Whole Exome Sequencing Reveals the Genetic Basis of a Case of Idiopathic Hemolytic Anemia and Suggests Candidate Rare Variants for ADHD in a Utah Pedigree

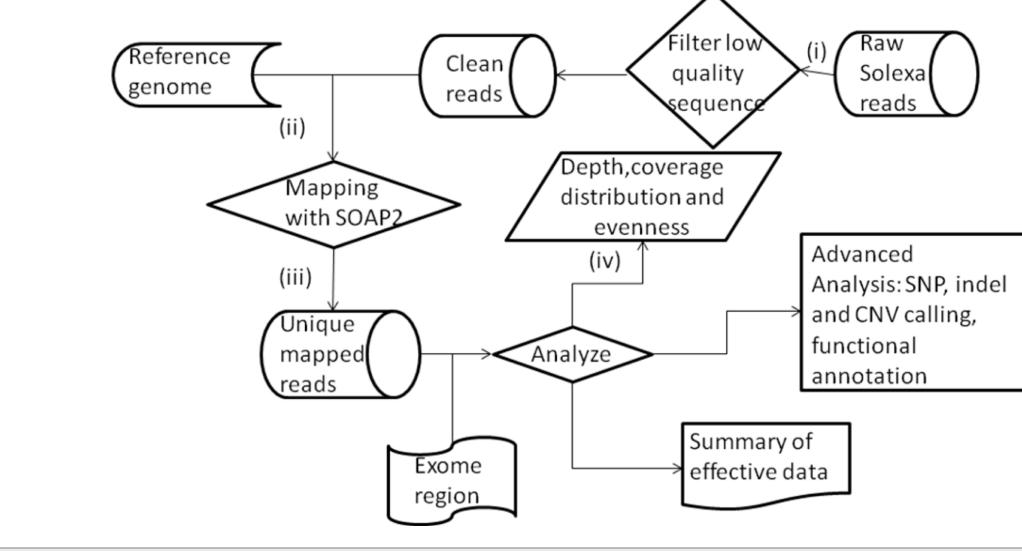
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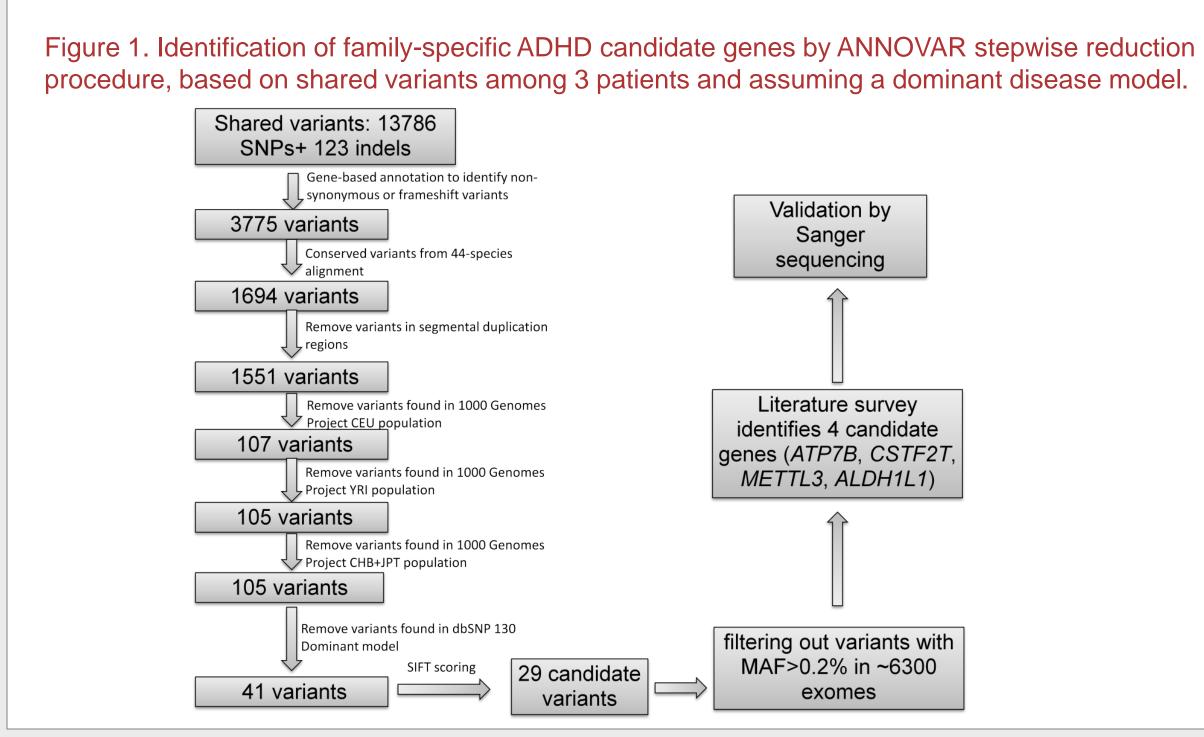
Introduction

