

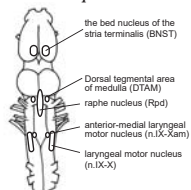
Call initiation in African clawed frogs

Ayako Yamaguchi
Biology Department, University of Utah, Salt Lake City, UT

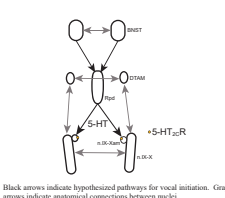
Introduction

Episodic behavior of animals are expressed only when appropriate. For example, escape behavior is expressed when an animal senses danger, and courtship behavior is expressed when the plasma levels of sex steroids are elevated and a potential mate is detected. How does the nervous system initiate episodic behavior? In our study, we use the vocalizations of African clawed frogs (*Xenopus laevis*) to address this question. During the breeding season, a male *Xenopus* generates advertisement calls. Previously, using a fictive preparation *in vitro*, we discovered that the advertisement call is generated by the central pattern generator (CPG) in the brainstem. The vocal CPG consists of a pair of premotor nuclei (DTAM, also known as parabrachial area) and a pair of laryngeal motor nuclei (n.IX-X) that are interconnected. Previously, we have shown that increasing the extracellular concentration of endogenous serotonin using a selective serotonin reuptake inhibitor elicits fictive advertisement calls by activating 5-HT_{2c} receptors expressed by the anterior-medial n.IX-X¹. The bed nucleus of the stria terminalis (BNST), a nucleus in the extended amygdala, directly projects to the dorsal raphe nucleus². A previous study showed that electrical stimulation of BNST elicits fictive advertisement calls *in vitro*, and lesioning of BNST decreases calling behavior *in vivo* in male *X. laevis*³. Based on these results, we hypothesize that advertisement calls of male *X. laevis* are initiated by BNST activating raphe nucleus, which releases serotonin that binds to 5-HT_{2c} receptors expressed by anterior-medial n.IX-X (nIX-Xam). To test this hypothesis, we obtained local field potential recordings (LFP) from n.IX-Xam.

The central vocal pathways of *Xenopus laevis*



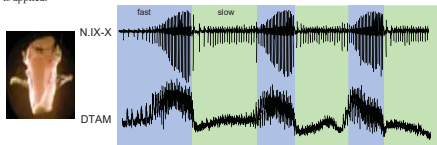
Hypothesized mechanism of call initiation



Black arrows indicate hypothesized pathways for vocal initiation. Gray arrows indicate anatomical connections between nuclei.

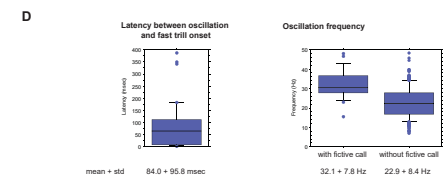
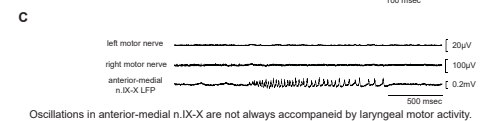
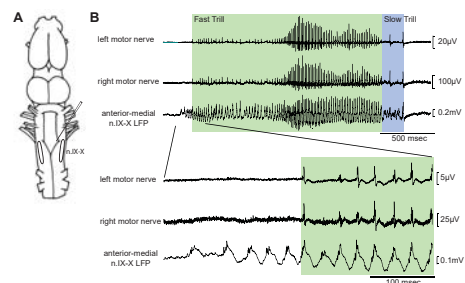
The Central Pattern Generator (CPG)

Fictive vocalizations (top trace, below) can be elicited from an isolated brainstem (left) *in vitro* when serotonin is applied.



Question: How are vocalizations initiated in the brains of male *Xenopus laevis*?

Anterior-medial n.IX-X generates oscillatory activity that precedes the onset of fictive fast trills.



When local field potential (LFP) was obtained from the anterior-medial n.IX-X (n.IX-Xam) of a brain (A) in response to 5-HT, oscillatory activity (B, top traces) was recorded that largely coincided with the fictive advertisement calls. Although the oscillation was phase-locked to motor CAPs during fictive calls, the onset of the oscillation typically preceded the onset of the fictive advertisement calls (B, bottom traces) by ~80msec on average (D left). Oscillatory activity that preceded the fictive calls had a mean frequency of ~30Hz (D right). Furthermore, the oscillatory activity was sometimes recorded in the absence of the fictive advertisement calls (C) with initial oscillatory frequency similar to those that accompany fictive calls (D, right), although the amplitude is smaller. We hypothesize that the initial part of these oscillation in the n.IX-Xam represents neural activity that initiates advertisement calls in male *X. laevis* brains.

