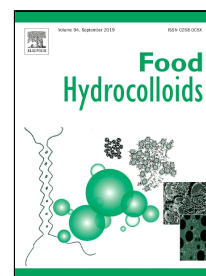


Accepted Manuscript

Rheological Properties and Structural Features of Coconut Milk Emulsions Stabilized with Maize Kernels and Starch

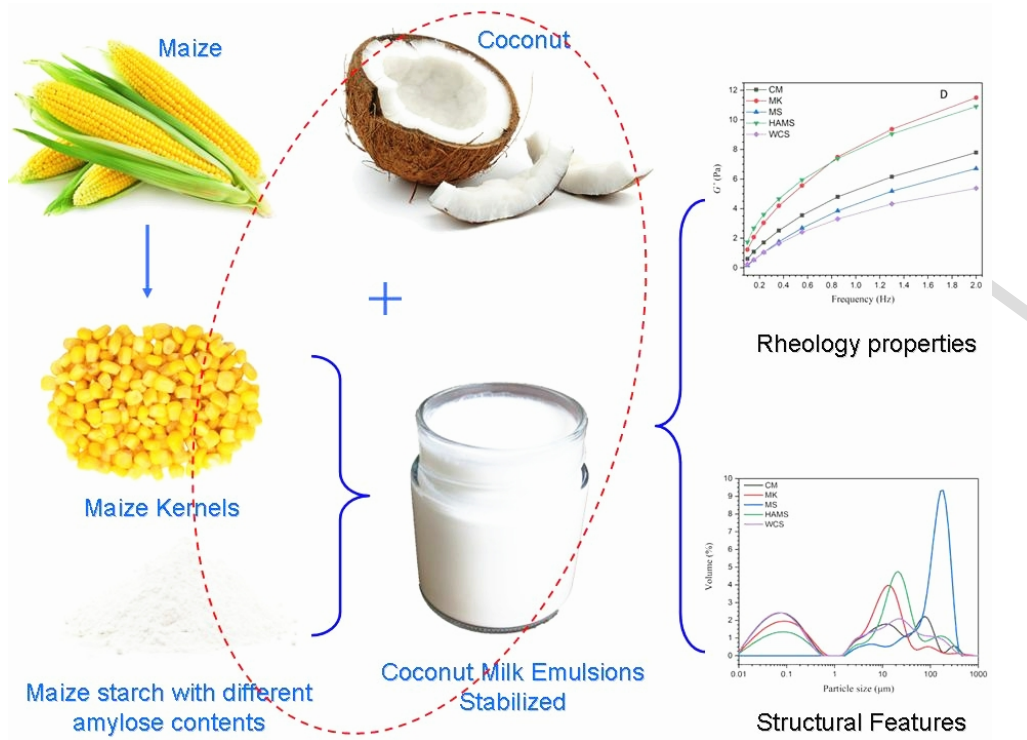
Xu Lu, Han Su, Juanjuan Guo, Jinjin Tu, Yi Lei, Shaoxiao Zeng, Yingtong Chen, Song Miao, Baodong Zheng



PII: S0268-005X(18)31777-6
DOI: 10.1016/j.foodhyd.2019.05.027
Reference: FOOHYD 5126
To appear in: *Food Hydrocolloids*
Received Date: 10 September 2018
Accepted Date: 15 May 2019

Please cite this article as: Xu Lu, Han Su, Juanjuan Guo, Jinjin Tu, Yi Lei, Shaoxiao Zeng, Yingtong Chen, Song Miao, Baodong Zheng, Rheological Properties and Structural Features of Coconut Milk Emulsions Stabilized with Maize Kernels and Starch, *Food Hydrocolloids* (2019), doi: 10.1016/j.foodhyd.2019.05.027

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



1 **Rheological Properties and Structural Features of Coconut** 2 **Milk Emulsions Stabilized with Maize Kernels and Starch**

3 Xu Lu^{a,b,c,d,e}, Han Su^{a,d,e}, Juanjuan Guo^{a,e}, Jinjin Tu^{a,d}, Yi Lei^a, Shaoxiao

4 Zeng^{a,c,d,e}, Yingtong Chen^{a,d}, Song Miao^{b,a,d,*}, Baodong Zheng^{a,c,d,e,*}

5 ^a College of Food Science, Fujian Agriculture and Forestry University, 15 Shangxiadian Road,
6 350002 Fuzhou, China;

7 ^b Teagasc Food Research Centre, Food Chemistry and Technology Department, Moorepark,
8 Fermoy, Co. Cork, Ireland;

9 ^c Institute of Food Science and Technology, Fujian Agriculture and Forestry University, 18 Simon
10 Pit Road, 350002 Fuzhou, China;

11 ^d China-Ireland International Cooperation Center for Food Material Science and Structure Design,
12 Fujian Agriculture and Forestry University, 350002 Fuzhou, China;

13 ^e Fujian Provincial Key Laboratory of Quality Science and Processing Technology in Special
14 Starch, Fujian Agriculture and Forestry University, Fuzhou 350002, China

15
16
17
18
19
20
21
22
23
24
25
26

***Corresponding Author and Email Address:** song.miao@teagasc.ie (S. Miao) and
zdbufst@163.com (B.D. Zheng)

27 **Abstract:** In this study, maize kernels and starch with different amylose contents at
28 the same concentration were added to coconut milk. The nonionic composite
29 surfactants were used to prepare various types of coconut milk beverages with optimal
30 stability, and their fluid properties were studied. The steady and dynamic rheological
31 property tests show that the loss modulus (G'') of coconut milk is larger than the
32 storage modulus (G'), which is suitable for the pseudoplastic fluid model and has a
33 shear thinning effect. As the droplet size of the coconut milk fluid changed by the
34 addition of maize kernels and starch, the color intensity, ζ -potential, interfacial tension
35 and stability of the sample significantly improved. The addition of the maize kernels
36 significantly reduced the size of the droplets ($p < 0.05$). The potential values of zeta (ζ)
37 and the surface tension of the coconut milk increased. Based on the differential
38 scanning calorimetry (DSC) measurement, the addition of maize kernels leads to an
39 increase in the transition temperature, especially in samples with a high amylose
40 content. The higher transition temperature can be attributed to the formation of some
41 starches and lipids and the partial denaturation of proteins in coconut milk, but phase
42 separation occurs. These results may be helpful for determining the properties of
43 maize kernels in food-containing emulsions (such as sauces, condiments, and
44 beverages) that achieve the goal of physical stability.

45 **Keywords:** coconut milk; starch; maize; amylose; emulsion; stability

47 1. Introduction

48 Coconut milk, which is a **type of oil-in-water** natural emulsion, is a coconut
49 processed product prepared by crushing, squeezing, extracting and other processing of
50 mature coconut endosperm. Coconut milk consists of coconut proteins (globulin and
51 albumin) and phospholipids. In coconut milk emulsions, the coconut protein adheres
52 to the surface of coconut oil as an emulsifier, preventing the coconut oil from
53 **flocculating** and merging. Coconut milk is rich in proteins, vitamins, sugars, amino
54 acid compounds, minerals, etc. Coconut milk contains 35.2% fat and 3.8% protein
55 (Ariyaprakai & Tananuwong, 2015; Raghavendra & Ksms, 2010). Coconut milk has
56 become **both** an important food emulsion and cooking material due to its unique
57 flavor and rich nutritional value (Iguttia, Pereira, Fabiano, Silva, & Ribeiro, 2011).
58 However, natural coconut milk, **like** most emulsions, is unstable and exhibits
59 stratification. During storage, due to its high fat content, natural coconut milk often
60 exhibits a floating fat layer, protein flocculation, sediment, etc. Even after repeated
61 homogenization, multiple layers (whey layer, cream layer, oil layer, etc.) appear **once**
62 **the product is left to stand**. The stratification of water and emulsion in coconut milk
63 has always been an unacceptable physical defect in **the processing and production of**
64 coconut milk (S. P. Ng, Lai, Abas, Hong, & Tan, 2014).

65 Studies have shown that the fat particle size, droplet size and **the degree of**
66 homogenization **have** significant effects on the stability of coconut milk. In addition,
67 the denaturation of coconut milk proteins caused by heating temperatures above 80°C
68 is among **one of** the factors affecting the stability of the coconut milk system

69 (Jirapeangtong, Siriwatanayothin, & Chiewchan, 2008; Nattapol & Johnn, 2009).
70 ChanthimaPhungamngoen, et al. studied the influence of homogenization steps
71 (before and after high temperature sterilization) on the quality of coconut milk, and
72 the results showed that compared with homogenization before single sterilization,
73 rehomogenization after sterilization can reduce the fat particle size of a coconut milk
74 system, thus improving the stability of coconut milk(Phungamngoen, Asawajinda,
75 Santad, & Sawedboworn, 2016). Jiang, et al. studied the effects of different
76 temperature treatments on the stability of coconut milk emulsions. The results showed
77 that the freezing treatment of coconut meat could effectively reduce the droplet size of
78 the emulsion, thus improving the stability of the emulsion. However, chemical
79 stabilizers are still commonly used to maintain the stability of beverage products due
80 to their advantages of low cost and simplicity of operation(Jiang, Xiang, & Wang,
81 2016). Yalegama et al. applied pasteurization technology at a low sterilization
82 temperature to coconut milk processing and searched for the optimum technology and
83 stabilizer to reduce the stratification of coconut milk under this processing condition.
84 The results showed that the stability of coconut milk could be effectively improved by
85 adding 0.5% sodium caseinate and 0.5% sodium stearylactylate as stabilizers and
86 sterilizing for 20-30 min at 72°C(Phungamngoen, et al., 2016). Currently, the variety
87 of coconut milk beverages is small, and compound beverages involving grain (corn,
88 rice, the seed of Job's tears, wheat, etc.) represent a breakthrough in dairy beverage
89 development. Understanding and controlling the stability of grains in coconut milk
90 systems are critical for the development of such beverages.

91 Corn, which is also known as Yumai, has a high nutritional value and is often
92 used as an ideal food for the elderly, patients and infants. Research by the German
93 Nutrition and Health Association shows that corn is currently the most nutritious and
94 healthful cereal among all staple foods. In addition, corn contains lutein and
95 zeaxanthin, which can delay visual aging, is rich in dietary fiber and has a mild taste.
96 In the food field, corn is mainly used in fresh foods; quick-freezing, starch-making
97 and corn oil; and corn fresh-pressed and corn-processed beverages; however, the sale
98 of fresh-pressed beverages is limited because of their high cost. Research
99 investigating corn beverage processing is limited, and corn coconut milk or other corn
100 drinks have not entered large-scale production to date. Furthermore, few studies have
101 investigated the stability and texture of this cereal compound beverage. Therefore, it
102 is of great significance to study the stability and texture of corn in coconut milk
103 systems for the development of compound cereal dairy products(L. Lin, et al., 2016).

