Serum anti-glycan antibodies in paediatric-onset Crohn's disease: association with disease phenotype and diagnostic accuracy

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Abstract

Introduction: Antibodies reacting with various microbial epitopes have been described in inflammatory bowel disease (IBD) and are associated with a specific diagnosis and clinical presentation.

Aim: To evaluate the profile of new anti-glycan antibodies, their potential association with disease phenotype and diagnostic accuracy in paediatric Crohn's disease (CD).

Material and methods: Blood samples from 134 paediatric IBD patients (109 CD, 25 ulcerative colitis (UC)) and 67 controls were blindly analysed for *anti-Saccharomyces cerevisiae* (ASCA), anti-chitobioside carbohydrate (ACCA), anti-laminaribioside carbohydrate (ALCA), and anti-mannobioside carbohydrate (AMCA) antibodies using commercially available assays. The serological response to glycans was correlated with clinical disease characteristics.

Results: At least one of the tested anti-glycan antibodies was present in 75% of CD patients. Despite the high frequency of reactivity to glycan epitopes, a limited overlap of serological markers was observed. In total, 49% of ASCA-negative patients presented with one of the following: ACCA, ALCA, or AMCA. The occurrence of one antibody from the anti-glycan panel was independently associated with complicated disease phenotype and ileocolonic disease location. A higher level of immune response as assessed by the quartile sum scores for ACCA, ALCA, and AMCA was linked with older age at diagnosis (10–17 years) and ileocolonic disease location. The ASCA had the greatest accuracy for diagnosis and differentiation of CD.

Conclusions: Qualitative and quantitative serologicalal response to glycan epitopes was associated with distinct clinical presentation in paediatric CD patients. This raises the possibility for the use of these markers to differentiate subgroups of CD patients with more sever clinical presentation. The ASCA was the most accurate serological marker for CD; however, testing for the new anti-glycan antibodies may constitute an adjunctive tool in a specific group of patients to aid in the differentiation of CD with absent ASCA from ulcerative colitis.

Introduction

Crohn's disease (CD) and ulcerative colitis (UC), the two major subtypes of inflammatory bowel disease (IBD), represent heterogeneous groups of chronic, systemic inflammatory disorders, mainly affecting the gastrointestinal tract. Although the exact aetiology still remains a subject of debate, there is clear evidence that an aberrant immune response to intestinal microbiome components affected by environmental factors plays a role in genetically predisposed individuals [1, 2]. Both UC and CD present with a wide range of pathogenetic pathways that have diverse and often nonspecific clinical presentations, a variable course, and an unpredictable response to treatment [3–6]. Establishing the diagnosis and differentiation of CD and UC is a complex and sometimes challenging process, based on a combination of clinical, laboratory, imaging, endoscopic, and pathological criteria [7, 8]. Despite impressive progress in the evaluation tools, in some patients the diagnosis is delayed or missed, and a subset of patients with isolated colitis remains unclassified. In the data recently published by de Bie *et al.* on the European paediatric IBD registry, 9% of 2087 newly diagnosed patients were identified with an unclassified type of the disease (IBD-U) [5].

Aberrations in immune response are well known to be present in IBD, and antibodies directed against a variety of microbial and self antigens have been proposed as useful tools in differentiating patients with IBD from non-IBD individuals, and in distinguishing the subtypes of IBD [9-11]. The first described and yet the most extensively studied CD-associated serological marker directed to mannose epitopes of the cell wall of the yeast Saccharomyces cerevisiae (ASCA) is present in 29-69% of CD patients, 0–29% of UC subjects, and 0–16% of healthy controls [9, 10, 12]. For UC, DNase-sensitive "atypical" antibodies directed to perinuclear components of neutrophils (pANCA), present in 41-73% of UC, 6-38% of CD subjects, and 0–8% of controls, remain a recognisable serological marker [9, 10]. The results presented by numerous studies indicate that both ASCA and pANCA individually have only modest accuracy in detecting and differentiating inflammatory bowel disease, and concerns about the sensitivity of the reported wide range extending from 37% to 72% limit their usability as a screening tool [7, 13]. After pANCA and ASCA, many additional types of antibodies have been described as being linked with CD, including those against Pseudomonas fluorescens-associated sequence I2 (Anti-I2), outer membrane porin C (OmpC) of Escherichia coli, and bacterial flagellin cBir1 (Anti-CBir1) [10, 14]. The correlation of the presence (qualitative) and level of serological (quantitative) response to those various protein and carbohydrate antigens with the disease phenotype, disease severity, and the course of disease and disease-related surgery requirements, have been reported for CD patients [9, 10, 15–18]. More recently, the presence of members of the anti-glycan antibodies family other than ASCA, including anti-mannobioside carbohydrate IgA antibodies (AMCA), anti-laminaribioside carbohydrate IgA antibodies (ALCA), and anti-chitobioside carbohydrate IgG antibodies (ACCA), has been recognised as being associated with CD in adults, but limited data are available with respect to paediatric patients [19–22].

