





Article

Late Onset of Estrogen Therapy Impairs Carotid Function of Senescent Females in Association with Altered Prostanoid Balance and Upregulation of the Variant ER α 36

Tiago Januário Costa ^{1,2,3,4}, Francesc Jiménez-Altayó ², Cinthya Echem ¹,
Eliana Hiromi Akamine ¹, Rita Tostes ⁴ , Elisabet Vila ², Ana Paula Dantas ^{3,*}  and
Maria Helena Catelli de Carvalho ¹

¹ Department of Pharmacology, Institute of Biomedical Sciences, University of Sao Paulo, Sao Paulo 05508-900, Brazil; tiago.januario@uol.com.br (T.J.C.); cinthya.echem@gmail.com (C.E.); eliakamine@usp.br (E.H.A.); mhccarvalho@uol.com.br (M.H.C.d.C.)

² Facultat de Medicina, Departament de Farmacologia, Terapèutica i Toxicologia, Institut de Neurociències, Universitat Autònoma de Barcelona, 08193 Bellaterra, Spain; Francesc.Jimenez@uab.cat (F.J.-A.); elisabet.vila@uab.cat (E.V.)

³ Group of Atherosclerosis and Coronary Disease, Institut Clinic del Torax, Institut d'Investigacions Biomèdiques August Pi I Sunyer (IDIBAPS), 08036 Barcelona, Spain

⁴ Pharmacology Department, Ribeirao Preto Medical School, University of Sao Paulo, Sao Paulo 14049-900, Brazil; rtostes@usp.br

* Correspondence: adantas@clinic.cat

Received: 22 August 2019; Accepted: 4 October 2019; Published: 8 October 2019



Abstract: Recent analysis of clinical trials on estrogen therapy proposes the existence of a therapeutic window of opportunity for the cardiovascular benefits of estrogens, which depend on women's age and the onset of therapy initiation. In this study, we aimed to determine how vascular senescence and the onset of estrogen treatment influence the common carotid artery (CCA) function in senescent and non-senescent females. Ovariectomized female senescence-accelerated (SAMP8) or non-senescent (SAMR1) mice were treated with vehicle (OVX) or 17 β -estradiol starting at the day of ovariectomy (early-onset, E₂E) or 45 days after surgery (late-onset, E₂L). In SAMR1, both treatments, E₂E and E₂L, reduced constriction to phenylephrine (Phe) in CCA [(AUC) OVX: 193.8 \pm 15.5; E₂E: 128.1 \pm 11.6; E₂L: 130.2 \pm 15.8, p = 0.004] in association with positive regulation of NO/O₂⁻ ratio and increased prostacyclin production. In contrast, E₂E treatment did not modify vasoconstrictor responses to Phe in OVX-SAMP8 and, yet, E₂L increased Phe vasoconstriction [(AUC) OVX: 165.3 \pm 10; E₂E: 183.3 \pm 11.1; E₂L: 256.3 \pm 30.4, p = 0.005]. Increased vasoconstriction in E₂L-SAMP8 was associated with augmented thromboxane A₂ and reduced NO production. Analysis of wild-type receptor alpha (ER α 66) expression and its variants revealed an increased expression of ER α 36 in E₂L-SAMP8 in correlation with unfavorable effects of estrogen in those animals. In conclusion, estrogen exerts beneficial effects in non-senescent CCA, regardless of the initiation of the therapy. In senescent CCA, however, estrogen loses its beneficial action even when administered shortly after ovariectomy and may become detrimental when given late after ovariectomy. Aging and onset of estrogen treatment are two critical factors in the mechanism of action of this hormone in CCA.

Keywords: estrogen; aging; estrogen receptor variants; cerebrovascular function; menopause; hormone replacement therapy

1. Introduction

The female aging process and associated disorders may be linked to changes in the hormonal milieu [1]. Of all aging-related disorders, cardiovascular disease is the leading cause of morbidity and mortality in older women [2]. Although estrogen modulates several pathways that are closely associated with aging, it remains unknown whether estrogen can delay cardiovascular senescence, or how estrogen-mediated responses occur in a senile vasculature.

In 1985, the observational study Nurses' Health Study reported that estrogen therapy reduces the risk of cardiovascular disease in postmenopausal women [3]. In agreement with these clinical data, many experimental studies demonstrated that estrogen has beneficial effects in several signaling pathways in the vascular system [4–9]. However, double-blind, randomized clinical trials—such as Woman's Health Initiative (WHI) and Heart and Estrogen/progestin Replacement Study (HERS I and II)—showed that estrogen therapy increases cardiovascular risk, and more specifically the risk for stroke in postmenopausal women [10,11]. Stroke is the leading cause of prolonged disability and the third leading cause of death among women [2]. Despite this, there is still a concerning gap in the comprehension of the mechanisms involved in the estrogen regulation of the cerebrovascular function of aged women.

Among the explanations for the discrepancies between clinical trial and observational studies, is the fact that women included in these trials were on average 60 years of age when they started receiving estrogen therapy, typically more than 10 years after the onset of menopause. While in the first observational study, women were younger, approximately 50 years of age, and closer to menopause when they initiated hormonal therapy. Detailed analysis by aging groups suggests that early initiation of estrogen therapy produces more favorable results than the average of late-onset used in most trials [12]. These analyzes led scientists to conceive the so-called “timing hypothesis”, which relies on the concept that estrogen has beneficial effects if taken before, or close to, the onset of menopause when the detrimental aging effects or lack of estrogen in the vasculature have not yet been established [13]. In 2016, the clinical trial ELITE (Early versus Late Intervention Trial with Estradiol) strengthened this theory by showing that estrogen therapy is associated with less progression of subclinical atherosclerosis only when it is initiated within six years after menopause onset [14]. Aging per se is known to cause a series of alterations in the endogenous mechanisms that control vascular function, leading to subsequent increased risk of cerebrovascular disease [15]. Moreover, vascular aging can alter estrogen-mediated signaling pathways [16] and, as a consequence, estrogen therapy may become harmful rather than beneficial to the vasculature.

The classical effects of estrogen in the vasculature are mediated via interactions with two subtypes of nuclear receptors belonging to a superfamily of hormone-inducible transcription factors (ER α and ER β), and by a seven-transmembrane G protein-coupled ER, termed GPER, associated to activate rapid signaling cascades after estrogen binding (for review, see [17]). Although the three estrogen receptors are known to promote cardiovascular effects, ER α is the most associated with the beneficial effects of estrogen in the endothelial and smooth muscle cells [17]. Furthermore, molecular studies have identified and cloned alternative splicing variants of ER α , which have been associated with modifications in the activation of the major signaling pathways by this receptor [17–19]. These splicing variants of ER α dimerize with the wild-type 66Kd receptor (ER α 66), and may serve as a competitive inhibitor of ER α 66 binding to DNA [18,19] and, therefore, an increase of ER α splicing may interfere with ER α 66 transcriptional activity function. Upregulation of this variant has been associated with rapid estrogen and anti-estrogen signaling in gynecological cancer [20], although little is known about their correlation with cardiovascular disease.

In this study, we interrogate the complicated relationship between the timing of estrogen therapy and senescence in the protection or detriment of female cerebrovasculature. The common carotid arteries (CCA) carry the main supply of blood to the brain, and inadequate circulation of blood through these vessels may result in cerebrovascular events. We hypothesized that vascular senescence and the onset of estrogen treatment modifies arterial function in association with changes in the expression

of variants of classical ER α . For this purpose, we analyzed vascular reactivity of the CCA from ovariectomized (OVX) senescence-accelerated prone (SAMP) mice following early- or late-onset of estrogen treatment. Our previous studies have established OVX-SAMP as a model that closely mimics menopause in women, as it can concomitantly determine vascular changes by estrogen withdrawal (or replacement) in a senescent environment [21].

2. Material and Methods

2.1. Animal Models and Hormonal Treatment

Female senescence-prone inbred (SAMP8, aging model $n = 80$) and senescence-resistant inbred strain (SAMR1, young control $n = 80$) mice, were obtained from the breeding stock at Parc Científic de Barcelona. The senescence-accelerated mouse model was developed as a result of selective inbreeding of mice showing a phenotype of severe exhaustion (SAM-prone) and inbreeding of a normal phenotype (SAM-resistant). SAMP strains are known to manifest spontaneously various pathobiological phenotypes, including vascular senescence. SAM offers several advantages in aging cardiovascular research as it ages fast and predictably, allowing the execution of experimental work in a convenient and standard time course [21]. The animals were housed at the Animal Facility of the University of Barcelona according to institutional guidelines (constant room temperature 22 °C, 12-h light/dark cycles, 60% humidity, standard mice chow, and water ad libitum). All the procedures used in this study were approved and performed following the guidelines of the Ethics Committee of the University of Barcelona (Protocol 272/12), the Institute of Biomedical Sciences, University of São Paulo (ICB-USP—Protocol 64, page 20, book 3. 27.05.2014), and in agreement with the *Guide for the Care and Use of Laboratory Animals* published by the US National Institute of Health (NIH Publication No.85-23, revised 1996). At six months of age, SAMR1 and SAMP8 mice were ovariectomized under controlled inhalant anesthesia with isoflurane (4% induction and 1.5–2% maintenance). After ovariectomy, mice were divided into three groups: (1) ovariectomized treated with vehicle (OVX); (2) early onset of estrogen treatment, initiated in the first day of ovariectomy (E₂E); and (3) late onset of estrogen treatment, initiated 45 days after ovariectomy (E₂L). Cyclic estrogen therapy (5 μ g/kg of 17 β -estradiol diluted in mineral oil), was administered by subcutaneous injections every third day in order to provide a more physiological hormonal milieu [16]. The efficacy of ovariectomy and estrogen treatment was determined by the uterine weight and plasma estrogen concentrations. Sixty days following the ovariectomy, all mice were euthanized with sodium pentobarbitone (85 mg/Kg, I.P.) and the CCAs were dissected and kept in ice-cold physiological salt solution and prepared for different experiments mainly as described [22].

