

Angiogenesis and pro-angiogenic factors in uterine fibroids — facts and myths

MONIKA KONARSKA¹, ANNA NATALIA WRONA², VERONIKA ALEKSANDROVYCH³,
TOMASZ BEREZA¹, MAREK SAJEWICZ⁴, BARBARA GACH-KUNIEWICZ⁵, MACIEJ LIS⁴,
KINGA KOMNATA¹, MATEUSZ PAZIEWSKI¹, ALEKSANDRA MALESZKA⁶, PAWEŁ DEPUKAT¹,
BERNARD SOLEWSKI¹, ŁUKASZ WARCHOŁ¹

¹Department of Anatomy, Jagiellonian University Medical College
ul. Kopernika 12, 31-034 Kraków, Poland

²Szpital Specjalistyczny im. J. Śniadeckiego w Nowym Sączu, Oddział Ginekologiczno-Położniczy
ul. Młyńska 10, 33-300 Nowy Sącz, Poland

³Department of Pathophysiology, Jagiellonian University Medical College
ul. Czysza 18, 31-121 Kraków, Poland

⁴Clinic of Obstetrics and Perinatology, The University Hospital
ul. Kopernika 23, 31-501 Kraków, Poland

⁵Szpital Specjalistyczny im. L. Rydygiera w Krakowie, Oddział Anestezjologii i Intensywnej Terapii
os. Złotej Jesieni 1, 31-826 Kraków, Poland

⁶Department of Diagnostics, Jagiellonian University Medical College
ul. Kopernika 15A, 31-501 Kraków, Poland

Corresponding author: Tomasz Bereza, MD, PhD; Department of Anatomy
Jagiellonian University Medical College
ul. Kopernika 12, 31-034 Kraków, Poland
Phone/Fax: +48 12 422 95 11; E-mail: t.bereza@wp.pl

Abstract: Uterine leiomyomata present major problem for females. Although they are benign tumors their frequency is associated with many symptoms like infertility, abdominal pain, menorrhagia. Authors based on their own morphological studies and review of the literature try to indicate main factors causing angiogenesis within leiomyomata and its influence on tumor growth. The strongest proangiogenic factor seems to be hypoxia, which stimulates up- and down-regulation of numerous genetically determined substances. Also mechanical pressure acting upon newly growing vessels is one of the factors which may determine formation of so called “vascular pseudocapsule” around the lesion.

Key words: uterine leiomyomata, microvessel density, angiogenesis.

Introduction

Angiogenesis is currently considered to be a very important factor controlling growth and ability to metastasize of malignant tumors [1]. Role of the angiogenesis in the growth of benign tumors still remains unclear. However if we consider the model of development of uterine fibroids based on adaptation of the vessels of normal vascular bed — their further regression and subsequent vascularization “de novo” — it may appear that angiogenesis plays key role also in the development of benign tumors. Formation of “vascular pseudocapsule” in the periphery of the fibroid requires undoubtedly involvement of pro-angiogenic factors. Although studies which consider angiogenic activity of uterine leiomyomata and surrounding myometrium, bring unanimous results by far.

It was proved that uterine fibroids are source of the whole palette of growth factors, which seem to be probable or to be angiogenic factors, too. Family of factors associated with uterine leiomyomata consists of: vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), platelet derived endothelial cells growth factor (PD-ECGF), epidermal growth factor (EGF), transforming growth factor (TGF alpha and beta), basic fibroblast growth factor (bFGF), insulin-like growth factor (IGF), adnenuomedullin (ADM) [2–12]. Some of mentioned factors, i.e. VEGF-A [6], bFGF [3, 11] and IGF-I [7] show increased expression in the uterine leiomyomata, comparing to the surrounding myometrium. Other factors, i.e. EGF [6], TGF-beta [13] show decreased expression in fibroids. It is generally assumed that VEGF and ADM show increased activity in uterine myomas [2]. From another hand however we have to cope with contrary reports. Lee and Novak described increased level of TGF-beta mRNA [10], while Vollenhoven *et al.* [14] did not show significant changes in expression of EGF and TGF-beta of fibroids and surrounding myometrium. Similar results were achieved by Harrisson-Woolrych *et al.* in EGF mRNA [9].

