Received: 31 May 2019 Accepted: 22 August 2019

DOI: 10.1002/mrd.23262

REVIEW ARTICLE

brought to you by T CORE

Molecular Reproductic Development

Could circRNA be a new biomarker for pre-eclampsia?

Rana Shafabakhsh¹ | Naghmeh Mirhosseini² | Shala Chaichian^{3,4} | Bahram Moazzami⁴ | Zahra Mahdizadeh⁵ | Zatollah Asemi¹

¹Research Center for Biochemistry and Nutrition in Metabolic Diseases, Kashan University of Medical Sciences, Kashan, Iran

²School of Public Health, University of Saskatchewan, Saskatoon, Saskatchewan, Canada

³Minimally Invasive Techniques Research Center in Women, Tehran Medical Sciences Branch, Islamic Azad University, Tehran, Iran

⁴Pars Advanced and Minimally Invasive Medical Manners Research Center, Pars Hospital, Iran University of Medical Sciences, Tehran. Iran

⁵Firoozabadi Clinical Research Development Unit, Iran University of Medical Sciences, Tehran, Iran

Correspondence

Zatollah Asemi, Research Center for Biochemistry and Nutrition in Metabolic Diseases, Kashan University of Medical Sciences, Kashan, PO Box: 81151-87159, Iran. Email: asemi_r@yahoo.com

Shala Chaichian, Minimally Invasive Techniques Research Center in Women, Tehran Medical Sciences Branch, Islamic Azad University, Tehran, Iran. Email: shchaichian@gmail.com

1 | INTRODUCTION

Abstract

Pre-eclampsia is a devastating complication of pregnancy which is characterized by hypertension and proteinuria in pregnant women. Pre-eclampsia is important as it is the leading cause of death. Moreover, untreated pre-eclampsia might lead to other lethal complications, for both fetus and mother. Pre-eclampsia can also affect the quality of life in affected women. Despite a large number of risk factors for preeclampsia, these risk factors are able to detect just 30% of women who are susceptible to pre-eclampsia. Heterogeneous manifestations of pre-eclampsia necessitate the discovery of potential biomarkers required for its early detection. Circular RNAs (circRNAs) are a type of RNA which are more abundant, specific, and highly organized compared with other types of RNA. Accordingly, circRNAs have been suggested as one of the potential biomarkers for different diseases. Recently, researchers have shown interest in the effects of circRNAs in pre-eclampsia, although the current evidence is limited. The majority of obstetricians are probably not aware of circRNAs as a useful biomarker. Here, we aimed to summarize recent supporting evidence and assess the mechanisms by which circRNAs are involved in preeclampsia.

KEYWORDS

circRNA, diagnosis, prediction, pre-eclampsia, pregnancy

Pre-eclampsia is a pregnancy complication which affects about 3–5% of pregnancies. It is characterized by the occurrence of hypertension and proteinuria in pregnant women (Mol et al., 2016). Hypertension in pre-eclampsia is defined as systolic blood pressure ≥140 mmHg and diastolic blood pressure ≥90 mmHg occurred after 20 weeks of gestation (Damian, Njau, Lisasi, Msuya, & Boulle, 2019). This disease leads to 70,000–80,000 maternal and 500,000 perinatal mortality per year worldwide (Hutcheon, Lisonkova, & Joseph, 2011). This pregnancy-related syndrome is a major cause of neonatal, fetal, and maternal death (Saleem et al., 2014). Untreated pre-eclampsia might have other lethal consequences, including stroke, renal failure, liver rupture, and pulmonary edema as well as fetal growth retardation and preterm birth (Souza et al., 2013). Preterm birth caused by pre-eclampsia may result in bronchopulmonary dysplasia in children

(Hansen, Barnés, Folkman, & McElrath, 2010). In addition, preeclampsia increases the risk of postpartum depression and reduces the quality of life in women (Blom et al., 2010; Prick et al., 2015). In developing countries, pre-eclampsia is the leading cause of maternal mortality, while in developed countries, enhanced diagnostic methods and earlier detection make pre-eclampsia less devastating (Ngoc et al., 2006).

There is no approved preventive method for pre-eclampsia, though screening and early detection of high-risk women may improve its management. The major screening is the assessment of clinical risk factors, including age, body mass index (BMI) and family history, especially in the first half of the pregnancy (Hyde & Thornton, 2013). Delivery is the only treatment that is thought to be effective for this disorder. Identifying the exact cause of preeclampsia is likely to result in a significant decrease in maternal and perinatal morbidity and mortality. However, since its etiology remains unknown, acting effectively for preventing its development (primary prevention) is not possible. Several factors such as genetic changes have been suggested to play a key role in the pathogenesis of pre-eclampsia. Several gene variants which are involved in some metabolic processes such as inflammation, thrombophilia, rennin–angiothensin system, and oxidative stress have been associated with pre-eclampsia (Jebbink et al., 2012; Rana, Karumanchi, & Lindheimer, 2014). The common symptoms of pre-eclampsia include headache, epigastric pain, visual impairments, nausea, and vomiting as well as some neurological, hepatic, renal, and cardiovascular complications such as the ischemic neurological deficit, liver dysfunction, acute renal disease, myocardial ischemia, and pulmonary edema. Moreover, severe pre-eclampsia may lead to fetal complications, including growth retardation, neonatal death, stillbirth, and prematurity (Mol et al., 2016).

