

MAJOR CLADES IN SOLANUM BASED ON ndhF SEQUENCE DATA

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ABSTRACT. Analysis of sequence data from the chloroplast gene *ndh*F identifies at least 12 major well-supported clades within the genus *Solanum*. These are briefly described, given informal clade names, and compared with the groups recognized by previous *Solanum* workers. Non-molecular synapomorphies are proposed for many of the clades. Continued use of informal taxonomic designations is advocated for new infrageneric groups within *Solanum*.

Key words: ndhF, phylogeny, Solanaceae, Solanum.

olanum L., with approximately 1400 species, is the largest and most diverse genus in the Solanaceae. Solanum is distinguished from most of the other genera in the tribe Solaneae by its poricidal anther dehiscence, a character present in nearly all Solanum species and shared only with the related genus Lycianthes. Although some previous authors considered Lycianthes to be part of Solanum, recent molecular studies have confirmed the distinction between the two genera (Bohs & Olmstead, 1997; Olmstead & Palmer, 1997; Olmstead et al., 1999). Morphologically, Lycianthes is differentiated from Solanum by differences in calyx structure (D'Arcy, 1986).

Although poricidal anther dehiscence is a relatively striking synapomorphy that allows Solanum to be recognized as a genus, its division into infrageneric subunits is less clear. Early workers attempted to divide Solanum into two large groups based on spininess, anther morphology, or hair type. Linnaeus, for instance, divided Solanum into two groups, Spinosa and Inermia, based on the presence or absence of spines (Linnaeus, 1753). Dunal, in his early treatments (Dunal, 1813, 1816), maintained this distinction as his categories Aculeata and Inermia, but in his Solanum treatment for DeCandolle's Prodromus

(Dunal, 1852) he established two major infrageneric divisions ("sections") based on anther shape as well as presence or absence of spines. His group Pachystemonum encompassed species that lack spines and have relatively short, broad anthers with large terminal pores which often enlarge into longitudinal slits, whereas Leptostemonum included prickly species with relatively narrow, distally tapered anthers with small terminal pores that do not elongate with age. Bitter (1919) also recognized two major infrageneric groups, the subgenera Eusolanum and Leptostemonum, based on the same characters as Dunal (1852). Seithe (1962), in contrast, divided Solanum into two groups based not on spininess or anther morphology, but rather on hair type. She recognized two categories in Solanum at the rank of "Chorus subgenerum," distinguished by the presence of unbranched or dendritically branched hairs (Chorus subgenerum Solanum) versus stellate hairs (Chorus subgenerum Stellatipilum). Danert (1970) integrated characters of branching patterns and shoot morphology with previous systems, and, along with Gilli (1970), summarized the infrageneric groups recognized by Bitter and Seithe.

These works provided the elements of D'Arcy's (1972) classification scheme and conspectus, which is the most widely used system today.

TABLE 1.

Solanum subgenera according to D'Arcy (1972, 1991).

- 1. Solanum subg. Archaesolanum Marzell
 - ca. 8 species, Australian region
- 2. Solanum subg. Bassovia (Aubl.) Bitter
 - ca. 15 species, New World
- 3. Solanum subg. Leptostemonum (Dunal) Bitter
 - ca. 250-450 species, worldwide
- Solanum subg. Lyciosolanum Bitter
 species, South Africa

- 5. Solanum subg. Minon Raf. [subg. Brevantherum (Seithe) D'Arcy, in D'Arcy (1972)]
 - ca. 70 species, New World
- 6. Solanum subg. Potatoe (G. Don) D'Arcy
 - ca. 300 species, worldwide
- 7. Solanum subg. Solanum

200 species, worldwide

D'Arcy's scheme recognizes seven subgenera in Solanum (Table 1; D'Arcy, 1972, 1991). These range in size from the monotypic subgenus Lyciosolanum to the subgenera Solanum, Leptostemonum, and Potatoe, each of which contains hundreds of species. In his 1972 paper, D'Arcy lectotypified all subgeneric names and provided a provisional conspectus of Solanum. In this conspectus, Solanum subgenera, sections, and series are listed along with their respective type species, but all the component species of each infrageneric group are not listed, nor are the characters given that circumscribe each of the groups. D'Arcy (1991) made minor modifications to this system. Whalen (1984) provided a detailed conspectus of Solanum subg. Leptostemonum (the spiny solanums). Subsequently, both Nee (1999) and Child and Lester (2001) provided infrageneric schemes for Solanum. Nee (1999) listed the species that belong to each of his taxonomic categories, but his system includes only New World taxa. Child and Lester (2001), like D'Arcy (1972), listed only the type species for each of their infrageneric groups. Hunziker (2001) modified D'Arcy's (1972) system and provided descriptions and commentary for each recognized section. All of these classifications relied completely on morphological data and, except for Whalen (1984), none utilized techniques of cladistic analysis.

The advent of molecular data has revolutionized the field of plant systematics and has led to new insights into phylogenetic relationships at all taxonomic levels. In the Solanaceae, Olmstead and colleagues have used restriction site and sequence data to examine phylogenetic relationships across the entire family (Olmstead & Palmer, 1992; Olmstead et al., 1999). Molecular studies above the sectional level in Solanum include the works of Spooner et al. (1993), Olmstead and Palmer (1997), and Bohs and Olmstead (1997, 1999, 2001). These studies provide information on major clades within Solanum, but none have sampled from all the subgenera recognized by morphological systematists such as Bitter, Seithe, Danert, and D'Arcy.

This paper presents results of a molecular phylogenetic study designed to identify major clades

within Solanum using sampling from a broad spectrum of Solanum subgroups. Results are presented from an analysis of sequence data from the chloroplast gene ndhF. Sampling includes members of all seven of D'Arcy's subgenera and over 40 of the 62 sections listed in D'Arcy (1991). All the sections listed in D'Arcy's (1972) conspectus as well as many sections described after 1972 are discussed in context of the major ndhF clades. Major lineages are described with informal clade names and their component sectional groups are listed. Possible non-molecular synapomorphies are suggested for most of the identified clades. These characters have been taken from the general references listed above and from the personal observations of the author. Although they may provide general guidelines for the recognition of clades, this is not intended to be a substitute for thorough morphological analyses, as many of the suggested characters are variable within clades and may be found in more than one clade. A few overall recommendations are made for taxonomic rearrangements within the genus Solanum. Results of analyses using data from nuclear genes such as ITS and waxy (Bohs, in prep.) and from combined chloroplast and nuclear sequence data sets will be presented in a future publication.

MATERIALS AND METHODS

Sampling comprised 120 species of Solanaceae, including five outgroup genera from the tribe Solaneae. Outgroup taxa were chosen on the basis of previously published results of Olmstead et al. (1999) and Bohs and Olmstead (2001). Solanum taxa sampled included representatives of all seven of D'Arcy's subgenera and a number of sections or species groups thought to represent distinctive clades based on morphology. Collection and voucher information is given in Table 2.

DNA was extracted from fresh or silica-dried leaves or, in rare cases, from herbarium specimens, using either the modified CTAB procedure of Doyle and Doyle (1987) or a microextraction protocol that used QiaQuick columns and buffer (Qiagen, Inc.) in place of the isopropanol precipitation step in the CTAB procedure. Samples

TABLE 2.

