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## A conceptual framework for restoration of threatened plants: the effective model of American chestnut (Castanea dentata) reintroduction

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#### Summary

*New Phytologist* (2013) **197:** 378–393 **doi**: 10.1111/nph.12020

Key words: biotechnology, *Castanea dentata*, ecosystem restoration, genetic engineering, introduced pathogen, reintroduction, socio-ecological system, tree improvement.

We propose a conceptual framework for restoration of threatened plant species that encourages integration of technological, ecological, and social spheres. A sphere encompasses ideas relevant to restoration and the people working within similar areas of influence or expertise. Increased capacity within a sphere and a higher degree of coalescing among spheres predict a greater probability of successful restoration. We illustrate this with Castanea dentata, a foundation forest tree in North America that was annihilated by an introduced pathogen; the species is a model that effectively merges biotechnology, reintroduction biology, and restoration ecology. Because of *C. dentata*'s ecological and social importance, scientists have aggressively pursued blight resistance through various approaches. We summarize recent advancements in tree breeding and biotechnology that have emerged from C. dentata research, and describe their potential to bring new tools to bear on socio-ecological restoration problems. Successful reintroduction of C. dentata will also depend upon an enhanced understanding of its ecology within contemporary forests. We identify a critical need for a deeper understanding of societal influences that may affect setting and achieving realistic restoration goals. Castanea dentata may serve as an important model to inform reintroduction of threatened plant species in general and foundation forest trees in particular.

\*The first two authors are co-first authors. All three authors contributed equally to the development of the conceptual framework.

### Tansley review

A conceptual framework for restoration of threatened plants: the effective model of American chestnut (*Castanea dentata*) reintroduction

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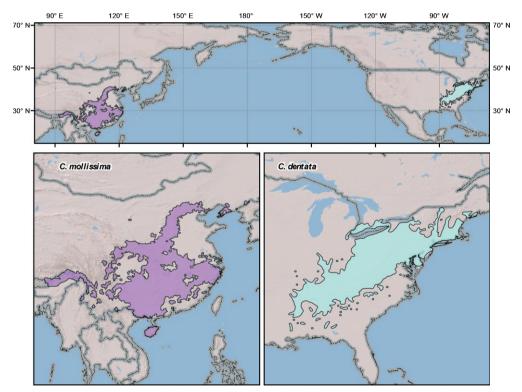
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## I. *Castanea dentata* as a model at the intersection of reintroduction biology and restoration ecology

Since the discovery of chestnut blight caused by Cryphonectria parasitica (Murr.) Barr (= Endothia parasitica (Murr.) Anderson & Anderson) in the early 1900s, many groups have worked to develop blight-resistant American chestnut (Castanea dentata (Marsh.) Borkh.); and a large-scale reintroduction program is being considered for implementation (Steiner & Carlson, 2006). The establishment of blight-resistant C. dentata in eastern forests resides at the intersection of two young disciplines: reintroduction biology and restoration ecology. Reintroduction biology has a single species focus (often a charismatic mammal or bird) and is used as a conservation tool to prevent extinction by re-establishing populations of a target species within its historical range (Seddon et al., 2007a; Armstrong & Seddon, 2008). Restoration ecology has often been botanically focused and emphasizes community and ecosystem structure and function (Young, 2000). Castanea dentata represents the reintroduction of a single, charismatic species that, because it is also a foundation species, may simultaneously restore community and ecosystem function. Recent calls for a more systematic, scientific approach to reintroduction biology and for a merging of reintroduction biology and restoration ecology (Lipsey & Child, 2007; Seddon, 2010; Polak & Saltz, 2011) can both be demonstrated in the case of C. dentata. In addition, reintroduction of C. dentata can serve as a catalyst for public discourse about the appropriate use of biotechnology (including genetic engineering (GE)) for restoration and commercial applications. Collectively, these aspects implicate *C. dentata* as an informative model that merges biotechnology, reintroduction biology, and restoration ecology (Lipsey & Child, 2007; Seddon *et al.*, 2007b; Seddon, 2010). Below, we introduce *C. dentata* and its devastation by the blight. We then present a conceptual framework for restoration that merges societal, technological, and ecological spheres of expertise and influence. Lastly, we describe the objectives of this review and how we will use the conceptual framework to synthesize *C. dentata* biology, genetics, and ecology to create a foundation for identifying restoration goals and strategies for implementation.

## 1. Loss of a foundation species caused by an introduced pathogen

Before the accidental introduction of chestnut blight, *C. dentata* was a dominant tree species throughout much of its 800 000 km<sup>2</sup> range in eastern North America (Fig. 1), sometimes comprising the majority of biomass within an exceptionally diverse forest community (Braun, 1950; Stephenson, 1986; Russell, 1987; Foster *et al.*, 2002). Particularly in the Appalachian region, *C. dentata* filled an important ecological niche and has been called a 'foundation species' because of its influence over population, community, and ecosystem processes (Youngs, 2000; Ellison *et al.*, 2005). For example, *C. dentata* was a major seed resource, producing large and consistent seed crops that drove consumer population dynamics (Dalgleish & Swihart, 2012). Because of its



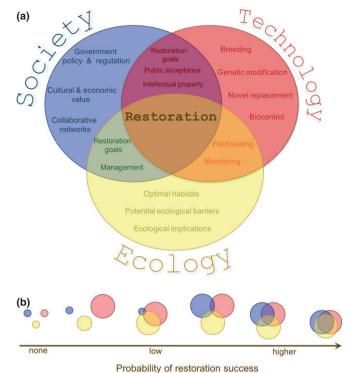
**Fig. 1** Native ranges of American chestnut (*Castanea dentata*) and Chinese chestnut (*C. mollissima*), the primary source of blight resistance genes. Frost sensitivity (Parker *et al.*, 1993) may have limited the proliferation of *C. dentata* at higher latitudes (Russell, 1987). An outbreak of the introduced soil-borne oomycete pathogen *Phytophthora cinnamomi* during *c.* 1825–1875 may have permanently retracted the southern portion of its range, which once extended as far south as Florida (Anagnostakis, 2001). By the late 1800s, most *C. dentata* in the Piedmont region of North Carolina had disappeared, while its natural range was still expanding before the introduction of the blight in other areas, for example, into Michigan (Russell, 1987; Brewer, 1995).

fast growth, high-tannin wood, and low carbon (C) : nitrogen (N) ratio leaves, *C. dentata* affected ecosystem functions such as productivity, decomposition, and nutrient cycling (Ellison *et al.*, 2005). *Castanea dentata* also filled an important economic and social niche. The strong wood of *C. dentata* is highly decay resistant and was used for telephone poles, posts, and railroad ties, as well as for furniture, construction lumber, siding, and roofing (Smith, 2000; Youngs, 2000). The wood and bark were used to produce tannin for leather production. The nuts, which are edible raw or roasted, were collected for household consumption and to sell (Youngs, 2000).

Chestnut blight caused by C. parasitica, a filamentous ascomycete fungus, rapidly annihilated C. dentata throughout its range (Anagnostakis, 1987) in the first half of the 20th Century (Hepting, 1974). The necrotrophic pathogen, first discovered in 1904 in New York City (Murrill, 1906; Anderson & Rankin, 1914), enters chestnut trees through stem wounds as germinating conidia or ascospores and colonizes the cambial zone, causing cell and tissue death, vascular dysfunction, and stem dieback (Beattie & Diller, 1954). Because it forms a perennial canker, the fungus has ample opportunity to sporulate. Conidia (asexual spores transported by water or animal movement) can form within 1 month of infection and ascospores (wind-disseminated sexual spores) within 4 months, resulting in rapid disease spread. Indeed, in < 50 yr the disease spread from New York throughout the range and killed aboveground stems of nearly every mature tree (Hepting, 1974). Castanea dentata is still a common component of eastern forests, but nearly all individuals are root-collar sprouts of blight-killed trees, which mostly originated from trees that were seedlings or saplings when blight arrived (Stephenson *et al.*, 1991; Paillet, 2002). Cycles of sprouting, infection, dieback, and re-infection may persist for decades (Paillet, 1984), yet sprouts rarely grow out of the understory or to reproductive maturity (Paillet, 2002). Since the discovery of chestnut blight, many groups have worked to develop blight-resistant C. dentata through diverse strategies including biocontrol of the fungus with mycoviruses, breeding and selection of large surviving C. dentata trees, hybrid breeding with resistant Asian chestnut species, and GE. Efforts to develop blight resistance have largely evolved separately, but are coming to fruition such that a large-scale reintroduction program is being considered for implementation (Steiner & Carlson, 2006).

#### 2. A conceptual framework for restoration

In this review, we present *C. dentata* as a model species and use it to illustrate a general conceptual framework for restoration (Fig. 2a), which contains three major spheres: *society, technology,* and *ecology.* We use 'sphere' to describe similar ideas relevant to restoration and the people working within similar areas of interest, influence, or expertise. The *society* sphere includes structures such as governmental policy and regulation, collaborative networks, and cultural or economic valuation. The *technology* sphere encompasses solutions to mitigate deficiencies in a given socioecological system. The *ecology* sphere includes targeting optimal habitats for application of the technology as well as forecasting



**Fig. 2** Conceptual framework for restoration. (a) We identify three key spheres of expertise that underpin the process of using (re)introduction of a species as a tool in restoration: the society (blue), technology (red), and ecology (yellow) spheres. The area corresponds to the capacity of each sphere. Within each sphere and area of overlap (green, orange, and purple areas), we illustrate several key roles. The degree of overlap among all three spheres, illustrated in brown, represents the probability of achieving restoration success, which we define as meeting the restoration goals set through collaboration among members in all three spheres. (b) The framework predicts that restoration success increases as the capacity of the spheres and the degree of overlap (collaboration) among the spheres increase. If only one of the spheres has large capacity, for example, restoration success remains low because the degree of overlap is low. Similarly, restoration success will be low even when all spheres are large if the

community and ecosystem effects. We identify areas that require collaboration among spheres, perhaps most critically the setting of realistic, attainable goals that are central to defining restoration success (Hobbs & Norton, 1996). *Castanea dentata*'s vast cultural and ecological significance has driven development of multiple technologies in pursuit of reintroduction (e.g. biocontrol, breeding, and GE) along with a recent initiative to improve ecological knowledge of the species.

degree of overlap is low.

