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Multimodal Dispersion of Nanoparticles: A Comprehensive Evaluation of Size Distribution with 9 Size Measurement Methods

Fanny Varenne¹ · Ali Makky¹ · Mireille Gaucher Delmas² · Frédéric Violleau^{3,4} · Christine Vauthier¹

ABSTRACT

Purpose Evaluation of particle size distribution (PSD) of multimodal dispersion of nanoparticles is a difficult task due to inherent limitations of size measurement methods. The present work reports the evaluation of PSD of a dispersion of poly(isobutylcyanoacrylate) nanoparticles decorated with dextran known as multimodal and developed as nanomedicine.

Methods The nine methods used were classified as batch particle i.e. Static Light Scattering (SLS) and Dynamic Light Scattering (DLS), single particle i.e. Electron Microscopy (EM), Atomic Force Microscopy (AFM), Tunable Resistive Pulse Sensing (TRPS) and Nanoparticle Tracking Analysis (NTA) and separative particle i.e. Asymmetric Flow Field Flow Fractionation coupled with DLS (AsFFFF) size measurement methods.

Results The multimodal dispersion was identified using AFM, TRPS and NTA and results were consistent with those provided with the method based on a separation step prior to on line size measurements. None of the light scattering batch methods could reveal the complexity of the PSD of the dispersion.

Conclusions Difference between PSD obtained from all size measurement methods tested suggested that study of the PSD of multimodal dispersion required to analyze samples by at

least one of the single size particle measurement method or a method that uses a separation step prior PSD measurement.

KEY WORDS light scattering · microscopy · nanoparticle tracking analysis · particle size distribution · tunable resistive pulse sensing

ABBREVIATIONS

AFM	Atomic force microscopy
AsFFFF	Asymmetrical flow field flow fractionation
DCS	Differential centrifugal sedimentation
DLS	Dynamic light scattering
EM	Electron microscopy
IBCA	Isobutylcyanoacrylate
NTA	Nanoparticle tracking analysis
PCCS	Photon cross correlation spectroscopy
PIBCA	Poly(isobutylcyanoacrylate)
PSD	Particle size distribution
PTA	Particle tracking analysis
QELS	Quasi elastic light scattering
SdFFF	Sedimentation field flow fractionation
SEM	Scanning electron microscopy
SLS	Static light scattering
TEM	Transmission electron microscopy
TRPS	Tunable resistive pulse sensing

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INTRODUCTION

Nanoparticles have been introduced in many applications including transport i.e. additives for fuels (1), industrial production i.e. catalysis (2), cosmetics i.e. sun blocks (3) and medicine i.e. drug delivery (4), contrast agents for imaging technique (5–7) or adjuvants potentializing effects of radiotherapy (8,9). Physicochemical properties of nanoparticles are strongly size dependent and their safety as well (10). In industries, physical

properties knowledge is important to understand and optimize processes. For instance, particle size and particle size distribution (PSD) measurements are paramount to investigate the repeatability and the efficiency of various industrial processes and products (11). In medicine, desired properties of the nanoparticles could be drawn by controlling nanoparticle size among other physico chemical characteristics (12–20). Although size and PSD of nanomaterials and nanomedicines have been identified as critical for a given application, it is paramount to be able to measure accurately these parameters having reliable size measurement methods (20). In the industry also the size characterization of nanomaterials is a critical parameter for the property and safety. A wide choice of methods based on different physical principles is available to measure nanoparticle size and PSD that can be applied on different types of particle. Microscopy is a direct method based on the analysis of images of the nanoparticles. In contrast, all the other methods are indirect. Although they are generally quite accurate to determine size characteristics of homogeneously distributed monomodal nanoparticle dispersions (21–26), the determination of the PSD of a dispersion of nanoparticles having distinct populations with different sizes (multimodal) having their own PSD remains extremely challenging (19). The different methods can be classified the way they are evaluating the PSD. This can be achieved directly including the dispersion on the whole population, i.e. “in batch”, or analyzing each nanoparticle individually (single size measurement method) (27). In general, signal produced from the application of physical methods are analysed based on mathematical models to deduce size characteristics of the dispersion. Several methods include a stage of separation enabling a fractionation of the particle as the function of their size prior to the determination of their size (27). The Tables I,

II and III summarize each method, the measurand and the type of obtained size distribution from the raw data for the different classes of methods i.e. batch, single and using a separative particle size measurement method respectively. While commercial apparatus are available to achieve size measurements with these methods that are based on different modalities, evaluating PSD of multimodal particles remains challenging. In general, it is recommended to investigate PSD of complex samples using different size measurement methods (24,33–40).

Several works have investigated the PSD of multimodal dispersions prepared intentionally by mixing particles of different size in different known proportions. The most advanced work was proposed by Anderson *et al.* investigating a multimodal mixture of 220, 330 and 400 nm polystyrene particles using different methods from the three classes of methods defined above: (i) batch particle size measurement method i.e. Dynamic Light Scattering (DLS), (ii) single particle size measurement method i.e. Transmission Electron Microscopy (TEM), Tunable Resistive Pulse Sensing (TRPS) and Particle Tracking Analysis (PTA) and (iii) separative particle size measurement method i.e. Differential Centrifugal Sedimentation (DCS) (33). TRPS and DCS were able to discriminate the three populations present in the mixed multimodal sample due to sufficient resolution. In contrast, light scattering methods were only able to resolve a single population. PTA resolved the largest population whereas DLS resolved the smallest population in the multimodal mixture. It is noteworthy that each light scattering method was able to resolve and provide accurate mean particle size within 10% of TEM reference value for particles taken independently of each other. Other work proposed by Cascio *et al.* investigated PSD of a bimodal mixture of 40 and 70 nm monodisperse

Table I Main Characteristics of Batch Particle Size Measurement Methods

Method	Principle	Measurand	Type of size distribution of raw data
Acoustic Techniques (28)	Measurement of the attenuation of the acoustic wave produced within a certain frequency range crossing through a dispersion	Volume based diameter	Volume based
Dynamic Light Scattering (DLS) (19,27,28)	Measurement of nanomaterial movements due to Brownian motion	Autocorrelation function of the scattered light intensity, Translational diffusion coefficient, Hydrodynamic diameter (Stokes Einstein analysis)	Scattering intensity based
Small Angle X ray Scattering (SAXS) (27)	Measurement scattered X rays light of nanomaterials with electron density diluted in dispersant with different electron density	Gyration diameter (Guiner analysis)	Scattering intensity based
Static Light Scattering (SLS) (28)	Measurement of the angular dependence of the scattered light (variation of the intensity of scattered light depending on nanomaterial size and detecting angle)	Gyration diameter (Rayleigh analysis)	Scattering intensity based
X ray diffraction (XRD) (27)	Measurement of diffraction rings at various angles	Scherrer's diameter	No distribution measured

Table II Main Characteristics of Single Particle Size Measurement Methods

Method	Principle	Measurand	Type of size distribution of raw data
Atomic Force Microscopy (AFM) (19,27,28)	Visualisation of nanomaterial images	Height of nanomaterial (used as equivalent to diameter) or diameter determined by analyzing images in the lateral (x y) dimension	Number based
Electron Microscopy (EM) (19,27,28)	Visualisation of nanomaterial images	Equivalent spherical diameter corresponding to the diameter of a circle with the same area than the projected area of the particle or Feret's diameter corresponding to the mean value of the distance between parallel tangents of the projected shape of the position of the particle	Number based
Nanoparticle Tracking Analysis (NTA) (27)	Measurement of ability of individual nanomaterials to scatter light and nanomaterials movement due to Brownian motion	Diffusion length obtained with tracking of the particle on 2 dimension, Translational diffusion coefficient, Hydrodynamic diameter (Stokes Einstein analysis)	Number based
single particle Inductively Coupled Plasma Mass Spectrometry (spICP MS) (27)	Individually atomization and ionisation of nanomaterials through plasma producing an increase of detected pulse which is proportional to the size of detected nanomaterial	Height of intensity of the pulse (used as equivalent to diameter)	Mass based
Tunable Resistive Pulse Sensing (TRPS) (29-31)	Measurement of resistive pulse sensing induced by nanomaterials crossing through a size tunable pore (based on the Coulter counter principle)	Raw diameter	Number based

silver nanoparticles using (i) batch particle size measurement method i.e. DLS and separative particle size measurement methods i.e. DCS and Sedimentation Field Flow Fractionation (SdFFF) (37). The separative particle size measurement methods were able to discriminate the two

populations. However, the resolution was different because in one case, the direction of the separation was orthogonally performed to the elution (SdFFF), and in the other case, the directions of the separation and the displacement were parallel (DCS). In contrast, the investigated batch particle size