Sources of taxa sequenced for *ndh*F. ^aDNA extracts provided by: (1) L. Bohs, University of Utah, Salt Lake City, Utah; (2) R. G. Olmstead, University of Washington, Seattle, Washington; (3) T. Mione, Central Connecticut State University, New Britain, Connecticut; (4) D. Spooner, University of Wisconsin, Madison, Wisconsin; (5) A. Bruneau, McGill University, Montreal, Canada. ^bAccording to D'Arcy (1972, 1991) unless noted. ^cAccession numbers given for cultivated collections. BIRM = cultivated at University of Birmingham, U.K.; NIJ = cultivated at University of Nijmegen, The Netherlands; PI = U.S.D.A. Plant Introduction number; D'Arcy collection = cultivated at MO. ^dCollector, number, and herbarium acronym (if known) of herbarium vouchers. ^eChild (1998). ^dBohs (1990). ^gNee (1999). ^hSymon (1981). ⁱD'Arcy (1992). ^jKnapp (2000).

Taxon*	Subgenus ^b	Section ^b	Collection locality ^c	Voucher ^d	GenBank accession number
Capsicum baccatum L. var. pendulum (Willd.) Eshbaugh ²			Bolivia	Eshbaugh 1584 (MU)	U08916
Capsicum chacoense Hunz. ²			Bolivia	Eshbaugh 1586A (MU)	AF500809
Jaltomata procumbens (Cav.) J. L. Gentry ³			Mexico	Davis 1189A	U47429
Jaltomata sinuosa (Miers) Mione ¹			Bolivia	Nee et al. 51830 (NY)	AF500835
Lycianthes heteroclita (Sendtn.) Bitter 1			Costa Rica	Bohs 2376 (UT)	U72756
Lycianthes rantonnei (Carrière) Bitter ²			BIRM S.0928	RGO S-96 (WTU)	AF500840
Physalis alkekengi L. ²		_	D'Arcy collection	D'Arcy 17707 (MO)	U08927
Solanum abutiloides (Griseb.) Bitter & Lillo ²	Minon	Brevantherum	BIRM S.0655	RGO S-73 (WTU)	U47415
Solanum accrescens Standl. & C. V. Morton ¹	Leptostemonum	Erythrotrichum ^e	Costa Rica	Bohs 2556 (UT)	AF500795
Solanum adhaerens Roem. & Schult. 1	Leptostemonum	Micracantha	Costa Rica	Bohs 2473 (UT)	AF224061
Solanum adscendens Sendtn. ¹	Solanum	Gonatotrichum	Bolivia	Bohs & Nee 2738 (UT)	AF500796
Solanum aethiopicum L. ²	Leptostemonum	Oliganthes	BIRM 5.0344	RGO S-74 (WTU)	AF500797
Solanum aggregatum Jacq. ²	Lyciosolanum	Lyciosolanum	South Africa	RGO 99-25 (WTU)	AF500798
Solanum aligerum Schltdl. ¹	Minon	Holophylla	Bolivia	Nee et al. 51822 (NY)	AF500799
Solanum allophyllum (Miers) Standl. ¹	None ^f	Allophyllum ^f	Panama	Bohs 2339 (UT)	U47416
Solanum amygdalifolium Steud. ¹	Potatoe	Jasminosolanum	Argentina	Nee & Bohs 50840 (NY)	AF500800
Solanum aphyodendron S. Knapp ²	Solanum	Geminata	Colombia	RGO S-92 (WTU)	AF500801
Solanum appendiculatum Dunal ²	Potatoe	Basarthrum	Mexico	Anderson 1401 (CONN)	AF224062
Solanum arboreum Dunal 1	Sol an um	Geminata	Costa Rica	Bohs 2521 (UT)	U47417

Solanum argentinum Bitter & Lillo ¹	Minon	Holophylla	Argentina	Bohs 2539 (UT)	U72752
Solanum aviculare G. Forst. ²	Archaesolanum	Archaesolanum	BIRM 5.0809	none	U47418
Solanum betaceum Cav. 1	Genus Cyphomandra	Pachyphylla	Bolivia	Bohs 2468 (UT)	U47428
Solanum brevicaule Bitter ⁴	Potatoe	Petota	Bolivia PI 498115	Hawkes et al. 6701	AF500803
Solanum bulbocastanum Dunal ⁴	Potatoe	Petota	Mexico PI 347757	Tarn 153	AF500804
Solanum caesium Griseb. ¹	Solanum	Solanum .	Bolivia	Bohs et al. 2815 (UT)	AF500805
Solanum calileguae Cabrera 1	Solanum ^g	Dulcamara ^g	Argentina	Nee & Bohs 50809 (NY) AF500806_
Solanum campanulatum R. Br. ²	Leptostemonum	Campanulata	BIRM 5.0387	RGO 5-78 (WTU)	AF500807
Solanum campechiense L. ¹	Leptostemonum	Unclear	Costa Rica	Bohs 2536 (UT)	AF224071
Solanum candidum Lindl. ²	Leptostemonum	Lasiocarpa	BIRM 5.0975	RGO 5-100 (WTU)	AF224072
Solanum capsicoides All. 1	Leptostemonum	_Acanthophora	Peru	Bohs 2451 (UT)	AF500808
Solanum carolinense L. ²	Leptostemonum	Lathyrocarpum	BIRM 5.1816	RGO S-77 (WTU)	AF500811
Solanum chenopodinum F. Muell. ²	Leptostemonum	Graciliflora	BIRM 5.0813	none	AF500812
Solanum cinereum R. Br. 1	Leptostemonum	Melongena ^h	NIJ 904750120	Bohs 2852 (UT)	AF500813
Solanum citrullifolium A. Braun ²	Leptostemonum	Androceras	BIRM 5.0127	RGO S-79 (WTU)	AF500814
Solanum cleistogamum Symon ²	Leptostemonum	Oliganthes	BIRM 5.0844	RGO S-80 (WTU)	AF500815
Solanum conditum C. V. Morton ¹	Leptostemonum	Unclear	Bolivia	Bohs & Nee 2733 (NY)	AF500816
Solanum cordovense Sessé & Moç. 1	Minon	Extensum	Costa Rica	Bohs 2693 (UT)	U72751 .
Solanum crinitipes Dunal ²	Leptostemonum	Torva	Colombia	RGO 5-81 (WTU)	AF500817
Solanum crinitum Lam. ¹	Leptostemonum	Crinitum ^e	NIJ 924750049	Bohs 2850 (UT)	AF500818
Solanum crispum Ruiz & Pav. ²	Minon	Holophylla	BIRM 5.0486	none	AF500819
Solanum deflexum Greenm. ¹	Solanum	Gonatotrichum	Costa Rica	Bohs 2715 (UT)	AF500820
Solanum delitescens C. V. Morton ¹	Minon	Holophylla	Argentina	Nee & Bohs 50810 (NY)	AF500821
Solanum diploconos (Mart.) Bohs ¹	Genus Cyphomandra	Pachyphylla	Brazil	Bohs 2335 (UT)	AY049014
Solanum doddsii Correll ⁴	Potatoe	Petota	Bolivia	Spooner et al. 6651	AF500822
Solanum drymophilum O. E. Schulz ¹	Leptostemonum	Persicariae	Puerto Rico	Bohs 2461 (UT)	AF500823
Solanum dulcamara L. ²	Potatoe	Dulcamara	U.S.A.	none	U47419
Solanum echinatum R. Br. ¹	Leptostemonum	Leprophora	NIJ 954750052	Bohs 2727 (UT)	AF500824
Sol a num elaeagnifolium Cav. ²	Leptostemonum	Leprophora	U.S.A.	RGO S-82 (WTU)	AF224067
Solanum etuberosum Lindl. ⁴	Potatoe	Petota	Chile Pl 498311	UAC 1322	AF500825

TABLE 2 CONTINUED.

