

Fish propagule dispersal and patch cohesiveness

1 **PROPAGULE DISPERSAL AND LARVAL PATCH COHESIVENESS IN A**
2 **MEDITERRANEAN COASTAL FISH**

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13 **ABSTRACT**

14 The assessment of fish dispersal has a fundamental importance for both conservation and
15 management of fisheries resources as it can provide crucial information for the establishment of
16 effective marine protected areas (MPAs) and networks of MPAs. In this study we investigated
17 otolith elemental composition in early life stages of the saddled sea bream *Obalda melanura*
18 (Linnaeus, 1758) (Perciformes: Sparidae) in order to obtain information on its propagule (egg -
19 larva) dispersal in the south-western Mediterranean Sea. Specifically, considering late stage larvae
20 and early post-settlers, we investigated: (1) number of potential natal sources; (2) propagule
21 dispersal distances and (3) larval patch cohesiveness during the last phase of the larval life. Seven
22 natal sources were found to replenish, with different proportions, almost all the sampling sites along
23 a stretch of coastline of ~180 km. This outcome suggests that propagule dispersal can take place at
24 least up to ~90 km. We show also that different larval patches can merge in the pelagic environment
25 after having travelled separately for some days. This information can provide important insights for

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26 the understanding of fish dispersal process and support the correct establishment of spatially
27 explicit conservation strategies such as MPAs and MPA networks in the south-western
28 Mediterranean sea.

29

30 **KEYWORDS:** propagule dispersal, natal origins, patch cohesiveness, saddled sea bream,
31 Mediterranean Sea

32 INTRODUCTION

33

34 Dispersal is the process by which organisms expand actively or passively the space where they live
35 affecting patterns of connectivity (i.e. the link between sub-populations through the exchange of
36 individuals), meta-population dynamics and ultimately species persistence (Shima & Swearer 2010)
37 (Cote et al. 2010). The study of dispersal patterns is thus of paramount importance for marine
38 biodiversity conservation and fisheries management as it provides fundamental information for the
39 establishment of effective marine protected areas (MPAs) and networks of MPAs (Grüss et al. 2011)
40 (Green et al. 2014). In many coastal fish with a bipartite life cycle (i.e. composed by a larval and a
41 juvenile/adult phase), the life stages after settlement (i.e. when the pelagic larval phase ends with
42 the metamorphosis into juvenile) are relatively sedentary, so that species dispersal potential is
43 mostly determined by the propagule (i.e. egg and larva) phase (Leis 2015). Despite the huge effort
44 being currently accomplished, the empirical measure of propagule dispersal is still extremely
45 challenging for fish ecologists due to several issues, such as the difficulty to mark small planktonic
46 larvae and/or to recapture them (given the high mortality rates) (Barbee & Swearer 2007, Fontes et
47 al. 2009).

48 In recent years, a number of studies have been conducted using natural environmental markers for
49 investigating fish natal origins and evaluate species dispersal capacity (Leis et al. 2011, Calò et al.
50 2013 for reviews). Otoliths are particularly useful for their unique feature of recording the chemical
51 characteristics of the surrounding environment experienced by fishes at all the life stages (Barbee &
52 Swearer 2007). Otoliths grow by a continuous deposition of layers of calcium carbonate, mainly
53 aragonite, in a protein matrix (Campana 1999). During the deposition, trace-elements from the
54 ambient water can substitute for calcium remaining permanently in the matrix (Campana 1999). The
55 physical and chemical characteristics of the environment (primarily temperature, salinity and trace-
56 elements concentration) influence the incorporation rates of some specific elements (e.g. Sr and Ba)

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57 into the otolith matrix (Campana 1999, Walther & Thorrold 2006, Barnett-Johnson et al. 2008,
58 Muhlfeld et al. 2012). Variations in otolith elemental composition can be used to discriminate larval
59 groups that experienced different environmental conditions and obtain information on important
60 early life history traits such as the number of potential natal sources, larval dispersal distances and
61 pathways (Hamilton et al. 2008, Fontes et al. 2009, Di Franco et al. 2012, Shima & Swearer 2010,
62 Miller et al. 2014, Shima et al. 2015, but see also Berumen et al. 2010). The analysis of the
63 chemical composition of otolith cores (i.e. the inner portion of otoliths that forms during the
64 embryogenesis) allows to assess the number of potential sources of propagules that supply different
65 settlement sites along a stretch of coastline (Ruttenberg et al. 2008). From this information it is
66 possible to infer the spatial scale over which propagule dispersal takes place, considering the
67 distance between settlement sites replenished by the same natal sources (*sensu* Di Franco et al.
68 2012). The analysis of the chemical profile from the otolith edge toward its core provides
69 information on the different environmental conditions faced by a larva backwards in time (Sandin et
70 al. 2005). The comparison of otolith chemical profiles of larvae coming from the same patch (i.e. an
71 aggregation of larvae) can be used to investigate the consistency of the patch during the pelagic
72 phase (i.e. cohesiveness), that is, if larvae travelled together to the sampling site or experienced
73 different dispersal pathways, that would result in different individual otolith chemical profiles
74 (Shima & Swearer 2009, Ben-Tzvi et al. 2012). The knowledge of the ontogenetic cohesiveness of a
75 larval patch can provide important insights on post-settlement demographic dynamics (Shima et al.
76 2015) as different larval trajectories, potentially shaped by a variable dispersal environment, can
77 have crucial implications on the post-settlement persistence of the larval patch (Shima & Swearer
78 2009, Shima & Swearer 2010, Shima et al. 2015).

79 In this study we investigated geochemical signatures in the otoliths of early life stages of the
80 saddled sea bream *Obalda melanura* (Linnaeus, 1758) (Perciformes: Sparidae) in order to obtain
81 information on its propagule dispersal characteristics in the south-western Mediterranean Sea. This

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82 fish was chosen as a model species as it is widely distributed and generally abundant in
83 Mediterranean coastal ecosystems (Bauchot & Hureau 1986, García-Charton et al. 2004) and for its
84 relatively high commercial value for artisanal and recreational fisheries (Harmelin-Vivien et al.
85 1995, Lloret et al. 2008, Félix-Hackradt et al. 2014). Specifically, considering early post-settlers and
86 late stage larvae of *O. melanura*, we investigated: (1) the number of potential natal sources
87 replenishing our study area; (2) propagule dispersal distances and (3) the cohesiveness of larval
88 patches during the last phase of the larval life. This information, can provide important insights for
89 the understanding of fish dispersal process and support the correct establishment of spatially
90 explicit conservation strategies such as MPAs and MPA networks.

91 MATERIALS AND METHODS

92

93 *Study area and sampling design*

94 The study was conducted along the south-east Spanish coast (SW Mediterranean Sea). This sector is
95 particularly relevant from a conservation perspective hosting an important biodiversity hotspot
96 (Calvin Calvo et al. 1999).

