Lab. A1 microscope equipped with a digital camera. The measurements were obtained by using the software ZEN lite. Classification was based on Eisenback, Jepson, Hewlett and Tarjan, Karssen, and Kazachenko & Mukhina [12,14,23,24,25]. The specific primers were used to confirm the morphological identification. The meaning for the indices *sensu de Man* [26,27] that were used: n = number of specimens; L = body length; V = % distance of vulva from anterior / body length; a = L / maximum body diameter; b = L / distance from anterior to pharyngo-intestinal junction; b' = L / distance from anterior to base of pharyngeal

glands; c = L / tail length; c' = tail length / tail diameter at anus or cloaca; Stylet = stylet length; DGO = distance from dorsal gland orifice to basal knob of stylet; VBD = diameter of body at vulva; ABD = diameter of body at anus.

## 3. RESULTS AND DISCUSSION

### 3.1 *Meloidogyne incognita* (Kofoid & White, 1919) Chitwood, 1949

See Fig. 1 and Table 1.

**Females:** Pear-shaped, no posterior terminal protuberance. Stylet, basal knobs rounded, offset. Perineal pattern oval to rounded, typically with high, squared, dorsal arch, striae usually wavy, lateral field absent or weakly demarcated by forked striae. **Male:** Labial region not offset, labial disc elevated, lateral lips usually absent. Stylet, basal knobs offset, rounded to transversely elongate. **J2:** Hemizonid anterior or adjacent to excretory pore, tail long hyaline region, rounded tail tip.

**Remarks:** The measurements of *M. incognita* on carrots in Vietnam are mostly in agreement with the original description of Chitwood, (1949). Few variations were observed such as: slightly larger body length and stylet length (350 - 411 vs 360 - 393 µm; 10 – 12.2 vs 10 µm, respectively), smaller a and c values (23 - 33 vs 29 - 33; 7.8 - 9 vs 8 - 9.4, respectively). Compared to the description of Nguyen & Nguyen (2000) the a value is larger (43 - 59 vs 39 - 48) and Stylet length of the males is smaller (20 - 23 vs 23 - 26 µm; 14 - 16,6 vs 15 - 15 µm). [18].

**Host:** following CABI (2018) [28].

**Distribution:** following CABI (2018) [28].

### 3.2 *Meloidogyne arenaria* (Neal, 1889) Chitwood, 1949

See Fig. 2, Table 2.

**Female:** Pear-shaped, no posterior terminal protuberance; stylet cone curved dorsally, gradually tapering to blunt tip anteriorly; shaft broad, cylindrical, gradually widening posteriorly; basal knobs rounded to teardrop-shaped, offset; perineal pattern variable, rounded to ovoid with fine to coarse striae; dorsal arch low, flattened with striae smooth or slightly wavy, continuous or broken, slightly bent towards tail tip at lateral line; generally forming shoulders on lateral portion of arch; dorsal and ventral striae often meeting at an angle at lateral lines; lateral field distinct, slightly irregular. **Male:** Labial region

**Table 1. Morphometrics of juveniles, females and males of *Meloidogyne incognita* from different populations. All measurements in µm (except for ratio) and in the format: mean ± S.D. (range)**