Taxon a	Subgenus ^b	Section ^b	Collection locality ^c	Voucher ^d	GenBank accession number
Solanum evolvulifolium Greenm. ¹	Bassovia	Herpystichum ⁹	Pana ma	Knapp & Mallet 9178 (BM)) AF500826
Solanum ferocissimum Lindl. ²	Leptostemonum	Graciliflora	BIRM 5.0819	RGO 5-83 (WTU)	AF500827
Solanum fiebrigii Bitter ¹	Solanum	Solanum	Bolivia	Bohs et al. 2784 (UT)	AF500828
Solanum fraxinifolium Dunal ¹	Potatoe	Basarthrum	Costa Rica	Bohs 2558 (UT)	AF500810
Solanum furfuraceum R. Br. ²	Leptostemonum	Unclear	BIRM 5.1442	RGO 5-84 (WTU)	AF500829
Solanum glaucophyllum Desf. ²	Solanum	Glaucophyllum	D'Arcy collection	none	U72753
Solanum havanense Jacq. 1	Solanum	Diamonon ^e	NIJ 904750122	Bohs 3076 (UT)	AF500830
Solanum herculeum Bohs ²	genus <i>Triguera</i>	 	Morocco	Jury 13742 (RNG)	AF224065
Solanum hindsianum Benth. 1	Leptostemonum	Unclear	Mexico	Bohs 2975 (UT)	AF500831
Solanum hoehnei C. V. Morton ¹	Leptostemonum	Nemorense	Brazil	Folli 1668 (MO)	AF500832
Solanum inelegans Rusby 1	Minon	Holophylla	Bolívia	Nee et al. 51813 (NY)	AF500833
Solanum ipomoeoides Chodat & Hassl. 1	Solanum ^g	Dulcamarag	Bolivia	Bohs & Nee 2766 (UT)	AF500834
Solanum jamaicense Mill. ²	Leptostemonum	Eriophylla	BIRM S.1209	RGO S-85 (WTU)	AF224073
Solanum jasminoides Paxton ²	Potatoe	Jasminosolanum	Colombia	RGO S-86 (WTU)	AF500836
Solanum juglandifolium Dunal ⁴	Potatoe	Petota	Colombia	LA 2788	AF500837
Solanum laciniatum Aiton ¹	Archaesolanum	Archaesolanum	New Zealand	Bohs 2528 (UT)	U47420
Solanum lepidotum Dunal ¹	Minon	Lepidotum	Costa Rica	Bohs 2621 (UT)	AF500838
Solanum lidii Sunding ¹	Leptostemonum	Nycterium	NIJ 934750022	Bohs 2903 (UT)	AF500839
Solanum luteoalbum Pers.1	Genus Cyphomandra	Cyphomandropsis	BIRM 5.0042	Bohs 2337 (UT)	U72749
Solanum lycopersicum L. ²	Genus Lycopersicon	Lycopersicon	U.S.A. (cult.)	none	U08921
Solanum macrocarpon L. ²	Leptostemonum	Melongena	BIRM 5.0133	RGO S-88 (WTU)	AF224068
Solanum mahoriense D'Arcy & Rakot. ¹	Leptostemonum	Cryptocarpum ⁱ	Madagascar	Bohs 2576 (UT)	AF500841
Solanum mammosum L. ²	Leptostemonum	Acanthophora	BIRM 5.0983	RGO S-89 (WTU)	AF224074
Solanum mapiriense Bitter ¹	None ^f	Allophyllum ^f	Bolivia	Nee & Solomon 30305 (UT)	AF500842
Solanum mauritianum Scop. ²	Minon	Brevantherum	BIRM S.0860	RGO 5-90 (WTU)	AF500843

Solanum melongena L. ²	Leptostemonum	Melongena	BIRM S.0657	RGO S-91 (WTU)	AF224069
Solanum montanum L. ¹	Potatoe	Regmandra	NIJ 904750205	Bohs 2870 (UT)	AF500844
Solanum multifidum Ruiz & Pav. 1	Potatoe	Regmandra	NIJ 904750205	Bohs 2863 (UT)	AF500845
Solanum muricatum Aiton ²	Potatoe	Basarthrum	Colombia	RGO S-93 (WTU)	AF500846
Solanum nemorense Dunal 1	Leptostemonum	Nemorense	Bolivia	Bohs & Nee 2757 (UT)	AF500847
Solanum nitidum Ruiz & Pav. ¹	Minon	Holophylla	Bolivia	Nee 31944 (NY)	AF224075
Solanum ochrophyllum Van Heurck & Müll. Arg. 1	Solanum	Geminata	Bolivia	Bohs & Nee 2805 (UT)	AF500848
Solanum palitans C. V. Morton ¹	Solanum	Parasolanum	BIRM 5.0837/70	Bohs 2449 (UT)	AF224064
Solanum phaseoloides Pol. ¹	Bassovia	Herpystichum	Costa Rica	Bohs 2485 (UT)	AF500849
Solanum physalifolium Rusby var. nitidibaccatum (Bitter) Edmonds ¹	Solanum	Solanum	U.S.A.	Bohs 2467 (UT)	U47421
Solanum pinnatisectum Dunal ⁴	Potatoe	Petota	Mexico PI 347766	Tarn 205A	AF500850
Solanum piurae Bitter ⁴	Potatoe	Petota	Peru Pl 310997	Hawkes 2443	AF500851
Solanum prinophyllum Dunal ¹	Leptostemonum	Oliganthes ^h	NIJ 904750171	Bohs 2725 (UT)	AF500852
Solanum pseudocapsicum L. ²	Minon	Pseudocapsicum	BIRM 5.0870	none	U47422
Solanum ptychanthum Dunal ²	Solanum	Solanum	U.S.A.	RGO S-94 (WTU)	U47423
Solanum pubigerum Dunal ¹	Minon	Holophylla	NIJ 904750104	none	AF500853
Solanum pyracanthum Lam. ²	Leptostemonum	Oliganthes	cult. (UCSB Bot. Gard.)	RGO S-95 (WTU)	AF500854
Solanum guadrangulare L.f. ²	Solanum	Quadrangulare	South Africa	RGO 99-46 (WTU)	AF500855
Solanum riojense Bitter ¹	Solanum	Episarcophyllum	Argentina	Nee & Bohs 50843 (NY)	AF500856
Solanum rostratum Dunal ¹	Leptostemonum	Androceras	U.S.A.	none	U47424
Solanum rovirosanum Donn. Sm. 1	Solanum	Geminata	Costa Rica	Bohs 2919 (UT)	AF500857
Solanum rugosum Dunal ¹	Minon	Brevantherum	Costa Rica	Bohs 3011 (UT)	AF500858
Solanum sandwicense Hook. & Arn. ¹	Leptostemonum	Irenosolanum_	Hawaii	Bohs 29 92 (UT)	AF500859
Solanum schimperianum Hochst. ²	Leptostemonum	Torva	BIRM 5.1538	RGO S-97 (WTU)	AF500860
Solanum schlechtendalianum Walp. ¹	Minon	Extensum	Costa Rica	Bohs 2915 (UT)	AF500861
Solanum seaforthianum Andrews ²	Potatoe	Jasminosolanum	BIRM 5.0051	none	U47425
Solanum sisymbriifolium Lam. ¹	Leptostemonum	Cryptocarpum	Argentina	Bohs 2533 (UT)	AF500862
Solanum stenophyllidium Bitter ¹	Potatoe	Petota	NIJ 904750079	Bohs 2855 (UT)	AF500802
Solanum stramonifolium Jacq. ⁵	Leptostemonum	Lasiocarpa	Peru	Whalen 860 (HUT)	AF500863

