

1 **Drivers of survival in a small mammal of conservation concern: an**
2 **assessment using extensive genetic non-invasive sampling in fragmented**
3 **farmland**

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36 **Abstract**

37 Although important to guide conservation management, detailed demographic studies on rare
38 or elusive species inhabiting fragmented, human-dominated landscapes are often hampered by
39 the species' low densities, and the logistic and ethical constraints in obtaining reliable
40 information covering large areas. Genetic non-invasive sampling (gNIS) provides cost-effective
41 access to demographic information, though its application to small mammals is still scarce. We
42 used gNIS to infer on the demography of an endemic small mammal, the Cabrera vole (*Microtus*
43 *cabreræ*), occurring as a spatially-structured population in a 462-ha Mediterranean farmland
44 landscape. We intensively sampled fresh vole feces in four seasons, extracted the DNA, and
45 performed individual identification based on genotypes built using nine microsatellites. We then
46 estimated population size and individual survival relative to environmental variables, controlling
47 for heterogeneity in capture probabilities using capture-mark-recapture modelling. Population
48 size increased during the wet season and decreased during the dry season, while survival
49 remained constant across the study period. Individuals captured along road-verges and around
50 water-bodies survived longer than those captured near agricultural fields. The use of gNIS on a
51 heterogeneous landscape such as our study area allowed us to demonstrate that human land-
52 use activities affect Cabrera vole demographic parameters in Mediterranean farmland, with
53 implications for conservation planning towards its long-term persistence. Our approach can be
54 widely applied to other elusive small mammals of conservation concern, but for which
55 informative demographic data are still scarce.

56

57 **Keywords**

58 Agricultural intensification; *Capwire*; Cormack-Jolly-Seber; Mediterranean farmland; *Microtus*
59 *cabreræ*; Road effects.

60

61 **1. Introduction**

62 Estimating large-scale demographic patterns (e.g. abundance, population growth, survival) of
63 animal species in relation to both individual traits (e.g. sex, age, weight) and environmental
64 factors (e.g. climate or land-use change) is a difficult but necessary goal to understand species
65 ecology and sustain conservation policies (Smallwood and Schonewald, 1998; Williams et al.,
66 2002). This is particularly true for species occurring in agricultural landscapes where major
67 declines in biodiversity due to agricultural intensification have been reported worldwide
68 (Tschamntke et al., 2012). However, achieving these goals is often difficult due to a number of
69 technical, ethical, and logistic constraints in data collection, particularly for species that are rare,
70 elusive, or otherwise hard to capture or observe.

71 Capture-Mark-Recapture (CMR) is one of the most popular methods to assess demographic
72 parameters in animal populations (Lebreton et al., 1992), and hence to understand species'
73 biology and ecology in different environments (Smallwood and Schonewald, 1998). Traditional
74 CMR studies have been mostly based on live-trapping techniques, which are usually logistically
75 difficult to implement over large spatial and temporal scales and often expensive (Cheng et al.,
76 2017). In addition, because live-trapping implies both physical confinement and handling of
77 animals, it often involves behavioural and physiological responses due to trapping-induced
78 stress (Beja-Pereira et al., 2009; De Bondi et al., 2010). Stress responses can be reduced with the
79 use of minimally invasive techniques such as camera-trapping, which is expected to be more
80 time-efficient than live-trapping and does not require physical capturing and handling of animals
81 (De Bondi et al., 2010; Mondol et al., 2009). However, camera trapping is unsuitable for CMR
82 studies in species that are difficult to morphologically identify at the individual level, which is
83 the case of most small mammal species (Glen et al., 2013). Furthermore, in the case for rare and
84 elusive species, both live- and camera-trapping often yield insufficient data to be used in CMR
85 models, thus hampering proper evaluation of their population status and trends (Burgar et al.,
86 2018; Mondol et al., 2009).

87 Genetic non-invasive sampling (gNIS) has been increasingly used to estimate demographic
88 parameters of species that are difficult to trap, mainly due to decreased field sampling effort,
89 ever decreasing lab costs, and increasing DNA amplification success (Beja-Pereira et al., 2009;
90 Marucco et al., 2011). Despite its limitations in retrieving information on relevant individual
91 traits like age, body mass, or reproductive condition, gNIS can provide a more cost-effective

92 solution than traditional live-trapping (Cheng et al., 2017; Ferreira et al., 2018). DNA extracted
93 from non-invasive samples (e.g. feces, hairs, feathers) allows the identification of individuals,
94 providing data that can be easily combined with CMR methods to obtain population parameters
95 that otherwise would be difficult to obtain over large spatial scales (Cheng et al., 2017; Petit and
96 Valiere, 2006). However, to date, applications of gNIS in CMR studies have mostly focused on
97 large and medium-sized mammal species, and often provide snapshots of population size
98 estimates rather than variations over time (but see Brøseth et al., 2010 for an example).
99 Furthermore, very few studies have used gNIS to estimate other important population
100 parameters such as survival (Lampa et al., 2015; Marucco et al., 2012; Zielinski et al., 2013). In
101 the case of small mammals, while some recent studies have used gNIS to estimate population
102 density (DeMay et al., 2017; Gillet, 2016; Sabino-Marques et al., 2018) or to infer dispersal
103 (Ferreira et al., 2018; Gillet, 2016), to our knowledge no study has yet explored the application
104 of this method to understand how demographic parameters relate to large-scale environmental
105 variation.

106 In this study, we combined gNIS and CMR methods to assess the seasonal variations in
107 abundance, and to evaluate factors affecting survival probability of an elusive small mammal
108 species in a Mediterranean farmland landscape. We focused on the 'near-threatened', Iberian
109 endemic Cabrera vole (*Microtus cabreræ*, Thomas 1906), for which genotyping protocols based
110 on faecal samples have been recently optimized (Barbosa et al., 2013; Ferreira et al., 2018).
111 Additionally, previous studies have also shown the ability of gNIS to provide reliable density
112 estimates for this species (Sabino-Marques et al., 2018). Based on repeated surveys of Cabrera
113 vole feces, we explored the potential of gNIS to (i) assess the seasonal variation in population
114 abundance; and (ii) estimate capture and survival probabilities in relation to variables reflecting
115 survey conditions (genotyping success and season), individual traits (sex), and local and
116 landscape environmental features. We considered variables that might affect survival both
117 positively (e.g. patch area and presence of water) and negatively (e.g. isolation, patch
118 persistence, interactions with the competitor *Arvicola sapidus*, and human disturbances) (Pita et
119 al., 2014) (see Table 1 for a full description and rationale of covariates considered). Overall, our
120 study illustrates the use of gNIS within a CMR framework, demonstrating its application to
121 retrieve demographic data from elusive small mammals, thus enhancing conservation planning
122 in areas that have been highly modified by human activities.

123

124 2. Material and Methods

125 2.1. Study area and species

126 The study was carried out in a 461.8 ha area within the coastal plateau of south-western
127 Europe, Portugal (37° 21' - 38° 04' N, 08° 51' - 08° 30' W) (Fig. 1). The region is included in the
128 thermo-Mediterranean bioclimatic zone (Rivas-Martínez, 1981), with a mean annual
129 temperature of 16.5°C (monthly temperatures ranging from 6 to 29°C), and an annual rainfall of
130 about 650 mm (of which >80% falls between October and March) (Pita et al., 2007; 2006). The
131 landscape is mostly flat (56-76m above sea level) and land cover is dominated by pastures and
132 annual irrigated crops (Pita et al., 2007). Forest cover is limited to a few woodlots and hedges
133 with pines and eucalyptus, while natural woodlands, shrubs, and marshy vegetation are most
134 frequent along road verges (mostly dirt-roads with low traffic), and in the surroundings of
135 extensive agricultural fields (Pita et al., 2009). Over the past three decades, agricultural practices
136 have been strongly intensified, particularly through the expansion of cultivated land, associated
137 with the frequent use of pesticides and chemical fertilizers, with detrimental impacts on
138 biodiversity (Ferreira and Beja, 2013; Peralta et al., 2016; Pita et al., 2009).

139 Previous studies in the region have shown that Cabrera voles typically show a metapopulation-
140 like spatial structure (Pita et al., 2007), occurring within damp habitat patches densely covered
141 by tall wet herbs and shrubs along small streams, temporary ponds, field margins, and roadside
142 verges (Pita et al., 2014; 2006). Population densities within patches are typically low (Pita et al.,
143 2011; 2010), and individuals are rarely present in the same area for more than 4 months
144 (Fernández-Salvador et al., 2005b). Within habitat-patches, the Cabrera vole is often organized
145 in monogamous breeding pairs and tend to exhibit home-ranges of only a few hundred square
146 meters (Pita et al., 2010).

