

1 **Nest cover and faecal glucocorticoid metabolites are linked to hatching success and**  
2 **telomere length in breeding eiders (*Somateria mollissima*)**

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21

22 **ABSTRACT**

23 Habitat-associated crypsis may affect perceived predation vulnerability, selecting for different predator  
24 avoidance strategies. Glucocorticoids could mediate the adjustment of escape responses to the extent of  
25 crypsis, introducing an overlooked source of variation in glucocorticoid-fitness relationships. However,  
26 prolonged exposure to elevated glucocorticoids may be costly leading to accelerated telomere loss and  
27 consequently senescence. Here, we examined how nest cover and immunoreactive faecal glucocorticoid  
28 metabolite levels (fGCM) are linked to hatching success and telomere length in breeding female eiders  
29 (*Somateria mollissima* Linnaeus, 1758). We hypothesized that the degree of nest crypsis, reflecting  
30 differences in perceived predation risk, would moderate the relationship between reproductive success  
31 and fGCM levels. We also expected that telomere length would be shorter in birds with higher  
32 glucocorticoid concentration. Results showed that individuals with high fGCM levels had higher  
33 hatching success in nests with low cover, while low fGCM levels were more successful in well-  
34 concealed nests. We found that shorter telomeres were associated with high fGCM in nesting sites  
35 offering little cover and with low fGCM in well-concealed ones. This study provides the first evidence of  
36 habitat-dependent moderation of the relationships between stress physiology, telomere length and  
37 hatching success.

38 **Keywords:**

39 Cost of reproduction, eider, glucocorticoids, nesting habitat, *Somateria mollissima*

40

41 **INTRODUCTION**

42 Glucocorticoid (GC) stress hormones are considered to play a key role in integrating information about  
43 environmental challenges and in mediating inter-individual differences in fitness (Ricklefs and Wikelski  
44 2002; Boonstra 2013). So far, however, no consensus has emerged on the direction of the relationship  
45 between GCs and fitness in natural populations (Wingfield and Sapolsky 2003; Bonier et al. 2009a, b,  
46 Crossin et al. 2012; Jaatinen et al. 2013). This uncertainty is perhaps not surprising given that a negative  
47 relationship between GCs and fitness is often assumed *a priori*, based on evidence from biomedical  
48 research (Boonstra 2013). Thus, prolonged exposure to high concentrations of GCs is considered to be  
49 costly and subject individuals to pathologies (Ricklefs and Wikelski 2002). In contrast to laboratory  
50 models, however, wild animals are exposed to a diverse array of stressors, including predation risk. A  
51 short-term increase in GC levels is a vital response immediately before, during and after a predatory  
52 attack (reviewed by Wingfield et al. 1998). Less known and appreciated is the fact that circulating  
53 higher baseline GC concentrations may be adaptive whenever the risk of stress exposure is high  
54 ('preparative hypothesis'; Romero 2002), as GCs may prepare the organism to perform better under  
55 such circumstances (Sapolsky et al. 2000). Thereby, long-term elevation of GCs may serve to prepare  
56 prey for attacks by predators, for instance, by increasing vigilance (Scheuerlein et al. 2001; Cockrem  
57 and Silverin 2002; Hawlena and Schmitz 2010). If the benefits of such preparative responses outweigh  
58 the costs, an increase in GCs can be adaptive and continue to promote fitness (Boonstra 2013).  
59 Therefore, the ambiguity in the relationship between GCs and fitness may partly reflect our incomplete  
60 understanding of stress coping strategies in the wild.

61

62 The ability of the breeding habitat to provide protection has received little attention as a contextual  
63 factor affecting the relationship between GCs and fitness (Crespi et al. 2013; but see D'Alba et al. 2011).  
64 The degree of crypsis provided by the nest site is inexorably linked to the optimal predator avoidance

65 tactic. The optimal solution in well-concealed nest sites may be to rely on crypsis and down-regulate  
66 escape behaviour (Amat and Masero 2004; Albrecht and Klvaňa 2004), whereas preparing for predatory  
67 attacks by maintaining escape performance at a high level may maximize breeding success in less-  
68 concealed nests (Merilaita et al. 1999). Such increased vigilance and escape performance has been  
69 linked to high baseline GC levels (Sapolsky et al. 2000; Chin et al. 2009; Thaker et al. 2010).  
70 Experimental evidence suggests that the nest characteristics themselves may not directly affect the stress  
71 hormone levels of breeding birds, but rather that the breeders adopt different strategies of nest-site  
72 selection depending on their phenotypic traits (D'Alba et al. 2011). Although a parent selecting a poorly  
73 concealed nest site may have high GC levels due to high perceived predation risk, this physiological  
74 response may in fact represent an appropriate predatory avoidance strategy enhancing reproductive  
75 success (Boonstra 2013). While such a response may be adaptive, increased GCs may still have  
76 detrimental effects on the condition and future reproductive potential of individuals in long-lived species  
77 (Johnson 2007; Haussmann and Marchetto 2010).

78

79 The ability to cope with external and internal challenges varies widely between individuals (Wilson and  
80 Nussey 2010) and this variation may be associated with habitat choice (e.g., D'Alba et al. 2011). Despite  
81 this, only a few studies have considered the intrinsic stress tolerance quality of individuals occupying  
82 different habitats (e.g., Germain and Arcese 2014). Thus, we still know very little about fitness value  
83 associated with a given breeding habitat. Telomeres, nucleoprotein structures located at the ends of  
84 chromosomes, hold promise as a composite indicator of physiological stress associated with internal and  
85 external challenges (Mizutani et al. 2013; Young et al. 2015; LeVaillant et al. 2015). In general,  
86 individuals with longer than average telomeres for their age have longer life expectancy (Heidinger et al.  
87 2012; Barrett et al. 2013; Angelier et al. 2013), a higher number of functional cells (Monaghan and  
88 Haussmann 2006; Monaghan 2014), and higher stress resistance (Kotrschal et al. 2007). However,

89 chronically elevated GCs can accelerate telomere shortening (von Zglinicki 2002; Epel et al. 2004; Choi  
90 et al. 2008; Hausmann and Marchetto 2010). Because the typically high reproductive investment by  
91 good-quality individuals may be facilitated by elevated GC levels (Crossin et al. 2012), such investment  
92 may incur costs in terms of accelerated telomere attrition (Reichert et al. 2014, Schultner et al. 2014;  
93 Sudyka et al. 2014). However, breeding animals interact with their chosen breeding habitat and also  
94 telomere dynamics have been linked to habitat choice (Angelier et al. 2013). This adds a previously  
95 unappreciated level of complexity to the interrelationships between reproductive success, glucocorticoid  
96 stress physiology and telomere dynamics. Increased GC levels may facilitate reproduction and offspring  
97 care and can help individuals adjust their antipredatory behavior depending on the habitat-specific risk  
98 of predation. Thus, while the immediate benefits of elevated GCs are evident, elevated GC levels have  
99 also been shown to carry long-term costs in the form of telomere shortening and subsequently lowered  
100 survival (Kotrschalet al. 2007).