In the era of personalised medicine, identification of patients with complicated disease and a poor prognosis is becoming a challenging issue, with special importance attached to decision-making strategy to optimise the therapy for individual patients.

Aim

Therefore, it has been our objective in the present study to assess the profile of serological response to the panel of glycan epitopes (ASCA, ACCA, ALCA, and AMCA) and to evaluate a new set of anti-glycan antibodies, their diagnostic and differential usefulness, and their potential association with disease phenotype in paediatric-onset CD patients.

Material and methods Patient population

Blood samples were obtained from 109 CD patients and 25 UC patients, who were referred for investigation to our tertiary-care referral centre. As the controls, we recruited 67 children (34 boys, 33 girls, mean age (± SD) 10.63 ±4.56 years) diagnosed with acute diseases (acute gastroenteritis, acute ear, nose, or throat disorders). The exclusion criteria for the controls included the following: suspicion of inflammatory bowel disorders, gastrointestinal bleeding, presence of any chronic diseases, and positive family history for immunological and gastrointestinal disorders. Diagnosis and differentiation of CD and UC were performed based on clinical, endoscopic, histological, and imaging investigations according to the Porto criteria [7]. The phenotype of the disease was assigned using the Paris classification [8]. Clinical and demographic characteristics of the IBD patients are presented in Table I. Informed consent was obtained from all parents/ legal guardians and from patients above 16 years of age. The study was approved by the Institutional Research Ethics Board at the Jagiellonian University Medical College.

Serological analysis

All blood samples were drawn at the time of the enrolment, and further analysis was performed in a random and blind fashion. To detect ACCA IgA, ALCA IgG, and AMCA the IgG sera were immediately separated by centrifugation and stored at -20°C until analysis, which was performed with commercially available, specific enzyme-immunosorbent assay (ELISA) kits (Glycominds LTD, Lod, Israel) according to the manufacturer's protocol (Glycominds LTD, Lod, Israel). The results were presented as arbitrary ELISA units (EU), which were calculated based on the sample and calibrator optical density and documented both in absolute values and frequency of positivity. The cut-off values for positive tests were assessed according to the manufacturer's recommendations as follows: 90 EU/ml, 60 EU/ml, and 100 EU/ml for ACCA, ALCA, and AMCA, respectively. In addition to the qualitative analysis, to evaluate the level of immune response of ACCA, ALCA, and AMCA and possible association with distinct CD phenotypes, guartile scores for each of these serologicalal markers were calculated. In brief, for each antigen, the patients whose antibody levels were in the first, second, third, and fourth quartile of the distribution were assigned a quartile score of 1, 2, 3, and 4, respectively. By adding individual quartile scores for each of the glycan antigens a quartile sum score (range Q3–Q12) representing the cumulative immune response toward all the three new anti-glycan antigens for each individual patient was calculated.

