

Title:

N4-Methylation Changes the Conformation of (3*S*,6*S*)-3-Alkyl-6-benzylpiperazine-2,5-diones from Folded to Extended

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Abstract:

N4-Methylation of (3*S*,6*S*)-3-alkyl-6-benzylpiperazine-2,5-diones (*S,S*)-**1a–c** was found to change their folded conformation to an extended conformation. Conformational aspects of N1- and/or N4-methylated (*S,S*)-**1a–c** were revealed by single crystal X-ray crystallography and ¹H NMR spectroscopy.

Keywords:

Piperazine-2,5-dione

N-methylation

Folded conformation

Extended conformation

CH/ π interaction

X-ray crystal structure

1. Introduction

Piperazine-2,5-diones (2,5-DKPs) are an important class of heterocyclic compounds because of their wide range of biological activities [1–5]. In addition, 2,5-DKPs could be regarded as an attractive scaffold for functional molecules including pharmaceuticals due to the structural diversity achieved by introducing substituents on the 2,5-DKP ring [6–12]. Therefore, the regulation of their conformation by chemical modification has recently attracted much attention. In general, C3- or C6-mono-benzylated 2,5-DKPs are known to adopt a folded conformation, in which the benzyl moiety is folded over the 2,5-DKP ring [13–21]. Previously, we reported that 2,5-DKPs derived from L- or D-phenylalanine and α -substituted L-serine adopted a folded conformation based on their ^1H NMR spectra [22]. In addition, our recent study has demonstrated the importance of intramolecular CH/ π interaction in the folded conformation of mono-benzylated 2,5-DKPs due to the electronic effects of *para*-substituents on the benzyl group in the ^1H NMR spectra [23]. Intriguingly, similar folded conformations were also observed in chiral 5-benzylimidazolidin-4-ones, which are known as effective organocatalysts [24–30]. As the next step in our ongoing research on the conformation of 2,5-DKPs, we herein investigated the possibility of using N1- and/or N4-methylation to change the conformation of (3*S*,6*S*)-3-alkyl-6-benzylpiperazine-2,5-diones (*S,S*)-**1a–c** from a folded to an extended one (Figure 1). In this context, we note that *N*-methylation of amide bonds is considered to be one of the important chemical modifications for control of the conformation and the biological function of peptides or proteins [31].

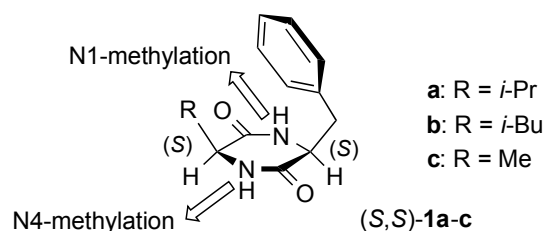


Fig. 1. N1- and N4-methylation of (3*S*,6*S*)-3-alkyl-6-benzylpiperazine-2,5-diones (*S,S*)-**1a–c**.

2. Experimental section

All reactions were monitored by TLC employing 0.25 mm silica gel plates (Merck 5715; 60 F₂₅₄). Column chromatography was carried out on silica gel [Kanto Chemical 60N (spherical, neutral); 63-210 mm]. Anhydrous CH₂Cl₂ and DMF were used as purchased from Kanto Chemical. Triethylamine was distilled prior to use. All other reagents were used as purchased. All melting points were determined on a Yanagimoto micro melting point apparatus and are uncorrected. IR spectra were obtained using a JASCO FT/IR-6200 IR Fourier transform spectrometer. ^1H NMR (500 MHz) and ^{13}C NMR (125 MHz) spectra were recorded on a Bruker AV500 spectrometer. Chemical shifts are given in δ values (parts per million)

using tetramethylsilane (TMS) as an internal standard. Electron spray ionization mass spectra (ESIMS) were recorded on a Waters LCT Premier spectrometer. Elemental combustion analyses were performed using a Yanagimoto CHN CORDER MT-5 and a J-SCIENCE LAB JM10. The microwave-assisted was performed utilizing an automated single-mode microwave synthesizer (InitiatorTM 60; Biotage AB). Single crystal X-ray diffraction experiments were performed on a Rigaku RAXIS-RAPID diffractometer using graphite monochromated Mo-K α ($\lambda = 0.71075 \text{ \AA}$, 50 kV, 40 mA) radiation. Data were corrected for Lorentz and polarization effects. The structure was solved by direct methods [32] and expanded using Fourier techniques. The non-hydrogen atoms were refined anisotropically. Hydrogen atoms were refined using the riding model. All calculations were performed using the CrystalStructure [33] crystallographic software package.

