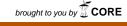
GENERAL VIROLOGY

ICTV VIRUS TAXONOMY PROFILE

Sato et al., Journal of General Virology 2019;100:1269–1270 DOI 10.1099/jgv.0.001297



MICROBIOLOGY

ICTV Virus Taxonomy Profile: Megabirnaviridae

Yukiyo Sato¹, Naoyuki Miyazaki², Satoko Kanematsu³, Jiatao Xie⁴, Said A. Ghabrial⁵†, Bradley I. Hillman⁶, Nobuhiro Suzuki^{1,*} and ICTV Report Consortium

Abstract

Megabirnaviridae is a family of non-enveloped spherical viruses with dsRNA genomes of two linear segments, each of 7.2–8.9 kbp, comprising 16.1 kbp in total. The genus *Megabirnavirus* includes the species *Rosellinia necatrix megabirnavirus* 1, the exemplar isolate of which infects the white root rot fungus (*Rosellinia necatrix*) to which it confers hypovirulence. Megabirnaviruses are characterized by their bisegmented genome with large 5'-untranslated regions (1.6 kb) upstream of both 5'-proximal coding strand ORFs, and large protrusions on the particle surface. This is a summary of the ICTV Report on the family *Megabirnaviridae*, which is available at ictv.global/report/megabirnaviridae.

This Profile is dedicated to the memory of our valued colleague Professor Said A. Ghabrial.

Table 1. Characteristics of members of the family Megabirnaviridae

Typical member:	Rosellinia necatrix megabirnavirus 1-W779 (RNA1: AB512282; RNA2: AB512283), species Rosellinia necatrix megabirnavirus 1, genus Megabirnavirus
Virion	Isometric, non-enveloped particles, 52 nm in diameter; dsRNA segments may be separately encapsidated
Genome	Two linear dsRNAs of 7.2-8.9 kbp, 16.1 kbp in total; large untranslated regions (over 1.6 kb) upstream of both 5'-proximal coding strand ORFs
Replication	Possibly within virus particles, as observed for other dsRNA viruses
Translation	Possible internal ribosomal entry site translation of the 5'-proximal ORFs on the mRNAs from dsRNA1and dsRNA2. The 3'-proximal ORF of dsRNA1 is translated via -1 ribosomal frameshifting
Host range	Fungi
Taxonomy	Realm Riboviria, one genus including a single species

VIRION

Megabirnaviruses form rigid spherical particles with a diameter of 52 nm (Table 1, Fig. 1a) [1, 2]. Each capsid with a T=1 lattice is composed of 60 asymmetric homodimers of the capsid protein, P1 (Fig. 1b), which are presumed to produce 120 protrusions on the virus surface. The genomic segments, dsRNA1 and dsRNA2, appear to be packaged into separate particles. The major capsid protein is encoded by the 5'-proximal open reading frame (ORF) on dsRNA1, which also encodes an RNA-directed RNA polymerase (RdRP) in

the 3'-proximal ORF (Fig. 2). Purified virions include capsid protein-RdRP fusion protein, which is translated from polycistronic mRNA transcribed from dsRNA1 [1, 3]. Purified virions are infectious to protoplasts of natural and experimental fungal hosts [1, 3, 4].

GENOME

The genome of Rosellinia necatrix megabirnavirus 1-W779 consists of two segments, dsRNA1 and dsRNA2 (8.9 kbp and

*Correspondence: Nobuhiro Suzuki, nsuzuki@okayama-u.ac.jp

†Deceased

Received 05 June 2019; Accepted 19 June 2019; Published 16 August 2019

Author affiliations: ¹Institute of Plant Science and Resources, Okayama University, Chuo 2-20-1, Kurashiki 710-0046, Japan; ²Life Science Center for Survival Dynamics, Tsukuba Advanced Research Alliance, University of Tsukuba, Tsukuba, Ibaraki 305-8577, Japan; ³National Agriculture and Food Research Organization (NARO) Headquarters, 3-1-1 Kannondai, Tsukuba 305-8517, Japan; ⁴College of Plant Science and Technology, Huazhong Agricultural University, Wuhan, PR China; ⁵Department of Plant Pathology, University of Kentucky, Lexington, KY 40546, USA; ⁶Department of Plant Biology and Pathology, Rutgers University, New Brunswick, NJ 08901, USA.

Keywords: ICTV Report; taxonomy; Megabirnaviridae.

Abbreviations: RdRP, RNA-directed RNA polymerase.

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

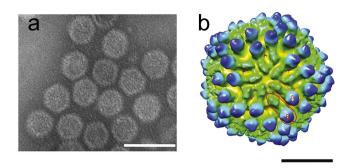


Fig. 1. Megabirnavirus particle structure – Rosellinia necatrix megabirnavirus 1-W779. (a) Transmission electron micrograph of negatively-stained virions. Scale bar, 100 nm. Reproduced with permission from [1]. (b) Surface representation of virion reconstructed by three-dimensional cryo-EM at a resolution of 15.7 Å. Scale bar, 20 nm. Reproduced with permission from [2].