104 The starch content in the corn dry base is as high as 71~74% and includes both
105 amylose and amylopectin. Due to the low gelatinization temperature and
106 retrogradation characteristics of starch, the dispersion of maize starch into a uniform
107 and stable system in aqueous solution is difficult and has become an important factor
108 affecting the stability of corn cereal beverages(L. Lin, et al., 2016; Xu Lu, et al.,
109 2019). Therefore, in this study, coconut milk was used as a liquid system, and maize
110 granules, common maize starch, waxy corn starch and high-amylose maize starch
111 were added to the coconut milk system. By studying the static and dynamic
112 rheological properties of coconut milk products, the effects of corn additives on the

113 texture (particle size, color difference, zeta potential, DSC, surface tension and
114 stability) of a coconut milk system were analyzed, providing a reference for studies
115 investigating the structure of cereal beverage solution systems.

116 **2. Materials and methods**

117 *2.1. Materials*

118 UHT-processed and aseptically packed coconut cream (Kara brand, Pt
119 PulauSambuGuntung Ltd., Indonesia, 30.8% fat, 3.9% protein, and 1.1%
120 carbohydrate), maize kernels (artificial stripping of fresh corn cobs; corn was
121 purchased from a local supermarket in Fuzhou, China), maize starch (Xinxiang Liang
122 Run Grain Foods Co., Ltd., Henan, China), high-amylose maize starch (Penford
123 Australia Pty Ltd., Lane Cove, NSW, Australia), and waxy corn starch (Qinhuangdao
124 Lihua Starch Co., Ltd., Hebei, China) were used. All chemicals and solvents were of
125 analytical grade.

126 *2.2. Preparation of coconut milk complex*

127 The sample types are divided into the following five groups: (1) pure coconut
128 milk without maize kernels and starch made of coconut cream (CM); (2) coconut milk
129 with the addition of corn products (MK); (3) coconut milk with the addition of maize
130 starch products (MS); (4) coconut milk with the addition of high-amylose maize
131 starch products (HAMS); and (5) coconut milk with the addition of waxy corn starch
132 products (WCS).

133 The process used adopted the following commercialized coconut milk formula:

134 6% (v/v) UHT coconut cream, sucrose (7%, w/v), sodium bicarbonate (0.038%, w/v),

135 sodium citrate (0.05%, w/v), D-sodium erythorbate (0.05%, w/v), EDTA (0.025%,
136 w/v) and 1 L water with the addition of a certain quantity of fresh corn kernels and
137 maize starch with different amylose contents (the nutritional components of each
138 maize starch sample are shown in Table 1). The determination of the starch content
139 added to MS, HAMS or WCS in coconut milk was carried out using samples with **the**
140 same starch content in coconut milk with maize kernels based on the AOAC Method
141 2002.02. The amount of starch released from the corn coconut milk was 3.956 g/L.
142 Based on the amount of maize starch, different amylose contents in the MS, HAMS or
143 WCS groups and compound emulsifiers were applied (the proportions are shown in
144 Table 2). The optimal stability formula of the five groups of samples was obtained by
145 optimizing the ratio of the composite emulsifier using the response surface method.
146 Subsequently, amylose and the compound emulsifiers were mixed in a high-shear
147 blender (13500 rpm, 3 min, Ultra Turrax T25, IKA, Germany) to prepare the coconut
148 milk emulsion oil-in-water beverage. Then, the coconut milk was canned, sealed, and
149 sterilized at 115°C for 20 min under high pressure in an autoclave (Zealway,
150 G154DWS, Xiamen, China). Furthermore, testing was performed after cooling.

151 *2.3. Rheological analysis*

152 The rheological properties of the coconut milk were assessed using a rotational
153 rheometer (Physical MCR 301; Anton Paar, Co., Ltd., Stuttgart, Germany) with a
154 parallel plate sensor (60 mm diameter, 1 mm gap). The temperature was maintained at
155 25 °C during these rheological measurements.

156 *2.3.1. Steady rheology measurements*

157 The steady-state shear experiments were carried out at a shear rate ($\dot{\gamma}$) range of
158 0.001–100 s⁻¹ for 300 s in the CR mode (controlled shear rate) at 25 °C using different
159 annular gap sizes between the parallel plate geometry with gaps of 1.0 mm in parallel
160 plate geometries with grooved surfaces. The steady-state shear experiments
161 investigating the coconut milk were carried out at a shear rate range of 0.01–
162 100 s⁻¹ for 300 s in the CR mode at 25 °C with an annular gap size of 1.0 mm in
163 parallel plate geometries with smooth surfaces. The experimental data of the flow
164 curve were obtained and fitted using the Herschel–Bulkley model.

$$165 \tau = \tau_0 + K\dot{\gamma}^n \quad (1)$$

166 where τ is the shear stress (Pa), τ_0 is the yield stress (Pa), $\dot{\gamma}$ is the shear rate
167 (s⁻¹), K is the consistency coefficient (Pa·sⁿ) and n is the flow behavior index.

168 2.3.2. *Dynamic rheology measurements*

169 Frequency sweep measurements: The frequency sweep tests were performed at a
170 frequency range of 0.1–2 Hz with constant deformation (0.5% strain) within the linear
171 viscoelastic range. The 0.5% strain was within the linear viscoelastic region according
172 to the strain sweep results. The mechanical spectra, which recorded the storage
173 modulus (G'), loss modulus (G''), and loss tangent ($\tan \delta = G''/G'$) as functions of the
174 frequency (Hz), were obtained.

175 2.4. *Particle size distribution (PSD) analysis*

176 The particle size was determined using the laser diffraction particle size analyzer
177 MasterSizer 3000 (Malvern Instruments Ltd., Malvern, Worcestershire, UK) equipped
178 with a wet sample dispersion unit (Malvern Hydro MV, UK). The background and

179 sample integration times were 20 and 10 s, respectively. The optical properties were
180 defined as a refractive index of 1.549 (olive oil) and 1.330 (dispersant water) and
181 absorption index of 0.001 using a normal instrument. Across a dynamic spanning
182 range of 0.01-3500 μm , before the analysis, all samples were shaken sufficiently to
183 ensure sample uniformity. In the sample port, the samples were dispersed in distilled
184 water at 2000 rpm until an obscuration of 8–25% and polarization intensity
185 differential scattering of 8~15% were achieved. The analysis of the samples was
186 performed in triplicate. The surface-area-based mean diameter (D[3,2]) and
187 volume-based mean diameter (D[4,3]) were also obtained. Among these particle
188 diameters, the mode diameter represents the most common particle size observed;
189 regarding the mean diameters, D[4,3] is highly influenced by large particles, and
190 D[3,2] is more influenced by smaller particles. The size distribution was expressed as
191 the surface-weighted mean diameter as follows:

$$192 \quad D[4,3] = \frac{\sum_i^{(j)} n_i d_i^4}{\sum_i^{(j)} n_i d_i^3} \quad (2)$$

$$193 \quad D[3,2] = \frac{\sum_i^{(j)} n_i d_i^3}{\sum_i^{(j)} n_i d_i^2} \quad (3)$$

194 where n_i is the number of droplets of diameter d_i . Additionally, the dispersion
195 index (Span) was calculated.

196 2.5. Color analysis

197 The color measurements were performed using a CS-200 Spectrophotometer
198 (Hangzhou CHNSpec Technology Co., Ltd., Hangzhou, China). White calibration
199 was used for the instrument standardization. The samples were placed in glass cells,

200 and the measurement was performed. The L^* , a^* , b^* color space was used for the
201 measurement. The L^* , a^* , and b^* values indicate luminosity, chromaticity on a green
202 (–) to red (+) axis, and chromaticity on a blue (–) to yellow (+) axis, respectively. For
203 each treatment, the color of three samples was measured. The total color change (ΔE)
204 was calculated using the following formula:

$$205 \quad \Delta E = \sqrt{(L^* - L_0^*)^2 + (a^* - a_0^*)^2 + (b^* - b_0^*)^2} \quad (4)$$

206 where subscript '0' indicates the initial color of the coconut milk without maize.

207 2.6. Surface tension analysis

208 The surface tension was measured by a BZY-1 automatic surface tension meter
209 (Shanghai Equity Instruments Co., Ltd., Shanghai, China) at 25°C. The BZY-1 meter
210 employs the Wilhelmy plate principle, *i.e.*, the maximum tensile force competing with
211 the surface tension is measured when the bottom edge is parallel to the interface and
212 touches the liquid. The temperature and surface tension measurement ranges are
213 (268.15–383.15) K and (0.1–400.0) mN·m⁻¹, respectively. The uncertainty is
214 ±0.1 mN·m⁻¹. The size-volume of the different samples used in BZY-1 m was 20 mL.
215 During the experiments, the copper pan in the host of BZY-1 m was connected to a
216 thermostatic bath (CH-1006, uncertainty is ±0.1 K). Via the circulation of the water,
217 the temperature of the water in the copper pan was kept the same as that in the
218 thermostatic bath. The aqueous solution was placed in a solution container immersed
219 in the copper pan, and its temperature was measured by a thermocouple. The scale
220 reading of the thermocouple had been well calibrated by a mercury thermometer.

221 2.7. ζ -potential analysis

222 Each sample (1 mL) was added to the sample pool, and the particle charge of the
223 coconut milk (ζ -potential) was determined using a Zetasizer Nano-ZS (Mastersizer X,
224 Malvern Instrument Ltd., Worcestershire, U. K.) based on laser Doppler velocimetry.
225 All experiments were carried out at 25°C. The analysis of the samples was performed
226 in triplicate.

227 *2.8. Differential scanning calorimetry (DSC)*

228 The coconut milk was also analyzed using Netzsch DSC 204 F1 Phoenix®
229 (Netzsch Group, Selb, Germany) equipment, and the phase transitions were tracked.
230 Aliquots of protein dispersions (20 μ L) were accurately injected into aluminum pans
231 that were sealed with a Tzero hermetic lid. An empty pan was used as a reference.
232 The DSC sample pans were heated from 30 to 100°C at 10°C/min. All DSC analyses
233 were performed under nitrogen at dynamic purge and protective flows of 20 mL/min.
234 Proteus® Software for Thermal Analysis ver. 6.1.0 (Netzsch Group, Selb, Germany)
235 was used for the acquisition and processing of the DSC data.