101

102 To bring clarity to these issues, we explored potential links between the degree of visual nest  
103 concealment and stress physiology, telomere length and breeding success. We hypothesized interactive  
104 effects of nest cover and GCs on breeding success: low nest cover is associated with higher perceived  
105 predation risk than covered nests and thus the optimal antipredatory response may differ between  
106 degrees of nest concealment. We predicted that individuals with higher GC, and thereby presumably  
107 enhanced anti-predator responsiveness, would have the greatest reproductive output in poorly-concealed  
108 nests facilitating rapid escape, whereas individuals attaining high reproductive success in concealed  
109 nests would exhibit lower GC levels and rely on crypsis instead of escape. We also hypothesized that  
110 high levels of reproductive performance, either in association with elevated GC levels or independently,  
111 may be linked to shorter telomeres (see Bauch et al. 2013).

112

113 As a model system, we used female eider ducks (*S. mollissima*) from a well-studied population in south-  
114 western Finland. The eider is an excellent study species. Female eiders rely to a large extent on stored  
115 energy resources during incubation (Parker and Holm 1990; Bolduc and Guillemette 2003; but see  
116 Hobson et al. 2015; Jaatinen et al. 2016). These limited resources can be mobilized by GCs and  
117 consistent individual differences in baseline GC profiles are associated with individual differences in  
118 current reproductive success (Jaatinen et al. 2013), and thus potentially also long-term fitness. Also,  
119 females show fidelity to nest sites (Öst et al. 2011; Ekroos et al. 2012) and the degree of nest cover is  
120 repeatable between years (Öst and Steele 2010; Selmann et al. 2014). Further, incubating females  
121 encounter a spatially and temporally varying risk of attack by predators, posing a considerable threat for  
122 this ground-nesting bird (Ekroos et al. 2012). The relationship between the acute (handling-induced)  
123 stress response and reproductive investment has previously been shown to be modulated by predation  
124 risk (Jaatinen et al. 2014). Finally, the number of years of maternal experience, a proxy for age, does not  
125 explain the variability in telomere length observed among adult eider females (this study), and thus age  
126 is not likely to confound or mask the associations under focus here.

127

128 A previous experimental study on female eiders showed that nest shelter did not affect baseline plasma  
129 GC levels (D'Alba et al. 2011). However, this study also suggested that nest habitat was not independent  
130 of individual quality and that the relationship between hatching success and CORT was affected by  
131 female body condition, i.e., it was state-dependent. This study explores these possibilities further by first  
132 assessing the relationships between nest cover, female hatching success and faecal glucocorticoid  
133 metabolite (fGCM) level, an accumulative index of stress (Möstl et al. 2005). fGCMs provide a more  
134 integrated measure of adrenocortical activity than point serum samples and thus diminish the influence  
135 of temporal changes in GC secretion (Whitten et al. 1998). Thereafter, we examined the associations  
136 between individual biological state, as quantified by telomere length, nest-site cover and fGCM. In all

137 analyses we also controlled for other potentially important predictors of hatching success and telomere  
138 length, including female breeding experience, body condition and timing of breeding.

139

## 140 **MATERIALS AND METHODS**

141

### 142 *Field methods*

143

#### 144 (a) Study area and population

145 The study was conducted at Tvärminne Zoological Station (59°50'N, 23°15'E), in the western Gulf of  
146 Finland, in 2009-2011. The 23 study islands were represented by 9 forested island and 14 open rocky  
147 islets. Nest cover is highly variable on both island types as females readily nest under trees, bushes such  
148 as junipers (which are often abundant also on open islands), rock outcrops or concealed in grassy  
149 vegetation. Annually all study islands are searched through with equal thoroughness so that all nesting  
150 events are recorded. The number of nests on the islands ranged between 0-94 (mean  $\pm$  SD = 15.7  $\pm$  2.1)  
151 during the study period (Jaatinen et al. 2014). Female eiders in the study population nest at low densities  
152 and previous evidence suggests that nest-site selection is not affected by female competition or nest-site  
153 limitation (Öst et al. 2008; Öst and Steele 2010; Ekroos et al. 2012; Selmann et al. 2014), which  
154 contrasts with the situation described for eiders in other populations that nest in dense colonies (e.g.,  
155 D'Alba et al. 2011). Thus, in our current sample of individuals, there was no association between nest  
156 cover and the onset of breeding (linear mixed model:  $b = 0.001$ ,  $SD = 0.002$ ,  $t = 0.303$   $p > 0.05$ ,  $N$   
157 (observations/females) = 472/346) or between nest cover and body condition (linear mixed model:  $b =$   
158  $0.018$ ,  $SD = 0.012$ ,  $t = 1.478$ ,  $p > 0.05$ ,  $N$  (observations/females) = 472/346). Therefore, it is reasonable  
159 to assume that individual quality does not create a significant confounding effect on initial nest-site  
160 selection, which instead represents the outcome of an active decision-making process when females

161 come to the area to breed for the first time. In contrast, nest-site choices in subsequent breeding seasons  
162 may to some degree be constrained by high fidelity to the particular breeding island (Öst et al. 2011).  
163 The low breeding dispersal has been identified as a putative ecological trap for females (Ekroos et al.  
164 2012). Thus, despite an increased propensity to switch nest sites after events of nest depredation (Öst et  
165 al. 2011), females still exhibit high breeding philopatry regardless of the increased predation pressure on  
166 adults by white-tailed sea eagles (*Haliaeetus albicilla* Linnaeus, 1758), eagle owls (*Bubo bubo*  
167 Linnaeus, 1758) and several mammalian predators. On average open islands are subjected to higher  
168 predation pressure than islands with a forest cover (Ekroos et al. 2012). In a short term, spatial predation  
169 patterns may change due to predator movement between islands and this type of variation is more  
170 pronounced than temporal variation in predation pressure (Öst et al. 2011)

171

#### 172 **(b) Female trapping and measurements**

173 We captured nesting eider females (535 captures of 381 individuals) on their nests by using hand nets.  
174 Captured females were weighed with a spring balance to the nearest 10g, measured for structural size  
175 (length of the radius-ulna to the nearest 1mm), ringed, and their clutch size was recorded. Clutch size  
176 varied between 2-7 eggs (mean  $\pm$  *SD* = 4.69  $\pm$  1.16). Ducklings are not ringed in this population and  
177 hence female age could not be directly determined; therefore the ringing information was used to  
178 calculate the number of years since the bird was first trapped, indicating minimum years of maternal  
179 experience (Öst et al. 2008; Öst and Steele 2010; Jaatinen and Öst 2011; Jaatinen et al. 2012). This is a  
180 reasonably good proxy for female age in the population due to the high breeding philopatry and the fact  
181 that more than half of the breeding females in the population are captured annually, with a relatively  
182 constant annual trapping effort since 1996 (Jaatinen and Öst 2011).

