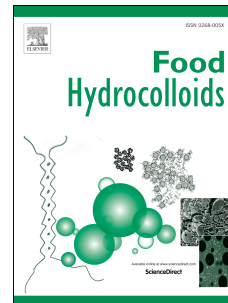


# Accepted Manuscript

Multi-scale structure, pasting and digestibility of adlay (*Coixlachryma-jobi L.*) seed starch

Jicheng Chen, Yazhen Chen, Huifang Ge, Chunhua Wu, Jie Pang, Song Miao



PII: S0268-005X(18)31522-4

DOI: <https://doi.org/10.1016/j.foodhyd.2018.11.050>

Reference: FOOHYD 4791

To appear in: *Food Hydrocolloids*

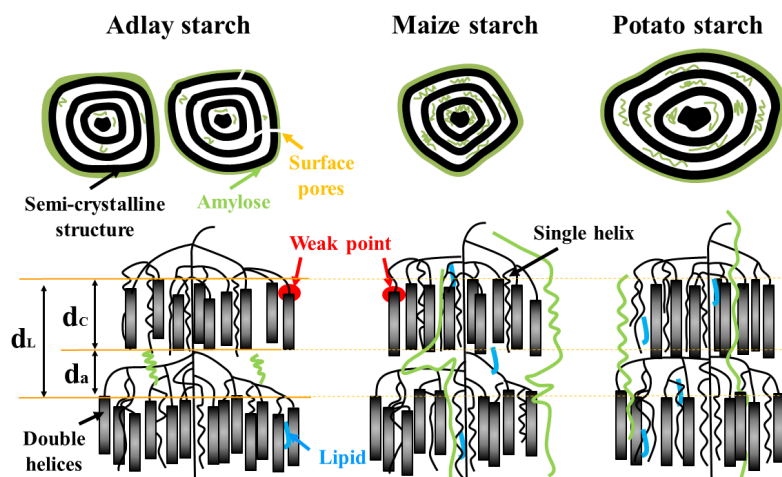
Received Date: 13 August 2018

Revised Date: 27 November 2018

Accepted Date: 27 November 2018

Please cite this article as: Chen, J., Chen, Y., Ge, H., Wu, C., Pang, J., Miao, S., Multi-scale structure, pasting and digestibility of adlay (*Coixlachryma-jobi L.*) seed starch, *Food Hydrocolloids* (2018), doi: <https://doi.org/10.1016/j.foodhyd.2018.11.050>.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.





16 **Abstract:** The hierarchical structure, pasting and digestibility of adlay seed starch  
17 (ASS) were investigated compared with maize starch (MS) and potato starch (PS). ASS  
18 exhibited round or polyglonal morphology with apparent pores/channels on the surface.  
19 It had a lower amylose content, a looser and more heterogeneous C-type crystalline  
20 structure, a higher crystallinity, and a thinner crystalline lamellae. Accordingly, ASS  
21 showed a higher slowly digestible starch content combined with less resistant starch  
22 fractions, and a decreased pasting temperature, a weakened tendency to retrogradation  
23 and an increased pasting stability compared with those of MS and PS. The ASS  
24 structure-functionality relationship indicated that the amylose content, double helical  
25 orders, crystalline lamellar structure, and surface pinholes should be responsible for  
26 ASS specific functionalities including pasting behaviors and *in vitro* digestibility.  
27 ASS showed potential applications in health-promoting foods which required low  
28 rearrangement during storage and sustainable energy-providing starch fractions.

29 **Keywords:** Adlay seed starch; multi-scales structures; pasting properties;  
30 digestibility

31

## 32 1. Introduction

33 Adlay (*Coixlachryma-jobi* L. var. *ma-yuen* Stapf), commonly known as adlay or  
34 Job's tears, is an annual crop widely cultivated in East and South-East Asia  
35 (Chaisiricharoenkul, Tongta, & Intarapichet, 2011; Chang, Huang, & Hung, 2003).  
36 Adlay seeds contain a great number of health-beneficial bioactive components (e.g.,  
37 protein, polysaccharide, polyphenols, coixenolide, coixol, oil, etc.) and have been  
38 considered as a traditional oriental medicine for centuries for the treatment of edema,  
39 rheumatism, and neuralgia (Liu et al., 2017; Tseng, Yang, Chang, Lee, & Mau, 2006;  
40 Zhu, 2017). Besides, numerous studies have been reported that adlay seeds have the  
41 ability to prevent the formation of tumors, reduce inflammation, ameliorate metabolic  
42 syndrome, and aid in gastrointestinal tract regulation (Chen, Lo, & Chiang, 2012; Tsai,  
43 Yang, & Hsu, 1999; Wenchang, Cheng, Mengtsan, & Kingthom, 2000). Owing to its  
44 perceived nutritional and health benefits, adlay seeds are increasingly utilized in the  
45 food industry.

46 The major component of adlay seeds is starch, accounting for approximately  
47 54.26%-58.15% of its dry mater (Chaisiricharoenkul et al., 2011; Liu, Han, & Sun,  
48 2012). Over the last decade, adlay seed starch (ASS) has been served as a food  
49 ingredient through several food products such as baked products, soups, broths,  
50 distilled liquor, etc. (Yang, Peng, Lui, & Lin, 2008; Zhu, 2017) . Besides, the  
51 relationships between supramolecular structures and pasting features of adlay seed  
52 starches have been revealed (Miao et al., 2018; Xu et al., 2017). However, studies  
53 focused on ASS physicochemical properties and functionalities such as the content of  
54 rapidly digestible starch (RDS), slowly digestible starch (SDS), and resistant starch  
55 (RS) are scarce (Chaisiricharoenkul et al., 2011; Li & Corke, 1999). Importantly,  
56 long-term consumption of starchy foods enriched with RDS was regarded as a

57 fundamental cause to a wide variety of metabolic complications such as obesity and  
58 type II diabetes (Brandmiller, Dickinson, Barclay, Celermajer, 2007; Brandmiller,  
59 2007).

60 The SDS fractions which provide a slow and prolonged release of glucose, and  
61 the RS that cannot be digested in human upper gastrointestinal tract but is used by  
62 microflora in the colon are more than encouraged for a health-promoting diet  
63 (Lehmann & Robin, 2007). With a never growing population and human health  
64 interests, there could be a shortage of common health-promoting starches for industrial  
65 food applications in the future. Therefore, it is essential to identify, characterize and use  
66 non-conventional starches for industrial applications. Based on the excellent  
67 functionalities of adlay, further insights into the ASS multi-scale structure and  
68 functionalities such as pasting properties and digestibility are inevitable and  
69 necessary.

70 Although most native starches are semi-crystalline (containing crystalline and  
71 amorphous lamellae) in nature, their granule size, shape and microstructures are  
72 diverse, depending on their botanical source, growing and harvesting conditions (Liu  
73 et al., 2017). The granule size of starches ranged from 1.5  $\mu\text{m}$  to 100  $\mu\text{m}$ , while the  
74 shape varied from irregular to elliptical, tetrahedral, polygonal and spherical forms.  
75 And the hierarchical structures of starch granules have been confirmed to be the  
76 critical determinant of starch functionalities for food processing and human nutrition  
77 (Chi et al., 2017). Thus, understanding the relationships between hierarchical  
78 structures and functional properties (e.g. digestibility and pasting properties) of starch  
79 is very important for optimizing food and industrial applications (Syahariza, Sar,  
80 Hasjim, Tizzotti, & Gilbert, 2013) .