147 2.2. Vole surveys and environmental variables

148 Following the studies by Ferreira et al. (2018) and Sabino-Marques et al. (2018) conducted
149 previously in a smaller area (78ha) within the same study region, we surveyed Cabrera voles on
150 four main occasions (seasons) along one year: early wet season (EWS, November-December
151 2013); late wet season (LWS, February-March 2014); early dry season (EDS, May-June 2014);
152 and late dry season (LDS, September-October 2014). In each season, we used a two-step
153 procedure, first identifying and mapping all habitat patches potentially used by Cabrera voles

154 (damp areas with vegetation patches dominated by dense and tall herbs) using both Bing
155 Maps™ aerial photographs (retrieved in 2012) and ground validation (Fig. 1). Then, a survey was
156 conducted over the whole area of each patch, to detect the presence of the species from its
157 characteristic signs (faecal pellets and grass clippings), and to collect fresh feces for genetic
158 analysis. Within each patch, two samples were collected (if available) every 5 m (in order to
159 maximize the chances of detecting different individuals) (Fig. 1), using sterilized tweezers, and
160 stored in the field in individual 2 mL microtubes with 96% alcohol, and later kept at -20°C until
161 DNA extraction. To minimize cross-contamination from conspecifics, feces were collected from
162 small latrines (<20 faecal pellets). The presence of the competitively superior southern water
163 vole (*Arvicola sapidus*) was recorded based on the presence of similar but larger faecal pellets
164 than those from the Cabrera voles, which are easily recognizable (Peralta et al., 2016). The
165 surveying of the study area followed the same direction (from NW to SE) in each sampling
166 season, such that habitat-patches were sampled following approximately the same order and
167 time-intervals.

168 All information regarding the voles' habitat, matrix land-uses, and sample geographic location
169 was stored in a vector-based Geographic Information System (GIS; QGIS, version 2.14.10 - Essen,
170 QGIS Development Team, 2016).

171 **2.3. DNA extraction and genotyping**

172 Due to budget restrictions, only a limited number of faecal samples could be analysed. The
173 selection of samples for genetic analyses followed a stepwise approach to reduce costs while
174 aiming to achieve a comprehensive spatial coverage of each patch, hence maximising the
175 number of captures and recaptures of individuals (Ferreira et al., 2018). In patches where less
176 than six samples were collected, all samples were analysed. In each of the remaining patches,
177 we selected at least 60% of the samples, evenly spread throughout the patch. When more than
178 one sample was collected every 5 m, only one was initially analysed. If genotyping failed for the
179 first sample, the second sample was analysed in order to obtain a minimum number of
180 genotypes per patch of at least 40% of all sampling sites.

181 Vole DNA was extracted using the E.Z.N.A.® Tissue DNA Kit (OMEGA bio-tek) following the
182 manufacturers' instructions, with an initial digestion step using a lysis washing buffer (Maudet
183 et al., 2004) for 15 minutes at 56°C. Samples were genotyped for a set of nine microsatellites
184 characterized by high levels of polymorphism ($H_o = 0.79$; $H_e = 0.81$), low probability of identity

185 of unrelated ($PI= 3.2 \times 10^{-12}$) and related individuals ($PIsibs= 9.2 \times 10^{-5}$), and high probability of
186 exclusion ($PE=0.99$), and two small sized sex-linked introns (Table A1, Appendix A). These
187 markers have been optimised for application to gNIS of Cabrera voles feces (Ferreira et al.,
188 2018), and provide accurate individual identifications and population estimates (Sabino-
189 Marques et al., 2018). We followed the protocol described by Ferreira et al. (2018), which
190 includes an initial screening of DNA quality using three species-specific microsatellite loci. The
191 samples that amplified for the three loci were then amplified for the additional six microsatellite
192 loci and two sex-linked introns. To account for genotyping errors (e.g. allele dropout and false
193 alleles) and obtain a consensus genotype, each multiplex reaction was replicated a minimum of
194 four times (three times for the sex-linked introns). To confirm species identification, a small
195 fragment of cytochrome-*b* gene was amplified in all genotyped samples following Barbosa et al.
196 (2013). To evaluate eventual biases in the estimation of genotyping success rate due to
197 misidentification of feces in the field, we also performed genetic species identification in at least
198 20% of the samples that failed during genotyping. The extractions and PCR reactions were
199 conducted in a physically isolated room, where all the equipment was sterilized with bleach and
200 ethanol, and exposed to UV light before and after usage. Negative controls were included in
201 each manipulation, maintaining conditions to monitor and reduce the risk of DNA
202 contamination (Barbosa et al., 2013; Beja-Pereira et al., 2009; Costa et al., 2017). All products
203 were sequenced on a ABI3130 Capillary Sequencer (Applied Biosystems). Allele calling of the
204 microsatellite loci and sex chromosome introns was performed using GeneMapper (v.4.0;
205 Applied Biosystems). Cytochrome-*b* gene sequences were analysed in Geneious 8 (Kearse et al.,
206 2012).

207 Consensus genotypes for the successfully genotyped samples were obtained by analysing all
208 replicate genotypes with Gimlet v.1.3.3 (Valiere, 2002). For genotypes differing only by one or
209 two loci or with up to two missing data, additional PCR replicates were performed to complete
210 genotypes with missing data, and to check for genotyping errors. Consensus genotypes for each
211 sample were then compared with each other to identify individuals. Following the criteria
212 detailed in Ferreira et al. (2018) and Sabino-Marques et al. (2018), only samples that differed in
213 more than two loci were assigned as new individuals. Genotyping error rates were estimated
214 using Pedant (Johnson and Haydon, 2007), with 10 000 search steps. Since the software only
215 compares two replicates at a time, we carried out all possible pairwise comparisons and then
216 averaged the results.

217 2.4. Data analysis

218 To estimate vole abundance (N), we considered each seasonal survey as a single sampling event
219 and used ‘continuous-occasion’ closed-population CMR models allowing for multiple captures of
220 the same individual within each survey. Specifically, we used the *capwire* estimator based on
221 urn models (Miller et al., 2005; Pennell et al., 2013), considering two alternative formulations:
222 the equal capture probability model (ECM) and the two innate rates model (TRIM). In ECM, all
223 individuals are considered equally likely to be captured on each survey, while in TRIM there is a
224 mixture of two types of individuals with different capture probabilities. A likelihood-ratio test
225 was used to evaluate the fit of both models and determine the best fit (Miller et al., 2005). We
226 used a parametric bootstrap test with 1000 samples to generate the 95% confidence interval for
227 population estimates of the best model for each season. Both ECM and TRIM were fitted and
228 compared in the package *capwire* (version 1.1.4) (Pennell et al., 2013) for R (version 3.3.2) (R
229 Core Team, 2016). For comparison purposes, we also used Chao’s lower bound estimator
230 assuming individual heterogeneity in capture probabilities (M_h -Chao) (Chao, 1989). Although the
231 M_h -Chao estimator assumes different capture occasions, it uses only the capture frequency, so
232 it may be applied to our data (Miller et al., 2005). This estimator is thought to outperform
233 *capwire* for large datasets ($N > 200$) (Miller et al., 2005), such as that used in this study (see
234 Results). For this, we used the R package *Rcapture* (version 1.4.2) (Baillargeon and Rivest, 2007).

235 To estimate monthly recapture (p) and survival (ϕ) probabilities we used the Cormack-Jolly-
236 Seber (CJS) open-population model approach (Lebreton et al., 1992), implemented in *RMark* (v
237 2.2.4; Laake, 2013), an R interface for software package MARK (White and Burnham, 1999). For
238 this, we first collapsed within-season capture histories for each genotyped individual into a
239 single value (0/1), denoting whether it was identified or not at each season (McCrea and
240 Morgan, 2015). We then tested the goodness-of-fit on a fully time-dependent CJS model
241 (McCrea and Morgan, 2015) using the *R2ucare* (v 1.0.0; Gimenez et al., 2017), which suggested a
242 good fit of the data ($\chi^2=2.34$, p-value=0.311). Potential predictors of CJS parameters included
243 the effects of genotyping success on p ; the effects of time, sex, and patch area on both p and ϕ ;
244 and the effects of patch isolation, patch permanence, presence of road-verges, presence of
245 permanent water-bodies, detection of southern water voles, distance to the nearest agricultural
246 area (vegetable gardens, orchards, or ploughed fields), and distance to the nearest urban area
247 (e.g. houses, social areas, buildings) on ϕ (see full description and rationale of covariates in Table

248 1). Environmental covariates were specified at the individual level, considering the moment of
249 first capture.

250 Before model building and selection, we assessed the correlations among predictors of p and
251 among predictors of ϕ , retaining in the analyses only the predictors with correlation coefficients
252 <0.30 (i.e. low correlations; Graham, 2003). Such a conservative criterion was chosen because in
253 CJS models, as the number of predictors increases, so will the number of model parameters, and
254 hence the number of possible models under evaluation, which decreases the ability to
255 distinguish between informative and spurious variables (Doherty et al., 2012). Among the
256 potential predictors of p , we found a negative correlation between genotyping success and
257 patch area (Table A2, Appendix A). Because p should be most critically affected by variables
258 more directly related to the sampling design, we retained genotyping success in the analyses to
259 the detriment of patch area. As regards to potential predictors of ϕ , we found some degree of
260 multicollinearity among the presence of road-verges, patch area, patch permanence, and patch
261 isolation (Table A2, Appendix), suggesting an association of road verge habitats to larger, more
262 stable, and less isolated patches. Because the presence of road-verges was considered to
263 provide a reliable descriptor of local environmental variations directly linked to human land-use
264 activity, this variable was carried out to the CJS modelling procedure. We also retained the
265 covariate distance to agricultural fields instead of the distance to urban areas (positively cross-
266 correlated; Table A2, Appendix A), as agricultural land-use was predominant, and potentially
267 most relevant land-use in the study area. Sex, presence of water, and detection of water voles
268 were only weakly correlated to other predictors, and were therefore also retained in the
269 analyses (Table A2, Appendix A).