Pre-eclampsia risk factors include history of pre-eclampsia in previous pregnancy, chronic kidney diseases, high blood pressure, diabetes, autoimmune disorders such as systematic lupus erythromatosus, first pre-eclampsia, age over 40 years, pregnancy interval of more than 10 years, BMI >35 kg/m², polycystic ovarian syndrome, and multiple gestation (National Collaborating Center for Women's and Children's Health, 2010). However, these risk factors are able to detect just 30% of women who are prone to pre-eclampsia (Saleem et al., 2014). Due to heterogeneous manifestations of pre-eclampsia, potential biomarkers are required for its early detection.

Current evidence shows the relationship between noncoding RNAs (ncRNAs) and pre-eclampsia (Ji et al., 2013). One of the wellknown types of ncRNAs is long noncoding RNAs (IncRNAs), which consist of more than 200 nucleotides. LncRNAs have many biological functions including DNA methylation, imprinting regulation, apoptosis, cell-cycle regulation, and angiogenesis (Wilusz, Sunwoo, & Spector, 2009). Genome-wide IncRNAs expression patterns indicate numerous IncRNAs extensively expressed in pre-eclampsia. Previous studies have reported that different expression of IncRNAs in preeclampsia is related to multiple pathways, including trophoblast invasion (Long et al., 2016), autophagy (Gao et al., 2011, 2012), apoptosis (Song et al., 2017; Xu et al., 2018; Zou & Sun, 2015), proliferation (Cao et al., 2017; Li et al., 2018; Xu et al., 2017), migration, and angiogenesis (Chen et al., 2015; Yu et al., 2018; Zhang et al., 2015). Circular RNAs (circRNAs), as one type of ncRNAs, have been demonstrated to function as a potential regulator of other RNAs transcription like microRNAs (miRNAs). MiRNAs affect almost all cellular processes from developmental pathways to oncogenesis. Indeed, miRNAs are able to regulate the expression of the majority of human proteins (Bartel, 2004). Thus, circRNAs can be involved in a wide range of pathways in the body through regulating miRNA expression. CircRNAs as one type of ncRNAs function as a potential regulator of other RNAs transcription like miRNAs. CircRNAs are single-stranded RNAs which are produced via different posttranscriptional processes and then become circular (Ojha, Nandani, Chatterjee, & Prajapati, 2018). Previously, circRNAs were believed to be produced by errors in alternative splicing and were considered as genetic accidents (Cocquerelle, Mascrez, Hétuin, & Bailleul, 1993).

SHAFABAKHSH ET AL.

Recently, by advancement in RNA sequencing and bioinformatics methods, potential biological activities of circRNAs have been approved (Danan, Schwartz, Edelheit, & Sorek, 2012). circRNAs have a regulatory role in gene expression as well as a pivotal role in biological development mechanisms, including regulating miRNA expression, endogenous RNAs, and act as a biomarker in the diagnosis of different diseases (Liu, Pan, Mao, Liu, & Chen, 2019). For instance, a large number of circRNAs have been shown to be involved in many normal processes including development of embryos, and sperm as well as pathological conditions like myocardial infarction, Alzheimer's disease, diabetes, and carcinomas such as ovarian, breast, and endometrial tumors (Greene et al., 2017; Liu et al., 2019; Szabo et al., 2015; Vausort et al., 2016; Zheng et al., 2016).

CircRNAs do not have 3' and 5' ends and are protected against RNA exonucleases. This unique structure has produced a highly stable marker in nature. CircRNAs have many miRNA binding sites which help them to act as a miRNA sponge leading to a reduction in the inhibitory effect of miRNAs on target genes and thereby enhancing the expression of these genes (Memczak et al., 2013). CircRNAs are more abundant, more specific, and highly organized compared with other RNAs. Accordingly, researchers have suggested that circRNAs could be strong biomarkers for different diseases; however, the exact mechanisms of circRNAs function are not clear. Recent evidence has indicated that these molecules may play important roles in various diseases from cardiovascular and neurological disorders to multiple neoplasms (Qu et al., 2015). Recently, researchers have shown interest in the role of circRNAs in pre-eclampsia and several studies have been conducted herein. However, most obstetricians are probably not aware of circRNAs. Here, we aimed to summarize recent supporting evidence and evaluate the mechanisms by which circRNAs are involved in pre-eclampsia.

2 | CircRNAs BIOGENESIS

During the alternative splicing process, some sequences which are named noncoding introns, are removed from pre-mRNA to form the mature linear RNA (Wang et al., 2008). CircRNAs are produced during a noncanonical form of alternative splicing, although its detailed mechanism has not been understood yet. Based on recent findings, back splicing could be the possible mechanism for circularization of RNA (Liu et al., 2017). Back splicing leads to the formation of exonic circRNAs (EcircRNAs), circular intronic RNAs (ciRNAs), and exon-intron circRNAs (ElcircRNAs; Li et al., 2015; Zhang et al., 2013). CircRNA is produced by special alternative splicing through exon circularization and intron circularization. There are two models for circRNA formation via exon circularization, including intron-pairing-driven circularization and lariat-driven circularization (Jeck et al., 2013). In the lariat-driven circularization, a covalent splice is formed from the 3' end of the splice donor to the 5' end of the splice recipient, resulting in a lariat structure which then is circularized to form an exonic circle. The intron-pairing-driven

Development

depends on the pairing of complementary motifs existing in the transcriptome (Ye, Chen, Liu, Zhu, & Fan, 2015).