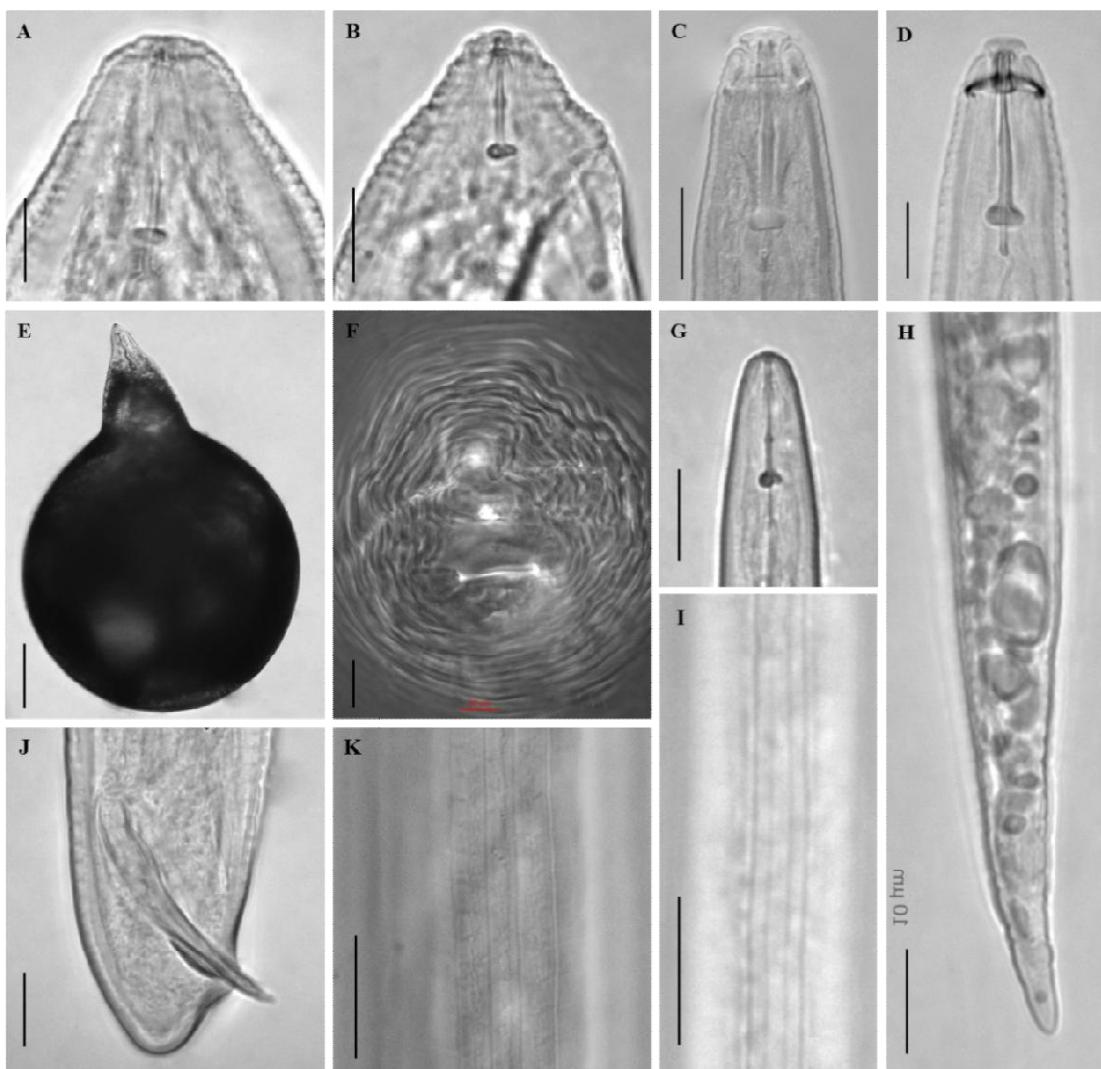
Measurements	<i>Meloidogyne incognita</i>											
	M4859 (Ha Noi)			M3656 (Hai Duong)			M4907 (Hung Yen)			M5269 (Lam Dong)		
	Female	Male	Juvenile	Female	Male	Juvenile	Female	Male	Juvenile	Female	Male	Juvenile
n	10	3	10	10	3	10	10	5	10	10	5	10
L	574 ± 30 (521 - 612)	1208 ± 121 (1081 - 1323)	385 ± 11.4 (379 - 409)	596 ± 84 (494 - 857)	1218 ± 194 (1037 - 1423)	402 ± 14.5 (380 - 412)	627.8 ± 37.3 (594 - 668)	1398 ± 64 (1309 - 1474)	370 ± 10.2 (350 - 386)	687 ± 61 (510 - 705)	1504 ± 94 (1430 - 1667)	392 ± 12.2 (362 - 409)
a	1.3 ± 0.2 (1 - 1.6)	49 ± 4.6 (44 - 53)	27.8 ± 1.5 (25 - 29.4)	2 ± 0.2 (1.7 - 2.2)	52 ± 6.4 (47 - 59)	31 ± 2.2 (26.4 - 33)	1.3 ± 0.3 (1.2 - 1.6)	52 ± 5.2 (43 - 57)	25.6 ± 1.8 (23 - 28.7)	1.6 ± 0.2 (1.4 - 2)	47 ± 2.8 (43 - 50)	29.3 ± 1.5 (26.3 - 31)
b	-	10.4 ± 0.7 (9.7 - 11)	5.4 ± 0.1 (5.2 - 5.6)	-	10.5 ± 1.2 (9.5 - 11.8)	5.7 ± 0.1 (5.4 - 6)	-	12.4 ± 1 (10.8 - 13.5)	5.2 ± 0.4 (5 - 5.4)	-	13.3 ± 1.2 (12.4 - 15.3)	5.5 ± 0.1 (5.4 - 5.7)
b'	-	4.4 ± 0.4 (4 - 4.6)	2 ± 0.1 (2 - 2.3)	-	4.5 ± 0.7 (3.8 - 5.3)	2.2 ± 0.1 (2 - 2.5)	-	5.2 ± 0.3 (4.8 - 5.5)	2 ± 0.2 (2 - 2.4)	-	6 ± 0.4 (5.3 - 6.5)	2 ± 0.1 (2 - 2.2)
c	-	100 ± 7.6 (91 - 106)	8.3 ± 0.4 (7.8 - 8.8)	-	112 ± 22 (98 - 138)	8.2 ± 0.4 (7.8 - 8.7)	-	117 ± 17.7 (92 - 131)	8 ± 0.2 (7.7 - 8.2)	-	129 ± 14 (112 - 144)	8.3 ± 0.4 (7.7 - 9)
c'	-	0.6 ± 0.2 (0.6 - 0.7)	4.5 ± 0.3 (4 - 5)	-	0.6 ± 0.1 (0.6 - 0.7)	4.8 ± 0.4 (4.3 - 5.6)	-	0.6 ± 0.2 (0.5 - 0.6)	4.8 ± 0.7 (3.8 - 6)	-	0.5 ± 0.1 (0.5 - 0.6)	4.6 ± 0.3 (4 - 5)
Stl	15 ± 0.5 (14.6 - 15)	20.4 ± 0.7 (20 - 21.2)	10.2 ± 0.4 (10 - 11)	15.5 ± 1 (15 - 16.2)	21.2 ± 0.5 (20.8 - 21.8)	11 ± 0.6 (10.4 - 12)	15.8 ± 0.4 (15.2 - 16.6)	22.6 ± 0.8 (21.5 - 23.3)	10.8 ± 0.4 (10 - 11.6)	14.6 ± 0.8 (14 - 16.4)	21.7 ± 0.3 (21.2 - 22)	10.5 ± 0.5 (10 - 11.2)
E.P. (%)	-	-	19.8 ± 0.3 (19 - 20.3)	-	-	20 ± 0.5 (19.3 - 21)	-	-	20.3 ± 1.2 (19 - 22.6)	-	-	20.2 ± 0.4 (19.7 - 21)