236 The onset (T_0) and offset (T_e) temperatures are defined as the intersections
237 between the tangents of the peak and the extrapolated baseline, and the peak
238 denaturation temperature (T_p) is defined as the temperature at the maximum/minimum
239 of the thermal event. These temperatures were obtained from a corresponding
240 thermogram, while the denaturation enthalpy (ΔH) was obtained from an integration
241 of the thermogram. The analyses were carried out in triplicate.

242 *2.9. Compound system stability*

243 The creaming stability of the emulsions was evaluated using a multisample

244 analytical centrifuge (Lumifuge, LUM GmbH, Berlin, Germany). Coconut milk (425
245 μL) was transferred to rectangular cells (2×8 mm) and analyzed by a light beam
246 emitted at a near infrared wavelength (880 nm) that scanned the sample cells over the
247 total length. The charge-coupled device (CCD) line sensor received the light
248 transmitted through the sample, which showed the pattern of light flux as a function of
249 the radial position, providing a macroscopic fingerprint of the sample at a given time;
250 based on this pattern, emulsion instability, including creaming, sedimentation, and
251 droplet aggregation, could be detected. In the current study, the samples were
252 centrifuged at 4000 rpm ($2320 \times g$) and 25°C at a scanning rate of once every 60 s for
253 1 h, simulating approximately 4 months of separation under normal gravity.

254 The separation rates were determined using the software package SepView 4.1
255 (L.U.M GmbH). The velocities of the separation of individual particles (mm/d) were
256 measured from the measurement results. Following the test, curves of the integrated
257 transmitted light against time were plotted, and the slope of each curve was
258 considered the Creaming Index (*CI*).

259 2.10. Statistical analysis

260 The statistical analysis was performed using OriginPro 8.0 (Origin Lab
261 Corporation, USA), and the data are expressed as the mean values \pm standard
262 deviation. All measurements were repeated three times. All statistical analyses were
263 performed using Data Processing System software (DPS, V9.05, Science Press,
264 Beijing, China). One-way ANOVA, followed by Duncan's post hoc test, was used to
265 determine the significant differences among the treatment groups. $p < 0.05$ was

266 considered statistically significant; all tests were two-sided, and no corrections were
267 applied for multiple significance testing.

268 **3. Results and discussion**

269 *3.1. Formula of composite stabilizer*

270 The results shown in Table 1 highlight that the amylose content was the highest
271 in the high-amylose maize starch, followed by the amylose content in the kernels and
272 common corn starch, while the amylose content in the waxy corn starch was the
273 lowest, and its main component was amylopectin. Compared with common corn
274 starch, the lipid and protein content in the corn kernels is higher than that in the
275 common corn starch.

276 The results shown in Table 1 highlight that the amylose content in the
277 high-amylose maize starch is the highest, followed by the amylose content in the
278 maize kernels and common corn starch, while the amylose content in the waxy corn
279 starch is the lowest, and its main component is amylopectin. Compared with the
280 common corn starch, the lipid and protein content in the maize kernels is higher than
281 that in the common corn starch. During the early stage of the study, based on the
282 response surface optimization experiment, the optimum stabilizer formula of different
283 corn additives and coconut milk was determined.

284 According to the emulsifier formula (Table 2), the ideal amount of carrageenan
285 that should be added to coconut milk with common corn starch and waxy corn starch
286 is 0.03%, while the amount of carrageenan that should be added to coconut milk with
287 maize kernels is 0.05%. The influencing factor may be that the maize kernels contain

288 up to 70% starch in addition to 4% oil, while carrageenan is a highly charged
289 emulsifier with a negative charge that can form a layer of high charge density
290 interfacial film around oil droplets, protecting the oil droplets and stabilizing the
291 emulsion. Therefore, more carrageenan is needed in corn coconut milk to stabilize the
292 system. Sodium tyrosinate is an amphiphilic surfactant with better thermal stability
293 than other emulsifiers (Z. Long, et al., 2012). The results showed that the maize
294 kernel-coconut milk needed the least amount of surfactant, while the waxy corn
295 starch-coconut milk required the greatest amount, indicating that the maize
296 kernel-coconut milk system was more stable.

297 The amount of sucrose fatty acid ester in the waxy corn starch coconut milk was
298 0.06% more than that in the common corn starch coconut milk. We speculated that the
299 sucrose fatty acid ester was a strong hydrophilic emulsifier that could wrap
300 amylopectin maize starch more effectively, which is then not soluble in water
301 (Watanabe, Kawai, & Nonomura, 2017). Furthermore, the content of distearin was
302 higher in the coconut milk system with the maize kernels, common corn starch and
303 high-amylose maize starch, while the content of glyceryl monostearate was the
304 highest in the high-amylose maize starch-coconut milk system. Glyceryl stearate is a
305 polyol type nonionic surfactant that inhibits the aging of starch. Glycerin
306 monostearate easily forms a starch-lipid complex with amylose, which improves the
307 stability of the composite system (Garcia & Franco, 2015). Therefore, the higher the
308 amylose content, the more stearin is required to stabilize the system.

309 *3.2. Steady rheological properties of coconut-corn systems*

310 The apparent viscosity curve of a coconut milk system is determined by setting
311 the shear rate at 0.001-100 s⁻¹. The shear rate is measured by using a groove with a
312 geometric plane to avoid surface slipping. As shown in Figure 1A, under the same
313 preparation, sterilization and measurement conditions, the apparent viscosity of the
314 five samples initially increases and then decreases as the shear rate increases and **the**
315 **rate of decrease slows down.**

316 The maize starch with different amylose **concentrations** increased the apparent
317 viscosity of the coconut milk, and the difference in the rheological properties was
318 partially due to the solubilization of macromolecular carbohydrates, such as starch.
319 Here, solubilization indicates that the starch macromolecular aggregates are
320 gelatinized and decomposed into colloids. The apparent viscosity of the coconut milk
321 containing the high-amylose maize starch was the highest, which might be due to the
322 gelation of high-amylose starch, increasing the apparent viscosity (Shao, Tseng,
323 Chang, Lin, & Lii, 2007). A similar phenomenon is observed in the corn starch fluid
324 with different amylose ratios of solvent in pure water systems (Xie, et al., 2009). **The**
325 maize kernel and common corn starch-coconut milk systems have a similar apparent
326 viscosity. Under high-pressure and high-temperature sterilization conditions, the
327 linear amylose molecule might dissolve first, followed by the mild amylopectin
328 molecule, and finally, amylopectin **would then dissolve.** The short-time
329 high-temperature sterilization resulted in a higher amylose content.

330 The maize kernel and common corn starch contained a similar content of
331 amylose. Gelatinization of amylose requires a higher temperature, leading to a

332 multiphase transition between starches and an increase in viscosity and shear stress
333 due to the winding of macromolecules between linear polymer chains. The main
334 component of waxy corn starch is amylopectin. The entanglement and interconnection
335 between droplets of stable powder particles are very weak because there is almost no
336 amylose. The three-dimensional network formed by amylose is relatively weak. After
337 high temperature sterilization, amylopectin is mainly composed of short-branched gel
338 spheres that are mainly composed of chains from the same subchain. Compared with
339 amylose, a gel network structure is not easily formed, and the size and length of the
340 chain (only 4-6 glucose) and the molecular entanglement between gel spheres are
341 much smaller than those between linear polymer chains (Han, Campanella, Mix, &
342 Hamaker, 2002; Y. Long & Christie, 2005). Once the mechanical deformation is
343 strengthened, the network undergoes a gradual decomposition into smaller clusters;
344 thus, the apparent viscosity is smaller (Xuanxuan Lu, Wang, Li, & Huang, 2018). In
345 summary, the higher the amylose content in a coconut milk system, the greater the
346 apparent viscosity.

347 The fluidity of coconut milk is affected by the addition of corn and affects its
348 shear thinning characteristics. As shown in Fig. 1B, the shear stress of the five
349 coconut milk samples increased with the rate of what, indicating that all samples have
350 the characteristics of pseudoplastic fluid with shear thinning. This finding may be
351 explained by the non-Newtonian behavior (shear-induced alignment) of the droplet
352 network or shear continuous phase (Liu & Tang, 2011), which is consistent with the
353 results of the apparent viscosity.

354 At the same shear rate, the shear stress of the coconut milk samples containing
355 the waxy corn starch was always lower than that of the other samples. We inferred
356 that the other four coconut milk samples were more easily shear thinned and had
357 strong pseudoplasticity. The shear stress of the coconut milk containing the
358 high-amylose maize starch and maize kernels was always greater than that of the pure
359 coconut milk. The high yield stress proved that there was a strong network. Studies
360 have shown that potato starch with a high amylose content gelled more easily than
361 potato starch with a low amylose content at a high temperature; thus, the shear stress
362 is stronger (Zhou, et al., 2015). During the process of autoclaving, starch granules
363 burst, and amylose precipitates first (Ji, et al., 2017). However, due to the influence of
364 the spatial structure of starch, amylose has a stronger ability to form hydrogen bonds
365 than amylopectin and solvent molecules, and the structure after binding is more
366 stable, reducing the fluidity of the coconut milk system and increasing the shear
367 stress. Compared with common corn starch, there are more lipids and protein
368 substances in maize kernels, and the fat is easily adsorbed into agglomerates due to
369 the oil-water layer structure; thus, the shear thinning effect of maize kernels is larger
370 than that of common corn starch (Schröder, Berton-Carabin, Venema, & Cornacchia,
371 2017).

372 The flow behavior index (n) indicates the degree of the pseudoplasticity of an
373 emulsion as follows: as the pseudoplasticity increases, the n value decreases. Native
374 maize starch and waxy maize starch have more pseudoplasticity because they exhibit
375 a lower flow behavior index. If the amylopectin content is high, the amylopectin

376 solution does not exhibit effective entanglement because there is no entanglement
377 between amylose chains, and the n value is higher (Y. Long, et al., 2005). The flow
378 characteristic index of all coconut milk systems is less than 1 (Table 3), and the
379 apparent viscosity decreases as the shear rate increases, reflecting non-Newtonian
380 flow and shear thinning flow behavior (Tanner & Rivlin, 1985). All fluids are
381 pseudoplastic fluids, and the addition of corn kernels or starch does not alter the fluid
382 type of coconut milk. The consistency coefficient (k) is an index of the emulsion
383 viscosity. Compared with pure coconut milk, maize starch, common starch and
384 high-amylose maize starch can increase the consistency coefficient of a system, and a
385 higher amylose content leads to a higher K value.