ADHD is a common disorder affecting more than 1 in 20 children in the U.S., with as many as 50% remaining symptomatic into adulthood. Genetic factors are thought to play a large role in the etiology of the disorder, but studies thus far have been inconsistent, accounting for a small portion of the risk for ADHD. It is hypothesized that rare, family-specific genetic variants may account for the remaining missing heritability of ADHD, and advances in next-generation sequencing have made whole exome sequencing a potentially viable option for gene variant finding in complex disease pedigrees. Exome sequencing has identified the causes of nearly a dozen Mendelian disorders. Whether exome sequencing can be used to find risk variants for complex traits and has clinical utility as a diagnostic tool remain to be explored. Here we examined the utility of exome sequencing to identify candidate genes for attention deficit/hyperactivity disorder (ADHD) in a Utah pedigree in which ADHD clustered in an apparently Mendelian pattern.

Methods

Whole exome sequencing was conducted on a father and two sons with severe ADHD, combined hyperactive and inattentive subtype, all of whom improved with treatment with stimulant medication in an adult ADHD clinical trial. Exome capture was done using a commercially available Agilent SureSelect in solution method. Paired end sequencing was performed using the Illumina Genome Analyzer IIx platform with read lengths of 76 base pairs, providing at least 20x-coverage. Sequence reads were aligned to human reference genome builds hg18 and hg19 using both Novoalign 2.07 and SOAPaligner 2.20. Single nucleotide polymorphism (SNP) calling was conducted using both Maq 0.5.0 and SOAPsnp 1.03. SNP results were filtered as followed: Base quality > 20, depth from 4-200, copy number estimate < 2, and distance between two adjacent SNPs no less than 5. Nonsynonymous SNPs (nsSNPs) were identified using ANNOVAR and USeq Alleler, and insertions/deletions (indels) were detected using SOAPindel. Illumina 610k SNP microarrays were also run on each of the three samples, with copy number variant (CNV) detection using PennCNV.





Results

Table 1. Summary of SNPs for exome capture sample

ExomeCapture Number of genomic positions for calling SNPs Number of high-confidence genotypes* Number of high-confidence genotypes in target reg Number of known SNPs in target region Coverage of known SNPs

Total number of novel SNPs Total number of novel indels

Table 2. Biochemical assays of enzyme activities in the patier affected with idiopathic hemolytic anemia confirmed PKLR de

PK, pyruvate kinase; HK, hexokinase; G6PD, glucose-6-phosphate dehyd

Analysis and filtering of shared novel variants among three family members with ADHD identified 29 candidate variants. Comparison to variant data from ~6,300 exomes revealed that 12 of these variants have minor allele frequency >0.2%. In addition, one family member, who had undergone a splenectomy for idiopathic hemolytic anemia, was found to have two rare non-synonymous mutations in PKLR, consistent with a diagnosis of red blood cell pyruvate kinase deficiency. This finding was confirmed by functional biochemical testing the majority of the SNPs were validated by Sanger sequencing, and validation of the indels is underway.

Table 3. 29 candidate variants for ADHD and their population frequencies in other cohorts