Lip width	4.4 ± 0.4 (3.8 - 4.8)	10.8 ± 0.5 (10.4 - 11.3)	-	5 ± 1 (3.8 - 6.5)	11 ± 0.2 (10.8 - 11)	-	4.7 ± 0.1 (4.5 - 4.8)	11.2 ± 0.5 (10.6 - 11.6)	-	4.7 ± 0.7 (3.7 - 5.5)	12 ± 0.6 (11.3 - 13)	
Lip height	2 ± 0.2 (1.7 - 2.3)	6 ± 0.3 (5.6 - 6)	-	3 ± 0.2 (2.8 - 3.2)	6 ± 0.3 (5.8 - 6.4)	-	2 ± 0.2 (1.7 - 2.1)	5.8 ± 0.7 (5 - 6.7)	-	2.6 ± 0.4 (2.2 - 3.2)	7 ± 0.3 (6.5 - 7.3)	
DGO	4 ± 0.8 (3 - 5)	3 ± 0.2 (2.8 - 3.2)	2.6 ± 0.3 (2.3 - 3)	3.6 ± 0.1 (3.6 - 3.8)	2.7 ± 0.2 (2.5 - 3)	3 ± 0.1 (2.7 - 3.2)	4.8 ± 0.3 (4.5 - 5)	3 ± 0.2 (2.7 - 3.3)	2.5 ± 0.2 (2.2 - 3)	5.5 ± 0.3 (5.2 - 6)	3 ± 0.3 (2.6 - 3.3)	2.8 ± 0.3 (2.5 - 3.2)
Median bulb width	33 ± 3.4 (28 - 38)	8.8 ± 1 (8.2 - 10)	7.3 ± 0.3 (6.8 - 7.8)	36 ± 4.7 (31 - 41)	9.2 ± 0.3 (9 - 9.6)	7.6 ± 0.6 (7 - 8.6)	35 ± 2.3 (33 - 38)	9.7 ± 1 (8.4 - 11)	7.4 ± 0.7 (6 - 8.3)	41 ± 4.2 (37 - 47)	10 ± 0.6 (9.4 - 11)	7.2 ± 0.3 (6.8 - 7.8)
Median bulb length	35 ± 3 (30 - 40)	16.2 ± 0.5 (15.8 - 16.7)	11 ± 0.6 (10 - 12)	31 ± 3.1 (30 - 38)	17.7 ± 0.7 (17 - 18.2)	11.6 ± 0.7 (10.8 - 13)	34 ± 3.7 (30 - 37)	16.7 ± 1 (15.8 - 18.2)	11.6 ± 0.5 (12 - 12.4)	51 ± 6.0 (43 - 58)	18.3 ± 1.7 (16.7 - 21)	11 ± 0.7 (10 - 12)
Median bulb valve length	12.2 ± 1.4 (10.5 - 14)	-	-	30 ± 0.7 (29.3 - 31)	-	-	11 ± 1 (10.5 - 12.4)	-	-	12.7 ± 0.4 (12.2 - 13)	-	-
Median bulb valve width	10.3 ± 1.3 (8 - 12)	-	-	27 ± 5.5 (20 - 31.5)	-	-	9.7 ± 2 (8 - 12)	-	-	10 ± 0.3 (9.7 - 10.3)	-	-
Distance from anterior end to median bulb	74 ± 11 (58 - 87)	76 ± 1.2 (75 - 77)	52 ± 1.5 (50 - 55)	79 ± 6.6 (72 - 85)	77 ± 4 (72 - 80)	52 ± 1.7 (50 - 55)	76 ± 12.2 (63 - 87)	80 ± 3.4 (75 - 83)	50 ± 2.6 (46 - 55)	86 ± 12.4 (69 - 97)	85 ± 2.3 (82 - 88)	53 ± 1.8 (50 - 55)
Distance from anterior end to nerve ring	-	98 ± 3.3 (94 - 100)	66 ± 1.6 (62 - 68)	-	98 ± 4.2 (94 - 103)	64 ± 2 (61 - 66)	-	100 ± 5.3 (92 - 106)	63 ± 2.6 (60 - 69)	-	103 ± 6.4 (92 - 108)	67 ± 2.5 (62 - 71)