The formation process of intronic circRNA is completely different from EcircRNA. GU-rich sequences near the 5' splice site and the C-rich sequences adjacent to the branch point bind together to form a circle structure followed by the function of spliceosome which cuts out the intronic and exonic sequences. Recent investigations have indicated that back splicing is regulated by splicing factors and *cis*-elements (Wang & Wang, 2015). Eventually, introns bind together to produce ciRNAs (Talhouarne & Gall, 2014). Extracellular vesicles transport circRNAs same as ncRNAs to the extracellular space. Moreover, other cells take up extracellular vesicles showing that circRNAs enhance cell-cell communication (Lasda & Parker, 2016). By the progression of gestation in each pregnancy, the levels of extracellular vesicles increase in the maternal plasma (Adam et al., 2017). These extracellular vesicles contain several proteins, including messenger RNAs (mRNAs) and miRNAs which have various biological functions and some of them are involved in preeclampsia (Salomon, Yee, Mitchell, & Rice, 2014). Recently, the role of circRNAs and extracellular vesicles in the pathogenesis of pre-eclampsia has been attractive for researchers (Figure 1).

3 | THE PATHOPHYSIOLOGY OF PRE-ECLAMPSIA

There are many investigations in terms of the pathophysiology of pre-eclampsia, though the certain mechanisms remain unclear. The pathophysiology of pre-eclampsia seems to be multifactorial and heterogeneous. The common feature of pre-eclampsia is abnormal placentation. Normally, during pregnancy the villous cytotrophoblasts occupy the inner layer of the myometrium, while maternal spiral arteries lose their endothelium and muscle fibers with subsequent conversion into low resistance vessels. The invasion of cytotrophoblast cells into the maternal spiral arteries is not similar in pre-eclampsia. Currently, there are three theories for the etiology of pre-eclampsia; one theory emphasizes the role of ischemia-reperfusion which leads to oxidative stress and vascular disorders. Hypoperfusion of the fetoplacental unit with subsequent hypoxia leads to increased production of oxygen species and inflammatory cytokines by the placenta resulting in inflammation and endothelial dysfunction as well as the expression of clinical features of the disease, especially in the third trimester.



FIGURE 1 CircRNA formation. CircRNA, circular RNA; ciRNA, circular intronic RNA; EcircRNA, exonic circRNA; ElcircRNA, exon-intron circRNA; mRNA, messenger RNA [Color figure can beviewed at wileyonlinelibrary.com]

Endothelial dysfunction is the most frequent pathological feature of pre-eclampsia presents as increased endothelial cell permeability and aggregation of platelet (Wang, Gu, Zhang, & Lewis, 2004). Endothelial dysfunction lowers the production and activation of prostaglandins and nitric oxide which are the two famous vasodilators. Subsequently, decreased production of vasodilators (prostaglandins and nitric oxide) may enhance the activation of platelets on the inner lining of spiral arteries. Endothelial dysfunction and platelet aggregation may lead to increased production of thrombin and fibrin especially in utero-placental circulation (Redman, 1990). On the other hand, the suppression of nitric oxide synthesis in the mother leads to the prevention of embryo implantation (Durán-Reyes et al., 1999). Additionally, disruption of endothelial integrity impairs sodium volume homeostasis and consequently many cardiovascular parameters such as elevated cardiac output and intravascular volume. Moreover, alterations in circulating angiogenic and antiangiogenic factors have been reported including increased levels of soluble endoglin and soluble fms-likely tyrosine kinase-1 as antiangiogenic factors, besides reduced levels of vascular endothelial growth factor (VEGF) and placental growth factor (PIGF) as proangiogenic factors (Nilsson, Salonen Ros, Cnattingius, & Lichtenstein, 2004).

The second theory is immune system alterations. Although immune system changes are associated with the origin of preeclampsia, other factors, including proinflammatory cytokines, neutrophil activation, and endothelial dysfunction, are also related to the pathophysiology of this syndrome. Increased generation of immune cells leads to the production of tumor necrosis factor-a (TNF- α) which stimulates apoptosis in the extravillous cytotrophoblasts (Genbacev, DiFederico, McMaster, & Fisher, 1999). Furthermore, the human leukocyte antigen (HLA) is involved in the invasion of the spiral arteries. In normal gestation, these cells interact with the trophoblast due to the production of VEGF and PIGF by natural-killer cells. In pre-eclampsia, these factors are elevated while other HLAs such as HLA-G and HLA-E have been reduced (Colbern, Chiang, & Main, 1994). Moreover, recent evidence has demonstrated that heme oxygenase-1 and its metabolite, carbon monoxide, have a protective role in pregnancy which are identified as appropriate targets for the treatment of pre-eclampsia (Ahmed, 2011). Furthermore, immunologists believe that pre-eclampsia is a maternal-fetal immune maladaptation. Fetal or maternal immune maladaptation leads to the induction of apoptosis as well as increasing of various cytokines such as interleukin-2 (IL-2), interferon- γ (IFN- γ), TNF- α , and FAS ligand which are famous mediators of apoptosis (Mor, Straszewski, & Kamsteeg, 2002).