Variables	Crohn's disease patients (N = 109)	Ulcerative colitis patients (N = 25)
Male/female, n (%)	64/45 (58.7)	8/17 (32.0)
Median age at diagnosis (IQR) [years]	11.2 (7.4–15.2)	12.8 (10.4–15.3)
Mean age of symptoms onset (± SD) [years]	10.0 ±4.8	11.8 ±3.5
Mean time of symptoms duration before diagnosis (± SD) [months]	12.4 ±18.9	3.05 ±4.1*
CD age (A) at diagnosis, n (%): A1a (< 10 years) A1b (10–17 years) A2 (17–40 years)	46 (42.2) 47 (43.1) 16 (14.7)	- - -
CD location (L), <i>n</i> (%): L1 (ileal) L2 (colonic) L3 (ileocolonic) L4 _a + L4 _b (upper GI proximal to 1/3 distal ileum)	8 (7.3) 41 (37.6) 60 (55.0) 53 (48.6)	- - - -
CD behaviour (B), <i>n</i> (%): B1 (inflammatory) B2 (stricturing) B3 (penetrating) p (perianal disease)	91 (83.5) 15 (13.7) 3 (2.8) 28 (25.7)	- - - -
CD growth retardation (G), <i>n</i> (%): G0 (no evidence) G1 (growth delay)	78 (71.6) 31 (28.4)	- -
UC extension (E), <i>n</i> (%): E1 (proctitis) E2 (distal to splenic flexure) E3 (distal to hepatic flexure) E4 (proximal to hepatic flexure)	- - -	1 (4.0) 2 (8.0) 3 (12.0) 19 (76.0)
UC severity (S), n (%): S0 (never sever; PUCAI < 65) S1 (ever severe; PUCAI \geq 65)	-	17 (68.0) 8 (32.0)

Table I. Characteristics and clinical phenotypes of the inflammatory bowel disease cohort

CD – Crohn's disease, UC – ulcerative colitis, SD – standard deviation, IQR – interquartile range, PUCAI – Paediatric Ulcerative Colitis Activity Index, *p < 0.001

Determination of ASCA IgA and IgG was performed by the indirect immunofluorescence technique using a commercially available kit according to the manufacturer's instructions (EUROIMMUN, Germany). The cut-off values for positivity were set at 1/100 for IgG and 1/1000 for IgA, as recommended by the manufacturer (EUROIMMUN, Germany). Diagnostic parameters (sensitivity, specificity, positive and negative predictive values) were calculated individually for each of the serological markers. To determinate the accuracy of ACCA, ALCA, and AMCA in differentiation between CD, UC, and the controls independently of the cut-off values, receiver operating characteristic (ROC) curves were generated by plotting sensitivity versus 1-specificity.

Statistical analysis

Data were analysed using StatSoft Statistica software system version 10. All the quantitative variables were presented as means \pm standard deviations (SD)

for symmetrically distributed data, or as median interquartile range (INQ) for skewed data. All categorical variables were presented as numbers of patients and percentages. Unpaired categorical data were compared using the χ^2 or Fischer's exact test, as appropriate. The unpaired *t*-test, Wilcoxon's rank-sum test, or Kruskal – Wallis test were used to compare continuous variables, as appropriate for the data normality and number of contrasting groups. The Mann-Whitney test was used for comparison of medians. Multivariate logistic regression analysis was performed to define independent predictors of seropositivity for each individual marker and age at diagnosis, location, and phenotype of CD.

For the analysis of quantitative immunological response, the Cochran-Armitage test was used to determine whether there was a linear trend between the ordered categories of antibody response defined by quartiles and the proportion of patients with a specific disease. Results were expressed as odds ratios (Odds) and 95% confidence intervals (CIs). Statistical significance was defined as a two-tailed *p*-value < 0.05.

Results

Presence and profile of anti-glycan antibodies in Crohn's disease patients

A significant proportion of CD patients responded to glycan antigens. The prevalence of ASCA, ACCA, ALCA, and AMCA in CD and UC patients as well as the controls is presented in Table II. ASCA were positive in 59.6% of CD patients and in 10.5% of the controls (OR = 12.38, 95% CI: 5.19–29.57, *p* < 0.001). There was no difference in the prevalence of ASCA in UC compared to the control group (OR = 0.745, 95% CI: 0.144–3.857, p = 0.726), but it was significantly lower as compared to CD patients (OR = 0.001, 95% CI: 0.0003–0.006, *p* < 0.001). Among the new anti-glycan antibodies, ALCA were significantly more prevalent in CD than in UC (OR = 14.614, 95% CI: 2.858–240.979, *p* < 0.007) or in the controls (OR = 9.176, 95% CI: 2.092–40.254, *p* < 0.001). For ACCA, the values approximated significance for the controls (OR = 2.078, 95% CI: 0.987–4.375, p = 0.055) but were not different in comparison to UC patients (OR = 2.380, 95% CI: 0.783–7.470, p = 0.137).