183

184 We obtained female blood samples by extracting approximately 1 ml of peripheral blood from the



185 brachial vein. Faecal samples were collected in Whirl-Paks (Nasco) directly from the female or by  
186 gathering fresh faeces from the nest, and both blood and faecal samples were immediately stored on ice  
187 in a cool box and transported to the laboratory within 2–4 *h*. Blood samples were centrifuged in a cold  
188 centrifuge (Sigma 3K12, B. Broun, Germany) for 10 min at 1500×*g* to separate blood serum and cells.  
189 Blood cells and faecal samples were stored frozen in –20°C until further analyses. Faecal samples  
190 collected during 2009-2011 (*N*=514/369) were used for immunoreactive fGCM measurement, whereas  
191 blood cells were collected only in 2011 (*N*=197) and subsequently used for telomere measurement.

192

193 We used egg floatation to determine the incubation stage at female capture (Kilpi and Lindström 1997).  
194 Information on incubation stage was used to calculate a body condition index since eider females refrain  
195 completely from feeding during the incubation period and lose up to 40% of their pre-laying body mass  
196 (e.g., Parker and Holm 1990). Body condition indices were determined for all trapped females that had  
197 been incubating eggs for at least 8 days (egg laying may otherwise not have been completed; Öst et al.  
198 2008). The index was given by the standardized residuals of a regression of log-transformed projected  
199 weight at hatching (response variable) on log-transformed radius–ulna length, and indices were derived  
200 separately for each year (Öst and Steele 2010). A female’s body mass at hatching was estimated by  
201 subtracting an estimate of the expected body mass loss during the remaining incubation time from her  
202 measured incubation body mass. Females were weighed once, but as females abstain from feeding  
203 during incubation and females were captured at different times in their incubation, we can derive an  
204 estimate of average mass loss rate during incubation as the slope of the regression of log-transformed  
205 body mass (response variable) on log-transformed incubation time and projected hatching date (Öst et  
206 al. 2008). The assumption of continued mass loss after female capture applies to our study population,  
207 which makes this index reliable for estimating body condition (Öst and Steele 2010).

208

209 Hatching success was determined upon subsequent nest visits that were timed to coincide with the  
210 expected hatching of the clutch, based on estimated incubation stage at trapping. Successful hatching  
211 was determined by observing live ducklings in the nests. If the female had left the nest with her brood  
212 prior to our arrival, we observed whether the egg shells remaining in the nest had intact egg membranes,  
213 indicating successful hatching (Öst and Steele 2010). Such egg shell remnants can be distinguished from  
214 those left after nest depredation. Thus, we were able to precisely determine the fate of the nests for the  
215 majority of trapped females ( $N=449$  observations (333 females) out of 535 (381 females), 83.9%). Of the  
216 nests with known fates in 2009-2011 (Supplementary Table 1), the majority were successful (at least one  
217 egg hatched; annual mean  $\pm SD = 63.1 \pm 15.1\%$ ), nearly one-third were depredated ( $29.2 \pm 13.3\%$ ),  
218 while only a small fraction were abandoned ( $7.7 \pm 6.2\%$ ). For all nests, we recorded the number of  
219 successfully hatched eggs (duckling has hatched and survived to leave the nest) and unhatched eggs (the  
220 number of eggs that failed to hatch due to depredation and abandonment, and to a lesser extent  
221 inviability). Hatching success of undisturbed eider nests is high at ca 90 %, showing low variability  
222 among clutches (Swennen 1989), and thus the small fraction of inviable eggs is unlikely to  
223 systematically bias our results. All nests with known fates were included in subsequent analyses (2009-  
224 2011;  $N=449/333$ ). To quantify spatial and temporal variation in predation risk, we calculated an annual  
225 island-specific predation index. This index was given by dividing the number of depredated nests with  
226 the total number of censused nests on a given island in each year (Öst et al. 2011).

227 Nestcover was quantified by taking hemispherical digital photographs with a Olympus C-740 camera  
228 equipped with a 42-mm Opteka fisheye lens. All nest photographs were taken right after females had  
229 hatched their broods to reduce bias caused by vegetation growth. Hemispherical images were taken by  
230 placing an upward facing camera on a stable surface in the nest (Öst and Steele 2010). We used the  
231 program Image Tool (v. 3.00; University of Texas Health Science Center, San Antonio) to process nest  
232 cover photographs. Firstly, images were converted to grey scale and pixels assigned as black or white so

233 that vegetation and other elements such as rocks providing cover were coded as black pixels whereas  
234 areas of open sky were coded as white pixels. Nest cover was then calculated as the proportion of black  
235 pixels in the image.

236

237 *Laboratory methods*

238 **(a) Telomere measurement**

239 Relative telomere length normalized for a non-variable copy gene (T/S), was measured in red blood  
240 cells (RBC), and corresponds to the average telomere length across chromosomes (Cawthon 2002). The  
241 length of telomeres in RBC reflects the telomere length of hematopoietic stem cells (Vaziri et al. 1994)  
242 and has been shown to correlate with telomere length in other tissues (Reichert et al. 2013). We obtained  
243 a relative telomere length measure (T/S) by using the real-time quantitative PCR (qPCR) method as  
244 proposed by Cawthon (2002) and previously validated for use in birds (Criscuolo et al. 2009). Results  
245 from the qPCR correlate well with the results obtained by terminal restriction fragment (TRF) analysis,  
246 the conventional method of telomere measurement, and qPCR has successfully been used in a growing  
247 number of studies (e.g., Criscuolo et al. 2009; Bize et al. 2009; Aviv et al. 2011; Heidinger et al. 2012).

248

249 Genomic DNA for the assay was extracted from 5µl of RBC following the protocol by Aljanabi and  
250 Martinez (1997). DNA integrity was determined by agarose gel electrophoresis where 50ng of  
251 undigested DNA were resolved in 1.5% agarose gel at 120V for 90min, and DNA purity and  
252 concentration was measured spectrophotometrically using a NanoDrop 2000 (ThermoFisher Scientific).  
253 Only intact samples, appearing as a tight crown migrating in parallel, and with a A260/A280 ratio >1.7  
254 were accepted for further analyses. Because telomere length has not previously been measured in eiders,  
255 we first validated the assay. We selected the *gapdh* gene (glyceraldehyde 3-phosphate dehydrogenase) to  
256 test whether it could function as the non-variable copy gene in the qPCR assay. Primers which were