In the pathogenesis of pre-eclampsia, increased placental apoptotic debris may play an important role through deteriorating inflammatory and immune responses. Also, syncytiotrophoblast microparticles can attract monocytes and neutrophiles which lead to increased levels of TNF, IL-12, and superoxide radicals (Aly et al., 2004; Sacks, Redman, & Sargent, 2003). In addition, increased apoptosis leads to the activation of macrophages (Mor et al., 2002). IL-12 which is derived from macrophages or monocytes has the main

role in promoting T-helper-1 reactions during pre-eclampsia. IL-12 can significantly stimulate IFN- γ production from natural-killer cells and naive T cells. Interestingly, IFN-y induces monocytes to more release of IL-12 which leads to rapid decadence in severe preeclampsia (Sacks et al., 2003). Taken together, maternal-fetal immune maladaptation may increase the risk of pre-eclampsia by triggering a systematic inflammatory response in mothers. The third theory is the genetic theory which suggests multiple susceptible genes which might affect cardiovascular and hemostatic systems and inflammation. These genes including HLA-G, the angiotensin-converting enzyme (I/D) and Type 1 angiotensin II receptor (AGTR1 A1166C) are also associated with several other genes such as angiotensinogen and endothelial nitric oxide synthase (Mütze, Rudnik-Schoneborn, Zerres, & Rath, 2008; Nilsson et al., 2004). Polymorphism in some genes such as inflammatory regulators leads to altered transcription of cytokines or some antiangiogenic agents (Story & Chappell, 2017). Altogether, pre-eclampsia is a multifactorial disease in which a wide range of factors are involved from genetics to inflammatory and immune responses.

4 | CircRNAs AND PRE-ECLAMPSIA: LATEST EVIDENCE

Zhang, Yang, Long, & Li (2016) investigated the expression of circRNAs in the blood of 82 pregnant women who were between 8 and 20 gestation weeks with the aim of identifying the role of circRNAs in early diagnosis of pre-eclampsia. Pre-eclampsia was detected in 41 subjects while other 41 individuals had a healthy pregnancy. Human circRNA microarray was used to analyze the samples. In addition to circRNA, protein factor endoglin concentration was also measured. The results showed that the blood concentration of circ_101222 in patients with pre-eclampsia was remarkably greater than healthy pregnant women. Circ_101222 in combination with endoglin had the sensitivity of 0.7073 and the specificity of 0.8049 for prediction of pre-eclampsia. This study introduced a novel method for prediction of pre-eclampsia, using a specific circRNA and a protein factor.

Another recent study was conducted with the same purpose using the placental tissues of pregnant women with pre-eclampsia (Qian et al., 2016). The results of microarray analysis detected three circRNAs which were highly upregulated in pre-eclampsia, including hsa_circR-NA_100782, hsa_circRNA_102682, and hsa_circRNA_104820. This study suggested that circRNAs might take part in the pathogenesis of pre-eclampsia through activating miRNA sponges (Qian et al., 2016). Indeed, identified circRNAs have several binding sites for miRNA-17 suggesting that these circRNAs are able to regulate the expression of miRNA-17 in human placental tissues. miRNA-17 has been introduced as an angiogenesis-related miRNA, which is expressed highly in preeclampsia (Wang et al., 2012). A previous study indicated that increased expression of miRNA-17 in the placenta enhanced developing preeclampsia through contributing to trophoblast invasion (Chen & Wang, 2013). Thus, differential expression of circRNAs in the placenta may lead to upregulation of miRNA-17 via activating miRNA sponge method, thereby enhancing the pathogenesis of pre-eclampsia (Qian et al., 2016). Bai et al. (2018) investigated the profile of circRNAs in placental tissues of pre-eclampsic women, also examined the potential effects of circRNAs dysregulation on the progression of pre-eclampsia. They identified 300 circRNAs differently expressed between women with pre-eclampsia and healthy pregnant women. The results of real-time quantitative polymerase chain reaction showed that in all patients with pre-eclampsia, hg38_circ_0014736 and hsa_circ_0015382 were highly upregulated and hsa_circ_0007121 was downregulated. Data revealed that these three circRNAs are significantly associated with the regulation of transcription, proliferation, response to hypoxia, and protein binding. This study introduced hsa_circ_0007121 as the circRNA which was expressed differently between pre-eclampsia and normal pregnancy before 20 gestation weeks. So, hsa circ 0007121 could be an appropriate noninvasive biomarker for the prediction of pre-eclampsia.

In another analysis, Zhou et al. (2018) found that 49 circRNAs differently expressed in placental tissues of women with pre-eclampsia, compared to healthy pregnant women. Two of them were upregulated and others downregulated in pre-eclampsic cases. These three circRNAs, including circRNA_3286, circRNA_5593, and circRNA_3800, participated in several cellular functions in pre-eclampsia through activating miRNA sponge (Zhou et al., 2018). In the latest related investigation, the expression of several mRNAs, ncRNAs, and circRNAs was profiled in women with pre-eclampsia and healthy pregnant women to determine a potential prediction marker for pre-eclampsia. Among all of the detected markers, hsa_circ_0036877 was observed to be a potent blood biomarker for pre-eclampsia (Hu et al., 2018). Recently, a bioinformatic analysis reported hsa_circ_0004904 and hsa_circ_0001855 as the two biomarkers involved in the pathogenesis of pre-eclampsia, through activating miRNA sponges which directly target pregnancy-associated plasma protein-A (PAPP-A). According to previous evidence, low levels of PAPP-A have been documented in the serum of pre-eclampsic women (Jiang et al.,

2018). There are evidence demonstrating the existence of PAPP-A in the serum of pre-eclampsic women (Jiang et al., 2018). Taken together, the above data indicates that circRNAs are differently expressed in pre-eclampsia which provides new insight into the novel biomarkers for the prediction of pre-eclampsia. Currently, the exact reason why circRNAs levels change during pre-eclampsia is not completely understood. One of the main functions of circRNAs is the activation of miRNA sponge which is involved in the pathogenesis of several diseases. On the other hand, some studies have suggested that circRNAs might play an important role in the pathogenesis of pre-eclampsia through activating miRNA sponge method (Jiang et al., 2018; Qian et al., 2016; Zhou et al., 2018). Thus, we hypothesize that alterations in the levels of circRNAs may be involved in the pathogenesis of pre-eclampsia and associated with its incidence. However, further investigations are required to evaluate possible pathways in pre-eclampsia related to circRNAs (Table 1; Figure 2).