97 With the aim to investigate the number of potential natal sources and propagule dispersal distances
98 of *O. melanura* in the study area, early post-settlement individuals were sampled in July 2013 after
99 the peak of settlement (Calò et al. submitted). Post-settlers were collected in 17 sites (stretches of 1
100 km of coastline), scattered within 3 zones (stretches of 30 km of coastline), as follows, from North
101 to South: southern Alicante province (SAP, 6 sites), central Murcia region (CMR, 5 sites) and
102 western Murcia region (WMR, 6 sites) (Fig 1). SAP and CMR host 2 of the most effective MPAs of
103 the Western Mediterranean Sea, Tabarca MPA and Cabo de Palos e Islas Hormigas MPA (Di Franco
104 et al. 2014a) (Fig 1). In each site 11 to 15 post-settlers (i.e. individuals of 1.1-1.8 cm in total length)
105 were collected during snorkeling using a hand net. Specimens were sampled in shallow water
106 habitats (i.e. <2m of depth) characterized by shadowed overhangs and steep rocks, as these are
107 considered to be the main habitat requirements of *O. melanura* early post-settlers (Bussotti &
108 Guidetti 2011, Félix-Hackradt et al. 2014) for evidence from our study area). All the specimens
109 were firstly euthanized, by immersing them in a sea water solution with few drops of 96% alcohol
110 for minimizing their suffering (Leary et al. 2013) and, after cessation of opercular movements,
111 preserved in 70% ethanol.

112 Larval patch cohesiveness was investigated considering late stage larvae (pre-settlers) of *O.*
113 *melanura*. Specimens were caught using light-traps that were deployed during the period of larval
114 supply to the coast in July 2013 (Calò et al. submitted). Six light-traps were deployed in 2 sites (S1
115 in the SAP zone and S2 in the WMR zone, Fig. 1) and placed on an imaginary line parallel to the

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116 coast using a buoy moored at 20 –30 m depth, depending on how close the sampling site was to the
117 coast (distance ranging 0.3 –0.8 km). Three light-trap samples from site S1 (here named T1, T2 and
118 T3), sampled during the same night, and one light-trap sample from site S2 (T4) contained from 8
119 to 11 pre-settlers each, and thus were processed in the laboratory. All the other light-trap samples
120 were not included in the successive analyses, because they contained only 1-2 individuals and thus
121 it was impossible to investigate any inter-individual variability in dispersal, in order to assess patch
122 cohesiveness.

123

124 *Otolith preparation and analysis*

125 In the laboratory, the right sagittal otolith was removed from each pre-settler and post-settler,
126 cleaned of soft tissue using plastic pins and rinsed with ultra-pure water ($18.2\text{M}\Omega\text{cm}^{-1}$). Then,
127 otoliths were mounted sulcus side up onto a glass slide using Crystalbond™ thermoplastic adhesive.
128 Otoliths were polished with 3 μm and 1 μm Imperial lapping film through the nucleus. After
129 polishing, otoliths were rinsed with an ultrasonic bath for 10 minutes in ultra-pure water in order to
130 remove surface contamination.

131 Elemental analysis was performed using a Thermo Elemental X7 inductively coupled plasma mass
132 spectrometer (ICP-MS) coupled to a New-Wave Research UP213 with aperture imaging laser
133 ablation (LA) system. Otolith sections were viewed remotely on a computer screen where the area
134 for ablation was selected. The laser was focused on the sample surface and fired through the
135 microscope objective lens. Helium gas was flushed into the ablation cell to reduce the deposition of
136 ablated aerosols and to improve signal intensities. The ablated aerosol was then mixed with Argon
137 before entering the ICP torch.

138 Otoliths of post-settlers were used for obtaining information on the number of potential natal origins
139 and larval dispersal distances of *O. melanura*. Post-settler otoliths were analysed in 2 different
140 regions: the core and the edge. The core elemental composition was investigated to obtain

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141 information about natal origins and was analysed through three vertical pits with a diameter of 30
142 μm (approximate size of the core) and 10 μm deep each. The otolith edge was investigated to
143 quantify recent elemental incorporation (i.e. material laid down just before capture) that was later
144 tested for site discrimination (see *Data analysis*) and to account for within otolith variability (Di
145 Franco et al. 2012, Di Franco et al. 2014b). Edge portion was analysed by three horizontal pits
146 using the same laser spot dimensions as specified before. Each spot run consisted of 62 s
147 acquisition: 25 s blank to correct for background which was subtracted from each sample, 2 second
148 of pre-ablation to remove surface contamination (laser at 30% power), 10 s ablation and 25 s for
149 washout. Otoliths were placed in the ablation chamber in groups of 6, randomly selected from the
150 17 sampling sites, to prevent sample batch bias.

151 Otoliths of pre-settlers were analysed for the investigation of larval patch cohesiveness.
152 Specifically, we compared the variability of otolith chemical profiles among individuals coming
153 from the same light-trap sample (here considered as a larval patch). Pre-settlers otoliths were
154 analysed through a series of 7 laser spots, following the major axis of the sagitta, and positioning
155 each spot on a different daily growth ring from the edge of the otolith (corresponding to the otolith
156 ring laid down during the sampling day) to the seventh last growth ring. In this way, for each pre-
157 settler, we obtained the chemical profile of the last 7 days of larval life. We chose not to analyse the
158 first days after hatching to avoid biased interpretation of chemical composition variability. In fact,
159 the consideration of the earliest larval stages could have added an additional source of variation
160 (ontogenetic) hiding or confounding the spatial variability of otolith chemical signatures, on which
161 the patch cohesiveness analysis is based (see *Data analysis* for the complete rationale of the
162 statistical method). The mean pelagic larval duration of *O. melanura* individuals sampled by light
163 traps is ~ 11 days (Calò et al. submitted), thus we reasonably excluded from the analysis the first 3-4
164 days after hatching, corresponding to the yolk sack phase of the species (Antolović et al. 2010).
165 Spot size was set to 15 μm in order to fit it with the daily rings width. Each spot run consisted of 42

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166 s acquisition: 10 s blank to correct for background, 2 second of pre-ablation, 10 s ablation and 20 s
167 for washout.

168 For both post-settlers' and pre-settlers' otoliths, instrumental precision was maintained by analysing
169 solid glass standard material from the National Institute of Standards and Technology (NIST 610
170 and NIST 612) every 6 samples, carrying out a linear interpolation between the 2 consecutive sets
171 of standards. Calcium was used as internal standard to take into account variation in ablation and
172 aerosol efficiency. All the 7 elements analysed (^7Li , ^{24}Mg , ^{45}Mn , ^{66}Zn , ^{88}Sr , ^{138}Ba , ^{208}Pb) were
173 expressed as ratios relative to ^{44}Ca . Detection limits were calculated from the concentration of
174 analyte yielding a signal equivalent to $3\times$ the standard deviation of the blank signal for each of the
175 elements (Tab. 1). Recoded values of Li, Mn, Zn and Pb were consistently below the detection
176 limits and thus were excluded from the analysis.