Material and methods

The study was carried out on 51 human uteri of females aged between 32–66 years, which died because of diseases not associated with the internal female genital tracts, obtained during autopsies. The study was approved by local Ethical Committee. The whole material was collected upon 12–22 hrs from the moment of decease. The protocol used was described in details in [14, 15]. Additionally 5 specimens were injected with water solution of acrylic emulsion (Liquitex R, Binney and Smith, USA) [16]. Thus obtained material was cut into slides, which after dehydration in rising concentrations of ethyl alcohol and immersed in methyl benzoese were paraffinized and dyed with hematoxylin and eosin. The material was studied under light microscope (magnification 5–40x). Some of tissue blocs were deparaffinized and hydrated to perform immunohistochemical reaction for von Willebrandt factor.

Results and discussion

Most of proangiogenic factors are located in cytoplasm of smooth muscle cells — of both myometrium and cells of blood vessels. TGF- α , EGF and FGF-2 were observed also within endothelial cells, and IGF-1 in the fibroblasts of connective tissue [6]. It is interesting that bFGF is located mainly within intercellular matrix, and its relatively high level was associated with its condensation [11].

Growth factors present in fibroids may influence all its factors: smooth muscle cells, vascular endothelial cells and connective tissue fibroblasts. Their specific participation in angiogenesis hasn't been sufficiently elicited yet. It is interesting that so far any relationship between increased vascular density and level of VEGF expression hasn't been proved, and VEGF is considered to be the main factor which stimulates angiogenesis [17, 18]. That kind of positive correlation was observed according to PD-ECGF, TGF- α and ADM. Expression of this last factor correlates positively also with the endothelial cells proliferation index [2]. ADM, opposite to VEGF has limited mitogenic action, limited almost to endothelial cells only. Furthermore, it shows relatively wide influence on proliferation and growth of tumor cells, both malignant and benign, and may play key role in angiogenesis of uterine leiomyomata.

Regulation of angiogenic factors probably follows few complicated mechanisms. Genetic aberrations of chromosomes 6, 7, 12, and 14 may cause increased tolerance considering angiogenic growth factors or decreased ability of production or response to angiogenic factors with the growth of the tumor. Genetic failures leading to increased sensitivity to angiogenic inhibitors, i.e. angiostatin [19], may be responsible for clinical picture of necrosis within certain leiomyomata.

Factors which stimulate angiogenesis, i.e. VEGF and substances from group of FGF, prove frequently increased activity in hypoxia, tissue damage and growth of new tissues. ADM and VEGF prove certain similarities, because both these strong angiogenic factors are induced by hypoxia, and both increase vascular permeability. It has been well documented that on induction both these factors require hypoxia stadium [2]. It is postulated that similar to most of the neoplasms, uterine leiomyomata show decreased level of oxygen.

In our studies one could observe relatively dense network of vessels penetrating the lesions, and in the specimens injected with Liquitex R we could see compressed blood vessels — even not filled with dye which were visible by staining using von Willebrandt factor. These compressed by growing tumor masses vessels were useless — and such condition led to significant hypoxia, a strong proangiogenic condition.

It is obvious that angiogenesis in the uterine fibroids may be influenced by ovarian steroids. Estrogens regulate expression of VEGF in the uterus at the transcription level [20]. In physiology they promote endometrial angiogenesis through regulation of VEGF expression in glandular cells and stroma [21]. Regulation of bFGF and its

receptors seems to be also dependent on the level of ovarian hormones [12]. PDGF and bFGF levels decrease in patients treated with agonists of gonadotropin releasing hormone (GnRH) [22, 23]. From another hand however in these patients nobody observed specific changes of microvessel density nor VEGF expression in fibroids [8, 24]. However the endothelial cells proliferation index of fibroids, myometrium and endometrium did not differ in particular phases of the menstruation cycle [2]. Xu *et al.* postulated that progesterone receptor modulator CDB-2914 down-regulates vascular endothelial growth factor, adrenomedullin and their receptors and modulates progesterone receptor content in cultured human uterine leiomyoma cells [25]. Also Catherino *et al* [26] indicate new potential targets for genetic researches.

In this context differences in vascular density observed between leiomyomata and the normal tissue may at least partially be result of different level of estrogen receptors (ER), which was shown in fibroids, comparing to the level of ER in regular myometrium [25, 27, 28]. The level of ER alpha RNA and immunoreactivity ER alpha are higher in leiomyomata than in surrounding myometrial tissue. Myometrial cells and in vitro cultured fibroid cells show increased expression of ER alpha receptors, and do not show changes in expression of ER beta. On the other hand endothelium of microvessels shows significant changes in expression of ER beta receptors [29]. Estrogens play vital role in survival of endothelial cells [30], so on this path they can stimulate changes which support or/and cause angiogenic response of microvessel endothelial cells, thus promoting growth of the new vessels within a tumor.