The presence of antibodies directed to one of the glycan epitopes (ASCA, ACCA, ALCA, or AMCA) was significantly associated with CD (Table II). At least one positive antibody of the panel was more prevalent in CD patients compared both to UC subjects (OR = 9.617, 95% CI: 3.482–26.562, p < 0.001) and the controls (OR = 7.137, 95% CI: 3.614–14.095, p = 0.001). Seventy-five percent of CD patients were

positive for one of the anti-glycan antibodies, while 76% of UC patients and 70% of the controls did not show any antibodies. A higher level of the immune response as expressed by the number of positive anti-glycan antibodies was significantly associated with CD (p = 0.0097); however, a limited overlap of antibodies was observed. The sera of 33.9% of CD patients reacted with two glycan epitopes, while 11.9% of CD subjects were positive for three antigens and only 1 patient expressed all four anti-glycan antibodies (Table II). Testing for ACCA, ALCA, and AMCA allowed for serological identification of an additional subgroup of CD patients, who did not express immunoreactivity toward mannan epitopes of Saccharomyces cerevisiae. Among 45 AS-CA-negative CD patients, serological response to one of the new glycan epitopes was found in almost half (49%) of them, including ACCA present in 31%, ALCA in 13%, and AMCA in 4% of the subjects.

Diagnostic accuracy and use for diagnosis and differentiation

Based on the manufacturer-recommended cut-off values the sensitivity and specificity, positive and negative predictive values of the serological markers were calculated for clinical situations differentiating CD from the controls and distinguishing CD from UC. Table III shows the results for each of the individual markers. All of the anti-glycan antibodies were highly specific (80–100%) for CD, with the highest values characterising ALCA (97–100%). ASCA presented the highest sensitivity for differentiation of CD from both the controls (59%) and UC (59%). The sensitivity of the other an-

Factor	Crohn's disease patients (N = 109)	Ulcerative colitis patients (N = 25)	Controls (N = 67)	
Median (IQR) levels, EU:				
ACCA	37.7 (21.5–54.7)	33.2 (24.7–40.3)	30.9 (23.3–46.1)	
ALCA	25.8 (18.4–57.0)	24.0 (19.1–30.7)	25.4 (18.8–34.1)	
AMCA	42.8 (31.33–60.2)	37.4 (30.1–54.9)	39.1 (29.2–59.1)	
Antibody positivity, n (%)				
ASCA*,**	65 (59.6)	2 (8)	7 (10.5)	
ACCA (≥ 90 EU)	34 (31.1)	4 (16)	12 (17.9)	
ALCA (≥ 60 EU)*.**	24 (22)	0 (0)	2 (2.9)	
AMCA (≥ 100 EU)	11 (10.1)	2 (8)	4 (5.9)	
Number of positive AGABs, n (%)*,**				
1	82 (75.2)	6 (24)	20 (29.9)	
2	37 (33.9)	2 (8)	5 (7.5)	
3	13 (11.9)	0 (0)	0 (0)	
4	1 (1)	0 (0)	0 (0)	

Table II. Anti-glycan antibodies distribution in the studied groups

CD – Crohn's disease, UC – ulcerative colitis, ACCA – anti-chitobioside carbohydrate antibody, ALCA – anti-laminaribioside carbohydrate antibody, AMCA – anti-mannobioside carbohydrate antibody, ASCA – anti-Saccharomyces cerevisiae antibody, AGABs – anti-glycan antibodies (positive ASCA, ACCA, ALCA, or AMCA), IQR – interquartile range, EU – ELISA units, *p < 0.05 CD vs. controls, **p < 0.05 CD vs. UC