177

178 *Data analysis*

179 Otolith elemental concentration data were converted to molar concentrations and $\log(x+1)$
180 transformed.

181 The number of post-settler potential natal origins was investigated analysing otolith core elemental
182 concentrations (as a proxy for identifying the existence of single or multiple areas of origin (Di
183 Franco et al. 2012) for a total of 230 individuals. No Mn:Ca spike, generally considered as an
184 indicator of the core location (Brophy et al. 2004, Ruttenberg et al. 2005), was recorded in almost
185 all the otolith cores analysed. For this reason the centroid of the three core pits for each otolith was
186 considered for the subsequent analysis, as it has been done in other species where no Mn:Ca spike
187 seems to not be an effective “core localizer” (Papetti et al. 2013, Guidetti et al. 2013). The
188 SIMPROF ('clustsig' package, R software) test was carried out on the output of a cluster analysis
189 (based on Euclidean distance) to determine which clusters were significantly different at the 5%
190 level. Similarity percentage (SIMPER, 'vegan' package, R software) was used to assess which

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191 Elemental/Ca ratios mainly contributed to the differences among the significant clusters identified
192 by the SIMPROF. Because homogeneity in otolith chemical composition may simply reflect
193 environmental similarity, we used permutational multivariate analysis of variance (PERMANOVA)
194 to test for differences between the 17 sampling sites by analysing the otolith edge of post-settlers
195 (i.e. post-settlement portion laid down just before capture) (Di Franco et al. 2012). 'Site' (Si) was
196 treated as a random factor (17 levels), 'Otolith' (Ot) was treated as a random factor nested in (Si)
197 (11-15 levels). There were three replicate ablations for each otolith (total n=690). Once different
198 natal origins were identified (see Results), we tested for possible differences in zone and site
199 replenishment for each identified natal source with a univariate two-way PERMANOVA using natal
200 origin number as a variable. 'Zone' (Zn) was treated as a random factor (3 levels); 'Site' (Si) was
201 treated as a random factor (5 to 6 levels) nested in (Zn). Finally, a Mantel test (based on 10^4
202 permutations, 'ade4' package, R software) was performed to assess if closer sites were more likely
203 to be replenished by the same natal origin. Thus, we measure the correlation between the abundance
204 of post-settlers coming from the same natal origin and the geographic distance between pairs of
205 sites. The distance matrix of abundances was created considering the 17 sites as observations and
206 the major natal sources identified as variables. Thus, each site was associated to n values of
207 abundance, corresponding to the number of post-settlers coming from each of the n major natal
208 sources.

209 Larval patch cohesiveness was investigated analysing the within patch (i.e. light-trap) variability of
210 pre-settler otolith chemical profiles. In particular we compared the chemical composition of otoliths
211 day by day, grouping together chemical data from chronologically homologous rings of different
212 otoliths (i.e. rings corresponding to the same day of life for all the fish). Based on the proviso that
213 the chemical composition of otoliths is similar for individuals that experienced the same
214 environmental conditions, we assumed that for individuals caught in the same light-trap, the
215 variability of the chemical composition would have increased moving from the first ring analysed

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216 (i.e. the ring laid down immediately before the sampling) toward the last ring analysed, in the case
217 in which fish travelled different pathways (i.e. starting from different sites and then reaching the
218 same light-trap). Otherwise, if pre-settlers travelled together to the site where we collected them
219 (light-trap), a similar variability would have been found between edge and the previous rings (days
220 of larval life) of the otolith. In order to perform the analysis, otolith chemical data were normalised
221 and grouped by ring analysed (i.e. day of life) from the last (1, the edge of the otolith) to the seventh
222 last (7) and considering the different light-traps samples separately. Then, individual deviations
223 from the centroids for each otolith ring, across individuals coming from the same light-trap, were
224 calculated using PERMDISP. Centroid distances were finally analysed through a PERMANOVA in
225 which 'light-trap' (LT) was treated as a random factor with 4 levels, 'specimen' (SP) was treated as
226 random factor with 8-11 levels, nested within LT and 'otolith ring' (OR) was treated as a fixed factor
227 with 7 levels, crossed to SP. This model, considering OR as a factor crossed to SP, was adopted as a
228 solution to repeated measures (the repeated measure effect is removed by fitting it as a factor). For
229 each light-trap, a means plot of distance from centroid per each otolith ring was used to show the
230 variation in fingerprints variability passing from day 1 to day 7. PERMANOVA analyses were run
231 using Primer 6 PERMANOVA+ software package. All the other analyses were run using R software
232 (R core development team).

233

234 RESULTS

235

236 SIMPROF test segregated 7 statistically different clusters, i.e. core chemical 'finger-prints'
237 potentially corresponding to seven different post-settler natal sources (Fig. 2). The five major natal
238 sources (A, C, D, E and G) accounted together for the 92.3% (212 individuals) of all the post-
239 settlers with single contribution ranging from 12.2% to 32.3%. The remaining natal sources, B and
240 F, consisted, respectively of 5 (2.1%) and 13 (5.6%) individuals. SIMPER analysis indicated that
241 Mg:Ca ratio contributed, on average, for ~50% to the total dissimilarity in pairwise comparisons
242 among five major natal sources identified, while Ba:Ca and Sr:Ca contributed with similar
243 proportion to the remaining 50% of dissimilarity. Cluster 'G' (that included 1/3 of all the post-
244 settlers) was characterized by a 2 times higher concentration of Mg:Ca than the remaining clusters.
245 Cluster 'A' and 'D' showed lower values of Ba:Ca than C, E and G. Sr:Ca ratios resulted different in
246 all the 5 groups (Fig. 3).

247 Post-settlement replenishment from the natal origins identified was found to be statistically different
248 among the 3 zones (PERMANOVA $p < 0.01$), with the southernmost zone (WMR) highly different
249 from the northernmost zone (SAP) (pairwise comparison $p < 0.01$) and weakly different from the
250 central zone (CMR) (pairwise comparison $p < 0.05$). No significant differences were found between
251 SAP and CMR. No differences were detected at the scale of site. The 5 major natal sources
252 replenished, with different proportions, almost all the sampling sites. Post-settlers from source 'G'
253 were mainly found in the WMR zone, with similar, lower, abundances recorded in the other two
254 zones. Source 'A' similarly replenished north (SAP) and central (CMR) sites. Source 'C' was mainly
255 composed by individuals from sites of SAP (north of the study area). Individuals from source 'E'
256 were mainly sampled in CMR, while a comparable number of post-settlers coming from the source
257 'D' was found in each site (Fig. 1 and 4). Regarding the last two natal sources, individuals from 'F'
258 were found in WMR, CMR and the southernmost site of SAP. Source 'B' replenished five sites (1, 2,

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259 5, 13 and 14) with one individual. The abundance of post-settlers sharing the same natal origin and
260 the geographic distance between sites were significantly positively correlated (Mantel test, $r=0.38$
261 and $p=0.0001$), this indicating that geographically closer sites had, on average, higher abundances
262 of post-settlers coming from the same natal origin, than distant sites. Focusing on natal sources G, A
263 and C (the three major natal sources clustered, representing more than 2/3 of all post-settlers), the
264 two higher values of abundance were recorded in pairs of adjacent and less distant sites (17-16 for
265 source 'G', 7-8 for source 'A' and 3-4 for source 'C' (Fig. 4). The chemical composition of the
266 juvenile portion of post-settlers was significantly different among the sampling sites
267 (PERMANOVA, $p<0.01$). Significant differences among otoliths were also found ($p<0.01$)
268 suggesting within-site differences among individuals.

269 Regarding the analysis of pre-settlers' patch cohesiveness, a significant difference between otolith
270 rings (days of life) was recorded (PERMANOVA, $p<0.01$), with an increasing variability moving
271 from day 1 to day 7 (Fig. 5) for all the 4 light-trap samples. Pairwise comparisons between days
272 indicated that statistically significant increments in chemical fingerprint variability were not
273 simultaneous across light-traps samples (i.e. did not happen between the same pair of days across
274 light-traps), resulting in a significant interaction Lt x OR ($p<0.05$): for pre-settlers from light-trap
275 T4 the first significant increment was recorded between the 2nd and the 3rd ring, in T1 between the
276 4th and the 5th ring, while in T2 and T3 a significant increment was recorded between the 1st and the
277 2nd ring (Fig. 5).