5 | CONCLUSIONS

Pre-eclampsia is a serious complication of pregnancy which is one of the main causes of neonatal, fetal, and maternal death. Untreated pre-eclampsia can develop many other lethal complications such as fetal growth retardation and preterm birth. However, several risk factors have been determined for this disease, which might help with its diagnosis

Still, its early diagnosis and prediction is the main issue for obstetricians. In recent years, circRNAs have attracted more attention for the prediction of diseases and also several studies have conducted to evaluate their roles in pre-eclampsia. Researchers could successfully identify several circRNAs, which are differently expressed during pre-eclampsia. These data revealed that circRNAs could be promising biomarkers in the prediction and diagnosis of

TABLE 1 Experimental studies that investigated the role of circRNAs in pre-eclampsia

circ_101222Between 8 and 20 weeksUpregulated2016Zhang et al. (2016)hsa_circRNA_100782Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hsa_circRNA_104820Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hsa_circRNA_102682Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hg38_circ_0014736Before 20 weeksUpregulated2018Bai et al. (2018)hsa_circ_0015382Before 20 weeksUpregulated2018Bai et al. (2018)
hsa_circRNA_100782Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hsa_circRNA_104820Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hsa_circRNA_102682Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hg38_circ_0014736Before 20 weeksUpregulated2018Bai et al. (2018)hsa_circ_0015382Before 20 weeksUpregulated2018Bai et al. (2018)
hsa_circRNA_104820Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hsa_circRNA_102682Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hg38_circ_0014736Before 20 weeksUpregulated2018Bai et al. (2018)hsa_circ_0015382Before 20 weeksUpregulated2018Bai et al. (2018)
hsa_circRNA_102682Between 30 and 34 weeksUpregulated2016Qian et al. (2016)hg38_circ_0014736Before 20 weeksUpregulated2018Bai et al. (2018)hsa_circ_0015382Before 20 weeksUpregulated2018Bai et al. (2018)
hg38_circ_0014736 Before 20 weeks Upregulated 2018 Bai et al. (2018) hsa_circ_0015382 Before 20 weeks Upregulated 2018 Bai et al. (2018)
hsa_circ_0015382 Before 20 weeks Upregulated 2018 Bai et al. (2018)
hsa_circ_0007121 Before 20 weeks Downregulated 2018 Bai et al. (2018)
circRNA_3286 Undergone cesarean section Downregulated 2018 Zhou et al. (2018)
circRNA_5593 Undergone cesarean section Downregulated 2018 Zhou et al. (2018)
circRNA_3800 Undergone cesarean section Downregulated 2018 Zhou et al. (2018)
hsa_circ_0036877 24 weeks Downregulated 2018 Hu et al. (2018)
hsa_circ_0004904 Before 20 weeks Upregulated 2018 Jiang et al. (2018)
hsa_circ_0001855 Before 20 weeks Upregulated 2018 Jiang et al. (2018)

Abbreviation: circRNA, circular RNA.



FIGURE 2 Factors related to circRNAs and their impact on the development of pre-eclampsia. CircRNA, circular RNA. [Color figure can beviewed at wileyonlinelibrary.com]

pre-eclampsia. However, further studies are necessary to elucidate certain mechanisms of action for circRNAs in pre-eclampsia.

CONFLICT OF INTERESTS

The authors declare that there are no conflict of interests.

AUTHOR CONTRIBUTIONS

Z. A. contributed to the conception, design, and drafting of the manuscript. R. S., S. C., and B. M. contributed in reviewing the relevant literature. N. M. was involved in drafting and editing the revised version. All authors approved the final version for submission. Z. M. was involved in the revised version. Z. A. oversaw the study.

ORCID

Zatollah Asemi 🕞 http://orcid.org/0000-0001-5265-4792

REFERENCES

- Adam, S., Elfeky, O., Kinhal, V., Dutta, S., Lai, A., Jayabalan, N., ... Salomon, C. (2017). Review: Fetal-maternal communication via extracellular vesicles -Implications for complications of pregnancies. *Placenta*, 54, 83–88.
- Ahmed, A. (2011). New insights into the etiology of preeclampsia: Identification of key elusive factors for the vascular complications. *Thrombosis Research*, 127(Suppl 3), S72–S75.
- Aly, A. S., Khandelwal, M., Zhao, J., Mehmet, A. H., Sammel, M. D., & Parry, S. (2004). Neutrophils are stimulated by syncytiotrophoblast microvillous membranes to generate superoxide radicals in women with preeclampsia. *American Journal of Obstetrics and Gynecology*, 190(1), 252–258.