	Sensitivity [%]	95% CI [%]	Specificity [%]	95% CI [%]	PPV	95% CI [%]	NPV [%]	95% CI [%]
CD vs. controls:								
ASCA	59	49–68	90	80–96	90	81–96	57	47–67
ACCA (≥ 90 EU)	31	23–41	82	71–90	74	59–86	42	34–51
ALCA (≥ 60 EU)	27	18–37	97	90–99	92	75–99	50	41–59
AMCA (≥ 100 EU)	10	5-17	92	74–99	85	55–98	19	12-27
CD vs. UC:								
ASCA	59	49–68	92	74–99	97	90–99	34	23–46
ACCA (≥ 90 EU)	33	24–43	80	59–93	88	74–96	22	14–31
ALCA (≥ 60 EU)	22	15-31	100	86–100	100	86–100	23	15-32
AMCA (≥ 100 EU)	10	5–17	92	74–99	85	55–98	19	12-27

Table III. Diagnostic accuracy of each studied marker for differentiation of CD (n = 109) with the controls (n = 67) and UC (n = 25)

CD – Crohn's disease, UC – ulcerative colitis, ACCA – anti-chitobioside carbohydrate antibody, ALCA – anti-laminaribioside carbohydrate antibody, AMCA – anti-mannobioside carbohydrate antibody, ASCA – anti-Saccharomyces cerevisiae antibody, PPV – positive predictive value, NPV – negative predictive value, CI – confidence interval, EU – ELISA unit

ti-glycan antibodies was low and had values from 10% to 33% (Table III). The best performance for positive predictive values was shown by ALCA (92–100%) and ASCA (90–97%). For clinical situations, when negative predictive values are of importance to avoid invasive diagnostic procedures, all the new anti-glycan antibodies presented poor levels that ranged from 19% to 50%, as shown in Table III.

To differentiate CD from the controls and UC independently of the cut-off levels, we constructed ROC curves for ACCA, ALCA, and AMCA. All the anti-glycan antibodies showed a weak discriminatory capacity for differentiation of CD from UC (AUC_{ACCA} – 0.554, 95% CI: 0.445–0.664; AUC_{ALCA} – 0.611, 95% CI: 0.506– 0.716; AUC_{AMCA} – 0.568, 95% CI: 0.443–0.693) and the controls (AUC_{ACCA} – 0.5596, 95% CI: 0.474–0.645; AUC_{ALCA} – 0.572, 95% CI: 0.488–0.656, AUC_{AMCA} – 0.553, 95% CI: 0.464–0.642). The only significant difference in antibodies level was found for ALCA, which presented higher values in the older (10 to 17 years) age group (46.3 ±31.54 vs. 31.9 ±24.25 EU, p = 0.012).

Serological response association with disease phenotype

In the univariate analysis, no association was found between the prevalence of each individual marker and disease phenotype, including age at diagnosis, disease behaviour, and disease location (p > 0.05). To determine which antibody was independently associated with specific disease characteristics, multivariate logistic regression analysis was performed. Taking into account separately age at diagnosis, disease localisation, and disease behaviour as potential confounders, we found an independent association between one of the newer anti-glycan antibodies (ACCA, ALCA, AMCA) and ileocolonic disease location (L3; OR = 4.444, 95% CI: 1.670–11.830, p = 0.0147). Among antibodies, only ALCA was significantly more likely to be linked with this disease location (OR = 4.429, 95% CI: 0.5733–34.211, p = 0.018). Isolated ileal disease was omitted from this analysis because of the low number of patients with this location. The multivariate logistic regression model revealed that the presence of one antibody of the entire anti-glycan panel (ASCA, ACCA, ALCA, AMCA) was associated with complicated disease behaviour (B2, B3) (p = 0.043).

To evaluate a possible association between the level of antibody response to the new anti-glycan antigens and CD phenotype, the quartile scores analysis individually for ACCA, ALCA, and AMCA (Table IV) and cumulative quartile scores of the sum (Figure 1) for all the antibodies were calculated as stated in the Material and methods section. We found that an increasing cumulative response to glycans was associated with older age at diagnosis (p = 0.0467) and a decreasing likelihood for disease localisation limited to the colon (p = 0.0295). For individual antibodies, an increasing response associated with older age at diagnosis was significant for ALCA (p = 0.017), and decreasing disease localisation in the colon was significant for ACCA (p = 0.0014). There was no trend among the quartile scores for any of the individual antibodies or for the quartile sum scores and disease behaviour (p > 0.05).