Conclusions

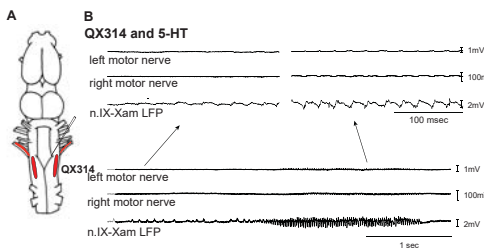
We conclude that there is a population of neurons medial to the anterior n.IX-X that likely play a role in call initiation in response to 5-HT in the *Xenopus* brainstem. Their oscillatory activity does not depend on the laryngeal motoneuron activity; the oscillations are elicited in response to 5-HT even in the absence of laryngeal motoneuron activity, or when the motoneurons are selectively silenced. In addition, these neurons are unlikely to show intrinsic oscillatory activity directly in response to 5-HT because pharmacological blockade of synaptic transmission blocks the 5-HT-induced oscillation. The anterior-medial n.IX-X expresses 5-HT_{2c} receptors that are known to initiate fictive vocalizations *in vitro*. Thus, we suggest that activation of 5-HT_{2c} receptors by the neurons in IX-Xam results in oscillatory activity that initiates the fictive vocalizations in ~80msec. We plan to use otogenetic tools (shown on the right column of this poster) to dissect the call initiation pathways further in the future.

References

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- Moreno et al. (2012) Characterization of the bed nucleus of the stria terminalis in the forebrain of anuran amphibians. *J. Comp. Neurol.* 2012 Feb 15; 520(2):330-63.
- Hall IC, Ballagh III, Kelley DB (2013) The *Xenopus* amygdala mediates socially appropriate vocal communication signals. *J. Neurosci.* 33:14534-48.

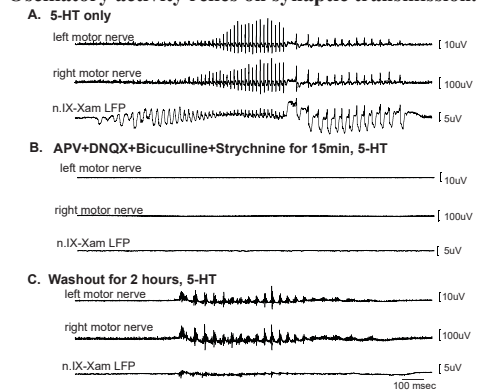
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Laryngeal motoneuron activity is not critical for generating 5-HT-induced oscillations in the anterior-medial n.IX-X.



To determine if 5-HT induced oscillations in the anterior-medial n.IX-X depend on motoneuron activity, we applied QX-314 (an intracellular Na⁺ channel blocker) to the laryngeal motor nerves bilaterally and selectively blocked the activity of all the laryngeal motoneurons (A). Application of 5-HT still elicited oscillations from the anterior-medial n.IX-X (B) although the amplitude of the oscillations was smaller compared to those obtained from control brains.

Oscillatory activity relies on synaptic transmission.



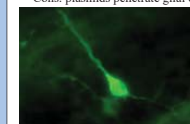
Pharmacological blockade of all synaptic transmission resulted in a loss of oscillations from the anterior-medial n.IX-X indicating that the activity is not mediated by pacemaker-like neurons responding directly to 5-HT.

Techniques: Transgene expression in *Xenopus* nervous system.

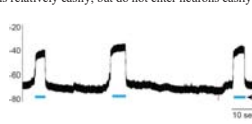
Optogenetic tools have revolutionized the field of neuroscience in recent years. However, these tools have not been available for *Xenopus laevis* because it is not a genetic model. To overcome this obstacle, we have been developing a reliable technique to express transgenes at high levels in *Xenopus* central vocal pathways. Specifically, our goal is to express optogenetic actuators (i.e., channelrhodopsin, halorhodopsin) and sensors (i.e., GCaMP6s, voltage-sensitive fluorescent proteins) in the *Xenopus* vocal neurons using either targeted electroporation or viral injection techniques.

1. Electroporation of plasmids into the *Xenopus* nervous system

Pros: plasmid can be easily obtained, fast gene expression
Cons: plasmids penetrate glial cells relatively easily, but do not enter neurons easily



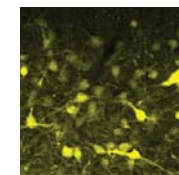
Electroporated *Xenopus* adult neuron expressing EGFP.



Electroporated *Xenopus* adult glial cell expressing a reporter gene that depolarize in response to blue light.

2. Recombinant adeno-associated virus (rAAV) injection into the *Xenopus* nervous system

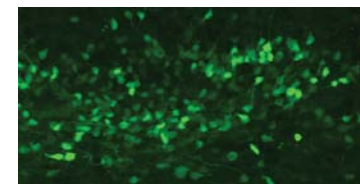
Pros: rAAV with a variety of transgenes can be readily purchased
Cons: low success rates in *Xenopus* adult brains (less than 10%), take a long time to (>3 weeks)



Xenopus adult neurons expressing reporter gene (EYFP) after injection of pAAV-hsChR2(H134R)-EYFP.

3. Vesicular stomatitis virus injection into the *Xenopus* nervous system

Pros: fast expression (6 hours at room temperature), trans-synaptic virus with presynaptic direction-specificity (anterograde or retrograde).
Cons: BSL2, appear to have high toxicity to neurons.



Xenopus adult neurons expressing reporter gene (EGFP) after injection of VSV-G VSV.