81 The aim of this study was to investigate the physicochemical, micro-structural,  
82 thermal, pasting and digestibility properties of ASS. The relationships of structures  
83 and functionalities of ASS were also discussed to state whether the ASS is suitable for  
84 the health-promoting foods or other specific industrial applications. The results were  
85 compared with commercial maize starch (MS) and potato starch (PS). This study will  
86 provide a scientific basis to extend the commercial applications of ASS.

## 87 **2. Materials and Methods**

### 88 **2.1. Materials**

89 The adlay was planted in May and harvested in December in Pucheng, the county  
90 of Fujian in China. The county showed an average temperature of 17.4 °C, annual  
91 rainfall of 1780 mm, and annual sunshine time of 1893 h.

92 Commercial potato and maize starch were purchased from Kang yuan Co., Ltd,  
93 (Henan, China) and used for comparison with ASS. Pancreatin from porcine pancreas  
94 and amyloglucosidase were purchased from Sigma-Aldrich Co., Ltd. The D-glucose  
95 assay kit (GOPOD, K-GLUC) was purchased from Megazyme International Ireland  
96 Co., Ltd. (Wicklow, Ireland). All chemical reagents were of analytical reagent grade.  
97 Commercial starches and chemicals were used directly without further purification.

### 98 **2.2. Isolation of ASS**

99 Starch was isolated from adlay seeds following the previously published methods  
100 with some modifications (Kim et al., 2008). The seeds were steeped in excess water for  
101 4 h at 25 °C, grounded in an organization broking machine (JJ-2B, Four Red  
102 Instrument, Co., Ltd., Shanghai, China) at full speed for 1 min and filtered through  
103 200-mesh sieves. The NaOH (0.05g/L) was added into the filtrate, with stirring for 10  
104 min and then after stewing for 3 h at 25 °C, the supernatant and the top, yellowish layer

105 of protein was removed. The sediment was washed several times with deionized water.  
106 The ethanol and diethyl ether were added to the starch suspension to eliminate  
107 non-starch polysaccharide and lipid. Then the starch suspension was centrifuged at  
108 5000×g for 10 min. After centrifugation, the supernatant and dark tailing layer were  
109 discarded and the residue was washed several times with deionized water until the  
110 supernatant was clear. Then the residue was dried at 40 °C and milled below 50 °C to  
111 yield the starch and stored in a sealed plastic bag.

### 112 **2.3. Chemical Composition Analysis of starches**

113 The starch content, moisture, protein, lipid and ash of starch were determined by  
114 the standard methods of AOAC (Scott & Helrich, 1990) procedures. The amylose  
115 contents were determined by the method of iodine colorimeter at 620 nm using a potato  
116 starch standard mixture (Yu, Ma, Menager, & Sun, 2012) . The results were reported on  
117 a dry weight basis. All the experiments were performed at least in triplicate and results  
118 were presented as the mean value.

### 119 **2.4. Crystal structure analysis**

120 The crystal structure of starches was determined with an Xpert PRO diffractometer  
121 (Rigaku, Corp., Tokyo, Japan), operated at 40 mA and 40 kV with an X-ray source of  
122 Cu K $\alpha$  radiation ( $\lambda= 0.1542$  nm). The range of the diffraction angle ( $2\theta$ ) was from 5° to  
123 60° with a scanning speed of 10°/min and scanning step of 0.033°. The moisture  
124 content of each sample was equilibrated at 40 °C and all were approximately 10%. The  
125 crystallinity of starch was calculated by following equation:

$$126 \text{Crystallinity}(\%) = \frac{A_c}{A_c + A_a} \times 100$$

127 where  $A_c$  is the crystalline area and  $A_a$  is amorphous on the X-ray diffractogram.



## 128 2.5. Lamellar structure

129 Lamellar structure of starches was detected by a synchrotron small angle X-ray  
130 scattering (SAXS) system at Shanghai Synchrotron Radiation Facility (SSRF, China).  
131 A monochromatic beam of 0.124 nm was used and the sample-to-detector distance  
132 was 1860 mm, which provided a  $q$ -range from 0.10 to 1.5 nm<sup>-1</sup>. Samples were  
133 presented in 2 mm sealed quartz capillaries as suspensions containing excess water  
134 and scattering was measured for 60 s. A sealed 2 mm quartz capillary filled with  
135 water was used as a background. SAXS curves were normalized to sample  
136 transmission and background-subtracted using fit 2D software. The Bragg spacing  $d$ ,  
137 i.e. the thickness of starch lamellar structure, was calculated from the position of the  
138 peak ( $q$ ) according to  $d=2\pi/q$ .

139 In order to further clarify the structural parameters of semi-crystalline lamellae,  
140 one-dimensional correlation function profiles were calculated according to following  
141 equation (Chi et al., 2017; Kuang et al., 2017):

$$142 \quad f(r) = \frac{\int_0^{\infty} I(q)q^2 \cos(qr) dq}{\int_0^{\infty} I(q)q^2 dq}$$

143 where  $r$  represents the distance in real space.

## 144 2.6. Morphology observation and particle size analysis

145 Granule micrographs were observed at 3000 × magnification under a scanning  
146 electron microscope (XL30, Philips, Holand), and the light property of granule were  
147 observed viewed under the Olympus BX53 polarized light microscope according to the  
148 method of Man et al. (2012). The granule size analysis carried out with JEDA-801D  
149 particle size analyzer (Jiangsu JEDA Science-Technology Development Co., Ltd.,  
150 Nanjing, China).

## 151 **2.7. Gelatinization properties**

152 The gelatinization properties of the samples were studied by using a differential  
153 scanning calorimeter (DSC-200F3 NETZSCH-Gerätebau GmbH, Germany). Indium  
154 was used as the calibration standard. Starch slurries were prepared at 1:3 dry  
155 starch/ratios and sealed, reweighed. Samples then were allowed to heat from 10 °C to  
156 110 °C at a heating rate of 10 °C per minute. The onset temperature ( $T_o$ ), peak  
157 temperature ( $T_p$ ), conclusion temperature ( $T_c$ ), gelatinization temperature range ( $T_c-T_o$ )  
158 temperatures, as well as enthalpy ( $\Delta H_{gel}$ ), were calculated. All thermal analyses were  
159 conducted in triplicate for each starch.

## 160 **2.8. Pasting properties**

161 The pasting properties of starches were analyzed by using Brabender  
162 Visco-Analyser (Brabendviscograph-E, Brabender GmbH & Co. KG, Germany).  
163 Briefly, the starch sample (8% w/w, d.b.) were subjected to the following heating and  
164 cooling program: equilibrated at 35 °C for 5 min, heated to 95 °C in 40 min, held at  
165 95 °C for 30 min, cooled to 50 °C in 40 min, and held at 50 °C for 30 min. All  
166 measurements were performed in triplicate.