Discussion

In the present study of a well-defined paediatric IBD cohort, we evaluated a potential association between

Markers		Q1	Q2	Q3	Q4	Value of <i>p</i>
ACCA	A1a	15 (33%)	13 (28%)	7 (15%)	11 (24%)	0.15
	A1b	9 (19%)	11 (23%)	15 (32%)	12 (26%)	-
ALCA	A1a	16 (35%)	14 (30%)	9 (20%)	7 (15%)	0.017
	A1b	9 (19%)	10 (21%)	14 (30%)	14 (30%)	-
AMCA	A1a	16 (35%)	9 (20%)	11 (24%)	10 (22%)	0.380
	A1b	11 (23%)	11 (23%)	14 (30%)	11 (23%)	-
ACCA	B1	24 (26%)	19 (21%)	24 (26%)	24 (26%)	0.388
	B2 + B3	4 (22%)	8 (44%)	3 (17%)	3 (17%)	_
ALCA	B1	25 (27%)	23 (25%)	21 (23%)	22 (24%)	0.329
	B2 + B3	3 (17%)	4 (22%)	6 (33%)	5 (28%)	-
AMCA	B1	24 (26%)	21 (23%)	23 (25%)	23 (25%)	0.863
	B2 + B3	4 (22%)	6 (33%)	4 (22%)	4 (22%)	_
ACCA	L1	3 (43%)	3 (43%)	0 (0%)	1 (14%)	
	L2	14 (34%)	9 (22%)	11 (27%)	7 (17%)	0.001
	L3	11 (18%)	15 (25%)	15 (25%)	19 (32%)	-
ALCA	L1	2 (29%)	4 (57%)	0 (0%)	1 (14%)	
	L2	12 (29%)	9 (22%)	13 (32%)	7 (17%)	0.119
	L3	13 (22%)	14 (23%)	14 (23%)	19 (32%)	-
AMCA	L1	3 (43%)	2 (29%)	0 (0%)	2 (29%)	
	L2	13 (32%)	11 (27%)	8 (20%)	9 (22%)	0.117
	L3	12 (20%)	14 (23%)	18 (30%)	16 (27%)	

Table IV. Quartile analysis of the Crohn's disease cohort (n = 109) for the three markers (ACCA, ALCA, and AMCA) for age at diagnosis (A), disease behavior (B), and disease location (L)

CD – Crohn's disease, Q – quartile, ACCA – anti-chitobioside carbohydrate antibody, ALCA – anti-laminaribioside carbohydrate antibody,

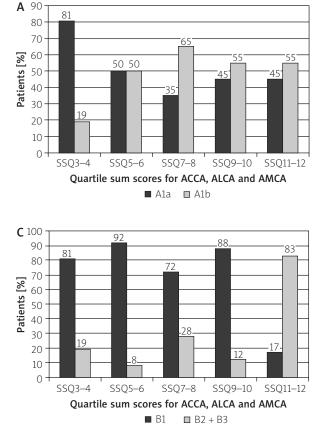
AMCA – anti-mannobioside carbohydrate antibody, ASCA – anti-Saccharomyces cerevisiae antibody, A1a – age of diagnosis < 10 years, A1b – age of diagnosis 10–17 years, B1 – inflammatory disease, B2 – stricturing disease, B3 – penetrating disease, L1 – ileal disease, L2 – colonic disease, L3 – ileocolonic disease

an adaptive response to the panel of glycan epitopes, including ASCA, ACCA, ALCA, and AMCA, and the clinical presentation of CD. Moreover, we tested whether the new markers (ACCA, ALCA, and AMCA) could be helpful in disease differentiation and stratification. We confirmed the association between some anti-glycan antibodies (ACCA, ALCA) and CD, detecting a correlation between the presence of antibodies and complicated disease presentation and ileocolonic disease location. In addition, a higher level of adaptive response was associated with older age at diagnosis and ileocolonic disease. Although newer anti-glycan antibodies were highly specific for CD, their capability as a screening tool for diagnosis of primary CD was limited, and ASCA antibodies remain the most accurate single marker for paediatric CD.