386 3.3. *Dynamic rheological properties of coconut-corn systems*

387 The dynamic rheological property (dynamic viscoelasticity) refers to the
388 mechanical response law of the material under the action of alternating stress and is
389 detected by elastic (G') and viscous (G'') moduli as a function of frequency. Coconut
390 milk is a beverage with a higher fat and vegetable protein content; the fat is easily
391 aggregated, and the system is stratified (Jiang, et al., 2016). Once maize starch
392 additives are introduced to the system, the rheological properties of the whole liquid
393 system changes under the action of high-speed shearing and high-temperature
394 processing, thereby affecting the quality. The storage modulus G' can reflect the
395 elasticity of the fluid, the recovery ability of the reaction fluid after deformation, the
396 energy modulus G'' , the size of the viscous component and the ability of the fluid to
397 release energy, which reflects the performance of the fluid in resisting the flow

398 (Barbieri, et al., 2018).

399 At a scanning range of 0.1-2 Hz, G' and G'' of the coconut milk samples increase
400 as oscillation frequency increases. After adding corn starch additives, G' and G'' both
401 showed a significant increasing trend (Fig. 1D and E), indicating that the dynamic
402 modulus depends on a strong frequency, and the loss modulus exceeds the storage
403 modulus, reflecting fluid-like behavior. Moreover, G' and G'' of the high-amylose
404 maize starch were larger than those of the other samples. The waxy corn starch
405 sample exhibited the smallest G' and G'' , and the emulsion had the weakest structure.
406 It's lowest G' value proved this finding and resulted in a WCS with a very high
407 droplet fluidity. The gel network structure formed by amylose increases the elasticity
408 of the liquid system, while amylopectin does not easily form a gel network structure
409 (Yu, Jing, Zhang, & Kopparapu, 2014). Therefore, the higher the amylose content, the
410 greater the dynamic viscoelasticity of a coconut milk system. Studies have shown that
411 the rheological viscoelasticity and viscosity of rice starch are closely related to its
412 amylose content, and the greater the amylose content, the greater the viscoelasticity
413 (Shao, et al., 2007; Takahashi & Fujita, 2017).

414 In addition, both the complex viscosity η^* and the tangent loss $\tan\delta$ of the sample
415 showed a tendency to decrease as the scanning oscillation frequency increased (Fig.
416 1C and F), which is consistent with the trend observed in the static rheological
417 research. The tangent loss of all coconut milk systems is related to the scanning
418 frequency, is greater than 1, and gradually approaches 1 as the frequency increases.
419 The tangent loss $\tan\delta=G''/G'$ reflects the relationship between the elastic and viscous

420 characteristics of reactive fluids. If $\tan\delta < 1$, the elastic properties of fluids dominate,
421 and if $\tan\delta > 1$, the viscous properties of fluids dominate. Following the addition of
422 maize kernels or high-amylose maize starch, coconut milk fluid tends to be rigid in a
423 three-dimensional network, and MS and WCS systems do not exhibit obvious gel
424 properties (Yu, et al., 2014). The decrease in the complex viscosity (η^*) may be
425 related to a shrinkage of starch swelling particles, depolymerization of protein or the
426 destruction of protein networks caused by protein thermal coagulation (Vanin,
427 Michon, & Lucas, 2013). In addition, the decrease in energy storage and loss modulus
428 results in a decrease in the hydrogel network strength.

429 *3.4. Changes in the particle size distribution*

430 Figure 2 shows the distribution of the particle size and particle radius of the
431 coconut milk system after the addition of the maize starch supplement. The
432 measurements of the area-based mean particle diameter ($D[3,2]$) and volume-based
433 mean diameter ($D[4,3]$) are shown in Table 4. $D[4,3]$ reflects the size of flocculated
434 droplets or solid aggregates. The pure coconut milk system showed three peaks in the
435 particle size distribution, and was mainly concentrated in the range of 0.01-100 μm . A
436 few particles were larger than 100 μm , and the average particle size was 28.6 μm .
437 Coconut milk is mainly composed of proteins and phospholipids, which is similar to
438 most emulsions. Fresh coconut milk is unstable and easy to be stratification. After
439 processing, coconut milk can be divided into cream and whey, which are known as
440 coconut cream and coconut skim milk, respectively. Even after emulsification,
441 coconut milk contains 3 to 4 phases, including an oil phase, cream phase, whey phase

442 and sediment layer. Coconut protein can be used as a natural emulsifier in coconut
443 milk, but following thermal denaturing, coconut protein loses the ability to stabilize
444 emulsion droplets (C. Y. Ng, Mohammad, Ng, & Jahim, 2014; Raghavendra, et al.,
445 2010). The size distribution of pure coconut milk exhibits three peaks at 0.01~1 μm ,
446 4~40 μm , and 40~110 μm , and the second group corresponds to fat globules at a
447 lower particle size (0.01~10 μm). Pure coconut milk emulsion droplets of
448 approximately 100 μm may be white solid particles produced by the condensation of
449 denatured proteins (Nattapol, et al., 2009), while with a high-amylose maize starch
450 and high-amylose maize kernel content, the emulsion particles aggregate in small size
451 directions, indicating that these additions can reduce the denaturation process of the
452 corresponding protein in coconut milk. The addition of both can enhance the heating
453 stability of coconut milk.

454 After the addition of the maize starch additives to the coconut milk, the D[3,2]
455 and D[4,3] of the system were significantly affected ($p<0.05$). Among these additives,
456 the common corn starch had the most significant impact on the particle size
457 distribution and particle size of the coconut milk ($p<0.05$), and D[4,3] and D[3,2] both
458 sharply increased. However, the high-amylose maize starch and maize kernels had
459 similar effects on the particle size distribution of the coconut milk system. Under the
460 influence of both additives, the peak of the D[3,2] value between 40 and 110 μm
461 disappeared, and the peak between 4 and 40 μm increased, while the peak of D[4,3]
462 obviously increased. The waxy corn starch had no significant effect on the distribution
463 of D[4,3] in the coconut milk system ($p>0.05$), but the distribution of D[3,2] became

464 more uniform and concentrated.

465 Studies have shown that D[3,2] is more susceptible to small molecular particles,
466 while D[4,3] is more susceptible to macromolecular particles (Ped, Ibarz, &
467 Cristianini, 2013). The grain size of common corn starch is significantly larger than
468 that of high-amylose maize starch (L. Lin, et al., 2016), and the grain size of waxy
469 corn starch is 16.4 μm (J. H. Lin, Kao, Tsai, & Chang, 2013), which is less than the
470 average particle size of coconut milk. Therefore, once maize grain or maize starch is
471 added to a coconut milk system, the average particle size of the system is affected to
472 varying degrees. Among these additives, common corn starch increases the size of the
473 system particles. The larger the particle size, the higher the number of starch granules
474 required to cover the same interface area probably because more starch granules are
475 required to stabilize a larger interface area. As the surface coverage of the droplets
476 increased, and the starch granules showed monolayer compact accumulation after
477 gelatinization. The dense accumulation of starch particles at the interface can enhance
478 the coalescence stability of adjacent droplets by forming a strong spatial barrier (Li,
479 Li, Sun, & Yang, 2013). Waxy corn starch can reduce the size of coconut milk
480 particles, resulting in a larger specific surface area, and as the average distance
481 between the particles decreased, the better the dispersion of the smaller particles in the
482 coconut milk. In addition, the surface active component adsorbed on the oil droplets
483 easily generates a noncovalent interaction with the hydrophobicity and corn bonding
484 between the waxy corn starch molecules, leading to a stronger interparticle
485 interaction, and the formed emulsion exhibited good anti-coalescence stability during

486 storage (Bi, Hemar, Balaban, & Liao, 2015; B. Wang, Wang, Li, Adhikari, & Shi,
487 2011; Yusoff & Murray, 2011).

488 3.5. Changes in color, surface tension and ζ -potential

489 The L^* value of the coconut milk system decreased after the addition of the
490 maize supplement, while the a^* value showed the opposite result (Table 5). The
491 deepening of the color caused by the addition of maize kernels may be attributed to
492 the migration of pigments from the maize kernels, the oxidation of carotenoids and
493 the increase in free phenolics during processing, causing coconut milk nonenzymatic
494 browning of the Maillard type (Rochavillarreal, Hoffmann, Vanier, Sernasaldivar, &
495 Garcíalara, 2018). In addition, the waxy corn starch significantly increased the a^*
496 value of the system ($p < 0.05$), while the high-amylose maize starch and maize kernels
497 had a negligible effect on the a^* value of the system. Most likely, this result is due to
498 the large number of Maillard reactions between the added reducing sugars and the
499 amino groups in the protein that occur under high temperature sterilization
500 (Rodsamran & Sothornvit, 2018). Amylopectin caused the loss of the luminosity of
501 the system and increased the red color intensity of the system, while the high-amylose
502 content had less of an effect on the red color of the system. Most likely, due to the
503 large particle size of the amylopectin particles, the macromolecular particles in
504 coconut milk are easily susceptible to precipitation, affecting the depth of the system.
505 In addition, the ΔE value of the corn supplement system significantly increased
506 compared to that of the pure coconut milk system.