## 167 **2.9. *In vitro* starch digestibility**

168 *In vitro* starch digestibility was analyzed according to the Englyst method  
169 (Englyst, Kingman, & Cummings, 1992) with slight modifications. Enzyme working  
170 solution containing 780 USP porcine pancreatin and 3 units amyloglucosidase was  
171 freshly prepared before use. Starches (1.0 g, dsb) were dispersed in 20.0 mL acetate  
172 buffer solution (0.1 M, pH 5.2) with 4 mM  $CaCl_2$ , then six glass balls were added to  
173 the starch suspension and incubated with 5mL enzyme solution under continuous  
174 shaking (190 rpm) at 37 °C. An aliquot (0.5 mL) of the hydrolysate was removed at

175 time intervals of 20 min and 120 min, and then mixed with 20 mL 70% ethanol to  
176 denature the enzymes. The samples were centrifuged at  $5000\times g$  for 5 min and the  
177 glucose content in the supernatant was measured with the Megazyme glucose assay  
178 kit (GOPOD method). The glucose content at intervals of 20 and 120 min was labeled  
179 as G20 and G120, and the contents of rapidly digestible starch (RDS), slowly  
180 digestible starch (SDS) and resistant starch (RS) content were calculated by the  
181 following equations:

$$182 \quad RDS = G20 \times 0.9 / TS \times 100\%$$

$$183 \quad SDS = (G120 - G20) \times 0.9 / TS \times 100\%$$

$$184 \quad RS = [TS - RDS - SDS] / TS \times 100\%$$

185 where the TS means the total starch (TS) content of the complexes used for digestibility  
186 measurement. Herein, the TS equals to 1 g.

## 187 **2.10. Statistical analysis**

188 One-way analysis of variance (ANOVA) was performed with Tukey's HSD test  
189 ( $*p < 0.05$ ) using SPSS (20.0 version, IBM). The significance level was set as  $*p < 0.05$ .

## 190 **3. Results and Discussion**

### 191 **3.1. Proximate composition analysis of ASS**

192 The chemical components of starches are presented in Table S1. The starch  
193 content of adlay seed (yield of  $(43.2 \pm 0.13)\%$ , d.b.s) is lower than that extracted from  
194 adlay seed planted in Japan, Burma and Thailand (Wu, Charles, & Huang, 2007),  
195 which may be attributed to the variation in different cultivation climates and regions.  
196 Careful isolation and washing procedures resulted in clean ASS (the purity reached  
197  $97.54 \pm 0.61\%$ ). The range of moisture contents in these starches varied from 7.74% to

198 9.37%. ASS showed slightly higher moisture content ( $8.10\pm 0.17$ ) % than maize starch  
199 ( $7.74\pm 0.06$ %). The residual protein and lipid contents of ASS were ( $0.38 \pm 0.05$ ) % and  
200 ( $0.07 \pm 0.01$ ) %, respectively, which is lower than that of MS and PS, indicating protein  
201 and lipid are extracted extensively from ASS. The ash contents of ASS, potato starch  
202 and maize starch are ( $0.14\pm 0.01$ ) %, ( $0.20\pm 0.04$ ) % and ( $0.19\pm 0.03$ ) %, respectively.

203 The amylose content of these starches ranged from 2.25 to 24.60%. It was  
204 observed that the amylose content ( $2.25\pm 0.76$ %) of ASS was much lower than that of  
205 potato starch ( $23.32\pm 0.42$ %) and maize starch ( $24.60\pm 0.25$ %), respectively. The  
206 amylose content of ASS is considerably different to published data in a previous  
207 literature (Li et al., 1999), i.e., the amylose content of normal ASS and waxy ASS is  
208 15.9%-25.8% and 0.7%-1.1% respectively. These results indicated that the  
209 physicochemical properties of ASS maybe significantly different to other cereal  
210 starches.

### 211 3.2. Crystal properties of starches

212 X-ray diffraction patterns of ASS, MS, and PS are shown in Fig.1. The MS and  
213 PS displayed typical A- and B-type patterns, respectively. It can be seen that ASS  
214 exhibited diffraction peaks at  $5.6^\circ$ ,  $15.2^\circ$ ,  $17.2^\circ$ ,  $18.4^\circ$  and  $23.6^\circ$  ( $2\theta$ ), suggesting that  
215 ASS showed a C-type (hybrid of A-type and B-type) X-ray pattern (Man et al., 2012).  
216 This observation was consistent with the result reported by a previous literature (Kim  
217 et al., 2008). The degree of crystallization of ASS was 35.79% (Table 1), which was  
218 higher than that of PS (29.83%) and MS (31.25%). ASS contained a vast number of  
219 branched short chains, which was more readily packed into double helices and  
220 arranged to form starch crystals. Therefore, ASS had the highest degree of  
221 crystallization which follows the orders of ASS > MS > PS. According to previous  
222 studies (Chi et al., 2017; Lopez-Rubio, Flanagan, Shrestha, Gidley, & Gilbert, 2008),

223 starch crystalline structures, especially the crystallinity, were significantly related to  
224 starch gelatinization properties, pasting behaviors and digestibility. Understanding  
225 crystalline structures would be of help to accelerate ASS-based food development and  
226 applications.

### 227 3.3. Lamellar structure of starches

228 As the alternating stack of amorphous and crystalline regions, semi-crystalline  
229 lamellae assembled with a repeat distance of 9-10 nm. A characteristic peak at  
230 approximately  $0.6 \text{ nm}^{-1}$  was observed for all starches from Fig.2, indicating the  
231 existence of starch semi-crystalline lamellae. To be more accurate, ASS had a peak at  
232  $0.6624 \text{ nm}^{-1}$  and PS, MS showed peaks at  $0.6503$  and  $0.6528 \text{ nm}^{-1}$ , respectively,  
233 corresponding to the Bragg distances of 9.48, 9.83 and 9.62 nm calculated from  
234 Woolf-Bragg's equation ( $d=2\pi/q$ ) (Table 1). Starch lamellar thickness was highly  
235 associated with the susceptibility of enzymes attack and hydrothermal treatment  
236 (Wang et al., 2018). To further understand the characteristics of starch lamellar  
237 structures such as the thickness of crystalline lamella ( $d_c$ ), amorphous lamella ( $d_a$ ) and  
238 long repeated distance ( $d_L=d_a+d_c$ ), the one-dimensional function profile was also used  
239 in this work (Fig. S1). Adopting the method, long repeated distance of semi-crystalline  
240 lamellae ( $d_L$ ) can be calculated as the value of  $r$  at the second maximum of  $f(r)$ ,  $d_a$  is  
241 representing the solution of linear regression in the auto correlation triangle at  $f(r) =$   
242 value of the flat minimum (Fig.S1). Hence, the average thickness of the crystalline  
243 lamellae  $d_c$ , equals to  $(d_L-d_a)$ . As seen from Table 1,  $d_L$  showed similar changes to  
244 that of  $d$  and always showed a smaller value when calculated from Woolf-Bragg's  
245 equation. It could have resulted from the different starch model hypothesis (Fan et al.,  
246 2014), i.e.,  $d$  value was obtained based on a paracrystalline model (the three-phase  
247 model which contained amorphous background, crystalline and amorphous phases)

248 and the  $d_L$  linear correlation function approach which only concerned the  
249 crystalline/amorphous phases. Notably, although ASS had smallest  $d$  or  $d_L$ , it  
250 possessed the largest  $d_a$  (2.99 nm), which was higher than that of PS (2.59 nm) and  
251 MS (2.84 nm). This observation could be attributed to the differences in the fine  
252 structure of amylose and amylopectin from different botanical origins.

### 253 3.4. Morphology and size distribution of starch granules

254 The scanning electron micrographs (SEM) and polarized light microscope (PLM)  
255 of MS, PS and ASS are presented in Fig.3. It can be seen that MS and ASS granules  
256 are round or polyglonal in shape with smooth surfaces (Fig.3a, e), while PS had a  
257 different morphology (oval or polyglonal) with larger granules (Fig.3b). According to  
258 the SEM photos, MS and PS did not showed any surface pinholes, while ASS  
259 apparently observed with channels or pinholes on granular surface. To our knowledge,  
260 channels always provide direct access of reagents to a loosely organized region at the  
261 hilum, which makes it possible to increase the accessibility of enzymes to starch  
262 (Bul on, Colonna, Planchot, & Ball, 1998). This observation indicated that the surface  
263 pinholes should be one of the critical factors determined the differential digestibility  
264 among MS, PS and ASS.