278 DISCUSSION

279

280 For the most of marine fish, the pelagic phase before settlement represents a fundamental period of
281 dispersion shaping connectivity patterns between populations (Burgess et al. 2014). Knowledge on
282 species dispersal characteristics (e.g. number of natal origins, dispersal distances) is crucial for
283 understanding fish populations dynamics and connectivity and ultimately helping the design of
284 effective marine protection strategies (Green et al. 2014). In this study we analysed otolith chemical
285 composition of early life stages of *O. melanura* in order to provide information on the dispersal
286 characteristics of its propagule pelagic phase in the south-western Mediterranean Sea.

287 The analysis of post-settler core elemental composition indicate the presence of multiple sources of
288 propagules, replenishing several sampling sites with different proportions along the south-eastern
289 coast of Spain. In recent works carried out in the Eastern Atlantic and in the Mediterranean Sea
290 similar results were recorded, with a high diversification of natal sources in early post-settler
291 individuals sampled in coastal sites (Fontes et al. 2009, Di Franco et al. 2012). The significant
292 spatial variability in the chemical composition of the juvenile portion (i.e. otolith edge) of post-
293 settler otoliths highlight the ability of the method to discriminate spatially distinct chemical
294 signatures, allowing us to reasonably draw inferences on the core portion. The segregation of
295 different core chemical signatures is very likely to be related to spatial differences in chemical-
296 physical characteristics of the water masses where the spawning took place. Different chemical
297 fingerprints in otolith cores could be associated also to a temporal variability in water masses
298 characteristics associated to temporally distinct spawning events. In 2013 the study area was
299 replenished by simultaneous spawning events that took place in a temporal window of 2 weeks
300 (Calò et al. submitted), so we can reasonably assume that the different chemical fingerprints
301 recorded are unlikely to be a consequence of a temporal variation.

302 The methodology we used does not allow to geographically locate the natal origins identified,

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303 impeding to have the exact range and the frequency distribution (i.e. kernel) of larval dispersal
304 distances. On the other hand, we can have an estimation of the spatial scale over which dispersal
305 can take place in the area. All the sites were supplied by more than one natal source. Two sources
306 replenished the extreme sites of the study area, spaced ~180 km from each other. This information
307 allows us to infer that the most conservative value of maximum dispersal distance is ~90 km (i.e.
308 half of the study area). In fact, supposing the case in which two of the natal sources identified were
309 adjacent and in the centre of the study area, propagules would have travelled at least 90 km. In all
310 the other cases, the dispersal distance would be greater. This estimate of propagule dispersal
311 distance is in accordance with the results of other studies conducted in the Mediterranean Sea on
312 other sparid species, using otolith natural tags (Di Franco et al. 2012, Di Franco et al. submitted)
313 and, in general, in the range of propagule dispersal suggested for other temperate fish species
314 (Anadón et al. 2013).

315 Even though it is not possible to locate the natal sources, the results suggest that the pattern of larval
316 supply to the sampling sites has a certain spatial dependence. Post-settlers from the same natal
317 origin were more likely to be found in closer sites with the highest abundances recorded in pairs of
318 adjacent sites and lower values both in northernmost and southernmost sites. This output was
319 consistent among almost all the major natal origins identified. Moreover, although the major natal
320 origins supply almost all the sampling sites, significant differences in post-settlers replenishment
321 was recorded at the scale of zone suggesting that post-settler demographic composition can change
322 at the scale of 50-100 km. A similar result was found by (Fontes et al. 2009) analysing otolith core
323 composition of the temperate wrasse *Coris julis* in the Azores. The spatial difference in natal source
324 composition between the northernmost and the southernmost zones of the study is also in
325 accordance with the difference in pelagic larval duration of *O. melanura* between the same zones
326 (Calò et al. submitted). This concordant results could indicate that the geographic origin of
327 propagules and early life history traits (e.g. the duration of the larval phase) could have a common

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328 source of variability or be non-independent. Patterns of post-settlers distribution along the coast are
329 likely to be a consequence of both pre- and/or post-settlement processes as well as both physical
330 (i.e. current dynamics during the larval dispersal phase) or biological ones (e.g. behaviour, inter-
331 and intra-specific competition for food and space, mortality). From this perspective the different
332 natal origin composition found between the southernmost zone (WMR) and the other two zones of
333 the study area (SAP and CMR) could be a consequence of the mesoscale oceanographic pattern that
334 characterizes the region. In a recent work, Rossi et al (2014) pointed out that the separation between
335 the Alboran sea and the rest of the Mediterranean sea, generally thought to have place along the
336 Almeria-Oran front, could actually have its Spanish coastal extreme around Cartagena, that in our
337 study separate the CMR zone from the WMR zone. The presence of this oceanographic boundary,
338 already suggested to be a biogeographic barrier in the Western Mediterranean sea (Pérez-Ruzafa
339 2010), could reduce the dispersal of propagules from the WMR zone toward the northernmost zones
340 and vice versa. However, it is very hard to draw conclusions on the forces that determined the
341 spatially dependent outcomes recorded in this study, as no further information are available to
342 elucidate them, especially considering the lack of information on fish spawning grounds. From this
343 perspective, further investigation are to be conducted using other methodologies that would permit
344 to directly track the movements of propagules from their source to the settlement site. These
345 methods (e.g. otolith artificial tagging or genetic parentage analysis), although undoubtedly
346 powerful, have been rarely or never applied so far in the Mediterranean context given the lack of
347 data about spawning ground locations and the huge sampling effort needed for obtaining
348 representative results (Calò et al. 2013).

349 The analysis of larval patch cohesiveness suggests that groups of larvae can merge in the pelagic
350 environment after having travelled separately for some days. In recent works a similar larval patch
351 formation was evidenced in a small triplefin fish of New Zeland reef, using otolith chemical
352 analysis (Shima & Swearer 2009, Shima et al. 2015). However, cases of protracted larval patch

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353 cohesiveness were also documented, with larvae experiencing the same dispersal pathway during
354 the entire pelagic phase (Ben-Tzvi et al. 2012, Bernardi et al. 2012). In our study, in two light-traps,
355 an increasing otolith chemical variability was recorded between the sampling day and the day
356 immediately before, suggesting that, during the sampling night two or more discrete groups of
357 larvae were attracted toward the same trap, after having travelled different dispersal pathways. In
358 other two light-traps, a stable chemical composition between the sampling day and the previous
359 ones was observed, suggesting that the aggregation of distinct patches can take place in different
360 moments of the larval phase. It is difficult to establish if this aggregation tendency is a result of
361 larval behaviour or particular oceanic features (e.g. gyres) that can mix together patches previously
362 separated. Kiparissis et al (2008) found evidences of aggregation behaviour in post-settlement
363 individual of *O. melanura*, differently from other Mediterranean sparid fishes (Harmelin-Vivien et
364 al. 1995). In particular, post-settlers arrive in small shoals and then actively aggregate in larger
365 shoals with increasing fish size after various aggregation steps (Kiparissis et al. 2008). From this
366 perspective, the same aggregation behaviour could characterize the larval phase, determining a mix
367 of distinct larval patches, potentially coming from different natal sources, during the pelagic phase.
368 This patch mixing tendency could be an adaptive strategy that would guarantee a higher genetic
369 diversity, and thus higher survival probability, when facing the environmental uncertainty of the
370 settlement habitat and the high mortality rates associated to it (Nanninga & Berumen 2014, Shima
371 et al. 2015). In the light of these considerations, it resulted evident that post-settler natal sources
372 diversity recorded here in each sampling site could be a consequence of multiple aggregation steps
373 that can occur both during the propagule dispersal phase and early after the settlement process and
374 would contribute together to demographic heterogeneity, fundamental in driving meta-population
375 dynamics and the persistence of species (Kritzer & Sale 2004).