Table III Main Characteristics of Separative Particle Size Measurement Methods

Method	Principle	Measurand	Type of size distribution of raw data
Capillary Electrophoresis (CE) (19,32)	Separation achieved by electrophoresis phenomenon and based on diameter and charge density	Apparent mobility (migration time)	Detector dependent
Differential Centrifugal Sedimentation (DCS) known as Centrifugal Liquid Sedimentation (CLS) (27)	Measurement of sedimentation time. Separation according the sedimentation rate before detection	Light extinction as a function of sedimentation time Sedimentation diameter (Stokes analysis)	Extinction intensity based
Field Flow Fractionation (FFF) (19,27,28,32)	Separation achieved through the interaction of nanomaterials with an external physical field (no stationary phase) and based on elution mode. <i>Flow: hydrodynamic diameter, Sedimentation: equivalent spherical volume diameter, Thermal: diameter and chemical composition, Electrical: diameter and charge density</i>	Retention time	Detector dependent
Hydrodynamic Chromatography (HDC) (27,32)	Separation achieved by flow velocity and the velocity gradient across the particle and based on hydrodynamic diameter	Retention time	Detector dependent
Size Exclusion Chromatography (SEC) (27,32)	Separation achieved through differential partitioning between the mobile and porous stationary phase and based on hydrodynamic diameter	Retention time	Detector dependent

measurement method provided with a monomodal distribution with an intermediate value of hydrodynamic diameter between the two real sizes of the nanoparticles included in the model multimodal dispersion but closer to the largest particles. These examples underlined the limits of the DLS method to resolve nanoparticles having less than a factor of 3 in size difference as described previously in the literature (41). While the previous experiences were all performed with model multimodal dispersions of known composition and size distribution, Sokolova *et al.* pointed out the difficulty to obtain relevant PSD measurement from a sample of nanoparticles of the “real life” (34). The authors characterized size of exosomes from three different cell types using methods from two classes: batch particle size measurement method i.e. DLS and single particle size measurement method i.e. Scanning Electron Microscopy (SEM) and PTA. They reported that the size of exosomes was not accurately determined using DLS because of the polydispersity of the sample and the likely weak scattering contrast provided by the nature of the nano objects. The work of Ingebrigtsen *et al.* dedicated to the analysis of the PSD of a liposomes preparation as a “real life” sample, explored the relevancy of the separative particle size measurement method as Size Exclusion Chromatography before measuring PSD using DLS (39). The reports also proved the usefulness to employ a separative particle size measurement method to resolve the difficult task evaluating the PSD of heterogeneous samples. All size measurement methods may provide biased representation of the true PSD of a given sample (28). A careful interpretation of their raw data is needed to investigate PSD measurement. Application of the electron microscopy (EM) that is an established direct method based on measurements done on images of particles and that is recommended as reference method in the standard ISO (42) and by the Health Agencies (18) also shows limitations. It requires an homogeneous and representative deposit of well individualized particles on the grids to allow measurements on a minimum of thousand particles to obtain a statistically significant PSD (43). The stringent quality of deposition of particles on the grid needed to perform relevant size and PSD measurement by TEM could not be fulfilled considering a sample of poly(isobutylcyanoacrylate) nanoparticles (PIBCA nanoparticles) decorated with dextran having a multimodal distribution (44). A segregation between large and small particles occurred systematically during deposit on grids for TEM as shown in Fig. 1. Also reported elsewhere (43), this hampers the establishment of unbiased PSD analyzing images obtained by EM. With this sample even the most straightforward method is unsuitable to achieve accurate evaluation of the PSD of the multimodal dispersion. Consequently, the evaluation of PSD of this sample appeared as a complex task. Thus, the aim of the present work was to achieve a comparative analysis using 9 different commercially available methods and to provide with a comprehensive critical evaluation of the results provided by

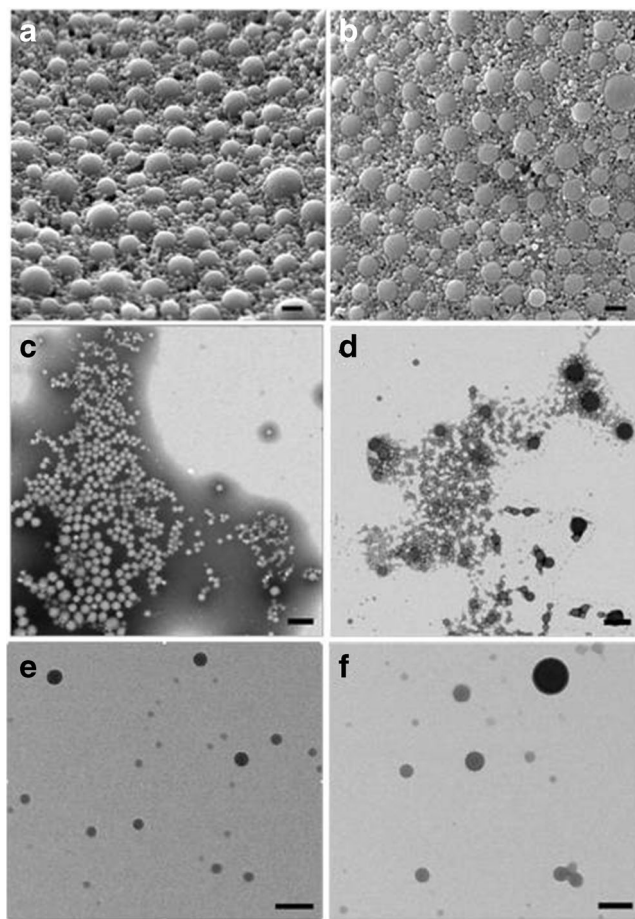


Fig. 1 Electron micrographs obtained from the analysis of the dispersion. (a) and (b): scanning electron microscopy performed on lyophilizates obtained from two independent preparation of the dispersions. (c, d, e and f): Transmission electron micrographs obtained after negative staining of sample deposited on an ionized grid (c) and a non ionized grid (d, e, f). Scale bar: 200 nm.

the methods that were taken from the different classes defined above. It was aimed to establish advised based on experimental facts to resolve the difficult task of size and PSD evaluation of unknown dispersions based on the analysis of a real sample of polymer nanoparticles obtained from a synthesis based on emulsion polymerization and developed as nanomedicine.

PRINCIPLE OF INVESTIGATED METHODS FOR PARTICLE SIZE DETERMINATION

Single Particle Size Measurement Methods

Direct Methods

Electron Microscopy. Electron Microscopy (EM) is a direct method described in the standards ISO (42) and recognized by Health Agencies to perform size and PSD measurement

(18). EM measures equivalent spherical diameter or Feret's diameter of nanometer sized nanoparticles by imaging samples based on the diffraction of electron beam passing through thin section of samples. EM is a counting method taking account of polydispersity of samples by producing a size value for each nanoparticle selected on images for analysis. Therefore, size distribution by number is mounted. It is noteworthy that capturing images implying both larger and smaller nanoparticles with unbiased and establishment of a representative distribution is difficult. Many hundreds or thousands particles depending on the broadness of the PSD should be measured to provide a statistically significant particle size distributions as suggested by the standard ISO (43). To increase number of screened nanoparticles, automated analysis software can be used. It requires to define measurements parameters that may induce bias in particle size distributions (45,46).

Atomic Force Microscopy. Atomic Force Microscopy (AFM) belongs to the family of scanning probe microscopes. AFM is recognized by Health Agencies to perform size and PSD measurement (18). Since its invention in 1986 by Binnig *et al.* (47), AFM has been widely used in nanomaterials science where microstructure or nanostructure needs to be determined. A standard AFM is composed of a flexible cantilever with a sharp tip (typically tip radius around 10 nm) at its end and a piezoelectric scanner which can move the probe very precisely in x, y and z axes and thus controls the interaction between the tip and sample. The deflection of the cantilever normal to the sample surface is monitored by means of a laser reflection on the back side of the cantilever based on the optical lever method proposed by G Meyer *et al.* (48). The small changes in cantilever deflection are thus detected with a position sensitive photodiode detector. This deflection is then processed by the system electronics by a feedback mechanism in order to maintain the same tip sample distance and thus to determine topological changes on the sample surface. The resolution of AFM in Z direction is very high in the sub Angstrom range; however lateral resolution is limited by the tip radius which is usually in the order of 10 nm. AFM is a counting method taking account of samples polydispersity by producing a size value for each nanoparticle selected on images for analysis. Therefore, size distribution by number is mounted. As for EM, capturing images implying both larger and smaller nanoparticles with unbiased and representative distribution is difficult. A large number of well separated particles are needed to provide a statistically significant particle size distribution. Automated image processing software may be used. But, it requires some assumptions about the nanoparticle shape to increase number of screened nanoparticles over time (49). Size information may be provided by means of two different approaches. The most commonly used approach consists in considering the height of the spherical nanoparticles as diameter. Such approach may be applied to determine the size of

well separated nanoparticles. However, precautions should be taken into account when analyzing AFM images with this approach due to the elastic deformation in Z direction of the imaged features when applying high forces and to the interaction of nanoparticles with the underlying substrate which may induce changes in the polymeric nanoparticle shape. The other approach consists in analyzing AFM images in the lateral dimension (x y). However, it should be noted that the quality of AFM images highly depends on the (i) sample preparation method to get well separated features, (ii) the smoothness of the underlying substrate, (iii) the applied force and (iv) the shape and the radius of the tip. Indeed, the shape of the tip may affect the accuracy of the lateral information for very small nanoparticles.