Our framework predicts that the larger the capacity of each sphere (represented by the size of the circle) and the higher the degree of integration and collaboration among spheres (overlap of the circles), the greater the probability of successful restoration (Fig. 2b). Restoration programs generally are comprised of diverse groups addressing specific facets of the restoration process, sometimes covering more than one sphere, and thereby yielding a degree of overlap. We argue, however, that, for complex restoration problems to be successfully addressed at a large scale, all three spheres must be pursued collectively, simultaneously, and ideally in a coordinated manner. In the case of *C. dentata*, several groups are currently working within subsets of our conceptual model. For example, the Forest Health Initiative (FHI), building on the technical successes of the National Science Foundation's Fagaceae Project, is using *C. dentata* as a 'test tree' to create a roadmap for evaluating the use of biotechnology (including GE) in forests through an approach that combines consideration of scientific, social, regulatory and environmental issues (http://foresthealthini tiative.org/). The American Chestnut Foundation (TACF) has primarily focused on the use of breeding technologies to produce a blight-resistant tree and reflects a network of devoted members aimed at reintroducing C. dentata. Use of our conceptual framework enables a more holistic approach to restoration that integrates collaborative networks such as FHI and TACF, and underscores the centrality and necessity of explicitly stated common goals. Although C. dentata is among the best-known examples of virtual extirpation caused by an exotic pest, numerous other tree species face a similar fate (e.g. emerald ash borer (Agrilus planipennis Fairmaire) on ash (Fraxinus spp. L.) in North America; hemlock wooly adelgid (Adelges tsugae (Annand)) on eastern hemlock (Tsuga canadensis (L.) Carr.) in North America; and Dutch elm disease (Ophiostoma ulmi (Buism.) Nannf.) on elm (Ulmus spp. L.) in Europe and North America). Through effective merging of biotechnology, reintroduction biology, and restoration ecology, C. dentata may serve as an important model to inform reintroduction of threatened plant species in general and foundation forest trees in particular.

#### 3. Objectives of this review

Using our conceptual framework as a guide, we synthesize C. dentata biology, genetics, and ecology to serve as a foundation for identifying reintroduction goals and strategies for implementation. Recent reviews have detailed the C. dentata inter-species backcross breeding program as well as social and management considerations toward C. dentata restoration (Steiner & Carlson, 2006; Jacobs, 2007) and genetic implications of the backcross breeding program for restoration (Worthen et al., 2010). Others have highlighted the potential for the use of GE technologies in reintroduction programs (Merkle et al., 2007), a call for a broader use of species introductions as a tool for restoration (Seddon, 2010), and the need for a stronger connection between socioeconomic research and restoration practice (Collier, 2011). By merging ideas from across diverse literature, we place C. dentata reintroduction within the broader context of biotechnology, reintroduction biology, and restoration ecology. We trace the paths of the three main technologies at the leading edge of developing blight-resistant C. dentata (the technology sphere) and examine the key ecological changes in post-blight deciduous forests that may influence the role of C. dentata in contemporary forests (the *ecology* sphere). We then present some of the critical challenges remaining for C. dentata reintroduction, the majority of which reside within the *social* sphere of our conceptual framework. These three facets should be well developed individually, yet must effectively converge to ensure successful restoration. The importance of restoration as a conservation tool is increasing as ecosystem

degradation as a consequence of anthropogenic influences becomes more prevalent; while our framework is based upon the model of *C. dentata*, we also suggest how it applies to reintroduction of threatened plants in general.

## II. Technological approaches for recovery of *Castanea dentata*

Scientists have pursued three main approaches for mitigating chestnut blight: (1) inoculation of chestnut blight infections with hypovirulent strains of *C. parasitica*, (2) breeding *C. dentata* using both intra- and inter-species methods, and (3) GE of *C. dentata*. Below, we review these approaches and outline how they inform and are enhanced by genomic technologies. Though these efforts have largely developed independently, their ability to coalesce through technological development as well as social structures has the potential to increase the probability of restoration success (Fig. 2a).

#### 1. Biological control with hypovirulence

Hypovirulence is the reduction in virulence caused by a mycovirus (family Hypoviridae) infection of the blight fungus (Milgroom & Cortesi, 2004). The reduced virulence causes sublethal or healing cankers (Griffin, 2000; Milgroom & Cortesi, 2004). In many areas of Europe, hypovirulence has effectively controlled blight (Griffin, 2000; Milgroom & Cortesi, 2004). Hypoviruses in Europe have spread both naturally and through artificial inoculations of cankers, though it remains unclear whether human-aided deployment has significantly increased spread (Milgroom & Cortesi, 2004). The incidence of blight infection, hypovirulence, and tree mortality all vary with management, environmental conditions, and the age of the trees (Milgroom & Cortesi, 2004).

The discovery of hypoviruses within blight cankers in C. dentata populations established in Michigan (outside the C. dentata native range) fueled hopes for using hypovirulence to control blight throughout North America (Jaynes & Elliston, 1980; Fig. 3). In some Michigan populations, trees grow large and reproduce via seed with healing cankers (Milgroom & Cortesi, 2004). Michigan populations are the exception, however, and not the rule: in experimental trials in Virginia, West Virginia, Connecticut, and Wisconsin, hypoviruses failed to spread among trees and sometimes even among cankers within a tree, severely limiting the use of hypovirulence for biocontrol (Griffin, 2000; Milgroom & Cortesi, 2004). Vegetative incompatibility (vic) is commonly observed between blight fungus cultures and several vic genes have been identified and mapped (Anagnostakis, 1982; Cortesi & Milgroom, 1998; Kubisiak & Milgroom, 2006). When any one of the vic genes does not match between cultures, anastomosis and, therefore, hypovirus transfer, is largely prevented (Liu & Milgroom, 1996; Cortesi et al., 2001). Although vic is considered a leading cause of the failure of hypovirus spread in North America, questions remain concerning the effects of environmental and biological conditions on the establishment and spread of hypoviruses (Milgroom & Cortesi, 2004). It appears that biocontrol with hypovirulence alone is unlikely to control blight in North America, but it may be used as a tool in concert with host genetic resistance and optimized

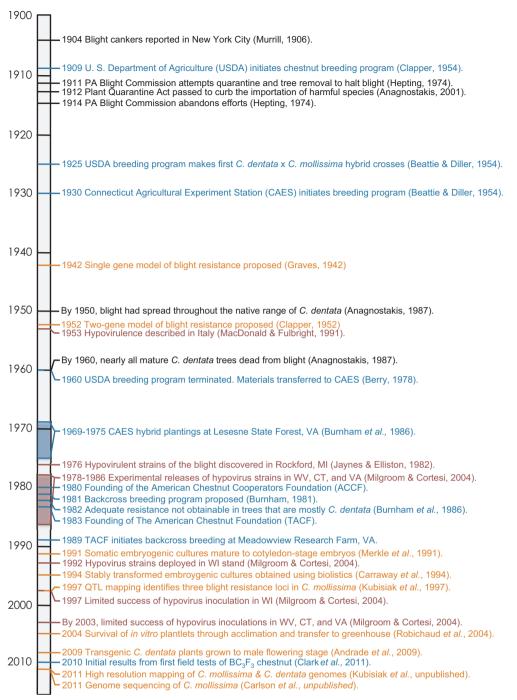


Fig. 3 Timeline showing key events in the development of blight resistance for *Castanea dentata*. Brown, blue, and orange text indicates events in the biocontrol, breeding, and genetic engineering approaches, respectively.

silviculture to achieve a high degree of blight control (Griffin, 2000).

#### 2. Breeding for blight resistance

Breeding for blight resistance has focused on (1) intra-species breeding to increase the very low frequency of naturally occurring blight resistance in populations of *C. dentata* or (2) inter-species breeding primarily with *Castanea mollissima* (Blume) (Fig. 1) to introgress Asian resistance into *C. dentata*. The American Chestnut Cooperators Foundation (ACCF) is actively pursuing intra-species breeding to complement their hypovirulence and silviculture programs. TACF is leading a large group of partners focusing on the inter-species strategy using backcross breeding while supporting research on biocontrol, silviculture, and GE methods. Breeding programs focusing primarily on inter-species approaches began shortly after the discovery of blight and have been conducted by the US Department of Agriculture (USDA; 1909–1960), the Connecticut Agricultural Experiment Station (CAES; 1930–present), and TACF (1983–present).

The USDA breeding program began in 1909 (Fig. 3) with the goal of producing blight-resistant forest trees for timber, tannins, and wildlife as well as horticultural trees for nut production (Clapper, 1954). By 1925, the USDA program made *C. dentata* × C. mollissima hybrid crosses utilizing materials collected in Asia (Beattie & Diller, 1954; Diller & Clapper, 1965). In 1960 the USDA program was discontinued and some materials were transferred to the ongoing breeding program at CAES (Berry, 1978). Work at CAES was highly collaborative with the USDA program during the years when both programs were active, using similar strategies of species hybridization and resistance testing in anticipation of finding and cloning the ideal combination of resistance from Asian chestnut species and fast growth and forest tree form from C. dentata. In 1983, TACF was founded to breed C. dentata using the backcross breeding method (Burnham, 1981; Burnham et al., 1986). Several selected genotypes from both the USDA and the CAES breeding programs formed the basis of TACF's program.

The specific steps of the backcross breeding method include making three backcross generations with selection for resistance at each generation to ensure retention of Asian resistance genes (Fig. 4), intercrossing the selected  $BC_3F_1$  trees to produce  $BC_3F_2$ populations fully segregating for resistance, selecting in the BC<sub>3</sub>F<sub>2</sub> populations for high resistance, and establishing the selections in seed orchards to produce seeds (BC3F3 and beyond) for forest planting (Hebard, 1994, 2006). As widely recognized in plant breeding, it is important to use many unrelated C. dentata trees at each generation to properly sample the native species alleles (Hebard, 2006; Worthen et al., 2010) and parent trees should originate within the region where the progeny trees will be planted to promote local adaptation. Thus, throughout the original C. dentata range, an extensive network has developed consisting of partnering institutions, C. dentata mother trees, and breeding and seed orchards to help ensure production of locally adapted C. dentata germplasm for reintroduction (Fig. 5). Ideally, sources of resistance should include parent trees of C. mollissima, Castanea seguinii



**Fig. 4** (a) BC<sub>3</sub>F<sub>1</sub> *Castanea dentata* 2.5 yr after inoculation with chestnut blight showing canker response indicating susceptibility; tree is dead above point of inoculation. (b) The first moderately blight-resistant BC<sub>3</sub>F<sub>1</sub> *C. dentata* produced by the Indiana chapter of the American Chestnut Foundation also shown 2.5 yr after inoculation (IN-96-01A, a cross from IW2 × GR226). Photographs courtesy of James McKenna.