270 We then evaluated a set of CJS models translating different combinations of hypotheses
271 regarding uncorrelated factors possibly affecting p and ϕ (Lebreton et al., 1992). Given the
272 relatively large number of possible models considering all possible combinations of main effects
273 in both p and ϕ (i.e., 512 models), we combined the most plausible submodels found separately
274 for each parameter (Bromaghin et al., 2013). Plausible submodels for both p and ϕ were
275 identified in each case, by first building and ranking all possible submodels defined by all
276 possible additive combinations of main effects (three in the case of p and six in the case of ϕ).
277 This resulted in eight submodels describing p , and 64 submodels describing ϕ . For the final set of
278 candidate models to be evaluated, we retained in each case the set of submodels with a
279 difference of Akaike's Information Criteria corrected for small sample size (ΔAIC_c) smaller than 2

280 relative to the respective top-ranked submodel, regarded as the most supported (Burnham and
281 Anderson, 2002). This plausible combination approach reduced the number of candidate models
282 to about 3% (n=15) in relation to the all possible combinations strategy (see Results), thus
283 reducing the potential incidence of spurious results (Doherty et al., 2012). Finally, from the most
284 supported models (i.e. $\Delta AIC_c < 2$ relative to the top-ranking model) within this final set of 15
285 candidate models, we discarded those including uninformative covariates [i.e. with 85%
286 confidence intervals of effect size estimates including zero; Arnold (2010)]. This resulted in the
287 selection of one single best model, from which we estimated the predicted monthly p and ϕ
288 relative to each informative covariate included in that model.

289 3. Results

290 The amount of suitable habitat increased from the early wet season (EWS; 36 ha) to the late wet
291 season (LWS; 46 ha), and declined both in the early dry season (EDS; 41 ha), and in the late dry
292 season (LDS; 29 ha) (Fig. 1). The percentage of occupied patches was of 45% (n= 131) in EWS,
293 51% (n=138) in LWS, 61% (n= 137) in EDS, and 54% (n=149) in LDS (Fig. 1).

294 We collected a total of 2 711 faecal samples (mean \pm SE per season = 678 \pm 54), of which 48.4%
295 (n=1312; 328 \pm 24 per season) were selected for DNA extraction and genotyping. Average
296 genotyping success rate was 33.9% (444 samples), with the highest values obtained for samples
297 collected in EDS (42.4%, n=153), followed by EWS (38.4%, n=140), LWS (26.8%, n=87), and LDS
298 (24.4%, n=64). Overall genotyping errors were low (dropout rate: 0.68-2.8%; false allele rate: 0-
299 0.18%; Table A1, Appendix A), with higher genotyping errors recorded in the seasons with lower
300 genotyping success. From the randomly selected 23% of samples that failed amplification (i.e.
301 163 out of 704 samples), about 86% were identified as Cabrera voles, while the others belonged
302 to other rodent species (7.3%) or were contaminated with human or ungulate DNA (6.7%).
303 Another 164 samples were also contaminated despite being successfully amplified. The 444
304 samples that were successfully amplified and were not contaminated were assigned to a total of
305 307 individuals, with 81 (EWS), 77 (LWS), 122 (EDS), and 64 (LDS) individuals per season. From
306 the 137 recaptures identified across all surveys, 120 (87.6%) were in the same habitat patch of
307 the previous (re)capture. The mean seasonal sex-ratio was even (M:F = 1.08), with no marked
308 seasonal changes (between 1.03 and 1.13).

309 The likelihood-ratio tests indicated that ECM was more supported than TRIM for estimating
310 abundances in all seasons ($p > 0.16$). Abundances estimated by ECM (range: 116-353) were very

311 similar to those obtained using M_h -Chao (range: 125-370). There was an over 4-fold increase in
312 vole abundance from EWS to LWS, with little change through to the EDS, followed by about a 2-
313 fold decrease until the LDS (Fig. 2). Except for EWS, abundance estimates derived from both
314 ECM and M_h -Chao had relatively wide confidence intervals.

315 The PC model selection approach resulted in the retention of five recapture probability
316 submodels and three survival probability submodels (Table 2). After applying the model
317 selection procedure on the final set of 15 plausible models and excluding models with
318 uninformative covariates, the most supported model retained no covariates affecting recapture
319 and included four covariates affecting survival (Table 3) Monthly recapture probability was
320 estimated as 0.54 (0.20-0.85 CI 95%), while monthly survival was estimated as 0.52 (0.39-0.65 CI
321 95%), being 1.5-times higher in males than in females (Fig. 3A). Survival was also affected by
322 habitat conditions, being 2-times higher on road-verges than elsewhere (Fig. 3B), 1.5-times
323 higher in the presence of (or bordered by) water-bodies (Fig. 3C), and 2-times higher 300m
324 away from agricultural areas (Fig. 3D).

325 **4. Discussion**

326 We demonstrated for the first time the usefulness of large-scale genetic non-invasive sampling
327 combined with capture-mark-recapture methods to estimate and identify the factors affecting
328 small mammal demographic parameters and infer their population dynamics. Using the near-
329 threatened Cabrera vole in Mediterranean farmland, we showed that our approach provides
330 key information to improve conservation planning of elusive small mammals, especially those
331 threatened by human activities and that are difficult to sample using traditional methods (Cheng
332 et al., 2017).

333 Our approach showed that Cabrera vole abundance varied greatly across an annual cycle,
334 confirming a large increase in vole numbers along the wet season (67%), and a substantial
335 decline through the dry season (44%), as reported elsewhere from live-trapping data collected
336 at more confined scales (Fernández-Salvador et al., 2005b; Rosário, 2012). These changes
337 seemed largely unrelated to seasonal genotyping success, which lowered by the end of the wet
338 season and by the end of the dry season, likely due to increased DNA degradation under higher
339 rainfall and sunlight exposure, respectively (Santini et al., 2007). Seasonal variation in vole
340 numbers was consistent with the described breeding period for this species, which suggests a
341 lower activity of individuals during the dry season, when reproduction may even cease

342 completely (Fernández-Salvador et al., 2005b; Pita et al., 2006). Comparable seasonal
343 fluctuations have also been found in other rodent species inhabiting highly seasonal
344 Mediterranean environments (Cohen-Shlagman et al., 1984; Gomez et al., 2016), as well as in
345 other Iberian endemic herbivores, like the Iberian rabbit (*Oryctolagus cuniculus algirus*)
346 (Gonçalves et al., 2002). This pattern is thought to be related to variations in habitat and food
347 availability, which in our study area are generally reduced during the dry season (Pita et al.,
348 2014). This was reflected in our data by the 37% decrease in habitat availability from the end of
349 the wet season to the end of the dry season, in accordance with the natural seasonal variation
350 in climate, and was also observed for other voles in intensively managed Mediterranean
351 farmlands [e.g. common vole *Microtus arvalis*; Rodríguez-Pastor et al. (2016)]. Pedigree and
352 sibship analyses indicated that our gNIS was able to detect animals from different generations
353 within each season (Ferreira et al., unpublished data), suggesting no serious bias towards any
354 particular age-class of the population. While we acknowledge that the two-month duration of
355 our seasonal surveys warrants some caution regarding abundance estimates obtained within a
356 closed CMR framework, these estimates likely describe the broad patterns of seasonal
357 population change at the surveyed landscape and its surroundings (Boulanger and McLellan,
358 2001). Therefore, our study suggests that estimating vole population size based on gNIS and
359 CMR modelling allows for the drawing of inferences on abundance variation across time.

360 Estimates of apparent survival of Cabrera voles based on gNIS in Mediterranean farmland were
361 relatively low (0.39-0.65 CI 95%), though still within the range usually observed in other semi-
362 fossorial *Microtus* species living in agricultural landscapes across different geographical regions.
363 For instance, the survival of *M. agrestis* in field margins in fragmented farmland from northeast
364 Scotland ranged between 0.42 and 0.69 (Renwick and Lambin, 2011), while that of *M. arvalis* in
365 agricultural landscapes from central western France varied between 0.22-0.69 (Bonnet et al.,
366 2013). Similar survival rates (0.25-0.64) were also reported for *M. pennsylvanicus* in forage crops
367 from Illinois (Getz et al., 2007). Surprisingly, despite the presumed lower habitat quality in our
368 study area during the dry season, and the observed temporal variation in Cabrera vole
369 abundance, survival was virtually constant across time. Variation in population abundance,
370 despite constant survival rates, suggests that population recruitment (newborn and immigrants)
371 increased during the wet season (when abundance increased), and decreased during the dry
372 season, probably until it became insufficient to compensate the low survival rates, thereby
373 resulting in pronounced decreases in vole numbers. Although we did not assess recruitment

374 explicitly and our gNIS approach inherently prevents the identification of individuals' age, the
375 observed seasonal variations in abundance agrees with the monthly fluctuations in recruitment
376 rates observed in Cabrera vole populations from other Mediterranean areas (Fernández-
377 Salvador et al., 2005b), as well as in other vole species from Mediterranean-like climates
378 (Cockburn and Lidicker, 1983; Cohen-Shlagman et al., 1984; Paradis and Guédon, 1993).