507 One of the most important parameters of the stability of food emulsions is the

508 droplet size because emulsion coalescence with larger droplets is faster, and a smaller
509 particle size is more stable. The coalescence rate constant is inversely proportional to
510 the ζ -potential, and the ζ -potential is related to the surface charge of the emulsion
511 droplet. The larger the net surface charge, the less coalescence (Das & Kinsella,
512 1990). After the different corn starch additives were added to the system, the
513 ζ -potential of the pure coconut milk became less negative and even became positive
514 (Table 5). The ζ -potential refers to the mutual exclusion or attraction between
515 particles. The smaller the dispersed particles, the larger the ζ -potential absolute value
516 and the more stable the system. The smaller the ζ -potential absolute value, the easier
517 the condensation or agglomeration. The maize kernels and high-amylose maize
518 kernels reduced the ζ -potential of the coconut milk to -7.93 mV and -0.7 mV,
519 respectively, suggesting that the system began to appear unstable, and the attraction
520 between the particles exceeded the rejection. After the common corn starch and waxy
521 corn starch were added, the ζ -potential of the system became positive at 1.38 mV and
522 1.39 mV, respectively; thus, a part of the system produced condensation or
523 agglomeration phenomenon, which may have affected the stability of the system. The
524 increase in the ζ -potential of the coconut milk after the addition of the maize kernels
525 and maize starch may be due to (1) the potential change in the starch granule surface
526 protein from hydrophilicity to hydrophobicity after heating, and the hydrophobicity of
527 starch is also improved (Li, et al., 2013). (2) In addition, electrostatic interactions
528 between adsorbed and unadsorbed proteins and polysaccharides in the coconut milk
529 could also be induced (Evans, Ratcliffe, & Williams, 2013). (3) Once a surfactant is

530 added to a coconut milk emulsion before emulsification, the small molecule surfactant
531 completely replaces the coconut protein adsorbed onto the newly created interface,
532 resulting in a decrease in negative charge on the surface protein and a change in the
533 ζ -potential of the emulsion droplet. The addition of a nonionic surfactant can provide
534 space stability and eliminate the electrostatic effects of the protein. If the protein on
535 the emulsion droplets is completely replaced, the nature of the emulsion should be the
536 same as that of the surfactant being replaced (McClements, 2005).

537 Many products with oil-in-water (O/W) or water-in-oil (W/O) emulsions contain
538 emulsifiers or stabilizers to reduce the interfacial tension between the oil and water
539 phases, thereby increasing the stability of the system. Surface tension plays an
540 important role in determining the ability to form and stabilize an emulsion. The
541 surface tension of water is 72 mN/m, and corn oil has a surface tension value of 30.96
542 mN/m at 25 °C (Nitschke, et al., 2010). Compared with natural proteins, the partial
543 denaturation of proteins in pure coconut milk and the interaction between protein
544 molecules can cause the formation of protein aggregates. When such autoclaving
545 aggregation occurs at a sufficiently high protein concentration, oil can be embedded in
546 the three-dimensional matrix of the aggregates. Moreover, heat-treated proteins
547 exhibit a lower surface tension due to protein denaturation and the increase in surface
548 hydrophobicity (J. Wang, et al., 2012). The surface tension and interfacial elasticity of
549 the emulsion protein film were decreased by the addition of the surfactants of glyceryl
550 monostearate, distearin, sucrose fatty acid ester and sodium caseinate corresponding
551 to the ratio of surfactant to protein. Thus, the addition of maize kernels and starch

552 increased the surface tension of the system. There was no significant difference in the
553 surface tension between the maize kernels and coconut milk with different amylose
554 contents ($p>0.05$). Starch polysaccharides have no surface activity, but they bind
555 surfactants, and their binding capacity is independent of the type of starch. The
556 combination of surfactant and amylose is synergistic, while the combination of
557 surfactant and amylopectin is a Langmuir type (Lundqvist, Eliasson, & Olofsson,
558 2002).

559 3.6. Changes in thermal properties

560 The endothermic conversion of pure coconut milk systems occurs between 58
561 and 71°C. The endothermic enthalpy produced by the initial gelatinization
562 temperature may be related to the denaturation temperature of some globulins in
563 coconut milk. Reheating after high pressure sterilization may cause the inner part of
564 the protein to continue denaturing (Nattapol, et al., 2009). The addition of corn starch
565 additives changed the endothermic conversion of the system to different degrees.
566 HAMS significantly increased the T_0 of the system ($p<0.05$), followed by maize
567 kernels, and common corn starch and waxy corn starch slightly increased the T_0 .
568 HAMS and MK increased the T_p of the system, whereas the common corn starch and
569 waxy corn starch slightly decreased the T_p (Table 6). In addition, the T_e value of the
570 system increased following the addition of maize starch, and the order
571 was HAMS>MK>WCS>MS. Compared with the pure coconut milk system, the ΔH
572 of HAMS and MK increased by 6.8/J·g⁻¹ and 6.56/J·g⁻¹, respectively. Granules of high
573 amylose maize starch, especially small size granules, still retain some Maltese cross

574 characteristics when heated at 120°C(Xu, et al., 2017).Therefore, a wide range of
575 endothermic gelation temperatures of high amylose is still obtained in the temperature
576 range of 65 to 125°C (Haralampu, 2000), and the wider peak value is attributed to the
577 overlapping of transitions, which is related to starch gelatinization and the
578 denaturation of some proteins.The superfluous endothermic enthalpy of coconut milk
579 with corn kernel, maize starch and high-amylose maize starch may represent the phase
580 transition of the amylose-lipid complex. The type I composite is formed at or below
581 60°C and results in the formation of a single spiral segment with a random
582 orientation.However, the complex formed at 70°C may be a mixture of type I and type
583 II crystals.Reaching 90°C results in the formation of a starch-CLA complex mainly
584 composed of type II crystals. The detection of a higher temperature in starch rich in
585 amylose can be used to explain its higher viscosity and reduced Newtonian
586 behavior.Regarding MS and WCS, there is no significant impact on the ΔH of the
587 system ($p>0.05$). The gel spheres formed by amylopectin require less energy to move
588 than long straight chains, showing increased Newtonian behavior (Karkalas, Ma,
589 Morrison, & Pethrick, 1995).

590 The effect of the system's T_e-T_0 on the maize starch additives decreased, except
591 when waxy corn starch was used. The results show that the heat treatment increased
592 the temperature of the endothermic conversion and narrowed the temperature range of
593 the starch. Higher crystallinity and a more orderly crystalline structure increase the
594 temperature of endothermic conversion. However, the relative crystallinity of
595 heat-treated (annealed) starch does not increase; thus, the increase in the thermal

596 transition temperature and the decrease in the temperature range are attributed to an
597 increase in the crystallization order. The improvement of order is related to the
598 agglomeration of amylopectin caused by amylose recombination. The waxy corn
599 starch-coconut milk system exhibited the least variation due to the lack of amylose
600 molecules in the system. There was no significant difference in ΔH between the
601 waxy corn starch and common corn starch ($p > 0.05$) mainly because the starch
602 granules had no crystal or double helix structures. Therefore, the amylose composition
603 of the system has an important influence on the endothermic conversion and
604 temperature transformation range of the system.

605 3.7. Effects of maize starch additives on CM stability

606 Fig. 3 shows a typical Lumifuge diagram of the transmission light evolution
607 during the centrifugation of the coconut milk after high temperature sterilization. If
608 the oil droplet density is small and the sample solution is clarified, the light intensity
609 of the detection curve is strong. The light intensity on the right side of the X-axis is at
610 the bottom of the sample colorimetric dish, and the light intensity on the left side of
611 the X-axis is on the left side of the sample colorimetric dish. The initial transmission
612 distribution (red line) shows the movement of the particles, and the final transmission
613 distribution (green line at the top of the picture) represents the emulsion layer of the
614 oil droplets. Colloidal instability appears as emulsification or precipitation (Lerche &
615 Sobisch, 2011). After centrifugation, colloidal instability can be observed in the tube
616 and the transmission profile. The high-density red line in the profile indicates a slower
617 separation and higher stability, which can be observed in the emulsions with the

618 different amylose contents of maize starch (Fig. 3C, D, E). The profile of the pure
619 coconut milk and coconut milk with added maize kernels showed progressive
620 movement, rapid particle separation and a fat layer (Fig. 3A, B). According to the
621 transmission curve, compared with the coconut milk with the added starch, the
622 thickness of the emulsified layer in the pure coconut milk and the stable emulsion
623 with the added maize kernels **was** reduced. The larger gap between the red lines
624 represents slow emulsification. Compared with the starch coconut milk, the
625 clarification rate of the pure coconut milk and coconut milk with the added maize
626 kernels **was** higher. Gravity often affects the movement of liquid droplets, causing
627 precipitation and coagulation and resulting in a higher clarification rate (Piorkowski &
628 McClements, 2014). During the whole centrifugation process, the destruction of
629 condensed large liquid particles led to the destruction of the emulsion structure, but
630 the transmission distribution of all coconut milk samples clearly showed that there
631 was no phase separation phenomenon.

632 This study uses the creaming index (*CI*) (%/hr) to describe the rotational rate of
633 light intensity, and the *CI* is calculated by integrating the slope of the curve of
634 transmitted light over time (Mao, Boiteux, Roos, & Miao, 2014). As shown in Table 5,
635 the coconut milk without the maize starch additives had the highest *CI* value,
636 indicating that the system was the most unstable and a few layers existed. In the
637 coconut milk emulsion, due to the presence of phospholipids, proteins and other
638 components, even though the sample was homogenized, there were different phases;
639 thus, the emulsion state is unstable (Jiang, et al., 2016). The combination of the four

640 kinds of starch additives reduced the *CI* value of the mixed system; thus, maize starch
641 can effectively improve the stability of the system. Among the systems, the *CI* value
642 of the corn additive system slightly decreased, while the other three types of maize
643 starches significantly **reduced** the system *CI* value ($p < 0.05$). Banana starch is also
644 often used as an emulsion stabilizer through a hydrophilic interaction with the oil
645 phase (Bellopérez, Belloflores, Nuñezsantiago, Coronelaguilera, & Alvarezramirez,
646 2015). However, maize kernels also contain complex components, such as fats and
647 proteins. During the process of high pressure sterilization, the starch granules in maize
648 kernels swell and cause some mechanical damage to corn granules, leading to the
649 leaching of amylose **from** starch granules. The leached amylose can enhance the
650 viscosity and stability of the emulsion by forming a three-dimensional network in the
651 system (Lii, Tsai, & Tseng, 1996). Such cell disruption and further fragmentation
652 results in the release of cell wall components (e.g., starch and protein) into the
653 coconut milk, which can enhance particle-particle and particle-emulsion interactions
654 through Van der **Waals** forces, electrostatic forces, and/or water forces (Ped, et al.,
655 2013). The MS, with the added large particles, may be adsorbed onto the surface of
656 the oil droplet and form a strong network due to its flocculation. The network acts as a
657 mechanical barrier to prevent droplet coalescence, which is the result of stronger
658 network formation; thus, the emulsion remains stable. HAMS and networks with a
659 high viscosity move at slower speeds and inhibit extensive coalescence, resulting in
660 better emulsion stability. Proteins can improve the stability of the emulsion by
661 reducing the interfacial tension and forming a protective film around the fat

662 droplets. Polysaccharides can act as emulsion stabilizers by increasing the viscosity or
663 gel strength of the continuous phase and inducing flocculation of emulsion droplets
664 through bridging or consumption mechanisms depending on the adsorption properties
665 of polysaccharides (Eric, 2009). In accordance with Stokes' law, the apparent
666 viscosity and yield stress of coconut milk with maize kernels or amylose starch
667 increased, while the droplets in the coconut milk with the waxy corn starch probably
668 have a smaller particle size and specific surface area due to the more dependent
669 surfactants (more sodium caseinate and sucrose fatty acid esters), both of which
670 contribute to improving the stability of pure coconut milk.