(Dode), and *Castanea crenata* (Siebold & Zucc.), for example, because it is likely that trees within and among species will carry different resistance genes. To achieve these goals, TACF is using multiple genotypes of primarily *C. mollissima* as resistance sources and breeding lines should contain at least 20 *C. dentata* parents, providing necessary genetic diversity (Hebard, 2006). Putatively blight-resistant BC<sub>3</sub>F<sub>3</sub> seed was first harvested by TACF in 2005 (Diskin *et al.*, 2006); germplasm availability from the breeding program should increase exponentially with time (Jacobs, 2007).

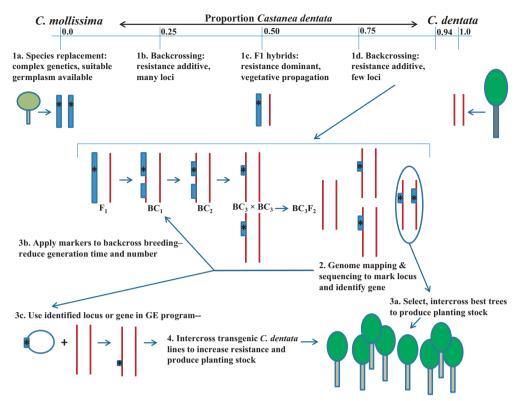
#### 3. Genetic engineering

The first application of genetically engineered (GE) forest trees will probably be for reintroduction of species decimated by invasive pathogens or pests (Adams et al., 2002; Merkle et al., 2007). An important consideration for GE trees is the source and tissue specificity of resistance genes as well as their regulatory elements. Both transgenic and cisgenic approaches draw from the same recombinant DNA toolbox, but they differ in the source of the pest resistance genes. Transgenic approaches use genes from nonplant organisms or from plants that are not closely related (i.e. sexually incompatible) to the target plant. Cisgenic approaches use genes from closely related or even the same species and cisgenics has been noted for its similarity to inter-species backcross breeding (Fig. 6; Schouten & Jacobsen, 2008; Han et al., 2010). GE technologies have progressed rapidly (Fig. 3) and may offer new opportunities for restoring species on the verge of extirpation. The exotic hemlock woolly adelgid, for example, is devastating T. canadensis and no crossable species with resistance exists; however, congenerics with resistance do exist (i.e. Tsuga chinensis (Franch.) Prixtel ex Diels.; Montgomery et al., 2009), opening the possibility for cisgenics. Both transgenic and cisgenic approaches are viable options for



**Fig. 5** Distribution of field sites (*Castanea dentata* mother trees, breeding plantations, orchards, and research sites) and major partnering organizations (university, government, and corporate) contributing toward *C. dentata* reintroduction. Data were provided by Sara Fitzsimmons (The American Chestnut Foundation).

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**Fig. 6** Approaches for attaining resistance to chestnut blight, ranging from species replacement to various forms of hybridization and selection (1a to 1d). These methods represent a continuum in proportions of donor genome in new or recipient species, ranging from 100% under species replacement (1a) to *c*. 6% in backcross breeding to the  $BC_3F_2$  generation (1d). The backcross breeding program provides optimal materials for genetically mapping blight resistance gene loci (2). Selected  $BC_3F_2$  trees can be used in seed orchards as relatively true breeding parents to produce seeds for forest planting (3a). Once genetically mapped, markers can be used to facilitate backcross breeding in other crosses for increasing genetic diversity of the foundation species (3b). In addition, fine mapping and quantitative trait locus (QTL) sequencing can identify candidate genes for testing in genetic engineering (GE) lines of the foundation species (3c). Resistant GE lines can be further propagated and intercrossed to increase diversity and provide seeds for forest planting (4).

*C. dentata* and make it a valuable test case for the use of these GE technologies for forest restoration.

Substantial progress has been made using transgenic approaches to developing blight resistance in C. dentata. Several small proteins with antimicrobial activity, including antifungal activity, show little or no toxicity to *Castanea*, *Malus*, or *Salix* spp. pollen (Powell et al., 1995, 2000, 2006). The oxalate oxidase (OxO) gene provides another promising lead for chestnut blight resistance (Polin et al., 2006; Welch *et al.*, 2007). When inserted into poplar (*Populus*  $\times$ euramericana (Dode) Guinier), OxO increases tissue tolerance to oxalate and enhances resistance to the fungal pathogen Septoria musiva (Liang et al., 2001). Co-transformation of three genes is routine in C. dentata – a visual selectable marker (e.g. GFP), antibiotic resistance (e.g. *npt*II, neomycin phosphotransferase II) for selection in culture, and the candidate resistance gene (Newhouse et al., 2010) – further facilitating work in this species. In this case, because the marker and selection genes are not linked to the resistance gene, they can be removed from the segregating breeding population while retaining progeny that contain resistance genes. Co-transformation has limitations, however, including high variation in gene expression and gene silencing (Halpin et al., 2001), although the co-expression of multiple genes in a single open reading frame may provide a solution to this problem (Liang et al., 2005).

genes in closely related species. The most definitive research on blight resistance genetics in Castanea supports a three gene model as detected by quantitative trait locus (QTL) mapping in a C. mollissima × C. dentata cross (Kubisiak et al., 1997). The three-locus model accounted for c. 70% of the genetic variation, implicating a combination of major and minor genes contributing to resistance. New higher density maps using short sequence repeat (SSR or microsatellite) and single nucleotide polymorphism (SNP) markers developed from large-scale expressed gene sequencing (Barakat et al., 2009) have confirmed and refined genomic locations of blight resistance QTLs (Kubisiak et al., in press). Such maps enable increased precision in locating blight resistance loci and greater sensitivity in finding smaller effect loci. At present, candidate genes for cisgenic GE applications are identified by their presence in genomic regions identified as QTLs for resistance, their presence or absence in suppressive subtraction hybridization (SSH) libraries (Baier, 2009), and their differential expression in inoculated vs noninoculated C. mollissima stems or in noninoculated C. mollissima vs C. dentata stems (Barakat et al., 2009, 2012). For example, one promising candidate resistance gene, a laccase gene, is highly expressed in C. mollissima stem tissues, expressed at very low levels in C. dentata, and appears to map to a blight resistance QTL (Baier, 2009).

A cisgenic approach relies on the identification of resistance

#### 4. Genome-informed genetics

Molecular markers have improved our understanding of C. dentata genetics by delineating patterns of genetic diversity and dissecting quantitative trait variation (Pigliucci et al., 1990; Kubisiak et al., 1997; Huang et al., 1998; Casasoli et al., 2006). Isozyme studies show that C. dentata contains low to moderate levels of genetic diversity relative to other species with large geographic ranges and similar life history traits (Dane et al., 2003), while DNA markers show C. dentata having similar levels of diversity to these species (Kubisiak & Roberds, 2006). It is unclear whether the low genetic diversity as observed using isozyme markers predisposed C. dentata to rapid population decline in response to the blight epidemic or whether it is a consequence of blight-induced population decline (Dane et al., 2003). Most of the neutral genetic variation observed in C. dentata resides within populations (>~90% for isozymes;  $>\sim$  95% for DNA markers), with evidence of clinal trends in overall allele diversity and allele frequencies for some loci (Huang et al., 1994; Kubisiak & Roberds, 2006). Relatively early in the DNA marker era, Bernatzky & Mulcahy (1992) and Ellingboe (1992) suggested using markers to map resistance genes in C. mollissima and thereby facilitate their introgression into C. dentata through backcross breeding. Conceptually, this is an excellent idea that has been shown to be effective in numerous systems (Collard & Mackill, 2008; Moose & Mumm, 2008), yet early markers (e.g. restriction fragment length polymorphism (RFLP), random amplified polymorphic DNA (RAPD), amplification fragment length polymorphism (AFLP), and isozymes) proved to be suboptimal and cost prohibitive for operational use in chestnut breeding programs.

The recent development of large sets of SSR and SNP markers (Kubisiak et al., in press;) is likely to provide the practical application envisioned (Fig. 6; Nance et al., 1992; Wheeler & Sederoff, 2009). SSRs and SNPs are codominant and have much higher sequence specificity (providing data on the same loci across parents), effectively overcoming the two major problems encountered with RAPD and AFLP. The candidate genes are being isolated from C. mollissima and C. seguinii and used to transform C. dentata to directly test their effectiveness in providing blight resistance. Additionally, the highly informative maps enable tracking of introgressed C. mollissima genes within inter-species backcross families as well as estimating the remaining C. mollissima genome at various generations, facilitating the dual selection for resistance (from *C. mollissima*) and recurrent type (from *C. dentata*). The use of markers in selection for recurrent type can greatly improve its recovery (Tanksley & Rick, 1980; Soller & Beckmann, 1983). Many backcross programs plan for six backcross generations (Allard, 1960); however, with informative markers two generations provide similar results (Visscher et al., 1996). Markers may reduce the three backcross generations of the TACF program to only one, allowing additional resistance sources to be introgressed with a similar level of effort. Such integration and collaboration among technological developments will expand the capacity of the technological sphere with the potential to increase restoration success (Fig. 2).

Tansley review

## III. Ecology of the historically dominant *Castanea dentata* in contemporary forests

To ensure that efforts associated with development of blightresistant C. dentata are fully realized, silvicultural management based on a thorough ecological understanding of the species is essential (Griffin, 2000). Our incomplete understanding of the basic biology and ecology of C. dentata (Paillet, 2002; Jacobs, 2007) originates primarily from historical observations, growth of stump sprouts, and studies of a few extant populations that were initiated by plantings outside the native range (Paillet, 1982, 1984; Paillet & Rutter, 1989). Collectively, this information has provided important insight into the historical role of C. dentata. However, over the past c. 100 yr, during which C. dentata has been functionally extirpated from the canopy, several major landscapelevel changes have occurred across the species range. Eastern deciduous forests were heavily impacted by extensive land clearing and large-scale fires during the late 19th to early 20th Centuries, which has since been replaced by management favoring fire suppression and clearcutting over progressively smaller units, selective cutting, or single tree selection (Abrams, 2003). Concurrent with a decrease in disturbance, dramatic increases in whitetailed deer (Odocoileus virginianus Zimmermann) populations have intensified herbivory (Abrams, 2003; McEwan et al., 2011). Analysis of climatic data for the period from the early 16th Century onwards suggests that the 20th Century exhibited increased soil moisture availability and decreased severity of drought (McEwan et al., 2011). The result is a distinctly different forest composition in contemporary forests, characterized by decreased regeneration of oaks (Quercus spp.) and an increase in the importance of maples (Acer spp.) (Abrams, 1998, 2003; Fei et al., 2011). Considering these broad-scale changes in land-use patterns and forest dynamics within the context of a recent burst of research into C. dentata ecology can improve our ability to forecast the niche of blightresistant C. dentata in modern-day forests. Within our conceptual framework, ecological knowledge informs both where and how to reintroduce blight-resistant C. dentata, and the capacity to forecast ecological barriers to and implications of reintroduction (Fig. 2a).