2.2. Vascular Function Study

Segments (2 mm) of CCA with intact endothelium were mounted on an isometric wire myograph (model 410 A; J.P. Trading, Aarhus, Denmark), as previously described [22]. The myograph was filled with modified Krebs solution ((in mM): NaCl 130; NaHCO₃ 14.9, KCl 4.7, KH₂PO₄ 1.18, MgSO₄ 1.17; CaCl₂.2H₂O 1.56, EDTA 0,026 and glucose 5.5), and kept at 37 °C, 95% O₂ and 5% CO₂. After 60 min (min) of equilibration, CCA segments were stimulated three times (10-min interval) with a KCl 60 mM solution until the contraction reached a stable plateau (~15 min). After washout and return to a stable baseline, consecutive concentration–response curves—with acetylcholine (ACh, 10⁻¹⁰ to 3 \times 10⁻⁵ M), phenylephrine (Phe, 10⁻⁹ to 10⁻⁵ M), and sodium nitroprusside (SNP, 10⁻¹⁰ to 3 \times 10⁻⁵ M)—separate by washout and 30 min intervals, were performed in the absence or the presence of several inhibitors. In some experiments, contraction to the thromboxane A₂ analog (U46619, 10⁻⁹ to 10⁻⁵ M), instead of Phe was performed. Vasodilation to ACh and SNP were performed in U46619 (10⁻⁷ M) pre-contracted vessels. The contribution of the different endothelium-derived factors to the vascular responses was determined by treating isolated CCA segments with one of the following inhibitors: (1) nonselective nitric oxide synthase (NOS) inhibitor N ω -nitro-L-arginine methyl ester (L-NAME; 10⁻⁴ M); (2) O₂⁻

scavenger (Tempol, 10^{-5} M); (3) non-selective COX inhibitor (Indomethacin, 10^{-6} M); (4) selective COX-1 inhibitor (SC560, 10^{-5} M); or (5) selective COX-2 inhibitor (NS398, 10^{-6} M). Experiments were done in parallel with segments from the same animal. Contractions to Phe and U46619 are shown as a percentage (%) of the contractile response to 60mM KCl. Relaxations to ACh and SNP were expressed as the percentage (%) of U46619 precontraction. The area under the concentration–response curve (AUC) was used as an overall measure of cumulative responses induced by agonists in the presence of vehicle or specific inhibitors.

2.3. Quantitative Real-Time PCR (qPCR) for Detection of COX and eNOS Expression

Total RNA was isolated from CCA using TRIZOL[®] reagent (Sigma-Aldrich, St Louis, MI, USA) according to the manufacturer's instructions. The amount of mRNAs encoding the COX isoforms (COX-1 and COX-2) and downstream enzymes (PGI2S and TXA2S), as well as eNOS, were quantified by qPCR based on SYBR[®] Green fluorescence (Applied Biosystems, Carlsbad, CA, USA). β -actin was used as an internal control. Primer sequences are described in Table S3 (Major Resources Tables). qPCR reactions were performed, recorded, and analyzed using the Corbett Research system (Corbett Life Sciences, Sydney, Australia). The conditions for qPCR were as follows: 95 °C for 2 min, 40 cycles of 95 °C for 15 s (s), and 60 °C for 1 min. Cycle threshold (Ct) values obtained for each gene were referenced to β -actin (Δ Ct) and converted to the linear form using the term $2^{-\Delta\Delta Ct}$ as a value directly proportional to the copy number of complementary DNA and the initial amount of mRNA.

2.4. Measurement of Prostanoids Release from Vascular Segments

The release of prostanoids following Phe stimulation was determined by enzyme immunoassay (ELISA). Aliquots of Krebs–Henseleit solution were collected after the concentration–response curves to Phe and kept at room temperature for 30 min for complete hydrolysis of prostacyclin (PGI₂) and thromboxane A₂ (TXA₂) into their stable metabolites, 6-Keto-PGF_{1 α} and TXB₂, respectively. Analysis of specific contributions of COX-1 and COX-2 to prostanoids release was carried out in organ baths containing vehicle-treated CCA or CCA treated with selective inhibitors of COX-1 or COX-2. Levels of 6-Keto-PGF_{1 α} and TXB₂ were assessed in duplicate and calculated according to the ELISA kit manufacturer (Cayman Chemical, Ann Arbor, MI). The results were expressed as pM for vehicle-treated CCA and as Log₂ of fold change in arteries treated with selective inhibitors of COX-1 (SC396) or COX-2 (NS398).

2.5. Western Blot Analysis

SDS-PAGE resolved equal amounts of protein from each CCA sample (30 μ g) on 4–12% gels and electroblotted onto nitrocellulose membranes. Membranes were incubated for 1 h (h) with casein-based blocking buffer, following overnight incubation at 4 °C with specific primary antibodies as follows: polyclonal rabbit anti-ER α 36, 1:1000 (Alpha Diagnostic, San Antonio, TX, USA); polyclonal rabbit anti-ER α 66 1:1000 (Santa Cruz Biotechnology, Sta Cruz, CA, USA). After washes, membranes were incubated with specific horseradish peroxidase-labeled secondary antibodies in phosphate-buffered saline (PBS) containing 1% casein-based buffer. Following additional washes, the chemiluminescent signal was visualized by LAS4000 imaging system (GE Healthcare, Chicago, IL, USA). All membranes were reblotted using a mouse monoclonal antibody anti- α -actin (1:2000; Agilent Dako, Sta Clara, CA, USA) as the loading control. Densitometric analyses of western blots were performed using ImageJ software, and data were normalized to corresponding values of α Actin densitometry.

2.6. Methylation Status of Gene Encoding ER α and Expression of Splicing Variants

The methylation status of different CpG enriched regions of the gene encoding ER α was determined by methylation-sensitive qPCR, as described [16]. Following mock digestion or enzymatic digestion with HpaII (methylation-sensitive) or MspI (no methylation-sensitive), resulting DNA was amplified using qPCR with Power SYBR[®] Green master mix, as described by the manufacturer (Applied

Biosystems). PCR primers were designed using Methyl Primer Software (Applied Biosystems) to amplify five distinct CpG enriched regions of ER α gene or to amplify a region that is devoid of any of the restriction sites of the enzymes used in the design of the experiment, as an internal control (Table S4, Major Resources Tables). Methylation at CpG sites prevents HpaII, but not MspI, digestion and allows the amplification of the fragment, resulting in a low cycle threshold (Ct) value. In contrast, if the CpG island is not methylated, HpaII cleaves DNA and prevents the amplification of the fragment, resulting in higher Ct values. The percentage (%) of methylation of a given site was determined from the change in Ct value (Δ Ct) of methylated-sensitive and mock digested samples. Taking into account the fundamental principle that each successive round of PCR amplification results in approximately a 2-fold increase in the amount of product, the % Methylation was calculated as $100 \times (2^{-0.7(\Delta Ct)})$. Each sample was analyzed in duplicate. The expression of wild type ER α and C-terminally truncated ER α product (CTERP) was determined in mice CCA by SYBR green-based qPCR. For relative quantification, the amount of CTERP splicing expressed in mice CCA was normalized by the expression of the very same wild type gene (ER α) and the very same sample, as previously described.

2.7. Data Analysis and Statistics

Data are expressed as mean \pm SEM of the number (n) of mice indicated in the figure legends. The extra sum-of-squares F determined the differences in the fit of concentration–response curves in all groups. The AUC was calculated for individual contractile or relaxing concentration–response curves and expressed as arbitrary units. The contribution of different endothelium-derived factors to Phe-induced contractions was calculated by subtracting the AUC for Phe curves in the presence of inhibitors from the AUC for control Phe curves (Δ Change). Brown–Forsythe testing determined equality in variances among experimental groups (OVX, E₂E, and E₂L). One-way ANOVA analyzed the dependence of data on the onset of estrogen therapy (none, early, or late) in SAMR1 or SAMP8 with Bonferroni’s post-test. The analysis was carried out using the Prism software (GraphPad Software V7.0, San Diego, CA, USA), and statistical significance was accepted at $p < 0.05$.

3. Results

3.1. Basic Parameters for Ovariectomy and Estrogen Treatment Efficacy

Ovariectomy resulted in reduced uterine weight and plasma 17 β -estradiol concentrations in SAMR1 and SAMP8 female mice (data not shown). Those parameters were restored by both early (E₂E) and late (E₂L) onset of 17 β -estradiol treatment (Table 1).

Table 1. Basic parameters (uterus weight and estrogen plasma concentration).

	SAMR1			SAMP8		
	OVX	E ₂ E	E ₂ L	OVX	E ₂ E	E ₂ L
Uterus wet weight (mg/cm)	30.1 \pm 4.5	49.5 \pm 5.4 *	56.1 \pm 7.1 *	14.0 \pm 1.7	42.4 \pm 5.7 *	40.3 \pm 5.5 *
Uterus dry weight (mg/cm)	7.9 \pm 1.1	12.3 \pm 1.0 *	13.9 \pm 1.6 *	4.3 \pm 0.8	12.1 \pm 1.8 *	11.0 \pm 1.2 *
17β-estradiol (pg/mL)	2.1 \pm 0.5	4.2 \pm 0.9 *	7.0 \pm 1.2 *	1.2 \pm 0.3	9.8 \pm 1.4 *	7.4 \pm 0.9 *

Values are means \pm SEM of samples from 10 animals per group. The uterus weight was normalized by tibia length (mg tissue/cm tibia). The analysis was obtained in untreated ovariectomized mice (OVX), and ovariectomized mice under early-onset (E₂E) or late-onset of 17 β -estradiol treatment (E₂L). * $p < 0.05$ vs. OVX.