379 Survival was however not constant across space. Voles detected closest to farmed areas showed
380 a 53% lower survival probability than voles captured furthest away, suggesting a negative
381 impact of agriculture management on voles. Agriculture activities in this area are highly
382 dynamic, involving for instance the conversion of fallow areas to farmed areas, resulting in the
383 destruction of habitat-patches. While the putative negative effects of agricultural intensification
384 on Cabrera vole populations have been widely suggested (Fernández-Salvador et al., 2005b;
385 2005a; Pita et al., 2006), our study provides the first quantitative evidence that the distance of
386 grass-rich fragments occupied by voles to unsuitable farmed habitat affects individual survival
387 probability. Further studies analysing fecundity variation across space and time are however
388 needed to fully understand how such effect impacts overall population persistence in farmland
389 areas. According to our initial predictions, voles captured in road-verge habitats showed higher
390 survival probabilities than voles captured elsewhere. While road-verge habitats can be viewed
391 as suboptimal for voles (Santos et al., 2007) it is likely that in intensively-used Mediterranean
392 farmland, these habitats provide refuges for the species, as their vegetation is often left
393 undisturbed for relatively long-time periods compared to surrounding fields (Ruiz-Capillas et al.,
394 2013). This is supported by the association of road-verge habitats to larger and more stable
395 habitats in our study area, as well as by the low traffic volume associated to the roads. The value
396 of road-verges as refuges for small mammals in farmlands has also been noted in other
397 Mediterranean environments with varying levels of land management (Ascensão et al., 2012;
398 Ruiz-Capillas et al., 2013; Sabino-Marques and Mira, 2011), being generally attributed to
399 increased habitat quality, and/or to predation release effects, in the case of roads with more
400 intensive traffic (Rytwinski and Fahrig, 2007).

401 The positive effects of the presence of permanent water-bodies within or bordering habitat-
402 patches on voles' survival was in accordance to the prediction that damper habitats provide
403 better resources for Cabrera voles (Pita et al., 2011). This may be associated with the presence
404 of fresh green vegetation providing high-quality food and shelter across the dry landscape
405 (Santos et al., 2007). While the presumed superior competitor water vole also prefers habitats

406 with dense and wet vegetation, and may affect Cabrera voles' occupancy patterns (Pita et al.,
407 2016), we found no evidence for inter-specific effects on Cabrera voles' survival. It is also
408 interesting to note that, despite their influence in shaping Cabrera voles' occupancy dynamics
409 and abundance (Pita et al., 2007), patch size and isolation were poor predictors of individual
410 survival. This has also been reported in other small mammal species, for which habitat quality
411 was also a better surrogate for survival than patch size (Mortelliti et al., 2014).

412 Besides environmental factors, and contrary to our expectations based on the predominantly
413 monogamous mating systems of Cabrera voles (Pita et al., 2014), apparent survival was also
414 related to sex, with support for higher survival in males. In the case of monogamous species that
415 are not sexually dimorphic and with both males and females sharing parental care, survival
416 generally tends to be similar across sexes or in some cases male-biased, thus highlighting an
417 interesting exception to the general rule of male-biased mortality, typical in polygynous
418 mammals (Clutton-Brock and Isvaran, 2007). Higher male survival in monogamous species may
419 result from reduced competition among males compared to polygynous animals, and the fact
420 that even in the presence of parental care, the costs of raising off-spring are likely to be endured
421 primarily by lactating females (Clutton-Brock and Isvaran, 2007; Lukas and Clutton-Brock, 2013).
422 Testing this hypothesis explicitly would require information on individuals' reproductive
423 condition and age, which are not obtainable from gNIS approaches such as ours. However,
424 because male-biased survival did not result in any male-biased sex-ratios, and apparent survival
425 is the product of true survival and site fidelity (Sandercock, 2006), we cannot rule out the
426 possibility that the observed male-biased survival could also reflect a female-biased dispersal (or
427 permanent emigration), which is also common in monogamous species (Mabry et al., 2013).
428 Clearly, more studies are needed to fully elucidate on possible male-biased survival and/or
429 female-biased dispersal in the Cabrera vole. This warrants the recommendation that future
430 gNIS-based studies aiming to assess survival of elusive species, should combine other methods
431 providing information on individual reproductive status and age, so as to improve inferences on
432 their population dynamics.

433

434 **5. Conclusions**

435 Overall, our study provides empirical evidence that gNIS is a useful tool to monitor small
436 mammal population parameters, and to identify management actions that may prove necessary

437 to maintain their populations. Regarding the Cabrera vole, our results support the idea that
438 conservation measures aimed to increase its survival in Mediterranean farmland, should
439 promote low intensity agricultural management near occupied patches (encompassing longer
440 fallow periods, low-disturbed margins, and high density of permanent water-bodies), in order to
441 allow the continued existence of suitable habitats for the species, particularly during the
442 stressful dry-season, when habitat availability is lower. We consider that our approach may be
443 applied to other elusive small or medium mammals requiring conservation action, but for which
444 informative demographic data across large spatial and temporal scales are still lacking.

445

446 **Author contributions**

447 Conception (PCA, PB, AM, RP); Design (XL, PCA, PB, AM, RP); Data Collection (APF, IL, CF, JP,
448 HSM, SB); Data Analysis (APF, CF, RP); Writing (APF, RP); Revision (APF, CF, JP, PCA, HSM, SB, XL,
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469

470 **References**

471 Arnold, T.W., 2010. Uninformative parameters and model selection using akaike's in- formation
472 criterion. *J. Wildl. Manag.* 74, 1175–1178. <https://doi.org/10.1111/j.1937-2817.2010.tb01236.x>.

473 Ascensão, F., Clevenger, A.P., Grilo, C., Filipe, J., Santos-Reis, M., 2012. Highway verges as
474 habitat providers for small mammals in agrosilvopastoral environments. *Biodivers. Conserv.* 21,
475 3681–3697. <https://doi.org/10.1007/s10531-012-0390-3>.

476 Baillargeon, S., Rivest, L.-P., 2007. Rcapture: loglinear models for capture-recapture in R. *J. Stat.*
477 *Softw.* 19. <https://doi.org/10.18637/jss.v019.i05>.

478 Barbosa, S., Paupério, J., Searle, J.B., Alves, P.C., 2013. Genetic identification of Iberian rodent
479 species using both mitochondrial and nuclear loci: application to noninvasive sampling. *Mol.*
480 *Ecol. Resour.* 13, 43–56. <https://doi.org/10.1111/1755-0998.12024>.

481 Beja-Pereira, A., Oliveira, R., Alves, P.C., Schwartz, M.K., Luikart, G., 2009. Advancing ecological
482 understandings through technological transformations in noninvasive ge- netics. *Mol. Ecol.*
483 *Resour.* 9, 1279–1301. <https://doi.org/10.1111/j.1755-0998.2009.02699.x>.

484 Bonnet, T., Crespin, L., Pinot, A., Bruneteau, L., Bretagnolle, V., Gauffre, B., 2013. How the
485 common vole copes with modern farming: insights from a capture— mark—recapture
486 experiment. *Agric. Ecosyst. Environ.* 177, 21–27. <https://doi.org/10.1016/j.agee.2013.05.005>.

487 Boulanger, J., McLellan, B., 2001. Closure violation in DNA-based mark-recapture esti- mation of
488 grizzly bear populations. *Can. J. Zool.* 79, 642–651. <https://doi.org/10.1139/z01-020>.

489 Bromaghin, J.F., McDonald, T.L., Amstrup, S.C., 2013. Plausible combinations: an im-
490 proved method to evaluate the covariate structure of Cormack-Jolly-Seber mark-re-
491 capture models. *Open J. Ecol.* 2013, 11–22. <https://doi.org/10.4236/oje.2013.31002>.

492 Brøseth, H., Flagstad, Ø., Wårdig, C., Johansson, M., Ellegren, H., 2010. Large-scale noninvasive
493 genetic monitoring of wolverines using scats reveals density dependent adult survival. *Biol.*
494 *Conserv.* 143, 113–120. <https://doi.org/10.1016/j.biocon.2009.09.012>.

495 Burgar, J.M., Stewart, F.E.C., Volpe, J.P., Fisher, J.T., Burton, A.C., 2018. Estimating density for
496 species conservation: comparing camera trap spatial count models to genetic spatial capture-
497 recapture models. *Glob. Ecol. Conserv.* 15, e00411.
498 <https://doi.org/10.1016/j.gecco.2018.e00411>.