#### 1. Identifying optimal reintroduction habitats

Current insight into the physiological ecology of *C. dentata* combined with evidence of its historical range and dominance suggests that the species is an opportunistic generalist, well adapted to conform to a broad range of environmental and climatic conditions. However, *C. dentata* is absent from or grows poorly on high-pH, limestone-derived soils (Russell, 1987; Tindall *et al.*, 2004). *Castanea dentata* exhibits characteristics of both a late-successional (relative shade tolerance and extended longevity) and pioneer species (aggressive stump sprouting and juvenile competitiveness), thereby distinguishing *C. dentata* from oaks and other co-occurring species.

*Castanea dentata* exhibits high plasticity to varying light conditions and has been classified as both an intermediate shade-tolerant (Joesting *et al.*, 2007, 2009) and a shade-tolerant (Wang *et al.*, 2006) species. Shading produces either a neutral (Wang *et al.*, 2006;

Rhoades et al., 2009) or a positive (McCament & McCarthy, 2005; Anagnostakis, 2007) effect on germination and/or juvenile survival. Once established, seedlings and saplings may persist for years under low light conditions beneath the canopy (Paillet & Rutter, 1989; McEwan et al., 2006), exhibiting plasticity by increasing leaf mass per unit area with greater light availability (King, 2003; Wang et al., 2006; Joesting et al., 2009). Castanea dentata seedlings, saplings, and mature trees exhibited light compensation points, quantum efficiency, leaf mass per unit area, and nitrogen concentrations similar to those of shade-tolerant species (Joesting et al., 2009). Understory trees, however, have high maximum rates of photosynthesis representative of fast-growing, shade-intolerant species such as yellow-poplar (Liriodendron tulipifera L.) and eastern cottonwood (Populus deltoides Bartram ex Marsh.) (Joesting et al., 2009). Furthermore, C. dentata growth and photosynthesis are promoted by increased light availability (McCament & McCarthy, 2005; Wang et al., 2006; Joesting et al., 2007), exceeding or equaling those of many shade-intolerant pioneer species (Boring et al., 1981; Griffin, 1989; King, 2003). Although C. dentata may survive for prolonged periods as stump sprouts or seedlings beneath the canopy, it retains the ability to rapidly respond to release following disturbance (Paillet & Rutter, 1989; Paillet, 2002; McEwan et al., 2006). Sprout growth may exceed that of any other hardwood species following clearcutting (Mattoon, 1909) and radial growth rates approach  $5 \text{ mm yr}^{-1}$  in plantation or natural stand settings, with maximum reported values of  $10-12 \text{ mm yr}^{-1}$  (Zeigler, 1920; Paillet & Rutter, 1989; Jacobs & Severeid, 2004).

The former dominance of C. dentata in upland habitats suggests greater drought tolerance compared with co-occurring species (Jacobs, 2007). Castanea dentata exhibited higher instantaneous water use efficiency under controlled drought conditions relative to several species of upland oaks and dry-site red maples (Acer rubrum L.) (Bauerle et al., 2006) and C. dentata sprouts had higher leaf water potential than several species of upland oaks during an early summer drought (Abrams et al., 1990). In spite of its capacity to persist on dry sites, C. dentata was reported to have comprised 25-40% of the basal area in some riparian zones in the southern Appalachians (Vandermast & Van Lear, 2002). Although C. dentata has been shown to increase leaf, shoot, and root biomass with greater availability of specific nutrients, including nitrogen, potassium, and magnesium (Latham, 1992; Rieske et al., 2003; McCament & McCarthy, 2005), responses to specific nutrient limitations or nutrient interactions are less well studied. In glasshouse experiments, however, C. dentata outperformed all other species tested in traits associated with competitive ability over a wide range of nutrient and light combinations (Latham, 1992). In addition, leachate from C. dentata litter may have allelopathic properties (Vandermast et al., 2002). In combination, the evidence regarding eco-physiological parameters and historical occurrence suggests that C. dentata should perform well over a broad range of sites targeted for reintroduction.

#### 2. Management to promote establishment and spread

Based on the apparent generalist character of *C. dentata*, Jacobs (2007) outlined several promising paths toward reintroduction,

namely through afforestation of degraded sites, such as former minelands, and reforestation by underplanting in extant forests. Despite the competitiveness of juvenile C. dentata, aggressive silvicultural management will probably be necessary to ensure vigorous establishment of high-value blight-resistant seedlings (Griffin, 2000; Rhoades et al., 2009). Specific recommendations for underplanting (Wang et al., 2006), thinning and burning (McCament & McCarthy, 2005), and herbicide application (Selig et al., 2005; Robertson & Davis, 2011) have been proposed to promote establishment in natural stands and field plantations. The co-occurring species, yellow-poplar, is a particularly serious potential competitor that may necessitate use of prescribed fire to promote establishment of C. dentata, particularly for reforestation in large openings on mesic sites (Griscom & Griscom, 2012), along with consistent silvicultural management through the stem exclusion stage of stand development (Morrissey et al., 2008). Reforestation of C. dentata in the Appalachian Mountains may also be hindered by competitive exclusion from dense thickets of understory ericaceous shrubs, such as Rhododendron maximum L. and Kalmia latifolia L. (Vandermast & Van Lear, 2002), which may require mechanical removal to facilitate seedling establishment and to ensure safe implementation of prescribed fire. Mycorrhizal colonization has been shown to increase C. dentata seedling survival and total biomass in afforestation plantings on mine reclamation sites (Bauman et al., 2012). Management actions, such as plowing or disking, increased the percentage colonization of ectomycorrhizas in planted C. dentata seedlings, and seedling survival was enhanced with nursery inoculation of mycorrhizas before planting (Bauman, 2010). A broad-scale restoration program will probably combine reforestation and afforestation approaches, and variable management techniques will be required to promote initial establishment of chestnut populations in different settings.

Ultimately, however, all reintroduction approaches will rely on effective seed dispersal and natural regeneration to promote population growth and spread of blight-resistant C. dentata. The little we know about natural regeneration dynamics of C. dentata comes largely from an isolated stand of C. dentata in southwestern Wisconsin located over 600 km from the native range that developed in the absence of blight. In 70 yr, nine original planted C. dentata trees supplied sufficient regeneration to spread the species over 1 km, outcompeting oaks and hickories (Carya spp.) (McEwan et al., 2006). Within c. 0.5 km from the source trees, C. dentata comprised at least 25% of total canopy basal area and predominated among advanced saplings entering the canopy (Paillet & Rutter, 1989). Regeneration and dispersal of C. dentata in this stand was a multi-step process, including (1) establishment of pioneer trees following seed dissemination in light gaps, (2) development of advance regeneration in the understory, and (3) persistence of advance regeneration underneath the established canopy until released by disturbance to assume canopy dominance (Paillet & Rutter, 1989; Jacobs, 2007). Reintroduction of blightresistant chestnut will probably rely upon management techniques that promote success at each of these three steps. Selecting appropriate sites for reintroduction and applying effective silvicultural management will minimize environmental stresses, thereby enhancing the expression of blight resistance (Jones et al., 1980; Griffin, 2000). As discussed above, open pollination and natural seeding, facilitated by silvicultural treatments, should allow for reproduction in stands of reintroduced *C. dentata* and may also provide complementary materials for the current breeding programs.

#### 3. Potential ecological barriers

Unfortunately, chestnut blight is not the only pathogen or pest that threatens C. dentata reintroduction. Emerging as perhaps the most serious obstacle in the southern portion of its range, the introduced soil-borne Oomycete pathogen Phytophthora cinnamomi Rands causes ink disease lesions that form on roots (i.e. root rot) and inhibit water and nutrient uptake (Maurel et al., 2001a,b), leading to reduced tree vigor and eventual mortality (Anagnostakis, 2001; Rhoades et al., 2003, 2009). Strategies to mitigate effects of P. cinnamomi include careful site selection (e.g. well-drained sites), identification of ectomycorrhizas that confer protection to roots, and additional resistance breeding using Asian chestnuts (Anagnostakis, 2001; Rhoades et al., 2003). The recently imported oriental gall wasp (Dryocosmus kuriphilus Yasumatsu) forms galls on actively growing chestnut shoots that may lead to severe damage or morality (Anagnostakis, 2001), although the severity of gall wasps may be regulated by their own pests such as parasitoid wasps or pathogens (Cooper & Rieske, 2007). Additional pests that may hamper C. dentata reintroduction include gypsy moth (Lymantria dispar L.), and ambrosia beetles (Xylosandrus crassiusulus Mot. and Xylosandrus saxeseni Blandford); the magnitude of these threats is still unclear (Oliver & Mannion, 2001; Rieske et al., 2003). Compared with blight resistance, however, less is known about the genetics of resistance to other pathogens and pests. Chestnuts (C. crenata, C. dentata, and C. mollissima) are apparently more susceptible to gall wasps relative to the chinkapins (Castanea pumila and Castanea pumila var. ozarkensis) and Castanea henryi (Anagnostakis et al., 2009). Thus, resistance to chestnut blight, ink disease, and gall wasps is available through other Castanea spp. and much of the variation seems to be controlled by only a few genes. Whether these resistance responses are controlled by the same or similar genes in the different host species remains to be seen, but emerging genomic technologies (Wheeler & Sederoff, 2009) should facilitate the manipulation of the effective genes in breeding and GE programs as restoration goals dictate.

Herbivores and seed predators may limit both the establishment and spread of reintroduced populations of *C. dentata.* While-tailed deer densities have increased greatly since chestnut was a dominant species in eastern forests, and both exclosure and modeling studies have shown that deer herbivory can have a profound effect on the structure and composition of forest tree communities (Rooney & Waller, 2003; Côté *et al.*, 2004; Didion *et al.*, 2009). Deer browse has been implicated in limiting establishment of many tree species including northern red oak (*Quercus rubra* L.), yellow birch (*Betula alleghaniensis* Britt.), northern white cedar (*Thuja occidentalis* L.), and eastern hemlock (*Tusga canadensis* (L.) Carrière) (Rooney & Waller, 2003). For *C. dentata* and the oaks that largely replaced them, scatterhoarding small mammals and birds are the major agents of seed dispersal (Steele *et al.*, 2005). Scatterhoarding animals, however, may also function as seed predators, particularly in low-mast years, when all cached seeds are recovered and consumed (Steele *et al.*, 2005). While acorns, hickories, and walnuts all contain a higher percentage of lipids, *C. dentata* nuts have a higher percentage of carbohydrates and much lower levels of tannins, which may lead to preferential consumption of *C. dentata* (Steele *et al.*, 2005). Other potentially significant seed predators for reintroduced *C. dentata* include weevils (Dalgleish *et al.*, 2012), turkey, deer, and black bear.