Indirect Methods

Nanoparticle Tracking Analysis. Nanoparticle Tracking Analysis (NTA) utilizes the properties of both light scattering and direct evaluation of the Brownian motion of particle in order to analyze particle size distributions of samples in liquid dispersion. A laser beam is passed through a prism edged glass flat within the sample chamber containing the nanoparticle dispersion. Once in contact with the laser, the nano objects scatter light. This signal is then collected *via* a microscope pathway mounted with a camera working in the black field mode. The camera records the movement of the particles under the Brownian motion. The NTA software simultaneously tracks particles as they move in the plane of focus and calculates the mean square displacement (in two dimensions) of each particle to obtain its translational diffusion coefficient. The translational diffusion coefficient is converted into a size *via* the Stokes Einstein Equation while temperature and viscosity of the dispersing medium are known. Since the method is particle by particle and not a batch method, the size distribution profiles display a higher peak to peak resolution. In addition, this microscope based technology allows the rapid measurement of the number of particle based concentration of the sample. The tracking method should resolve individual population present in multimodal samples due to the ability to individually follow particles reducing the influence of the scattered light intensity of larger particles hence bias of the measured PSD (50).

Tunable Resistive Pulse Sensing. Tunable Resistive Pulse Sensing (TRPS) is a method based on a size tunable pore, to measure particle size, charge and concentration by resistive pulse sensing. One of the first reported works on the technology, as described by Sowerby *et al.* (29), shows that it is possible to make a resizable aperture by penetrating an elastomeric membrane with a coned shaped needle probe. Based on the Coulter counter principle, the method involves measuring the increased electrical resistance induced by the passage of nano

and micro particles through the pore, immersed in a conductive liquid. The larger the volume of the particle is, the higher the displaced volume of electrolyte and hence resistance, will be. The rate of particle translocation is a function of concentration and electrophoretic mobility, being the main driving force of particles in aqueous solution as they pass through the pore, which also gives precise concentration and charge measurements on each counted particle, respectively, as an electric field is applied across electrodes (30).

This method requires a calibration with carboxylated polystyrene particles of known size, concentration and charge, based on the linear relationship between the variation of resistance and particle volume. Hence a single point of calibration is needed to provide an accurate size measurement, if thereafter measurement conditions remain constant (31).

Batch Particle Size Measurement Methods

Dynamic Light Scattering

Dynamic Light Scattering (DLS) or Photon Correlation Spectroscopy (PCS) is described in the standard ISO 22412:2008(E) (51) and recognized by Health Agencies to perform size and PSD measurement (18). It enables the evaluation of hydrodynamic diameter of particles dispersed in liquid by the measurement of translational diffusion coefficient characterizing the Brownian motion. The hydrodynamic diameter is then calculated using the Stokes Einstein equation. In a usual DLS setup, a laser light illuminates the sample which scatters light in all direction with fluctuations due to the displacement of the nanoparticles depending on the time. The analysis over the time of these fluctuations leads to an autocorrelation function, fitted with an exponential behavior. Parameters of the decrease of the exponential decay rate are related to the diffusion of the particles in the dispersion medium. Two popular algorithms are usually used: Cumulants and Contin. Cumulants, described in the standard ISO (51) is suited for sample composed of particles homogeneous in size with a low standard deviation value. This means that the autocorrelation function is fitted with a single exponential assuming that the sample is monodisperse. Contin, based on the Non Negative Least Squares (NNLS) method, is applied for multimodal samples but limited by the characteristics of the sample (range, mono/polydispersity). Usual DLS instruments are designed to perform measurements at a fixed detection angle (90 or 173° with respect to the laser illumination axis) or on a wide range of angles depending on the settings. Varying the detection angle is interesting for the analysis of heterogeneous samples as small and large particles scattered light proportionally to the power six of their radius. The intensity of the scattered light contribution of large and small particles depends on the angle of measurement allowing to privilege detection of different size according to the angle of measurement. DLS is suitable for light being

scattered once. Multiple scattered light can lead to bias results and misinterpretations. To overcome this technical limitation, novel instruments based on dynamic light scattering followed by cross correlation of photons (Photon Cross Correlation Spectroscopy, PCCS) have been developed. Measurements can be achieved on concentrated samples as multiple scattering effects are considerably reduced in accordance with recommendation of the standard ISO (51). In practice, two laser beams illuminate the same scattering volume and intensity fluctuations are recorded by two independent detectors. Multiple scattered lights are discarded by cross of the two separated autocorrelation functions obtained by each detector.

Static Light Scattering

Static Light Scattering (SLS) also called Laser Diffraction (LD) enables to evaluate the radius of gyration of nanoparticles dispersed in liquid by measuring the angular dependence of the intensity of scattered light. Larger particles scatter much light at small angles whereas smaller particles scatter light at greater angles. In a usual SLS setup, a laser light illuminates the sample which scatters light at one or many angles. The evolution of the scattered intensity as a function of detected angles depends on particle size. The analysis of the optical signal leads to the deconvolution of scattering pattern into a series of individual number (one per size classification). The relative amplitude of individual number corresponds to the relative volume of equivalent spherical particles of that size. This deconvolution may be performed by means of either the Fraunhofer or Mie theories of light scattering.

Separative Particle Size Measurement Methods: Asymmetrical Flow Field-Flow Fractionation Coupled With Dynamic Light Scattering

Field Flow Fractionation is another method recommended by Health Agencies to perform size and PSD measurement (18). Asymmetrical Flow Field Flow Fractionation (AsFFFF) is a liquid phase separation method that enables to fractionate nanoparticles based on their hydrodynamic size and appears as a powerful tool for obtaining high resolution information on the size distribution of a nanoparticle dispersion (52,53). Briefly, AsFFFF is performed in stationary phase free channel in which the sample i.e. a dispersion of nanoparticles is carried through a narrow channel in a laminar parabolic flow profile with eluent. A perpendicular flow field is applied to this carrier flow that drives nanoparticles to the accumulation wall of the channel where they accumulate and move slower. The separation of nanoparticles is then achieved according to their translational diffusion coefficient. Due to Brownian motion, the smaller nanoparticles having higher translational diffusion coefficient tend to reach an equilibrium position (diffusion against the applied field) at a larger distance from the

accumulation wall where they move faster. Thus, the gradient of velocity that establishes in the channel enables the separation of nanoparticles with different sizes. In the normal mode, the smaller particles move faster in the channel than the larger particles hence they are eluted before the latter ones. Combining the Stokes Einstein Equation and the relationship between the translational diffusion coefficient and the retention time, the hydrodynamic diameter may be evaluated. Different detectors may be coupled to AsFFFF for unambiguous identification and quantification of nanoparticles.

MATERIALS AND METHODS

Materials

Deionized and ultrapure water (MilliQ®) were obtained from Millipore water system.

PIBCA nanoparticles were prepared with isobutylcyanoacrylate (IBCA) from Orapi, Dextran (66.7 kDa) from Sigma, Cerium (IV) ammonium nitrate and nitric acid (purity between 61.5 and 65.5%) from Prolabo.

Preparation of Poly(Isobutylcyanoacrylate) Nanoparticles Decorated with Dextran

Synthesis of PIBCA nanoparticles decorated with dextran was performed according to Vauthier *et al.* by redox radical emulsion polymerization (44). Briefly, dextran (0.0502 g) was dissolved in 9.3 mL of aqueous 0.2 N nitric acid in a glass tube at 40°C under argon bubbling. Vigorous stirring was applied to create a vortex. After 10 min, 0.7 mL of cerium (IV) ammonium nitrate 8.10^{-2} M in aqueous nitric acid 0.2 N was added followed immediately by 0.5 mL of IBCA. The reaction was left to continue for 1 h. After cooling down in an ice bath, milky dispersion of PIBCA nanoparticles was purified by dialysis (Spectra/Por® membrane Biotech, molecular weight cutoff of 100,000 Da, Spectrum Laboratories) twice against 1 L of deionized water for 30 min, once for 6 h and the last overnight. The purified dispersions were stored at 4°C before using. The concentration in nanoparticle of the dispersion was 51.1 ± 1.2 mg.mL⁻¹ as determined by gravimetry.

Evaluation of the Size Characterization

Single Particle Size Measurement Methods

Direct methods.

Electron Microscopy

For SEM, a sample of the nanoparticle dispersion was freeze dried and deposited on a sample holder for SEM with

a double faced tape. The sample was then coated with a layer of 2 nm of Pt/Pd using a Cressington Sputter Coater 208HR instrument (Cressington). Observations were performed using a scanning electron microscope MEB LEO 1530 (LEICA) equipped with a Gemini type column at the CNRS CECM (Vitry sur Seine, France).

For TEM, nanoparticles were deposited on a formvar/carbon coated copper grid (Agar Scientific) either by flotation for 3 min on dilution of the dispersion adjusted at a concentration in nanoparticles of 0.5 mg.mL⁻¹ or by direct deposition of 3 µL of the diluted dispersion on the grid. Both ionized and non ionized grids were used. After drying of the sample, grids were stained by flotation over a solution of phosphotungstate at 1% for 30 s. The nanoparticles were imaged using a JEOLJEM1400 electron microscope equipped with a camera ORIUS SC1000 1.