3.2. Early- and Late-Onset Estrogen Treatments Promote Different Vasoconstrictor Effects in Response to Adrenergic Stimulus in SAMR1 and SAMP8 Ovariectomized Female Mice

To determine how the onset of estrogen therapy affects the vascular function of senescent and non-senescent females, we tested the responses of CCA to different vasoactive agents. All agonists studied induced concentration-dependent responses in CCA rings in either group. In SAMR1, treatments with either E₂E or E₂L reduced the contractile response to Phe, compared to untreated OVX (Figure 1A), as demonstrated by the smaller area under the curves (Figure 1C). In SAMP8, responses to Phe after E₂E treatment were similar to those observed in the OVX group. However, E₂L treatment increased Phe vasoconstriction when compared to the OVX group (Figure 1B,C). On the other hand, there were no differences in the vasoconstrictor responses to the thromboxane A₂ analog U46619 (Figure S1). Neither did we observe significant differences in vasodilatation responses to both endothelium-dependent (ACh) and endothelium-independent (SNP) agents (Figure S2).

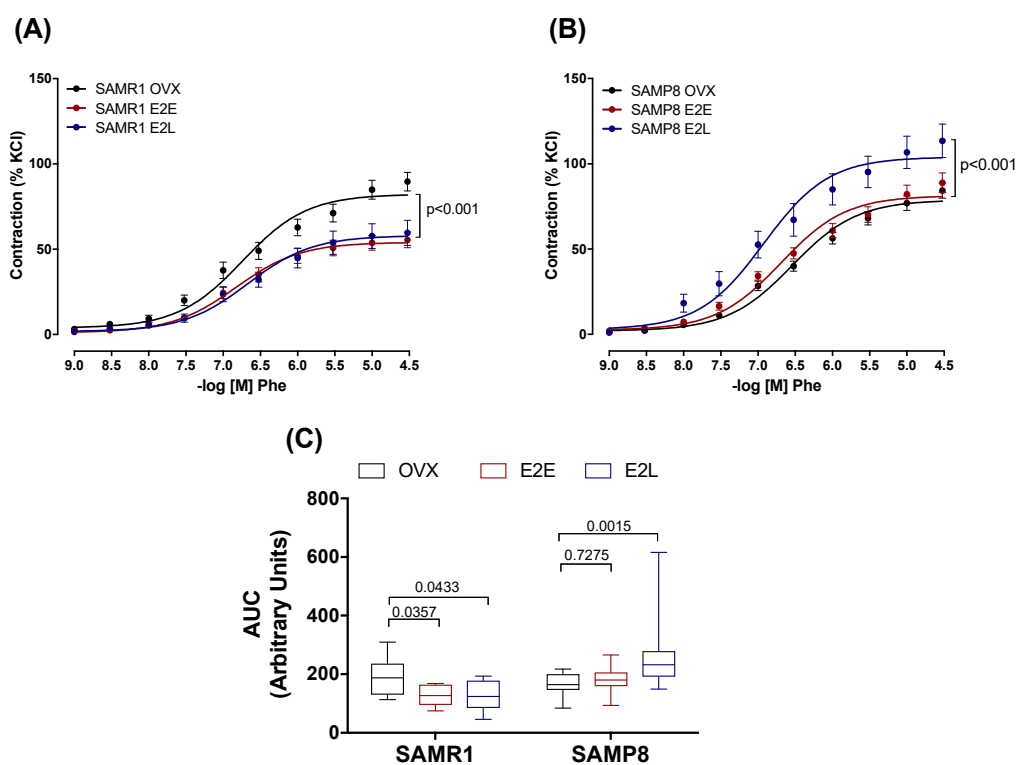


Figure 1. Phenylephrine-induced contractions. Concentration–effect curves to Phenylephrine (Phe) determined in endothelium-intact common carotids arteries from SAMR1 (A) and SAMP8 (B) female mice. Curves were obtained in vessels from untreated ovariectomized mice (OVX), and ovariectomized mice receiving early-onset (E₂E) or late-onset (E₂L) of 17 β -estradiol treatment. Differences in vascular responses are expressed as the area under the curve (AUC) (C). Each point of the curve and bar graphs represents the mean \pm SEM from 10–18 independent experiments. The extra sum-of-squares F. One-way ANOVA determined differences in the fit of concentration–response curves analyzed the dependence of data on the onset of estrogen therapy (none, early, or late) in SAMR1 or SAMP8 with Bonferroni’s post-test. *p*-values and comparisons are expressed next to the curves and on top of bar graphs. Significance is considered when *p* < 0.05.

3.3. Estrogen Differently Regulates the Contribution of NO/Superoxide Anion (O₂⁻) Pathways to Phe Contraction in Non-Senescent (SAMR1) and Senescent (SAMP8) Carotids

Inhibition of NO production by L-NAME increased the vasoconstrictor responses to Phe in all groups of SAMR1 (Figure 2A) and SAMP8 (Figure 2B) female mice. The Δ Change in the AUC (Figure 2C) shows that there is a more significant contribution of NO to attenuate contractile responses in arteries of non-senescent females (SAMR1) treated with both E₂E and E₂L than in untreated

OVX-SAMR1. In SAMP8, the Δ Change in the AUC of E₂E was similar to untreated mice. In opposition, E₂L displayed a lower Δ Change in the AUC compared to untreated OVX-SAMP8, suggesting a smaller contribution of NO to Phe contraction in this group (Figure 2C). Variations in the contribution of NO to vascular reactivity could be related to changes in both NO production, either via eNOS upregulation or downregulation, and modulation of NO scavenging by O₂⁻. Analysis of mRNA expression showed an increase in eNOS expression in estrogen-treated groups in SAMR1, but not in SAMP8 (Figure 2D). The influence of O₂⁻ on Phe-induced contractions was assessed by performing contractile curves in the presence of O₂⁻ scavenger, tempol. Incubation with tempol decreased Phe contractions in CCA of non-senescent untreated OVX, without affecting vasoconstrictor responses in estrogen-treated SAMR1 (Figure S3). In senescent females, scavenging of O₂⁻ by tempol decreased contractions to Phe in all SAMP8 groups (Figure S3).

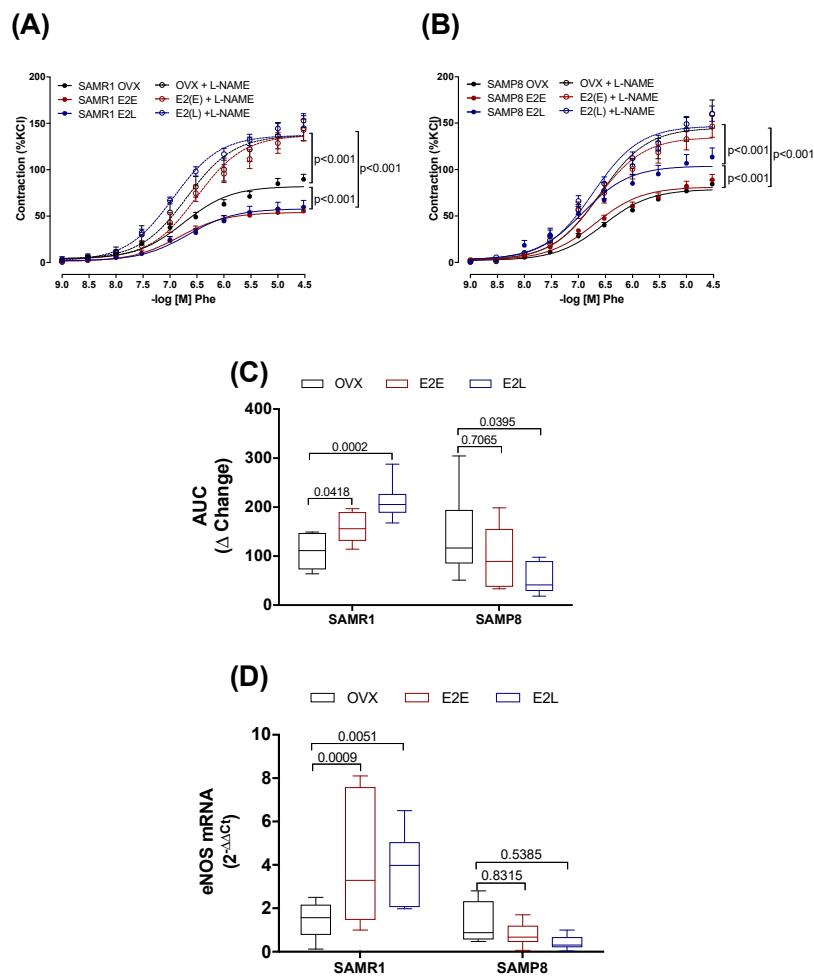


Figure 2. Role of NO in phenylephrine-induced contractions. Concentration–response curves to phenylephrine (Phe) in endothelium-intact common carotid artery from SAMR1 (A) and SAMP8 (B). Arteries were isolated from untreated ovariectomized mice (OVX), or OVX treated with early-onset of 17 β -estradiol treatment (E₂E) or late-onset 17 β -estradiol treatment (E₂L). Curves were obtained in the absence or the presence of L-NAME (10⁻⁴ M). The contribution of NO to Phe contraction is expressed as the change (Δ Change) in the area under the curve in L-NAME treated arteries relative to untreated arteries (C). mRNA levels of eNOS (D) normalized by β -actin mRNA and relative to the expression of untreated OVX mice. Each point represents the mean \pm SEM from 8–17 independent experiments. The extra sum-of-squares F determined differences in the fit of concentration–response curves. One-way ANOVA analyzed the dependence of data on the onset of estrogen therapy (none, early or late) in SAMR1 or SAMP8 with Bonferroni’s post-test. *p*-values and comparisons are expressed next to the curves and on top of bar graphs. Significance is considered when *p* < 0.05.