Taxon*	Sub	ogenus ^b	Section ^b	Collection locality °	Voucher d	GenBank accession number
Solanum terminale Forssk. ¹	Sola	num	Afrosolanum	NIJ 814750072	Bohs 2719 (UT)	AF500864
Solanum thelopodium Sendtn. ¹	Unc	lear ^j	Unclear ^j	Bolivia	Nee & Bohs 50858 (NY)) AF500865
Solanum toliaraea D'Arcy & Rakot. 1	Lep	tostemonum	Unclear ¹	Madagascar	Bohs 2574 (UT)	AF500866
Solanum torvum Sw. ²	Lep	tostemonum	Torva	BIRM 5.0839	RGO S-101 (WTU)	L76286
Solanum tridynamum Dunal ²	Lep	tostemonum	Nycterium	BIRM 5.1831	RGO S-102 (WTU)	AF500867
Solanum triflorum Nutt. ¹	Sola	num	Parasolanum	U.S.A.	Bohs 3062 (UT)	AF500868
Solanum tripartitum Dunal ¹	<u>Sola</u>	num	Parasolanum	BIRM 5.0708/71	Bohs 2465 (UT)	U72750
Solanum trisectum Dunal ¹	Pota	itoe	Normania	France	Bohs 2718 (UT)	AF224063
Solanum trizygum Bitter ¹	Bass	ovia	Pteroidea	Costa Rica	Bohs 2511 (UT)	U72754
Solanum tuberosum L. ²	Pota	atoe	Petota	U.S.A. (cult.) PI (245793 X 245796)	WRF 1610 USDA NRSP-6	L762 8 7
Solanum turneroides Chodat ¹	Sola	num	Gonatotrichum	Bolivia	Nee et al. 51716 (NY)	AF500 8 69
Solanum uleanum Bitter 1	Bass	ovia	Pteroidea	D'Arcy collection	Bohs 2720 (UT)	AF500870
Solanum vespertilio Aiton ²	Lept	tostemonum	Nycterium	BIRM 5.2091	RGO S-103 (WTU)	AF224070
Solanum villosum Mill. ¹	Sola	num	Solanum	Iran Pl 304600	Bohs 2553 (UT)	AF224066
Solanum wallacei (A. Gray) Parish ¹	Pota	toee	Californisolanum ^e	U.S.A.	Bohs 2438 (UT)	U47426
Solanum wendlandii Hook. f. ²	Lept	tostemonum	Aculeigerum	BIRM 5.0488	none	U47427
Witheringia solanacea L'Her. 1				Costa Rica	Bohs 2416 (UT)	U72755

extracted with the modified CTAB method were purified using cesium chloride density gradient centrifugation or a phenol-chloroform protocol. Amplification and sequencing of the ndhF gene used the primers and PCR program given in Bohs and Olmstead (1997). PCR products were cleaned using QiaQuick spin columns and sequenced on an ABI automated sequencer. Sequences were edited and contigs assembled using the program Sequencher (Gene Codes Corp.). After a consensus sequence was obtained, it was aligned by eye to a template sequence (Nicotiana tabacum L.). Base changes relative to the template sequence were then double-checked against the chromatograms. No alignment difficulties were encountered in assembling the sequences into a data set in NEXUS file format. All new sequences reported here have been submitted to GenBank (Table 2). The data set and resultant phylogenetic trees have been submitted to TreeBASE (accession numbers \$735 and M1167).

The data matrix was analyzed using unweighted parsimony with the program PAUP*4.0b10 (Swofford, 2002). The analysis used the heuristic search algorithm with the TBR and MulTrees options, 714 random addition replicates with rearrangements limited to 100,000 per replicate, and gaps treated as missing data. Trees were rooted using *Physalis alkekengi* as the outgroup. Bootstrap analyses were performed with 500 replicates using the heuristic search option, TBR and MulTrees, MaxTrees set to 1000, and 1,000,000 rearrangements per replicate.

The data were also analyzed using the parsimony ratchet (Nixon, 1999) as implemented in the program PAUPRat (Sikes & Lewis, 2001). Five replicate searches of 200 iterations each were performed. The shortest trees from all searches were retained and combined into a single consensus tree.

The same data matrix was analyzed by maximum likelihood using the program fastDNAml (Olsen et al., 1994) on a UNIX platform computer. Parameters used in the analysis were a transition/transversion ratio of 1.0006 (estimated using

ML in PAUP from a neighbor-joining tree of the 120-taxon data set), empirical base frequencies (A = 0.27665, C = 0.15518, G = 0.18366, T = 0.38450), and random addition order.

RESULTS

The ndhF sequences obtained for all taxa except Lycianthes heteroclita, Solanum wendlandii, S. diploconos, and S. deflexum were 2086 base pairs long, corresponding to positions 24 through 2109 in the tobacco ndhF sequence. Lycianthes heteroclita had a 15 bp insertion, S. wendlandii had a 33 bp insertion, and S. diploconos had a 24 bp insertion between positions 1476 and 1477. Solanum deflexum had a 9 bp deletion between positions 1703 and 1711.

Of 2119 total characters in the data set, 541 were variable and 288 of these were parsimony-informative. Pairwise sequence divergence calculated using the Kimura 2-parameter model ranged from 3.4% between *S. candidum* versus *Lycianthes heteroclita* to 0.048% in the closely related species pairs *S. ferocissimum* versus *S. chenopodinum*, *S. vespertilio* versus *S. liddii*, *S. doddsii* versus *S. stenophyllidium*, and *S. piurae* versus *S. doddsii*. *Solanum schlechtendalianum* and *S. lepidotum* had identical *ndh*F sequences.

The available memory capacity of PAUP on a Power Macintosh G4 was reached after saving 18,200 most parsimonious trees from 714 random addition replicates. These trees were 1053 steps long with a CI (excluding uninformative characters) of 0.497 and RI of 0.819. PAUPRat saved 992 trees of 1053 steps out of 1000 iterations. The strict consensus trees from the heuristic parsimony and the PAUPRat searches were nearly identical, differing only in greater resolution at two of the branch tips in the PAUPRat consensus tree (not shown). Likewise, the maximum likelihood topology (not shown) was virtually identical to the parsimony trees and included the same taxa in the major clades described below. This analysis was completed overnight, examined 39,626 trees, and resulted in a tree with a log likelihood of -13487.40739.

In these trees, Solanum forms a monophyletic clade, with members of the former genera Lycopersion, Cyphomandra, Normania, and Triguera nested within it (Fig. 1). Species of all these genera have been transferred to Solanum (Spooner et al., 1993; Bohs, 1995; Bohs & Olmstead, 2001). Capsicum plus Lycianthes emerges as the sister group to the Solanum clade with bootstrap support of 70%. Solanum plus the genera Jaltomata, Lycianthes, and Capsicum form a well-supported clade (bootstrap = 100%), and Lycianthes plus Capsicum form a well-supported group (bootstrap = 89%).

At least 12 major clades can be discerned within Solanum (Fig. 1, see pp. 48–49). These clades are supported with bootstrap values ranging from 51% (Leptostemonum s.l.) to 100% (the Regmandra, Archaesolanum, and Normania clades). However, the relationships among these major clades are unclear, because for the most part they form a polytomy at the base of Solanum. Several of these clades conform to infrageneric groups recognized by previous systematists, but others do not.

These clades have been given informal clade names and are briefly described below with a list of their constituent sections and non-molecular synapomorphies that may define them. Asterisks (*) indicate sections or species groups that have been sampled in the present analysis. Other groups listed under each clade are inferred to belong there due to morphological similarity. Brief comparisons are made with reference to D'Arcy's (1972) classification and with several other schemes.