Measurements	<i>Meloidogyne incognita</i>											
	M4859 (Ha Noi)			M3656 (Hai Duong)			M4907 (Hung Yen)			M5269 (Lam Dong)		
	Female	Male	Juvenile	Female	Male	Juvenile	Female	Male	Juvenile	Female	Male	Juvenile
Distance from anterior end to secretory-excretory pore	21 ± 4 (15.6 - 22)	128 ± 5.3 (122 - 131)	78 ± 2 (74 - 81)	17.8 ± 0.3 (17.5 - 18)	126 ± 5.3 (120 - 131)	79 ± 1.5 (78 - 83)	20.6 ± 5.8 (15.6 - 24)	126 ± 3.8 (121 - 130)	76 ± 3 (72 - 81)	18.5 ± 1 (18 - 20.2)	125 ± 6 (118 - 134)	79 ± 2.7 (74 - 83)
Distance from anterior end to pharyngo-intestinal junction	-	116 ± 4.4 (111 - 120)	74 ± 2 (69 - 78)	-	116 ± 5.7 (109 - 120)	77 ± 2.5 (73 - 81)	-	113 ± 7 (105 - 121)	76 ± 6.7 (67 - 92)	-	114 ± 3.2 (109 - 117)	75 ± 2.6 (69 - 77)
ES	-	278 ± 10 (267 - 287)	190 ± 10.5 (178 - 205)	-	271 ± 2 (269 - 273)	176 ± 8 (163 - 186)	-	267 ± 9.3 (252 - 277)	175 ± 13 (157 - 195)	-	257 ± 9.4 (248 - 272)	187 ± 8 (178 - 199)
OPL	-	162 ± 10 (150 - 168)	117 ± 10 (104 - 131)	-	155 ± 5.4 (149 - 160)	99 ± 7.3 (86 - 108)	-	153 ± 12.2 (140 - 160)	100 ± 11.7 (78 - 118)	-	143 ± 8.4 (133 - 155)	112 ± 8 (104 - 123)
W	454 ± 56 (349 - 529)	24.6 ± 1.7 (23 - 26.4)	14.3 ± 0.7 (13 - 15.2)	311 ± 95 (237 - 494)	23.4 ± 1 (22.2 - 24)	13 ± 1 (12 - 14.8)	423 ± 67 (349 - 479)	27 ± 2.2 (25 - 30)	14.7 ± 0.8 (13.5 - 16)	403 ± 73 (262 - 465)	32 ± 1.5 (29.3 - 33)	13.4 ± 0.5 (12.7 - 4.2)
ABD	-	19.5 ± 2 (18.2 - 21.7)	10.7 ± 0.7 (9.4 - 12)	-	17 ± 0.4 (16.5 - 17.2)	10 ± 0.6 (8.8 - 10.8)	-	21.3 ± 2 (18 - 23.3)	10 ± 1.2 (7.5 - 11.2)	-	22.7 ± 1 (22 - 24.7)	10.2 ± 0.6 (9.2 - 11)
T	-	12 ± 0.4 (11.8 - 12.5)	47 ± 0.8 (46 - 49)	-	11 ± 0.8 (10.3 - 11.8)	48 ± 2.6 (44 - 52)	-	12 ± 1.7 (10.3 - 14.3)	47 ± 2.3 (42 - 50)	-	11.8 ± 1.2 (10.4 - 13)	47 ± 1 (46 - 49)
H	-	-	11.2 ± 1 (9 - 12.6)	-	-	11.4 ± 1 (10.2 - 13)	-	-	10.8 ± 0.6 (10 - 11.7)	-	-	11.5 ± 1 (9 - 13)
Testis length	-	730 ± 88 (630 - 799)	-	-	724 ± 94 (651 - 830)	-	-	760 ± 70 (702 - 868)	-	-	851 ± 43 (783 - 896)	-
Spicule length	-	31 ± 1.2 (29.2 - 34)	-	-	31 ± 1.4 (29 - 31)	-	-	34 ± 1 (32 - 36)	-	-	33 ± 1 (30 - 36)	-
Spicule width	-	3.8 ± 0.3 (3.6 - 4)	-	-	4 ± 0.5 (3.6 - 4.7)	-	-	3.4 ± 0.3 (3.0 - 3.8)	-	-	3.9 ± 0.5 (3.0 - 4.4)	-
Gubernaculum length	-	8.6 ± 0.6 (8 - 9.2)	-	-	8 ± 0.4 (7.8 - 8.5)	-	-	8.8 ± 0.6 (8 - 9.3)	-	-	9 ± 0.6 (8.4 - 9.7)	-
Neck length	127 ± 29.5 (75 - 172)	-	-	259 ± 77.4 (139 - 369)	-	-	158 ± 19.7 (137 - 175)	-	-	208 ± 30 (158 - 245)	-	-
L/ neck length	4.8 ± 1 (3.6 - 7)	-	-	2.4 ± 0.6 (2 - 3.6)	-	-	4.0 ± 0.4 (3.5 - 4.3)	-	-	3.1 ± 0.5 (2.5 - 4)	-	-
Vulva slit length	18.8 ± 1 (17 - 20.5)	-	-	18.2 ± 1.8 (16 - 19.8)	-	-	18.5 ± 2 (16 - 19.7)	-	-	17.8 ± 2.2 (14.8 - 21)	-	-
Vulva-anus distance	17.2 ± 1 (16 - 18.7)	-	-	18.2 ± 0.2 (18 - 18.5)	-	-	18.2 ± 0.2 (18 - 18.5)	-	-	15 ± 2.4 (12.2 - 20)	-	-
Anus-tail tip distance	11.7 ± 1.2 (9.7 - 13.4)	-	-	14.3 ± 0.7 (13.5 - 15)	-	-	14 ± 0.6 (13.5 - 14.5)	-	-	12.3 ± 1.4 (10 - 14)	-	-
Distance between two plasmids	19.5 ± 4 (14 - 24.2)	-	-	21 ± 3.2 (17 - 23.5)	-	-	22.2 ± 1 (21.5 - 23.5)	-	-	20 ± 2 (17 - 22.3)	-	-
Distance from anterior end to secretory-excretory pore/Stylet	1.6 ± 0.3 (1.4 - 1.8)	-	-	1.8 ± 0.4 (1.3 - 2)	-	-	1.7 ± 0.5 (1.4 - 2)	-	-	1.3 ± 0.1 (1.2 - 1.4)	-	-