3.4. Senescence and Onset of Estrogen Therapy Play an Important Role in Regulating the Contribution of Vasodilator and Vasoconstrictor Prostanoids to Phe Contraction

We next sought to determine the contribution of prostanoids metabolites to the regulation of Phe contraction by estrogen, and how their contributions are affected by vascular senescence and therapy onset. Accordingly, CCAs were treated with the unspecific inhibitor of cyclooxygenases (COX) indomethacin. Pre-incubation with indomethacin decreased Phe contraction in estrogen-untreated OVX-SAMR1 and all three groups of SAMP8 (Figure S4), suggesting a predominance of COX-derived vasoconstrictor metabolites. The specific role of COX isoenzymes to Phe contractions was then determined by treating CCA with the selective inhibitors of COX-1 (SC560) or COX-2 (NS398). In SAMR1, COX-1, but not COX-2, inhibition reduced vasoconstrictor responses in untreated OVX, indicating a contribution of COX-1-derived vasoconstrictor prostanoids to Phe-induced contractions (Figure 3A,D). On the contrary, Phe contraction in E₂E was modulated by a COX-2-derived vasodilator (Figure 3B,E). Late-onset of estrogen treatment (E₂L) modified prostanoids' contribution to Phe vasoconstriction by increasing both COX-1- and COX-2-derived vasodilators (Figure 3C,F). In the senescent CCA from SAMP8, both COX inhibitors reduced vasoconstriction to Phe only in the E₂L group (Figure 3I,L). Inhibition of either COX-1 or COX-2 did not affect contractions in arteries from untreated OVX-SAMP8 (Figure 3G,J) or SAMP8 E₂E (Figure 3H,K). mRNA expression analysis of the major enzymes involved in the prostanoids' biosynthesis cascade revealed differential regulation by estrogen in non-senescent and senescent CCA (Figure 4). Estrogen therapy increased the expression of COX-2 in SAMR1 (Figure 4B) and augmented the expression of TXA₂ synthase in SAMP8 (Figure 4D). In contrast, no changes in the expression of COX-1 (Figure 4A) or PGI₂ synthase (Figure 4C) were observed.

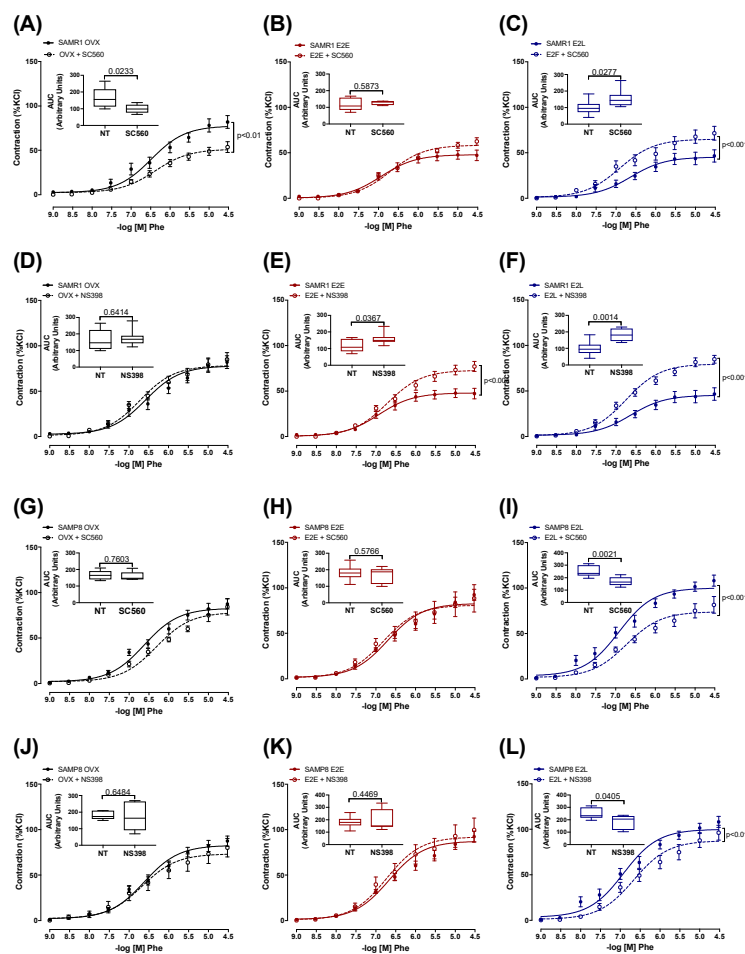


Figure 3. Role of COX-derived prostaglandins in phenylephrine-induced contractions. Concentration–response curves to phenylephrine (Phe) in the endothelium-intact common carotid artery

from SAMR1 (A–F) and SAMP8 (G–L). Arteries were isolated from untreated ovariectomized mice (OVX) or treated with early-onset of 17 β -estradiol treatment (E₂E) or late-onset 17 β -estradiol treatment (E₂L), as indicated. Curves were obtained in the absence or the presence of COX-1 (SC560, 10⁻⁶ M) or COX-2 (NS 398, 10⁻⁶ M) inhibitor. Inset graphs represent the areas under the curve (AUC) obtained from the curves of untreated carotids (NT) or those receiving selective COX inhibition. Each point represents the mean \pm SEM from 5–10 independent experiments. The extra sum-of-squares F determined differences in the fit of concentration–response curves. One-way ANOVA analyzed the dependence of data on the onset of estrogen therapy (none, early, or late) in SAMR1 or SAMP8 with Bonferroni's post-test. *p*-values and comparisons are expressed next to the curves and on top of bar graphs. Significance is considered when *p* < 0.05.

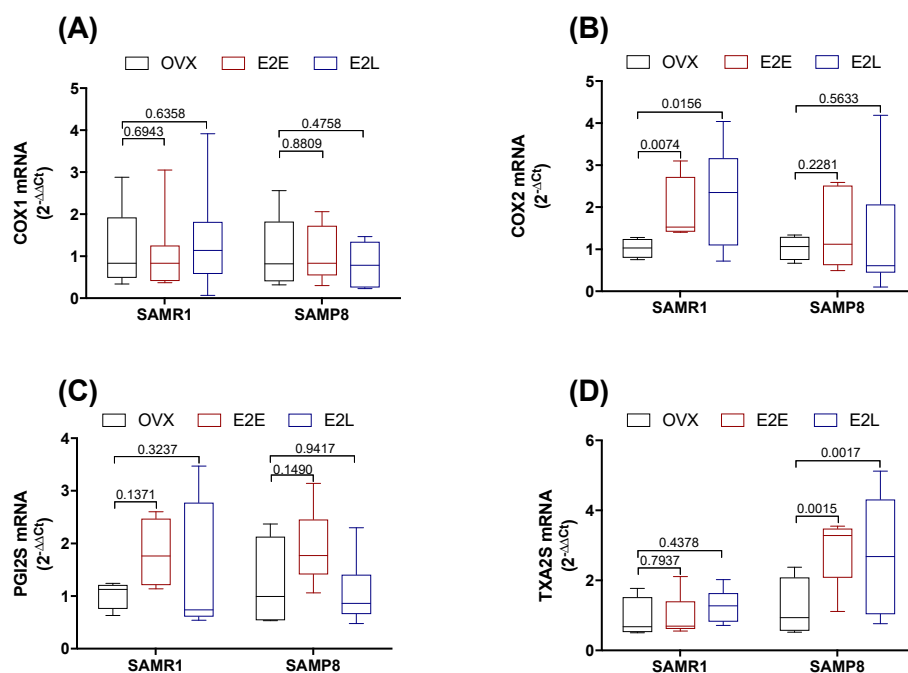


Figure 4. mRNA expression in cyclooxygenase cascade. COX-1 (A), COX-2 (B), PGI₂S (C), and TXA₂S (D) expression in common carotid artery of SAMR1 and SAMP8 normalized by β -actin mRNA and relative to the expression of untreated OVX mice. Total RNA was obtained in vessels from untreated ovariectomized female mice (OVX), OVX mice receiving early-onset 17 β -estradiol treatment (E₂E), and OVX mice receiving late-onset 17 β -estradiol treatment (E₂L). Each point represents the mean \pm SEM from 5–10 independent experiments. One-way ANOVA analyzed the dependence of data on the onset of estrogen therapy (none, early, or late) in SAMR1 or SAMP8 with Bonferroni's post-test. *p*-values and comparisons are expressed on top of bar graphs. Significance is considered when *p* < 0.05.