Atomic Force Microscopy

The AFM imaging of PIBCA nanoparticles was performed in ambient conditions using a JPK Nanowizard® 3 Ultraspeed AFM from JPK instruments in amplitude modulation AFM (AM AFM) with low force settings (80–90% of the free amplitude). In AM AFM modulation, the tip surface distance regulation is performed to maintain constant the amplitude to a precised setpoint. Gold coated silicon cantilevers PPP NCHAuD with a spring constant of ~ 42 N.m⁻¹ and a tip curvature radius of ~ 10 nm (Nanosensors) have been used. Nanoparticle samples were diluted at a concentration of 0.1 mg.mL⁻¹ of nanoparticles with filtered ultrapure water, and 100 µl of this solution was deposited on freshly cleaved mica substrates and kept to dry overnight at 22°C and ambient humidity. AFM images were processed and analyzed to evaluate PSD of nanoparticles including 393 counted nanoparticles using JPK Data processing software (JPK instruments). To reduce the convolution effect on the lateral size of imaged nanoparticles, the widths were measured manually at the full width at half maximum (FWHM) of the peak height using the line profile measurement option.

Indirect methods.

Nanoparticle Tracking Analysis

Evaluation of the size characterization of the PIBCA nanoparticle dispersion was performed by NTA using a Nanosight NS300HSB device (Malvern Instruments) equipped with a sCMOS camera, a 405 nm laser and a syringe pump. The software version used for capturing and analyzing the data was the NTA 3.0.

Six measurements of the same sample of PIBCA nanoparticles were performed. Measurements were performed under a regular flow (using the NanoSight Syringe Pump addon)

which allows the tracking of new particles all along the analysis. The sample of PIBCA nanoparticles was diluted in filtered ultrapure water using a 0.22 μm filter (Roth) with a 10^6 factor (v/v) ($51.1 \text{ ng}\cdot\text{mL}^{-1}$).

Tunable Resistive Pulse Sensing

All measurements were made using an Izon qNano (NZ). Polystyrene calibration particles with a concentration of $1.2 \times 10^{13} \text{ particles}\cdot\text{mL}^{-1}$ and mode size of 115 nm were purchased from Thermo Fischer Laboratories. Sample particle diameters were calculated using a calibration process (47). Mode diameter of the calibration particles stands for the position of the maximum of the size distribution. All samples were dispersed in phosphate buffered saline containing 0.03% of Tween 20 (IZON solution Q) for analysis at a dilution factor of $1/10^3$ (v/v) ($51.1 \text{ }\mu\text{g}\cdot\text{mL}^{-1}$). Particle concentration and size were calculated using Izon Control Suite Software v 3.2 on a minimum of 500 particle events.

Batch Particle Size Measurement Methods

Dynamic Light Scattering. Up to 4 different DLS were used to evaluate PSD of the PIBCA nanoparticle dispersion: single angle DLS working at 90 or 173°, multiangle DLS and PCCS.

Single Angle DLS

The hydrodynamic diameter of PIBCA nanoparticles, D_h , was measured at 25°C by DLS using two different Zetasizer Nano ZS from Malvern. The first one was a Zetasizer Nano ZS operating at fixed scattered angle at 173° using Zetasizer Software version 7.04. The second one was a Zetasizer Nano ZS 90 operating at fixed scattered angle at 90° using Zetasizer Software version 6.11. Both instruments were equipped with a laser source of wavelength of 633 nm. The sample of PIBCA nanoparticles was diluted in filtered ultrapure water using a 0.22 μm filter (Roth) by $1/200$ (v/v) ($0.256 \text{ mg}\cdot\text{mL}^{-1}$) at room temperature. Measurements were performed in macrocuvette with four optical faces (VWR) controlled for spotting surface scratches or coatings that could interfere with optical measurements, rinsed three times with filtered ultrapure water and stored in a dust free environment until use. Appropriate volume of samples (1 mL) was introduced into measurement cells and placed in the instrument for equilibration time of 300 s to ensure temperature homogeneity prior to making 3 measurements.

Multiangle DLS

Size measurements of PIBCA nanoparticles were performed at 25°C by multi angle DLS using a NanoDS (Cilas)

using NanoExpert version 10.36 with detection from 10 to 150° equipped with a laser source of wavelength of 638 nm. For measurements, the sample of PIBCA nanoparticles was diluted in filtered ultrapure water using a 0.2 μm filter (Whatman) by $1/3000$ (v/v) ($17.0 \text{ }\mu\text{g}\cdot\text{mL}^{-1}$) at room temperature. The obtained diluted dispersion (3 mL) was prepared in a standard quartz cell and placed in the instrument for an equilibration time of 120 s prior to the measurements.

Photon Cross Correlation Spectroscopy

Evaluation of the size characterization PIBCA nanoparticle dispersion was performed by photon cross correlation spectroscopy using a Nanophox (Sympatec) using Windox software version 5.8.7.0 at 25°C. The instrument was equipped with a laser source working at a wavelength of 658 nm and the detection was achieved at the fixed scattered angle at 90°. The sample was placed in macrocuvette with four optical faces (Sarstedt) inspected for any default, rinsed three times with filtered ultrapure water and stored in a dust free environment before use. Appropriate diluted sample of PIBCA nanoparticles in filtered ultrapure water by $1/20$ (v/v) ($2.56 \text{ }\mu\text{g}\cdot\text{mL}^{-1}$) at room temperature was introduced into measurement cells (volume of 1 mL) and placed in the instrument for an equilibration time of 30 s. Five measurements were carried out on the sample.

Static Light Scattering. Size measurements of the PIBCA nanoparticle dispersion were performed at 25°C by SLS using a LS 13 320 apparatus (Beckman Coulter) and LS 13 320 software v 6.01 at different scattering angles (0.03 to 35°) to determine the diameter of gyration, D_g . The instrument was equipped with Polarization Intensity Differential Scattering (PIDS) to measure differences between horizontally and vertically radiated light for several wavelengths (450, 600, and 900 nm), which were horizontally and vertically polarized at multiple angles. The instrument was equipped with a laser source of wavelength of 780 nm and measurements were performed in Universal Liquid module of 125 mL. Appropriate diluted sample of PIBCA nanoparticles in filtered ultrapure water (3 drops of the dispersion were added to 125 mL of filtered ultrapure water, dilution $\sim 1/83$, $61.3 \text{ }\mu\text{g}\cdot\text{mL}^{-1}$) was introduced into Universal Liquid module at room temperature. This module was then placed in the apparatus. Three measurements were carried out on the sample.

Separative Particle Size Measurement Methods: Asymmetrical Flow Field-Flow Fractionation Coupled With Dynamic Light Scattering

The Asymmetrical Flow Field Flow Fractionation (AsFFFF) were performed on an Eclipse 2 System (Wyatt Technology Europe). The AsFFFF channel was installed with a 350 μm

spacer and a regenerated cellulose membrane with a 10 kDa cutoff (Wyatt Technology). The carrier flow was delivered by an Agilent 1100 Series Isocratic Pump (Agilent Technologies) with an in line vacuum degasser. The carrier liquid composed of deionized water containing 0.02% sodium azide (Sigma Aldrich) was filtered using vacuum filtration system (Gelman filters of 0.1 μm). Samples of PIBCA nanoparticles were diluted at 1/100 with carrier liquid and mixed using a vortex (Scientific Industries) at 2500 rpm for 10 s prior injection into the channel. During focusing, the cross flow was fixed at 1.5 mL.min⁻¹ for 2 min. The sample (20 μL) was then injected into the channel using an Agilent Autosampler at 0.2 mL.min⁻¹ for 5 min. After injection, 2 min of focus was applied before starting the elution. PIBCA nanoparticles were eluted using cross flow rate fixed at 0.3 mL.min⁻¹, a channel flow fixed at 1.0 mL.min⁻¹ for 45 min and monitored by a 18 angles multi angle light scattering (MALS) instrument DAWN HELEOS II (Wyatt Technology) equipped with a Laser source at 633 nm and a QELS (DLS) at 99°, an Optilab Rex Refractometer (Wyatt Technology) and a UV detector Agilent 1100 operating at 254 nm. Measurements were performed at room temperature. The software version installed on AsFFFF chain was the Astra v 5.3.4.20.

RESULTS

Size characterization of a nanoparticle dispersion that appeared quite heterogeneous by EM was evaluated using different instruments working on different principles. Most of them are methods recommended by the standard ISO (42,51) and by the Health Agencies (18). The results are given below, method by method, considering whether they are single particle measurement methods, batch particle measurement methods or measurement methods applied after a separation of particles of the dispersion according to their size with an appropriate method. All results described the nanoparticle size expressed in diameter calculated from the measurand of the corresponding method using models explained in the Materials and Methods when required.