257 originally developed for chickens (genbank accession number: NW\_001471525): *gapdh*Fw.: (5'-  
258 TCCTGTGACTTCAATGGTGA-3') and *gapdh* Rev.: (5'-AAACAAGCTTGACGAAATGG-3') also  
259 successfully amplified the *gapdh* gene fragment in eiders, which resulted in a single DNA band of the  
260 expected size (80bp) when visualized on an agarose gel. Negative control reactions, without template  
261 DNA, showed no detectable product, suggesting that primer-dimer formation during qPCR was  
262 negligible. To confirm that the *gapdh* gene fragment is non-variable in copy number we compared  
263 whether the number of copies was stable at the inter-individual level ( $N=31$ ) as well as at the intra-  
264 individual (for repeatedly sampled individuals with at least a 10-year gap in sampling,  $N=4$ ).  
265 Examination by qPCR showed that the *gapdh* copy number did not differ between individuals and it also  
266 did not systematically change with age in the same individuals. These results are also supported by other  
267 evidence suggesting that the avian *gapdh* gene is usually a single copy gene found on autosomes and no  
268 pseudogenes have been identified (Alström et al. 2011). For amplifying telomeric repeats we used the  
269 universal primers developed by Cawton (2002): *tel1b* (5'-  
270 CGGTTTGGTTTGGGTTTGGGTTTGGGTTTGGGTTTGGGTT-3') and *tel2b* (5'-  
271 GGCTTGCCTTACCCTTACCCTTACCCTTACCCTTACCCT-3'). After amplification, the products  
272 were visualized on agarose gels and we observed a smear which was most intense around 78bp as  
273 reported in previous studies (Criscuolo et al. 2009).

274

275 The qPCR reactions were carried out in BIO-RAD X1000 real time thermal cyclers (BIO-RAD) in 384  
276 well microplates (BIO-RAD). For this purpose, we used iQ<sup>TM</sup> SYBR® Green qPCR mix (BIO-RAD)  
277 which includes iTaq<sup>TM</sup> DNA polymerase, dNTPs, MgCl<sub>2</sub> and fluorescein-SYBR. The reaction mix  
278 contained iQ<sup>TM</sup> SYBR® Green qPCR mix, and 400 nM of forward and reverse primers for either  
279 telomere and *gapdh* gene fragment amplification. 10 ng of sample DNA was added to each reaction and  
280 each sample was measured in triplicate on the same plate. Each plate also included serial doubling

281 dilutions (from 1.25 ng/well to 40 ng/well) of a standard sample DNA plus a no template control, also  
282 carried out in triplicate for both telomere and *gapdh* reactions and later used to construct standard  
283 curves. The thermal cycling conditions for both amplicons, were as follows: an initial denaturation at  
284 95°C for 3 min followed by 45 cycles of 95°C for 15s, 58°C for 18s and 72°C for 30s. The melt curve,  
285 used to determine the specificity of the qPCR amplification, was generated by slowly increasing  
286 temperature (0.1°C/s) from 65 to 95 °C.

287

288 Our mean qPCR efficiencies, determined from the standard curve, were 102% ( $\pm 6.31$  SD) and 92%  
289 ( $\pm 4.9$  SD) for telomere and *gapdh* reactions, respectively, and thus fell within the acceptable range (85-  
290 115%) for reliable telomere measurements (Criscuolo et al. 2009). Cq values (defined as the cycle  
291 number at which the fluorescence reached a fixed threshold value) were standardized for interplate  
292 variation using the software GenEx6 (MultiD). Replicates of samples were scanned for outliers ( $CV >$   
293 5%) and resulted in the exclusion of one sample. Intraplate coefficients of variation for telomeres and  
294 *gapdh* Cq values were 1.7% and 0.68%, respectively. Inter-plate  $CV$ 's were 2.4% for telomere and 1.15  
295 % for *gapdh* Cq values. We calculated the average of Cq values for the replicates and used them to  
296 calculate relative telomere length (T/S) for 172 females. It was calculated according using a formula  
297 taking qPCR efficiencies into consideration (Pfaffl 2001).

298

### 299 **(b) Faecal glucocorticoid metabolite analysis**

300 Faecal GCM concentrations provide an integrated measure of faecal hormone profiles accumulated over  
301 periods up to several days, since female alarm excreta start to accumulate in the intestinal tract  
302 immediately after incubation onset, typically occurring after the second egg is laid (e.g., Andersson and  
303 Waldeck 2006). Although females were trapped at different incubation stages, fGCM levels do not  
304 systematically change with advancing incubation (Jaatinen et al. 2013). We measured immunoreactive

305 fGCMs by radioimmunoassay (RIA) using a double antibody kit (ImmuChem™ Double Antibody,  
306 Corticosterone, 1251 RIA Kit, MP Biomedicals, Orangeburg, NY). The assay was carried successfully  
307 for 423 samples from 317 individuals, which were sampled from one to three times during the study  
308 period. As a biological validation of the assay, we note that immunoreactive fGCMs in eiders have  
309 previously been found to be elevated up to 3 weeks after surgical interventions (Latty, 2008) and  
310 repeatable within individuals (Jaatinen et al. 2013). A detailed description of the protocol for the use of  
311 this RIA kit for eider fGMC can be found in Jaatinen et al. (2013). Briefly, serial dilutions (1:2 to 1:256)  
312 of ten pooled faecal extracts were used for constructing a displacement curve which is parallel to the  
313 standard curve. This allowed to determine a faecal dilution (1:8) where binding was close to 60% and  
314 which was used for all test samples. Radioactivity of the bound portion was read in gamma counter  
315 (Gamma C12, Diagnostic Products, CA). The mean recovery rate of 3H-labeled CORT added to faecal  
316 samples pools was  $78 \pm 10\%$ . The cross-reactivities with other steroids were: desoxycorticosterone  
317 (0.34%), testosterone (0.1%), cortisol (0.05%), aldosterone (0.03%), progesterone (0.02%) and 0.01%  
318 for other steroids. The mean sensitivity of the assay for immunoreactive fGCM was 13.4 ng/g (range  
319 7.70 to 21.95 ng/g) and the mean ( $\pm SD$ ) fGCM level in samples was  $167.81 \pm 124.16$  ng/g (range 11.2 to  
320 757.65 ng/g). Immunoreactive fGCM levels were always above detection limit and our intra-assay *CV*  
321 was less than 10% and inter-assay *CV* was 15.28%.