3.5. Dual Effect of COX-1 and COX-2 in Prostanoids Production

Prostanoids release from CCA was determined in the Krebs–Henseleit solution collected after concentration–response curves to Phe. In SAMR1, E₂E increased the release of PGI₂ by ~2-fold, while it decreased the TXA₂ released by Phe by ~3-fold in comparison to untreated OVX. Late-onset estrogen therapy did not produce the inhibitory effects of estrogen on TXA₂ release in untreated OVX-SAMR1, even though it increased PGI₂ concentration. In senescent CCA, neither early nor late-onset estrogen treatments modified PGI₂ release (Supplementary Figure S5), while E₂L increased TXA₂ concentration induced by Phe stimulation (Supplementary Figure S6). Selective inhibition of COX-1 and COX-2 produced a differential regulation of prostanoids biosynthesis depending on the onset of estrogen therapy in non-senescent and senescent arteries. In SAMR1, increased release of PGI₂ by E₂E and E₂L was mainly dependent on COX-2, since the degree of change in PGI₂ production (Log Fold Change) upon estrogen treatment was only observed when the CCAs were inhibited with NS398. In SAMP8, both COX-1 and COX-2 inhibitors similarly decreased PGI₂ production in all groups (Figure S5). As for

TXA₂, COX-1 is the primary regulator of its production in CCA of OVX-SAMR1 (Figure S6). On the other hand, the increase in TXA₂ observed upon E₂L treatment in SAMP8 was regulated by COX-2 (Figure S6).

3.6. Late-Onset Estrogen Treatment Increases Alternative Splicing of Estrogen Receptor Alpha (ERα36) in Aging

Most of the vascular protective effects of estrogens are mediated through interaction with ERα. We next evaluated how senescence and therapy onset modify ERα expression. Analysis of methylation sites in the gene encoding ERα by methylation-sensitive quantitative PCR showed a marked increase of methylation in the region corresponding to Exon 8 in senescent arteries (Figure 5A). The early onset of estrogen treatment further increased the degree of methylation in the CAA of OVX-SAMP8. In order to determine whether changes in the degree of methylation of the ERα gene can affect the pattern of receptor expression, we analyzed mRNA and protein expression of wild-type ERα and its alternative splicing. The onset of estrogen therapy did not change gene (Supplementary Figure S7) or protein (Figure 5C) expression of the wild-type ERα (ERα-66kDa) in CCA. Gene expression of the variant splicing ERα-36kDa was marked lower relatively to the wild-type receptor in most groups, except SAMP8 E₂L, which exhibited a 6-fold increase over the expression of the wild type ERα (Figure 5B). Changes in mRNA expression of ERα-36kDa corresponded to the pattern of protein expression (Figure 5D). No changes in the expression of the variant ERα-46kDa were found (Figure 5C).

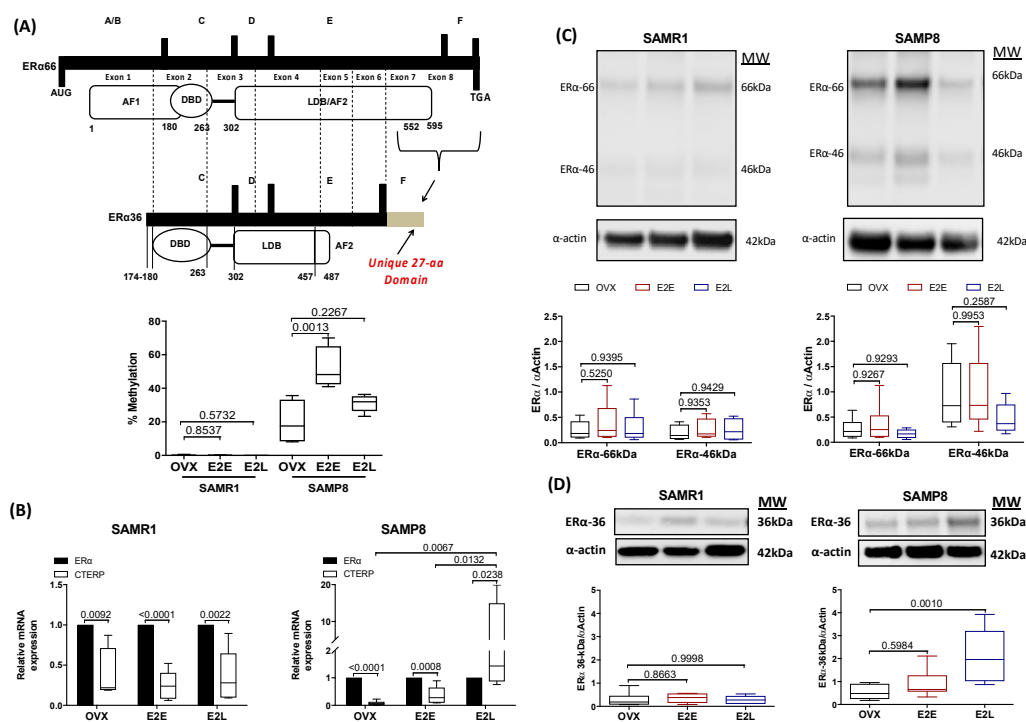


Figure 5. Transcriptional and post-transcriptional regulation of ERα. Percentage of specific DNA methylation in the gene encoding ERα in common carotid artery of SAMR1 and SAMP8 (A). Bar graphs show the percentage (%) of methylation in the region comprised between exon 7 and exon 8 and translated to the C-terminus. mRNA expression of C-terminally truncated ERα product (CTERP) relative to the expression of wild-type (ERα66) in mice carotid (B). Protein expression of the classical ERα66 and the alternative splicing variants ERα46 (C) and ERα36 (D) in common carotid normalized by the expression of α-actin. Samples of genomic DNA, total RNA or protein were obtained in vessels from untreated ovariectomized mice (OVX), ovariectomized mice under early-onset (E₂E) or late-onset (E₂L) of 17β-estradiol treatment. Each point represents the mean ± SEM from 5–7 independent experiments. One-way ANOVA analyzed the dependence of data on the onset of estrogen therapy (none, early, or late) in SAMR1 or SAMP8 with Bonferroni’s post-test. *p*-values and comparisons are expressed on top of bar graphs. Significance is considered when *p* < 0.05.

4. Discussion

In this study, we describe that the onset of estrogen therapy modifies the vascular function of ovariectomized senescent female mice (SAMP8). Our findings are consistent with new clinical studies suggesting that the beneficial effects of estrogen therapy in the cardiovascular system depend on the time when therapy is initiated [14]. Our data show that estrogen reduces constriction to the adrenergic stimulus by Phe in the CCA of non-senescent OVX mice, regardless of treatment onset, by favoring the release of endothelium-derived relaxing factors, such as NO and PGI₂. Nonetheless, onset timing plays a critical role when arteries are senescent, a more representative model of arteries in menopausal women. In CCA of senescent female mice (SAMP8), late-onset of estrogen treatment increases adrenergic vasoconstriction in association with augmented TXA₂ production and upregulation of ER α 36 expression, alternative splicing of the classical ER α .

Estradiol, also known as E₂ or 17 β -estradiol, has been widely used as a therapeutic choice for postmenopausal women seeking amelioration of many different menopause-associated symptoms, including vasomotor symptoms, vaginal atrophy, and prevention of osteoporosis [23]. Because premenopausal women are less likely to progress to cardiovascular disease, a protective cardiovascular effect by estrogen was proposed in the past involving a myriad of different mechanisms [24]. Nevertheless, important randomized and placebo-controlled trials—including the WHI and HERS—raised doubts and concerns about the use of estrogen in the primary and secondary prevention of cardiovascular disease [10,11]. More importantly, the WHI study alerted about the increased risk of stroke not offset by the lower risk of coronary heart disease. The surprising controversies on the risks and benefits of estrogen therapy are, in part, associated with the time of its initiation. Detailed analysis of the WHI showed a significantly lower risk of coronary heart disease by estrogen therapy among women who were less than 10 years past the onset of menopause, and therefore, less time devoid of estrogen actions [12]. Reinforcing this analysis are the results of the ELITE trial specifically designed to test the hormone-timing hypothesis concerning cardiovascular risk in postmenopausal women [14]. Despite growing evidence validating the existence of a window of therapeutic opportunity for the cardiovascular benefits by estrogen, we still need to understand the mechanisms behind it.

Vascular aging has been associated with a series of progressive functional alterations that can modify estrogen signaling in the arterial wall [25]. Considering that women enrolled in the clinical trials on hormone therapy and cardiovascular protections were middle-aged and had long been in menopause, it becomes difficult to separate the contribution of senescence from the long-term estrogen withdrawal to the adverse cardiovascular outcomes observed. In previous studies, we described that senescence dampens the beneficial effects of estrogen in aortas of OVX female mice [16]. However, we still do not know how senescence could modulate the risk-benefit ratio of estrogen use in arteries that are important for cerebral perfusion. Arteries of the distinct sizes and in distinct beds are regulated differently by intrinsic stimuli (including adrenergic stimulation) [26] and by the aging process [27]; therefore, it is of the utmost importance to understand the mechanisms of estrogens in the different senescent vascular beds, and more specifically in the cerebrovasculature. In this regard, we aimed to determine how the time to initiate estrogen treatment shapes estrogen-mediated effects in CCA of both non-senescent and senescent females. We used the OVX-SAMP8 as a model of menopause, based on our previous studies establishing OVX-SAMP8 as a suitable model to concomitantly study the effects of senescence and estrogen deficiency in middle-aged females [21].

In our preliminary studies in the carotid reactivity, we observed that basal response to estrogen might vary depending on mice strain (SAMR1 or SAMP8). When we compared the maximal contraction to Phe in carotid rings from intact (non-ovariectomized) SAMR1 and SAMP8 we observed that the maximal contractile response to Phe was markedly higher in intact SAMP8 ($106 \pm 5.2\%$) in comparison to intact SAMR1 ($81 \pm 1.8\%$, $p < 0.0001$), suggesting a detrimental effect of senescence in this vascular bed. Curiously, OVX did not modify the contractile response in SAMR1 ($82.2 \pm 2.3\%$, $p = 0.812$), while it decreased Phe contraction in SAMP8 ($78.8 \pm 1.8\%$, $p < 0.001$). Together, these results propose that

estrogen plays a differential role in the control of vascular reactivity in young and senescent carotid arteries, even when at physiological levels.