265 For starches with semi-crystalline granules, most of starch granules exhibited a  
266 Maltese cross under polarized light microscope (Sandhu, Singh, & Kaur, 2004) . The  
267 MS and ASS have typical Maltese cross in the central position (Fig.3a and c). However,  
268 the Maltese cross of PS is at one end of granule (Fig. 3b). It can also be seen that the  
269 size of PS is bigger than MS and ASS, which is in agreement with the scanning electron  
270 micrographs observed.

271 The size-distribution curves of starch granules are displayed in Fig.3g. It can be  
272 observed that ASS and MS showed similar uni-modal size distribution at 4-31  $\mu\text{m}$ , but

273 the PS exhibit a wider uni-modal size distribution at 17-104  $\mu\text{m}$ . The average particle  
274 sizes were 14.61  $\mu\text{m}$ , 14.15  $\mu\text{m}$  and 48.18  $\mu\text{m}$  for ASS, MS and PS, respectively. The  
275 variation of starch granules may be attributed to the biological origin, biochemistry of  
276 the amyloplast or chloroplast, as well as the physiology of the plant (Man et al., 2012;  
277 Singh, Singh, Kaur, Sodhi, & Gill, 2003). Starch granules with smaller particle size are  
278 less resistant to the permeation of water/digestive enzymes into the granules, which  
279 would critically influence the starch pasting properties or digestibility (Tan et al.,  
280 2015; Zhang & Hamaker, 2009).

### 281 3.5. Gelatinization properties of starches

282 The gelatinization properties of starches were investigated by DSC. The  
283 endothermic gelatinization properties are displayed in Table 2. The ASS, MS and PS  
284 illustrate endothermic peaks between 60 and 85  $^{\circ}\text{C}$ . The transition temperatures ( $T_o$ ,  $T_p$ ,  
285  $T_c$ ) and  $\Delta H_{\text{gel}}$  of starches from adlay seed, corn and potato starches were significantly  
286 different (Table 3). The  $T_o$  of ASS in water was slightly lower than that of MS and PS.  
287 However, the  $T_p$  and  $T_c$  of ASS in water were higher than that of MS and PS,  
288 respectively. Broader gelatinization temperature ranges were observed ( $T_c - T_o$ ) for ASS  
289 in comparison with that of MS or PS, which indicated the more heterogeneous  
290 crystalline structure of ASS. Generally, the orders of the starch double helices, size of  
291 crystallites and length of starch branch chain may contribute to these diverse views  
292 (Kim et al., 2008). Hence, ASS tends to have a looser double helices order, and  
293 thereby the  $\Delta H_{\text{gel}}$  of ASS was lower than that of MS and PS, respectively. These  
294 observations might be ascribed to the differences in the granular structure, amylose content,  
295 crystallinity and content or perfection of the double helices of starches (Singh et al.,  
296 2003; Xie, Liu, & Cui, 2006).

### 297 **3.6. Pasting properties of starches**

298 Starches from different plant sources exhibited their unique pasting behaviors,  
299 which were important for the evaluation and estimation of process design, unit  
300 operation and quality of the final starch products (Huang et al., 2014). The pasting  
301 profiles of three different starches were measured by Brabender viscometer and the  
302 parameters obtained from the pasting curve are listed in Table 3. Among the three  
303 starches, ASS had the lowest pasting temperature, which was in agreement with the  
304 DSC results. Moreover, ASS had the highest peak viscosities (1473.0 BU) and  
305 breakdown viscosities (1163.0 BU), but the lowest setback viscosities (395.0 BU), final  
306 viscosities (105.0 BU), and pasting temperatures (66.4 °C). The results indicated that  
307 the ASS is suitable for a long-time storage since its low retrogradation. Notably, the  
308 amylose content of ASS was negatively correlated to the peak and breakdown  
309 viscosities, but positively correlated to the setback viscosities, final viscosities, and  
310 pasting temperatures. These observations were well in accordance with the previous  
311 report that higher amylopectin contents resulted in a higher peak viscosity and lower  
312 setback viscosity (Ibanez et al., 2007). On the other hand, some researchers have  
313 reported that proteins and lipid influenced the pasting properties of rice starch and the  
314 forming of amylose-lipid complexes (Dautant, Simancas, Sandoval, & Muller, 2007;  
315 Marcoa & Rosell, 2008), which significantly affected the final viscosity, setback value,  
316 and pasting temperature of starch. In this work, lower lipid and protein content in ASS  
317 would contribute to the lower pasting temperature and higher breakdown viscosity.

### 318 **3.7. *In vitro* digestibility of starches**

319 Starch is the most important energy resource for humans, and the digestion  
320 behavior is critically related to human health. Depending on the rate and extent of



321 hydrolysis, starch fractions were classified as shown in Table 4. ASS had higher  
322 digestibility with less RS and higher RDS than those of MS and PS, indicating that  
323 ASS was likely to contribute to a higher glycemic response after ingestion. In addition,  
324 ASS showed higher SDS content than that of both MS and PS, suggesting ASS could  
325 provide more sustainable energy for the human body. Due to starch functionalities are  
326 highly correlated with its original multi-scale structures (Lopez-Rubio et al., 2008;  
327 Man et al., 2012; Wang et al., 2018), the differences in digestibility among ASS, MS  
328 and PS should be a result of the distinct hierarchical structures.

### 329 **3.8. Mechanism of starch structural properties and functionalities**

330 In starch granules, linear amylose and highly branched amylopectin are packed  
331 into the amorphous and crystalline starch regions with different length scales,  
332 including the molecular scale ( $\sim 0.1$  nm), lamellar structure (8-10 nm), growth rings  
333 ( $\sim 0.1$   $\mu\text{m}$ ), and whole granular morphology (1-100  $\mu\text{m}$ ) (Pikus, 2005; Zhang, Chen, Li,  
334 Li, & Zhang, 2015). These hierarchical structures always play key roles in starch  
335 functionalities, such as starch pasting properties and digestibility (Benmoussa,  
336 Moldenhauer, & Hamaker, 2007; Tan et al., 2015).

337 Schematic structural differences between ASS, MS and PS are illustrated in Fig.  
338 4. ASS had smaller granular size and less amylose content entangled on the granular  
339 surface compared with MS and PS. Besides, ASS showed a more heterogeneous  
340 crystalline structure with thicker amorphous lamellae, thinner crystalline lamellae and  
341 semi-crystalline lamellae. The less amylose content and heterogeneous crystalline  
342 features should be responsible for the weaker resistance to hydrothermal treatment  
343 (Table 3). The lipids content within starch granules always contributed to a higher  
344 pasting temperature/gelatinization temperature, since lipids would interact with  
345 amylose or long-branched chains of amylopectin to form starch-lipid inclusion

346 complexes which had a high thermostability. ASS had much lower lipid content  
347 (Table S1), which might significantly contribute to the lower pasting and  
348 gelatinization temperatures (Table 3 and Table 4). Although ASS showed higher  
349 crystallinity than those of MS and PS, a looser and more heterogeneous crystalline  
350 structure as well as thinner crystalline lamellae contributed to its lower  
351 thermostability. However, ASS showed lower setback viscosity than MS and PS,  
352 which ascribed to the higher amylopectin content and the difficulty of amylopectin  
353 rearrangement in a short timeframe. Actually, starch rearrangement and aging always  
354 decreased food quality which in turn leading to a deterioration of consumer  
355 acceptance. Therefore, ASS showed promising potential applications in the food  
356 industry such as instant soups, sauces and jelly, due to its low retrogradation during  
357 storage.