2.1. General procedure for the preparation of dipeptides (*S,S*)-**4a–c** and (*S,S*)-**6a–c**

2.1.1. Methyl (*S*)-2-[(*S*)-2-(*tert*-Butoxycarbonylamino)-*N*,3-dimethylbutanamido]-3-phenylpropanoate [(*S,S*)-**4a**]

To a solution of (*S*)-**2a** (172 mg, 0.793 mmol) and (*S*)-**3d** (200 mg, 0.872 mmol) in anhydrous CH₂Cl₂ (3 mL) was added HOBt (118 mg, 0.872 mmol), EDC • HCl (228 mg, 1.19 mmol) and triethylamine (122 μ L, 0.872 mmol) at 0 °C under argon. The reaction mixture was allowed to warm to room temperature and stirred for 24 h. The reaction mixture was treated with 5% citric acid (5 mL) and then extracted with CHCl₃ (10 mL \times 3). The extract was dried over anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The oily residue was purified by silica gel column chromatography [Silica Gel 60N: *n*-hexane–AcOEt (3:1)] to afford (*S,S*)-**4a** (192.1 mg, 62%). White solid; mp 87.0–88.0 °C; [α]_D²⁹ –69.9 (*c* 1.01, CHCl₃); ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 0.36 (d, *J* = 6.7 Hz, 0.4H), 0.66 (d, *J* = 6.7 Hz, 0.4H), 0.89 (d, *J* = 6.8 Hz, 2.6H), 0.95 (d, *J* = 6.8 Hz, 2.6H), 1.41 (s, 9H), 1.87–1.98 (m, 1H), 2.92/2.94 (s \times 2, 3H), 3.00 (dd, *J* = 10.1, 14.6 Hz, 1H), 3.39 (dd, *J* = 5.7, 14.6 Hz, 1H), 3.72/3.73 (s \times 2, 3H), 4.00–4.05 (m, 0.2H), 4.34 (dd, *J* = 6.1, 9.4 Hz, 0.8H), 4.94 (d, *J* = 9.8 Hz, 0.2H), 4.99 (dd, *J* = 4.4, 10.3 Hz, 0.2H), 5.05 (d, *J* = 9.3 Hz, 0.8H), 5.34 (dd, *J* = 5.7, 10.1 Hz, 0.8H), 7.15–7.33 (m, 5H); ¹³C NMR (125 MHz, CDCl₃, mixture of rotamers) δ 17.20, 17.27, 18.9, 19.5, 28.29, 28.34, 29.7, 30.7, 31.2, 32.8, 34.5, 35.2, 52.3, 52.6, 54.9, 55.0, 58.4, 61.4, 79.3, 79.4, 126.8, 127.1, 128.5, 128.8, 128.9, 129.2, 136.75, 136.80, 155.4, 155.7, 170.5, 171.1, 172.9, 173.1; IR (KBr) 2971, 1734, 1710, 1642, 1516, 1364 cm⁻¹; ESIMS *m/z*: calcd for C₂₁H₃₂N₂NaO₅ [M+Na]⁺, 415.2209; found, 415.2206.

2.1.2. Methyl (*S*)-2-[(*S*)-2-(*tert*-Butoxycarbonylamino)-*N*,4-dimethylpentanamido]-3-phenylpropanoate [(*S,S*)-**4b**]

White solid, mp 127–129 °C; $[\alpha]_D^{29}$ -79.4 (c 1.20, CHCl_3); ^1H NMR (500 MHz, CDCl_3 , mixture of rotamers) δ 0.69/0.71 (d \times 2, J = 6.6 Hz, 1H), 0.92/0.96 (d \times 2, J = 6.7, 6.5 Hz, 5H), 1.30–1.45 (m, 2H), 1.40/1.41 (s \times 2, 9H), 1.65–1.75 (m, 1H), 2.90/2.93 (s \times 2, 3H), 3.05 (dd, J = 10.5, 14.5 Hz, 1H), 3.38 (dd, J = 5.3, 14.5 Hz, 1H), 3.73/3.75 (s \times 2, 3H), 4.31 (dt, J = 3.8, 10.1 Hz, 0.2H), 4.53 (dt, J = 4.0, 9.6 Hz, 0.8H), 4.88 (d, J = 9.8 Hz, 0.2H), 4.96 (dd, J = 4.1, 10.8 Hz, 0.2H), 5.02 (d, J = 9.2 Hz, 0.8H), 5.20 (dd, J = 5.5, 10.4 Hz, 0.8H), 7.16–7.53 (m, 5H); ^{13}C NMR (125 MHz, CDCl_3 , mixture of rotamers) δ 21.5, 21.8, 23.2, 23.3, 24.2, 24.6, 28.28, 28.33, 29.9, 32.9, 34.5, 35.2, 41.1, 42.4, 47.8, 48.8, 52.2, 52.6, 58.9, 61.3, 79.3, 79.4, 126.8, 127.1, 128.5, 128.8, 129.0, 129.1, 136.76, 136.84, 155.2, 155.5, 170.5, 171.0, 173.5, 173.7; IR (KBr) 3385, 2978, 1731, 1705, 1645, 1517, 1363 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{22}\text{H}_{34}\text{N}_2\text{NaO}_5$ $[\text{M}+\text{Na}]^+$, 429.2365; found, 429.2343.