A correlation between senescence and vascular dysfunction has been extensively described in men and women and much associated with decreased NO-mediated vasodilation [15,28]. Intact (non-ovariectomized) SAMP8 female mice presented a faster and time-dependent decrease in NO and increased oxidative stress compared to SAMR1 [29]. Estrogen withdrawal suppresses the involvement of NO in both groups [30], suggesting an overlap between aging and hormonal levels to control NO-mediated vascular function, similar to what is observed in postmenopausal women [31]. When females are non-senescent (SAMR1), estrogen therapy improves the vascular function of CCA (decreased Phe vasoconstriction) and augment NO bioavailability, via both upregulation of eNOS and decreased oxidative stress. Corroborating our data in SAMR1, several experimental studies in different vascular systems of young ovariectomized animal models show protective effects by estrogens in the aorta [5,32–34], mesenteric microvessels [8] and veins [9].

On the other hand, when females are senescent, estrogen treatment does not fully restore NO bioavailability in CCA. The reasons for this effect are unclear but may be related to the age-associated phenotypic changes in the vascular wall that diminish responsiveness [6,35] or modify signaling for estrogens [16]. In previous studies, we found that estrogens lose its ability to modulate eNOS expression, while significantly increases NADPH oxidase 1 (NOX1) expression and, consequently, the production of O_2^- in aortas of OVX-SAMP8 mice [16].

In addition to NO, the cyclooxygenase (COX)-prostanoid cascade has also been implicated in the pathophysiology of vascular dysfunction during aging [36,37] and following estrogen withdrawal [7]. As revised by Hermenegildo and colleagues [38], estrogen increases the production of the COX-derived vasodilators PGI_2 in different vascular beds—such as uterine, mesenteric, cerebral, and aortic—supporting additional benefit by this hormone in the female vasculature, whereas PGI_2 metabolite levels decrease in postmenopausal women [39]. Moreover, PGI_2 has been associated with the atheroprotective effect of estrogen [40]. In CCA of non-senescent SAMR1, COX-1 inhibition decreases Phe contraction, revealing an imbalance towards COX-1-derived vasoconstrictor prostanoids generation when estrogen is withdrawn by OVX. In contrast, COX-2 inhibition increases vasoconstriction in estrogen-treated SAMR1, suggesting an increased COX-2-derived vasodilator production by estrogen. Estrogen increased COX-2 mRNA expression in CCA of SAMR1 female, regardless of the timing for therapy initiation, which is associated with a marked increase of PGI_2 concentration in the organ bath after Phe stimulation. Estrogen increases COX-2-mediated PGI_2 production in both human endothelial cells [41] and mice artery [40].

On the other hand, the senescent carotids of SAMP8 show a different relationship between estrogens and prostanoids production. Our preliminary data using nonspecific COX inhibitor (indomethacin) agree with other studies describing an increase of COX-derived vasoconstrictor products by aging and estrogen withdrawal, as indomethacin decreases Phe-induced vasoconstriction in all SAMP8 groups. However, when we used selective COX-1 or COX-2 inhibitors, we did not see any change in Phe responses in CCA of OVX and E_2 -treated SAMP8. Only in arteries of E_2 L females, both inhibitors decrease vasoconstrictor responses. These results suggest crosstalk between COX-1 and COX-2 in senescent arteries that maintain vasoconstriction when one or another is inhibited. Nevertheless, this compensation is missing in senescent females when they receive a late onset of estrogen therapy. The responses in E_2 L were associated with an increase of TXA_2 synthase expression and TXA_2 production after challenge with Phe. Augmented TXA_2 production was previously reported in intact six months old SAMP8 female mice, and nonspecific COX inhibition corrected this parameter [29]. In postmenopausal women, estrogen increases platelet activation via TXA_2 production [42]. In contrast, another study demonstrated that estrogen does not change TXA_2 production in cultured human endothelial cells [43]. Together, these results suggest that the effects of estrogen on TXA_2 production may depend on cell/tissue type and pathophysiological condition.

The primary receptor responsible for the vascular actions of estrogens is the ER α , despite many studies describing vascular actions of ER β and GPER [17]. Studies with ER α knockout [44] and Cre-lox recombinant endothelium-specific ER α knockout flox/flox (CreLoxP) mice [45] have demonstrated that ER α is necessary for estrogen-induced reendothelialization and vascular protection. Moreover, ER α activation has been associated with increased eNOS transcription and NO production [46], antioxidant effects [47], and regulation of prostanoids production [48], reinforcing that ER α activation is essential for most of the beneficial actions of estrogens in the vasculature. Expression of ER α is subject to complex regulation since the gene encoding this receptor subtype has a multiple promoter system, which produces distinct splicing variants during transcription. The two most-studied ER α isoforms are the 46Kd (ER α 46) and the 36Kd (ER α 36) receptors. ER α 46 lacks the transcriptional activation function-1 (AF-1) in the N-terminus, while ER α 36 misses both AF-1 and part of AF-2 in the C-terminal portion of the receptor. Thus, these variants may have limited transcriptional activity and can oppose genomic actions of the full-length 66Kd (ER α 66) [17]. Disruption in estrogen signaling by upregulation of these splicing has been associated with cancer [20]. The pattern of ER α 66 expression and its splicing might also be a determining factor in the aging process. It is known that alternative splicing is a typical post-transcriptional process in eukaryotes that can be associated with premature aging and associated diseases [49]. However, the contribution of alternative variants of ER α to women's vascular health and disease remains an open question.

In the present study, we described for the first time that the CCA of female mice expresses both ER α 46 and ER α 36 variants. The expression of ER α 36 mRNA was markedly lower relative to wild-type ER α 66 in all experimental groups of SAMR1 mice. However, in senescent CCA late onset of estrogen treatment increased ER α 36 expression to levels that were higher than the wild type receptors. The upregulation of ER α 36 in E₂L-SAMP8 was parallel by both increased TXA₂ production and Phe vasoconstriction. Although we cannot provide direct evidence on the role of ER α 36 on TXA₂ production, due to the lack of specific agonist/antagonist for ER36, the fact that the expression of this variant is increased only in the group where we observed increased TXA₂ release, points to a plausible relationship between these two pathways. Anti-ER α 36 antibody treatment was shown to reduce estrogen-induced vasodilation in middle cerebral arteries from male rats [50]. Even though this study supports the vascular role of ER α 36, the molecular mechanisms regulated by this specific ER α variant in the vascular wall warrant further examination.

In summary, this study demonstrates that timing to initiate the treatment is a determining factor for the detrimental or beneficial effects of estrogens. More specifically, when arteries are senescent, late-onset estrogen therapy may represent a risk to the vasculature for increasing the production of contractile factors and vasoconstriction. Although we do not have direct evidence, the detrimental effects of estrogen are in parallel with an increase in expression of ER α 36, which could plausibly modify estrogen signaling pathways towards vascular impairment. On average, women are living 20 years in menopause, and the use of estrogen in this population has been widely used to lessen problems associated with this natural process. This study draws our attention to the need for increasing our understanding of the mechanisms of action of estrogen receptors and their variants during cardiovascular aging in women. Because of the great controversy and the gap in the knowledge of the risks/benefits of estrogen use in the cardiovascular system, our results are of significant scientific impact. Besides, our results provide new insights into the regulation of estrogen receptors in aging and disease, which may eventually lead to the development of new regulatory molecules acting on estrogen receptors to improve vascular health in women and men.

5. Translational Perspective

The risk or benefit for estrogen use in menopausal women remain controversial, and the modulation of estrogen signaling in healthy, aged, and diseased vasculature is still unknown. In precision medicine times, we still do not know which population of women would benefit from the use of estrogen and in which one hormone therapy would pose a cardiovascular risk. This study shows that timing to

initiate estrogen treatment is a determining factor for the detrimental or beneficial effects of estrogens and provide new insights into the regulation of estrogen receptors in aging and disease, which may eventually lead to the development of new regulatory molecules acting on estrogen receptors to improve vascular health in postmenopausal women.

Supplementary Materials: The following are available online at <http://www.mdpi.com/2073-4409/8/10/1217/s1>, Figure S1: U46619-induced contraction; Figure S2: Acetylcholine (ACh) and Sodium Nitroprusside (SNP)-induced relaxation; Figure S3: Effects of reactive oxygen species (ROS) on Phenylephrine (Phe)-induced contractions; Figure S4: Effects of Cyclooxygenase (COX)-derived vasoactive metabolites on Phenylephrine (Phe)-induced contractions; Figure S5: Prostacyclin (PGI₂) production; Figure S6: Thromboxane A₂ (TXA₂) production; Figure S7: Expression of the classical estrogen receptor (ER α) in common carotid artery; Major Resources Table S1: Animal Model; Major Resources Table S2: Vascular Reactivity; Major Resources Table S3: Primer sequences for quantitative RT-PCR; Major Resources Table S4: Primer sequences for quantitative DNA methylation; Major Resources Table S5: Antibodies.

Author Contributions: T.J.C., E.V., A.P.D. and M.H.C.d.C. conceived the study; T.J.C., F.J.-A., C.E. and A.P.D., performed the experimental design and laboratory experiments; T.J.C., F.J.-A., E.H.A., R.T., E.V., A.P.D., M.H.C.d.C. performed the analyses and discussion of results; T.J.C. and A.P.D. wrote the manuscript; T.J.C., F.J.-A., E.H.A., R.T., E.V., A.P.D. and M.H.C.d.C., critically revised the manuscript. All authors read and approved the manuscript.

