

A Home-made Hybrid System for the Simultaneous Determination of Ergotamine, Dipyrone and Caffeine in Pharmaceutical Preparations

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Um novo sistema híbrido para detecção simultânea de cafeína (CAF), dipirone (DIP) e de ergotamina (ERG) foi desenvolvido para teste de preparações farmacêuticas. ERG foi determinada por um sinal fluorimétrico, enquanto para CAF e DIP foi utilizado o método PLS-1 para resolução de dados espectrométricos de UV-V. A curva de calibração da ERG foi linear na faixa de 2.5 - 10 mg L⁻¹. O conjunto de calibrações constuiu-se de 18 soluções com 0.5 a 4 mg L⁻¹ de CAF, 5 a 20 mg L⁻¹ de DIP e de 2.5 e 10 mg L⁻¹ de ERG. Não foi necessária a preparação de amostras anterior à análise. Foram realizados estudos de recuperação com amostras reais obtendo-se resultados altamente satisfatórios

A novel home-made hybrid detection system was designed for simultaneous determination of caffeine (CAF), dipyrone (DIP) and ergotamine (ERG) in pharmaceutical preparations. Thus, ERG was determined by a fluorimetric signal and PLS-1 method for resolving DIP and CAF UV-V spectral data was used. The calibration curve for ERG was linear over the range 2.5 - 10 mg L⁻¹. The calibration set consisted of 18 mixtures with 0.5 to 4 mg L⁻¹ for CAF, 5 to 20 mg L⁻¹ for DIP and 2.5 and 10 mg L⁻¹ for ERG. Sample preparation prior to analysis was not required. A recovery study with the real samples was carried out and the obtained results were highly satisfactory.

Keywords: hybrid system, ergotamine, dipyrone, caffeine, simultaneous determination, PLS-1

Introduction

Ergotamine tartrate [(5S)-12'-hydroxy-2'-methyl-3',6',18-trioxo-5-benzyl ergotaman (2R, 3R)-tartrate], (ERG); dipyrone [disodium salt of [(2,3-dihydro-1,5-dimethyl-3-oxo-2-phenyl-1H-pyrazol-4-yl) methylamino] methanesulfonic acid], (DIP) and caffeine [1,3,7-trimethylxanthine], (CAF) are usually joined in some pharmaceutical preparations and in several formulations. ERG is a sedative used to relieve migraine, DIP possesses analgesic and antipyretic properties. The effect of these drugs on relief pains can be enhanced when CAF is present.

Chromatographic techniques are widely applied for determining these analytes in pharmaceutical preparations, but these methods present the disadvantages of relative high cost, time consumption and use large volumes of toxic organic solvents.^{1, 2}

Methodologies such as FIA and SIA with spectrometric, fluorimetric and biamprometric detection has been reported for this purpose.³⁻⁹ Nevertheless, the simultaneous determination of ERG, DIP and CAF was not found in the literature.

The spectrophotometric and fluorimetric detection have been used in the analysis of pharmaceuticals and biomedical samples owing to lower cost of instruments, the simplicity of procedures, precision and accuracy, etc. Moreover, the simultaneous determination of multicomponents in complex samples by using these classical techniques is not possible due to overlapping spectra.

On the other hand, the application of chemometric techniques as PCR (Principal component regression), PLS1 and PLS2 (Partial least squares) to spectrophotometric and fluorimetric data are being used for the analysis of complex mixtures.¹⁰⁻¹⁶ In this paper a new analytical method with home-made hybrid detection system for simultaneous determination of CAF, DIP and ERG in pharmaceutical preparations is proposed.

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These kind of samples have such lower concentration of ERG than the other two analytes, so it is necessary to develop an original hybrid instrument that combines features from both fluorimetric and spectrophotometric detections.¹⁷ By this way it is possible the fluorimetric determination of ERG. CAF and DIP are determined by using UV-V absorption spectral data and PLS-1.

Experimental

Apparatus

The home-made hybrid detection system was built by using a Hewlett-Packard 8452A diode array spectrophotometer controlled by a computer for UV-V spectra acquisition. Moreover, four phototransistors, an operational amplifier and a digital voltmeter Aka M-3850 to register the fluorescence signal were used.