- Bai, Y., Rao, H., Chen, W., Luo, X., Tong, C., & Qi, H. (2018). Profiles of circular RNAs in human placenta and their potential roles related to preeclampsia. *Biology of Reproduction*, 98(5), 705–712.
- Bartel, D. P. (2004). MicroRNAs. Cell, 116(2), 281-297.
- Blom, E., Jansen, P., Verhulst, F., Hofman, A., Raat, H., Jaddoe, V., ... Tiemeier, H. (2010). Perinatal complications increase the risk of postpartum depression. The Generation R study. *BJOG*, 117(11), 1390–1398.
- Cao, C., Li, J., Li, J., Liu, L., Cheng, X., & Jia, R. (2017). Long non-coding RNA Uc.187 is upregulated in preeclampsia and modulates proliferation, apoptosis, and invasion of HTR-8/SVneo trophoblast cells. *Journal of Cellular Biochemistry*, 118(6), 1462–1470.
- Chen, D. B., & Wang, W. (2013). Human placental microRNAs and preeclampsia. *Biology of Reproduction*, *88*(5), 130.
- Chen, H., Meng, T., Liu, X., Sun, M., Tong, C., Liu, J., ... Du, J. (2015). Long non-coding RNA MALAT-1 is downregulated in preeclampsia and regulates proliferation, apoptosis, migration and invasion of JEG-3 trophoblast cells. *International Journal of Clinical and Experimental Pathology*, 8(10), 12718–12727.
- Cocquerelle, C., Mascrez, B., Hétuin, D., & Bailleul, B. (1993). Mis-splicing yields circular RNA molecules. *The FASEB Journal*, 7(1), 155–160.
- Colbern, G. T., Chiang, M. H., & Main, E. K. (1994). Expression of the nonclassic histocompatibility antigen HLA-G by preeclamptic placenta. *American Journal of Obstetrics and Gynecology*, 170(5 Pt 1), 1244–1250.
- Damian, D. J., Njau, B., Lisasi, E., Msuya, S. E., & Boulle, A. (2019). Trends in maternal and neonatal mortality in South Africa: A systematic review. *Systematic Reviews*, 8(1), 76.
- Danan, M., Schwartz, S., Edelheit, S., & Sorek, R. (2012). Transcriptomewide discovery of circular RNAs in Archaea. Nucleic Acids Research, 40(7), 3131–3142.
- Durán-Reyes, G., Rocío gómez-Meléndez, M., la Brena, G. M., Mercado-Pichardo, E., Medina-Navarro, R., & Hicks-Gómez, J. J. (1999). Nitric oxide synthesis inhibition suppresses implantation and decreases cGMP concentration and protein peroxidation. *Life Sciences*, 65(21), 2259–2268.
- Gao, W., Li, D., Xiao, Z., Liao, Q., Yang, H., Li, Y., ... Wang, Y. (2011). Detection of global DNA methylation and paternally imprinted H19 gene methylation in preeclamptic placentas. *Hypertension Research*, 34(5), 655–661.
- Gao, W., Liu, M., Yang, Y., Yang, H., Liao, Q., Bai, Y., ... Wang, Y. (2012). The imprinted H19 gene regulates human placental trophoblast cell proliferation via encoding miR-675 that targets Nodal Modulator 1 (NOMO1). RNA Biology, 9(7), 1002–1010.
- Genbacev, O., DiFederico, E., McMaster, M., & Fisher, S. J. (1999). Invasive cytotrophoblast apoptosis in pre-eclampsia. *Human Reproduction*, 14(Suppl 2), 59–66.
- Greene, J., Baird, A. M., Brady, L., Lim, M., Gray, S. G., McDermott, R., & Finn, S. P. (2017). Circular RNAs: Biogenesis, function and role in human diseases. *Frontiers in Molecular Biosciences*, 4, 38.
- Hansen, A. R., Barnés, C. M., Folkman, J., & McElrath, T. F. (2010). Maternal preeclampsia predicts the development of bronchopulmonary dysplasia. *The Journal of Pediatrics*, 156(4), 532–536.
- Hu, X., Ao, J., Li, X., Zhang, H., Wu, J., & Cheng, W. (2018). Competing endogenous RNA expression profiling in pre-eclampsia identifies hsa_circ_0036877 as a potential novel blood biomarker for early preeclampsia. *Clinical Epigenetics*, 10, 48.
- Hutcheon, J. A., Lisonkova, S., & Joseph, K. S. (2011). Epidemiology of preeclampsia and the other hypertensive disorders of pregnancy. Best Practice & Research Clinical Obstetrics & Gynaecology, 25(4), 391–403.
- Hyde, C., & Thornton, S. (2013). Does screening for pre-eclampsia make sense? BJOG, 120(10), 1168–1170.
- Jebbink, J., Wolters, A., Fernando, F., Afink, G., van der Post, J., & Ris-Stalpers, C. (2012). Molecular genetics of preeclampsia and HELLP syndrome - A review. *Biochimica et Biophysica Acta (BBA)*, 1822(12), 1960–1969.