2.1.3. Methyl (S)-2-[(S)-2-(*tert*-Butoxycarbonylamino)-*N*-methylpropanamido]-3-phenylpropanoate [(S,S)-4c]

Colorless needles (CH_2Cl_2 -*n*-hexane), mp 88–89 °C; $[\alpha]_D^{19}$ -64.9 (c 1.00, CHCl_3); ^1H NMR (500 MHz, CDCl_3 , mixture of rotamers) δ 0.72 (d, J = 6.6 Hz, 0.5H), 1.25 (d, J = 6.8 Hz, 2.5H), 1.40/1.42 (s \times 2, 9H), 2.86 (s, 2.5H), 2.92 (s, 0.5H), 2.95–3.02 (m, 0.2H), 3.08 (dd, J = 10.6, 14.5 Hz, 0.8H), 3.38 (dd, J = 5.2, 14.5 Hz, 1H), 3.73/3.74 (s \times 2, 3H), 4.26–4.33 (m, 0.2H), 4.50 (brquint, 0.8H), 4.94 (brdd, 0.2H), 5.05–5.15 (m, 1H), 5.26 (brd, 0.8H), 7.15–7.31 (m, 5H); ^{13}C NMR (125 MHz, CDCl_3 , mixture of rotamers) δ 18.5, 18.6, 28.3, 28.4, 29.9, 32.4, 33.4, 34.4, 34.7, 35.2, 45.2, 46.5, 52.3, 52.6, 58.1, 59.5, 61.4, 79.4, 79.5, 126.8, 126.9, 127.2, 128.5, 128.77, 128.83, 128.9, 129.1, 136.6, 136.9, 154.8, 155.0, 170.4, 170.9, 173.2, 173.6; IR (KBr) 3387, 2980, 2946, 1737, 1710, 1644, 1513, 1360 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{19}\text{H}_{28}\text{N}_2\text{NaO}_5$ $[\text{M}+\text{Na}]^+$, 387.1896; found, 387.1916.

2.1.4. Methyl (S)-2-[(S)-2-(*tert*-Butoxycarbonylamino)-*N*-methyl-3-phenylpropanamido]-3-methylbutanoate [(S,S)-6a]

Colorless oil; $[\alpha]_D^{28}$ -68.9 (c 0.93, CHCl_3); ^1H NMR (500 MHz, CDCl_3 , mixture of rotamers) δ 0.55 (d, J = 6.7 Hz, 0.3H), 0.80 (d, J = 6.7 Hz, 2.7H), 0.91 (d, J = 6.5 Hz, 0.3H), 0.95 (d, J = 6.5 Hz, 2.7H), 1.38/1.40 (s \times 2, 9H), 2.07–2.18 (m, 1H), 2.72 (s, 2.7H), 2.86 (s, 0.3H), 2.93 (dd, J = 5.7, 13.1 Hz, 1H), 3.01 (dd, J = 8.6, 13.1 Hz, 1H), 3.66/3.69 (s \times 2, 3H), 4.02 (d, J = 10.4 Hz, 0.1H), 4.79–4.86 (m, 0.9H), 4.91 (d, J = 10.7 Hz, 0.9H), 4.95–5.01 (m, 0.1H), 5.13 (brd, 0.1H), 5.28 (brd, 0.9H), 7.16–7.29 (m, 5H); ^{13}C NMR (125 MHz, CDCl_3 , mixture of rotamers) δ 18.5, 19.6, 27.2, 28.3, 30.7, 39.4, 51.70, 51.75, 61.3, 79.8, 126.79, 126.84, 128.40, 128.46, 129.5, 129.6, 136.2, 155.3, 170.9, 173.0; IR (neat) 3315, 3029, 2972, 2876, 1740, 1711, 1659, 1642, 1513, 1494, 1453, 1411, 1391, 1367, 1294, 1249, 1170 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{21}\text{H}_{32}\text{N}_2\text{NaO}_5$ $[\text{M}+\text{Na}]^+$, 415.2209; found, 415.2210.

2.1.5.

Methyl

(*S,S*)-2-[(*S*)-2-(*tert*-Butoxycarbonylamino)-*N*-methyl-3-phenylpropanamido]-4-methylpentanoate [(*S,S*)-**6b**]
Colorless oil; $[\alpha]_{\text{D}}^{29} -21.8$ (*c* 1.34, CHCl₃); ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 0.80/0.83 (d \times 2, *J* = 6.4, 7.0 Hz, 0.5H), 0.89/0.91 (d \times 2, *J* = 6.5, 6.7 Hz, 5.5H), 1.37/1.39 (s \times 2, 9H), 1.42–1.49 (m, 1H), 1.60–1.74 (m, 2H), 2.79/2.81 (s \times 2, 3H), 2.90 (dd, *J* = 6.2, 13.5 Hz, 1H), 3.08 (dd, *J* = 7.5, 13.5 Hz, 0.9H), 3.15 (dd, *J* = 7.7, 12.9 Hz, 0.1H), 3.67 (s, 3H), 4.55–4.60 (m, 0.1H), 4.84 (brq, 0.9H), 5.11 (brd, 0.1H), 5.26 (brd, 0.9H), 5.30 (dd, *J* = 5.2, 10.5 Hz, 1H), 7.17–7.30 (m, 5H); ¹³C NMR (125 MHz, CDCl₃, mixture of rotamers) δ 21.4, 22.2, 22.7, 23.2, 24.66, 24.72, 28.1, 28.3, 31.0, 37.2, 38.2, 39.0, 39.7, 51.3, 51.7, 52.1, 52.3, 54.5, 57.8, 79.7, 126.7, 126.8, 128.37, 128.41, 129.5, 129.6, 136.3, 136.7, 155.2, 171.2, 171.9, 172.7; IR (neat) 3319, 3029, 2956, 2871, 1741, 1711, 1643, 1495, 1454, 1411, 1391, 1366, 1249, 1172 cm⁻¹; ESIMS *m/z*: calcd for C₂₂H₃₄N₂NaO₅ [M+Na]⁺, 429.2365; found, 429.2375.