By the other hand, home made cell holder was designed and a 10mm path length Hellma quartz cell was employed.

Reagents

Analytical grade reagents and ultra pure water Milli-Q quality ($18.3 \text{ m}\Omega \text{ cm}^{-1}$) were always used. Pure ergotamine, caffeine and dipyrone were obtained from Saporitti.

The stock solutions of ergotamine, caffeine and dipyrone were prepared by weighing 0.0025 g, 0.0025 g and 0.0253 g respectively, 2.5 mL of ethanol were added and then made up to 50 mL with water. For dissolving these solutions an ultrasonic bath was used for seven minutes.

The standard solutions were prepared by appropriate dilution of the stock solutions and making up to 10.0 mL with $10^{-4} \text{ mol L}^{-1}$ NaOH.

Calibration and validation sets

The calibration set was obtained by applying a random experimental design. A training set of 18 standard solutions were prepared. The concentration ranges were 5 to 20 mg L^{-1} and 0.5 to 4 mg L^{-1} for DIP and CAF, respectively. The ERG of 2.5 and 10 mg L^{-1} was added to the mixtures. Table 1 shows the composition of the calibration set. On the other hand, a six mixtures validation set was prepared in order to validate the chemometric models.

Sample preparation

The pharmaceutical samples were purchased from local drugstores. The trade names were Migra Dioxadol

Table 1. Concentration level of DIP, CAF and ERG for calibration set

Mixtures	Dipyrone/ (mg L^{-1})	Caffeine/ (mg L^{-1})	Ergotamine/ (mg L^{-1})
M1	5.00	0.50	2.50
M2	20.00	0.50	2.50
M3	5.00	4.00	2.50
M4	20.00	4.00	2.50
M5	12.50	2.25	2.50
M6	12.50	2.25	2.50
M7	5.00	0.50	10.00
M8	20.00	0.50	10.00
M9	5.00	4.00	10.00
M10	20.00	4.00	10.00
M11	12.50	2.25	10.00
M12	12.50	2.25	10.00
M13	5.00	0.50	2.50
M14	20.00	0.50	10.00
M15	5.00	4.00	2.50
M16	20.00	4.00	10.00
M17	12.50	2.25	2.50
M18	12.50	2.25	10.00

(BAGO), Tetralgil (CRAVERI) and Migral 500 (MONTPELLIER).

Twenty tablets of each one were weighed to calculate the average tablet weight. They were finely powdered in a mortar and homogenised. In order to obtain a solution of approximately 4.0 mg L^{-1} of ERG, a suitable amount of the powder was accurately weighed and dissolved in an appropriate amount of ethanol (5% v/v), assisted by an ultrasonic bath. Then, making up to an adequate volume with $10^{-4} \text{ mol L}^{-1}$ NaOH and finally it was filtered (Solution A).

In order to obtain approximately 12 mg L^{-1} of DIP and 2.5 mg L^{-1} of CAF a suitable dilution of Solution A was carried out with $10^{-4} \text{ mol L}^{-1}$ NaOH.

Results and Discussion

The relationship of DIP, CAF and ERG in commercial pharmaceutical products is 500:100:1 mg/tablet so all the analytical studies were carried out by considering this proportion.

In order to select the optimum medium for the simultaneous spectrophotometric determination of the three analytes, different solvents and pH (HCl, NaOH, NH_3 and 20% v/v ethanol) were tested. Every time, CAF and DIP presented higher absorbances than ERG, which gave hardly absorption (Figure 1). As the CAF and DIP spectra overlapped the low ERG signal prevented the simultaneous determination of the three analytes.

The same study was carried out by using fluorimetric technique. CAF and DIP were not fluorescent in the tested

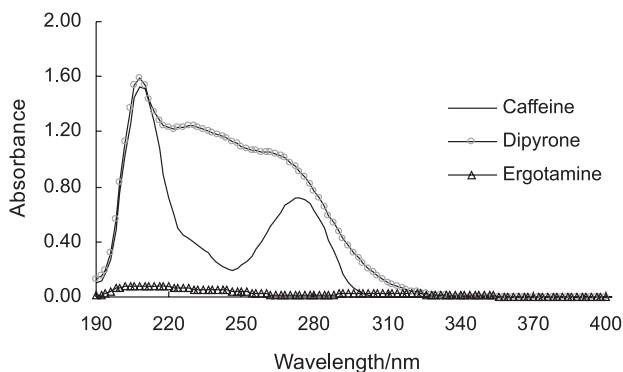


Figure 1. CAF (10 mg L⁻¹), DIP (50 mg L⁻¹) and ERG (1 mg L⁻¹) spectra.

experimental conditions, while ERG has an important signal. The best results for ERG were obtained by using 10⁻⁴ mol L⁻¹ NaOH solution.