Profile of anti-glycan antibodies and association with disease phenotype

In this study, qualitative and quantitative immune response to glycan epitopes was strongly associated

with CD and specific disease phenotype. Notably, the serological response to glycans was complex, and despite a relatively high frequency of antibodies present in CD patients, a limited overlap was found. The observed wide variation in both the number and magnitude of immune responses as expressed by the quartile sum scores to glycans indicates that the degree of reactivity to microbial components is not uniform in all paediatric CD patients. Testing for the new set of antibodies, which allowed for serological identification of an additional subgroup of CD patients, may facilitate the discrimination of indeterminate IBD-phenotype. Forty-nine percent of those who did not show immune reactivity toward mannan epitopes of Saccharomyces cerevisiae were positive for at least one other marker from the panel. A similar distribution of anti-glycan antibodies with respect to the status of the markers as well as diversification of the response to anti-glycan epitopes have been published in studies performed in adults [17, 19, 21, 22]. Rieder et al. reported 40.9% ASCA-negative



CD children to be positive for at least one other marker from the anti-glycan panel [20]. This illustrates the importance of combined testing of newer glycan antibodies in a specific group of patients, and their potential usefulness as an additional tool in cases of diagnostic uncertainty.

There is a growing body of evidence indicating that the pathogenesis of CD is strongly associated with aberrant recognition of resident intestinal microbial components [1]. Glycans are cell wall surface carbohydrate components that can be found on various pathogenic and saprophytic microorganisms playing a role in diversified immunological processes and stimulating immune response in a Syk-dependent way [19, 23]. The non-uniform nature of the serological response to glycan epitopes may reflect different levels of the altered host-microbial relationship and might be related to diverse immunological mechanisms (e.g. antigen sampling, antigen recognition, antigen clearance) affected by genetic heterogeneity and/or varied microbiome components [1]. Therefore, specific antibodies may be the sign of an underlying pathomechanism associated with a characteristic disease presentation. The inflammatory behaviour is dominantly diagnosed in CD paMałgorzata Sładek, Agata Wasilewska, Agnieszka Świat, Adam Ćmiel

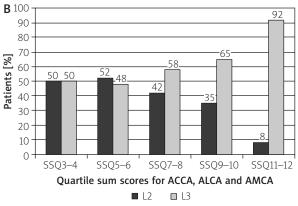


Figure 1. Cumulative quartile sum analysis of the Crohn's disease cohort for the three tested anti-glycan antibodies (ACCA, ALCA, and AMCA). The distribution of quartile sum scores in relation to age at diagnosis (p = 0.47) (**A**), disease localisation (p = 0.047) (**B**) and disease behavior (p = 0.94) (**C**)

CD – Crohn's disease, SSQ – quartile sum scores (after aggregation by two), ACCA – anti-chitobioside carbohydrate antibody, ALCA – anti-laminaribioside carbohydrate antibody, AMCA – antimannobioside carbohydrate antibody, ASCA – anti-Saccharomyces cerevisiae antibody, A1a – age of diagnosis < 10 years, A1b – age of diagnosis 10–17 years, L1 – ileal localisation, L2 – colonic localisation, L3 – ileocolonic localisation, B1 – inflammatory disease, B2 – stricturing form, B3 – penetrating form

tients at disease onset, but the disease phenotype is not stable and progression to a complicated form can evolve in most patients during years of disease duration [3, 4]. After initial observations concerning ASCA, several other serologicalal markers have been linked with more complicated disease presentation and severe disease course [15]. Studies in adults indicate that patients with complicated CD disease were more likely to show the presence of anti-glycan antibodies; however, an inconsistent association between individual anti-glycan antibodies and disease phenotype has been reported [17-22, 24-29]. We confirmed that in paediatric patients, anti-glycan antibodies were associated with a complicated disease already at disease onset. Because it was a single time point study with no follow-up, we cannot draw a conclusion addressing a predictive association with the disease course. The only paediatric study to date suggests an association between anti-glycan antibodies and complicated disease phenotype, and a predictive role of the more severe course of the disease [20]. Once the diagnosis of CD is established, anti-glycan antibodies might be an important serologicalal marker for disease stratification and could be considered as an additional tool to identify

high-risk patients. This may have a significant clinical implication because the introduction of a more potent therapeutic regimen (e.g. biologic and/or immunomodulators) early in the disease course may be of benefit in patients with a higher risk of complicated CD.