376 To conclude, with the present study we provided insights on the propagule dispersal traits of a
377 temperate coastal fish. This is the first study aimed to estimate dispersal distances in the south-

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378 western Mediterranean Sea and, to our knowledge, the first to provide information on larval patch
379 dispersal cohesiveness during the pelagic phase of a Mediterranean coastal fish. Information on
380 propagule dispersal patterns are fundamental for the correct planning of MPAs and networks
381 (Planes et al. 2009, Gaines et al. 2010). From this perspective, our results are particularly relevant
382 considering the environmental value of the region and the strong effort that is being accomplished
383 to build an effective network of MPAs in the sector of Mediterranean sea considered. Certainly, the
384 study of one species is not sufficient for drawing conclusions on the effective MPAs spatial
385 arrangement, but it represents an important piece of knowledge, that together with information on
386 other species and/or using different approaches, can provide the fundamental background for
387 correct marine spatial conservation actions.

388

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390

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396 BIBLIOGRAPHY

397

- 398 Anadón JD, Mancha-Cisneros M, Best B, Gerber LR (2013) Habitat-specific larval dispersal and
399 marine connectivity: implications for spatial conservation planning. *Ecosphere* 4:1–15
- 400 Antolović N, Kozul V, Safner R, Glavić N, Bolotin J (2010) Embryonic and yolk-sac larval
401 development of saddled bream, *Oblada melanura* (Sparidae). *Cybium* 34:381–386
- 402 Barbee N, Swearer S (2007) Characterizing natal source population signatures in the diadromous
403 fish *Galaxias maculatus*, using embryonic otolith chemistry. *Mar Ecol Prog Ser* 343:273–282
- 404 Barnett-Johnson R, Pearson TE, Ramos FC, Grimes CB, MacFarlane RB (2008) Tracking natal
405 origins of salmon using isotopes, otoliths, and landscape geology. *Limnol Oceanogr* 53:1633–
406 1642
- 407 Bauchot ML, Hureau JC (1986) Sparidae. In: *Fishes of the North-eastern Atlantic and the*
408 *Mediterranean*. Vol. II.p 883–907
- 409 Ben-Tzvi O, Abelson A, Gaines SD, Bernardi G, Beldade R, Sheehy MS, Paradis GL, Kiflawi M
410 (2012) Evidence for cohesive dispersal in the sea. *PLoS One* 7:e42672
- 411 Bernardi G, Beldade R, Holbrook SJ, Schmitt RJ (2012) Full-sibs in cohorts of newly settled coral
412 reef fishes. *PLoS One* 7:e44953
- 413 Berumen ML, Walsh HJ, Raventos N, Planes S, Jones GP, Starczak V, Thorrold SR (2010) Otolith
414 geochemistry does not reflect dispersal history of clownfish larvae. *Coral Reefs* 29:883–891
- 415 Brophy D, Jeffries TE, Danilowicz BS (2004) Elevated manganese concentrations at the cores of
416 clupeid otoliths: possible environmental, physiological, or structural origins. *Mar Biol*
417 144:779–786
- 418 Burgess SC, Nickols KJ, Griesemer CD, Barnett LK, Dedrick AG, Satterthwaite E V, Yamane L,
419 Morgan SG, White JW, Botsford LW (2014) Beyond connectivity: how empirical methods can
420 quantify population persistence to improve marine protected-area design. *Ecol Appl* 24:257–
421 270
- 422 Bussotti S, Guidetti P (2011) Timing and habitat preferences for settlement of juvenile fishes in the
423 Marine Protected Area of Torre Guaceto (south-eastern Italy, Adriatic Sea). *Ital J Zool* 78:243–
424 254
- 425 Calò A, Félix-Hackradt FC, Garcia J, Hackradt CW, Rocklin D, Treviño Otón J, García Charton JA
426 (2013) A review of methods to assess connectivity and dispersal between fish populations in
427 the Mediterranean Sea. *Adv Oceanogr Limnol* 4:150–175
- 428 Calvin Calvo JC, Franco Navarra I, Marín Atucha A, Belmonte Ríos A, Ruiz Fernandez JM (1999)
429 El litoral sumergido de la region de murcia. *Cartografía bionómica y valores ambientales*.
- 430 Campana SE (1999) Chemistry and composition of fish otoliths: pathways, mechanisms and

Fish propagule dispersal and patch cohesiveness

- 431 applications. *Mar Ecol Prog Ser* 188:263–297
- 432 Cote J, Clobert J, Brodin T, Fogarty S, Sih a (2010) Personality-dependent dispersal:
433 characterization, ontogeny and consequences for spatially structured populations. *Philos Trans*
434 *R Soc Lond B Biol Sci* 365:4065–76
- 435 Di Franco A, Bodilis P, Piante C, Di Carlo G, Thiriet P, Francour P, Guidetti P (2014a) Fishermen
436 engagement in Mediterranean marine protected areas. A key element to the success of artisanal
437 fisheries management. MedPAN North Project. WWF France. 135 pages
- 438 Di Franco A, Bulleri F, Pennetta A, De Benedetto G, Clarke KR, Guidetti P (2014b) Within-otolith
439 variability in chemical fingerprints: implications for sampling designs and possible
440 environmental interpretation. *PLoS One* 9:e101701
- 441 Di Franco, Gillanders BM, De Benedetto G, Pennetta A, De Leo GA, Guidetti P (2012) Dispersal
442 Patterns of Coastal Fish: Implications for Designing Networks of Marine Protected Areas.
443 *PLoS One* 7
- 444 Félix-Hackradt FC, Hackradt CW, Treviño-Otón J, Pérez-Ruzafa A, García-Charton JA (2014)
445 Habitat use and ontogenetic shifts of fish life stages at rocky reefs in South-western
446 Mediterranean Sea. *J Sea Res* 88:67–77
- 447 Fontes J, Caselle J, Sheehy M, Santos R, Warner R (2009) Natal signatures of juvenile *Coris julis* in
448 the Azores: investigating connectivity scenarios in an oceanic archipelago. *Mar Ecol Prog Ser*
449 387:51–59
- 450 Gaines SD, White C, Carr MH, Palumbi SR (2010) Designing marine reserve networks for both
451 conservation and fisheries management. *Proc Natl Acad Sci U S A* 107:18286–93
- 452 García-Charton JA, Pérez-Ruzafa Á, Sánchez-Jerez P, Bayle-Sempere JT, Reñones O, Moreno D
453 (2004) Multi-scale spatial heterogeneity, habitat structure, and the effect of marine reserves on
454 Western Mediterranean rocky reef fish assemblages. *Mar Biol* 144:161–182
- 455 Green AL, Maypa AP, Almany GR, Rhodes KL, Weeks R, Abesamis R a., Gleason MG, Mumby PJ,
456 White AT (2014) Larval dispersal and movement patterns of coral reef fishes, and implications
457 for marine reserve network design. *Biol Rev*: doi: 10.1111/brv.12155
- 458 Grüss A, Kaplan DM, Guénette S, Roberts CM, Botsford LW (2011) Consequences of adult and
459 juvenile movement for marine protected areas. *Biol Conserv* 144:692–702
- 460 Guidetti P, Petrillo M, Benedetto G De, Albertelli G (2013) The use of otolith microchemistry to
461 investigate spawning patterns of European anchovy: A case study in the eastern Ligurian Sea
462 (NW Mediterranean). *Fish Res* 139:1–4
- 463 Hamilton SL, Regetz J, Warner RR (2008) Post-settlement survival linked to larval life in a marine
464 fish. *Proc Natl Acad Sci U S A* 105:1561–6
- 465 Harmelin-Vivien M, Harmelin J-G, Leboulleux V (1995) Microhabitat requirements for settlement
466 of juvenile sparid fishes on Mediterranean rocky shores. *Hydrobiologia* 300/301:309–320