- Jeck, W. R., Sorrentino, J. A., Wang, K., Slevin, M. K., Burd, C. E., Liu, J., ... Sharpless, N. E. (2013). Circular RNAs are abundant, conserved, and associated with ALU repeats. RNA, 19(2), 141–157.
- Ji, L., Brkić, J., Liu, M., Fu, G., Peng, C., & Wang, Y. L. (2013). Placental trophoblast cell differentiation: Physiological regulation and pathological relevance to preeclampsia. *Molecular Aspects of Medicine*, 34(5), 981–1023.
- Jiang, M., Lash, G. E., Zhao, X., Long, Y., Guo, C., & Yang, H. (2018). CircRNA-0004904, CircRNA-0001855, and PAPP-A: Potential novel biomarkers for the prediction of preeclampsia. *Cellular Physiology and Biochemistry*, 46(6), 2576–2586.
- Lasda, E., & Parker, R. (2016). Circular RNAs co-precipitate with extracellular vesicles: A possible mechanism for circRNA clearance. *PLOS One*, 11(2), e0148407.
- Li, J. L., Li, R., Gao, Y., Guo, W. C., Shi, P. X., & Li, M. (2018). LncRNA CCAT1 promotes the progression of preeclampsia by regulating CDK4. European Review for Medical and Pharmacological Sciences, 22(5), 1216–1223.
- Li, Z., Huang, C., Bao, C., Chen, L., Lin, M., Wang, X., ... Shan, G. (2015). Exon-intron circular RNAs regulate transcription in the nucleus. *Nature Structural & Molecular Biology*, 22(3), 256–264.
- Liu, K. S., Pan, F., Mao, X. D., Liu, C., & Chen, Y. J. (2019). Biological functions of circular RNAs and their roles in occurrence of reproduction and gynecological diseases. *American Journal of Translational Research*, 11(1), 1–15.
- Liu, L., Wang, J., Khanabdali, R., Kalionis, B., Tai, X., & Xia, S. (2017). Circular RNAs: Isolation, characterization and their potential role in diseases. RNA Biology, 14(12), 1715–1721.
- Long, W., Rui, C., Song, X., Dai, X., Xue, X., Lu, Y., ... Ding, H. (2016). Distinct expression profiles of IncRNAs between early-onset preeclampsia and preterm controls. *Clinica Chimica Acta*, 463, 193–199.
- Memczak, S., Jens, M., Elefsinioti, A., Torti, F., Krueger, J., Rybak, A., ... Rajewsky, N. (2013). Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature*, 495(7441), 333–338.
- Mol, B. W. J., Roberts, C. T., Thangaratinam, S., Magee, L. A., de Groot, C. J. M., & Hofmeyr, G. J. (2016). Pre-eclampsia. *The Lancet*, 387(10022), 999–1011.
- Mor, G., Straszewski, S., & Kamsteeg, M. (2002). Role of the Fas/Fas ligand system in female reproductive organs: Survival and apoptosis. *Biochemical Pharmacology*, 64(9), 1305–1315.
- Mütze, S., Rudnik-Schöneborn, S., Zerres, K., & Rath, W. (2008). Genes and the preeclampsia syndrome. *Journal of Perinatal Medicine*, 36(1), 38–58.
- National Collaborating Centre for Women's and Children's Health (2010). National Institute for Health and Clinical Excellence: Guidance. Hypertension in Pregnancy: The Management of Hypertensive Disorders During Pregnancy. London: RCOG Press Royal College of Obstetricians and Gynaecologists.
- Ngoc, N. T., Merialdi, M., Abdel-Aleem, H., Carroli, G., Purwar, M., Zavaleta, N., ... Villar, J. (2006). Causes of stillbirths and early neonatal deaths: Data from 7993 pregnancies in six developing countries. Bulletin of the World Health Organization, 84(9), 699–705.
- Nilsson, E., Salonen Ros, H., Cnattingius, S., & Lichtenstein, P. (2004). The importance of genetic and environmental effects for pre-eclampsia and gestational hypertension: A family study. *BJOG: An International Journal of Obstetrics and Gynaecology*, 111(3), 200–206.
- Ojha, R., Nandani, R., Chatterjee, N., & Prajapati, V. K. (2018). Emerging role of circular RNAs as potential biomarkers for the diagnosis of human diseases. Advances in Experimental Medicine and Biology, 1087, 141–157.
- Prick, B. W., Bijlenga, D., Jansen, A. J. G., Boers, K. E., Scherjon, S. A., Koopmans, C. M., ... Duvekot, J. J. (2015). Determinants of healthrelated quality of life in the postpartum period after obstetric complications. European Journal of Obstetrics & Gynecology and Reproductive Biology, 185, 88–95.