Single Particle Size Measurement Methods

Direct Methods

EM can be used to determine size characteristics of nanomaterials based on the measurement of the size of each nanoparticle contained in the dispersion and seen on samples observed by EM. It is considered as a direct size particle measurement method. To achieve these determination, deposition of the nanoparticles on the sample holder should fulfill some requirements. Particles should be well dispersed on the holder and randomly distributed. Figure 1 showed electron

micrographs of nanoparticles contained in the dispersion studied in the present work. Observed freeze dried samples by SEM obtained from two independent preparations of the dispersion revealed the presence of at least two populations of nanoparticles. Size measurement was not performed because of the high density of the deposit of particles on the sample holder found on the electron micrographs. The presence of large and small particles was confirmed from TEM observations on samples deposited by different methods on either ionized or non ionized grids. Areas on grids showed well isolated nanoparticles as required by the standard ISO to perform measurements of nanoparticle size and to access the size distribution of the dispersion (28) (Fig. 1e and f). However, by investigating sample for TEM observation on a systematic manner, it clearly appeared that a segregation between large and small particles occurred during the preparation of the grids compromising an objective determination of the size distribution. Although present in a lower number, larger nanoparticles were almost not found on areas included isolated particles. They were almost exclusively seen in clusters (Fig. 1d) while the majority of the isolated particles corresponded to the smallest nanoparticles (Fig. 1e and f). The larger particles appeared in the center of clusters while small particles accumulated at the periphery of large particles with the smallest nanoparticles located at the external border (Fig. 1d). These systematically observed arrangements may results of an artifact that occurred during the drying of samples on grids whatever was the method of deposition. As the deposition of the nanoparticles on the grid could not be random, it compromised the evaluation of the size characteristics of the dispersion which appeared highly heterogeneous on a qualitative basis. From EM observations of the sample, it could be concluded that the dispersion contained various populations of nanoparticles that differed in their size.

PSD of PIBCA nanoparticles contained in the sample was investigated using AFM. AFM images showed well dispersed particles on the holder that allowed to determine the number based size distribution of in the dispersion (Fig. 2). In contrast with EM, apparently no segregation between large and small particles occurred during deposition of the dispersion and acceptable density of particles was found on images enabling an accurate determination of the size distribution. The PSD highlighted several populations of nanoparticles with a major population in number having a diameter of 95.0 nm and an asymmetric Gaussian showing several secondary peaks at 135, 150, 175 and 195 nm. Minor separated populations appeared at 245, 275 and 335 nm. A few large particles were present in the multimodal dispersion around 400–600 nm. No particles with size lower than 60 nm was highlighted, indicating thus that the radius of the AFM tips sufficiently small to resolve such features. The dispersion contained various populations of nanoparticles with different size consistently with the quite complex polydispersity of the dispersion revealed by TEM on a qualitative basis.

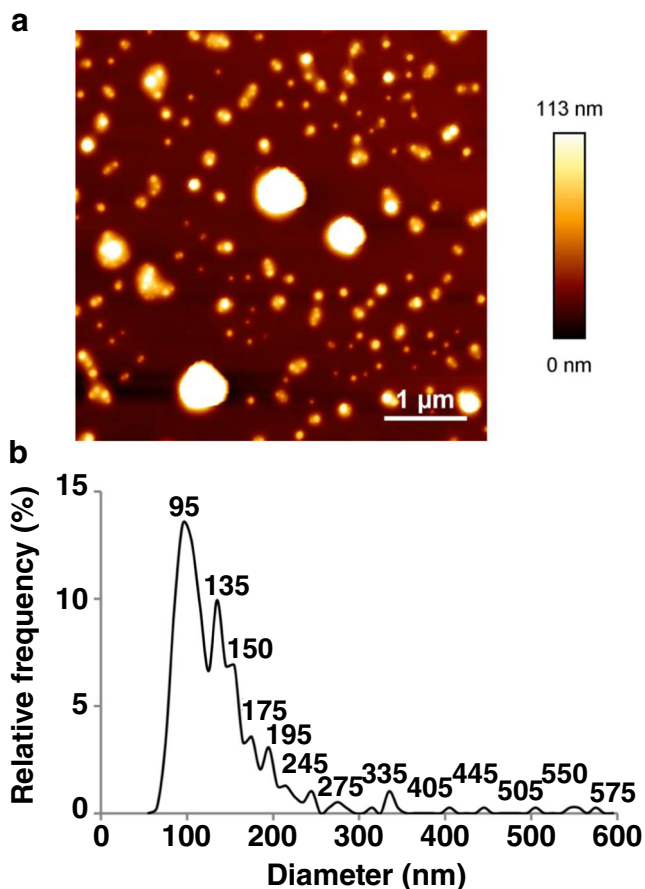


Fig. 2 AFM image (a) and number based size distribution (b) obtained from the analysis of the dispersion. Size distribution obtained from the measurement of 393 nanoparticles.

Indirect Methods

The PSD of PIBCA nanoparticles was recorded using NTA. The Fig. 3a showed the corresponding number based distribution of nanoparticles contained in the dispersion. The PSD highlighted one population of nanoparticles around 67.0 nm with an asymmetric Gaussian with several secondary peaks at 97.0, 143 and 217 nm. A few large particles were present in the multimodal dispersion in the range 355 to 500 nm.

The PSD of PIBCA nanoparticles was investigated using TRPS. The Fig. 3b showed the corresponding number based distribution of nanoparticles contained in the dispersion. The PSD highlighted one population of nanoparticles around 109 nm with an asymmetric Gaussian with several secondary peaks at 122, 135 and 188 nm. A few large particles were present in the multimodal dispersion around 300 nm. The diameter of 10% (D10) and 90% (D90) of the population of the multimodal dispersion was less than 97 and 190 nm respectively. The ratio D90/D10 was equal to 2 showing the polydispersity of the dispersion.

For both methods, the PSD of PIBCA nanoparticles was consistent with the PSD provided by AFM.

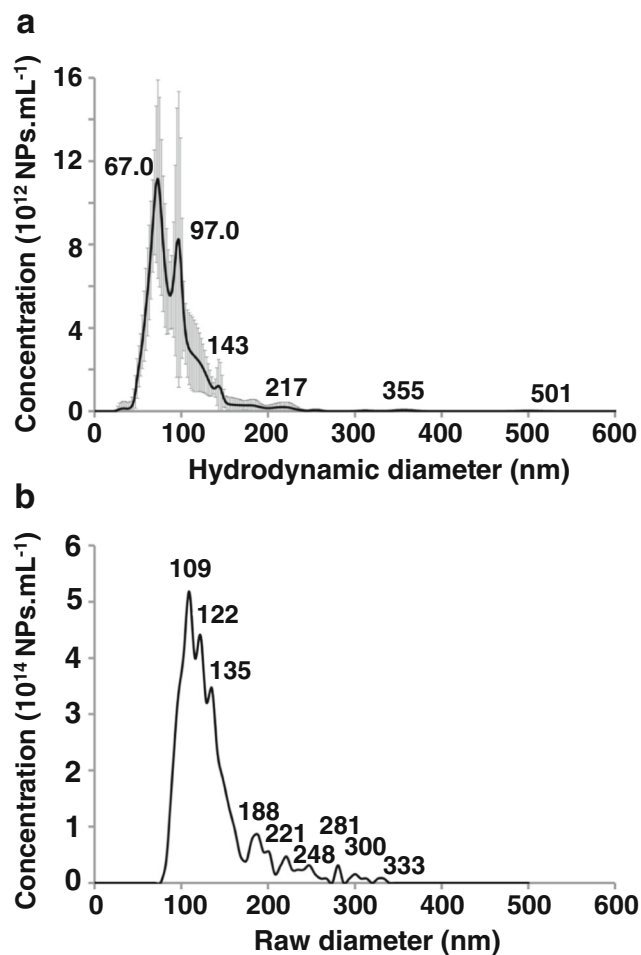


Fig. 3 Number based size distribution obtained from the analysis of the dispersion for NTA (a) and TRPS (b).

Batch Particle Size Measurement Methods

PSD of PIBCA nanoparticles was investigated using different DLS. The Fig. 4a and b showed the intensity based distribution of nanoparticles contained in the dispersion obtained from DLS working at fixed angle of 90 and 173° respectively. The PSD provided by the DLS method working at fixed angle showed only one population of nanoparticles with narrow dispersity suggesting a monomodal PSD. Mean D_h determined from the two apparatus were 220 and 190 nm, for the angle of measurement of 90 and 173° respectively. The results were consistent with known performances of the measurement method. At the low angle, the smallest particles were predominantly detected. The contrary was observed with the detection at 90°. The Fig. 4c showed the intensity based distribution of nanoparticles contained in the dispersion obtained from DLS multi angle. The optimum angle was found at 120° to perform size measurement that would be closed to the iso intensity between the different populations and thus show smaller populations. Two populations that differed in their size (35.0 and 199 nm) were observed on the PSD. It is noteworthy that the peak of the lower