Funding: This work was supported by Fundação de Amparo a Pesquisa do Estado de São Paulo – FAPESP (2014/03758-4), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES/DGU 269/12). Ministerio de Economía y Competitividad, Instituto Carlos III (FIS PI13/00091, PI16/00742 and RIC RD12/0042/0006). Programa Hispano-Brasileño de Cooperación Interuniversitaria (HBP-2011-0054 PC). Tiago J. Costa was supported by FAPESP (2013/19423-9; 2015/26690-9) fellowship.

Acknowledgments: The authors are grateful to Nadia Castillo Machado, Sonia Maria Leite, and Marta Rodrigues for technical support in the experiments. Tiago J. Costa is grateful to Yara Onetti, Joaquim Bobi, Cira Rubies, Manel Garabito, and Renée de Nazaré Oliveira da Silva, graduate students at Universitat Autònoma de Barcelona, IDIBAPS and the University of Sao Paulo, for scientific support and discussions during the development of this project.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Abbreviations

CCA	Common carotid artery
OVX	Ovariectomized
SAMP8	Senescence-accelerated mice prone-8
SAMR1	Senescence-accelerated mice resistant-1
E2	Estrogen
E2E	Early estrogen treatment
E2L	Late estrogen treatment
Phe	Phenylephrine
ACh	Acetylcholine
NPS	Sodium nitroprusside
ER α	Estrogen receptor alpha
COX-1	Cyclooxygenase 1
COX-2	Cyclooxygenase 2
PGI ₂	Prostacyclin
TXA ₂	Thromboxane A ₂

References

1. Comhaire, F. Hormone replacement therapy, and longevity. *Andrologia* **2016**, *48*, 65–68. [[CrossRef](#)] [[PubMed](#)]
2. Benjamin, E.J.; Virani, S.S.; Callaway, C.W.; Chamberlain, A.M.; Chang, A.R.; Cheng, S.; Chiuve, S.E.; Cushman, M.; Dellinger, F.N.; Deo, R.; et al. Heart Disease and Stroke Statistics-2018 Update: A Report From the American Heart Association. *Circulation* **2018**, *137*, e67–e492. [[CrossRef](#)] [[PubMed](#)]

3. Stampfer, M.J.; Willett, W.C.; Colditz, G.A.; Rosner, B.; Speizer, F.E.; Hennekens, C.H. A prospective study of postmenopausal estrogen therapy and coronary heart disease. *N. Engl. J. Med.* **1985**, *313*, 1044–1049. [[CrossRef](#)] [[PubMed](#)]
4. Filgueira, F.; Lobato, N.; Ceravolo, G.; Dantas, A.P.; Fortes, Z.; Webb, C.; Tostes, R.; Carvalho, M.H. Characterization of the relaxant response to equilin in rat mesenteric arteries. *FASEB J.* **2010**, *24*, 575–577.
5. Ceravolo, G.S.; Filgueira, F.P.; Costa, T.J.; Lobato, N.S.; Chignalia, A.Z.; Araujo, P.X.; Tostes, R.C.; Dantas, A.P.; Fortes, Z.B.; Carvalho, M.H.C. Conjugated equine estrogen treatment corrected the exacerbated aorta oxidative stress in ovariectomized spontaneously hypertensive rats. *Steroids* **2013**, *78*, 341–346. [[CrossRef](#)]
6. Novella, S.; Heras, M.; Hermenegildo, C.; Dantas, A.P. Effects of estrogen on vascular inflammation: A matter of timing. *Arterioscler. Thromb. Vasc. Biol.* **2012**, *32*, 2035–2042. [[CrossRef](#)]
7. Dantas, A.P.V.; Scivoletto, R.; Fortes, Z.B.; Nigro, D.; Carvalho, M.H.C. Influence of Female Sex Hormones on Endothelium-Derived Vasoconstrictor Prostanoid Generation in Microvessels of Spontaneously Hypertensive Rats. *Hypertension* **1999**, *34*, 914–919. [[CrossRef](#)]
8. Dantas, A.P.V.; Tostes, R.C.; Nigro, D.; Fortes, Z.B.; Carvalho, M.H.C. In vivo evidence for the antioxidant potential of estrogen in spontaneously hypertensive rats. *Hypertension* **2002**, *39*, 405–411. [[CrossRef](#)]
9. Araujo, P.X.; Costa, T.J.; Echem, C.; de Oliveira, M.; Santos-Eichler, R.A.; Colli, L.G.; Jiménez-Altayó, F.; Vila, E.; Akamine, E.H.; Dantas, A.P.; et al. Treatment with Standard and Low Dose of Conjugated Equine Estrogen Differentially Modulates Estrogen Receptor Expression and Response to Angiotensin II in Mesenteric Venular Bed of Surgically Postmenopausal Hypertensive Rats. *J. Pharmacol. Exp. Ther.* **2017**, *362*, 98–107. [[CrossRef](#)]
10. Hulley, S.; Grady, D.; Bush, T.; Furberg, C.; Herrington, D.; Riggs, B.; Vittinghoff, E. Randomized trial of estrogen plus progestin for secondary prevention of coronary heart disease in postmenopausal women. Heart and Estrogen/progestin Replacement Study (HERS) Research Group. *JAMA* **1998**, *280*, 605–613. [[CrossRef](#)]
11. Rossouw, J.E.; Anderson, G.L.; Prentice, R.L.; LaCroix, A.Z.; Kooperberg, C.; Stefanick, M.L.; Jackson, R.D.; Beresford, S.A.; Howard, B.V.; Johnson, K.C.; et al. Risks and benefits of estrogen plus progestin in healthy postmenopausal women: Principal results From the Women’s Health Initiative randomized controlled trial. *JAMA* **2002**, *288*, 321–333. [[PubMed](#)]
12. Grodstein, F.; Manson, J.E.; Stampfer, M.J. Hormone therapy and coronary heart disease: The role of time since menopause and age at hormone initiation. *J. Womens Heal.* **2006**, *15*, 35–44. [[CrossRef](#)] [[PubMed](#)]
13. Harman, S.M.; Vittinghoff, E.; Brinton, E.A.; Budoff, M.J.; Cedars, M.I.; Lobo, R.A.; Merriam, G.R.; Miller, V.M.; Naftolin, F.; Pal, L.; et al. Timing and duration of menopausal hormone treatment may affect cardiovascular outcomes. *Am. J. Med.* **2011**, *124*, 199–205. [[CrossRef](#)] [[PubMed](#)]
14. Hodis, H.N.; Mack, W.J.; Henderson, V.W.; Shoupe, D.; Budoff, M.J.; Hwang-Levine, J.; Li, Y.; Feng, M.; Dustin, L.; Kono, N.; et al. Vascular Effects of Early versus Late Postmenopausal Treatment with Estradiol. *N. Engl. J. Med.* **2016**, *374*, 1221–1231. [[CrossRef](#)] [[PubMed](#)]
15. Izzo, C.; Carrizzo, A.; Alfano, A.; Virtuoso, N.; Capunzo, M.; Calabrese, M.; De Simone, E.; Sciarretta, S.; Frati, G.; Olivetti, M.; et al. The impact of aging on cardio and cerebrovascular diseases. *Int. J. Mol. Sci.* **2018**, *19*, 481. [[CrossRef](#)] [[PubMed](#)]
16. Novensa, L.; Novella, S.; Medina, P.; Segarra, G.; Castillo, N.; Heras, M.; Hermenegildo, C.; Dantas, A.P. Aging Negatively Affects Estrogens-Mediated Effects on Nitric Oxide Bioavailability by Shifting ER alpha/ER beta Balance in Female Mice. *PLoS ONE* **2011**, *6*, e25335. [[CrossRef](#)] [[PubMed](#)]
17. Arnal, J.-F.; Lenfant, F.; Metivier, R.; Flouriot, G.; Henrion, D.; Adlanmerini, M.; Fontaine, C.; Gourdy, P.; Chambon, P.; Katzenellenbogen, B.; et al. Membrane and Nuclear Estrogen Receptor Alpha Actions: From Tissue Specificity to Medical Implications. *Physiol. Rev.* **2017**, *97*, 1045–1087. [[CrossRef](#)]
18. Wang, Z.; Zhang, X.; Shen, P.; Loggie, B.W.; Chang, Y.; Deuel, T.F. Identification, cloning, and expression of human estrogen receptor-alpha36, a novel variant of human estrogen receptor-alpha66. *Biochem. Biophys. Res. Commun.* **2005**, *336*, 1023–1027. [[CrossRef](#)]
19. Flouriot, G.; Brand, H.; Denger, S.; Metivier, R.; Kos, M.; Reid, G.; Sonntag-Buck, V.; Gannon, F. Identification of a new isoform of the human estrogen receptor-alpha (hER-alpha) that is encoded by distinct transcripts and that is able to repress hER-alpha activation function 1. *EMBO J.* **2000**, *19*, 4688–4700. [[CrossRef](#)]
20. Hua, H.; Zhang, H.; Kong, Q.; Jiang, Y. Mechanisms for estrogen receptor expression in human cancer. *Exp. Hematol. Oncol.* **2018**, *7*, 24. [[CrossRef](#)]