DISCUSSION

Major clades defined by ndhF data:

1. Thelopodium clade

3 spp., South America

Included taxa:

Solanum thelopodium species group sensu Knapp (2000)*

This group is morphologically distinctive due to its enlarged roots, single-stemmed growth habit, reduced number of sympodia, and narrow, tapered, dimorphic anthers. It was revised recently by Knapp (2000), who recognized three species. One of them, S. thelopodium, was included in the ndhF analysis, where it forms a single branch at the very base of Solanum. This placement is surprising and has not been suggested by recent Solanaceae systematists, although Bitter thought that S. thelopodium was sufficiently distinct to merit generic rank (Knapp, 2000). Dunal (1852) and Seithe (1962) placed S. thelopodium into Solanum sect. Anthoresis (Dunal) Bitter, but this means little, as section Anthoresis is a catch-all group of disparate taxa. D'Arcy did not include it in either of his summary classifications (D'Arcy, 1972, 1991). Nee (1999) put this species into Solanum sect. Pteroidea (Potato clade), but the ndhF data do not support this placement. Further sampling is needed to determine if the basal position of this clade in Solanum is correct or is perhaps a long branch artifact.

2. Regmandra clade

ca. 7 spp., South America

Included taxa:

Solanum subg. Potatoe (G. Don) D'Arcy pro parte

Solanum sect. Regmandra (Dunal) D'Arcy*

D'Arcy (1972, 1991) placed this small group of species from Pacific coastal deserts of South America into Solanum subg. Potatoe. Nee (1999) also allied this section with the potatoes, whereas Child and Lester (2001) put it into Solanum subg. Solanum, and Hunziker (2001) considered its subgeneric position uncertain. Taxa of Solanum sect. Regmandra included in the ndhF data set are S. montanum and S. multifidum, and they fall out together on a well-supported but isolated clade near the base of Solanum.

Non-molecular characters that may distinguish this clade include herbaceous habit and usually pinnately dissected and rather thick leaves, sometimes with winged petioles and stems. Plants of Solanum montanum and S. multifidum grown in the University of Utah greenhouse had nearly rotate corollas and notably expanded stigmas. Solanum montanum is reported to bear tubers (Dunal, 1852; Macbride, 1962), but the ndhF results do not suggest a direct relationship between the Regmandra clade and the tuber-bearing members of the Potato clade.

3. Archaesolanum clade

ca. 8 spp., Australia, New Guinea, New Zealand

Included taxa:

Solanum subg. Archaesolanum Marzell

Solanum sect. Archaesolanum (Marzell) Danert*

This is a distinctive group with no obvious close relatives within Solanum. It is distinguished by its aneuploid chromosome number based on n=23, a number unique in the genus. All species of this group occur in Australia and the South Pacific (New Guinea, Australia, Tasmania, New Zealand). Aside from its chromosome number, possible non-molecular synapomorphies of this clade include plurifoliate sympodial units, rotate corollas with abundant interpetalar tissue, loosely erect anthers on relatively long filaments, and fruits with abundant stone cell aggregates. The basal position of this clade may indicate a relatively old radiation in the South Pacific.

The Archaesolanum clade has been recognized as distinct by virtually all previous *Solanum* workers, including D'Arcy (1972, 1991), Bitter in Marzell (1927), Danert (1970), and Symon (1994). Olmstead and Palmer (1997) included *S. aviculare* in their analysis of *Solanum* using chloroplast restriction site data, and it formed a clade with 76% bootstrap support along with *S. ptychanthum*, *S. crispum*, *S. dulcamara*, and *S. jasminoides*. However, sampling within non-spiny *Solanum* taxa was sparse in their study, with 17 non-spiny representatives out of 36 total *Solanum* species. Bohs and Olmstead (2001)

found that *S. aviculare* and *S. laciniatum* formed a well-supported basal clade in *Solanum* in analyses using nuclear ITS sequence data as well as ITS combined with *ndh*F data. It seems safe to say that the Archaesolanum clade represents an isolated group whose closest relatives have not yet been identified

4. Normania dade

3 spp., Macaronesia, Spain, NW Africa

Included taxa:

Solanum sect. Normania (Lowe) Bitter [genus Normania Lowe]*
genus Triguera Cav.*

This clade includes two enigmatic groups endemic to Macaronesia and adjacent areas of Spain and northwestern Africa. Although these taxa have been recognized as the segregate genera Normania and Triguera, molecular data indicate that both are nested within Solanum and the three species of both genera have been transferred to Solanum (Bohs & Olmstead, 2001). Francisco-Ortega et al. (1993) made a thorough morphological analysis of Normania and Triguera and concluded that they were closely related.

Numerous non-molecular characters unite the species of the Normania clade, including herbaceous or weakly woody habit, foliaceous and accrescent calyces, zygomorphic corollas, subequal to very unequal stamens, anther dehiscence by both apical pores and longitudinal slits, anthers with horned projections, fruits dry or with sparse pulp, seeds large and few per fruit with the seed coat cell walls radially expanded, and pollen grains with colpi joined at the poles. Affinities of the Normania clade within Solanum are presently obscure. In combined analyses of ndhF and ITS data this clade forms a group with members of the Potato and Morelloid/ Dulcamaroid clade (Bohs & Olmstead, 2001), but this placement is poorly supported, with a bootstrap value of 17%. As with the Archaesolanum clade, the Normania clade appears to form an isolated group within Solanum without obvious close relatives.

5. African non-spiny clade

ca. 7 spp., Africa

Included taxa:

Solanum subg. Lyciosolanum Bitter*
Solanum subg. Solanum pro parte

Solanum sect. Afrosolanum Bitter*

Solanum sect. Quadrangulare Bitter*

Solanum sect. Benderianum Bitter

D'Arcy (1972, 1991) recognized Solanum subg. Lyciosolanum as monotypic, with S. aggregatum as its sole member, but the ndhF data indicate that probably this group should be expanded to include members of Solanum sects. Afrosolanum, Quadrangulare, and perhaps Benderianum, all placed by D'Arcy (1972, 1991) in Solanum subg. Solanum. This clade forms an isolated group within Solanum. It is poorly known taxonomically, but possible non-molecular synapomorphies may include shrubby or climbing habit, unbranched or dendritically branched hairs, and purple or white stellate corollas. This group needs better molecular sampling and morphological characterization.

No DNA samples are available from representatives of *Solanum* sects. *Lemurisolanum* Bitter and *Macronesiotes* Bitter, two non-spiny sections endemic to Madagascar. Their affinities may lie with the African non-spiny clade or with the Dulcamaroid clade.

6. Potato clade

ca. 200-300 spp., New World

Included taxa:

Solanum subg. Potatoe (G. Don) D'Arcy pro parte

Solanum sect. Petota Dumort.*

Solanum sect. Anarrhichomenum Bitter*

Solanum sect. Basarthrum

(Bitter) Bitter*

Solanum sect. Lycopersicon (Mill.) Wettst.*

Solanum sect. Neolycopersicon
Correll

Solanum sect. Juglandifolium (Rydb.) A. Child*

Solanum sect. Etuberosum (Bukasov & Kamaraz) A. Child*

Solanum sect. Articulatum (Correll) A. Child

Solanum sect. Taeniotrichum A Child

Solanum subg. Bassovia (Aubl.) Bitter pro parte

> Solanum sect. Herpystichum Bitter*

Solanum sect. Pteroidea Dunal*

This clade includes most of the groups of D'Arcy's subgenera Potatoe and Bassovia. Child's treatment of subgenus Potatoe (Child, 1990; Child & Lester, 2001) included these groups, but his concept also encompassed a number of disparate elements that are placed here in different clades, such as Solanum sect. Normania (here placed in the Normania clade), the dulcamaroid taxa sensu Child and Lester (2001; sects. Dulcamara, Jasminosolanum, and Californisolanum, here placed in the Dulcamaroid clade), and the "anomalously prickly" taxa sensu Child (1990; Aculeigerum. sects. Nemorense. and Herposolanum, here placed in the Wendlandii/Allophyllum and Leptostemonum clades). Nee's recent Solanum scheme (Nee, 1999) considered the taxa that here belong to the Potato clade to represent two distinct evolutionary lines. He included the potatoes and their relatives (sects. Petota, Anarrhichomenum, Basarthrum) in a large and morphologically diverse subgenus Solanum, along with other groups such as sections Dulcamara, Solanum,

Holophylla, Brevantherum, Regmandra, and Archaesolanum. He also included members of section Herpystichum in this clade. As Nee (1999) noted, the type of section Herpystichum is not known with certainty and the group is not well circumscribed, but he listed S. phaseoloides and S. evolvulifolium as members of the section. These species are sampled in the ndhF analyses, and they both fall out in the Potato clade.