358 On the other hand, starch digestibility was critically related to human health and  
359 was influenced by structural features. Amylose entangled at the starch granular  
360 surface increased starch compactness, which could resist enzymes attack. In addition,  
361 surface pores/channels on the starch granules also increased enzyme accessibility to  
362 starches, since enzymes could enter starch interior through pores/channels. According  
363 to a previous study (Jane, Wong, & McPherson, 1997), starch with a B type  
364 crystalline structure had most of the branch points (i.e.,  $\alpha$ -1,6-glycosidic linkages)  
365 clustered in the amorphous region and making them less susceptible to the enzymatic  
366 hydrolysis, while A or C (A+B) type crystalline structure had 'weak points'(i.e.,  
367 susceptible to enzymatic hydrolysis) due to its branch points scattered in both  
368 amorphous and crystalline regions. Therefore, ASS, a typical C type starch, had a  
369 higher RDS content compared with PS. Generally, starch crystalline lamellae showed  
370 a more compact structure than that of amorphous lamellae, and in turn, making the

371 amorphous lamellae is easily digested when enzymes were treated. ASS had a thinner  
372 crystalline lamellae and thicker amorphous lamellae than those of MS and PS,  
373 indicating ASS could be digested by enzymes such as  $\alpha$ -amylase easily. Importantly,  
374 a looser and more heterogeneous crystalline structure of ASS also contributed to a  
375 higher susceptibility of starch to enzymes. Herein, ASS had a higher RDS content and  
376 a lower RS fraction compared with MS and PS. Nevertheless, ASS showed a higher  
377 crystallinity, which should be responsible for its higher SDS content than MS and PS..  
378 It could be concluded that structural features such as amylose content, surface pores,  
379 crystalline structure and lamellar structures contributed to ASS digestion behaviors.  
380 ASS, which had a higher SDS content, could be used for health-promoting foods  
381 which required a slow and prolonged release of glucose.

#### 382 **4. Conclusions**

383 In this work, ASS was extracted by alkaline steeping and its physicochemical and  
384 functionalities were investigated in comparison with commercial MS and PS. Although  
385 the ASS used in this study cannot represent the starches from coix in China, the  
386 structural features and functionalities relationships of ASS from the ASS major  
387 planting area were revealed. ASS had low amylose content and apparent surface  
388 pores/channels. It exhibited relatively high crystallinity, but a loose and heterogeneous  
389 C type crystalline structure. In addition, ASS had a thicker amorphous lamellae and  
390 thinner crystalline lamellae. All these structural features contributed to ASS lower  
391 pasting temperature, lower setback viscosity, higher RDS content and higher SDS  
392 fractions compared with those of MS and PS. It can be concluded that ASS can be  
393 used as an additive in health-promoting foods which required high stability during  
394 storage and are rich energy-providing starch fractions.

395 **Acknowledgments**

396       The authors would like to thank the support of Development Project of Marine  
397 High-Tech Industry in Fujian Province (Min Hai Yu Gao Xin No. [2004] 03) and  
398 Project of High-level University Construction of Fujian Agriculture and Forestry  
399 University (No. 612014042).

400

401 **References**

- 402 Benmoussa, M., Moldenhauer, K. A. K., & Hamaker, B. R. (2007). Rice Amylopectin  
403 Fine Structure Variability Affects Starch Digestion Properties. *Journal of*  
404 *Agricultural and Food Chemistry*, 55(4), 1475-1479.  
405 <http://dx.doi.org/10.1021/jf062349x>.
- 406 Buléon, A., Colonna, P., Planchot, V., & Ball, S. (1998). Starch granules: structure and  
407 biosynthesis. *International Journal of Biological Macromolecules*, 23(2),  
408 85-112. [http://dx.doi.org/10.1016/S0141-8130\(98\)00040-3](http://dx.doi.org/10.1016/S0141-8130(98)00040-3).
- 409 Brandmiller J., Dickinson S., Barclay A., Celermajor D (2007). The glycemic index  
410 and cardiovascular disease risk. *Current Atherosclerosis Reports*, 9(6),  
411 479-485. <http://dx.doi.org/10.1007/s11883-007-0064-x>.
- 412 Brandmiller J (2007). The glycemic index as a measure of health and nutritional  
413 quality: an Australian perspective. *Cereal Foods World*, 52(2), 41-44.  
414 <http://dx.doi.org/10.1007/s10900-012-9584-6>.
- 415 Chaisiricharoenkul, J., Tongta, S., & Intarapichet, K.-O. (2011). Structure and  
416 chemical and physicochemical properties of Job's tear (*Coix lacryma-jobi* L.)  
417 kernels and flours. *Suranaree J. Sci. Technol*, 18, 109-122.
- 418 Chang, H. C., Huang, Y. C., & Hung, W. C. (2003). Antiproliferative and  
419 chemopreventive effects of adlay seed on lung cancer in vitro and in vivo.  
420 *Journal of Agricultural and Food Chemistry*, 51(12), 3656-3660.  
421 <http://dx.doi.org/10.1021/jf021142a>.
- 422 Chen, H. J., Lo, Y. C., & Chiang, W. C. (2012). Inhibitory effects of adlay bran (*Coix*  
423 *lachryma-jobi* L. var. *ma-yuen* Stapf) on chemical mediator release and  
424 cytokine production in rat basophilic leukemia cells. *Journal of*  
425 *Ethnopharmacology*, 141(1), 119-127.  
426 <http://dx.doi.org/10.1016/j.jep.2012.02.009>.
- 427 Chi, C. D., Li, X. X., Zhang, Y. P., Chen, L., Li, L., & Wang, Z. J. (2017).  
428 Digestibility and supramolecular structural changes of maize starch by  
429 non-covalent interactions with gallic acid. *Food & Function*, 8(2), 720-730.  
430 <http://dx.doi.org/10.1039/c6fo01468b>.
- 431 Dautant, F. J., Simancas, K., Sandoval, A. J., & Muller, A. J. (2007). Effect of  
432 temperature, moisture and lipid content on the rheological properties of rice  
433 flour. *Journal of Food Engineering*, 78(4), 1159-1166.  
434 <http://dx.doi.org/10.1016/j.jfoodeng.2005.12.028>.
- 435 Englyst, H. N., Kingman, S., & Cummings, J. (1992). Classification and measurement  
436 of nutritionally important starch fractions. *European Journal of Clinical*  
437 *Nutrition*, 46, S33-50.
- 438 Fan, D., Wang, L., Chen, W., Ma, S., Ma, W., Liu, X., Zhao, J., & Zhang, H. (2014).  
439 Effect of microwave on lamellar parameters of rice starch through small-angle  
440 X-ray scattering. *Food Hydrocolloids*, 35, 620-626.  
441 <http://dx.doi.org/10.1016/j.foodhyd.2013.08.003>.