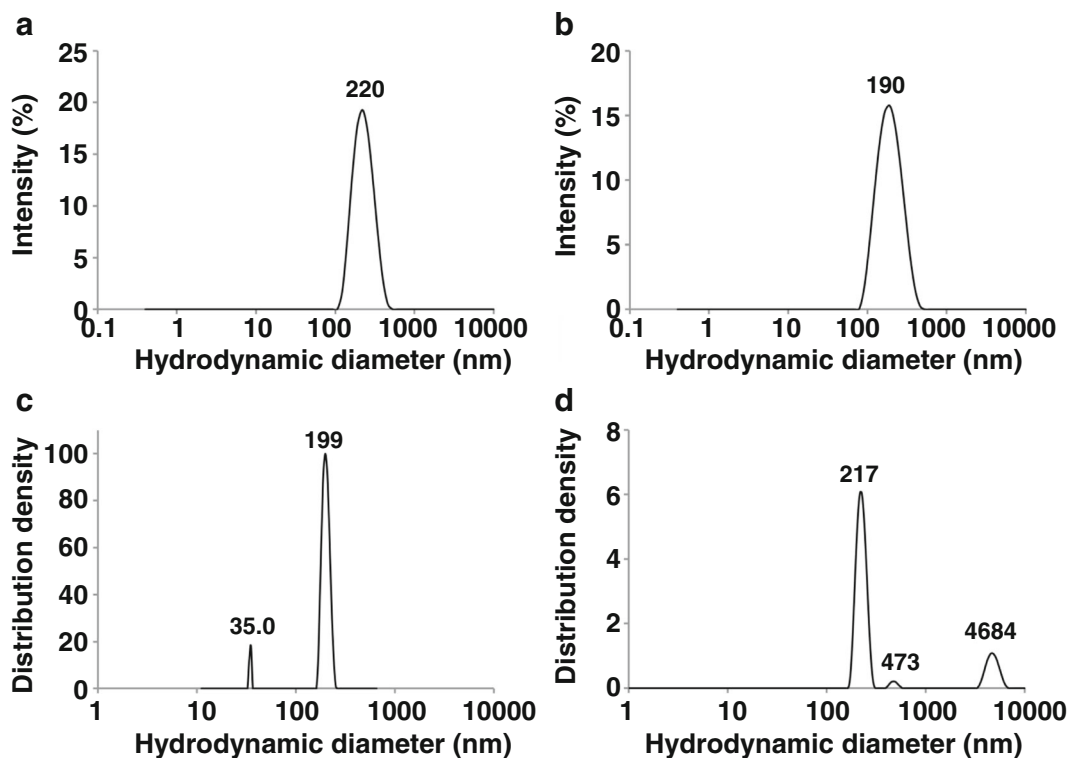


Fig. 4 Intensity based size distribution obtained from the analysis of the dispersion for (a) DLS 90° (NNLS), (b) DLS 173° (NNLS), (c) Multi angle DLS, optimal angle: 120° (Contin) and (d) PCCS (NNLS).

population was not repeatable that of the larger population appeared at a size that was consistent to that revealed by the two previous DLS methods working at fixed angle. The Fig. 4d showed the intensity based distribution of nanoparticles contained in the dispersion obtained from PCCS. Three populations that differed in their size (217, 473 and 4684 nm) were detected on the PSD. This instrument revealed populations of larger size while the population at 217 nm could correspond to that also detected with other DLS apparatus. Results obtained by light scattering methods in batch were provided with very simple PSD compared with those that could deduced from AFM and other single particle size measurements methods.

PSD of PIBCA nanoparticles was recorded using SLS. The Fig. 5 showed the corresponding volume based distribution of nanoparticles contained in the dispersion. The PSD showed interestingly three well separated populations of nanoparticles characterized by Gaussian at 63.8, 195 and 496 nm. SLS revealed a quite complex PSD characteristics of the PIBCA nanoparticles although it appeared more simple than that depicted by AFM and the other single particle size measurement methods.

Separative Particle Size Measurement Methods: Asymmetrical Flow Field-Flow Fractionation Coupled With Dynamic Light Scattering

Separative particle size measurement methods may provide with a more resolute PSD while analysing unknown

dispersions of nanoparticles. PIBCA nanoparticle dispersion was then analysed by AsFIFFF coupled with a DLS detector. The Fig. 6a showed the obtained fractogram. The elution profile of PIBCA nanoparticles recorded by DLS detector showed a broad PSD with 4 peaks having maxima at elution time of 18.0, 23.9, 32.4 and 36.2 min. These peaks indicated the presence of particles of significantly different size as shown by their measured hydrodynamic diameter (97.0, 168, 417 and 446 nm). The multimodal dispersion was well identified by the AsFIFFF. The number based size distribution (Fig. 6b)

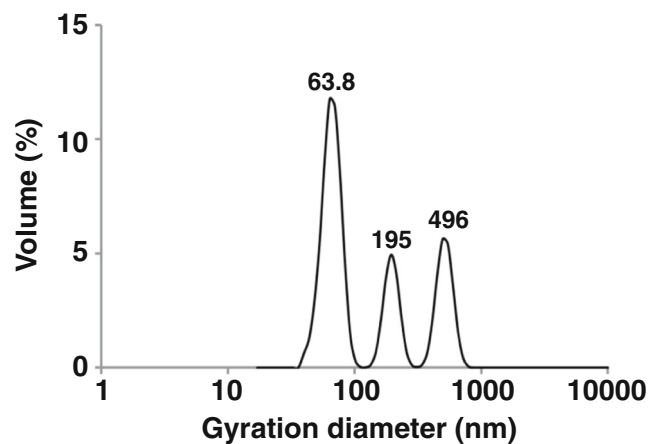


Fig. 5 Volume based distribution obtained from the analysis of the dispersion for SLS (Mie theory).

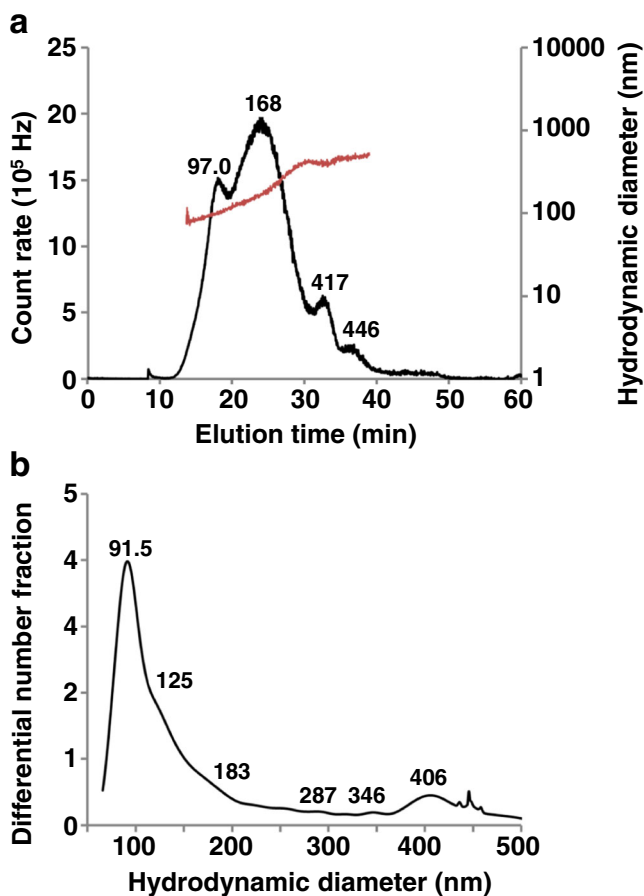


Fig. 6 AsFIFFF fractogram of PIBCA nanoparticles recorded by DLS detector working at 99° (a) intensity based size distribution and (b) number based size distribution (Unit of differential number fraction: $1/\log(\text{nm})$).

deduced from the analysis was consistent with images of microscopy and PSD provided by AFM and single particle size measurement method.

DISCUSSION

A dispersion of PIBCA nanoparticles prepared by emulsion polymerization appeared quite heterogeneous from observations by electron microscopy. The size characteristic of the dispersion was first evaluated by DLS which is a widely used method in laboratories due to its apparent ease of use, the short duration of the measurements and the availability of accessible marketed apparatus. The dispersion was characterized by DLS working at two detection angles, 90 (DLS 90°) and 173° (DLS 173°). Both have provided with a single population of particles with mean hydrodynamic diameters at 220 and 190 nm respectively. Results provided by the DLS instruments were not satisfied as it seemed that they did not reflect the heterogeneity of the dispersion shown on electron micrographs. Although not satisfying, they agreed with the known fact that determination of size characteristics of

complex dispersions by DLS is problematic (33,34,38) while the method is powerful to evaluate size characteristics of monomodal dispersions with a high accuracy (21). The limitation comes from the fact that the intensity of scattered light depends on particle size being proportional to the power six of the particle diameter. Thus, intensity of scattered light due to large particles can hinder the detection of populations of particles having a smaller diameter. Due to a segregation that occurred during preparation of the samples for observation by EM, PSD of the PIBCA nanoparticle dispersion could also not be determined by this direct method of measurement. Size characteristics of this dispersion was then studied by different methods including a series of light scattering methods that can be classified as batch methods, a series of methods that are based on the determination of the size characteristics of single particles and a method that include a separation process as the function of the nanoparticle size prior to achieve size measurements by a light scattering method that gives accurate results on homogeneously dispersed monomodal dispersion.