671 4. Conclusion

672 The purpose of this study was to obtain an understanding of the effect of maize
673 starch with different amylose contents on the stability of coconut milk emulsions.
674 Maize grains and maize starch additives do not change the type of fluids in coconut
675 milk. Due to the higher the amylose content the greater the dynamic viscoelasticity.
676 The addition of maize grain can reduce the average particle size of the system, making
677 it difficult for the composite system to coagulate or precipitate. The addition of maize
678 kernels and starch resulted in a significant decrease in the white value of the coconut
679 milk system, and the ζ -potential and surface tension significantly increased
680 ($p < 0.05$). The high-amylose maize starch significantly improved the system's T_0 , T_e , T_p
681 and ΔH ($p < 0.05$) but shortened the temperature range of the system and enhanced
682 the thermal stability of the coconut milk. Maize kernels and maize starch increased the
683 viscosity of the emulsion, waxy maize starch improves the stability of coconut milk

684 emulsions by the action of the surfactant. The stability of the formed system of maize
685 kernels is similar to that of coconut milk formed by the addition of high-amylose
686 maize starch in terms of the fluid properties. The results obtained in this study can
687 provide a useful insight into the addition of maize or maize flour as stabilizers or
688 nutritional enhancers to foods containing emulsions, such as flavoring agents,
689 condiments and beverages.

690 **Acknowledgements**

691 This study was financially supported by the Research Fund for the China-Ireland
692 International Cooperation Centre for Food Material Science and Structure Design (No.
693 KXGH17001), Projects for Scientific and Technological Development of Fujian
694 Agriculture and Forestry University (KF2015099), the Program for Innovative
695 Research Team in Science and Technology in Fujian Province University (2012-03),
696 and the Scientific and Technological Innovation Team Support Plan of Fujian
697 Agriculture and Forestry University (cxt12009).

698 **Conflict of interest**

699 All co-authors declare no conflict of interest.

700 **Reference**

- 701 Ariyaprakai, S., & Tananuwong, K. (2015). Freeze–thaw stability of edible oil-in-water emulsions
702 stabilized by sucrose esters and Tweens. *Journal of Food Engineering*, 152, 57-64.
- 703 Barbieri, S. F., de Oliveira Petkowicz, C. L., de Godoy, R. C. B., de Azeredo, H. C. M., Franco, C.
704 R. C., & Silveira, J. L. M. (2018). Pulp and jam of gabirola (*Campomanesia xanthocarpa*
705 Berg): Characterization and rheological properties. *Food chemistry*, 263, 292-299.
- 706 Bellopérez, L. A., Belloflores, C. A., Nuñezsantiago, M. C., Coronelaguilera, C. P., &
707 Alvarezramirez, J. (2015). Effect of the degree of substitution of octenyl succinic

- 708 anhydride-banana starch on emulsion stability. *Carbohydrate Polymers*, 132(2), 17-24.
- 709 Bi, X., Hemar, Y., Balaban, M. O., & Liao, X. (2015). The effect of ultrasound on particle size,
710 color, viscosity and polyphenol oxidase activity of diluted avocado puree. *Ultrasonics*
711 *Sonochemistry*, 27, 567-575.
- 712 Das, K. P., & Kinsella, J. E. (1990). Stability Of Food Emulsions: Physicochemical Role Of
713 Protein And Nonprotein Emulsifiers. *Advances in Food & Nutrition Research*, 34,
714 81-201.
- 715 Eric, D. (2009). Hydrocolloids as emulsifiers and emulsion stabilizers. *Food Hydrocolloids*, 23(6),
716 1473-14829.
- 717 Evans, M., Ratcliffe, I., & Williams, P. A. (2013). Emulsion stabilisation using polysaccharide-
718 protein complexes. *Current Opinion in Colloid & Interface Science*, 18(4), 272-282.
- 719 Garcia, M. C., & Franco, C. M. L. (2015). Effect of glycerol monostearate on the gelatinization
720 behavior of maize starches with different amylose contents. *Starch - Stärke*, 67(1-2),
721 107-116.
- 722 Han, X. H., Campanella, O. H., Mix, N. C., & Hamaker, B. R. (2002). Consequence of Starch
723 Damage on Rheological Properties of Maize Starch Pastes. *Cereal Chemistry*, 79(6),
724 897-901.
- 725 Haralampu, S. G. (2000). Resistant starch - a review of the physical properties and biological
726 impact of RS3. *Carbohydrate Polymers*, 41(3), 285-292.
- 727 Iguttia, A. M., Pereira, A. C. I., Fabiano, L., Silva, R. A. F., & Ribeiro, E. P. (2011). Substitution
728 of ingredients by green coconut (*Cocos nucifera* L) pulp in ice cream formulation.
729 *Procedia Food Science*, 1, 1610-1617.
- 730 Ji, Z., Yu, L., Liu, H., Bao, X., Wang, Y., & Chen, L. (2017). Effect of pressure with shear stress
731 on gelatinization of starches with different amylose/amylopectin ratios. *Food*
732 *Hydrocolloids*, 72, 331-337.
- 733 Jiang, P., Xiang, D., & Wang, X. (2016). Effect of different treatment on the properties of coconut
734 milk emulsions. *Food Science and Technology Research*, 22(1), 83-89.
- 735 Jirapeangtong, K., Siriwatanayothin, S., & Chiewchan, N. (2008). Effects of coconut sugar and
736 stabilizing agents on stability and apparent viscosity of high-fat coconut milk. *Journal of*
737 *food engineering*, 87(3), 422-427.

- 738 Karkalas, J., Ma, S., Morrison, W. R., & Pethrick, R. A. (1995). Some factors determining the
739 thermal properties of amylose inclusion complexes with fatty acids. *Carbohydrate*
740 *Research*, 268(2), 233-247.
- 741 Lerche, D., & Sobisch, T. (2011). Direct and Accelerated Characterization of Formulation
742 Stability. *Journal of Dispersion Science & Technology*, 32(12), 1799-1811.
- 743 Li, C., Li, Y., Sun, P., & Yang, C. (2013). Pickering emulsions stabilized by native starch
744 granules. *Colloids & Surfaces A Physicochemical & Engineering Aspects*, 431(33),
745 142-149.
- 746 Lii, C. Y., Tsai, M. L., & Tseng, K. H. (1996). Effect of amylose content on the rheological
747 property of rice starch. *Cereal Chemistry*, 73(4), 415-420.
- 748 Lin, J. H., Kao, W. T., Tsai, Y. C., & Chang, Y. H. (2013). Effect of granular characteristics on
749 pasting properties of starch blends. *Carbohydrate Polymers*, 98(2), 1553-1560.
- 750 Lin, L., Guo, D., Zhao, L., Zhang, X., Wang, J., Zhang, F., & Wei, C. (2016). Comparative
751 structure of starches from high-amylose maize inbred lines and their hybrids. *Food*
752 *Hydrocolloids*, 52, 19-28.
- 753 Liu, F., & Tang, C. H. (2011). Cold, gel-like whey protein emulsions by microfluidisation
754 emulsification: Rheological properties and microstructures. *Food Chemistry*, 127(4),
755 1641-1647.
- 756 Long, Y., & Christie, G. (2005). Microstructure and mechanical properties of orientated
757 thermoplastic starches. *Journal of Materials Science*, 40(1), 111-116.
- 758 Long, Z., Zhao, Q., Liu, T., Kuang, W., Xu, J., & Zhao, M. (2012). Role and properties of guar
759 gum in sodium caseinate solution and sodium caseinate stabilized emulsion. *Food*
760 *Research International*, 49(1), 545-552.
- 761 Lu, X., Chen, J., Zheng, M., Guo, J., Qi, J., Chen, Y., Miao, S., & Zheng, B. (2019). Effect of
762 high-Intensity ultrasound irradiation on the stability and structural features of
763 coconut-grain milk composite systems utilizing maize kernels and starch with different
764 amylose contents. *Ultrasonics sonochemistry*, Doi: [https://doi.org/10.1016/j.ultsonch.](https://doi.org/10.1016/j.ultsonch.2019.03.003)
765 2019.03.003.
- 766 Lu, X., Wang, Y., Li, Y., & Huang, Q. (2018). Assembly of Pickering emulsions using milled
767 starch particles with different amylose/amylopectin ratios. *Food Hydrocolloids*, 84,

- 768 47-57.
- 769 Lundqvist, H., Eliasson, A. C., & Olofsson, G. (2002). Binding of hexadecyltrimethylammonium
770 bromide to starch polysaccharides. Part II. Calorimetric study. *Carbohydrate Polymers*,
771 49(2), 109-120.
- 772 Mao, L., Boiteux, L., Roos, Y. H., & Miao, S. (2014). Evaluation of volatile characteristics in
773 whey protein isolate–pectin mixed layer emulsions under different environmental
774 conditions. *Food Hydrocolloids*, 41(41), 79-85.
- 775 McClements, D. J. (2005). Protein-stabilized emulsions. *Current Opinion in Colloid & Interface*
776 *Science*, 9(5), 305-313.
- 777 Nattapol, T., & Johnn, C. (2009). Effect of thermal treatments on the properties of coconut milk
778 emulsions prepared with surface-active stabilizers. *Food Hydrocolloids*, 23(7),
779 1792-1800.
- 780 Ng, C. Y., Mohammad, A. W., Ng, L. Y., & Jahim, J. M. (2014). Membrane fouling mechanisms
781 during ultrafiltration of skimmed coconut milk. *Journal of Food Engineering*, 142(6),
782 190-200.
- 783 Ng, S. P., Lai, O. M., Abas, F., Hong, K. L., & Tan, C. P. (2014). Stability of a concentrated
784 oil-in-water emulsion model prepared using palm olein-based diacylglycerol/virgin
785 coconut oil blends: Effects of the rheological properties, droplet size distribution and
786 microstructure. *Food Research International*, 64, 919-930.
- 787 Nitschke, M., Costa, S. G. V. A. O., Haddad, R., Gonçalves, L. A. G., Eberlin, M. N., & Contiero,
788 J. (2005). Oil wastes as unconventional substrates for rhamnolipid biosurfactant
789 production by *Pseudomonas aeruginosa* LBI. *Biotechnology Progress*, 21(5), 1562-1566.
- 790 Ped, A., Ibarz, A., & Cristianini, M. (2013). Effect of high pressure homogenization (HPH) on the
791 rheological properties of tomato juice: Viscoelastic properties and the Cox–Merz rule.
792 *Food Research International*, 54(1), 169-176.
- 793 Phungamngoen, C., Asawajinda, T., Santad, R., & Sawedboworn, W. (2016). Feasibility study of
794 aseptic homogenization: Affecting homogenization steps on quality of sterilized coconut
795 milk. In *MATEC Web of Conferences* (Vol. 62, pp. 02010): Food Sciences.
- 796 Piorkowski, D. T., & McClements, D. J. (2014). Beverage emulsions: Recent developments in
797 formulation, production, and applications. *Food Hydrocolloids*, 42(42), 5-41.