- 442 Huang, J. R., Chen, Z. H., Xu, Y. L., Li, H. L., Liu, S. X., Yang, D. Q., & Schols, H. A.  
443 (2014). Comparison of waxy and normal potato starch remaining granules  
444 after chemical surface gelatinization: Pasting behavior and surface  
445 morphology. *Carbohydrate Polymers*, *102*, 1001-1007.  
446 <http://dx.doi.org/10.1016/j.carbpol.2013.07.086>.
- 447 Ibanez, A. M., Wood, D. F., Yokoyama, W. H., Park, I. M., Tinoco, M. A., Hudson, C.  
448 A., McKenzie, K. S., & Shoemaker, C. F. (2007). Viscoelastic properties of  
449 waxy and nonwaxy rice flours, their fat and protein-free starch, and the  
450 microstructure of their cooked kernels. *Journal of Agricultural and Food*  
451 *Chemistry*, *55*(16), 6761-6771. <http://dx.doi.org/10.1021/jf070416x>.
- 452 Jane, J. L., Wong, K. S., & McPherson, A. E. (1997). Branch-structure difference in  
453 starches of A- and B-type x-ray patterns revealed by their Naegeli dextrans.  
454 *Carbohydrate Research*, *300*(3), 219-227.  
455 [http://dx.doi.org/10.1016/s0008-6215\(97\)00056-6](http://dx.doi.org/10.1016/s0008-6215(97)00056-6).
- 456 Kim, M. J., Choi, S. J., Shin, S. I., Sohn, M. R., Lee, C. J., Kim, Y., Il Cho, W., &  
457 Moon, T. W. (2008). Resistant glutarate starch from adlay: Preparation and  
458 properties. *Carbohydrate Polymers*, *74*(4), 787-796.  
459 <http://dx.doi.org/10.1016/j.carbpol.2008.04.043>.
- 460 Kuang, Q., Xu, J., Liang, Y., Xie, F., Tian, F., Zhou, S., & Liu, X. (2017). Lamellar  
461 structure change of waxy corn starch during gelatinization by time-resolved  
462 synchrotron SAXS. *Food Hydrocolloids*, *62*, 43-48.  
463 <http://dx.doi.org/10.1016/j.foodhyd.2016.07.024>.
- 464 Lehmann, U., & Robin, F. (2007). Slowly digestible starch – its structure and health  
465 implications: a review. *Trends in Food Science & Technology*, *18*(7), 346-355.  
466 <http://dx.doi.org/10.1016/j.tifs.2007.02.009>.
- 467 Li, J., & Corke, H. (1999). Physicochemical properties of normal and waxy Job's  
468 Tears (*Coix lachryma-jobi* L.) starch. *Cereal chemistry*, *76*(3), 413-416.
- 469 Liu, X., Zhang, B., Xu, J. H., Mao, D. Z., Yang, Y. J., & Wang, Z. W. (2017). Rapid  
470 determination of the crude starch content of Coix seed and comparing the  
471 pasting and textural properties of the starches. *Starch-Starke*, *69*(1-2).  
472 <http://dx.doi.org/10.1002/star.201600115>.
- 473 Lopez-Rubio, A., Flanagan, B. M., Shrestha, A. K., Gidley, M. J., & Gilbert, E. P.  
474 (2008). Molecular rearrangement of starch during in vitro digestion: Toward a  
475 better understanding of enzyme resistant starch formation in processed  
476 starches. *Biomacromolecules*, *9*(7), 1951-1958.  
477 <http://dx.doi.org/10.1021/bm800213h>.
- 478 Man, J. M., J. W.Cai, Cai, C. H., Xu, B., Huai, H. Y., & Wei, C. X. (2012).  
479 Comparison of physicochemical properties of starches from seed and rhizome  
480 of lotus. *Carbohydrate Polymers*, *88*(2), 676-683.  
481 <http://dx.doi.org/10.1016/j.carbpol.2012.01.016>.
- 482 Marcoa, C., & Rosell, C. M. (2008). Effect of different protein isolates and  
483 transglutaminase on rice flour properties. *Journal of Food Engineering*, *84*(1),

- 484 132-139. <http://dx.doi.org/10.1016/j.jfoodeng.2007.05.003>.
- 485 Miao, L., Zhao, S., Zhang, B., Tan, M., Niu, M., Jia, C., & Huang, Q. (2018).  
486 Understanding the supramolecular structures and pasting features of adlay  
487 seed starches. *Food Hydrocolloids*, 83, 411-418.  
488 <https://doi.org/10.1016/j.foodhyd.2018.05.034>
- 489 Pikus, S. (2005). Small-angle X-ray scattering (SAXS) studies of the structure of  
490 starch and starch products. *Fibres and Textiles in Eastern Europe*, 13(5),  
491 82-86.
- 492 Qingjie, X. L. H. Z. S. (2012). The Comparison of Job' s-Tears Powder and Job'  
493 s-Tears Starch on Their Physicochemical Properties and Digestibility [J].  
494 *Journal of the Chinese Cereals and Oils Association*, 7, 010.
- 495 Sandhu, K. S., Singh, N., & Kaur, M. (2004). Characteristics of the different corn  
496 types and their grain fractions: physicochemical, thermal, morphological, and  
497 rheological properties of starches. *Journal of Food Engineering*, 64(1),  
498 119-127. <http://dx.doi.org/10.1016/j.jfoodeng.2003.09.023>.
- 499 Scott, P., & Helrich, K. (1990). Official methods of analysis of the Association of  
500 Official Analytical Chemists. *Official methods of analysis of the Association of*  
501 *Official Analytical Chemists*.
- 502 Singh, N., Singh, J., Kaur, L., Sodhi, N. S., & Gill, B. S. (2003). Morphological,  
503 thermal and rheological properties of starches from different botanical sources.  
504 *Food Chemistry*, 81(2), 219-231.  
505 [http://dx.doi.org/10.1016/S0308-8146\(02\)00416-8](http://dx.doi.org/10.1016/S0308-8146(02)00416-8).
- 506 Syahariza, Z. A., Sar, S., Hasjim, J., Tizzotti, M. J., & Gilbert, R. G. (2013). The  
507 importance of amylose and amylopectin fine structures for starch digestibility  
508 in cooked rice grains. *Food Chemistry*, 136(2), 742-749.  
509 <http://dx.doi.org/10.1016/j.foodchem.2012.08.053>.
- 510 Tan, X., Zhang, B., Chen, L., Li, X., Li, L., & Xie, F. (2015). Effect of planetary  
511 ball-milling on multi-scale structures and pasting properties of waxy and  
512 high-amylose cornstarches. *Innovative Food Science & Emerging*  
513 *Technologies*, 30, 198-207. <http://dx.doi.org/10.1016/j.ifset.2015.03.013>.
- 514 Tsai, C., Yang, L., & Hsu, H. (1999). Ingestion of adlay may reduce liver fat  
515 accumulation in hamsters fed high fat diets. *Food Sci*, 26, 265-276.
- 516 Tseng, Y. H., Yang, J. H., Chang, H. L., Lee, Y. L., & Mau, J. L. (2006). Antioxidant  
517 properties of methanolic extracts from monascal adlay. *Food Chemistry*, 97(3),  
518 375-381. <http://dx.doi.org/10.1016/j.foodchem.2005.04.022>.
- 519 Wang, H., Liu, Y., Chen, L., Li, X., Wang, J., & Xie, F. (2018). Insights into the  
520 multi-scale structure and digestibility of heat-moisture treated rice starch.  
521 *Food chemistry*, 242, 323-329.  
522 <http://dx.doi.org/10.1016/j.foodchem.2017.09.014>.
- 523 Wenchang, C., Cheng, C. Y., Mengtsan, C., & Kingthom, C. (2000). Effects of  
524 dehulled adlay on the culture count of some microbiota and their metabolism  
525 in the gastrointestinal tract of rats. *Journal of Agricultural & Food Chemistry*,