Despite expression of numerous proangiogenic factors within uterine leiomyomata, angiogenesis seems not to be specially outstanding during tumorigenesis, what is confirmed by relatively low vascular density (Fig. 1).

This hypothesis was lastly confirmed by studies of Weston *et al.*, which proved in the leiomyomata decreased expression of two factors promoting angiogenesis: connective tissue growth factor (CTGF) and cysteine rich factor inducing angiogenesis (CYR61), and also decreased expression of collagen 4 alpha2 (COL4A2), precursor of angiogenic inhibitor — canstatine. This is why fibroids comparing to the surrounding myometrium show antiangiogenic profile of gene expression.

For contrast, the most intense angiogenesis seems to occur at the border leiomyoma/myometrium, which leads to formation of vascular rich “pseudocapsule”, visible especially in greater uterine leiomyomata. It might be a result of release of proangiogenic factors through stimulated by fibroid angiogenesis within surrounding myometrium. As a matter of fact we do not know, if the arrangement of the vessels in the central region of the lesion is resulted by physical pressure of the new vessels ‘invading’ the tumor or is it result of activity of the factors which inhibit angiogenesis in these parts of the tumor? This problem remains in the sphere of speculations, by far.

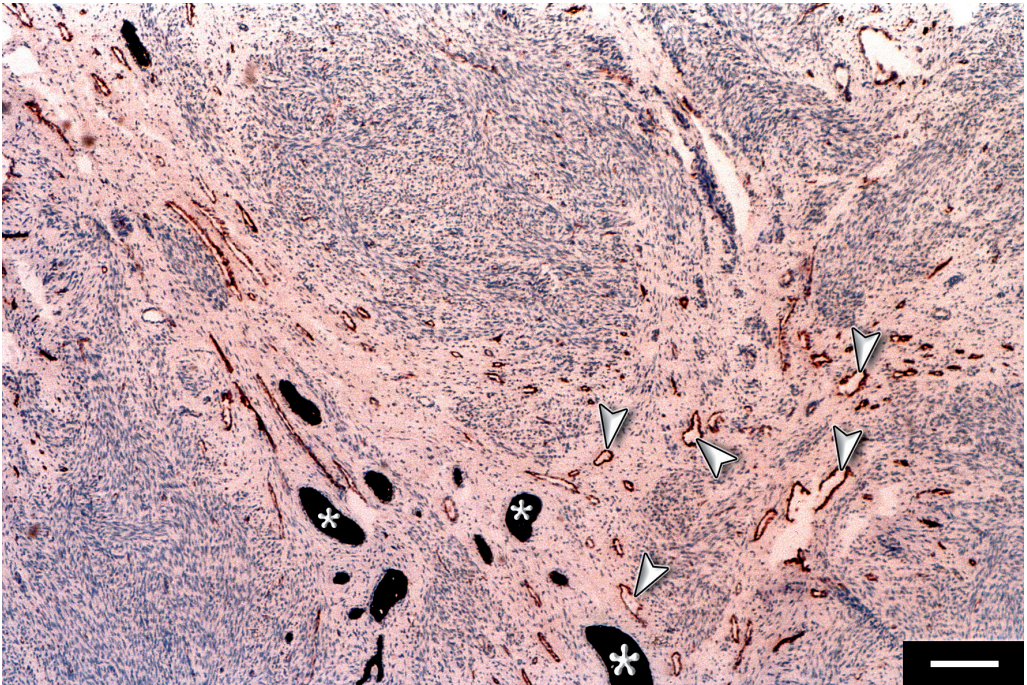


Fig. 1. Uterus of 43 years old female. Subserous leiomyoma injected through the arteries with water solution of Acrylic emulsion Liquitex R (Binney and Smith) (*). Immunostaining for von Willebrandt factor. Centrally visible numerous minute, not filled with emulsion vessels (arterioles and capillaries) (↑). Bar = 500 μ m.

Conflict of interest

None declared.