499 Burnham, K.P., Anderson, D.R., 2002. *Model Selection and Multimodel Inference: A Practical*
500 *Information-Theoretic Approach*, 2nd ed. Springer-Verlag.

501 Chao, A., 1989. Estimating population size for sparse data in capture-recapture experiments.
502 *Biometrics* 45, 427–438. <https://doi.org/10.2307/2531487>.

503 Cheng, E., Hodges, K.E., Sollmann, R., Mills, L.S., 2017. Genetic sampling for estimating density
504 of common species. *Ecol. Evol.* 7, 6210–6219. <https://doi.org/10.1002/ece3>.

505 Clutton-Brock, T.H., Isvaran, K., 2007. Sex differences in ageing in natural populations of
506 vertebrates. *Proc. R. Soc. Lond. B Biol. Sci.* 274, 3097–3104.
507 <https://doi.org/10.1098/rspb.2007.1138>.

508 Cockburn, A., Lidicker Jr., W.Z., 1983. Microhabitat heterogeneity and population ecology of an
509 herbivorous rodent, *Microtus californicus*. *Oecologia* 59, 167–177.
510 <https://doi.org/10.1007/BF00378834>.

511 Cohen-Shlagman, L., Yom-Tov, Y., Hellwing, S., 1984. The biology of the Levant vole,
512 *Microtus guentheri* in Israel. I. Population dynamics in the field. *Z. Säugetierk* 135–147.
513 http://www.zobodat.at/pdf/Zeitschrift-Saeugetierkunde_49_0135-0147.pdf.

514 Costa, V., Rosenbom, S., Monteiro, R., O'Rourke, S.M., Beja-Pereira, A., 2017. Improving DNA
515 quality extracted from fecal samples—a method to improve DNA yield. *Eur. J. Wildl. Res.* 63, 3.
516 <https://doi.org/10.1007/s10344-016-1058-1>.

517 De Bondi, N., White, J.G., Stevens, M., Cooke, R., 2010. A comparison of the effectiveness of
518 camera trapping and live trapping for sampling terrestrial small-mammal com- munities. *Wildl.*
519 *Res.* 37, 456–465. <https://doi.org/10.1071/WR10046>.

520 DeMay, S.M., Becker, P.A., Rachlow, J.L., Waits, L.P., 2017. Genetic monitoring of an endangered
521 species recovery: demographic and genetic trends for reintroduced pygmy rabbits (*Brachylagus*
522 *idahoensis*). *J. Mammal.* 98, 350–364. <https://doi.org/10.1093/jmammal/gyw197>.

523 Doherty, P.F., White, G.C., Burnham, K.P., 2012. Comparison of model building and se- lection
524 strategies. *J. Ornithol.* 152, 317–323. <https://doi.org/10.1007/s10336-010- 0598-5>.

525 Fernández-Salvador, R., García-Perea, R., Ventura, J., 2005a. Effect of climatic fluctua- tions on
526 body mass of a Mediterranean vole, *Microtus cabrerae*. *Mamm. Biol.* 70, 73–83.
527 <https://doi.org/10.1016/j.mambio.2004.06.002>.

528 Fernández-Salvador, R., Ventura, J., García-Perea, R., 2005b. Breeding patterns and de-
529 mography of a population of the Cabrera vole, *Microtus cabrerae*. *Anim. Biol.* 55, 147–161.
530 <https://doi.org/10.1163/1570756053993497>.

531 Ferreira, M., Beja, P., 2013. Mediterranean amphibians and the loss of temporary ponds: are
532 there alternative breeding habitats? *Biol. Conserv.* 165, 179–186.
533 <https://doi.org/10.1016/j.biocon.2013.05.029>.

534 Ferreira, C.M., Sabino-Marques, H., Barbosa, S., Costa, P., Encarnação, C., Alpizar-Jara, R., Pita,
535 R., Beja, P., Mira, A., Searle, J.B., Paupério, J., Alves, P.C., 2018. Genetic non-invasive sampling
536 (gNIS) as a cost-effective tool for monitoring elusive small mammals. *Eur. J. Wildl. Res.* 1–44.
537 <https://doi.org/10.1007/s10344-018-1188-8>.

538 Getz, L.L., Oli, M.K., Hofmann, J.E., McGuire, B., 2007. Vole population dynamics: factors
539 affecting peak densities and amplitudes of annual population fluctuations of *Microtus*
540 *pennsylvanicus*. *Acta Theriol.* 52, 159–170. <https://doi.org/10.1007/BF03194211>.

541 Gillet, F., 2016. Genetic monitoring of the endangered Pyrenean desman (*Galemys pyrenaicus*)
542 in the Aude River, France. *Belg. J. Zool.* 146, 44–52.

543 Gimenez, O., Lebreton, J.-D., Choquet, R., Pradel, R., 2018. R2ucare: an r package to perform
544 goodness-of-fit tests for capture–recapture models. *Methods Ecol. Evol.* 9, 1749–1754.
545 <https://doi.org/10.1111/2041-210X.13014>.

546 Glen, A.S., Cockburn, S., Nichols, M., Ekanayake, J., Warburton, B., 2013. Optimising camera
547 traps for monitoring small mammals. *PLoS One* 8, e67940.
548 <https://doi.org/10.1371/journal.pone.0067940>.

549 Gomez, M.D., Serafini, V., Coda, J., Priotto, J., 2016. Demographic dynamics of *Akodon azarae*
550 (*Cricetidae*: *Sigmodontinae*) in linear habitats of agricultural landscapes of central Argentina.
551 *Stud. Neotropical Fauna Environ.* 51, 10–18. <https://doi.org/10.1080/01650521.2015.1137167>.

552 Gonçalves, H., Alves, P.C., Rocha, A., 2002. Seasonal variation in the reproductive ac-
553 tivity of the wild rabbit (*Oryctolagus cuniculus algirus*) in a Mediterranean ecosystem. *Wildl. Res.* 29, 165–
554 173. <https://doi.org/10.1071/WR00048>.

555 Graham, M.H., 2003. Confronting multicollinearity in ecological multiple regression. *Ecology* 84,
556 2809–2815. <https://doi.org/10.1890/02-3114>.

557 Johnson, P.C.D., Haydon, D.T., 2007. Software for quantifying and simulating micro-
558 genotyping error. *Bioinf. Biol. Insights* 1, 71–75. <https://doi.org/10.4137/BBI.S373>.

559 Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton, S., Cooper,
560 A., Markowitz, S., Duran, C., Thierer, T., Ashton, B., Meintjes, P., Drummond, A., 2012. Geneious
561 Basic: an integrated and extendable desktop software platform for the organization and analysis
562 of sequence data. *Bioinformatics* 28, 1647–1649.
563 <https://doi.org/10.1093/bioinformatics/bts199>.

564 Laake, J.L., 2013. RMark: An R Interface for Analysis of Capture-Recapture Data With MARK.
565 Alaska Fish. Sci. Cent., NOAA, Natl. Mar. Fish. Serv., Seattle, WA.

566 Lampa, S., Mihoub, J.-B., Gruber, B., Klenke, R., Henle, K., 2015. Non-invasive genetic mark-
567 recapture as a means to study population sizes and marking behaviour of the elusive eurasian
568 otter (*Lutra lutra*). *PLoS One* 10, e0125684. <https://doi.org/10.1371/journal.pone.0125684>.

569 Lebreton, J.D., Burnham, K.P., Clobert, J., Anderson, D.R., 1992. Modeling survival and testing
570 biological hypotheses using marked animals: a unified approach with case studies. *Ecol.*
571 *Monogr.* 62, 67–118.

572 Lukas, D., Clutton-Brock, T.H., 2013. The evolution of social monogamy in mammals. *Science*
573 341, 526–530. <https://doi.org/10.1126/science.1238677>.

574 Mabry, K.E., Shelley, E.L., Davis, K.E., Blumstein, D.T., Van Vuren, D.H., 2013. Social mating
575 system and sex-biased dispersal in mammals and birds: a phylogenetic analysis. *PLoS One* 8,
576 e57980. <https://doi.org/10.1371/journal.pone.0057980>.

577 Marucco, F., Boitani, L., Pletscher, D.H., Schwartz, M.K., 2011. Bridging the gaps between non-
578 invasive genetic sampling and population parameter estimation. *Eur. J. Wildl. Res.* 57, 1–13.
579 <https://doi.org/10.1007/s10344-010-0477-7>.

580 Marucco, F., Vucetich, L.M., Peterson, R.O., Adams, J.R., Vucetich, J.A., 2012. Evaluating the
581 efficacy of non-invasive genetic methods and estimating wolf survival during a ten-year period.
582 *Conserv. Genet.* 13, 1611–1622. <https://doi.org/10.1007/s10592-012-0412-4>.

583 Maudet, C., Luikart, G., Dubray, D., von Hardenberg, A., Taberlet, P., 2004. Low geno- typing
584 error rates in wild ungulate faeces sampled in winter. *Mol. Ecol. Notes* 4, 772–775.
585 <https://doi.org/10.1111/j.1471-8286.2004.00787.x>.