- 526 48(3), 829-832. <http://dx.doi.org/10.1021/jf990473t>.
- 527 Wu, T. T., Charles, A. L., & Huang, T. C. (2007). Determination of the contents of the  
528 main biochemical compounds of Adlay (*Coxi lachrymal-jobi*). *Food*  
529 *Chemistry*, 104(4), 1509-1515.  
530 <http://dx.doi.org/10.1016/j.foodchem.2007.02.027>.
- 531 Xie, X., Liu, Q., & Cui, S. W. (2006). Studies on the granular structure of resistant  
532 starches (type 4) from normal, high amylose and waxy corn starch citrates.  
533 *Food Research International*, 39(3), 332-341.  
534 <http://dx.doi.org/10.1016/j.foodres.2005.08.004>.
- 535 Xu, L., Chen, L., Ali, B., Yang, N., Chen, Y., Wu, F., ... & Xu, X. (2017). Impact of  
536 germination on nutritional and physicochemical properties of adlay seed  
537 (*Coixlachryma-jobi* L.). *Food chemistry*, 229, 312-318.  
538 <https://doi.org/10.1016/j.foodchem.2017.02.096>
- 539 Yang, S. H., Peng, J., Lui, W. B., & Lin, J. (2008). Effect of adlay species and rice  
540 flour ratio on the physicochemical properties and texture characteristic of  
541 adlay-based extrudates. *Journal of Food Engineering*, 84(3), 489-494.  
542 <http://dx.doi.org/10.1016/j.jfoodeng.2007.06.010>.
- 543 Yu, S. F., Ma, Y., Menager, L., & Sun, D. W. (2012). Physicochemical Properties of  
544 Starch and Flour from Different Rice Cultivars. *Food and Bioprocess*  
545 *Technology*, 5(2), 626-637. <http://dx.doi.org/10.1007/s11947-010-0330-8>.
- 546 Zhang, B., Chen, L., Li, X., Li, L., & Zhang, H. (2015). Understanding the multi-scale  
547 structure and functional properties of starch modulated by glow-plasma: A  
548 structure-functionality relationship. *Food Hydrocolloids*, 50, 228-236.  
549 <http://dx.doi.org/10.1016/j.foodhyd.2015.05.002>.
- 550 Zhang, G., & Hamaker, B. R. (2009). Slowly digestible starch: concept, mechanism,  
551 and proposed extended glycemic index. *Critical Reviews in Food Science and*  
552 *Nutrition*, 49(10), 852-867. <http://dx.doi.org/10.1080/10408390903372466>.
- 553 Zhu, F. (2017). Coix: Chemical composition and health effects. *Trends in food science*  
554 *& technology*, 2017, 61: 160-175. <https://doi.org/10.1016/j.tifs.2016.12.003>
- 555



556 **Table 1.** Lamellar structural parameters and relative crystallinity of adlay seed  
 557 starch (AAS), normal maize starch (MS) and potato starch (PS) granules\*

Sample	$q(\text{nm}^{-1})$	$d$	$d_L(\text{nm})$	$d_a$	$d_c$	DC (%)
ASS	0.6624 <sup>a#</sup>	9.48 <sup>c</sup>	9.25 <sup>c</sup>	2.99 <sup>a</sup>	6.26 <sup>c</sup>	35.79 <sup>a</sup>
MS	0.6503 <sup>c</sup>	9.83 <sup>a</sup>	9.61 <sup>a</sup>	2.84 <sup>b</sup>	6.77 <sup>b</sup>	31.25 <sup>b</sup>
PS	0.6528 <sup>b</sup>	9.62 <sup>b</sup>	9.54 <sup>b</sup>	2.59 <sup>c</sup>	6.95 <sup>a</sup>	29.83 <sup>c</sup>

558 \* Parameters obtained by SAXS:  $q$ , peak position of semicrystalline lamellae;  $d$ , average thickness  
 559 of semicrystalline lamellae calculated by Woolf-Bragg's equation ( $d=2\pi/q$ ). Parameters calculated  
 560 from one-dimensional correlation function:  $d_L$ , long repeated distance of semicrystalline lamellae;  
 561  $d_a$ , average thickness of amorphous lamellae;  $d_c$ , average thickness of crystalline lamellae. DC,  
 562 degree of crystalline of starches obtained from XRD patterns.

563 # Values are means of three determinations ( $n = 3$ ). Values followed by the different letter within a  
 564 column differ significantly ( $*p < 0.05$ ).

565

**Table 2.** Gelatinization properties parameter for starches

Sample	$T_o$ (°C)	$T_p$ (°C)	$T_e$ (°C)	$T_e - T_o$ (°C)	$\Delta H_{gel}$ (J/g)
ASS	64.3±0.04c	71.2±0.32a	81.0±0.02a	16.7±0.23a	6.8±0.02c
MS	66.4±0.13a	70.9±0.01b	76.0±0.01b	9.6±0.12c	10.8±0.05b
PS	64.8±0.20b	69.3±0.09c	75.3± 0.12c	10.5±0.08b	12.7±0.02a

566  $T_o$ , on set temperature;  $T_p$ , peak temperature;  $T_e$ , end temperature;  $T_e - T_o$ , gelatinization range; values567 followed by the different letter within a column differ significantly ( $*p < 0.05$ ).

568

**Table 3.** Viscosity characteristics of starches

Sample	Pasting temperatures, °C	Peak viscosities, Bu	Final viscosities, Bu	Breakdown viscosities, Bu	Setback viscosities, Bu
ASS	66.4±0.09 <sup>b#</sup>	1473.0±3.8 <sup>a</sup>	395.0±1.4 <sup>c</sup>	1163.0±2.1 <sup>a</sup>	105.0±0.5 <sup>c</sup>
MS	67.4±0.14 <sup>a</sup>	514.0±3.1 <sup>c</sup>	732.0±1.9 <sup>b</sup>	441.0±1.6 <sup>c</sup>	441.0±0.8 <sup>b</sup>
PS	67.4±0.21 <sup>a</sup>	1191.1±2.3 <sup>b</sup>	1179.0±1.1 <sup>a</sup>	433.0±0.8 <sup>b</sup>	594.0±1.0 <sup>a</sup>

569 <sup>#</sup>Values followed by the different letter within a column differ significantly (\* $p < 0.05$ ).

570

571

**Table 4.** Digestibility of PS, MS and ASS.

Sample	RDS (%)	SDS (%)	RS (%)
ASS	10.5±1.6 <sup>a</sup>	20.6±1.7 <sup>a</sup>	68.9±1.1 <sup>c</sup>
MS	8.7±0.2 <sup>b#</sup>	15.2±1.8 <sup>b</sup>	76.0±2.1 <sup>b</sup>
PS	4.5±0.6 <sup>c</sup>	9.6±0.7 <sup>c</sup>	85.9±1.7 <sup>a</sup>

572 <sup>#</sup>Values followed by the different letter within a column differ significantly (\**p*< 0.05).