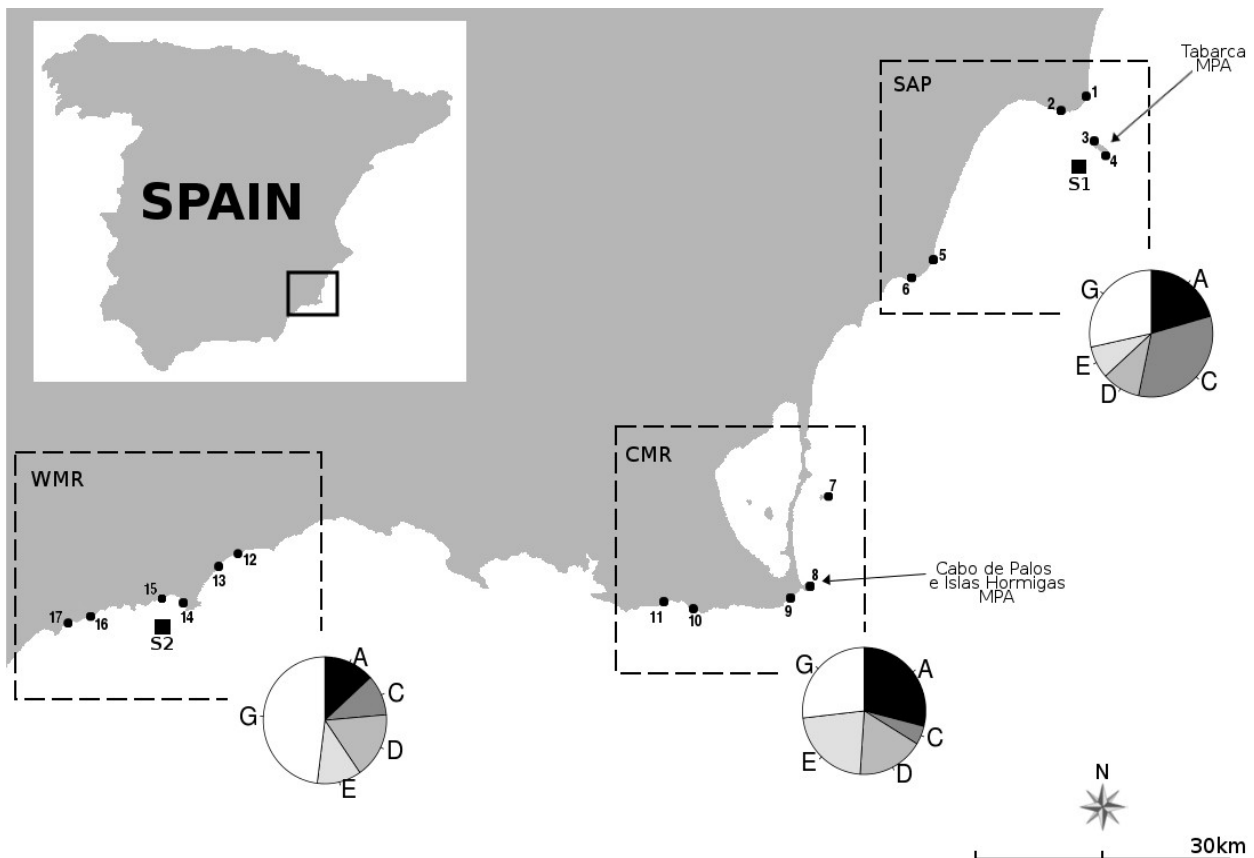
Fish propagule dispersal and patch cohesiveness

- 467 Kiparissis S, Tserpes G, Somarakis S, Economidis PS, Koutsikopoulos C (2008) Site-attachment
468 behaviour of *Oblada melanura* (Linnaeus, 1758) (Osteichthyes: Sparidae) benthic larvae: a
469 quantitative approach. *Sci Mar* 72:429–436
- 470 Kritzer JP, Sale PF (2004) Metapopulation ecology in the sea: from Levins' model to marine
471 ecology and fisheries science. *Fish Fish* 5:131–140
- 472 Leary S, Underwood W, Anthony R, Cartner S, Corey D, Grandin T, Meyer R, Shearer J (2013)
473 AVMA Guidelines for the Euthanasia of Animals : 2013 Edition.
- 474 Leis JM (2015) Is dispersal of larval reef fishes passive? In: Mora C (ed) *Ecology of fishes on Coral*
475 *Reefs*. Cambridge University Press, p 223–226.
- 476 Leis JM, van Herwerden L, Patterson HM (2011) Estimating connectivity in marine fish
477 populations: what works best? In: *Oceanography and Marine Biology: An Annual Review*. p
478 193–234
- 479 Lloret J, Zaragoza N, Caballero D, Font T, Casadevall M, Riera V (2008) Spearfishing pressure on
480 fish communities in rocky coastal habitats in a Mediterranean marine protected area. *Fish Res*
481 94:84–91
- 482 Miller JA, DiMaria RA, Hurst TP (2014) Patterns of larval source distribution and mixing in early
483 life stages of Pacific cod (*Gadus macrocephalus*) in the southeastern Bering Sea. *Deep Sea Res*
484 Part II Top Stud Oceanogr:1–13
- 485 Muhlfeld CC, Thorrold SR, McMahon TE, Marotz B, Gillanders B (2012) Estimating westslope
486 cutthroat trout (*Oncorhynchus clarkiilewisi*) movements in a river network using strontium
487 isoscapes. *Can J Fish Aquat Sci* 69:906–915
- 488 Nanninga GB, Berumen ML (2014) The role of individual variation in marine larval dispersal. *Front*
489 *Mar Sci* 1:1–17
- 490 Papetti C, Franco A Di, Zane L, Guidetti P, Simone V De, Spizzotin M, Zorica B, Cikes Kec V,
491 Mazzoldi C (2013) Single population and common natal origin for Adriatic *Scomber scombrus*
492 stocks: evidence from an integrated approach. *ICES J Mar Sci* 70:387–398
- 493 Pérez-Ruzafa Á (2010) Project final memory. Coastal oceanographic observatory of Murcia.
494 Murcia.
- 495 Planes S, Jones GP, Thorrold SR (2009) Larval dispersal connects fish populations in a network of
496 marine protected areas. *Proc Natl Acad Sci* 106:5693–5697
- 497 Rossi V, Ser-Giacomi E, López C, Hernández-García E (2014) Hydrodynamic provinces and
498 oceanic connectivity from a transport network help designing marine reserves. *Geophys Res*
499 *Lett* 41:2883–2891
- 500 Ruttenberg B, Hamilton S, Hickford M, Paradis G, Sheehy M, Standish J, Ben-Tzvi O, Warner R
501 (2005) Elevated levels of trace elements in cores of otoliths and their potential for use as
502 natural tags. *Mar Ecol Prog Ser* 297:273–281