Bearing in mind these considerations and taking into account that only ERG is fluorescent, it was decided to design an hybrid system providing simultaneously UV-Vis spectra to determine CAF and DIP and fluorimetric signals for ERG determination.

Hybrid system

The hybrid system was designed in our laboratory (Figure 2).

For this purpose, a polypropylene tube (length: 11.5 cm, inner diameter: 2.0 cm) was placed instead of the cell holder of the spectrophotometer. The tube is black to prevent radiation loss and it has a square hole (15×15 mm) to permit the insertion of a cuvette. Moreover, two phototransistors were placed on each side of the tube, near the sidewalls of the cell (Figure 2 b).

The radiation emitted by the deuterium lamp of the spectrophotometer went on the rail tube and through the cell. The UV-Vis spectra were recorded by the spectrophotometer while the fluorescence signal was detected by the phototransistors at 90 degree. The output of the latter is amplified with an operational amplifier, and the amplified output is read with a digital voltmeter connected to the device through panel jacks.

Analytical parameters

The calibration curve of ERG was $F = (0.105 \pm 0.002)X + (0.196 \pm 0.013)$ where **F** is the fluorescent signal and **X** is the concentration of ERG in mg L⁻¹, with a correlation coefficient of 0.995. The linear range was 2.50-10.00 mg L⁻¹.

The reproducibility (RSD%) was 1.97%, calculated from ten replicates containing 2.50 mg L⁻¹.

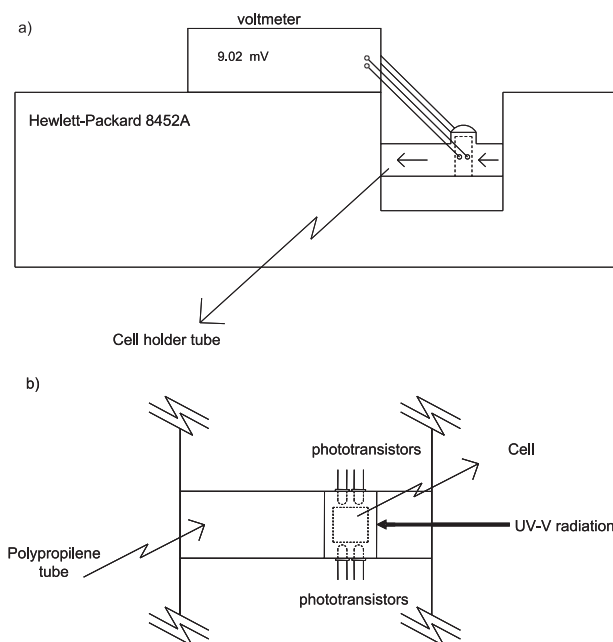


Figure 2. a) Hybrid system; b) Cell holder (top view).

PLS-1 method for resolving DIP and CAF spectral data, was used. The number of factors was selected by using the leave-one-out cross-validation method. The optimum number of factors was obtained following the statistics criterion of Haaland and Thomas.¹⁸ The appropriated wavelengths regions were selected by checking different wavelength ranges. In Table 2 were shown the different figures of merit: the spectral range, the number of factors for each analyte, the relative error of prediction (REP%), selectivity (SEL) and sensitivity (SEN).¹⁹

Table 2. PLS calibration figures

Figures	Dipyrone	Caffeine
Spectral region / (nm)	270-330	210-290
Optimal number of factors	3	3
REP (%)	6.8	19.3
SEN	0.01	0.145
SEL	0.13	0.45

Applications to real samples

The proposed method was applied for the determination of ERG, DIP and CAF in pharmaceutical formulations. In order to validate the obtained results, a recovery study was carried out. Table 3 shows these results and as it can be seen, there are very good recovery values, all in the recommended range by Pharmacopoeia.²⁰