Because the native range of C. dentataspans five US climatic zones (Fig. 1), reintroduction must also be locally adapted in order to promote successful establishment (Worthen et al., 2010). Kubisiak & Roberds (2006) found low but positive correlations between genetic and geographic distances, suggesting that C. dentata was a single metapopulation established by high gene flow and genetic drift. Neutral DNA markers support a southwest to northeast clinal trend in decreasing genetic diversity with no indication of regional boundaries (Kubisiak & Roberds, 2006). Although these trends are important to consider in breeding and reintroduction planning, new research to better understand adaptive trait variation is needed to clearly delineate zones of optimal adaptation. Recent evidence of uncertainty regarding cold tolerance of hybrid-backcross C. dentata used for reintroduction (Gurney et al., 2011) re-emphasizes the need for local adaptability. To achieve local adaptability, final crosses within backcross breeding and/or GE approaches must utilize C. dentata germplasm specifically adapted to the planting environments (Fig. 5). In addition to being locally adapted, it is necessary to ensure that reintroduced C. dentata has and maintains sufficient genetic diversity, which will be challenged by the continually declining vigor of extant sprouts (Huang et al., 1998). Furthermore, given that a relatively narrow range of genotypes has served as resistance donors for backcross breeding (Hebard, 2006), additional sources of resistance should be continuously integrated to protect against adaptation of the pathogen to the resistance genes (Ellingboe, 1992).

#### 4. Ecological implications of reintroduction

Because *C. dentata* was a foundation species in eastern forests, successful reintroduction has the potential to influence population, community, and ecosystem dynamics in these forests. For consumer populations, *C. dentata* may increase and stabilize the available seed resource (Dalgleish & Swihart, 2012). Previous research has shown that the size and frequency of the seed resource have far-reaching community effects: increasing small mammal populations can increase predation pressure on songbirds, decrease gypsy moth outbreaks, and increase Lyme disease risk to human populations (Jones *et al.*, 1998; Ostfeld *et al.*, 2006). While successful *C. dentata* reintroduction may offset seed losses due to limited oak regeneration, it also may exacerbate the oak recruitment problems as acorns compete with *C. dentata* for dispersers and establishment sites.

Successful reintroduction of *C. dentata* may also alter ecosystem functions such as productivity, decomposition, and nutrient cycling as a consequence of its fast growth, high tannin content of the wood, and low C : N ratio of the leaves (Jacobs & Severeid,

2004; Ellison *et al.*, 2005). Jacobs *et al.* (2009) found that *C. dentata* had greater aboveground biomass and bole C than other interplanted species and compared favorably in carbon sequestration ability with fast-growing species commonly used for forestry plantations. Furthermore, *C. dentata* wood is decay resistant: Wallace *et al.* (2001) found that > 24% of the large woody debris sampled in an Appalachian stream was *C. dentata*, although all the large *C. dentata*, however, decay more quickly and have a higher nutritional quality for stream macroinvertebrates than do oak leaves (Smock & MacGregor, 1988). The loss of *C. dentata* has probably altered nutrient cycling in Appalachian streams by changing the foliar inputs that form the energy base in these systems (Ellison *et al.*, 2005).

Deployment of blight-resistant C. dentata is intended by TACF and partners to be targeted toward the original C. dentata range and most activity is concentrated within this zone (Fig. 5). Nevertheless, in the wake of a rapidly changing climate the original native range may no longer reflect the zone to which C. dentata is optimally adapted (Harris et al., 2008; Potter & Hargrove, 2012). Additionally, much of the large-scale hardwood afforestation plantings in the USA for C sequestration, conservation, wildlife, and timber occur in the Midwest and along the Mississippi River Alluvial Valley, which encompass a limited portion of the original C. dentata range. Castanea dentata has demonstrated its ability to thrive when introduced far outside of its native range (Jacobs & Severeid, 2004; McEwan et al., 2006; Jacobs et al., 2009), raising ecological considerations regarding its potential to suppress indigenous vegetation (Jacobs, 2007). Additional research into the biotic interactions between chestnut and its dispersers, seed predators, plant competitors, and herbivores within the context of the current forest ecosystems is required to adequately forecast the ecological consequences of chestnut restoration, whether inside or outside the native range.

## IV. Societal challenges for successful reintroduction of *Castanea dentata*

Recently, the importance of understanding the connections and feedback between social and biological systems has come to the forefront in conservation (Pretty & Smith, 2004) and restoration (Aronson et al., 2010; Collier, 2011). While barriers to integrating sociological and ecological knowledge can be significant, effective restoration will require both (Stanturf, 2005; Hobbs, 2007; Miller & Hobbs, 2007). Socio-economic research, however, can serve to simultaneously gather and disseminate information among different stakeholders, and thereby increase cohesion and participation in restoration activities (Collier, 2011). In the case of C. dentata reintroduction, the social sphere within our conceptual framework remains, perhaps, the least well understood (Fig. 2a). We detail three key areas for further socio-ecological study, including understanding the social acceptability of different technologies for developing blight resistance, current policy issues that may influence reintroduction, and the crucial task of setting restoration goals.

An integral question is whether hybrid and GE *C. dentata* trees will prove socially acceptable for reintroduction (Jacobs, 2007;

Merkle et al., 2007). The backcross breeding program, which produces a backcross hybrid, relies upon traditional plant breeding techniques that are widely used in production agriculture, but may prove less publically acceptable for use in forest restoration (Jacobs, 2007). The efficacy of backcross breeding for forest trees also remains uncertain, which could necessitate a shift toward integration of GE technologies in C. dentata restoration. Whether the public will accept a GE C. dentata to ensure restoration of the species and its ecosystem is unclear (Merkle et al., 2007). The case of C. dentata (and other threatened forest tree species targeted for restoration) is further complicated by the notion that any GE tree deployed in North America must be sterile or have some other means to control flowering (Merkle et al., 2007), which conflicts with the underlying premise of repopulation through sexual reproduction. While pursuit of both backcross breeding and GE simultaneously may be the soundest strategy to obtain effective blight resistance, ultimately, public perception and acceptance may dictate which types of trees may be used for restoration, regardless of efficacy (Strauss et al., 2009).

Although public concerns regarding application of GE technologies have been increasing, many leading scientific groups espouse that the plant trait and not the GE method of obtaining the trait, as well as its associated ecological impacts, should determine whether and in what contexts GE trees should be used (Strauss et al., 2009). In addition, because of the diversity of GE methods, desired outcomes, potential applications, costs, and benefits, each use of GE trees should be evaluated on a case-by-case basis (Strauss et al., 2009). For example, recent advances in GE technology may allow the development of cisgenic C. dentata (Han et al., 2010), whereby the expressed genes along with their genetic control/regulatory elements are from Castanea species. The relative similarity of cisgenics to backcross breeding (Schouten & Jacobsen, 2008) suggests that this approach may garner the broadest social support, and ultimately prove to be the most biologically viable method to introduce a trait such as blight resistance in a GE context. Restoring a culturally important species, such as C. dentata, for the public good may be viewed as an acceptable use of GE. Conversely, allowing GE for restoration of C. dentata could be seen as a 'slippery slope' for the broader commercial use of GE trees (Strauss et al., 2009). Further education on GE methods, benefits and risks, as well as a better understanding of public opinion and concerns is needed (Bublea et al., 2009).

Governmental and regulatory policies relevant to restoration are shaped by both political and financial interests (Collier, 2011). GE technologies are regulated by multiple governmental agencies when the modified trait is pest resistance and the organism produces food for wildlife and human consumption (i.e. USDA Animal and Plant Health Inspection Service (APHIS)-Biotechnology Regulatory Service, Environmental Protection Agency (EPA), and Food and Drug Administration (FDA)). Societal influences may also drive silvicultural management policy options for reintroduction as public opinion regarding harvesting, fire, and other forms of disturbance exert direct influence on management on public lands (Jacobs, 2007). While targeting private land for reintroduction overcomes many of the political and policy restrictions specific to silvicultural manipulations on public lands, it creates other social obstacles. For example, the increasingly parceled nature of private forest land ownership may limit silvicultural options that promote establishment of *C. dentata* (McEwan *et al.*, 2006; Jacobs, 2007). In addition, Knoot *et al.* (2010) document a lack of support among private landowners for management that favors oak regeneration (e.g. disturbance), raising the question of whether similar management strategies will be supported for *C. dentata.* The reluctance of private forest landowners to use prescribed fire because of air quality issues and the risk of fire escape at the expanding wildland– urban interface (Stanturf & Goodrick, in press) has been identified as a serious obstacle for restoration of the fire-adapted longleaf pine (*Pinus palustris* Mill.) in the southeastern USA (Brockway *et al.*, 2005). Ultimately, the success of different management options to promote *C. dentata* on varying land types will also be balanced against their cost to implement (Stanturf *et al.*, 2009).

Restoration resides at the center of our conceptual framework as the overarching and central common goal among the three spheres. The Society for Ecological Restoration defines ecological restoration as the 'process of assisting the recovery of an ecosystem that has been degraded, damaged, or destroyed' (SER, 2004). Identifying the source of the ecosystem degradation and setting realistic, attainable goals are critical to the success of any restoration endeavor (Hobbs, 2007). In the case of C. dentata reintroduction, it is the loss of a foundation species that is the cause of degradation. Setting realistic and attainable goals for C. dentata reintroduction is arguably among the most critical and difficult tasks. Static reliance upon the historical distribution and abundance of C. dentata as the sole criterion for success may be neither feasible nor desirable (Hildebrand et al., 2005). We posit that some of the major determinants of C. dentata reintroduction goals will be (1) the ability to confer blight resistance, effectively and sustainably, in a socially acceptable manner, (2) the availability of sufficient blight-resistant plant material at an acceptable cost, (3) prioritization of restoration targets (locations and site types, e.g. urban landscapes, public forests, and private land) with capable partners and at appropriate scales, and (4) the capacity to accurately assess the viability of reintroduced populations (e.g. growth and spread) within the varied levels of management intensity. The large geographic scale, immense number of diverse interest groups involved (Fig. 5), and potential to influence many aspects of eastern forest ecology will require setting national, regional, and local goals that engage stakeholders as active participants (Hobbs & Cramer, 2008; Collier, 2011); ambiguity will set the stage for failure.

#### **V.** Conclusions

In the case of *C. dentata*, an introduced pathogen reduced a major component of the eastern forest canopy to an understory shrub within a single generation, illustrating how the rapid loss of a foundation species can dramatically change an ecosystem across a landscape. Because of its strong ecological and cultural significance, developing blight resistance has been aggressively pursued by a diverse contingency of supporters over several decades. The ultimate goal of producing blight-resistant *C. dentata* has advanced the fields of tree breeding and biotechnology, offering the possibility to bring new tools to bear on socio-ecological problems. We are now arguably on the cusp of perhaps the largest reintroduction effort to date for a plant species.