References

1. Folkman J.: Role of angiogenesis in tumor growth and metastasis. *Semin Oncol.* 2002; 29, Suppl. 16: 15–18.
2. Hague S., Zhang L., Oehler M.K., Manek S., MacKenzie I.Z., Bicknell R., Rees M.C.P.: Expression of hypoxically regulated angiogenic factor adrenomedullin correlates with uterine leiomyoma vascular density. *Clin Cancer Res.* 2000; 6: 2808–2814.
3. Anania C.A., Stewart E.A., Quade B.J., Hill J.A., Nowak R.A.: Expression of the fibroblast growth factor receptor in women with leiomyomas and abnormal uterine bleeding. *Mol Hum Reprod.* 1997; 3: 685–691.
4. Anderson J.: Factors in fibroid growth. *Baillière's Clin Obstet Gynecol.* 1998; 12: 225–243.
5. Arici A., Sozen I.: Transforming growth factor-beta3 is expressed at high levels in leiomyoma where it stimulates fibronectin expression and cell proliferation. *Fertil Steril.* 2000; 73: 1006–1011.

6. Dixon D., He H., Haseman J.K.: Immunohistochemical localization of growth factors and their receptors in uterine leiomyomas and matched myometrium. *Environ Health Perspect.* 2000; 108, Suppl. 5: 797–802.
7. Englund K., Lindblom B., Carlstrom K., Gustavsson I., Sjoblom P., Blanck A.: Gene expression and tissue concentrations of IGF-I in human myometrium and fibroids under different hormonal conditions. *Mol Hum Reprod.* 2000; 6: 915–920.
8. Gentry C.C., Okolo S.O., Fong L.W.F.T., Crow J.C., Maclean A.B., Perrett C.W.: Quantification of vascular endothelial growth factor—A in leiomyomas and adjacent myometrium. *Clin Sci.* 2001; 101: 691–695.
9. Harrison-Woolrych M.L., Charnock-Jones D.S., Smith S.K.: Quantification of messenger ribonucleic acid for epidermal growth factor in human myometrium and leiomyomata. *J Clin Endocrinol Metab.* 1995; 80: 1853–1858.
10. Lee B.S., Nowak R.A.: Human leiomyoma smooth muscle cells show increased expression of transforming growth factor-beta 3 (TGF beta 3) and altered responses to the antiproliferative effects of TGF beta. *J Clin Endocrinol Metab.* 2001; 86: 913–920.
11. Mangrulkar R.S., Ono M., Ishikawa M., Takashima S., Klagsbrun M., Nowak R.A.: Isolation and characterization of heparin-binding growth factors in human leiomyomas and normal myometrium. *Biol Reprod.* 1995; 53: 636–646.
12. Wu X., Blanck A., Olovsson M., Moller B., Lindblom B.: Expression of basic fibroblast growth factor (bFGF), FGF receptor 1 and FGF receptor 2 in uterine leiomyomas and myometrium during the menstrual cycle, after menopause and GnRHa treatment. *Acta Obstet Gynecol Scand.* 2001; 80: 497–504.
13. Hong T., Shimada Y., Uchida S., Itami A., Li Z., Ding Y., Kaganoi J., Komoto I., Sakurai T., Imamura M.: Expression of angiogenic factors and apoptotic factors in leiomyosarcoma and leiomyoma. *Int J Mol Med.* 2001; 8: 141–148.
14. Vollenhoven B.J., Pearce P., Herington A.C., Healy D.L.: Messenger ribonuclein acid expression of the insulin-like growth factors and their binding proteins in uterine fibroids and myometrium. *J Clin Endocrinol Metab.* 1995; 76: 1106–1110.
15. Walocha J.A., Miodoński A.J., Szczepański W., Skrzat J., Stachura J.: Two types of vascularisation of intramural uterine leiomyomata revealed by corrosion casting and immunohistochemical study. *Folia Morphol. (Warsz.)* 2004; 63, 1: 37–41.
16. Walocha J.A., Miodoński A.J., Nowogrodzka-Zagórska M., Kuciel R., Gorczyca J.: Application of a mixture of glycol polyethylenes for the preparation of microcorrosion casts — an observation. *Folia Morphol. (Warsz.)* 2002; 61, 4: 313–316.
17. Walocha J.A., Szczepański W., Miodoński A.J., Gorczyca J., Skrzat J., Bereza T., Ceranowicz P., Lorkowski J., Stachura J.: Application of acrylic emulsion Liquitex R (Binney and Smith) for the preparation of injection specimens and immunohistochemical studies—an observation. *Folia Morphol.* 2003; 62 (2), 157–161.
18. Poncelet C., Fauvet R., Feldmann G., Walker F., Madelenat P., Darai E.: Prognostic value of von Willebrand factor, CD34, CD31, and vascular endothelial growth factor expression in women with uterine leiomyosarcomas. *J Surg Oncol.* 2004; 86: 84–90.
19. O'Reilly M.S., Boehm T., Shing Y., Fukai N., Vasios G., Lane W.S., Flynn E., Birkhead R., Olsen B.R., Folkman J.: Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis Lung Carcinoma. *Cell.* 1994; 79: 315–328.
20. Hyder S.M., Huang J.C., Nawaz Z., Boettger-Tong H., Makela S., Chiapetta C., Stancel G.M.: Regulation of Vascular Endothelial Growth Factor Expression by Estrogens and Progestins. *Environm. Health Perspect.* 2000, 108: 785–790.
21. Albrecht E.D., Babischkin J.S., Lidor Y., Anderson L.D., Udoff L.C., Pepe G.J.: Effect of estrogen on angiogenesis in co-cultures of human endometrial cells and microvascular endothelial cells. *Hum Reprod.* 2003; 18, 10: 2039–2047.