322

### 323 *Data analysis*

324

325 To elucidate the effects of immunoreactive fGCM level and nest cover on female hatching success, we  
326 constructed a generalized linear mixed effect model (GLMM) with a binomial error distribution where  
327 hatched eggs of a clutch were considered a success and unhatched eggs a failure. In more detail, we  
328 combined the number of hatched and unhatched eggs in each clutch using the “*cbind*” function in R (R

329 Core Team 2013). This procedure combines the number of successes and failures for each clutch and  
330 thus produces a clutch-specific hatching proportion, which takes into account the total number of  
331 observations (i.e., eggs) used to produce the clutch-specific hatching proportion. This response variable  
332 describes the probability by which a given egg hatches (here after, P(hatch)) and we tested whether it  
333 was explained by nest cover and female fGCM level (Table 1). To reduce statistical bias arising from  
334 missing covariates, we included additional variables known to affect female breeding success: female  
335 minimum breeding experience, body condition and hatch date. Because predation pressure varies  
336 between islands and between years (Öst et al. 2011), we also included island-specific annual predation  
337 risk as a covariate in the model. Year was included to account for annual differences in hatching dates  
338 and hatching success, which may arise due to factors other than those explicitly considered in the model.  
339 To test our hypothesis that nest cover and immunoreactive fGCM may have interacting effects on  
340 hatching success, we included the interaction term between fGCM and nest cover in the model. Model  
341 selection was done by removing all non-significant variables ( $\alpha = 0.05$ ) using backward stepwise model  
342 reduction, where the least significant covariates were removed one at a time until the model contained  
343 only significant variables and interactions. The model was fitted using Laplace approximation and  
344 female identity was included as a random effect to correct for repeated measurements on the same  
345 female in different years ( $N = 423$  observations of 317 females; Table 1).

346

347 To study the associations between telomere length, stress physiology and breeding microhabitat, we  
348 constructed a linear model (LM) where relative telomere length was explained by immunoreactive  
349 fGCM, nest cover and the interaction between these two variables (Table 2). We included minimum  
350 maternal experience and body condition as covariates, to account for the potential telomere attrition with  
351 advancing age and potential links between telomere length and individual body condition. Telomere  
352 length was log transformed to ensure the normality of model residuals. Non-significant ( $\alpha = 0.05$ ) were

353 removed from the model using backward stepwise model reduction as described above.

354

355 To graphically illustrate significant interaction terms, these were analysed *post hoc* using the established  
356 method of simple slope analysis (Aiken and West 1991). Predictive trend lines depicted in graphs serve  
357 to illustrate significant interaction between two non-discrete predictors. Grouping of females into three  
358 categories depending on the concentration of immunoreactive fGCM (low– L, medium – M, high – H)  
359 was done after the statistical analyses therefore significance of the interaction is not affected by the  
360 grouping of females. In short, regression equations were restructured to reflect the regression of the  
361 criterion on one predictor and simple slope regressions were plotted to display the interactions at the  
362 mean and 1SD above and below the mean. All statistical analyses were performed in R2.13.0 (R Core  
363 Team 2011).

364

## 365 **RESULTS**

366

### 367 *Hatching success*

368 We found that the relationship between immunoreactive fGCM levels and hatching success varied with  
369 the degree of nest cover (fGCM  $\times$  nest cover interaction:  $b = -0.018$ ;  $SE = 0.004$ ;  $Z = -4.187$ ;  $p < 0.001$ ;  
370  $N$  (observations/individuals)=423/317;  $R^2_{\text{marg.}} = 0.18$ ; Table 1, Fig.1). For eider females with low fGCM,  
371 hatching success was positively associated with nest site cover. However, the opposite was observed for  
372 females with high fGCM levels; high proportional hatching success was associated with low nest cover.  
373 Hatching success decreased with advancing hatching date, as it did with increasing island-specific  
374 annual predation risk (Table 1). However, minimum maternal experience and body condition were not  
375 significantly associated with proportional hatching success, and there was no significant year effect



376 (Table 1).

377

378 *Telomere length*

379 Variation in telomere length was explained by an interaction between female immunoreactive fGCM  
380 level and nest cover (fGCM  $\times$  nest cover interaction:  $b = 0.002$ ;  $SE = 0.001$ ;  $t = 2.014$ ;  $p < 0.05$ ;  $df = 155$ ;  
381  $R^2_{adj.} = 0.03$ ; Table 2, Fig. 2). Longer telomeres were associated with high nest concealment for females  
382 with high fGCM while the opposite trend was observed for low fGCM females. Importantly, we did not  
383 detect a significant association between telomere length and female minimum years of maternal  
384 experience, and female body condition was likewise not significantly associated with telomere length  
385 (Table 2).

386

## 387 **DISCUSSION**

388 Consistent with the hypothesized role of GCs in adaptively regulating escape responses to the habitat-  
389 specific risk of detection by predators, we found that individuals with high immunoreactive fGCM  
390 levels had the highest hatching success in nests offering little cover, whereas females with low fGCM  
391 profiles had the highest hatching success in well-covered nests (Fig. 1). Thereby, variation in nest-site  
392 preferences may facilitate the coexistence of different baseline GC levels in populations subjected to  
393 habitat-specific risks of attack by predators (Rivers et al. 2014), cautioning against uncritically assuming  
394 a uniformly negative association between baseline GC levels and fitness (Bonier et al. 2009b). While we  
395 observed no link between telomere length and a proxy of age in female eiders, we found that shorter  
396 telomeres were associated with high fGCM in nest sites with little shelter and with low fGCM in well-  
397 concealed ones (Fig. 2). Interestingly, this habitat-associated pattern of telomere dynamics may imply a  
398 potential cost of reproduction, since the females with shorter telomeres also had higher reproductive  
399 success. This result agrees with that of a recent study on common terns (*Sterna hirundo* Linnaeus,

400 1758), showing that individuals with short telomeres had higher reproductive performance (Bauch et al.  
401 2013).

402

403 The interactive effects of immunoreactive faecal GCs and nest cover on hatching success are consistent  
404 with the presence of habitat-specific antipredator strategies. Cross-species comparisons have shown that  
405 ecologically similar co-inhabiting species may show contrasting escape tactics when at risk from  
406 predation (Lima 1990; Wirsing et al. 2010). Thus, some species always select dense vegetation because  
407 of the protection it provides against predators ('cover-dependent escape tactic'), whereas others prefer a  
408 clear path of escape to the air ('aerial escape tactic'; Lima 1990). These different antipredatory tactics  
409 may also be present within species (Cuadrado et al. 2001; Thaker et al. 2010; Brink et al. 2011). In the  
410 case of eiders, evidence suggests that well-concealed nest-sites may be associated with a reduced  
411 risk of detection by predators but also potentially higher costs of escape, favouring cover-dependent  
412 escape tactics at such nest-sites. First, predation pressure (number of killed females/nesting attempt) is  
413 lower and female survival is higher on forested islands than on open ones (Ekroos et al. 2012). Second,  
414 it has been experimentally shown that the risk of egg predation decreases with increasing nest cover (Öst  
415 et al. 2008), suggesting that concealed nests may attract less attention from visually hunting predators.  
416 Third, the presumed benefit of immobility in the presence of predators ('freezing') in densely vegetated  
417 habitat is enhanced by the fact that once detected by a predator, dense vegetation may prevent successful  
418 escape (Öst and Steele 2010). In contrast, the optimal strategy in poorly concealed nests may be to rely  
419 on early escape from predators in anticipation of the higher risk of predator detection (Amat and Masero  
420 2004; Albrecht and Klvaňa 2004).