7.2 kbp, respectively) (Fig. 2) [1]. Each segment has two ORFs; dsRNA1 encodes the capsid (P1) and RdRP (P2) proteins, while dsRNA2 encodes the hypothetical proteins, P3 and P4, with unknown functions. The 5'-proximal ORFs are preceded by large (over 1.6 kb) untranslated regions. There is a putative slippery sequence (5'-AAAAAAC-3') immediately before the stop codon of the P1 ORF, followed by a sequence that is predicted to form a potential stem–loop structure. This suggests that the capsid protein-RdRP fusion protein is likely to be produced via –1 ribosomal frameshifting.

TAXONOMY

The exemplar isolate of *Rosellinia nexatrix megabirnavirus* 1, a member of the genus *Megabirnavirus*, is Rosellinia necatrix megabirnavirus 1-W779, which was isolated from a phytopathogenic ascomycetous fungus that causes white root rot in many perennial crops worldwide. The mycoviruses Sclerotinia sclerotiorum megabirnavirus 1 and Rosellinia necatrix megabirnavirus 2 are closely related to Rosellinia

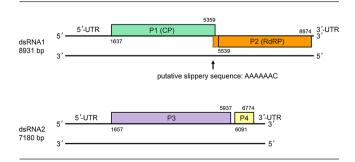


Fig. 2. Genomic organization of Rosellinia necatrix megabirnavirus 1-W779. Black lines represent genomic dsRNA segments. Boxes indicate ORFs on the positive-sense strands with dotted lines indicating the start of the -1 frameshifted reading frame .

necatrix megabirnavirus 1 but are currently unclassified [5, 6]. Other mycovirus sequences have been detected from diverse ascomycetous and basidiomycetous fungi [7–10], and have deduced RdRP amino acid sequences with 30–47 % identity to that of Rosellinia necatrix megabirnavirus 1.

RESOURCES

Full ICTV Report on the family *Megabirnaviridae*: ictv.global/report/megabirnaviridae.

Funding information

Production of this summary, the online chapter, and associated resources was funded by a grant from the Wellcome Trust (WT108418AIA).

Acknowledgements

Conflicts of interest

Members of the ICTV (10th) Report Consortium are Elliot J. Lefkowitz, Andrew J. Davison, Stuart G. Siddell, Sead Sabanadzovic, Donald B. Smith, Richard J. Orton and Peter Simmonds.

The authors declare that there are no conflicts of interest.

References

- Chiba S, Salaipeth L, Lin YH, Sasaki A, Kanematsu S et al. A novel bipartite double-stranded RNA mycovirus from the white root rot Fungus Rosellinia necatrix: molecular and biological characterization, taxonomic considerations, and potential for biological control. J Virol 2009;83:12801–12812.
- Miyazaki N, Salaipeth L, Kanematsu S, Iwasaki K, Suzuki N. Megabirnavirus structure reveals a putative 120-subunit capsid formed by asymmetrical dimers with distinctive large protrusions. *J Gen Virol* 2015;96:2435–2441.
- Salaipeth L, Chiba S, Eusebio-Cope A, Kanematsu S, Suzuki N. Biological properties and expression strategy of rosellinia necatrix megabirnavirus 1 analysed in an experimental host, *Cryphonectria* parasitica. J Gen Virol 2014;95:740–750.
- Kanematsu S, Shimizu T, Salaipeth L, Yaegashi H, Sasaki A et al. Genome rearrangement of a mycovirus Rosellinia necatrix megabirnavirus 1 affecting its ability to attenuate virulence of the host fungus. Virology 2014;450-451:308–315.
- Wang M, Wang Y, Sun X, Cheng J, Fu Y et al. Characterization of a novel megabirnavirus from *Sclerotinia sclerotiorum* reveals horizontal gene transfer from single-stranded RNA virus to doublestranded RNA virus. *J Virol* 2015;89:8567–8579.
- Sasaki A, Nakamura H, Suzuki N, Kanematsu S. Characterization of a new megabirnavirus that confers hypovirulence with the aid of a co-infecting partitivirus to the host fungus, *Rosellinia necatrix*. *Virus Res* 2016;219:73–82.
- Zhong J, Chen CY, Gao BD. Genome sequence of a novel mycovirus of *Rhizoctonia solani*, a plant pathogenic fungus. *Virus Genes* 2015;51:167–170.
- Nerva L, Ciuffo M, Vallino M, Margaria P, Varese GC et al. Multiple approaches for the detection and characterization of viral and plasmid symbionts from a collection of marine fungi. Virus Res 2016;219:22–38.
- Arjona-Lopez JM, Telengech P, Jamal A, Hisano S, Kondo H et al. Novel, diverse RNA viruses from Mediterranean isolates of the phytopathogenic fungus, *Rosellinia necatrix*: insights into evolutionary biology of fungal viruses. *Environ Microbiol* 2018;20:1464–1483.
- Zhang X, Gao F, Zhang F, Xie Y, Zhou L et al. The complete genomic sequence of a novel megabirnavirus from Fusarium pseudograminearum, the causal agent of wheat crown rot. Arch Virol 2018;163:3173–3175.