**Table 2. Morphometrics of *Meloidogyne arenaria* and *Meloidogyne graminicola* from different populations. All measurements in µm (except for ratio) and in the format: mean ± S.D. (range)**

Measurements	<i>Meloidogyne arenaria</i>			<i>Meloidogyne graminicola</i>		
	M4806 (Hai Duong)			M4856 (Ha Noi)		
	Female	Male	Juvenile	Female	Male	Juvenile
n	10	6	10	10	3	10
L	676 ± 41.2 (630 - 757)	1396 ± 110 (1227 - 1542)	407 ± 11.8 (388 - 428)	559 ± 61.4 (460 - 694)	1288 ± 394 (881 - 1668)	378 ± 11.4 (366 - 401)
a	2.2 ± 0.3 (1.6 - 2.6)	50 ± 3.7 (46 - 57)	28.7 ± 1.6 (25.7 - 31)	1.8 ± 0.3 (1.6 - 2.2)	45 ± 6 (39 - 51)	29.4 ± 0.7 (28.4 - 30)
b	-	12 ± 1 (10.6 - 13.2)	5.3 ± 0.2 (5 - 5.6)	-	12.2 ± 3.4 (8.7 - 15.4)	5.4 ± 0.2 (5 - 5.6)
b'	-	5 ± 0.5 (4.3 - 5.5)	2.3 ± 0.1 (2 - 2.4)	-	6.5 ± 1.8 (4.5 - 8)	2 ± 0.1 (2 - 2.2)
c	-	121 ± 9.7 (107 - 132)	8 ± 0.3 (7.7 - 8.3)	-	113 ± 21.6 (80 - 130)	6.5 ± 0.5 (5.4 - 7)
c'	-	0.5 ± 0.1 (0.5 - 0.6)	5 ± 0.3 (4.5 - 5.5)	-	0.6 ± 0.1 (0.5 - 0.7)	5 ± 0.3 (4.7 - 5.6)
Stylet	15.5 ± 2 (13.8 - 16.7)	22.7 ± 0.7 (21.5 - 23.6)	12.2 ± 0.5 (11.4 - 13)	12.8 ± 1.2 (11.4 - 13.2)	20 ± 2 (18.6 - 22.4)	11 ± 0.4 (10 - 12.2)
Lip width	5.5 ± 1 (4 - 6.2)	11.6 ± 0.3 (11.2 - 12)	-	5 ± 0.4 (4.5 - 5.5)	11.3 ± 0.3 (11 - 11.6)	-
Lip height	2.3 ± 0.7 (1.3 - 3)	6 ± 0.4 (5.3 - 6.5)	-	2.8 ± 0.3 (2.4 - 3.2)	5.7 ± 0.8 (5 - 6.5)	-
DGO	5 ± 0.7 (4 - 5.8)	4.8 ± 0.3 (4.4 - 5)	3 ± 0.2 (2.7 - 3.3)	3 ± 0.3 (2.8 - 3.3)	3 ± 0.3 (2.8 - 3.4)	2.8 ± 0.2 (2.5 - 3)
Median bulb width	30 ± 4.7 (27 - 39)	9.2 ± 0.6 (8.4 - 10)	7.5 ± 0.3 (7.2 - 8.3)	39 ± 5.3 (33 - 47)	10.8 ± 0.5 (10.2 - 11)	7.8 ± 0.4 (7.2 - 8.8)
Median bulb length	46 ± 5.8 (38 - 51)	17 ± 1.2 (15.8 - 18.4)	12 ± 0.7 (10.6 - 13)	50 ± 5.4 (43 - 58)	16.7 ± 0.8 (15.8 - 17.3)	11.5 ± 1 (9.8 - 13.2)
Median bulb valve width	8.5 ± 0.6 (7.8 - 9.6)	-	-	10 ± 0.5 (9.3 - 10.4)	-	-
Median bulb valve length	12.4 ± 2 (9.5 - 14.3)	-	-	13.6 ± 0.8 (12.7 - 14.6)	-	-
Distance from anterior end to median bulb	80 ± 4.8 (73 - 83)	85 ± 2 (83 - 88)	52 ± 2.5 (49 - 56)	97 ± 5.6 (90 - 105)	75 ± 3 (72 - 78)	49 ± 1.2 (48 - 51)
Distance from anterior end to nerve ring	-	101 ± 3.3 (97 - 106)	66 ± 2.3 (62 - 69)	-	91 ± 1.6 (90 - 93)	59 ± 1.8 (56 - 61)
Distance from anterior end to secretory-excretory pore	34 ± 1.6 (32 - 37)	128 ± 3 (124 - 131)	80 ± 2.3 (78 - 86)	32 ± 5.5 (26 - 42)	120 ± 9 (111 - 129)	75 ± 1.6 (72 - 77)

Measurements	<i>Meloidogyne arenaria</i>			<i>Meloidogyne graminicola</i>		
	M4806 (Hai Duong)			M4856 (Ha Noi)		
	Female	Male	Juvenile	Female	Male	Juvenile
Distance from anterior end to pharyngo-intestinal junction	-	116 ± 3 (111 - 120)	77 ± 2.4 (73.2 - 81.3)	-	105 ± 3 (102 - 108)	70 ± 2 (67 - 74)
Distance from anterior end to end of pharynx	-	277 ± 6.5 (268 - 286)	178 ± 6.3 (166 - 187)	-	197 ± 11.7 (185 - 209)	181 ± 6 (170 - 188)
Maximum body diameter	322 ± 61 (247 - 426)	28 ± 3 (24 - 32)	14.2 ± 0.7 (13.2 - 15)	276 ± 41 (227 - 353)	28.0 ± 5.2 (22.6 - 33)	13 ± 0.6 (12.2 - 13.5)
Neck length	241 ± 25.6 (193 - 270)	-	-	218 ± 71 (128 - 349)	-	-
Body length/ neck length	3 ± 0.5 (2.3 - 4)	-	-	2.8 ± 0.8 (1.4 - 4.4)	-	-
Vulva slit length	20.2 ± 2.4 (16 - 23.2)	-	-	20.8 ± 0.6 (20 - 21.8)	-	-
Vulva-anus distance	18.4 ± 2.3 (14.6 - 21)	-	-	17 ± 0.7 (16 - 17.8)	-	-
Anus-tail tip distance	15.5 ± 4 (12 - 23.2)	-	-	14.2 ± 1.4 (12.7 - 16)	-	-
Distance between two phasmids	17.2 ± 3 (13 - 23.8)	-	-	16.0 ± 1.5 (14.3 - 18.3)	-	-
Distance from anterior end to secretory-excretory pore/Stylet length	2.2 ± 0.3 (2 - 3)	-	-	2.3 ± 0.4 (1.9 - 2.9)	-	-
ABD	-	22.3 ± 1.7 (20 - 23.8)	10.4 ± 0.5 (9.5 - 11)	-	18.8 ± 3.6 (14.7 - 21)	9 ± 0.4 (8.3 - 9.6)
Tail length	-	11.6 ± 1 (10.7 - 13.5)	51 ± 1.3 (50 - 54)	-	11.3 ± 2.2 (10 - 13.8)	57 ± 1.6 (52 - 59)
Hyaline length	-	-	12.5 ± 0.7 (11.3 - 13.5)	-	-	16 ± 1 (14 - 18.5)
Testis	-	837 ± 54 (787 - 932)	-	-	722 ± 243 (462 - 943)	-
Spicule length	-	31 ± 1.2 (29.4 - 33)	-	-	34 ± 1 (33 - 34)	-
Spicule width	-	4 ± 0.2 (3.7 - 4.4)	-	-	3.7 ± 0.7 (3.2 - 4.5)	-
Gubernaculum length	-	8 ± 0.6 (7.4 - 9)	-	-	8.8 ± 0.8 (8 - 9.6)	-



**Fig. 1. *Meloidogyne incognita* (Kofoid & White, 1919) Chitwood, 1949**  
(LM ♀). A: Head region, B: Stylet, E: Entire body, F: Perriniel pattern. (LM ♂). C-D: Stylet, J: Tail region, K: Lateral field; (LM J2). G: Stylet, I: Lateral field, K: Tail region (Scalebar: A = 100  $\mu$ m, B-H = 10  $\mu$ m)