2.1.6. Methyl (*S,S*)-2-[(*S*)-2-(*tert*-Butoxycarbonylamino)-*N*-methyl-3-phenylpropanamido]propanoate [(*S,S*)-**6c**]

Colorless oil; $[\alpha]_{\text{D}}^{29} -19.1$ (*c* 0.60, CHCl₃); ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 0.91 (d, *J* = 7.0 Hz, 0.5H), 1.36 (d, *J* = 7.3 Hz, 2.5H), 1.40/1.41 (s \times 2, 9H), 2.73 (s, 0.5H), 2.81 (s, 2.5H), 2.91 (dd, *J* = 6.3, 13.6 Hz, 0.8H), 2.99 (d, *J* = 7.2 Hz, 0.4H), 3.06 (dd, *J* = 7.1, 13.6 Hz, 0.8H), 3.67 (s, 0.5H), 3.70 (s, 2.5H), 4.40 (q, *J* = 7.0 Hz, 0.2H), 4.80–4.88 (m, 1H), 5.14 (q, *J* = 7.3 Hz, 0.8H), 5.30 (brd, 0.8H), 5.37 (brd, 0.2H), 7.17–7.31 (m, 5H); ¹³C NMR (125 MHz, CDCl₃, mixture of rotamers) δ 14.2, 14.7, 28.3, 28.8, 31.4, 39.3, 40.5, 51.4, 51.6, 52.2, 52.5, 52.6, 54.9, 79.6, 126.8, 126.9, 128.4, 128.6, 129.4, 129.6, 136.2, 136.5, 154.9, 155.1, 171.2, 171.8, 172.0, 172.1; IR (neat) 3318, 2979, 1743, 1711, 1659, 1642, 1513, 1494, 1484, 1462, 1452, 1410, 1366, 1248, 1171 cm⁻¹; ESIMS *m/z*: calcd for C₁₉H₂₈N₂NaO₅ [M+Na]⁺, 387.1896; found, 387.1862.

2.2. General procedure for the preparation of piperazine-2,5-diones (*S,S*)-**5a–c** and (*S,S*)-**7a–c**

2.2.1. (3*S*,6*S*)-6-Benzyl-3-isopropyl-1-methylpiperazine-2,5-dione [(*S,S*)-**5a**]

A suspension of dipeptide (*S,S*)-**4a** (80 mg, 0.204 mmol) in a mixed solvent of H₂O (1.5 mL) with MeOH (0.5 mL) was irradiated at 170 °C for 10 min using an automated single-mode microwave synthesizer. The reaction mixture was treated with H₂O (2 mL) and then extracted with CHCl₃ (5 mL \times 2). The extract was dried over anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography [Silica Gel 60N: CHCl₃–MeOH (50:1)] to afford (*S,S*)-**5a** (47.3 mg, 89% yield). Colorless columns (CH₂Cl₂–AcOEt); mp 173–175 °C; $[\alpha]_{\text{D}}^{27} -53.8$ (*c* 1.00, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.49 (d, *J* = 6.8 Hz, 3H), 0.85 (d, *J* = 7.0 Hz, 3H), 1.35–1.46 (m, 1H), 2.96 (s, 3H), 3.26 (dd, *J* = 4.7, 14.2 Hz, 1H), 3.30 (dd, *J* = 5.0, 14.2 Hz, 1H), 3.60 (dd, *J* = 2.7, 5.9 Hz, 1H), 4.18 (brt, 1H),

6.34 (brs, 1H), 7.14–7.19 (m, 2H), 7.21–7.32 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 17.3, 19.3, 32.5, 33.6, 38.1, 60.8, 63.5, 127.4, 128.8, 129.9, 135.7, 165.5, 167.3; IR (KBr) 3247, 2972, 1687, 1661, 1456, 1301 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{15}\text{H}_{20}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$, 283.1422; found, 283.1401. Anal. Calcd for $\text{C}_{15}\text{H}_{20}\text{N}_2\text{O}_2$: C, 69.20; H, 7.74; N, 10.76. Found: C, 69.20; H, 7.88; N, 10.55%.

2.2.2. (3*S*,6*S*)-6-Benzyl-3-isobutyl-1-methylpiperazine-2,5-dione [(*S,S*)-**5b**]

Colorless columns (CH_2Cl_2 –AcOEt); mp 170–172 °C; $[\alpha]_{\text{D}}^{26}$ –11.8 (*c* 1.01, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ –0.22 (ddd, J = 4.5, 11.0, 13.7 Hz, 1H), 0.70 (d, J = 6.6 Hz, 3H), 0.75 (d, J = 6.6 Hz, 3H), 1.03 (ddd, J = 3.9, 10.3, 13.7 Hz, 1H), 1.28–1.40 (m, 1H), 3.07 (s, 3H), 3.16 (dd, J = 4.6, 14.0 Hz, 1H), 3.33 (dd, J = 3.7, 14.0 Hz, 1H), 3.74 (brdt, 1H), 4.20 (brt, 1H), 6.27 (brs, 1H), 7.09–7.15 (m, 2H), 7.25–7.34 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 20.4, 22.9, 23.8, 33.0, 36.6, 43.6, 53.3, 63.1, 127.7, 128.8, 130.2, 134.9, 166.7, 166.8; IR (KBr) 3609, 3520, 3328, 2956, 1681, 1649, 1468 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$, 297.1579; found, 297.1552. Anal. Calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_2$: C, 70.04; H, 8.08; N, 10.21. Found: C, 69.97; H, 8.11; N, 10.24%.