The Fig. 7 summarizes the position of peaks on a scale reporting the diameter of the corresponding nanoparticle populations given from the analysis of the PSD of the dispersion of PIBCA nanoparticles by the different particle size measurement methods considered in the present work.

Results obtained from single particle size analysis, i.e. AFM, TRPS, NTA, and based on a separative method, i.e. AsFIFFF, were plotted on the upper part of the graph while those produced by batch methods based on light scattering were plotted on the lower part of the graph. Methods considered in the upper part of the graph revealed the presence of several populations of nanoparticles of different diameters suggesting that the dispersion had a quite complex PSD. Graphs of size distributions obtained from all these methods showed a very similar shape of the size distribution curves consistently with what was expected from qualitative observations made by electron microscopy. Each method was based on a different principle and provided with slightly different ways to express the nanoparticle diameters. Despite these differences, several populations of size were detected by at least 3 methods of analysis giving a mean diameter within a $\pm 5\%$ range as underlined by the grey backgrounds shown on the graph at certain size positions (Fig. 7). It is noteworthy that results provided by these methods were rather consistent between them and with those of the AFM that is considered as a direct method of the evaluation of the size characteristics of dispersion. The consistency obtained between these methods supports that the deposit of the nanoparticle sample on the mica plate holder was random and representative to the size distribution characteristics of the dispersion. Despite slightly different results, all these methods have revealed that the dispersion was composed of several populations of nanoparticles that differed from their size making the PSD quite complex. Their consistency with AFM that is a direct method suggested

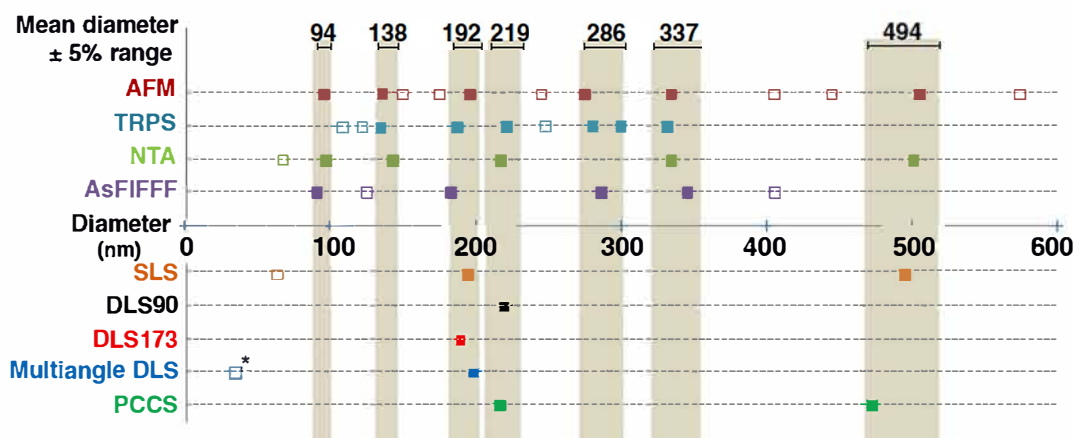


Fig. 7 Summary of the obtained PSD from the dispersion of PIBCA nanoparticles analyzed by different size measurement methods. Results from measurements performed with single size measurement methods (AFM, TRPS, NTA) and methods based on a separative process (AsFIFFF) are shown on the upper part of the graph. Results from measurements performed with batch size measurement methods comprising different light scattering methods (SLS, DLS, PCCS) are summarized on the lower part of the graph. * indicates a significant variation of the size position of the corresponding detected population. Mean diameters were calculated from the diameters of the population of nanoparticles with the corresponding size that was detected in a $\pm 5\%$ range (shown by the grey background) by at least 3 different methods.

that they provided with a quite representative view of the PSD of the analyzed dispersion. This is interesting because AFM is a very demanding method while the time required to perform one analysis by the other methods is much shorter. Indeed, times required to perform an analysis by NTA, TRPS and AsFIFFF are 18, 2 and 60 min respectively.

Methods of analysis used to provide with the results summarized on the lower part of the graph suggested that the PSD of the nanoparticle dispersion was composed of 1, 2 or a maximum of 3 populations of nanoparticles. Light scattering methods that formed this group of methods were all able to detect one population of nanoparticles with a diameter around 200 nm. It seemed that this population of particle was also detected by the other methods with peak positions slightly below, 192 nm, or slightly above, 219 nm. The three DLS methods only revealed this population of nanoparticles. In contrast, the SLS and the PCCS methods have detected a second population of nanoparticles with a diameter around 494 nm that corresponded to a population also detected by 2 methods of the other group, AFM and NTA. Nevertheless, PSD depicted by the five light scattering methods used to characterize the dispersion of PIBCA nanoparticles was rather simple compared with PSD revealed by methods based on single particle size measurement methods, i.e. AFM, NTA, TRPS, and a separation process, i.e. AsFIFFF. Thus, the different light scattering methods tested in this work have failed to provide with a representative view of the complex PSD revealed by the analysis performed with methods based on totally different principles and that was expected from the qualitative observations of the electron micrographs. It is noteworthy that the change in modalities to carry on light scattering measurements by using SLS and PCCS improved the sensibility of the method as witnesses by the detection of 2 or 3 populations. However, it remained insufficient compared

with methods based on single particle size measurement and methods including a separation by size prior to particle size measurement, i.e. AsFIFFF that have revealed a much complex PSD for the nanoparticle dispersion considered in this work in a consistent way. This confirms once more that light scattering methods should be applied carefully while characterizing unknown dispersions. Although they are well implemented in laboratories and achieved measurements in few minutes, they are unsuitable to depict size characteristics of dispersions having complex PSD (33,37–39). This is why several reports of the literature (19,33) recommend the use of at least two methods to characterize unknown dispersions or hyphenated separative particle size measurement methods with other particle size measurement methods (32,39). From the present work that have investigated size characteristics of a complex dispersion of polymer nanoparticles with 9 methods chosen among 3 classes of methods, it can be suggested that at least one of the two particle size measurement methods should be chosen among methods based on single particle size measurement or should include a method of separation by size. Applications of light scattering methods that are parts of the batch methods must only be applied to the characterization of dispersions with simple PSD once this has been established.

CONCLUSION

Taking account the inherent limitations of particle size measurement methods, the evaluation of PSD of complex dispersions of nanoparticles with multimodal distribution is a difficult task to provide an accurate and representative PSD. PSD of unknown dispersions of nanoparticles should be measured with two methods based on different physical principles including at least one method based on single particle size

measurement or on a prior separative step of nanoparticles according to their size before size measurement.