573

574 **Figure captions**575 **Fig.1** X - ray diffraction spectra analysis of different starches576 **Fig.2** SAXS curves of ASS, MS and PS.577 **Fig.3** Morphology (SEM: a, c, e; PLM: b, d, f) and size-distribution of starch granules

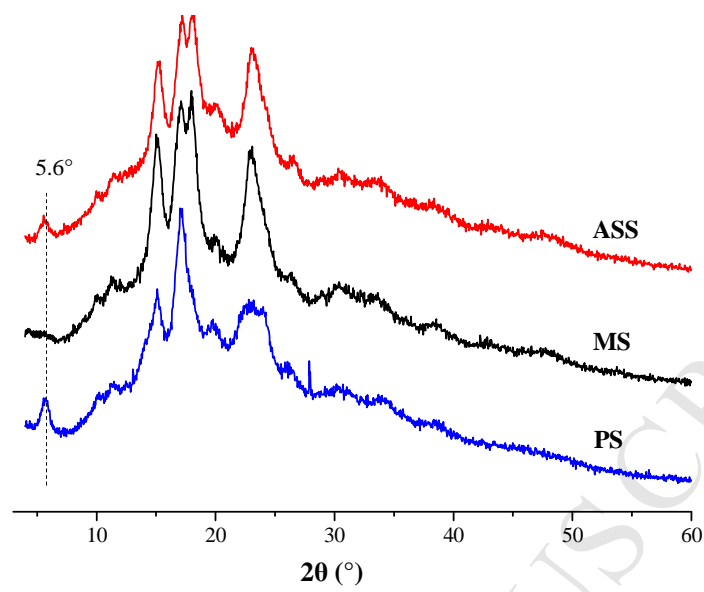
578 (g). ASS: a b; MS: c, d; PS: e, f. The yellow arrows showed in Figure 3e indicated the

579 channels or pinholes on starch granular surface.

580 **Fig. 4** Structural differences between ASS, MS and PS.  $d_L$ , long repeated distance of581 semicrystalline lamellae;  $d_a$ , average thickness of amorphous lamellae;  $d_c$ , average582 thickness of crystalline lamellae. The weak point indicates the  $\alpha$ -1,6-glycosidic

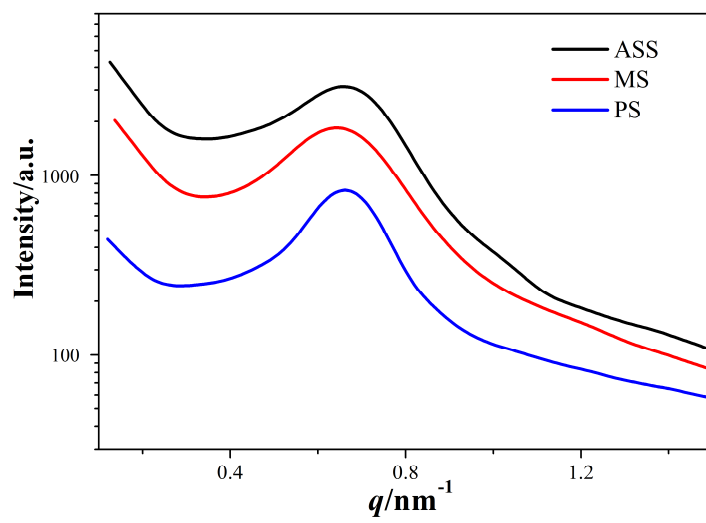
583 linkages which cluster in crystalline regions.

584

585 **Fig.1**

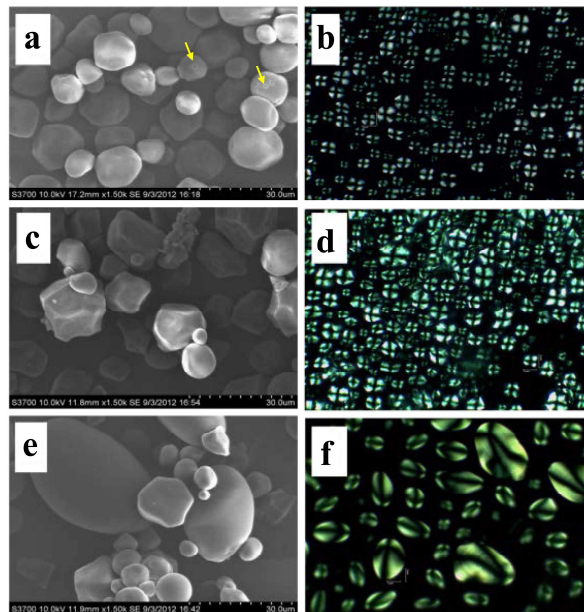
586

587

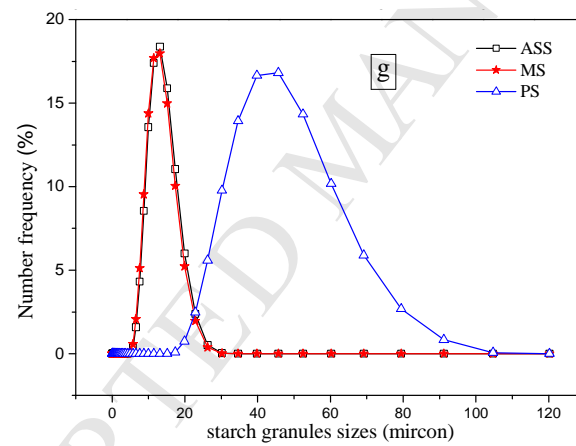
588 **Fig.2**

589

590

591 **Fig.3**

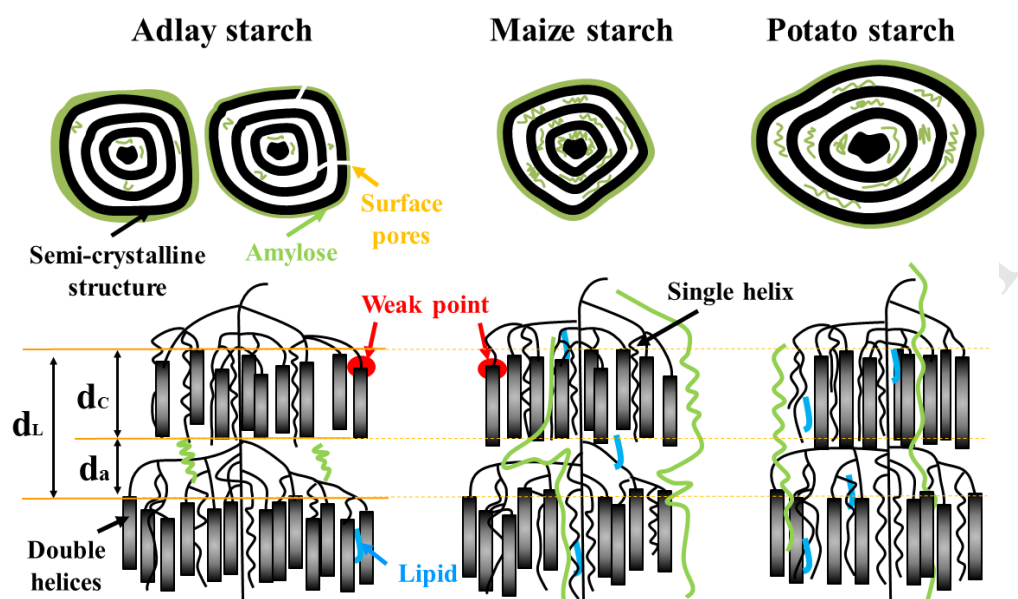
592



593

594



595 **Fig. 4**

596

**Highlights**

- ♦ Structural features of adlay seed starch were determined in comparison with normal maize starch and potato starch.
- ♦ Great differences were found comparing with normal maize and potato starches.
- ♦ Functionalities such as swelling power, solubility, pasting properties and *in vitro* digestibility were investigated.
- ♦ Factors determining starches functionalities were discussed.
- ♦ A comprehensive elucidation was proposed to reveal the structure-functionalities relationships of starches.