- Qian, Y., Lu, Y., Rui, C., Qian, Y., Cai, M., & Jia, R. (2016). Potential significance of circular RNA in human placental tissue for patients with preeclampsia. *Cellular Physiology and Biochemistry*, 39(4), 1380-1390.
- Qu, S., Yang, X., Li, X., Wang, J., Gao, Y., Shang, R., ... Li, H. (2015). Circular RNA: A new star of noncoding RNAs. *Cancer Letters*, 365(2), 141–148.
- Rana, S., Karumanchi, S. A., & Lindheimer, M. D. (2014). Angiogenic factors in diagnosis, management, and research in preeclampsia. *Hypertension*, 63(2), 198–202.
- Redman, C. W. G. (1990). Platelets and the beginnings of preeclampsia. New England Journal of Medicine, 323(7), 478-480.
- Sacks, G. P., Redman, C. W. G., & Sargent, I. L. (2003). Monocytes are primed to produce the Th1 type cytokine IL-12 in normal human pregnancy: An intracellular flow cytometric analysis of peripheral blood mononuclear cells. *Clinical and Experimental Immunology*, 131(3), 490–497.
- Saleem, S., McClure, E. M., Goudar, S. S., Patel, A., Esamai, F., Garces, A., ... Goldenberg, R. L. (2014). A prospective study of maternal, fetal and neonatal deaths in low- and middle-income countries. *Bulletin of the World Health Organization*, 92(8), 605–612.
- Salomon, C., Yee, S. W., Mitchell, M. D., & Rice, G. E. (2014). The possible role of extravillous trophoblast-derived exosomes on the uterine spiral arterial remodeling under both normal and pathological conditions. *BioMed Research International*, 2014, 1–10.
- Song, X., Rui, C., Meng, L., Zhang, R., Shen, R., Ding, H., ... Long, W. (2017). Long non-coding RNA RPAIN regulates the invasion and apoptosis of trophoblast cell lines via complement protein C1q. *Oncotarget*, 8(5), 7637–7646.
- Souza, J. P., Gülmezoglu, A. M., Vogel, J., Carroli, G., Lumbiganon, P., Qureshi, Z., ... Say, L. (2013). Moving beyond essential interventions for reduction of maternal mortality (the WHO multicountry survey on maternal and newborn health): A cross-sectional study. *The Lancet*, 381(9879), 1747–1755.
- Story, L., & Chappell, L. C. (2017). Preterm pre-eclampsia: What every neonatologist should know. Early Human Development, 114, 26–30.
- Szabo, L., Morey, R., Palpant, N. J., Wang, P. L., Afari, N., Jiang, C., ... Salzman, J. (2015). Statistically based splicing detection reveals neural enrichment and tissue-specific induction of circular RNA during human fetal development. *Genome Biology*, 16, 126.
- Talhouarne, G. J. S., & Gall, J. G. (2014). Lariat intronic RNAs in the cytoplasm of Xenopus tropicalis oocytes. RNA, 20(9), 1476-1487.
- Vausort, M., Salgado-Somoza, A., Zhang, L., Leszek, P., Scholz, M., Teren, A., ... Devaux, Y. (2016). Myocardial infarction-associated circular RNA predicting left ventricular dysfunction. *Journal of the American College of Cardiology*, 68(11), 1247–1248.
- Wang, E. T., Sandberg, R., Luo, S., Khrebtukova, I., Zhang, L., Mayr, C., ... Burge, C. B. (2008). Alternative isoform regulation in human tissue transcriptomes. *Nature*, 456(7221), 470–476.
- Wang, W., Feng, L., Zhang, H., Hachy, S., Satohisa, S., Laurent, L. C., ... Chen, D. (2012). Preeclampsia up-regulates angiogenesis-associated microRNA (i.e., miR-17, -20a, and -20b) that target ephrin-B2 and EPHB4 in human placenta. *Journal of Clinical Endocrinology and Metabolism*, 97(6), E1051–E1059.
- Wang, Y., Gu, Y., Zhang, Y., & Lewis, D. F. (2004). Evidence of endothelial dysfunction in preeclampsia: Decreased endothelial nitric oxide synthase expression is associated with increased cell permeability in endothelial cells from preeclampsia. American Journal of Obstetrics and Gynecology, 190(3), 817–824.
- Wang, Y., & Wang, Z. (2015). Efficient backsplicing produces translatable circular mRNAs. RNA, 21(2), 172–179.
- Wilusz, J. E., Sunwoo, H., & Spector, D. L. (2009). Long noncoding RNAs: Functional surprises from the RNA world. *Genes & Development*, 23(13), 1494–1504.
- Xu, Y., Ge, Z., Zhang, E., Zuo, Q., Huang, S., Yang, N., ... Sun, L. (2017). The IncRNA TUG1 modulates proliferation in trophoblast cells via epigenetic suppression of RND3. *Cell Death & Disease*, 8(10), e3104.

- Development
- Xu, Y., Lian, Y., Zhang, Y., Huang, S., Zuo, Q., Yang, N., ... Sun, L. (2018). The long non-coding RNA PVT1 represses ANGPTL4 transcription through binding with EZH2 in trophoblast cell. *Journal of Cellular* and Molecular Medicine, 22(2), 1272–1282.
- Ye, C. Y., Chen, L., Liu, C., Zhu, Q. H., & Fan, L. (2015). Widespread noncoding circular RNAs in plants. New Phytologist, 208(1), 88–95.
- Yu, L., Kuang, L. Y., He, F., Du, L. L., Li, Q. L., Sun, W., ... Chen, D. J. (2018). The role and molecular mechanism of long nocoding RNA-MEG3 in the pathogenesis of preeclampsia. *Reproductive Sciences*, 25(12), 1619–1628.
- Zhang, Y., Zhang, X. O., Chen, T., Xiang, J. F., Yin, Q. F., Xing, Y. H., ... Chen, L. L. (2013). Circular intronic long noncoding RNAs. *Molecular Cell*, 51(6), 792–806.
- Zhang, Y., Zou, Y., Wang, W., Zuo, Q., Jiang, Z., Sun, M., ... Sun, L. (2015). Down-regulated long non-coding RNA MEG3 and its effect on promoting apoptosis and suppressing migration of trophoblast cells. *Journal of Cellular Biochemistry*, 116(4), 542–550.
- Zhang, Y. G., Yang, H. L., Long, Y., & Li, W. L. (2016). Circular RNA in blood corpuscles combined with plasma protein factor for early prediction of pre-eclampsia. BJOG: An International Journal of Obstetrics & Gynaecology, 123(13), 2113–2118.

- Zheng, Q., Bao, C., Guo, W., Li, S., Chen, J., Chen, B., ... Huang, S. (2016). Circular RNA profiling reveals an abundant circHIPK3 that regulates cell growth by sponging multiple miRNAs. *Nature Communications*, 7, 11215.
- Zhou, W., Wang, H., Wu, X., Long, W., Zheng, F., Kong, J., & Yu, B. (2018). The profile analysis of circular RNAs in human placenta of preeclampsia. *Experimental Biology and Medicine*, 243(14), 1109-1117.
- Zou, Y. F., & Sun, L. Z. (2015). [Long noncoding RNA HOTAIR modulates the function of trophoblast cells in pre-eclampsia]. *Sichuan da xue xue bao*. Yi xue ban, 46(1), 113–117.

How to cite this article: Shafabakhsh R, Mirhosseini N, Chaichian S, Moazzami B, Mahdizadeh Z, Asemi Z. Could circRNA be a new biomarker for pre-eclampsia? *Mol Reprod Dev*. 2019;86:1773–1780. <u>https://doi.org/10.1002/mrd.23262</u>