421

422 D'Alba et al. (2011) argued that exposed nest sites are occupied by female eiders of lower phenotypic  
423 quality and that the effects of GCs on hatching success appear to vary independent of nest shelter. In line

424 with these conclusions, our own previous work indicated a consistently negative association between  
425 immunoreactive fGCMs and hatching success in eiders (Jaatinen et al. 2013). However, our present  
426 study showed that a detailed examination of nest-site preferences profoundly changes these conclusions,  
427 providing a more nuanced view of the interrelationships between baseline fGCM, reproductive success  
428 and breeding habitat. This discrepancy may relate to the orchestrating role of fGCM in simultaneously  
429 affecting both reproductive physiology and antipredator behaviours; both of which are intimately linked  
430 to reproductive success (Crossin et al. 2016). GCs are associated with the anticipation or awareness of  
431 danger when confronted with the threat of predation (e.g., Korte 2001; Cockrem and Silverin 2002).  
432 Thus, GCs enhance vigilance behavior (e.g., Romero and Butler 2007) and causally affect flight  
433 initiation distance (Thaker et al. 2010). In incubating female eiders, flight initiation distance increases  
434 with the magnitude of the acute handling-induced corticosterone (a major GC in birds) response  
435 (Seltmann et al. 2012), while handling-induced corticosterone responsiveness decreases with increasing  
436 nest cover (Schmidt et al. 2009; Jaatinen et al. 2014). This earlier work also suggests a positive link  
437 between enhanced GC responsiveness and reproductive success under high risk of predation (Jaatinen et  
438 al. 2014). Our result showing a positive correlation between high fGCM levels and hatching success in  
439 poorly concealed nests corroborates this notion, while also suggesting a nest-cover dependent nature of  
440 the association. Although the mechanisms underlying a positive association between corticosterone  
441 secretion and fitness under high risk of predation remain obscure, it is perhaps pertinent that  
442 minimization of incubation time may be particularly beneficial in microhabitats offering limited  
443 protection from predators. Thus, experimental evidence suggests that corticosterone shortens incubation  
444 time in birds (Schmidt et al. 2009) and female eiders having a long flight initiation distance,  
445 characterized by higher stress-induced corticosterone secretion, have a shorter incubation period  
446 (Seltmann et al. 2012).

447

448 We were unable to detect any effect of our proxy for female age on telomere length in adult female  
449 eiders. This result could be an artefact of selective disappearance, i.e. individuals with shorter telomeres  
450 disappearing earlier from the population (van de Pol and Verhulst 2006). However, body condition, a  
451 correlate of life expectancy (Ekroos et al. 2012) showing individual repeatability between years  
452 (Jaatinen and Öst 2011), was also not significantly associated with telomere length (Table 2). This lack  
453 of a relationship between telomere length and body condition adds credence to the possibility that  
454 telomere length may not be associated with age *per se* in adult eiders. Likewise, a lack of an association  
455 between age and telomere length in adulthood has been found in some other long-lived birds (e.g.,  
456 Mizutani et al. 2009; Pauliny et al. 2012; Rattiste et al. 2015), although there are exceptions (e.g., Bize  
457 et al. 2009). In this study, we quantified relative rather than absolute telomere length which could  
458 potentially mask some between-individual differences in telomere length (see Young et al. 2013).  
459 Nonetheless, some studies on long-lived birds where absolute telomere length was quantified also failed  
460 to observe telomere shortening with age (e.g., Hall et al. 2004). Potentially, this lack of correlation may  
461 be attributed to lifelong persistence of active telomerase (Hausmann et al. 2007), a possibility  
462 warranting further investigation.

463

464 How can we reconcile the finding that individuals with short telomeres had higher breeding performance  
465 (Fig. 2) with the widely-held notion that individuals with longer telomeres, after controlling for any age  
466 effects, are of higher phenotypic quality (Pauliny et al. 2006; Le Vaillant et al. 2015)? However, as  
467 argued by Bauch et al. (2013), increased investment in reproduction may induce telomere loss, and this  
468 effect may become particularly pronounced if some individuals consistently perform better than others  
469 throughout their lives. Viewed in this light, individual variation in telomere length may implicate long-  
470 term cumulative reproductive costs, rather than merely reflecting the current reproductive burden  
471 (Bauch et al. 2013). This argument may also be valid in the case of eiders. For example, female identity

472 explains more than half of the variation in nest fate (i.e., at least one egg hatched vs. all eggs unhatched)  
473 (Öst and Steele 2010). Nevertheless, some open questions remain, the solution of which will require  
474 further, preferably experimental evaluation. One particular challenge relates to the observation that  
475 females with low fGCM levels in covered nests had shorter telomeres, given the alleged role of  
476 glucocorticoids in accelerating telomere loss (Hausmann and Marchetto2010). Although this effect may  
477 seem small (Fig. 2), it deserves further longitudinal study, because our current, cross-sectional analysis  
478 inevitably only provides a snapshot of telomere length, and thus it cannot unveil the underlying  
479 complexity of the telomere shortening and restoration process (Monaghan and Hausmann 2006).

480

481 Because behavioural reactivity and physiological stress coping mechanisms are tightly linked (Koolhaas  
482 et al. 1999), the same forces are likely to maintain variation in both set of traits in the population.  
483 Theory predicts that individuals with higher GCs should perform better under unpredictable  
484 environmental conditions, whereas low GC levels are favoured under stable conditions (Cockrem 2005).  
485 These fundamental context-dependent differences in optima could serve to maintain phenotypic  
486 variability in the population under temporally or spatially fluctuating selection pressures. In line with  
487 this general expectation, our study demonstrates that short-term reproductive output tends to become  
488 equalized for individuals with different stress profiles if individually repeatable habitat choices are taken  
489 into account. Nest cover is an important habitat feature especially for ground-nesting birds, as it can  
490 influence adult and egg predation risk (Martin 1993), offer variable thermal conditions (e.g., Kilpi and  
491 Lindström 1997) and thereby influence habitat predictability. Our current results showed that females  
492 with high fGCM have higher reproductive success but shorter telomeres in open compared to concealed  
493 nest sites. Since concealed nest sites are less exposed to weather extremes (Kilpi and Lindström 1997;  
494 Fast et al. 2007) and predator attacks (Ekroos et al. 2012) and thereby likely to offer a more stable  
495 environment, our findings agree with the hypothesis that variation in environmental predictability can

496 promote the co-existence of different behavioural and physiological phenotypes within the same  
497 population (Cockrem 2005).