# chrom	Position in HG18	Reference allele	Mutant allele	Gene	Type of Mutation	Amino acid change	# variants in BGI exomes ²	% in BGI exomes	# variants in ~600 Baylor exomes	% in Baylor exomes
chr17	64384287	A	G	ABCA8	Nonsynonymous	C1387R	0	0.00%	0	0.00%
chr11	68323378	G	А	CPT1A	Nonsynonymous	L193F	0	0.00%	0	0.00%
chr8	101063450	A	G	RGS22	Nonsynonymous	I1084T	0	0.00%	0	0.00%
chr18	59805227	G	Т	SERPINB8	Nonsynonymous	G287V	0	0.00%	0	0.00%
chr1	205267500	-	Т	C1orf116	frameshift insertion		34	1.40%	0	0.00%
chr17	42571111	-	TGGC	CDC27	frameshift insertion	5		0.20%	0	0.00%
chr17	42589406	A	-	CDC27	frameshift deletion	31		1.30%	0	0.00%
chr1	109266693	TTC	-	GPSM2	amino acid deletion		9	0.40%	0	0.00%
chr9	8577604	AT	-	HNRNPK	frameshift deletion		37	1.60%	0	0.00%
chr11	5432010	-	CA	OR5112	frameshift insertion		15	0.60%	0	0.00%
chr17	36038691	С	-	SMARCE1	frameshift deletion		19	0.80%	0	0.00%
chr7	98839005	С	-	PDAP1	frameshift deletion		37	1.60%	1	0.20%
chr18	27355154	Т	G	DSG2	Nonsynonymous	V158G	1	~0.0%	1	0.20%
chr3	127359980	G	Α	ALDH1L1	Nonsynonymous	P107L	2	~0.0%	0	0.00%
chr13	51440681	Α	G	ATP7B	Nonsynonymous	V536A	1	~0.0%	1	0.20%
chr10	53128652	Α	С	CSTF2T	Nonsynonymous	C222G	4	0.10%	1	0.20%
chr14	21041859	G	Α	METTL3	Nonsynonymous	R36W	9	0.20%	1	0.20%
chr11	76632438	-	A	GDPD4	frameshift insertion		36	1.50%	6	1.00%
chr7	86998554	A	Т	ABCB1	Nonsynonymous	S893T	815	14.3% ¹	9	1.50%
chr11	133634133	С	G	ACAD8	Nonsynonymous	S171C	112	2.00%	20	3.30%
chr20	17904347	С	Т	C20orf72	Nonsynonymous	R178W	23	0.40%	8	1.30%
chr8	33438433	Т	С	FUT10	Nonsynonymous	Q27R	15	0.30%	3	0.50%
chr13	19695025	A	Т	GJB6	Nonsynonymous	S199T	68	1.20%	4	0.70%
chr16	69572830	G	Т	HYDIN	Nonsynonymous	P1491H	77	1.40%	dozens	>5.0%
chr10	22059861	G	А	MLLT10	Nonsynonymous	R713H	15	0.30%	6	1.00%
chr17	10355994	A	G	MYH1	Nonsynonymous	Y435H	99	1.70%	14	2.30%
chr1	143727234	G	Т	PDE4DIP	Nonsynonymous	L142I	1256	22.10%	hundreds	>30.0%
chr2	98175864	Т	С	VWA3B	Nonsynonymous	I513T	15	0.30%	16	2.70%
chr5	115230317	AAGA	-	AP3S1	frameshift deletion		185	7.80%	19	3.20%

les						
	84060	84615	9	92157		
	52,105,343	52,105	,343 5	52,105,343		
	46,651,026	,026 45,555,039		46,393,954		
gions	35,571,754	34,856	,090 3	35,742,301		
	192,415	192,415		192,415		
	148,795	142,933		148,396		
	(77.33%)	(74.28%)		(77.12%)		
	25,523	24,470		25,321		
	335	325		335		
ent eficiency.		Patient 84060	Control	Reference values		
	PK (U/gHb)	3.3 L	8.6	6.1 – 12.3		
/drogenase.	HK (U/gHb)	3.2 H	1.1	0.8 - 1.5		
-	G6PD (U/gHb)	15.8 H 9.2		6.4 – 10.5		

Figure 2: Pedigree of a Utah family affected with idiopathic hemolytic anemia and ADHD.

Discussion

Preliminary results show four interesting candidate rare nonsynonymous variants in brain-expressed genes that are shared among 3 individuals (father and two sons) and thus may be associated with ADHD in this family. Validation of these findings is underway in each individual, along with expansion of the pedigree to include DNA sequence and phenotype data from the unaffected mother and other siblings, along with functional characterization of the nsSNPs and PKLR mutations. These preliminary results show that exome sequencing can identify genetic causes of mendelian disorders, as in the case of pyruvate kinase deficiency in one member of this family, and also has potential to discover rare variants associated with complex neuropsychiatric conditions such as ADHD.

Conclusion

Collectively, our results identify new candidate genes for ADHD, and suggest that exome sequencing may be an efficient strategy for solving diagnostic unknowns.

Author Affiliations

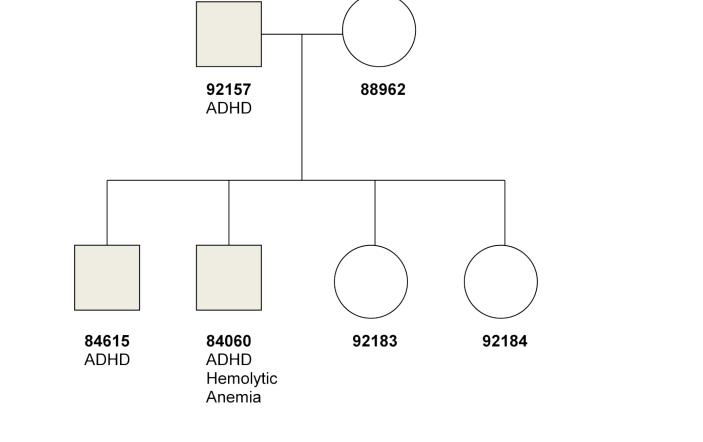
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