*Castanea dentata* restoration provides an effective model for restoration. Based upon *C. dentata*, we have proposed a conceptual framework, whereby maximizing restoration success requires increasing the capacity and integration among social, ecological, and technological spheres (Fig. 2a). While the technological and ecological spheres have been the primary targets of emphasis and are the areas for which the majority of breakthroughs have been realized for *C. dentata*, the critical need for progress in the social sphere has only recently been fully recognized; development in this area is in its infancy and remains perhaps the weakest link toward achieving restoration success.

Although *C. dentata* was among the first eastern deciduous tree species threatened with extirpation as a result of introduced pathogens or pests, many other tree species face a similar fate. Our conceptual framework can be broadly applied because the three spheres (technological, ecological, and social) are common to all restoration challenges, yet with the common denominator that restoration success is limited by the size and degree of overlap among spheres (Fig. 2b). Through effective merging of genetic technologies, reintroduction biology, and restoration ecology, *C. dentata* may serve as an important model to inform reintroduction of threatened species in general and foundation forest tree species in particular.

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#### References

Abrams MD. 1998. The red maple paradox. BioScience 48: 355-364.

Abrams MD. 2003. Where has all the white oak gone? BioScience 53: 927-938.

- Abrams MD, Schultz JC, Kleiner KW. 1990. Ecophysiological responses in mesic versus xeric hardwood species to an early-season drought in central Pennsylvania. *Forest Science* 36: 970–981.
- Adams JM, Piovesan G, Strauss S, Brown S. 2002. The case for genetic engineering of native and landscape trees against introduced pests and diseases. *Conservation Biology* 16: 874–879.
- Allard RW. 1960. Principles of plant breeding. New York, NY, USA: Wiley.
- Anagnostakis SL. 1982. Genetic analysis of *Endothia parasitica*: linkage data for four single genes and three vegetative compatibility types. *Genetics* 102: 25–28.
- Anagnostakis SL. 1987. Chestnut blight: the classical problem of an introduced pathogen. *Mycologia* 79: 23–37.
- Anagnostakis SL. 2001. The effect of multiple importations of pests and pathogens on a native tree. *Biological Invasions* 3: 245–254.

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Anagnostakis SL. 2007. Effect of shade on growth of seedling American chestnut. Northern Journal of Applied Forestry 24: 317–318.

- Anagnostakis SL, Clark S, McNab HW. 2009. Preliminary report on the segregation of resistance in chestnuts to infestation by oriental chestnut gall wasp. *Acta Horticulturae* 815: 33–35.
- Anderson PJ, Rankin WH. 1914. Endothia *canker of chestnut*. Bulletin 347 of the Department of Pathology, Cornell University, Agricultural Experiment Station of the College of Agriculture. Ithaca, NY USA: Cornell University.
- Andrade GM, Nairn CJ, Le HT, Merkle SA. 2009. Sexually mature transgenic American chestnut trees via embryogenic suspension-based transformation. *Plant Cell Reports* 28: 1385–1397.

Armstrong DP, Seddon PJ. 2008. Directions in reintroduction biology. Trends in Ecology & Evolution 23: 20–25.

- Aronson J, Blignaut JN, Milton SJ, Le Maitre D, Esler KJ, Limouzin A, Fontaine C, de Wit MP, Mugido W, Prinsloo P et al. 2010. Are socioeconomic benefits of restoration adequately quantified? A meta-analysis of recent papers (2000–2008) in Restoration Ecology and 12 other scientific journals. *Restoration Ecology* 10: 143–154.
- Baier KM. 2009. Interspecific suppression subtractive hybridization identifies a differentially expressed Chinese chestnut (*Castanea mollissima*) laccase-like gene. M.S. Thesis. Syracuse, NY, USA: State University of New York, College of Environmental Science and Forestry, 218p.

Barakat A, Diloreto DS, Zhang Y, Smith C, Baier K, Powell WA, Wheeler N, Sederoff R, Carlson JE. 2009. Comparison of the transcriptomes of American chesnut (*Castanea dentata*) and Chinese chestnut (*C. mollissima*) in response to chestnut blight infection. *BMC Plant Biology* 9: 51.

- Barakat A, Staton M, Cheng C-H, Park J, Yassin NBM, Ficklin S, Yeh C-C, Hebard F, Baier K, Powell W et al. 2012. Chestnut resistance to the blight disease: insights from transcriptome analysis. BMC Plant Biology 12: 38.
- Bauerle WL, Wang GG, Bowden JD, Hong CM. 2006. An analysis of ecophysiological responses to drought in American chestnut. *Annals of Forest Science* 63: 833–842.
- Bauman JM. 2010. Ectomycorrhizal communities associated with restoration plantings of American chesntut (Castanea dentata) seedlings on Ohio mine lands: planting methodologies to promote root colonization, PhD dissertation. Miami University, Oxford, OH, USA.
- Bauman JM, Keiffer CH, Hiremath S. 2012. Facilitation of American chestnut (*Castanea dentata*) seedling establishment by *Pinus virginiana* in mine restoration. *International Journal of Ecology* 2012: 257326.
- Beattie RK, Diller JD. 1954. Fifty years of chestnut blight in America. Journal of Forestry 52: 323–329.
- Bernatzky R, Mulcahy DL. 1992. Marker-aided selection in a backcross breeding program for resistance to chestnut blight in the American chestnut. *Canadian Journal of Forest Research* 22: 1031–1035.
- Berry FH. 1978. Chestnut breeding in the U.S. In: MacDonald WL, Cech FC, Luchok J, Smith C, eds. *Proceedings of the American Chestnut Symposium*. Morgantown, WV, USA: West Virginia University Books, Department of Agriculture, 39–40.

Boring LR, Monk CD, Swank WT. 1981. Early regeneration of clear-cut southern Appalachian forest. *Ecology* 62: 1244–1253.

Braun EL. 1950. Deciduous forests of eastern North America. New York, NY, USA: Hafner.

Brewer LG. 1995. Ecology of survival and recovery from blight in American chestnut trees (*Castanea dentata* (Marsh.) Borkh.) in Michigan. *Bulletin of the Torrey Botanical Club* 122: 40–57.

- Bublea T, Nisbet MC, Borchelt R, Brungers F, Critchley C, Einsiedel E, Geller G, Gupta A, Hampel J, Hyde-Lay R et al. 2009. Science communication reconsidered. *Nature Biotechnology* 27: 514–518.
- Burnham CR. 1981. Blight-resistant American chestnut: there's hope. *Plant Disease* 65: 459–460.
- Burnham CR, Rutter PA, French DW. 1986. Breeding blight-resistant chestnuts. *Plant Breeding Reviews* 4: 347–397.

- Carraway DT, Wilde HD, Merkle SA. 1994. Somatic embryogensis and gene transfer in American chestnut. *Journal of The American Chestnut Foundation* 8: 29–33.
- Casasoli M, Derory J, Morera-Dutrey C, Brendel O, Porth I, Guehl JM, Villani F, Kremer A. 2006. Comparison of quantitative trait loci for adaptive traits between oak and chestnut based on an expressed sequence tag consensus map. *Genetics* 172: 533–546.
- Clapper RB. 1952. Relative blight resistance of some chestnut species and hybrids. *Journal of Foresty* 50: 453–455.
- Clapper RB. 1954. Chestnut breeding, techniques and results I. Breeding material and pollination techniques. *Journal of Heredity* 45: 106–114.
- Clark SL, Schlarbaum SE, Saxton AM, Hebard FV. 2011. Making history: field testing of blight-resistant American chestnut (*Castanea dentata*) in the southern region. In: Fei S, Lhotka JM, Stringer JW, Gottschalk KW, Gary W, eds. *Proceedings, 17th central hardwood forest conference.* Lexington, KY, USA: USDA Forest Service Northern Research Station, 656–657.
- **Collard BCY, Mackill DJ. 2008.** Marker-assisted selection: an approach for precision plant breeding in the twenty-first century. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* **363**: 557–572.

Collier MJ. 2011. Incorporating socio-economic factors into restoration: implications from industrially harvested peatlands. *Restoration Ecology* 19: 559–563.

Cooper WR, Rieske LK. 2007. Community associates of an exotic gallmaker, Dryocosmus kuriphilus, (Hymenoptera: Cynipidae) in eastern North America. Annals of the Entomological Society of America 100: 236–244.

Cortesi P, McCulloch CE, Song H, Lin H, Milgroom MG. 2001. Genetic control of horizontal virus transmission in the chestnut blight fungus, *Cryphonectria parasitica. Genetics* 159: 107–118.

Cortesi P, Milgroom MG. 1998. Genetics of vegetative incompatibility in Cryphonectria parasitica. Applied and Environmental Microbiology64: 2988–2994.

Côté SD, Rooney TP, Tremblay J-P, Dussault C, Waller DM. 2004. Ecological impacts of deer overabundance. *Annual Review of Ecology and Systematics* 35: 113–147.

Dalgleish HJ, Shukle JT, Swihart RK. 2012. Weevil seed damage reduces germination and seedling growth of hybrid American chestnut. *Canadian Journal of Forest Research* 42: 1107–1114.

Dalgleish HJ, Swihart RK. 2012. American chestnut past and future: implications of restoration for resource pulses and consumer populations of eastern U. S. Forests. *Restoration Ecology* 20: 490–497.

Dane F, Lang P, Huang H, Fu Y. 2003. Intercontinental genetic divergence of *Castanea* species in eastern Asia and eastern North America. *Heredity* 91:314–321.

- Didion M, Kupferschmid AD, Bugmann H. 2009. Long-term effects of ungulate browsing on forest composition and structure. *Forest Ecology and Management* 258: S44–S55.
- Diller JD, Clapper RB. 1965. A progress report on attempts to bring back the chestnut tree in the eastern United States, 1954–1964. *Journal of Forestry* 63: 186–188.

Diskin M, Steiner KC, Hebard FV. 2006. Recovery of American chestnut characteristics following hybridization and backcross breeding to restore blightravaged *Castanea dentata*. Forest Ecology and Management 223: 439–447.

Ellingboe A. 1992. Breeding blight resistant American chestnut. In: Double ML, MacDonald WL, eds. *International Chestnut Conference*. Morgantown, WV, USA: West Virginia University, 47–51.