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REFERENCES

1. Jung H, Kittelson DB, Zachariah MR. The influence of a cerium additive on ultrafine diesel particle emissions and kinetics of oxidation. *Combust Flame*. 2005;142:276–88.
2. Jørgensen B, Kristensen SB, Kunov-Kruse AJ, Fehrmann R, Christensen CH, Rissager A. Gas-phase oxidation of aqueous ethanol by nanoparticle vanadia/anatase catalysts. *Top Catal*. 2009;52:253–7.
3. Wissing SA, Müller RH. Cosmetic applications for solid lipid nanoparticles (SLN). *Int J Pharm*. 2003;254:65–8.
4. Olivier J. Drug transport to brain with targeted nanoparticles. *NeuroRx*. 2005;2:118–9.
5. Cormode DP, Naha PC, Fayad ZA. Nanoparticle contrast agents for computed tomography: a focus on micelles. *Contrast Media Mol Imaging*. 2014;9(1):37–52.
6. Neuwelt EA, Varallyay P, Bago AG, Muldoon LL, Nesbit G, Nixon R. Imaging of iron oxide nanoparticles by MR and light microscopy in patients with malignant brain tumours. *Neuropathol Appl Neurobiol*. 2004;30:456–71.
7. Perlman O, Weitz IS, Azhari H. Copper oxide nanoparticles as contrast agents for MRI and ultrasound dual-modality imaging. *Phys Med Biol*. 2015;60(15):5767–83.
8. Galper MW, Saung MT, Fuster V, Roessl E, Thran A, Proksa R, *et al*. Effect of computed tomography scanning parameters on gold nanoparticle and iodine contrast. *Invest Radiol*. 2012;47(8):475–81.
9. Liu CJ, Wang CH, Chen ST, Chen HH, Leng WH, Chien CC, *et al*. Enhancement of cell radiation sensitivity by pegylated gold nanoparticles. *Phys Med Biol*. 2010;55(4):931–45.
10. Seaton A, Tran L, Aitken R, Donaldson K. Nanoparticles, human health hazard and regulation. *J R Soc Interface*. 2009;7:119–29.
11. Li C. Structure controlling and process scale-up in the fabrication of nanomaterials. *Front Chem Eng China*. 2010;4:18–25.
12. Organisation for Economic Co-operation and Development (OCDE), Regulatory frameworks for nanotechnology in foods and medical products: summary results of a survey activity, DSTI/STP/NANO(2012)22/FINAL, 21 March 2013. Available from: <http://search.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=DSTI/STP/NANO%282012%2922/FINAL&docLanguage=En> (consulted on November 2015). Available from.
13. Draft guidance from FDA, Considering Whether an FDA-Regulated Product Involves the Application of Nanotechnology, 14 June 2011. Available from: <http://www.fda.gov/RegulatoryInformation/Guidances/ucm257698.htm> (consulted on November 2015).
14. Reflection paper on the data requirements for intravenous liposomal products developed with reference to an innovator liposomal product, EMA/CHMP/806058/2009/Rev 02, 21 February 2013. Available from: http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2013/03/WC500140351.pdf (consulted on November 2015).
15. Joint MHLW/EMA reflection paper on the development of block copolymer micelle medicinal products, EMA/CHMP/13099/2013, 17 January 2013. Available from: http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2013/02/WC500138390.pdf (consulted on November 2015).
16. Report of the Joint Regulator -Industry Ad Hoc Working Group: Currently Available Methods for Characterization of Nanomaterials, 17 June 2011. Available from: http://ec.europa.eu/consumers/sectors/cosmetics/files/pdf/iccr5_char_nano_en.pdf (consulted on November 2015).
17. Organization for Economic Co-operation and Development (OCDE), Guidance manual for the testing of manufactured nanomaterials: OECD’s sponsorship programme; First revision ENV/JM/MONO(2009)20/REV, 2 June 2010. Available from: <http://search.oecd.org/officialdocuments/displaydocumentpdf/?cote=env/jm/mono%282009%2920/rev&doclanguage=en> (consulted on November 2015).
18. FDA advisory committee for pharmaceutical science and clinical pharmacology meeting Topic 2 Nanotechnology - Update on FDA Activities, 9 August 2012. Available from: <http://www.fda.gov/downloads/AdvisoryCommittees/CommitteesMeetingMaterials/Drugs/AdvisoryCommitteeForPharmaceuticalScienceandClinicalPharmacology/UCM314585.pdf> (consulted on November 2015).
19. Gaumet M, Vargas A, Gurny R, Delie F. Nanoparticles for drug delivery: the need for precision in reporting particle size parameters. *Eur J Pharm Biopharm*. 2008;69:1–9.
20. Shekunov BY, Chattopadhyay P, Tong HHY, Chow AHL. Particle size analysis in pharmaceuticals: principles, methods and applications. *Pharm Res*. 2006;24(2):203–27.
21. Varenne F, Botton J, Merlet C, Beck-Broichsitter M, Legrand F-X, Vauthier C. Standardization and validation of a protocol of size measurements by dynamic light scattering for monodispersed stable nanomaterial characterization. *Colloid Surf A*. 2015;486:124–38.
22. Braun A, Couteau O, Franks K, Kestens V, Roebben G, Lamberty A, *et al*. Validation of dynamic light scattering and centrifugal liquid sedimentation methods for nanoparticle characterisation. *Adv Powder Technol*. 2011;22:766–70.
23. Woodward RC, Heeris J, St Pierre TG, Saunders M, Gilbert EP, Rutnakornpituk M, *et al*. A comparison of methods for the measurement of the particle-size distribution of magnetic nanoparticles. *J Appl Crystallogr*. 2007;40:495–500.
24. Elizalde O, Leal GP, Leiza JR. Particle size distribution measurements of polymeric dispersions: a comparative study. *Part Part Syst Charact*. 2000;17:236–43.

25. Fielding LA, Mykhaylyk OO, Armes SP, Fowler PW, Mittal V, Fitzpatrick S. Correcting for a density distribution: particle size analysis of core-shell nanocomposite particles using disk centrifuge photosedimentometry. *Langmuir*. 2012;28:2536-44.
26. Bell NC, Minelli C, Tompkins J, Stevens MM, Shard AG. Emerging techniques for submicrometer particle sizing applied to Stoeber silica. *Langmuir*. 2012;28:10860-72.
27. Linsinger T, Roebben G, Gilliland D, Calzolari L, Rossi F, Gibson N, et al. Requirements on measurements for the implementation of the European Commission definition of the term "nanomaterial". *JRC Reference Reports*. 2012.
28. Powers KW, Brown SC, Krishna VB, Wasdo SC, Moudgil BM, Roberts SM. Research strategies for safety evaluation of nanomaterials. Part VI. Characterization of nanoscale particles for toxicological evaluation. *Toxicol Sci*. 2006;90(2):296-303.
29. Sowerby SJ, Broom MF, Petersen GB. Dynamically resizable nanometre-scale apertures for molecular sensing. *Sensors Actuators B*. 2007;123:325-30.
30. Willmott GR, Vogel R, Yu SSC, Groenewegen LG, Roberts GS, Kozak D, et al. Use of tunable nanopore blockade rates to investigate colloidal dispersions. *J Phys-Condens Mat*. 2010;22(45):1-11.
31. Vogel R, Willmott G, Kozak D, Roberts GS, Anderson W, Groenewegen L, et al. Quantitative sizing of nano/microparticles with a tunable elastomeric pore sensor. *Anal Chem*. 2011;83:3499-506.
32. Lespes G, Gigault J. Hyphenated analytical techniques for multidimensional characterisation of submicron particles: a review. *Anal Chim Acta*. 2011;692:26-41.
33. Anderson W, Kozak D, Coleman VA, Jämting ÅK, Trau M. A comparative study of submicron particle sizing platforms: accuracy, precision and resolution analysis of polydisperse particle size distributions. *J Colloid Interface Sci*. 2013;405:322-30.
34. Sokolova V, Ludwig A-K, Hornung S, Rotan O, Horn PA, Epple M, et al. Characterisation of exosomes derived from human cells by nanoparticle tracking analysis and scanning electron microscopy. *Colloid Surf B*. 2011;87:146-50.
35. Pace HE, Rogers NJ, Jarolimek C, Coleman VA, Gray EP, Higgins CP, et al. Single particle inductively coupled plasma-mass spectrometry: a performance evaluation and method comparison in the determination of nanoparticle size. *Environ Sci Technol*. 2012;46:12272-80.
36. van der Pol E, Coumans FAW, Grootemaat AE, Gardiner C, Sargent IL, Harrison P, et al. Particle size distribution of exosomes and microvesicles determined by transmission electron microscopy, flow cytometry, nanoparticle tracking analysis, and resistive pulse sensing. *J Thromb Haemost*. 2014;12:1182-92.
37. Cascio C, Gilliland D, Rossi F, Calzolari L, Contado C. Critical experimental evaluation of Key methods to detect, size and quantify nanoparticulate silver. *Anal Chem*. 2014;86:12143-51.
38. Calzolari L, Gilliland D, Garcia CP, Rossi F. Separation and characterization of gold nanoparticle mixtures by flow-field-flow fractionation. *J Chromatogr A*. 2011;1218:4234-9.
39. Ingebrigtsen L, Brandl M. Determination of the size distribution of liposomes by SEC fractionation, and PCS analysis and enzymatic assay of lipid content. *AAPS Pharm Sci Tech*. 2002;3(2):9-15.
40. Sitar S, Kejžar A, Pahovnik D, Kogej K, Tušek-Žnidarič M, Lenassi M, et al. Size characterization and quantification of exosomes by asymmetrical-flow field-flow fractionation. *Anal Chem*. 2015;87:9225-33.
41. Gun'ko VM, Klyueva AV, Levchuk YN, Leboda R. Photon correlation spectroscopy investigations of proteins. *Adv Colloid Interface*. 2003;105:201-328.
42. ISO/TS 10797:2012: Nanotechnologies - Characterization of single-wall carbon nanotubes using transmission electron microscopy.
43. ISO 13322-1:2004 Particle size analysis - Image analysis methods - Part 1: Static image analysis, methods.
44. Vauthier C, Persson B, Lindner P, Cabane B. Protein adsorption and complement activation for di-block copolymer nanoparticles. *Biomaterials*. 2011;32:1646-56.
45. Rasband W. ImageJ (Computer Program), National Institute of Health, 2013.
46. Cybernetics M. Image-Pro Plus (Computer Program), Roper Industries, 2013.
47. Binnig G, Quate CF, Gerber C. Atomic force microscope. *Phys Rev Lett*. 1986;56:930-3.
48. Meyer G, Amer NM. Novel optical approach to atomic force microscopy. *Appl Phys Lett*. 1988;53(12):1045-7.
49. Couteau O, Roebben G. Measurement of the size of spherical nanoparticles by means of atomic force microscopy. *Meas Sci Technol*. 2001;22(6):65101-8.
50. Filipe V, Hawe A, Jiskoot W. Critical evaluation of Nanoparticle Tracking Analysis (NTA) by NanoSight for the measurement of nanoparticles and protein aggregates. *Pharm Res*. 2010;27:796-810.
51. ISO 22412:2008(E): Particle size analysis - dynamic light scattering (DLS).
52. Cho TJ, Hackley VA. Fractionation and characterization of gold nanoparticles in aqueous solution: asymmetric-flow field flow fractionation with MALS, DLS, and UV vis detection. *Anal Bioanal Chem*. 2010;398:2003-18.
53. Rbii K, Violleau F, Guedj S, Surel O. Analysis of aged gelatin by AFIFFF-MALS: Identification of high molar mass components and their influence on solubility. *Food Hydrocoll*. 2009;23:1024-30.