Table 3. Determination of ergotamine, dipyrone and caffeine in pharmaceutical preparations (tablets)

Sample	Amount								
	Labeled mg/tablet			Found mg/tablet \pm SD ^a			Recoveries % ^b		
	ERG	DIP	CAF	ERG	DIP	CAF	ERG	DIP	CAF
Migradioxadol	1	500	100	1.08 \pm 0.02	512 \pm 30	101 \pm 3	108.0	102.4	101
Tetralgin	1	500	100	0.98 \pm 0.05	499 \pm 14	101 \pm 2	98.0	99.8	101
Migral 500	1	400	100	1.02 \pm 0.04	431 \pm 6	100 \pm 4	102.0	107.7	100

^a. Standard deviations (n=5), ^b. recoveries are based on the amount labeled.

Conclusions

The most important goal of this work was the possibility to carry out a simultaneous determination of the three analytes by using different techniques as fluorimetry for ERG and spectrophotometry assisted by chemometric tool for DIP and CAF.

For this purpose a home made hybrid system was developed. This system is simple, cheap, easy to built and with a fast data collection.

The results obtained applying this system showed a good accuracy and reproducibility for these kind of samples. So, this proposed method is a good alternative to those of the Pharmacopoeia.

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References

- Ramos-Martos, N.; Aguirre-Gómez, F.; Molina-Díaz, A.; Capitan-Vallvey, L. F.; *J. AOAC Intern.* **2001**, *84*, 676.
- Berzas Nevado, J. J.; Villaseñor Llerena, M. J.; Contento Salcedo, A. M.; Aguas Nuevo, E.; *J. Pharm. Biomed. Anal.* **2005**, *38*, 52.
- Medeiros, E. P.; Castro, S. L.; Formiga, F. O. M.; Santos, S. R. B.; Araujo, M. C. U.; Nascimento, B. V.; *Microchem. J.* **2004**, *78*, 91.
- Legnerová, Z.; Sklenáková, H.; Solich, P.; *Talanta* **2002**, *58*, 1151.
- Morelli, B.; *J. Pharm. Biomed. Anal.* **2003**, *33*, 423.
- Singh, D. K.; Sahu A.; *Anal. Biochem.* **2006**, *349*, 176.
- Abbaspour, A.; Mirzajani, R.; *J. Pharm. Biomed. Anal.* **2005**, *38*, 420.
- Moreira, A. B.; Dias, I. L. T.; Neto, G. O.; Zagatto E. A. G.; Ferreira M. C.; Kubota L. T.; *Talanta* **2005**, *67*, 65.
- Hooper, W. D.; Sutherland, J. M.; Eadie, M. J.; Tyrer, J. H.; *Anal. Chim. Acta* **1974**, *69*, 11.
- Vigneau, E.; Devaux, M.; Qannari, M.; Robert, P.; *J. Chemom.* **1997**, *11*, 239.
- Massart, D.; Vandeginste, B.; Buydens, L.; De Jong, S.; Lewi, P.; Smeyers-Verbeke, J.; *Handbook of Chemometrics and Qualimetrics*, Elsevier: Amsterdam 1998.
- Lavine, B. K.; *Anal. Chem.* **2000**, *72*, 91R.
- Haaland, D. M.; *Appl. Spectrosc.* **2000**, *54*, 246.
- Feam, T.; *J. Near Infrared Spectrosc.* **2001**, *9*, 229.
- Brereton, R.; *Chemometrics, Data Analysis for the Laboratory and Chemical Plant*, Wiley: Chichester, 2003.
- Geladi, P.; *Spectrochim. Acta, Part B*; **2003**, *58*, 767.
- Valcárcel, M.; Cárdenas, M. S.; *Automatización y Miniaturización en Química Analítica* Springer-Verlag Ibérica, 2000.
- Haaland, M.; Thomas, E.V.; *Anal. Chem.* **1988**, *60*, 1193.
- Lorber, A.; Faber, K.; Kowalski, B.R.; *Anal. Chem.* **1997**, *69*, 1620.
- British Pharmacopoeia*, H. M. Stationery Office: London, 1998, vol. 2.

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