- Ellison AM, Bank MS, Clinton BD, Colburn EA, Elliot K, Ford CR, Foster BD, Kloeppel BD, Knoepp JD, Lovett GM *et al.* 2005. Loss of foundation species: consequences for the structure and dynamics of forested ecosystems. *Frontiers in Ecology and the Environment* 9: 479–486.
- Fei S, Kong N, Steiner KC, Moser WK, Steiner EB. 2011. Change in oak abundance in the eastern United States from 1980 to 2008. Forest Ecology and Management 262: 1370–1377.
- Foster DR, Clayden S, Orwig DA, Hall B, Barry S. 2002. Oak, chestnut and fire: climatic and cultural controls of long-term forest dynamics in New England, USA. *Journal of Biogeography* 29: 1359–1379.

Griffin GJ. 1989. Incidence of chestnut blight and survival of American chestnut in forest clear-cut and neighboring understory sites. *Plant Disease* 73: 123–127.

Brockway DG, Outcalt KW, Tomczak D, Johnson EE. 2005. Restoring longleaf pine forest ecosystems in the Southern U.S. In: Stanturf JA, Madsen P, eds. *Restoration of boreal and temperate forests*. Boca Raton, FL, USA: CRC Press, 501–519.

Graves AH. 1942. Breeding work toward the development of a timber type of blightresistant chestnut. Report for 1941. *American Journal of Botany* 29: 622–626.

- Griffin GJ. 2000. Blight control and restoration of the American chestnut. *Journal of Forestry* 98: 22–27.
- Griscom HP, Griscom BW. 2012. Evaluating the ecological niche of American chestnut for optimal hybrid reintroduction sites in the Appalachian ridge and valley province. *New Forests* 43: 441–455.
- Gurney KM, Schabert PG, Hawley GJ, Shane JB. 2011. Inadequate cold tolerance as a possible limitation to American chestnut restoration in the northeastern United States. *Restoration Ecology* 19: 55–63.
- Halpin C, Barakate A, Askari BM, Abbott JC, Ryan MD. 2001. Enabling technologies for manipulating multiple genes on complex pathways. *Plant Molecular Biology* 47: 295–310.
- Han K, Dharmawardhana P, Arias R, Ma C, Busov V, Strauss S. 2010. Gibberellinassociated cisgenes modify growth, stature and wood properties in *Populus. Plant Biotechnology Journal* 9: 162–178.
- Harris JA, Hobbs RJ, Higgs E, Aronson J. 2008. Ecological restoration and global climate change. *Restoration Ecology* 14: 170–176.
- Hebard FV. 1994. The American Chestnut Foundation breeding plan: beginning and intermediate steps. *Journal of The American Chestnut Foundation* 8: 21–28.
- Hebard FV. 2006. The backcross breeding program of the American Chestnut Foundation. *Journal of The American Chestnut Foundation* 19: 55–77.
- Hepting GH. 1974. Death of the American chestnut. *Journal of Forest History* 18: 61–67.
- Hildebrand RH, Watts AC, Randle AM. 2005. The myths of restoration ecology. *Ecology and Society* 10: Article 19.
- Hobbs RJ. 2007. Setting effective and realistic restoration goals: key directions for research. *Restoration Ecology* 15: 354–357.
- Hobbs RJ, Cramer VA. 2008. Restoration ecology: interventionist approaches for restoring and maintaining ecosystem function in the face of rapid environmental change. *Annual Review of Ecology and Systematics* 33: 39–61.
- Hobbs RJ, Norton DA. 1996. Towards a conceptual framework for restoration ecology. *Restoration Ecology* 4: 93–110.
- Huang H, Dane F, Norton JD. 1994. Allozyme diversity in Chinese, seguin, and American chestnut (*Castanea* spp). *Theoretical and Applied Genetics* 88: 981–985.
- Huang HW, Dane F, Kubisiak TL. 1998. Allozyme and RAPD analysis of the genetic diversity and geographic variation in wild populations of the American chestnut (Fagaceae). *American Journal of Botany* 85: 1013–1021.
- Jacobs DF. 2007. Toward development of silvical strategies for forest restoration of American chestnut (*Castanea dentata*) using blight-resistant hybrids. *Biological Conservation* 137: 497–506.
- Jacobs DF, Seilig MF, Severeid LR. 2009. Aboveground carbon biomass of plantation-grown American chestnut (*Castanea dentata*) in absence of blight. *Forest Ecology and Management* 258: 288–294.
- Jacobs DF, Severeid LR. 2004. Dominance of interplanted American chestnut (*Castanea dentata*) in southwestern Wisconsin, USA. *Forest Ecology and Management* 101: 111–120.
- Jaynes RA, Elliston JE. 1980. Pathogenticity and canker control by mixtures of hypovirulent strains of *Endothia parasitica* in American chestnut. *Phytopathology* 70: 453–456.
- Joesting HM, McCarthy BC, Brown KJ. 2007. The photosynthetic response of American chestnut seedlings to different light conditions. *Canadian Journal of Forest Research* 37: 1714–1722.
- Joesting HM, McCarthy BC, Brown KJ. 2009. Determining the shade tolerance of American chestnut using morphological and physiological leaf parameters. *Forest Ecology and Management* 257: 280–286.
- Jones C, Griffin GJ, Elkins JR. 1980. Association of climatic stress with blight on Chinese chestnut in the eastern United States. *Plant Disease* 64: 1001–1004.
- Jones CG, Ostfelt RS, Richard MP, Schauber EM, Wolff JO. 1998. Chain reactions linking acorns to gypsy moth outbreaks and Lyme disease risk. *Science* 279: 1023–1026.
- King DA. 2003. Allocation of above-ground growth is related to light in temperate deciduous saplings. *Functional Ecology* 17: 482–488.
- Knoot TG, Schulte LA, Rickenbach M. 2010. Oak conservation and restoration on private forestlands: negotiating a social-ecological landscape. *Ecological Management* 45: 155–164.
- Kubisiak TL, Hebard FV, Nelson CD, Zhang JS, Bernatzky R, Huang H, Anagnostakis SL, Doudrick RL. 1997. Molecular mapping of resistance to blight in an interspecific cross in the genus *Castanea*. *Phytopathology* 87: 751–759.

- Kubisiak TL, Milgroom MG. 2006. Markers linked to vegetative incompatibility (vic) genes and a region of high heterogeneity and reduced recombination near the mating type locus (mat) in Cryphonectria parasitica. Fungal Genetics and Biology 43: 453–463.
- Kubisiak TL, Nelson CD, Staton ME, Zhebentyayeva T, Smith C, Olukolu BA, Fang GC, Hebard FV, Anagnostakis S, Wheeler N *et al.* In press. A transcriptome-based genetic map of Chinese chestnut (*Castanea mollissima*), and identification of regions of segmental homology with peach (*Prunus persica*). *Tree Genetics & Genomes.* doi: 10.1007/s11295-012-0579-3
- Kubisiak TL, Roberds JH. 2006. Genetic structure of American chestnut populations based on neutral DNA markers. In: Steiner KC, Carlson JE, eds. *Restoration of American chestnut to forest lands: proceedings of a conference and workshop*. Asheville, NC, USA: National Park Service, 109–122.
- Latham RE. 1992. Co-occurring tree species change rank in seedling performance with resources varied experimentally. *Ecology* 17: 482–488.
- Liang H, Gao H, Maynard CA, Powell WA. 2005. Expression of a self-processing, pathogen resistance-enhancing gene construct in *Arabidopsis. Biotechnology Letters* 27: 435–442.
- Liang H, Maynard CA, Allen RD, Powell WA. 2001. Increased Septoria musiva resistance in transgenic hybrid poplar leaves expressing a wheat oxalate oxidase gene. *Plant Molecular Biology* 45: 619–629.
- Lipsey MK, Child MF. 2007. Combining the fields of reintroduction biology and restoration ecology. *Conservation Biology* 21: 1387–1388.
- Liu YC, Milgroom MG. 1996. Correlation between hypovirus transmission and the number of vegetative incompatibility (vic) genes different among isolates from a natural population of Cryphonectria parasitica. Phytopathology 86: 79–86.
- MacDonald WL, Fulbright DW. 1991. Biological control of chestnut blight: use and limitations of transmissible hypovirulence. *Plant Disease* 75: 656–661.
- Mattoon WR. 1909. The origin and early development of chestnut sprouts. *Forest Quarterly* 7: 34–37.
- Maurel M, Robin C, Capdevielle X, Loustau D, Desperez-Loustau M-L. 2001a. Effects of variable root damage caused by *Phytophthora cinnamomi* on water relations of chestnut saplings. *Annals of Forest Science* **58**: 639–651.
- Maurel M, Robin C, Capron G, Desperez-Loustau M-L. 2001b. Effects of root damage associated with *Phytophthora cinnamomi* on water relations, biomass accumulation, mineral nutrition, and vulnerability to water deficit of five oak and chestnut species. *Forest Pathology* 31: 353–369.
- McCament CL, McCarthy BC. 2005. Two-year response of American chestnut (*Castanea dentata*) to shelterwood harvesting and fire in a mixed-oak forest ecosystem. *Canadian Journal of Forest Research* 35: 740–749.
- McEwan RW, Dyer JM, Pederson N. 2011. Multiple interacting ecosystem drivers: toward an encompassing hypothesis of oak forest dynamics across eastern North America. *Ecography* 34: 244–265.
- McEwan RW, Keiffer CH, McCarthy BC. 2006. Dendroecology of American chestnut in a disjunct stand of oak-chestnut forest. *Canadian Journal of Forest Research* 36: 1–11.
- Merkle SA, Andrade GM, Hair CJ, Powell WA, Maynard CA. 2007. Restoration of threatened species: a noble cause for transgenic trees. *Tree Genetics & Genomes* 3: 111–118.
- Merkle SA, Wiecko AT, Watson-Pauley AB. 1991. Somatic embryogenesis in American chestnut. *Canadian Journal of Forest Research* 21: 1698–1701.
- Milgroom MG, Cortesi P. 2004. Biological control of chestnut blight with hypovirulence: a critical analysis. *Annual Review of Phytopathology* 42: 311–338.
- Miller JR, Hobbs RJ. 2007. Habitat restoration—do we know what we're doing? *Restoration Ecology* 15: 382–390.
- Montgomery ME, Bentz SE, Olsen RT. 2009. Evaluation of hemlock (*Tsuga*) species and hybrids to *Adelges tsugae* (Hemiptera: Adelgidae) using artifical infestation. *Journal of Enonomic Entomology* **102**: 1247–1254.
- Moose SP, Mumm RH. 2008. Molecular plant breeding as the foundation for 21st century crop improvement. *Plant Physiology* 147: 969–977.
- Morrissey RC, Jacobs DF, Seifert JR, Fischer BC, Kershaw JA. 2008. Competitive success of natural oak regeneration in clearcuts during the stem exclusion stage. *Canadian Journal of Forest Research* **38**: 1419–1430.
- Murrill WA. 1906. A new chestnut disease. Torreya 6: 186-189.
- Nance WL, Tuskan GA, Nelson CD, Doudrick RL. 1992. Potential application of molecular markers for genetic analysis of host-pathogen systems in forest trees. *Canadian Journal of Forest Research* 22: 1036–1043.