498

499 In summary, we have demonstrated that the relationships between breeding microhabitat, telomere  
500 length and reproductive success may differ depending on individual stress coping strategies in a wild  
501 population, subject to temporally and spatially varying predation pressure. Our results are consistent  
502 with adaptive adjustment of GC levels to match local environmental conditions, thereby tending to  
503 equalize fitness across nests of different concealment. Accordingly, our results may help to explain the  
504 considerable variation in nest concealment at the intraspecific level (Öst and Steele 2010). Here we have  
505 argued that this adjustment may be driven by threat-sensitive predation avoidance, where different  
506 behavioural tactics are favoured in contrasting nest microhabitats. However, since our study is  
507 necessarily correlational, causality remains to be demonstrated (but see D'Alba et al. 2011). Equally  
508 unclear at this point is whether female eiders with high reproductive success, incurring an apparent cost  
509 in terms of telomere shortening, also have shorter lifespan, i.e., whether they actually pay a cost of  
510 reproduction. In fact, circumstantial evidence suggests a positive relationship between fecundity and  
511 survival in this species (Yoccoz et al. 2002). To address these open questions, we encourage future  
512 longitudinal studies investigating within-individual relationships between stress physiology, fitness and  
513 telomere dynamics, preferably involving experimental manipulations of predation risk.

514

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526

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737 **Table1**

738 Model selection and GLMM (binomial error distribution, log link function and female identity as a  
 739 random factor) testing the effects of a set of independent variables on proportional hatching success  
 740 (P(hatch)).

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<b>Independent variable</b>	<b>Parameter estimate (b)</b>	<b>SE</b>	<b>Z value</b>	<b>p</b>	<b>N</b>
Minimum maternal experience (years)	0.036	0.065	0.555	0.58	421/316
Body condition	-0.118	0.174	-0.675	0.50	408/309
Year	0.359	0.215	1.669	0.09	423/317
<b>Hatching date</b>	<b>-0.226</b>	<b>0.025</b>	<b>-8.981</b>	<b>&lt; 0.001</b>	<b>423/317</b>
<b>Island-specific predation</b>	<b>-4.812</b>	<b>1.433</b>	<b>-3.358</b>	<b>&lt; 0.001</b>	<b>423/317</b>
<b>Nestcover</b>	<b>2.485</b>	<b>0.933</b>	<b>2.662</b>	<b>&lt; 0.001</b>	<b>423/317</b>
<b>fGCM (ng/g)</b>	<b>0.007</b>	<b>0.003</b>	<b>2.434</b>	<b>0.01</b>	<b>423/317</b>
<b>fGCM × nestcover</b>	<b>-0.018</b>	<b>0.004</b>	<b>-4.187</b>	<b>&lt; 0.001</b>	<b>423/317</b>

742 The final model (in **bold**) was selected by removing all non-significant variables ( $\alpha = 0.05$ ). Variables  
 743 included in the initial model included the *a priori* defined two-way interaction between fGCM and nest  
 744 cover. Abbreviations: fGCM –faecal glucocorticoid metabolites; df-degrees of freedom; N:  
 745 observations/unique individuals; SE-standard error. Sample sizes differ because data were not available  
 746 for all independent variables.

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749 **Table 2**

750 Model selection and linear model (LM) testing the effects of the set of independent variables on  
751 telomere length.

<b>Independent variable</b>	<b>Parameter estimate (b)</b>	<b>SE</b>	<b>t value</b>	<b>df</b>	<b>p</b>
Minimum maternal experience (years)	0.008	0.011	0.697	154	0.49
Body condition	0.017	0.043	0.404	136	0.69
<b>fGCM (ng/g)</b>	<b>-0.001</b>	<b>0.001</b>	<b>-1.274</b>	<b>155</b>	<b>0.20</b>
<b>Nest cover</b>	<b>-0.365</b>	<b>0.310</b>	<b>-1.179</b>	<b>155</b>	<b>0.24</b>
<b>fGCM × nestcover</b>	<b>0.002</b>	<b>0.001</b>	<b>2.014</b>	<b>155</b>	<b>&lt;0.05</b>

752 The final model (in **bold**) was selected by removing all non-significant variables ( $\alpha = 0.05$ ). Variables  
753 included in the initial model included the *a priori* defined two-way interaction between fGCM and nest  
754 cover. Abbreviations: fGCM: faecal glucocorticoid metabolites; *df*: degrees of freedom; *SE*: standard  
755 error.

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760 Figure legends

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762 Fig. 1. Proportional hatching success is affected by an interaction between proportional nest  
763 cover and immunoreactive faecal glucocorticoid metabolite (fGCM, ng/g) level, so that the hatching  
764 success of females with low fGCM (mean - 1SD, solid line, L, black dots) positively correlates with  
765 increasing proportional nest cover, whereas females with high fGCM (mean + 1SD, dotted line, H, open  
766 circles) tend to have lower hatching success in concealed nests. Females with intermediate fGCM  
767 concentrations (mean, dashed line, M, grey dots) exhibit an intermediate response.