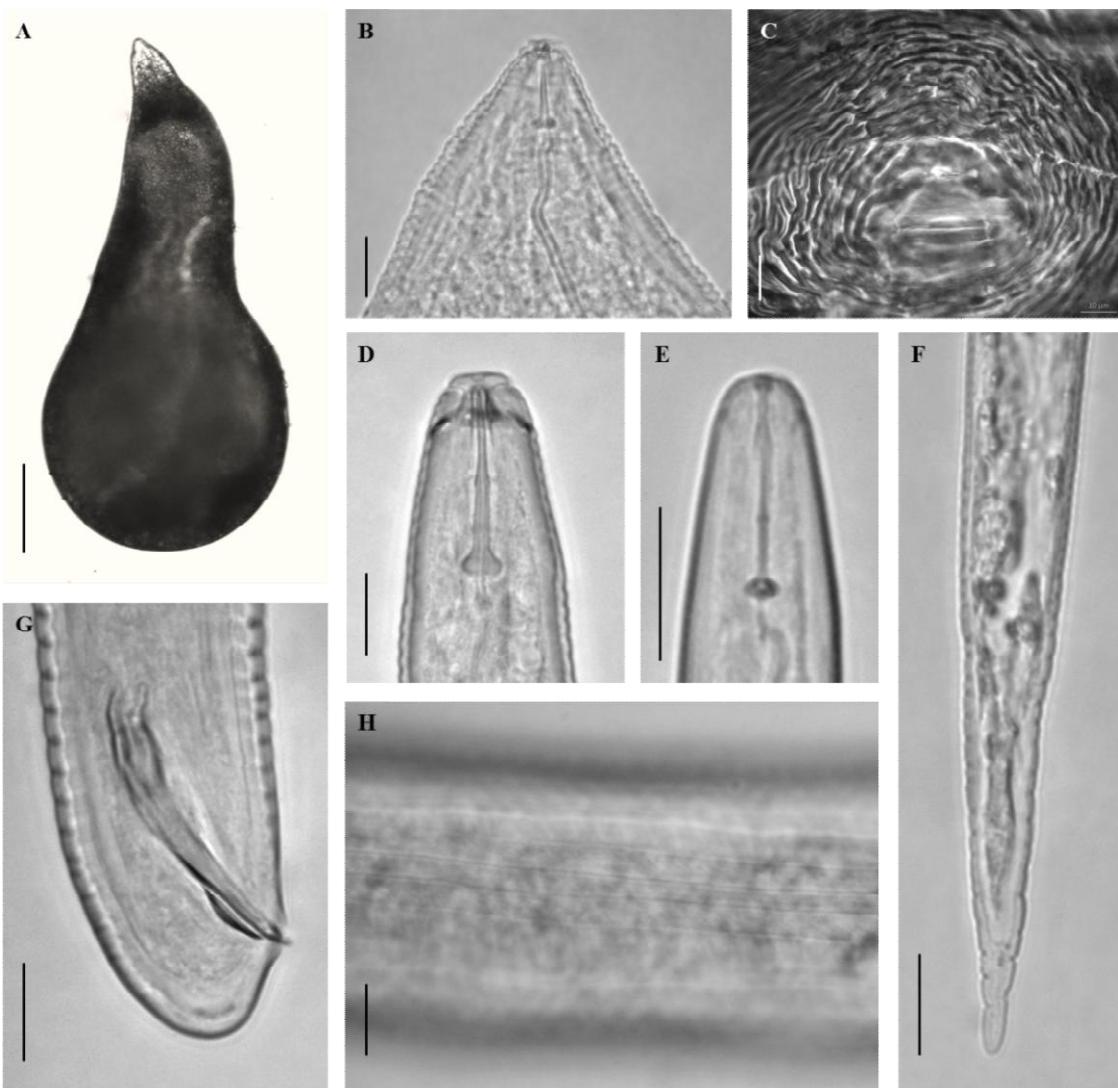
not offset, smooth, rarely with one or two incomplete annulations, labial disc more or less rounded, slightly raised above level of medial lips, lateral lips usually absent (remnants occasionally present); stylet basal knobs offset, angular or more amalgamated. J2: Hemizonid one to three annules anterior to excretory pore, tail rounded to pointed tail tip and indistinct long hyaline region.

**Remarks:** The morphology and morphometry of *M. arenaria* on carrots in Vietnam is mostly similar to the original description of Chitwood, (1949) except for the slightly smaller body length of females, males,

and juveniles (630 - 767 vs 510 - 1000  $\mu$ m; 1227 - 1542 vs 1270 - 2000  $\mu$ m; 388 - 428 vs 450 - 490  $\mu$ m, respectively), smaller b value in males and juveniles (10.6 - 13 vs 11 - 16, 5 - 5.6 vs 7.2 - 7.8, respectively), larger c value and stylet length in juveniles (7.7 - 8.3 vs 6 - 7.5, 11 - 13 vs 10  $\mu$ m, respectively) [29]. Compared to the description of Nguyen & Nguyen (2000), the stylet of this population is smaller (11 - 13 vs 13 - 14.5  $\mu$ m) [18].

**Host:** following CABI [28].

**Distribution:** following CABI [28].



**Fig. 2. *Meloidogyne arenaria* (Neal, 1889) Chitwood, 1949.**  
(LM ♀). A. Entire body; B. Stylet; C. Perineal pattern. (LM ♂). D: Stylet, G: Tail region, H: Lateral field. (LM J2). E: Stylet, F: Tail region (Scale bar: A = 100  $\mu$ m, B - H = 10  $\mu$ m)