2.2.3. (3*S*,6*S*)-6-Benzyl-1,3-dimethylpiperazine-2,5-dione [(*S,S*)-**5c**]

Colorless plates (CH_2Cl_2 –AcOEt), mp 143.5–145.5 °C; $[\alpha]_{\text{D}}^{28}$ +30.2 (*c* 1.02, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 0.50 (d, J = 7.1 Hz, 3H), 3.08 (s, 3H), 3.18 (dd, J = 4.6, 14.1 Hz, 1H), 3.33 (dd, J = 3.7, 14.1 Hz, 1H), 3.87 (dq, J = 2.6, 7.1 Hz, 1H), 4.20 (brt, 1H), 6.27 (brs, 1H), 7.10–7.14 (m, 2H), 7.24–7.33 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 20.7, 33.0, 36.7, 51.1, 63.2, 127.6, 128.9, 130.2, 134.8, 166.5, 166.8; IR (KBr) 3181, 3137, 1681, 1632, 1477 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{13}\text{H}_{16}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$, 255.1109; found, 255.1099. Anal. Calcd for $\text{C}_{13}\text{H}_{16}\text{N}_2\text{O}_2$: C, 67.22; H, 6.94; N, 12.06. Found: C, 67.21; H, 6.86; N, 11.88%.

2.2.4. (3*S*,6*S*)-3-Benzyl-6-isopropyl-1-methylpiperazine-2,5-dione [(*S,S*)-**7a**]

Colorless prisms (CH_2Cl_2 –AcOEt); mp 133–134 °C; $[\alpha]_{\text{D}}^{29}$ –155.0 (*c* 1.10, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 1.06 (d, J = 7.0 Hz, 3H), 1.19 (d, J = 7.0 Hz, 3H), 2.25 (dsept, J = 4.2, 7.0 Hz, 1H), 2.80 (dd, J = 11.4, 13.5 Hz, 1H), 3.03 (s, 3H), 3.54 (dd, J = 3.2, 13.5 Hz, 1H), 3.75 (brd, 1H), 4.17 (brt, 1H), 5.67 (brs, 1H), 7.19–7.24 (m, 2H), 7.26–7.31 (m, 1H), 7.32–7.38 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 18.1, 19.7, 31.6, 34.6, 41.5, 57.0, 67.7, 127.4, 129.1, 129.4, 136.1, 165.6, 165.8; IR (KBr) 3207, 3133, 2984, 2972, 1690, 1636, 1456, 1412, 1336 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{15}\text{H}_{20}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$, 283.1422; found, 283.1438. Anal. Calcd for $\text{C}_{15}\text{H}_{20}\text{N}_2\text{O}_2$: C, 69.20; H, 7.74; N, 10.76. Found: C, 69.05; H, 7.87; N, 10.71%.

2.2.5. (3*S*,6*S*)-3-Benzyl-6-isobutyl-1-methylpiperazine-2,5-dione [(*S,S*)-**7b**]

Colorless columns (CH₂Cl₂–AcOEt); mp 105–109 °C; [α]_D¹⁸ –82.5 (*c* 1.02, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.88 (d, *J* = 6.7 Hz, 3H), 0.94 (d, *J* = 6.5 Hz, 3H), 1.15 (ddd, *J* = 5.2, 8.7, 14.1 Hz, 1H), 1.33 (ddd, *J* = 4.2, 8.8, 14.1 Hz, 1H), 1.77–1.88 (m, 1H), 2.95 (s, 3H), 2.99 (dd, *J* = 8.7, 13.6 Hz, 1H), 3.26 (dd, *J* = 3.7, 13.6 Hz, 1H), 3.73 (dd, *J* = 4.2, 8.7 Hz, 1H), 4.22 (brdt, 1H), 5.94 (brs, 1H), 7.20–7.22 (m, 2H), 7.26–7.30 (m, 1H), 7.31–7.35 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 21.9, 23.1, 24.8, 33.0, 41.1, 42.4, 57.1, 60.2, 127.4, 128.9, 129.9, 135.8, 165.4, 168.0; IR (KBr) 3242, 2960, 1683, 1638, 1442, 1344 cm⁻¹; ESIMS *m/z*: calcd for C₁₆H₂₂N₂NaO₂ [M+Na]⁺, 297.1579; found, 297.1578. Anal. Calcd for C₁₆H₂₂N₂O₂: C, 70.04; H, 8.08; N, 10.21. Found: C, 69.92; H, 8.12; N, 10.15%.

2.2.6. (3*S*,6*S*)-3-Benzyl-1,6-dimethylpiperazine-2,5-dione [(*S,S*)-**7c**]

Colorless needles (CH₂Cl₂–AcOEt), mp 126–129 °C; [α]_D²⁷ –47.8 (*c* 1.04, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.01 (d, *J* = 7.1 Hz, 3H), 2.92 (s, 3H), 3.09 (dd, *J* = 7.5, 13.7 Hz, 1H), 3.17 (dd, *J* = 3.9, 13.7 Hz, 1H), 3.78 (q, *J* = 7.1 Hz, 1H), 4.27 (brquint, 1H), 6.14 (brs, 1H), 7.18–7.23 (m, 2H), 7.25–7.30 (m, 1H), 7.31–7.36 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 18.0, 32.2, 41.1, 56.8, 57.5, 127.4, 128.8, 130.1, 135.5, 164.8, 168.6; IR (KBr) 3242, 1687, 1643, 1455, 1340, 1322 cm⁻¹; ESIMS *m/z*: calcd for C₁₃H₁₆N₂NaO₂ [M+Na]⁺, 255.1109; found, 255.1119.