In agreement with studies in adults, we found that the magnitude of the immune response to glycans was associated with a more extensive disease localisation, involving the colon and terminal ileum [22, 25]. However, unlike Rieder et al., we were not able to show an association between anti-glycan antibodies and ileal CD [20]. It could be partially explained by the much lower proportion of patients with limited ileal localisation, which in our cohort was present in only 8 (7.3%) subjects (Table I). Paediatric CD is characterised by a large proportion of colonic involvement, ileitis is usually associated with colitis (55% of the patients in our cohort), and the cumulative probability of isolated ileal CD increases with age until puberty [30]. Differences in disease classification, since we applied the Paris classification and time of blood procurement in relation to the stage of disease duration, may influence the results and limit the possibility of comparing these two studies [8].

Anti-glycan antibodies diagnostic accuracy

The diagnostic differentiation between CD and UC has both therapeutic and prognostic implications. In the majority of IBD patients it can be made based on clinical, endoscopic, histopathological, and radiological features. However, in a subgroup of patients with strictly colonic disease, differentiation may be a challenge and serological tests could be supportive. As in previous studies, our results indicate that the new anti-glycan antibodies are specific for CD [9, 19]. However, no antibody demonstrated a sufficient sensitivity to support their usefulness as a screening tool for primary paediatric CD in the diagnostic workup algorithm. The diagnostic accuracy of the ASCA and ALCA for differentiation of CD versus UC has been reported to be similar between adults and children, while some discrepancies have been found for AMCA and ACCA [25, 26, 31]. As in the case of our data, most studies indicate that among the new anti-glycan antibodies, AMCA gives poor values for both sensitivity and specificity [9]. When analysing the differentiation capability of the markers, the new anti-glycan antibodies taken individually have not improved differentiation between CD and UC and the controls. ASCA may still be considered the most accurate single marker from the anti-glycan panel.

Analysis of the diagnostic accuracy of newer anti-glycan antibodies, independently of the recommended cut off values, showed no differences of titre levels between the studied groups. Moreover, the ROC curves have shown a weak discriminatory capability. Data from adult studies are inconsistent in terms of titres for individual markers, as well as a possible association between the serological response and age at the disease onset [20-22, 25, 26]. The only paediatric study indicates that the cut-off values for the new anti-glycan antibodies are lower than those for adults [20]. In our study, an increasing magnitude of the immune response to glycans has been associated with older age at diagnosis, which could be explained by a dependency in antibody production on the age at diagnosis, as previously described [32]. Therefore, the age-dependent differences in serological response should be taken into account and different cut-off values for optimal performance of the anti-glycan antibodies may apply as dependent on age. Further studies on larger patient populations to establish paediatric referral cut-off values are definitely needed.

This study is not without limitations. The first possible limitation is due to a tertiary centre referral bias and higher pre-test probability compared to the general population, due to the diagnosis of CD and UC already having been established at the time of sample procurement. The second limitation is the cross-sectional study design with single point antibody measurements and no longitudinal observation. Therefore, results in the conclusion regarding the prognostic values of the anti-glycan antibodies from this study should be formulated with caution. On the other hand, ASCA appears even years before CD diagnosis, and until now a predominant numbers of studies in adult patients have reported antibody responses to be stable over time, despite different therapeutic modalities being employed, including active and inactive disease [33-35]. As mentioned above, distinct cut-off values for the paediatric CD population, for the optimal performance of the markers, have to be taken into account although the optimal cut-off values for the glycan marker panel have not been determined to date.

Conclusions

Our results suggest that some new anti-glycan antibodies are associated with paediatric CD and specific disease phenotype, and may be supportive in stratifying disease subtypes. With the growing evidence of an association between serological response and a complicated and more extended disease phenotype, the most important clinical value of anti-glycan antibodies could be in the stratification of patients according to the risk of presentation a more severe disease. Therefore, further studies are definitely needed to evaluate the said risk. The diagnostic significance of individual new anti-glycan antibodies to differentiate IBD from non-IBD and CD from UC patients is of limited value and cannot replace conventional clinical evaluation. However, such tests may constitute an adjunctive tool in a specific group of patients, to aid in the differentiation of CD patients with absent ASCA from ulcerative colitis subjects.

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