### 3.3 *Meloidogyne graminicola* Golden & Birchfield, 1965

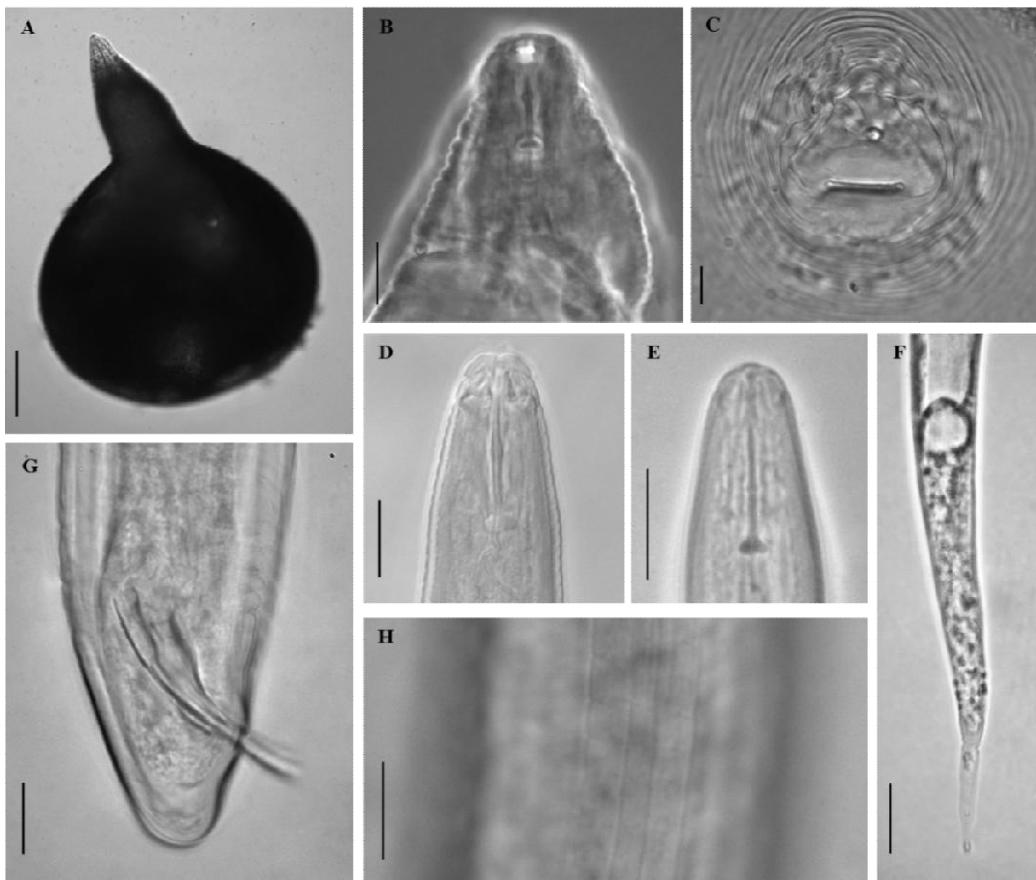
See Fig. 3, Table 2.

**Female:** Elongate, slight terminal protuberance present; stylet basal knobs ovoid, offset; perineal pattern rounded/oval, striae smooth, lateral field absent. **Male:** Labial region not offset, labial disc not elevated, lateral lips usually present, stylet basal knobs ovoid, offset. **J2:** Hemizonid anterior or adjacent to excretory pore, tail = 60–80mm, tail tip finely rounded.

**Remarks:** Few variation were observed from the population of *M. graminicola* on carrots in Vietnam compared to the description of Golden & Birchfield [30] such as: smaller body length in females (460 - 694 vs 445 - 765  $\mu$ m), larger stylet in females (11.4 – 13.2 vs 10 – 11.2  $\mu$ m); larger body length and stylet length in males (881 - 1668 vs 1020 - 1428  $\mu$ m; 18.6 – 22.4 vs 16.2 – 17.4  $\mu$ m, respectively); smaller body length and stylet in juveniles (366 - 401 vs 415 - 484  $\mu$ m).

**Host:** following CABI [28]. Carrot is a new host in this study.

**Distribution:** following CABI [28].



**Fig. 3. *Meloidogyne graminicola* Golden & Birchfield, 1965**  
(*LM ♀*). A: Entire body, B: Stylet, C: Perineal pattern. (*LM ♂*). D: Stylet, G: Tail region, H: Lateral field. (*LM J2*). E: Stylet, F: Tail region (Scalebar: A = 100  $\mu$ m; B-H = 10  $\mu$ m)

#### 4. CONCLUSION

This study provides the morphological and morphometric characterization of six populations from three species of the genus *Meloidogyne* namely *M. incognita*, *M. arenaria* and *M. graminicola*. This is the first time *M. graminicola* is reported to be present on carrots.

The morphology and morphometry of all the studied species were given and the classification was confirmed by PCR with specific primers following Kiewnick *et al.* (2013). The results of electrophoresis products of *M. incognita* was 300 bp long, *M. arenaria* species was 420 bp long, and *M. graminicola* was negative. Presently, the diversity of PPN on carrots is still poorly studied. The morphology and morphometry of the genus *Meloidogyne* in this study provide useful information for management and control of plant-parasitic nematodes on carrots. Furthermore, this study

provides good references for identification of the genus *Meloidogyne* in future.

*Meloidogyne graminicola* is previously known as aquatic root-knot nematode because their life cycle is stick to aquatic host plants. However, we have found these nematodes on carrots proving the ability of *M. graminicola* to survive on other host plants on dry land.