2.3. General procedure for the preparation of piperazine-2,5-diones (*S,S*)-**8a–c**

2.3.1. (3*S*,6*S*)-3-Benzyl-6-isopropyl-1,4-dimethylpiperazine-2,5-dione [(*S,S*)-**8a**]

NaH (50–72%, 6.9 mg, 0.144 mmol) was added to a solution of (*S,S*)-**7a** (25 mg, 0.096 mmol) in anhydrous DMF (2 mL) and stirred at 0 °C for 15 min under argon. After adding MeI (8.96 μ L, 0.144 mmol), the mixture was stirred at 0 °C for 30 min under argon. The reaction mixture was treated with 1N HCl (2 mL) and then extracted with AcOEt (5 mL \times 3). The extract was washed with sat. Na₂S₂O₃ (5 mL) and H₂O (5 mL). The organic layer was dried over anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography [Silica Gel 60N: CHCl₃–MeOH (30:1)] to afford (*S,S*)-**8a** (24.1 mg, 91% yield). White solid; mp 77–80 °C; [α]_D³⁰ –81.4 (*c* 0.24, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.95 (d, *J* = 6.8 Hz, 3H), 1.02 (d, *J* = 6.9 Hz, 3H), 1.61–1.73 (m, 1H), 2.67 (s, 3H), 3.01 (s, 3H), 3.09 (dd, *J* = 7.7, 14.1 Hz, 1H), 3.38 (dd, *J* = 4.4, 14.1 Hz, 1H), 3.55 (d, *J* = 7.1 Hz, 1H), 4.12 (dd, *J* = 4.4, 7.7 Hz, 1H), 7.20–7.35 (m, 5H); ¹³C NMR (125 MHz, CDCl₃) δ 19.6, 20.6, 33.5, 33.9, 35.8, 40.3, 65.1, 68.9, 127.3, 128.9, 129.5, 137.1, 165.8, 166.4; IR (KBr) 2934, 2874, 1662, 1477, 1456, 1402 cm⁻¹; ESIMS *m/z*: calcd for C₁₆H₂₂N₂NaO₂ [M+Na]⁺, 297.1579; found, 297.1578.

2.3.2. (3*S*,6*S*)-3-Benzyl-6-isobutyl-1,4-dimethylpiperazine-2,5-dione [(*S,S*)-**8b**]

White solid; mp 144.5–145.5 °C; [α]_D³⁰ –5.5 (*c* 0.38, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.28 (ddd, *J*

= 5.2, 9.2, 14.1 Hz, 1H), 0.66 (ddd, $J = 4.2, 9.3, 14.1$ Hz, 1H), 0.72 (d, $J = 6.7$ Hz, 3H), 0.83 (d, $J = 6.5$ Hz, 3H), 1.66–1.75 (m, 1H), 2.86 (s, 3H), 2.94 (s, 3H), 3.17 (dd, $J = 4.6, 14.0$ Hz, 1H), 3.29 (dd, $J = 4.9, 14.0$ Hz, 1H), 3.61 (dd, $J = 4.2, 9.2$ Hz, 1H), 4.18 (brt, 1H), 7.09–7.14 (m, 2H), 7.22–7.32 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 21.5, 22.7, 25.1, 32.5, 32.8, 37.7, 42.4, 60.1, 64.0, 127.6, 128.9, 130.0, 135.7, 165.2, 166.6; IR (KBr) 2953, 2866, 1651, 1491, 1455, 1404 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{17}\text{H}_{24}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$, 311.1735; found, 311.1737.

2.3.3. (3*S*,6*S*)-3-Benzyl-1,4,6-trimethylpiperazine-2,5-dione [(*S,S*)-**8c**]

White solid, mp 123.5–127 °C; $[\alpha]_{\text{D}}^{30} -2.0$ (c 0.74, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 0.48 (d, $J = 7.1$ Hz, 3H), 2.82 (s, 3H), 3.03 (s, 3H), 3.15 (dd, $J = 4.4, 14.0$ Hz, 1H), 3.32 (dd, $J = 4.2, 14.0$ Hz, 1H), 3.69 (q, $J = 7.1$ Hz, 1H), 4.21 (brt, 1H), 7.07–7.12 (m, 2H), 7.21–7.26 (m, 1H), 7.26–7.31 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 17.5, 31.7, 32.7, 37.3, 57.6, 63.5, 127.5, 128.9, 130.1, 135.3, 164.5, 166.8; IR (KBr) 3476, 3385, 2942, 1651, 1493, 1453, 1405, 1346 cm^{-1} ; ESIMS m/z : calcd for $\text{C}_{14}\text{H}_{18}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$, 269.1266; found, 269.1288.