586 McCrea, R.S., Morgan, B.J.T., 2015. Analysis of capture-recapture data. Chapman & Hall/ CRC,
587 Canterbury, UK.

588 Mestre, F., Pita, R., Paupério, J., Martins, F.M.S., Alves, P.C., Mira, A., Beja, P., 2015. Combining
589 distribution modelling and non-invasive genetics to improve range shift forecasting. *Ecol.*
590 *Model.* 297, 171–179. <https://doi.org/10.1016/j.ecolmodel.2014.11.018>.

591 Miller, C.R., Joyce, P., Waits, L.P., 2005. A new method for estimating the size of small
592 populations from genetic mark-recapture data. *Mol. Ecol.* 14, 1991–2005. <https://doi.org/10.1111/j.1365-294X.2005.02577.x>.

594 Mondol, S., Ullas Karanth, K., Samba Kumar, N., Gopaldaswamy, A.M., Andheria, A.,
595 Ramakrishnan, U., 2009. Evaluation of non-invasive genetic sampling methods for estimating

596 tiger population size. *Biol. Conserv.* 142, 2350–2360. [https://doi.org/10.](https://doi.org/10.1016/j.biocon.2009.05.014)
597 1016/j.biocon.2009.05.014.

598 Mortelliti, A., Sozio, G., Driscoll, D.A., Bani, L., Boitani, L., Lindenmayer, D.B., 2014. Population
599 and individual-scale responses to patch size, isolation and quality in the hazel dormouse.
600 *Ecosphere* 5, 1–21. <https://doi.org/10.1890/ES14-00115.1>.

601 Paradis, E., Guédon, G., 1993. Demography of a Mediterranean microtine: the Mediterranean
602 pine vole, *Microtus duodecimcostatus*. *Oecologia* 95, 47–53.
603 <https://doi.org/10.1007/BF00649505>.

604 Pennell, M.W., Stansbury, C.R., Waits, L.P., Miller, C.R., 2013. Capwire: a R package for
605 estimating population census size from non-invasive genetic sampling. *Mol. Ecol. Resour.* 13,
606 154–157. <https://doi.org/10.1111/1755-0998.12019>.

607 Peralta, D., Leitão, I., Ferreira, A., Mira, A., Beja, P., Pita, R., 2016. Factors affecting southern
608 water vole (*Arvicola sapidus*) detection and occupancy probabilities in Mediterranean farmland.
609 *Mamm. Biol.* 81, 123–129. <https://doi.org/10.1016/j.mambio.2015.10.006>.

610 Petit, E.J., Valiere, N., 2006. Estimating population size with noninvasive capture-mark-
611 recapture data. *Conserv. Biol.* 20, 1062–1073. [https://doi.org/10.1111/j.1523-](https://doi.org/10.1111/j.1523-1739.2006.00417.x)
612 1739.2006.00417.x.

613 Pita, R., Mira, A., Beja, P., 2006. Conserving the Cabrera vole, *Microtus cabrerae*, in in-
614 tensively used Mediterranean landscapes. *Agric. Ecosyst. Environ.* 115, 1–5.
615 <https://doi.org/10.1016/j.agee.2005.12.002>.

616 Pita, R., Beja, P., Mira, A., 2007. Spatial population structure of the Cabrera vole in
617 Mediterranean farmland: the relative role of patch and matrix effects. *Biol. Conserv.* 134, 383–
618 392. <https://doi.org/10.1016/j.biocon.2006.08.026>.

619 Pita, R., Mira, A., Moreira, F., Morgado, R., Beja, P., 2009. Influence of landscape char-
620 acteristics on carnivore diversity and abundance in Mediterranean farmland. *Agric. Ecosyst. Environ.* 132,
621 57–65. <https://doi.org/10.1016/j.agee.2009.02.008>.

622 Pita, R., Mira, A., Beja, P., 2010. Spatial segregation of two vole species (*Arvicola sapidus* and
623 *Microtus cabreræ*) within habitat patches in a highly fragmented farmland landscape. *Eur. J.*
624 *Wildl. Res.* 56, 651–662. <https://doi.org/10.1007/s10344-009-0360-6>.

625 Pita, R., Mira, A., Beja, P., 2011. Assessing habitat differentiation between coexisting species:
626 the role of spatial scale. *Acta Oecol.* 37, 124–132. <https://doi.org/10.1016/j.actao.2011.01.006>.

627 Pita, R., Mira, A., Beja, P., 2014. *Microtus cabreræ* (Rodentia: Cricetidae). *Mamm. Species* 912,
628 48–70. <https://doi.org/10.1644/912.1>.

629 Pita, R., Lambin, X., Mira, A., Beja, P., 2016. Hierarchical spatial segregation of two
630 Mediterranean vole species: the role of patch-network structure and matrix composition.
631 *Oecologia* 182, 253–263. <https://doi.org/10.1007/s00442-016-3653-y>.

632 QGIS Development Team, 2016. QGIS Geographic Information System. Open Source Geospatial
633 Foundation Project. <http://qgis.osgeo.org>.

634 R Core Team, 2018. R: A language and environment for statistical computing. R Foundation for
635 Statistical Computing, Vienna, Austria. <https://www.R-project.org>.

636 Renwick, A.R., Lambin, X., 2011. Abundance thresholds and the underlying ecological processes:
637 field voles *Microtus agrestis* in a fragmented landscape. *Agric. Ecosyst. Environ.* 144, 364–369.
638 <https://doi.org/10.1016/j.agee.2011.10.006>.

639 Rivas-Martínez, S., 1981. The vegetation of bioclimatic stages of Iberian Peninsula. *Anal. Jard.*
640 *Bot. Madr.* 37, 251–268.

641 Rodríguez-Pastor, R., Luque-Larena, J.J., Lambin, X., Mougeot, F., 2016. “Living on the edge”: the
642 role of field margins for common vole (*Microtus arvalis*) populations in recently colonised
643 Mediterranean farmland. *Agric. Ecosyst. Environ.* 231, 206–217.
644 <https://doi.org/10.1016/j.agee.2016.06.041>.

645 Rosário, I., 2012. Towards a Conservation Strategy for an Endangered Rodent, the Cabrera Vole
646 (*Microtus cabreræ* Thomas). *Insights From Ecological Data*. Universidade de Lisboa, Lisbon,
647 Portugal.

648 Ruiz-Capillas, P., Mata, C., Malo, J.E., 2013. Road verges are refuges for small mammal
649 populations in extensively managed Mediterranean landscapes. *Biol. Conserv.* 158, 223–229.
650 <https://doi.org/10.1016/j.biocon.2012.09.025>.

651 Rytwinski, T., Fahrig, L., 2007. Effect of road density on abundance of white-footed mice. *Landsc.*
652 *Ecol.* 22, 1501–1512. <https://doi.org/10.1007/s10980-007-9134-2>.

653 Sabino-Marques, H., Mira, A., 2011. Living on the verge: are roads a more suitable refuge for
654 small mammals than streams in Mediterranean pastureland? *Ecol. Res.* 26, 277–287.
655 <https://doi.org/10.1007/s11284-010-0781-4>.

656 Sabino-Marques, H., Ferreira, C.M., Paupério, J., Costa, P., Barbosa, S., Encarnação, C., Alpizar-
657 Jara, R., Alves, P.C., Searle, J.B., Mira, A., Beja, P., Pita, R., 2018. Combining genetic non-invasive
658 sampling with spatially explicit capture-recapture models for density estimation of a patchily
659 distributed small mammal. *Eur. J. Wildl. Res.* 64. <https://doi.org/10.1007/s10344-018-1206-x>.

660 Sandercock, B.K., 2006. Estimation of demographic parameters from live-encounter data: a
661 summary review. *J. Wildl. Manag.* 70, 1504–1520. [https://doi.org/10.2193/0022-
662 541X\(2006\)70\[1504:EODPFL\]2.0.CO;2](https://doi.org/10.2193/0022-541X(2006)70[1504:EODPFL]2.0.CO;2).

663 Santini, A., Lucchini, V., Fabbri, E., Randi, E., 2007. Ageing and environmental factors affect PCR
664 success in wolf (*Canis lupus*) excremental DNA samples. *Mol. Ecol. Notes* 7, 955–961.
665 <https://doi.org/10.1111/j.1471-8286.2007.01829.x>.

666 Santos, S.M., Mathias, M., Mira, A., Simões, M., 2007. Vegetation structure and compo- sition of
667 road verge and meadow sites colonized by Cabrera vole (*Microtus cabrae* Thomas). *Pol. J.*
668 *Ecol.* 55, 481.

669 Smallwood, K.S., Schonewald, C., 1998. Study design and interpretation of mammalian carnivore
670 density estimates. *Oecologia* 113, 474–491. <https://doi.org/10.1007/s004420050400>.

671 Tscharrntke, T., Clough, Y., Wanger, T.C., Jackson, L., Motzke, I., Perfecto, I., Vandermeer, J.,
672 Whitbread, A., 2012. Global food security, biodiversity conservation and the fu- ture of
673 agricultural intensification. *Biol. Conserv.* 151, 53–59.
674 <https://doi.org/10.1016/j.biocon.2012.01.068>.