Fish propagule dispersal and patch cohesiveness

- 503 Ruttenberg BI, Hamilton SL, Warner RR (2008) Spatial and temporal variation in the natal otolith
504 chemistry of a Hawaiian reef fish: prospects for measuring population connectivity. *Can J Fish*
505 *Aquat Sci* 65:1181–1192
- 506 Sandin S a., Regetz J, Hamilton SL (2005) Testing larval fish dispersal hypotheses using maximum
507 likelihood analysis of otolith chemistry data. *Mar Freshw Res* 56:725
- 508 Shima JS, Noonburg EG, Swearer SE (2015) Consequences of variable larval dispersal pathways
509 and resulting phenotypic mixtures to the dynamics of marine metapopulations. *Biol Lett* 11
- 510 Shima JE, Swearer SE (2009) Larval quality is shaped by matrix effects: implications for
511 connectivity in a marine metapopulation. *Ecology* 90:1255–1267
- 512 Shima JS, Swearer SE (2010) The legacy of dispersal: larval experience shapes persistence later in
513 the life of a reef fish. *J Anim Ecol* 79:1308–14
- 514 Walther B, Thorrold S (2006) Water, not food, contributes the majority of strontium and barium
515 deposited in the otoliths of a marine fish. *Mar Ecol Prog Ser* 311:125–130

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517 Figure 1. Map of the study area. Black dots indicate post-settlers sampling sites (numbers represent
 518 site names); dashed line rectangles represents zones: south Alicante province (SAP), central Murcia
 519 region (CMR) and west Murcia region (WMR). Black squares represent pre-settlers sampling sites.
 520 Pie charts indicate, for each zone, the proportion of post-settlers belonging to the five major natal
 521 sources clustered (see results).

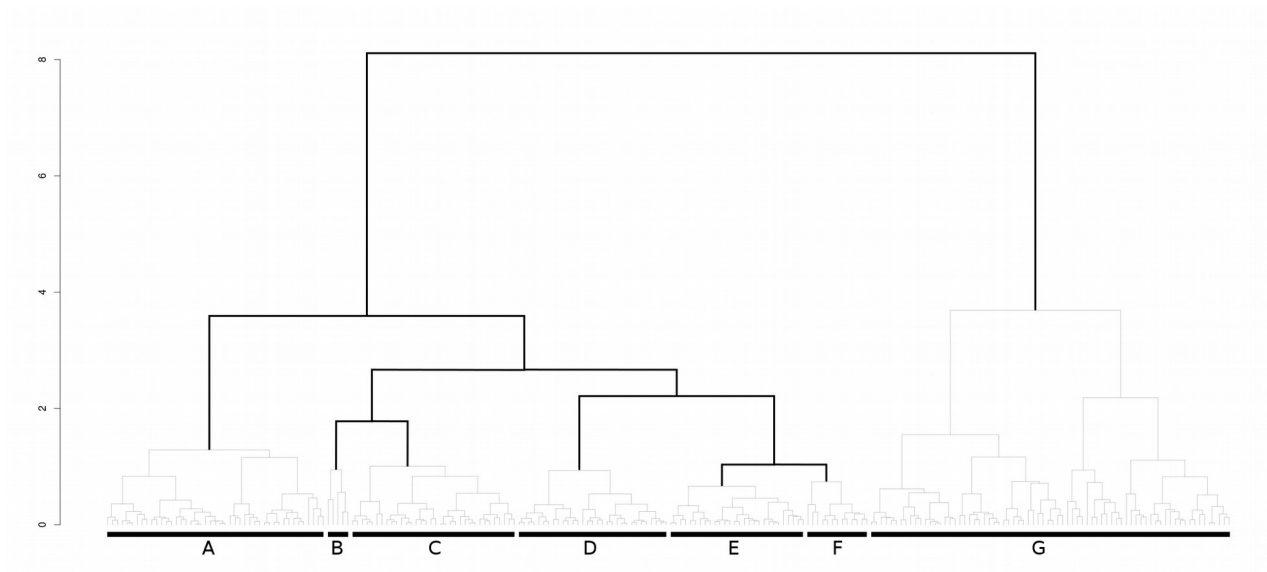
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Element ratio	NIST 610 %RSD	NIST 612 %RSD	NIST 610 %Accuracy	NIST 612 %Accuracy	LOD mmol/mol
Mg:Ca	9.8	11.1	84	92	0.03852
Sr:Ca	7.8	8.3	115	98	0.00148
Ba:Ca	7.1	9.4	91	89	0.00066

Element ratio	NIST 610 %RSD	NIST 612 %RSD	NIST 610 %Accuracy	NIST 612 %Accuracy	LOD mmol/mol
Mg:Ca	6.6	14.4	98	106	0.04568
Sr:Ca	7	9.3	98	93	0.01441
Ba:Ca	6.1	7	98	82	0.00076

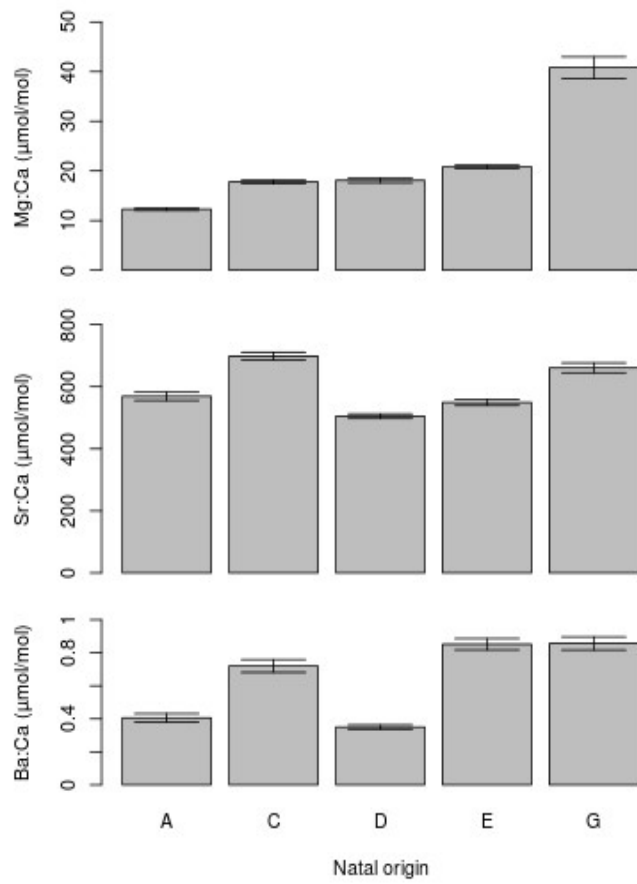
523 Table 1. Estimates of precision, accuracy and limits of detection (LOD) for post settlers' otoliths
524 (upper table) and pre settlers' otoliths (lower table). Values for %RSD (% relative standard
525 deviation) and % accuracy are dimensionless.