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Newhouse AE, Zhang AB, Northern L, Maynard CA, Powell WA. 2010. Analysis of transgenic American chestnut. *Phytopathology* 100: S1–S89.

Oliver JB, Mannion CM. 2001. Ambrosia beetles (Coleoptera: Scolytidae) species attacking chestnut and captured in ethanol-baited traps in middle Tennessee. *Environmental Entomology* **30**: 909–918.

Ostfeld RS, Canham CD, Oggenfuss K, Winchcombe RJ, Keesing F. 2006. Climate, deer, rodents, and acorns as determinants of variation in Lyme-disease risk. *PLoS Biology* 4: 1058–1068.

Paillet FL. 1982. The ecological significance of American chestnut (*Castanea dentata* (Marsh.) Borkh.) in the Holocene forests of Connecticut. *Bulletin of the Torrey Botanical Club* 109: 457–473.

Paillet FL. 1984. Growth-form and ecology of American chestnut sprout clones in northeastern Massachusetts. *Bulletin of the Torrey Botanical Club* 111: 316–328.

Paillet FL. 2002. Chestnut: history and ecology of a transformed species. *Journal of Biogeography* 29: 1517–1530.

Paillet FL, Rutter PA. 1989. Replacement of native oak and hickory tree species by the introduced American chestnut (*Castanea dentata*) in southwestern Wisconsin. *Canadian Journal of Botany* 67: 3457–3469.

Parker GG, Hill SM, Kuehnel LA. 1993. Decline of understory American chestnut (*Castanea dentata*) in a southern Appalachian forest. *Canadian Journal of Forest Research* 23: 259–265.

Pigliucci M, Benedettelli S, Villani F. 1990. Spatial patterns of genetic variability in Italian chestnut (*Castanea sativa*). *Canadian Journal of Botany* 68: 1962–1967.

Polak T, Saltz D. 2011. Reintroduction as an ecosystem restoration technique. Conservation Biology 25: 424–427.

Polin LD, Liang H, Rothrock RE, Nishii M, Diehl DL, Newhouse AE, Nairn CJ, Powell WA, Maynard CA. 2006. Agrobacterium-mediated transformation of American chestnut (*Castanea dentata* (Marsh.) Borkh.) somatic embryos. *Plant Cell, Tissue, and Organ Culture* 84: 69–78.

Potter KM, Hargrove WW. 2012. Determining suitable locations for seed transfer under climate change: a global quantitative method. *New Forests* 43: 581–599.

Powell WA, Catranis CM, Maynard CA. 1995. Synthetic antimicrobial peptide design. *Molecular Plant-Microbe Interactions* 8: 792–794.

**Powell WA, Catranis CM, Maynard CA. 2000.** Design of self-processing antimicrobial peptides for plant protection. *Letters in Applied Microbiology* **31**: 163–168.

Powell WA, Maynard CA, Boyle B, Seguin A. 2006. Fungal and bacterial resistance in transgenic trees. In: Fladung M, Ewald D, eds. *Tree transgenics, recent developments.* Berlin, Germany: Springer, 235–252.

Pretty J, Smith D. 2004. Social capital in biodiversity conservation and management. *Conservation Biology* 18: 631–638.

Rhoades C, Loftis D, Lewis J, Clark S, Serv UF. 2009. The influence of silvicultural treatments and site conditions on American chestnut (*Castanea dentata*) seedling establishment in eastern Kentucky, USA. *Forest Ecology and Management* 258: 1211–1218.

Rhoades CC, Brosi SL, Dattilo AJ, Vincelli P. 2003. Effects of soil compaction and moisture on incidence of *Phytophthora* root rot on American chestnut (*Castanea dentata*) seedlings. *Forest Ecology and Management* 184: 47–54.

Rieske LK, Rhoades CC, Miller SP. 2003. Foliar chemistry and gypsy moth, *Lymantria dispar* (L.), herbivory on pure American chestnut, *Castanea dentata* (Fam: Fagaceae), and a disease-resistant hybrid. *Environmental Entomology* **32**: 359–365.

Robertson ND, Davis AS. 2011. Influence of sulfometuron methyl on American chestnut seedling growth and leaf function. *Northern Journal of Applied Forestry* 28: 36–40.

Robichaud RL, Lessard VC, Merkle SA. 2004. Treatments affecting maturation and germination of American chestnut somatic embryos. *Journal of Plant Physiology* 161: 957–969.

Rooney TP, Waller DM. 2003. Direct and indirect effects of white-tailed deer in forest ecosystems. *Forest Ecology and Management* 181: 165–176.

Russell EWB. 1987. Pre-blight distribution of *Castanea dentata* (Marsh.) Borkh. Bulletin of the Torrey Botanical Club 114: 183–190.

Schouten H, Jacobsen E. 2008. Cisgenesis and intragenesis, sisters in innovative plant breeding. *Trends in Plant Science* 13: 260–261.

Seddon PJ. 2010. From reintroduction to assisted colonization: moving along the conservation translocation spectrum. *Restoration Ecology* 18: 796–802. Seddon PJ, Armstrong DP, Maloney RF. 2007a. Developing the science of reintroduction biology. *Conservation Biology* 21: 303–312.

Seddon PJ, Armstrong DP, Maloney RF. 2007b. Combining the fields of reintroduction biology and restoration ecology. *Conservation Biology* 21: 1388– 1390.

Selig MF, Seifert JR, Jacobs DF. 2005. Response of American chestnut to weed control treatments at plantation establishment. *Journal of The American Chestnut Foundation* 19: 33–41.

SER (Society for Ecological Restoration International Science and Policy Working Group). 2004. The SER international primer on ecological restoration. [WWW document] URL http://www.ser.org/ [accessed on 5 March 2012].

Smith DM. 2000. American chestnut: ill-fated monarch of the eastern hardwood forest. *Journal of Forestry* 98: 12–15.

Smock LA, MacGregor CM. 1988. Impact of the American chestnut blight on aquatic shredding macroinvertebrates. *Journal of The North American Benthological Society* 7: 212–221.

Soller M, Beckmann JS. 1983. Genetic polymorphism in varietal identification and genetic improvement. *Theoretical and Applied Genetics* 67: 25–33.

Stanturf JA. 2005. What is forest restoration? In: Stanturf JA, Madsen P, eds. Restoration of boreal and temperate forests. Boca Raton, FL, USA: CRC Press, 3–11.

Stanturf JA, Gardiner ES, Shepard JP, Schweitzer CJ, Portwood CJ, Dorris LC Jr. 2009. Restoration of bottomland hardwood forests across a treatment intensity gradient. *Forest Ecology and Management* 257: 1803–1814.

Stanturf JA, Goodrick SL. In press. Fire. In: Wear DN, Greis JG, eds. Southern Forest Futures Project. Asheville, NC: USDA Forest Service, Southern Research Station. [WWW document] URL http://www.srs.fs.usda.gov/futures/reports/ draft/Frame.htm [accessed on 17 August 2012].

Steele MA, McCarthy BC, Keiffer CH. 2005. Seed dispersal, seed predation, and the American chestnut. *Journal of The American Chestnut Foundation* 19: 47–54.

Steiner KC, Carlson JE, eds. 2006. Restoration of American chestnut to forest lands: Proceedings of a conference and workshop. Natural Resources Report NPS/NCR/ CUE/NRR - 2006/001, National Park Service. Washington, DC. [WWW document] URL http://sfr.psu.edu/public/chestnut/information/conference-2004/conference/conference-proceedings [accessed on 27 April 2012].

Stephenson SL. 1986. Changes in a former chestnut-dominated forest after a half century of succession. *American Midland Naturalist* 116: 173–179.

Stephenson SL, Adams HS, Lipford ML. 1991. The present distribution of chestnut in the upland forest communities of Virginia. *Bulletin of the Torrey Botanical Club* 118: 24–32.

Strauss SH, Tan H, Boerjan W, Sedjo R. 2009. Strangled at birth? Forest biotech and the Convention on Biological Diversity. *Nature Biotechnology* 27: 519–527.

Tanksley SD, Rick CM. 1980. Isozyme gene linkage map of the tomato: applications in genetics and breeding. *Theoretical and Applied Genetics* 57: 161– 170.

Tindall JR, Gerrath JA, Meizer M, McKendry K, Husband BC, Boland GJ. 2004. Ecological status of American chestnut (*Castanea dentata*) in its native range in Canada. *Canadian Journal of Forest Research* 34: 2554–2563.

Vandermast DB, Van Lear DH. 2002. Riparian vegetation in the southern Appalachian mountains (USA) following chestnut blight. *Forest Ecology and Management* 155: 97–106.

Vandermast DB, Van Lear DH, Clinton BD. 2002. American chestnut as an allelopath in the southern Appalachians. *Forest Ecology and Management* 165: 173–181.

Visscher PM, Haley CS, Thompson R. 1996. Marker-assisted introgression in backcross breeding programs. *Genetics* 144: 1923–1932.

Wallace JB, Webster JR, Eggert SL, Meyer JL, Siler ER. 2001. Large woody debris in a headwater stream: long-term legacies of forest disturbance. *International Review of Hydrobiology* 86: 501–513.

Wang GG, Bauerle WL, Mudder BT. 2006. Effects of light acclimation on the photosynthesis, growth, and biomass allocation in American chestnut (*Castanea dentata*) seedlings. *Forest Ecology and Management* 226: 173–180.

Welch AJ, Stipanovic AJ, Maynard CA, Powell WA. 2007. The effects of oxalic acid on transgenic *Castanea dentata* callus tissue expressing oxalate oxidase. *Plant Science* 172: 488–496.

- Wheeler N, Sederoff R. 2009. Role of genomics in the potential restoration of the American chestnut. *Tree Genetics & Genomes* 5: 181–187.
- Worthen LM, Woeste KE, Michler CH. 2010. Breeding American chestnuts for blight resistance. *Plant Breeding Reviews* **33**: 305–339.
- Young TP. 2000. Restoration ecology and conservation biology. *Biological Conservation* 92: 73–83.
- Youngs RL. 2000. "A right smart little jolt" Loss of the chestnut and a way of life. Journal of Forestry 98: 17–21.
- Zeigler EA. 1920. Problems arising form the loss of our chestnut. *Forest Leaves* 17: 152–155.

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