- 675 Valiere, N., 2002. gimlet: a computer program for analysing genetic individual identification
676 data. *Mol. Ecol. Notes* 2, 377–379. <https://doi.org/10.1046/j.1471-8286.2002.00228.x-i2>.
- 677 White, G.C., Burnham, K.P., 1999. Program MARK: survival estimation from populations of
678 marked animals. *Bird Study* 46, S120–S139. <https://doi.org/10.1080/00063659909477239>.
- 679 Williams, B.K., Nichols, J.D., Conroy, M.J., 2002. *Analysis and Management of Animal*
680 *Populations*. Academic Press.
- 681 Zielinski, W.J., Schlexer, F.V., George, T.L., Pilgrim, K.L., Schwartz, M.K., 2013. Estimating
682 abundance and survival in the endangered point arena mountain beaver using noninvasive
683 genetic methods. *Northwest Sci.* 87, 126–139. <https://doi.org/10.3955/046.087.0205>.

Accepted Article

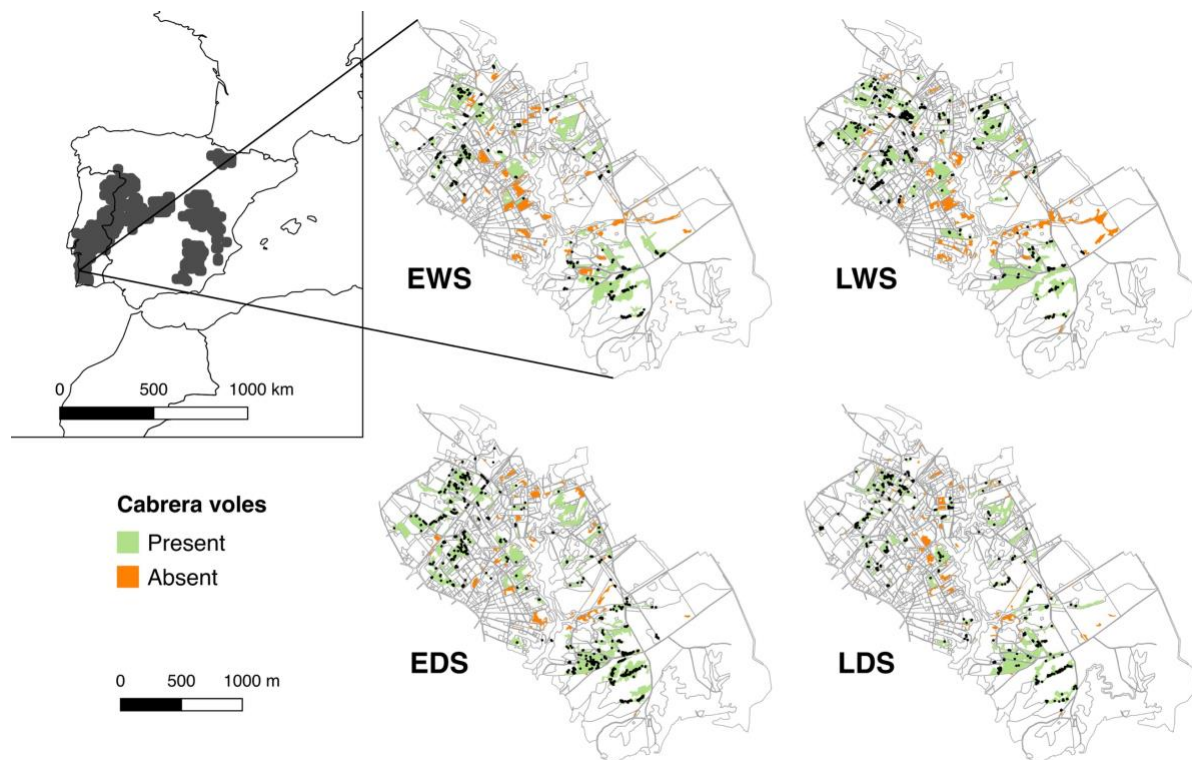


Fig. 1 – On the upper left, Cabrera vole distribution (in dark-grey, adapted from Mestre et al., 2015) and study area location. On the right, four seasonal surveys made in a 461.8 ha area within the coastal plateau of SW Portugal. EWS - early wet season, November and December 2013; LWS - late wet season, February and March 2014; EDS - early dry season, May and June 2014; LDS - late dry season, September and October 2014. Coloured polygons represent identified suitable habitat-patches for voles. Black dots represent fresh-faeces samples collected for genetic analyses.

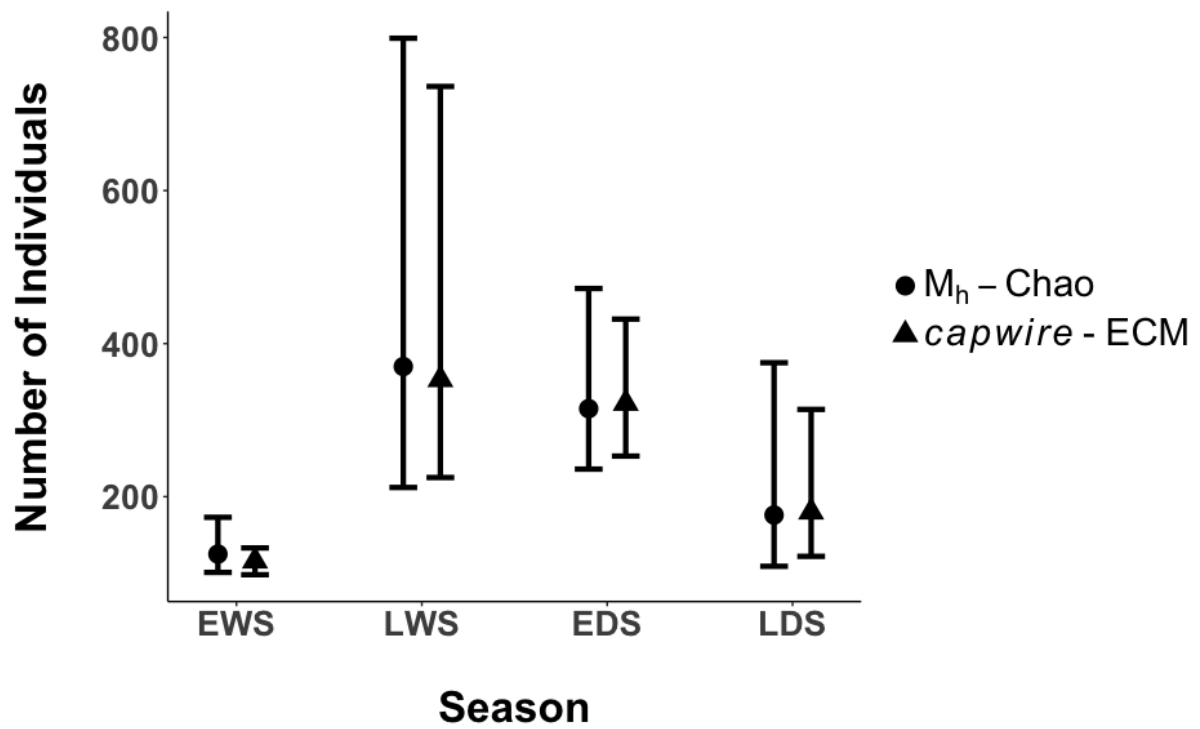


Fig. 2 – Seasonal population abundance of Cabrera voles in a Mediterranean farmland, based on genetic non-invasive sample, estimated using the M_h -Chao estimator (Chao) and the *capwire*'s equal capture probability model (ECM). EWS - early wet season, November and December 2013; LWS - late wet season, February and March 2014; EDS - early dry season, May and June 2014; LDS - late dry season, September and October 2014. Bars indicate 95% confidence intervals.