- 798 Raghavendra, S. N., & Ksms, R. (2010). Effect of different treatments for the destabilization of
799 coconut milk emulsion. *Journal of Food Engineering*, *97*(3), 341-347.
- 800 Rochavillarreal, V., Hoffmann, J. F., Vanier, N. L., Sernasaldivar, S. O., & Garcíalara, S. (2018).
801 Hydrothermal treatment of maize: Changes in physical, chemical, and functional
802 properties. *Food Chemistry*, *263*, 225-231.
- 803 Rodsamran, P., & Sothornvit, R. (2018). Physicochemical and functional properties of protein
804 concentrate from by-product of coconut processing. *Food Chemistry*, *241*, 364-371.
- 805 Schröder, A., Berton-Carabin, C., Venema, P., & Cornacchia, L. (2017). Interfacial properties of
806 whey protein and whey protein hydrolysates and their influence on O/W emulsion
807 stability. *Food Hydrocolloids*, *73*, 129-140.
- 808 Shao, Y. Y., Tseng, Y. H., Chang, Y. H., Lin, J. H., & Lii, C. Y. (2007). Rheological properties of
809 rice amylose gels and their relationships to the structures of amylose and its subfractions.
810 *Food Chemistry*, *103*(4), 1324-1329.
- 811 Takahashi, T., & Fujita, N. (2017). Thermal and rheological characteristics of mutant rice starches
812 with widespread variation of amylose content and amylopectin structure. *Food*
813 *Hydrocolloids*, *62*, 83-93.
- 814 Tanner, R. I., & Rivlin, R. S. (1985). *Engineering Rheology*: Clarendon Press.
- 815 Vanin, F. M., Michon, C., & Lucas, T. (2013). Effect of the drying rate on the complex viscosity
816 of wheat flour dough transforming into crust and crumb during baking. *Journal of Cereal*
817 *Science*, *58*(2), 290-297.
- 818 Wang, B., Wang, L. J., Li, D., Adhikari, B., & Shi, J. (2011). Effect of gum Arabic on stability of
819 oil-in-water emulsion stabilized by flaxseed and soybean protein. *Carbohydrate*
820 *Polymers*, *86*(1), 343-351.
- 821 Wang, J., Xia, N., Yang, X., Yin, S., Qi, J., He, X., Yuan, D., & Wang, L. (2012). Adsorption and
822 dilatational rheology of heat-treated soy protein at the oil–water interface: Relationship to
823 structural properties. *Journal of agricultural and food chemistry*, *60*(12), 3302-3310.
- 824 Watanabe, T., Kawai, T., & Nonomura, Y. (2017). Effects of Fatty Acid Addition to Oil-in-water
825 Emulsions Stabilized with Sucrose Fatty Acid Ester. *Journal of Oleo Science*, *67*(3),
826 307-313.
- 827 Xie, F. W., Long, Y., Bing, S., Peng, L., Wang, J., Liu, H. S., & Ling, C. (2009). Rheological

- 828 properties of starches with different amylose/amylopectin ratios. *Journal of Cereal*
829 *Science*, 49(3), 371-377.
- 830 Xu, C., Du, X., Chen, P., Li, G., Yang, X., & Zhou, X. (2017). Morphologies and gelatinization
831 behaviours of high-amylose maize starches during heat treatment. *Carbohydrate*
832 *Polymers*, 157, 637-642.
- 833 Yu, S. F., Jing, X., Zhang, Y. C., & Kopparapu, N. K. (2014). Relationship between intrinsic
834 viscosity, thermal, and retrogradation properties of amylose and amylopectin. *Czech*
835 *Journal of Food Sciences*, 32(5), 514-520.
- 836 Yusoff, A., & Murray, B. S. (2011). Modified starch granules as particle-stabilizers of oil-in-water
837 emulsions. *Food Hydrocolloids*, 25(1), 42-55.
- 838 Zhou, W., Yang, J., Yan, H., Liu, G., Zheng, J., Gu, Z., & Zhang, P. (2015). Impact of amylose
839 content on starch physicochemical properties in transgenic sweet potato. *Carbohydrate*
840 *Polymers*, 122, 417-427.
- 841

Figure captions

Figure 1. Rheology properties of coconut milk. (A) Influence of the shear rate on the apparent viscosity curve (B) and flow curve. (C) Strain dependence of the storage modulus G' (D), loss modulus G'' (E) and loss tangent ($\tan\delta$) as a function of frequency.

Figure 2. Particle size distribution and particle diameters of coconut milk supplemented with maize kernels and starch.

Figure 3. Evolution of transmission light signals of coconut milk measured by a Lumifuge. The bottom red line represents the first scanning profile, and the top green line represents the final scanning profile. A: CM; B: MK; C: MS; D: HAMS; E: WCS.