Fish propagule dispersal and patch cohesiveness



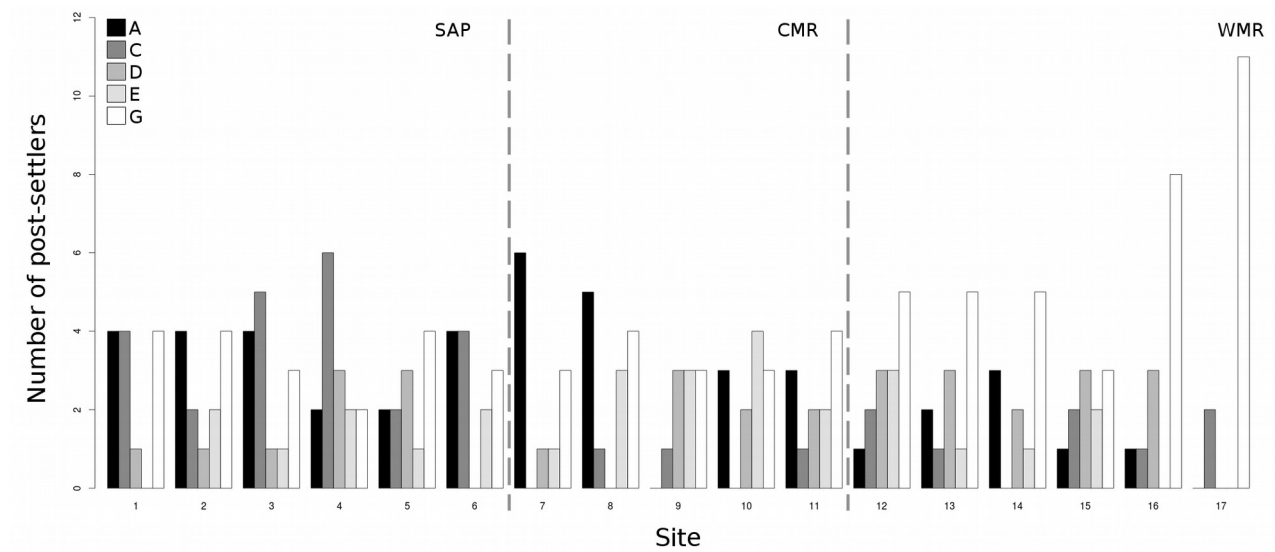
527 Figure 2. Cluster (based on Ward's approach) of post-settlers otolith cores into groups considering
528 Mg:Ca, Sr:Ca and Ba:Ca elemental concentrations. Black branches indicate statistically significant
529 groups resulted from SIMPROF (arbitrarily named from left to right). Grey branches indicate non-
530 significant differences.

Fish propagule dispersal and patch cohesiveness



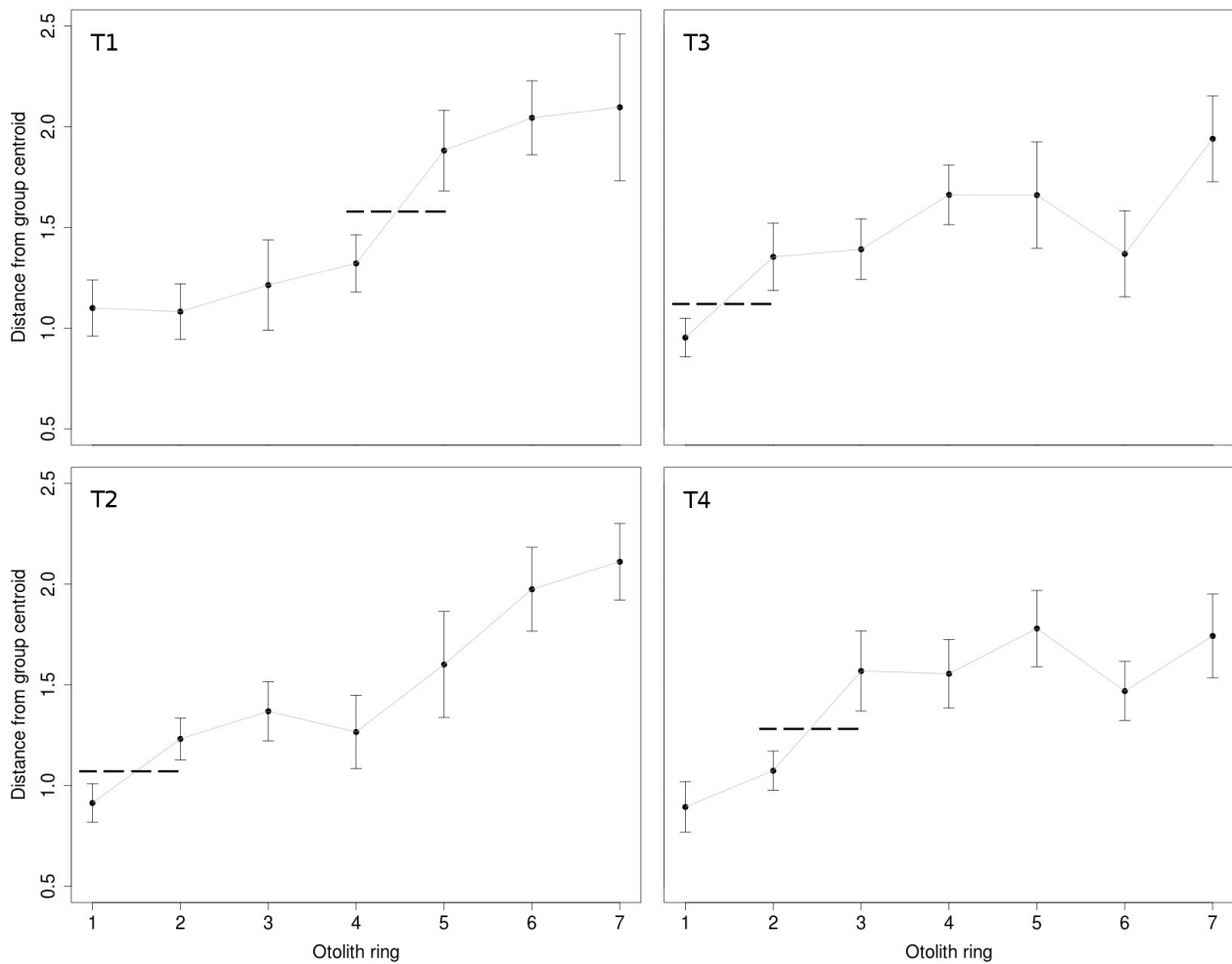
531 Figure 3. Average (\pm standard error) element/calcium ratios in the core region of the five major
532 clusters identified by SIMPROF (see Figure 2).

Fish propagule dispersal and patch cohesiveness



534 Figure 4. Total number of post-settlers, for each sampling site, coming from the five major natal
535 sources. Different colors represent the five major groups identified by SIMPROF. Sites are
536 numbered progressively from the northernmost to the southernmost (See Fig. 1). Vertical dashed
537 lines separate the three zones considered. Source B and F (minor sources) were omitted for enhance
538 readability.

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539 Figure 5. Plots of average distance (\pm standard error) from group centroid per each otolith ring (day
540 of life) analysed, as calculated by PERMDISP. Each plot represent a light-trap sample: T1 (10
541 specimens), T2 (8 specimens), T3 (11 specimens) and T4 (11 specimens). Dashed segments
542 represent statistically significant increments in chemical fingerprint variability between pairs of
543 consecutive rings.