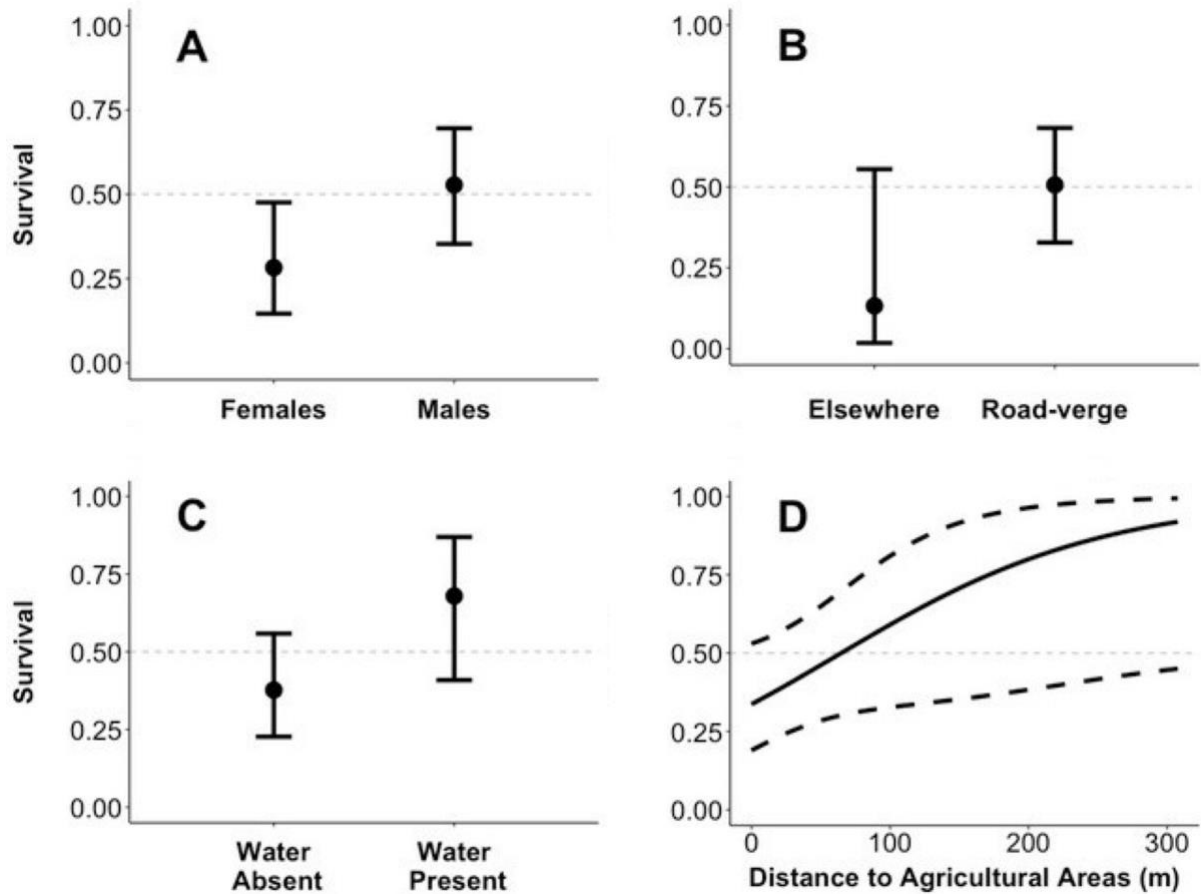


Fig. 3 – Apparent survival of Cabrera vole relative to sex (A); road verges (B); presence of water (C); and distance to agricultural areas (D), based in genetic non-invasive sampling in a Mediterranean farmland, during the period between November 2013 and October 2014. Black bars and dashed lines represent 95% confidence intervals. Grey dashed line represents average monthly apparent survival (0.52).

Table 1 – Definition and summary statistics of covariates used in Cormack-Jolly-Seber (CJS) models to estimate recapture probabilities (p) and survival (ϕ) of the Cabrera vole in a Mediterranean farmland landscape of SW Portugal. Underlying hypotheses regarding covariate effects tested on p and ϕ are also presented. All environmental covariates were measured for each individual at the time of first capture (see text). * indicates variables selected for the CJS modelling procedure (see *Data analyses*).

Covariate Code	Description	Type	N	Mean \pm SE [range]	Parameter	Underlying hypothesis (based on Pita et al., 2014, except where indicated)
<i>GenSuccess</i> *	Mean genotyping success of analysed samples in each patch	Proportion (0-1)	307	0.5 \pm 0.01 [0.1 – 1]	p	p is higher in patches where genotyping success was higher
<i>Time</i> *	Transition between seasons	Categorical (EWS-LWS; LWS-EDS; EDS-LDS)	307	-	p ϕ	p and ϕ differ among seasons, being greater in spring when animals are more active, and habitat and food availability are greater; and lower in the summer when habitat and food resources are scarcer, and animals are less active.
<i>Sex</i> *	Sex of individuals	Categorical (M/F)	M: 152 F: 155	-	p ϕ	p is similar between sexes, as home-ranges vary little between males and females, and both sexes are expected to equally mark their territories. ϕ is similar between sexes, as expected for monogamous mammals (Clutton-Brock and Isvaran, 2007)
<i>PArea</i>	Area of the patch where animals were captured (m ²)	Continuous (> 0)	307	9834.0 \pm 658.93 [106.2–51764.0]	p ϕ	p decreases with patch area, as animals tend to concentrate their territories within particular areas of habitat-patches. ϕ increases with patch area, as larger patches should provide more habitat and food resources.
<i>PIsolation</i>	Mean distance from the patch to the 3 nearest patches (m)	Continuous (> 0)	307	34.7 \pm 2.06 [3.4–187.9]	ϕ	ϕ decreases with increasing patch Isolation, due to increased mortality during movement attempts between patches further apart from each other.
<i>PPermanence</i>	Permanence of the patch in the following seasons	Categorical (0/1)	0: 73 1: 234	-	ϕ	ϕ is lower if patch disappears in the following seasons
<i>RoadVerge</i> *	Binary coding of whether the patch is a roadside habitat (<10m from a road)	Categorical (0/1)	0: 66 1: 241	-	ϕ	ϕ is higher in roadside habitats, as these are usually less disturbed by farming operations.
<i>Water</i> *	Presence of a permanent water-body within or at the border of the patch	Categorical (0/1)	0: 278 1: 29	-	ϕ	ϕ is higher where water is present, as the species prefers wet vegetation providing both refuge and fresh food
<i>Arvicola</i> *	Occurrence of <i>Arvicola sapidus</i> in the patch	Categorical (0/1)	0: 102 1: 205	-	ϕ	ϕ is lower where the competitively superior <i>A. sapidus</i> is present
<i>AgroDist</i>	Distance from patch to the nearest agricultural area (m)	Continuous (\geq 0)	307	27.9 \pm 3.20 [0–248.9]	ϕ	ϕ is lower in patches closer to agricultural fields, as these are associated to increased disturbance resulting from farming operations.
<i>UrbDist</i>	Distance from the patch to nearest urban area (m)	Continuous (\geq 0)	307	36.9 \pm 3.07 [0–247.6]	ϕ	ϕ is lower in patches closer to urban areas, as these are associated to increased human disturbance and/or predation by domestic cats and dogs.

Table 2 – Set of best plausible models (with a difference of Akaike’s Information Criteria corrected for small sample size < 2) obtained from the set of 8 submodels describing p (keeping ϕ fixed) and 64 submodels describing ϕ (keeping p fixed).

Varying p, fixed ϕ	AIC_c	k
~1	178.1	9
~Time	178.4	11
~Sex + Time	179.8	12
~Sex	179.9	10
~GenSuccess	179.9	10
Varying ϕ, fixed p		
~RoadVerge + AgroDist + Sex + Water	178.1	10
~RoadVerge + AgroDist + Sex + Water + Time	179.9	12
~RoadVerge + AgroDist + Sex + Water + Arvicola	180.0	11

AIC_c – Akaike’s Information Criteria corrected for small sample size; k – degrees of freedom; ϕ – apparent survival probability; p – recapture probability.

Table 3 – Ordered set of best ranked plausible models (n = 15), obtained by fitting models separately for recapture (p) and apparent survival (ϕ) probabilities (more details in the Material and Methods section and in Bromaghin et al., 2013). Among the three most supported models (with $\Delta AIC_c < 2$), the top-ranked model (in bold, $AIC_c = 172.24$) was the only one not including uninformative covariates, being thus retained as the best model. See definition of covariates in Table 1.

ϕ	p	ΔAIC_c	w_i	k
~AgroDist + RoadVerge + Sex + Water	~1	0	0.29	6
~AgroDist + RoadVerge + Sex + Water	~Sex	1.76	0.12	7
~AgroDist + RoadVerge + Sex + Water	~GenSuccess	1.80	0.12	7
~AgroDist + RoadVerge + Sex + Water + Arvicola	~1	2.05	0.10	7
~AgroDist + RoadVerge + Sex + Water	~Time	3.07	0.06	8
~AgroDist + RoadVerge + Sex + Water + Arvicola	~Sex	3.74	0.04	8
~AgroDist + RoadVerge + Sex + Water + Time	~1	3.83	0.04	8
~AgroDist + RoadVerge + Sex + Water + Arvicola	~GenSuccess	3.84	0.04	8
~AgroDist + RoadVerge + Sex + Water	~Sex + Time	3.89	0.04	9
~AgroDist + RoadVerge + Sex + Water + Time	~Time	4.00	0.04	10
~AgroDist + RoadVerge + Sex + Water + Arvicola	~Time	5.05	0.02	9
~AgroDist + RoadVerge + Sex + Water + Time	~Sex + Time	5.43	0.02	11
~AgroDist + RoadVerge + Sex + Water + Time	~GenSuccess	5.60	0.02	9
~AgroDist + RoadVerge + Sex + Water + Time	~Sex	5.65	0.02	9
~AgroDist + RoadVerge + Sex + Water + Arvicola	~Sex + Time	5.83	0.02	10

ΔAIC_c – Difference of Akaike's Information Criteria corrected for small sample size; w_i – model relative Akaike weight; k – degrees of freedom.