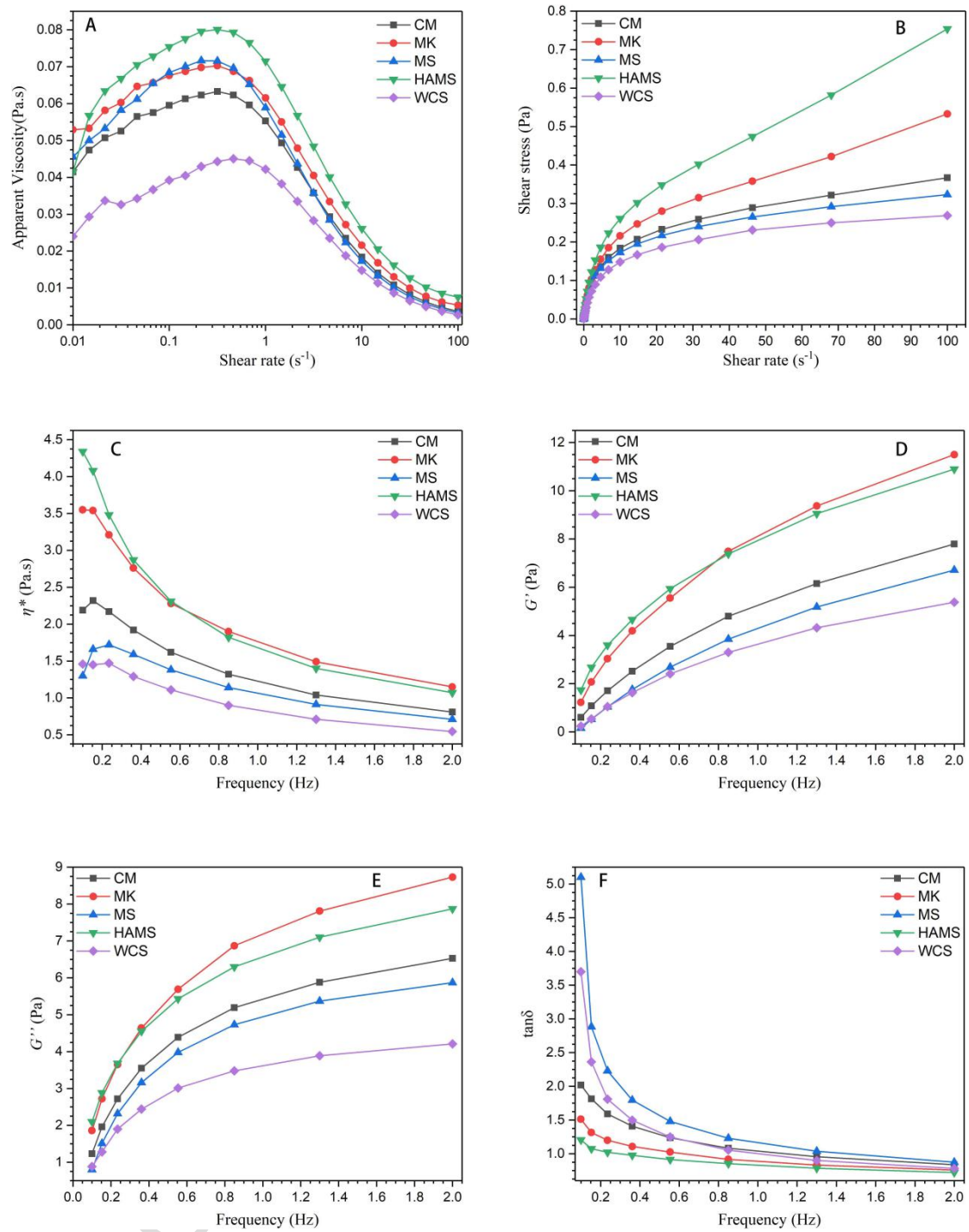


Fig.1

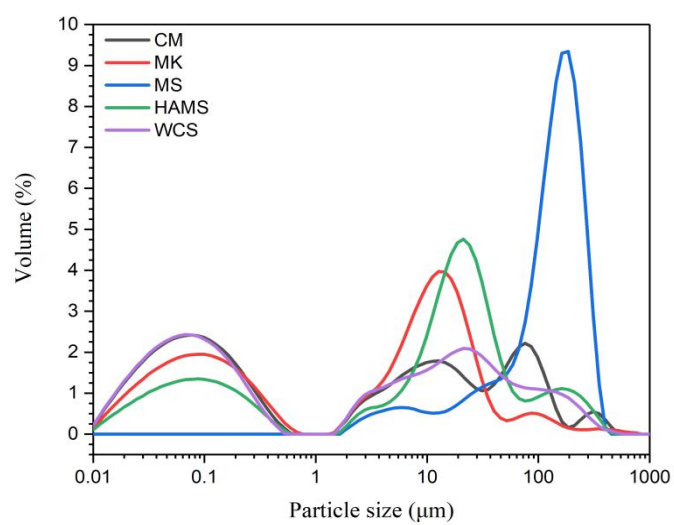


Fig.2

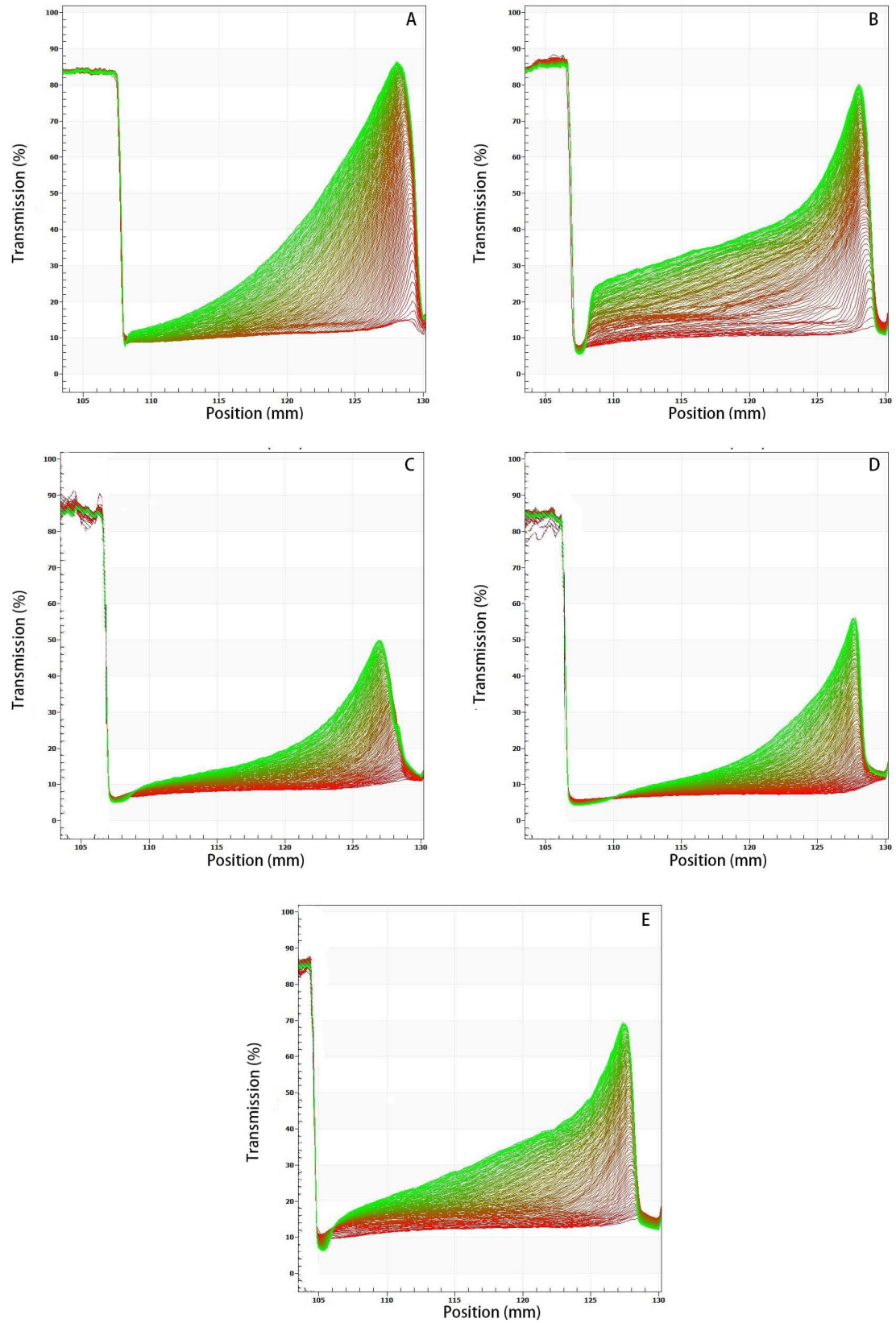


Fig.3

Highlights

- The amylose content increased the viscosity and yield stress of coconut milk.
- The coconut milk fluid of the maize kernel is similar to high-amylose maize starch.
- The addition of corn improves the stability of coconut milk.

Table 1 Nutrient composition of maize starch used in this study (%)

Sample	Amylose ^a	Water ^b	Lipid ^c	Protein ^d	Ash ^e
maize kernels	25.63%±0.26	13.49%±0.40	4.43%±0.03	8.76%±0.17	1.45%±0.06
maize starch	26.89%±0.04	9.46%±0.37	0.04%±0.03	0.21%±0.05	0.17%±0.02
high-amylose maize starch	89.06%±0.18	11.29%±0.35	0.13%±0.01	0.39%±0.32	0.08%±0.02
waxy corn starch	0.86%±0.14	10.03%±0.27	0.19%±0.04	0.15%±0.03	0.04%±0.01

Test methods: ^a amylose / amylopectin assay kit (Megazyme Int, Ireland), ^b AACC International Method 44-15.02 (1999), ^c AACC International Method 30-10.01 (1999), ^d AACC International Method 46-16.01 (1999), ^e AACC International Method 08-17.01 (1999).

Table 2 Formulation of composite stabilizers and additives under optimal stability in each group of coconut milk

Sample	Carrageenan (g/L)	Distearin (g/L)	Sodium caseinate (g/L)	Sucrose fatty acid ester (g/L)	Glyceryl monostearate (g/L)	Corn kernels and starch content (g/L)	Creaming rate (mm/d)
CM	0.5	0.5	1	1.5	10	0	0.0184
MK	0.5	0.7	0.8	1.1	10	100	0.0142
MS	0.3	0.7	1.2	1.5	10	3.956	0.0079
HAMS	0.3	0.8	1	1.3	12	3.956	0.0083
WCS	0.3	0.5	1.6	1.7	10	3.956	0.0133

Note: The velocities of the separation of individual particles (mm/d) are used as an indicator; the slower the particle separation rate is, the more stable the system.

Table 3 Modeling of the flow curve between 0.01 and 100 s⁻¹ of the shear rate of the coconut milk containing maize kernels and starch using the Herschel-Bulkley model

Sample	τ_0/Pa	$K/(\text{Pa}\cdot\text{S}^n)$	n	R^2
CM	-0.0448	0.0951	0.2900	0.9963
MK	-0.0301	0.1184	0.3700	0.9973
MS	-0.0500	0.1104	0.2600	0.9960
HAMS	-0.0228	0.1219	0.3500	0.9984
WCS	-0.0421	0.0987	0.2700	0.9935

Table 4 D[4,3], D[3,2] and the specific surface area of coconut milk supplemented with maize kernels and starch; D[3,2]: surface-area-based mean diameter; D[4,3]: volume-based mean diameter

Sample	D[4,3] (μm)	D[3,2] (μm)	Specific surface area (m^2/kg)
CM	28.64 \pm 1.1 ^a	0.11 \pm 0.00 ^a	35730 \pm 1589.12 ^a
MK	16.12 \pm 0.32 ^b	0.13 \pm 0.01 ^b	28000 \pm 1242.51 ^b
MS	150.35 \pm 6.71 ^c	40.97 \pm 0.32 ^c	91.63 \pm 1.00 ^c
HAMS	35.71 \pm 1.52 ^d	0.19 \pm 0.01 ^d	19370 \pm 561.15 ^d
WCS	27.44 \pm 0.91 ^a	0.11 \pm 0.01 ^a	36160 \pm 798.89 ^a

^{abc} Means in the same column without the same letter are significantly different ($p < 0.05$) according to a Duncan multiple range test.

Table 5 Changes in the color, surface tension, zeta-potential and creaming index of coconut milk stabilized by the five tested samples (CM, MK, MS, HAMS, and WCS)

Sample	L^*	a^*	b^*	ΔE	Zeta potential (mv)	Surface tension (mN/m)	Creaming index (%/h)
CM	78.12±0.81 ^a	-2.98±0.02 ^a	-2.89±0.01 ^a	0 ^a	-25.54±2.12 ^a	19.18±0.34 ^a	4.12±0.23 ^a
MK	72.73±0.19 ^b	-3.14±0.05 ^b	-3.29±0.03 ^b	5.4±0.38 ^b	-7.93±0.34 ^b	20.58±0.45 ^b	3.69±0.11 ^b
MS	74.48±0.91 ^c	-4.49±0.06 ^c	-5.35±0.07 ^c	4.64±0.33 ^c	1.38±0.57 ^c	20.46±0.51 ^b	2.47±0.04 ^c
HAMS	75.87±0.48 ^c	-3.32±0.03 ^d	-3.88±0.06 ^d	2.46±0.16 ^d	-0.7±0.35 ^d	20.58±0.38 ^b	2.94±0.09 ^d
WCS	74.76±1.01 ^c	-5.32±0.03 ^e	-6.25±0.01 ^e	5.29±0.29 ^b	1.39±0.99 ^c	20.12±0.21 ^b	2.76±0.06 ^e

^{abc} Means in the same column without the same letter are significantly different ($p < 0.05$) according to a Duncan multiple range test.

Table 6 DSC scans of coconut milk stabilized by the five tested samples (CM, MK, MS, HAMS, and WCS)

Sample	T_0 Initial gelatinization temperature /°C	T_p Peak of gelatinization temperature /°C	T_e Termination of gelatinization temperature /°C	ΔH Thermal enthalpy /J·g ⁻¹
CM	58.23±0.23 ^a	65.10±0.31 ^a	71.91±0.24 ^a	44.23±0.61 ^a
MK	64.99±0.17 ^b	70.60±0.29 ^b	76.88±0.16 ^b	50.79±0.63 ^b
MS	60.96±0.21 ^c	64.90±0.22 ^a	73.78±0.26 ^c	47.73±0.47 ^c
HAMS	67.21±0.45 ^d	71.9±0.31 ^c	79.12±0.36 ^d	51.03±0.24 ^b
WCS	60.13±0.19 ^c	63.70±0.34 ^d	76.21±0.26 ^b	45.88±0.58 ^d

^{abc} Means in the same column without the same letter are significantly different ($p < 0.05$) according to a Duncan multiple range test.