21. Novella, S.; Dantas, A.P.; Segarra, G.; Novensà, L.; Bueno, C.; Heras, M.; Hermenegildo, C.; Medina, P. Gathering of aging and estrogen withdrawal in vascular dysfunction of senescent accelerated mice. *Exp. Gerontol.* **2010**, *45*, 868–874. [[CrossRef](#)] [[PubMed](#)]
22. Onetti, Y.; Jiménez-Altayó, F.; Heras, M.; Vila, E.; Dantas, A.P. Western-type diet induces senescence, modifies vascular function in non-senescence mice and triggers adaptive mechanisms in senescent ones. *Exp. Gerontol.* **2013**, *48*, 1410–1419. [[CrossRef](#)] [[PubMed](#)]
23. Takahashi, T.A.; Johnson, K.M. Menopause. *Med. Clin. N. Am.* **2015**, *99*, 521–534. [[CrossRef](#)]
24. Xing, D.; Nozell, S.; Chen, Y.F.; Hage, F.; Oparil, S. Estrogen and mechanisms of vascular protection. *Arter. Thromb. Vasc. Biol.* **2009**, *29*, 289–295. [[CrossRef](#)] [[PubMed](#)]
25. Stice, J.P.; Lee, J.S.; Pechenino, A.S.; Knowlton, A.A. Estrogen, aging and the cardiovascular system. *Futur. Cardiol.* **2009**, *5*, 93–103. [[CrossRef](#)] [[PubMed](#)]
26. Gordan, R.; Gwathmey, J.K.; Xie, L.-H. Autonomic and endocrine control of cardiovascular function. *World J. Cardiol.* **2015**, *7*, 204–214. [[CrossRef](#)] [[PubMed](#)]
27. Mitchell, G.F. Effects of central arterial aging on the structure and function of the peripheral vasculature: Implications for end-organ damage. *J. Appl. Physiol.* **2008**, *105*, 1652–1660. [[CrossRef](#)]
28. Lakatta, E.G. Arterial and cardiac aging: Major shareholders in cardiovascular disease enterprises: Part III: Cellular and molecular clues to heart and arterial aging. *Circulation* **2003**, *107*, 490–497. [[CrossRef](#)]
29. Novella, S.; Dantas, A.P.; Segarra, G.; Novensà, L.; Heras, M.; Hermenegildo, C.; Medina, P. Aging enhances contraction to thromboxane A2 in aorta from female senescence-accelerated mice. *Age (Omaha)* **2013**, *35*, 117–128. [[CrossRef](#)]
30. Vidal-Gómez, X.; Novella, S.; Pérez-Monzó, I.; Garabito, M.; Dantas, A.P.; Segarra, G.; Hermenegildo, C.; Medina, P. Decreased bioavailability of nitric oxide in aorta from ovariectomized senescent mice: Role of cyclooxygenase. *Exp. Gerontol.* **2016**, *76*, 1–8. [[CrossRef](#)]
31. Virdis, A.; Ghiadoni, L.; Pinto, S.; Lombardo, M.; Petraglia, F.; Gennazzani, A.; Buralli, S.; Taddei, S.; Salvetti, A. Mechanisms responsible for endothelial dysfunction associated with acute estrogen deprivation in normotensive women. *Circulation* **2000**, *101*, 2258–2263. [[CrossRef](#)] [[PubMed](#)]
32. Rubanyi, G.M.; Freay, A.D.; Kauser, K.; Sukovich, D.; Burton, G.; Lubahn, D.B.; Couse, J.F.; Curtis, S.W.; Korach, K.S. Vascular estrogen receptors and endothelium-derived nitric oxide production in the mouse aorta. Gender difference and effect of estrogen receptor gene disruption. *J. Clin. Investig.* **1997**, *99*, 2429–2437. [[CrossRef](#)] [[PubMed](#)]
33. Rubanyi, G.M.; Johns, A.; Kauser, K. Effect of estrogen on endothelial function and angiogenesis. *Vasc. Pharmacol.* **2002**, *38*, 89–98. [[CrossRef](#)]
34. Costa, T.J.; Ceravolo, G.S.; Dos Santos, R.A.; De Oliveira, M.A.; Araújo, P.X.; Giaquinto, L.R.; Tostes, R.C.; Akamine, E.H.; Fortes, Z.B.; Dantas, A.P.; et al. Association of testosterone with estrogen abolishes the beneficial effects of estrogen treatment by increasing ROS generation in aorta endothelial cells. *Am. J. Physiol. - Hear. Circ. Physiol.* **2015**, *308*, 723–732. [[CrossRef](#)] [[PubMed](#)]
35. Sherwood, A.; Bower, J.K.; McFetridge-Durdle, J.; Blumenthal, J.A.; Newby, L.K.; Hinderliter, A.L. Age moderates the short-term effects of transdermal 17beta-estradiol on endothelium-dependent vascular function in postmenopausal women. *Arter. Thromb. Vasc. Biol.* **2007**, *27*, 1782–1787. [[CrossRef](#)] [[PubMed](#)]
36. Briones, A.M.; Montoya, N.; Giraldo, J.; Vila, E. Ageing affects nitric oxide synthase, cyclooxygenase and oxidative stress enzymes expression differently in mesenteric resistance arteries. *Aut. Autacoid Pharmacol.* **2005**, *25*, 155–162. [[CrossRef](#)] [[PubMed](#)]
37. Costa, G.; Garabito, M.; Jiménez-Altayó, F.; Onetti, Y.; Sabate, M.; Vila, E.; Dantas, A.P. Sex differences in angiotensin II responses contribute to a differential regulation of cox-mediated vascular dysfunction during aging. *Exp. Gerontol.* **2016**, *85*, 71–80. [[CrossRef](#)]
38. Hermenegildo, C.; Oviedo, P.J.; Cano, A. Cyclooxygenases regulation by estradiol on endothelium. *Curr. Pharm. Des.* **2006**, *12*, 205–215. [[CrossRef](#)]
39. Mueck, A.O.; Seeger, H.; Lüdtke, R.; Gräser, T.; Wallwiener, D. Effect on biochemical vasoactive markers during postmenopausal hormone replacement therapy: Estradiol versus estradiol/dienogest. *Maturitas* **2001**, *38*, 305–313. [[CrossRef](#)]
40. Egan, K.M.; Lawson, J.A.; Fries, S.; Koller, B.; Rader, D.J.; Smyth, E.M.; Fitzgerald, G.A. COX-2-derived prostacyclin confers atheroprotection on female mice. *Science (80-.)* **2004**, *306*, 1954–1957. [[CrossRef](#)]

41. Akarasereenont, P.; Techatraisak, K.; Thaworn, A.; Chotewuttakorn, S. The induction of cyclooxygenase-2 by 17beta-estradiol in endothelial cells is mediated through protein kinase C. *Inflamm. Res.* **2000**, *49*, 460–465. [[CrossRef](#)] [[PubMed](#)]
42. Oliveira, R.L.; Aldrighi, J.M.; Gebara, O.E.; Rocha, T.R.; D'Amico, E.; Rosano, G.M.; Ramires, J.A. Postmenopausal hormone replacement therapy increases plasmatic thromboxane beta 2. *Int. J. Cardiol.* **2005**, *99*, 449–454. [[CrossRef](#)] [[PubMed](#)]
43. Sobrino, A.; Mata, M.; Laguna-Fernandez, A.; Novella, S.; Oviedo, P.J.; García-Pérez, M.A.; Tarín, J.J.; Cano, A.; Hermenegildo, C. Estradiol stimulates vasodilatory and metabolic pathways in cultured human endothelial cells. *PLoS ONE* **2009**, *4*, e8242. [[CrossRef](#)] [[PubMed](#)]
44. Pare, G.; Krust, A.; Karas, R.H.; Dupont, S.; Aronovitz, M.; Chambon, P.; Mendelsohn, M.E. Estrogen receptor-alpha mediates the protective effects of estrogen against vascular injury. *Circ. Res.* **2002**, *90*, 1087–1092. [[CrossRef](#)] [[PubMed](#)]
45. Billon-Galés, A.; Fontaine, C.; Douin-Echinard, V.; Delpy, L.; Berges, H.; Calippe, B.; Lenfant, F.; Laurell, H.; Guéry, J.C.; Gourdy, P.; et al. Endothelial estrogen receptor-alpha plays a crucial role in the atheroprotective action of 17beta-estradiol in low-density lipoprotein receptor-deficient mice. *Circulation* **2009**, *120*, 2567–2576. [[CrossRef](#)]
46. Sumi, D.; Ignarro, L.J. Estrogen-related receptor alpha 1 upregulates endothelial nitric oxide synthase expression. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 14451–14456. [[CrossRef](#)]
47. Zhang, Q.G.; Raz, L.; Wang, R.; Han, D.; De Sevilla, L.; Yang, F.; Vadlamudi, R.K.; Brann, D.W. Estrogen attenuates ischemic oxidative damage via an estrogen receptor alpha-mediated inhibition of NADPH oxidase activation. *J. Neurosci.* **2009**, *29*, 13823–13836. [[CrossRef](#)]
48. Sobrino, A.; Oviedo, P.J.; Novella, S.; Laguna-Fernandez, A.; Bueno, C.; García-Pérez, M.A.; Tarín, J.J.; Cano, A.; Hermenegildo, C. Estradiol selectively stimulates endothelial prostacyclin production through estrogen receptor- α . *J. Mol. Endocrinol.* **2010**, *44*, 237–246. [[CrossRef](#)]
49. Tazi, J.; Bakkour, N.; Stamm, S. Alternative splicing and disease. *Biochim. Biophys. Acta* **2009**, *1792*, 14–26. [[CrossRef](#)]
50. Patkar, S.; Farr, T.D.; Cooper, E.; Dowell, F.J.; Carswell, H. V Differential vasoactive effects of oestrogen, oestrogen receptor agonists and selective oestrogen receptor modulators in rat middle cerebral artery. *Neurosci. Res.* **2011**, *71*, 78–84. [[CrossRef](#)]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).