22. Di Lieto A., De Rosa G., De Falco M., Ianotti F., Staibano S., Pollio F., Scaramellino M., Salvatore G.: Relationship between platelet-derived growth factor expression in leiomyomas and uterine volume changes after gonadotropin-releasing hormone agonist treatment. *Hum Pathol.* 2002; 33: 220–224.
23. Di Lieto A., De Falco M., Staibano S., Ianotti F., Scaramellino M., Mansueto G., Granata P., Pontillo M., Pollio F., De Rosa G.: Effects of gonadotropin-releasing hormone agonists on uteri volume and vasculature and on the immunohistochemical expression of basic fibroblast growth factor (bFGF) in uterine leiomyomas. *Int J Gynecol Pathol.* 2003; 22: 353–358.
24. Abulafia O., Kleinhaus K., Levi G., Lee Y.C., Sherer D.M.: Effect of gonadotropin-releasing hormone agonist treatment upon angiogenesis in uterine leiomyoma. *Gynecol Obstet Invest.* 2001; 52: 108–113.
25. Xu Q., Ohara N., Chen W., Liu J., Sasaki H., Morikawa A., Sitruk-Ware R., Johansson E.D.B., Maruo T.: Progesterone receptor modulator CDB-2914 down-regulates vascular endothelial growth factor, adrenomedullin and their receptors and modulates progesterone receptor content in cultured human uterine leiomyoma cells. *Hum Rep.* 2006; 21 (9): 2408–2416. doi: 10.1093/humrep/del159.
26. Catherino W., Salama A., Potlog-Nahari C., Leppert P., Tsibris J., Segars J.: Gene expression studies in leiomyomata: New directions for research. *Semin Rep Med.* 2004; 22 (2): 83–90.
27. Benassayag C., Leroy M.J., Rigourd V., Robert B., Honore J.C., Mignot T.M., Vacher-Lavenu M.C., Chapron C., Ferre F.: Estrogen receptors (ER α /ER β) in normal and pathological growth of the human myometrium: pregnancy and leiomyoma. *Am J Physiol.* 1999; 39: E1112–1118.
28. Wang H., Wu X., Englund K., Masironi B., Eriksson H., Sahlin L.: Different expression of estrogen receptors alpha and beta in human myometrium and leiomyoma during the proliferative phase of the menstrual cycle and after GnRHa treatment. *Gynecol Endocrinol.* 2001; 15: 443–452.
29. Gargett C.E., Bucak K., Zaitseva M., Chu S., Taylor N., Fuller P.J., Rogers P.A.W.: Estrogen receptors- α and - β expression in microvascular endothelial cells and smooth muscle cells of myometrium and leiomyoma. *Mol Hum Reprod.* 2002; 8, 8: 770–775.
30. Razandi M., Pedram A., Levin E.R.: Estrogen signals to the preservation of endothelial cell form and function. *J Biol Chem.* 2000; 275: 38540–38546.
31. Weston G., Trajstman A.C., Gargett C.E., Manuelpillai U., Vollenhoven B.J., Rogers P.A.W.: Fibroids display an anti-angiogenic gene expression profile compared with adjacent myometrium. *Mol Hum Reprod.* 2003; 9: 541–549.