2.4. X-ray diffraction studies of (*S,S*)-**5a** and (*S,S*)-**7a**

2.4.1. (3*S*,6*S*)-6-Benzyl-3-isopropyl-1-methylpiperazine-2,5-dione [(*S,S*)-**5a**]

A colorless prism crystal of (*S,S*)-**5a** having approximate dimensions of 0.400 × 0.300 × 0.300 mm was mounted on a glass fiber. The data were collected at a temperature of -180 ± 1 °C to a maximum 2θ value of 54.8°. A total of 44 oscillation images were collected. A sweep of data was done using ω scans from 130.0 to 190.0° in 5.0° step, at $\chi = 45.0^\circ$ and $\phi = 80.0^\circ$. The exposure rate was 130.0 [sec./°]. A second sweep was performed using ω scans from 0.0 to 160.0° in 5.0° step, at $\chi = 45.0^\circ$ and $\phi = 260.0^\circ$. The exposure rate was 130.0 [sec./°]. The crystal-to-detector distance was 127.40 mm. Readout was performed in the 0.100 mm pixel mode. The absolute configuration was deduced from the synthetic pathway.

Deposition number: CCDC-1429971

Empirical formula: $\text{C}_{15}\text{H}_{20}\text{N}_2\text{O}_2$

Formula weight: 260.34

Crystal system, Space group: orthorhombic, $P 2_12_12_1$ (No. 19)

Unit cell dimensions: $a = 8.1071(3)$ Å, $b = 12.3418(4)$ Å, $c = 13.9765(5)$ Å

Volume: 1398.44(8) Å³

Z, Calculated density: 4, 1.236 g/cm³

Absorption coefficient $\mu(\text{Mo-K}\alpha)$: 0.826 cm^{-1}

Final R indices [$I > 2.00\sigma(I)$]: R1 = 0.0340, wR2 = 0.1042.

2.4.2. (3*S*,6*S*)-3-Benzyl-6-isopropyl-1-methylpiperazine-2,5-dione [(*S,S*)-**7a**]

A colorless block crystal of (*S,S*)-**7a** having approximate dimensions of 0.300 × 0.250 × 0.200 mm was mounted on a glass fiber. The data were collected at a temperature of -150 ± 1 °C to a maximum 2θ value of 54.8°. A total of 44 oscillation images were collected. A sweep of data was done using ω scans from 130.0 to 190.0° in 5.0° step, at $\chi = 45.0^\circ$ and $\phi = 0.0^\circ$. The exposure rate was 130.0 [sec./°]. A second sweep was performed using ω scans from 0.0 to 160.0° in 5.0° step, at $\chi = 45.0^\circ$ and $\phi = 180.0^\circ$. The exposure rate was 130.0 [sec./°]. The crystal-to-detector distance was 127.40 mm. Readout was performed in the 0.100 mm pixel mode. The absolute configuration was deduced from the synthetic pathway.

Deposition number: CCDC-1429972

Empirical formula: C₁₅H₂₀N₂O₂

Formula weight: 260.34

Crystal system, Space group: orthorhombic, *P* 2₁2₁2₁ (No. 19)

Unit cell dimensions: a = 8.4741(5) Å, b = 12.8041(7) Å, c = 12.7997(9) Å

Volume: 1388.8(2) Å³

Z, Calculated density: 4, 1.245 g/cm³

Absorption coefficient $\mu(\text{Mo-K}\alpha)$: 0.832 cm⁻¹

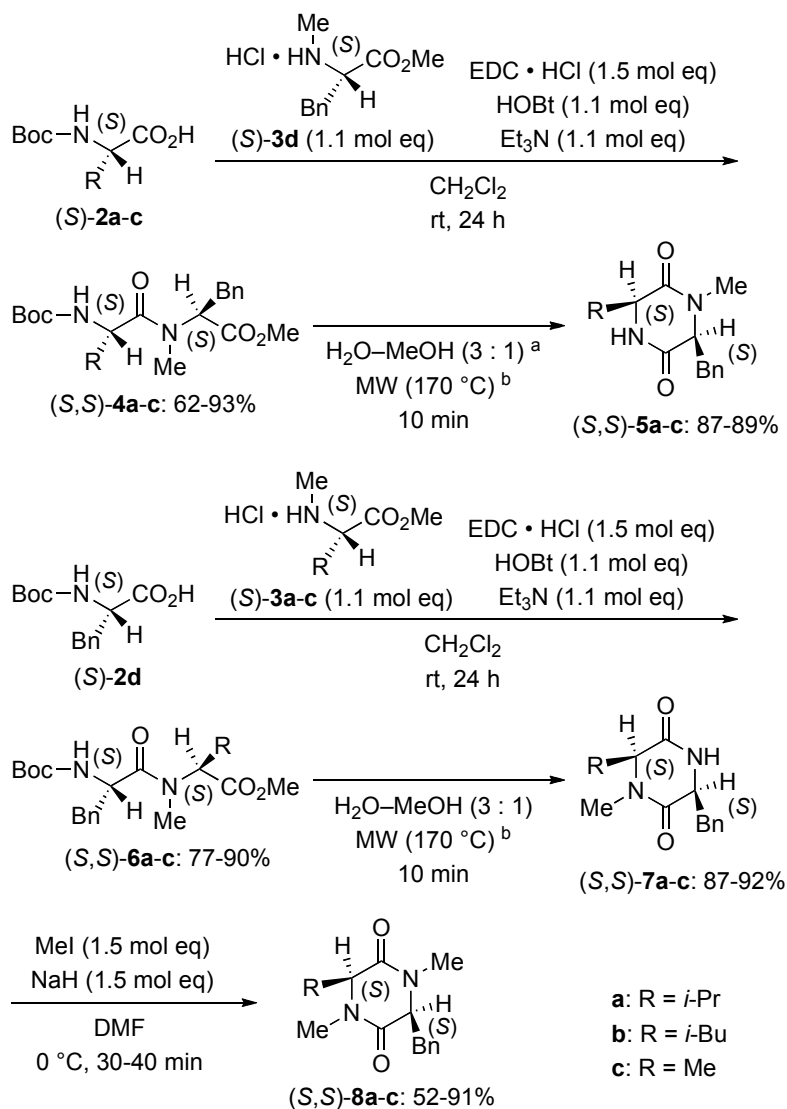
Final R indices [$I > 2.00\sigma(I)$]: R1 = 0.0326, wR2 = 0.0996.