On the other hand, Nee (1999) maintained Solanum subg. Bassovia, amplifying it to include sections Cyphomandropsis and Pachyphylla of the Cyphomandra clade and section Allophylla of the Wendlandii/Allophyllum clade along with section Pteroidea, which was placed in subgenus Bassovia by previous workers such as Bitter (1921), Seithe (1962), Danert (1970), and D'Arcy (1972). Knapp and Helgason (1997) revised the species of section Pteroidea, but they were unsure of the higher-level relationships of the section.

The ndhF data indicate that section Pteroidea belongs to the Potato clade, and that the sampled representatives of the subgenera Potatoe and Bassovia sensu D'Arcy (1972) each form monophyletic clades. Non-molecular synapomorphies that may unite both of these groups include herbaceous to weakly woody and often scandent habit, exclusively unbranched hairs, presence of rhizomes or tubers in many taxa, presence of compound leaves in most species, and lack of stone cell aggregates in the fruits. The presence of solanidine/tomatidine alkaloids may be the most consistent synapomorphy that defines the subgenus Potatoe. Whether members of the subgenus Bassovia possess these types of alkaloids is unknown.

Child (1990) placed Solanum evolvulifolium in section Anarrhichomenum, whereas Nee (1999) placed this species in section Herpystichum. The ndhF data show that S. evolvulifolium is more closely related to S. phaseoloides (sect. Herpystichum) than to S. appendiculatum (sect. Anarrhichomenum).

The placement of this monotypic Solanum sect. Rhynchantherum Bitter has been debated. Dunal (1852), D'Arcy (1972, 1991), and Hunziker (2001) assigned it to subgenus Potatoe, Bitter (1913a) proposed an affinity with S. reptans of section S. Herposolanum (cf. hoehnei Leptostemonum clade), and Miers (1855) and Child (1984b; Child & Lester, 2001) placed it in the genus Cyphomandra (Cyphomandra clade). Although no DNA data are available, its pinnately compound leaves and anther structure (described in Bohs, 1994) argue for placement in the Potato clade.

7. Morelloid/Dulcamaroid clade

This group comprises two subclades, which will be discussed separately. Bootstrap support for the association of the two groups is strong (94% of bootstrap replicates) in the *ndhF* data set, but additional molecular data from other genes are needed to ascertain whether this group should be better recognized as two separate clades. For instance, ITS data from a small subset of the taxa considered here provided weak support (19% of bootstrap replicates) for the association of the morelloid and dulcamaroid subgroups (Bohs & Olmstead, 2001).

7a. Morelloid clade

ca. 75 spp., worldwide

included taxa:

Solanum subg. Solanum pro parte

Solanum sect. Solanum*

Solanum sect.

Campanulisolanum Bitter*

Solanum sect. Parasolanum A. Child*

Solanum sect.

Episarcophyllum Bitter*

Solanum sect.

Chamaesarachidium Bitter

This clade includes the core of Solanum species often known as the morelloid taxa. The four sections exclusive of section Parasolanum are morphologically homogeneous, and sectional distinctions are not clear-cut. Three members of Solanum sect. Parasolanum (S. tripartitum, S. palitans, S. triflorum) were sampled in the ndhF analyses, and all are included in the morelloid clade. However, these three taxa do not fall out together, indicating that section Parasolanum as circumscribed by Child (1984a) may not be a monophyletic group. In the ndhF analyses, S. tripartitum and S. palitans form a strongly supported clade, which, in turn, is strongly associated with the rest of the Morelloid clade (95% bootstrap support). However, these two species form a separate group distinct from the rest of the Morelloid clade in trees based on ITS sequences (Bohs & Olmstead, 2001), More extensive ITS sampling along with molecular data from additional genes may enhance the circumscription and placement of section Parasolanum.

Some non-molecular characters that may serve to unite this clade include herbaceous or weakly woody habit, 2- to 3-foliate sympodial units, pubescent filaments and styles in many taxa, and small stone cell aggregates in the fruits.

7b. Dulcamaroid clade

ca. 40 spp., worldwide

Included taxa:

Solanum subg. Potatoe (G. Don) D'Arcy pro parte

Solanum sect. Dulcamara Dumort.*

Solanum sect. Jasminosolanum Seithe*

Solanum sect. Californisolanum A. Child*

Solanum subg. Solanum pro parte

Solanum sect. Lysiphellos (Bitter) Seithe Solanum subg. Minon Raf. pro parte

Solanum sect. Holophylla Walp, pro parte*

This clade consists of elements from three of D'Arcy's subgenera. Sectional limits are not well defined, and the majority of groups included here are in need of critical taxonomic revision and nomenclatural clarification. The ndhF results indicate that Solanum sect. Holophylla is not monophyletic as traditionally defined. Part of Solanum sect. Holophylla that includes the species S. aligerum, S. pubigerum, and members of the S. nitidum group [Knapp, 1989; equivalent to S. subsect. Nitidum A. Child (Child, 1998)] belongs to the Dulcamaroid clade. At least part of the remainder of Solanum sect. Holophylla, represented in the ndhF trees by S. argentinum, belongs to the Geminata clade. Morphological synapomorphies of the Dulcamaroid clade may include vining habit in many taxa, the presence of unbranched, dendritic, or echinoid hairs, 3- to many-foliate sympodial units, and fruits lacking stone cell aggregates.

The following clades form a large group in Solanum with 98% bootstrap support (Fig. 1). Although the majority of species in this group belong to the spiny Solanum subg. Leptostemonum (the Leptostemonum clade), four other predominantly non-spiny clades are represented here. This group is morphologically heterogeneous and has not been recognized formally at any rank.

8. Wendlandii/Allophyllum clade

ca. 10 spp., New World

Included taxa:

Solanum sect. Allophyllum (Child) Bohs*

Solanum subg. Leptostemonum pro parte

Solanum sect. Aculeigerum Seithe*

This clade is perhaps the most unusual and surprising in all of Solanum. Thus far it consists of two groups whose relationships to other Solanum taxa have been debated. Species of Solanum sect. Allophyllum were previously placed in the genus Cyphomandra (D'Arcy, 1973: Child, 1984b; Bohs, 1988), but Bohs (1989) showed that they did not have the characters of the Cyphomandra clade. The subgeneric placement of Solanum sect. Allophyllum, however, has been obscure (Bohs, 1990). Solanum sect. Aculeigerum has usually been placed in subgenus Leptostemonum because the plants bear spines (D'Arcy, 1972, 1991; Whalen, 1984). However, they lack stellate hairs, a hallmark of the subgenus, so some workers have placed this section in with the non-spiny species of Solanum in either subgenus Solanum (Seithe, 1962) or Potatoe (Child, 1990; Child & Lester, 2001). Molecular data of Bohs and Olmstead (1997, 1999, 2001) showed that Solanum sect. Aculeigerum probably does not belong in the spiny Solanum subq. Leptostemonum, but is instead allied to a spineless group, section Allophyllum. The ndhF analyses presented here continue to support that placement. Species of Solanum sections Allophyllum and Aculeigerum are morphologically distinctive, but both groups have narrow, tapered anthers that dehisce by small terminal pores, exclusively unbranched hairs, and frequently have pinnately lobed leaves.