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#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Heinonen MI. Carotenoids and provitamin A activity of carrot (*Daucus carota* L.) cultivars. *Journal of Agricultural and Food Chemistry*. 1990;38:609-612.
2. Chen ZG., Guo XY, Wu T. A novel dehydration technique for carrot slices implementing ultrasound and vacuum drying methods. *Ultrasonics sonochemistry*. 2016;30:28-34.
3. Agrios GN. Plant pathology, San Diego, California, Elsevier Academic Press2005.
4. Jones JT, Haegeman A, Danchin EG, Gaur HS, Helder J, Jones MG, Kikuchi T, Manzanilla-López R, Palomares-Rius JE, Wesemael WM, Perry RN. Top 10 plantparasitic nematodes in molecular plant pathology. *Molecular Plant Pathology*. 2013;14: 946-961.
5. Perry, R. & Moens, M. 2013. Plant nematology, CABI, 542 pp.
6. Ahmed M, Van De Vossenberg BT, Cornelisse C, Karssen G. On the species status of the root-knot nematode *Meloidogyne ulmi* Palmisano & Ambrogioni, 2000 (Nematoda: Meloidogynidae). *ZooKeys*; 2013:1-27.
7. Humphreys-Pereira DA, Elling AA, Gómez M, Flores-Chaves L, Gómez-Alpízar L, Salazar L. *Meloidogyne lopezi* n. sp.(Nematoda: Meloidogynidae), a new root-knot nematode associated with coffee (*Coffea arabica* L.) in Costa Rica, its diagnosis and phylogenetic relationship with other coffee-parasitising *Meloidogyne* species. *Nematology*. 2014;16: 643-661.
8. Tao Y, Xu C, Yuan C, Wang H, Lin B, Zhuo K, Liao J. *Meloidogyne aberrans* sp. nov.(Nematoda: Meloidogynidae), a new root-knot nematode parasitizing kiwifruit China. *PloS one*.2017;12, e0182627.  
DOI:10.1371/journal.pone.0182627
9. Trinh QP, Le TML, Nguyen TD, Nguyen HT, Liebanas G, Nguyen TAD. *Meloidogyne daklakensis* n. sp. (Nematoda: Meloidogynidae), a new root-knot nematode associated with Robusta coffee (*Coffea canephora* Pierre ex A. Froehner) in the Western Highlands, Vietnam. *Journal of Helminthology*. 2018;1-13.  
DOI:10.1017/S0022149X18000202
10. Hartman KM, Sasser JN. Identification of *Meloidogyne* species on the basis of differential hosts test and perineal pattern morphology. In: Barker, K. R., Carter, C. C. & Sasser, J. N. (Eds.) An advanced treatise on *Meloidogyne*. North Carolina State University Graphics, Raleigh, North Carolina: A cooperative publication of the Department of Plant Pathology and the United States Agency for International Development. 1985;69-77.
11. Eisenback JD, Hirschmann H, Sasser JN, Triantaphyllou AC. A guide to the four most common species of root-knot nematodes [*Meloidogyne* species] with a pictorial key (No. REP-7253. CIMMYT.); 1981.
12. Eisenback JD. Detailed morphology and anatomy of second-stage juveniles, males, and females of the genus *Meloidogyne* (root-knot nematodes). In: Sasser, J. N. & Carter, C. C. (Eds.) An advanced treatise on *Meloidogyne*. V. 1. Biology and control. Volume 2. Methodology. North Carolina State University, Raleigh, NC (EUA): Department of Plant Pathology AID. 1985;47-78.
13. Hirschmann, H. The genus *Meloidogyne* and morphological characters differentiating its species. An advanced treatise on *Meloidogyne* Volume 1: Biology and control. North Carolina State University, Raleigh, NC (EUA): Department of Plant Pathology AID. 1985:79-93.
14. Jepson SB. Identification of Root-Knot Nematodes CABI. 1987;265.
15. Taylor AL. Identification and estimation of root-knot nematode species in mixed populations; 1987.
16. Eisenback, JD, Triantaphyllou HH. Root-knot nematodes: *Meloidogyne* species and races. Manual of agricultural nematology.1991;1:191-274.
17. Janssen T, Karssen G, Verhaeven M, Coyne D, Bert W. Mitochondrial coding genome analysis of tropical root-knot nematodes (*Meloidogyne*) supports haplotype based diagnostics and reveals evidence of recent reticulate evolution. *Scientific Reports*. 2016;6:22591.
18. Nguyen NC, Nguyen VT. Plant parasitic nematodes in Vietnam. Sci. & Tech. Publ, Hanoi, Vietnam. 2000;403.
19. Whitehead AG, Hemming JR. A comparison of some quantitative methods of extracting small vermiciform nematodes from soil. *Annales of Applied Biology*. 1965;55:25-38.
20. Perry RN, Moens M, Starr JL. Root-knot nematodes, CABI. 2009;488.
21. Kiewnick S, Holterman M, van den Elsen S, van Meegen H, Frey JE, Helder J. Comparison of two short DNA barcoding loci (COI and COII) and two longer ribosomal DNA genes (SSU & LSU rRNA) for specimen identification among quarantine root-knot nematodes (*Meloidogyne* spp.) and their close relatives. *European Journal of Plant Pathology*. 2014;140:97-110.

22. Seinhorst JW. A Rapid Method for the Transfer of Nematodes From Fixative To Anhydrous Glycerin. *Nematologica*. 1959;4:67-69.
23. Hewlett TE, Tarjan AC. Monographs: Synopsis of the Genus *Meloidogyne* Goeldi, 1887. *Nematropica*. 1983;13:79-102.
24. Karssen G. The plant parasitic nematode genus *Meloidogyne* Göldi, 1892 (Tylenchida) in Europe, Brill. 2002;160.
25. Kazachenko IP, Mukhina TI. Root-knot nematodes of genus *Meloidogyne* Goeldi (Tylenchida: Meloidognidae) of the world, Vladivostok, Russia, Institute of Biology and Soil Sciences. 2013:306 pp.
26. de Man JG. Onderzoeken over vrij in de aarde levende Nematoden. *Tijdschrift der Nederlandsche Dierkundige Vereeniging* Ver. 1876;2:78-196.
27. de Man J.G. Die Einheimischen, frei in der reinen Erde und im süssen Wasser lebenden Nematoden Vorlaufiger Bericht und descriptiv-systematischer Theil. *Tijdschrift der Nederlandsche Dierkundige Vereeniging*. 1880;5:1-104.
28. Cabi. *Invasive Species Compendium*. Wallingford, UK; 2018.
29. Chitwood BG. Root-knot nematodes, part I. A revision of the genus *Meloidogyne* Goeldi, 1887. *Proceedings of the Helminthological Society of Washington*. 1949;16(2):90-104.
30. Golden AM, Birchfield W. *Meloidogyne graminicola* (Heteroderidae), a new species of root-knot nematode from grasses. *Proc. Helminthol. Soc. Wash.* 1965;32:228-231.

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