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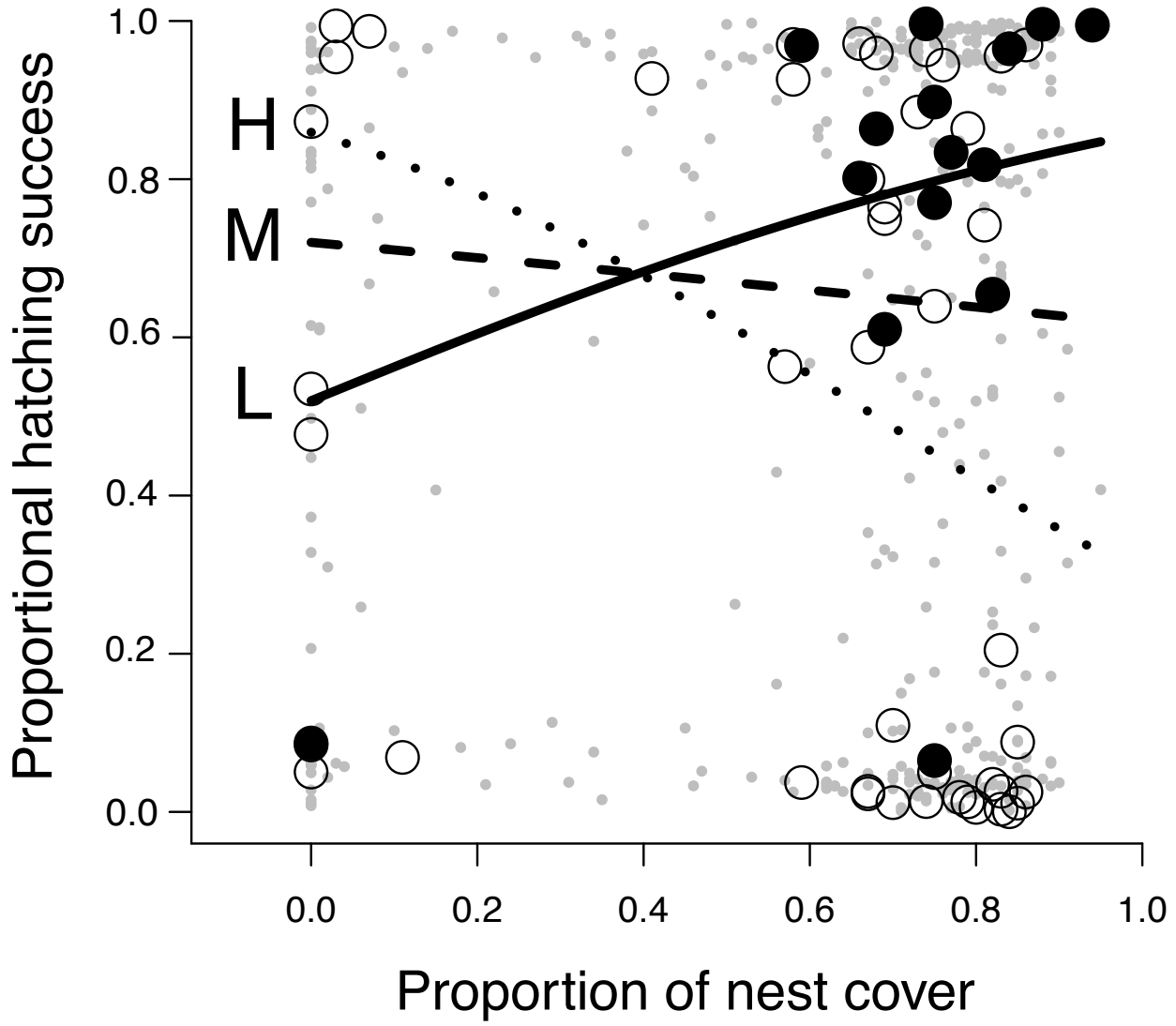
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771 Fig. 2. Female telomere length is connected to nest cover, but this relationship is modulated by the  
772 immunoreactive faecal glucocorticoid metabolite (fGCM, ng/g) level. Telomere length is positively  
773 associated with proportional nest cover for females with high fGCMs (mean+1SD, dotted line, H, open  
774 circles), whereas for nesting females with low fGCM (mean-1SD, solid line, L, black dots) this  
775 association is negative. Females exhibiting intermediate fGCM levels (mean dashed line, M, grey dots)  
776 show an intermediate response.

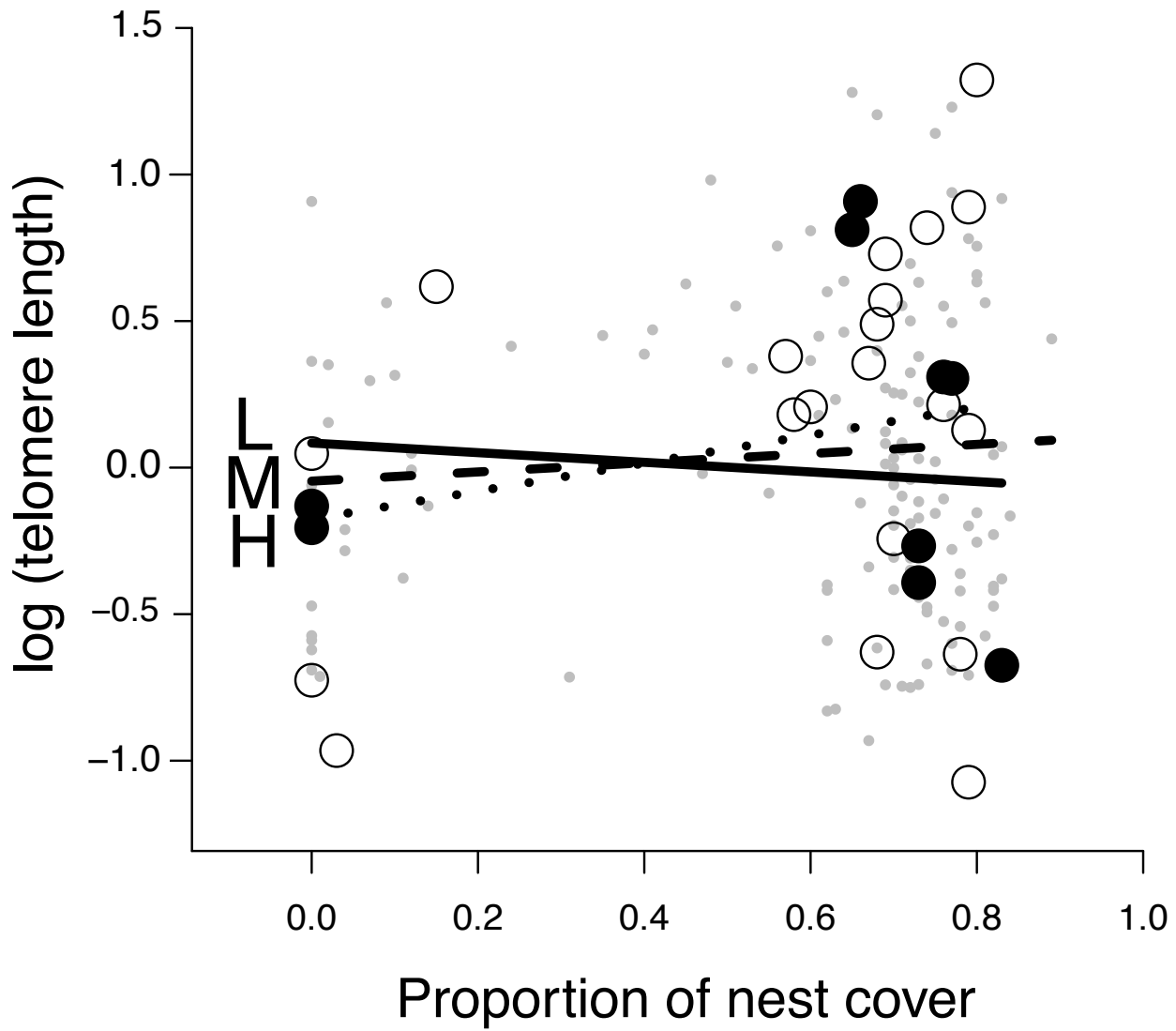
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784 **Supplementary Table1**

785 Nest fate during 2009-2011. Nest fate was categorized as either depredated, abandoned or successful (at  
 786 least one successfully hatched offspring) (see Methods).

<b>Year</b>	<b>Depredatednest s</b>	<b>Abandoned nests</b>	<b>Successful nests</b>	<b>Total number of known-fate nests</b>	<b>Total number of nests</b>
<b>2009</b>	22 (14.76%)	7 (4.70%)	120 (80.54%)	149	165
<b>2010</b>	43 (31.85%)	20 (14.81%)	72 (53.33%)	135	173
<b>2011</b>	67 (40.60%)	6 (3.64%)	92 (55.75%)	165	197

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