9. Cyphomandra clade

ca. 50 spp., New World

Included taxa:

Solanum sect. Pachyphylla (Dunal)

Dunal [genus Cyphomandra Sendtn.]*

Solanum sect. Cyphomandropsis Bitter*

Solanum sect. Glaucophyllum A. Child*

The association of these three sections and their relationship to *Solanum* have been controversial. From 1845 to 1995, *Cyphomandra* was recognized as a separate genus (Sendtner, 1845; Bohs, 1994, and references therein). However, molecular data establish that it is nested within *Solanum*,

and all species of Cyphomandra were transferred to Solanum in 1995 (Bohs, 1995). Solanum sect. Cyphomandropsis was considered to be part of Cyphomandra by some workers (D'Arcy, 1972; Child, 1984b; Child & Lester, 2001), whereas others maintained this group in Solanum (Bitter, 1913b; Seithe, 1962; Gilli, 1970; Danert, 1970; Morton, 1976). Within Solanum, its subgeneric placement has been debated, with Seithe (1962) placing it in subgenus Solanum and Smith and Downs (1966) and Morton (1976) placing it in subgenus Leptostemonum. Most authors have considered S. glaucophyllum to belong to Solanum sect. Cyphomandropsis, but Child (1986) removed it to its own monotypic section and placed it in subgenus Solanum. Hunziker (2001) disagreed with this view on morphological grounds and placed it within Solanum subg. Potatoe. Morphological, cytological, and molecular studies have confirmed the close association of Solanum sections Pachyphylla, Cyphomandropsis, and Glaucophyllum (Morton, 1976; Moscone, 1992; Bohs, 2001; Bohs & Olmstead, 2001), and molecular data indicate that they form a distinct clade within Solanum whose close relatives are unclear (Fig. 1).

Species of the Cyphomandra clade are woody shrubs or trees that often have enlarged or elaborated anther connectives or dorsal anther surfaces. The synapomorphy that unites this group is the presence of very large chromosomes, which have been found in all species of the clade investigated to date.

10. Geminata clade

ca. 140 spp., mainly New World

Included taxa:

Solanum subg. Solanum pro parte

Solanum sect. Geminata (G. Don) Walp.*

Solanum sect. Delitescens Hunz. & Barboza*

Solanum sect. Diamonon (Raf.)
A. Child*

Solanum subg. Minon Raf. pro parte

Solanum sect. Holophylla pro parte*

Solanum sect. Pseudocapsicum (Moench) Bitter*

Although placed by D'Arcy (1972, 1991) in separate subgenera of *Solanum*, both morphological studies (Knapp, 2002) and the *ndh*F analyses confirm that section *Geminata* and section *Pseudocapsicum* are closely related. Both groups have mainly leaf-opposed inflorescences and often 1- to 2-foliate sympodial units. Yet other elements belong to the Geminata clade, such as *S. argentinum*, *S. delitescens*, and *S. havanense*. *Solanum argentinum* has been placed in section *Holophylla*, but this group is apparently polyphyletic, with at least part of the section belonging to the Dulcamaroid clade.

The systematic position of *S. delitescens* has been unclear. Knapp (2002) includes it in her treatment of *Solanum* sect. *Geminata*, but lists it under taxa of uncertain placement. Nee (1999) included it within the heterogeneous *Solanum* sect. *Holophylla* within subgenus *Solanum*. Hunziker and Barboza (in Hunziker, 2000) created the monotypic *Solanum* sect. *Delitescens* to accommodate this species and also placed it within subgenus *Solanum*. The *ndh*F data indicate that *Solanum* sections *Geminata*, *Pseudocapsicum*, and *Delitescens* are closely related to each other and are not allied with the morelloid species that make up the core of subgenus *Solanum*.

Likewise, the affinities of Solanum havanense have been uncertain. This species occurs in Cuba and Jamaica and, according to Knapp (2002), is allied to the Jamaican species S. troyanum Urb. Knapp (2002) excluded these two species from Solanum sect. Geminata and regarded them as an isolated lineage in Solanum, which she called the S. havanense species group (Knapp, 2002). Child (1998) created the monotypic Solanum sect. Diamonon to accommodate S. havanense and hypothesized that it may belong near section Pseudocapsicum. In the ndhF trees, S. havanense belongs to the Geminata clade along with members of Solanum sections Geminata,

Pseudocapsicum, and Delitescens.

Characters that may unite the taxa of this clade include woody habit, unbranched to dendritically branched hairs, oblong anthers with large terminal pores, and fruits lacking stone cell aggregates.

11. Brevantherum clade

ca. 60 spp., New World

Included taxa:

Solanum subg. Brevantherum (Seithe) D'Arcy pro parte [Solanum subg. Minon pro parte in D'Arcy (1991)]

Solanum sect. Brevantherum Seithe*

Solanum sect. Extensum D'Arcy*

Solanum sect. Lepidotum Seithe*

Solanum sect. Stellatigeminatum A. Child*

Solanum sect. Cernuum Carvalho & G. J. Sheph.

Solanum subg. Solanum pro parte

Solanum sect. Gonatotrichum Bitter*

For the most part, this clade consists of a number of morphologically similar groups that often have stellate hairs or lepidote scales, oblong anthers with large terminal pores, and green, yellow, or purple fruits. D'Arcy (1991) used the subgeneric name *Minon* to refer to an analogous group in *Solanum*, which, however, also included elements such as sections *Holophylla* and *Pseudocapsicum* that are here referred to different clades. Since the type species of subgenus *Minon* is *S. pseudocapsicum*, which belongs to the Geminata clade, the appropriate name for the Brevantherum clade at subgeneric rank would be *Solanum* subg. *Brevantherum*.

The sections of Solanum subg. Brevantherum are not well demarcated. The three members of Solanum sect. Brevantherum (S. abutiloides, S. mauritianum, S. rugosum) sampled in the ndhF

trees do not form a monophyletic group, but additional data and sampling are needed to resolve relationships in the Brevantherum clade. There are a number of species that fall outside the traditional limits of the established sections listed above. One example is *Solanum inelegans*, placed by Nee (1999) in the polymorphic and ill-defined *Solanum* sect. *Holophylla* and evidently a member of the Brevantherum clade according to the *ndh*E data.

The odd group out from a morphological perspective is Solanum sect. Gonatotrichum (S. adscendens, S. turneroides, S. deflexum), Its placement here is surprising, because Solanum sect. Gonatotrichum has few of the characters listed above for the Brevantherum clade and has been thought to be more closely related to the Morelloid clade (D'Arcy, 1972, 1991; Nee, 1999; Child & Lester, 2001) or to Solanum sect. Pseudocapsicum of the Geminata clade (Hunziker, 2001). Molecular data indicate that Solanum sect. Gonatotrichum forms a distinct subclade within the Brevantherum clade (Fig. 1), but it clearly does not belong to the Morelloid clade. The names S. adscendens and S. deflexum may be synonymous (Nee, 1989, 1999; D'Arcy, 2001) but the two species exhibit a fair amount of sequence divergence in ndhF (1.0%) and are apparently allopatric (Bitter, 1912).

12. Leptostemonum clade

ca. 450 spp., worldwide

Includes all spiny sections and species groups except *Solanum* sect. *Aculeigerum* Seithe

Possibly includes *Solanum* sect. *Herposolanum* Bitter

Sampling to date includes at least 20 sections and 20 species groups sensu Whalen (1984)

This is the largest and most complex of the major clades of *Solanum* and encompasses the vast majority of species traditionally placed in *Solanum* subg. *Leptostemonum*. Data thus far

indicate that all the species of Solanum that bear spines form a clade with the exception of section Aculeigerum mentioned above. Nearly all members of this group have stellate hairs as well as spines. The anthers are narrow and tapered with small terminal pores that do not enlarge into longitudinal slits. Much work is still needed to reveal the phylogenetic structure within Leptostemonum clade and to interpret patterns of character evolution and biogeography within the group. A more detailed analysis of the Leptostemonum clade using ndhF and nuclear ITS sequence data is under way (L. Bohs, unpublished data) and will be summarized in a later publication.

The ndhF data indicate members of Solanum sections Nemorense ls. nemorense) and Herposolanum (S. hoehnei) may represent the basalmost branches in the Leptostemonum clade, but the bootstrap support for this grouping is low (51%). These taxa are similar to Solanum sect. Aculeigerum in that they have spines but lack stellate hairs. The placement of Solanum sect. Herposolanum has been particularly problematic; D'Arcy (1972, 1991) put it into Solanum subg. Bassovia, whereas Child (1983) suggested a relationship with Solanum sect. Aculeigerum (the Wendlandii/Allophyllum clade above) and provisionally placed it in Solanum subg. Potatoe (Child, 1990; Child & Lester, 2001). Whalen (1984) merged Solanum sections Herposolanum and Nemorense into his S. nemorense species group, which he considered to belong to Solanum suba. Leptostemonum. Nee (1999) included Solanum sect. Aculeigerum in section Herposolanum and regarded both as members of subgenus Leptostemonum. The ndhF data do not fully resolve these questions, but Solanum sections Herposolanum and Nemorense apparently do not belong to the Potato clade and are not closely related to section Aculeigerum.

Solanum sect. Acanthophora (S. capsicoides, S. mammosum) also appears to be relatively basal in the Leptostemonum clade. This group often has unbranched or weakly stellate hairs in addition to spines. These have been interpreted as being

reduced stellate hairs (Nee, 1979), but a thorough examination of the ontogeny of hairs in this clade should be undertaken with a phylogenetic perspective to determine if these simple hairs represent an ancestral rather than derived state in the Leptostemonum group.

GENERAL RECOMMENDATION

This is not the last word on phylogenetic structure or evolutionary relationships in Solanum. The major clades identified here, although well supported from ndhF data, need to be corroborated by data from other genes. Additional sampling, especially from morphologically unusual, underrepresented, and/or putatively isolated groups, is needed to test the distinctiveness of the major ndhF clades and to ascertain the phylogenetic position of enigmatic taxa. For instance, no molecular data are available for the two species placed in Solanum sect. Solanocharis (Bitter) A. Child. The two species may not be closely related (M. Nee, pers. comm.), and they may not belong to Solanum. The type of the section is S. albescens (Britton) Hunz., which apparently has longitudinal anther dehiscence and has been regarded by some as belonging to the genera Solanocharis, Poecilochroma, or Saracha (Rusby, 1896; Bitter, 1918; M. Nee, pers. comm.). Molecular data will certainly aid in the interpretation of this puzzling group.

Morphological and biochemical characters also should be examined, especially in the light of molecular findings, in order to identify non-molecular synapomorphies that support the *ndhF* clades. Taxonomic studies at lower levels to demarcate species limits are desperately needed for many subgeneric groups. Many nomenclatural issues also need careful clarification.

In light of these uncertainties, new formal taxonomic designations for infrageneric categories in *Solanum* are strongly discouraged without more extensive data and sampling. Progress will not be facilitated by the creation of yet more formal names that must be sifted through by all subsequent workers in the group. Informal names for species groups or clades (e.g., Whalen, 1984; Knapp, 1989, 2000, 2002; Bohs, 1994, 2001) are

encouraged until enough data have accumulated to positively demarcate and define distinct evolutionary units within *Solanum*.

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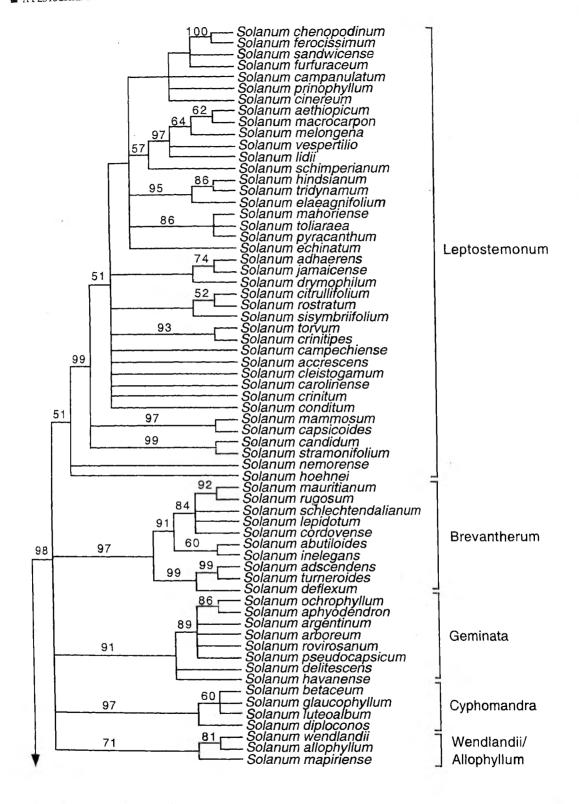


Figure 1. Strict consensus of 18,200 trees of 1053 steps from parsimony analysis of *ndh*F data. Numbers above branches are bootstrap values (500 replicates). Major clades in *Solanum* discussed in the text are labeled.

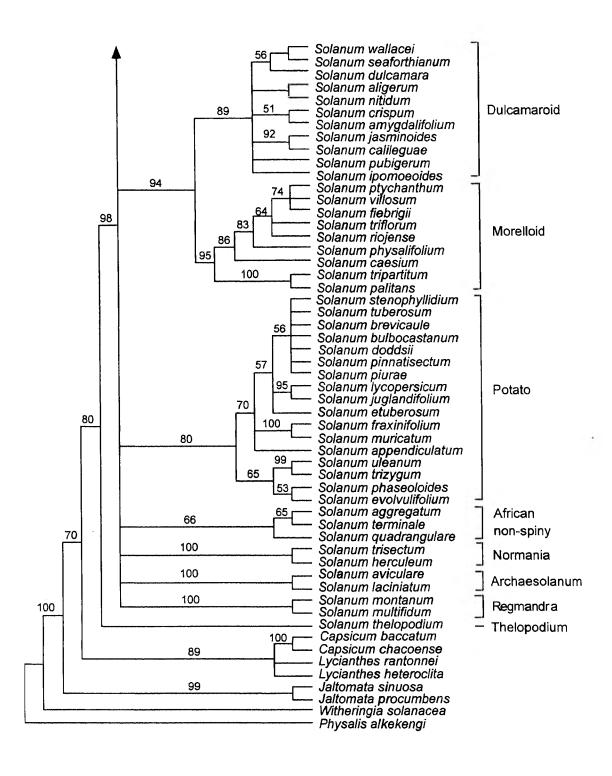


Figure 1 continued.