3. Results and discussion

3.1. Synthesis of N-methylated and N,N'-dimethylated 2,5-DKPs

N1-methylated 2,5-DKPs (*S,S*)-**5a–c**, N4-methylated 2,5-DKPs (*S,S*)-**7a–c**, and *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a–c** were synthesized as shown in Scheme 1. Methyl, isopropyl, and isobutyl groups, which came from L-alanine, L-valine, and L-leucine, were chosen as 3-alkyl substituents of (*S,S*)-**1a–c**. Condensation of Boc-L- α -amino acids (*S*)-**2a–c** with *N*-methyl-L-phenylalanine methyl ester hydrochloride [(*S*)-**3d**] using 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDC•HCl) as a coupling reagent in the presence of 1-hydroxybenzotriazole (HOBt) and triethylamine afforded the dipeptides (*S,S*)-**4a–c**. Subsequently, microwave-assisted removal of the Boc group followed by intramolecular cyclization furnished the N1-methylated 2,5-DKPs (*S,S*)-**5a–c** by a one-pot reaction [34].

N4-methylated 2,5-DKPs (*S,S*)-**7a-c** were also synthesized in a similar way. Furthermore, *N*-methylation of (*S,S*)-**7a-c** afforded *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a-c**.



Scheme 1. Synthesis of N1-methylated 2,5-DKPs (*S,S*)-**5a-c**, N4-methylated 2,5-DKPs (*S,S*)-**7a-c**, and *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a-c**.

^a H₂O was used alone as the solvent in the reaction of (*S,S*)-**4a**.

^b InitiatorTM 60 (Biotage AB).

3.2. Crystallographic studies of N1- and N4-methylated 2,5-DKPs

Conformations of the 2,5-DKPs (*S,S*)-**5a** and (*S,S*)-**7a** in the solid state were examined by single crystal X-ray crystallography as shown in Figures 2 and 3. Unfortunately, the results showed that N1-methylated 2,5-DKP (*S,S*)-**5a** adopted a folded conformation, and the perpendicular distance (2.3778 Å) of the methine hydrogen atom [H(10)] of the isopropyl group from the plane of the benzene ring (P_{benzene}) was clearly shorter than the conventional van der Waals limit (2.9 Å: 1.2 Å for C-H plus 1.7 Å for a half

thickness of the aromatic molecule) (Figure 2) [35–38]. The C(12)–H(10)•••C_{benzene} (the centroid of the benzene ring) angle of 173.74° was also observed. Therefore, the crystal structure of N1-methylated 2,5-DKP (*S,S*)-**5a** strongly suggested the existence of an intramolecular CH/π interaction [35–43] as a stabilizing factor for the folded conformation of (*S,S*)-**5a**. Intermolecular CH/π interactions of (*S,S*)-**5a** leading to a molecular packing were not observed. However, N4-methylation of (*S,S*)-**1a** successfully changed the folded conformation to the extended one. As shown in Figure 3, the crystal structure of N4-methylated 2,5-DKP (*S,S*)-**7a** was found to exist in an extended conformation in which the benzyl moiety was apart from the 2,5-DKP ring [44].

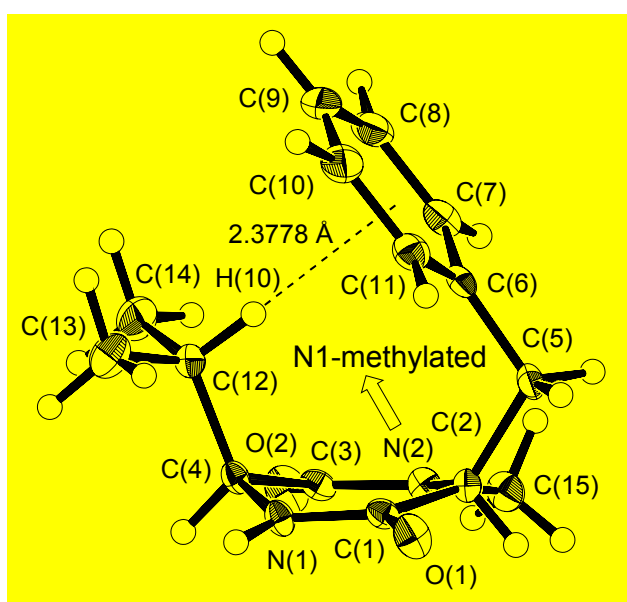


Fig. 2. ORTEP drawing of N1-methylated 2,5-DKP (*S,S*)-**5a** with 50% probability ellipsoids. Selected distances (Å) and angles (°) are: H(10)•••P_{benzene} = 2.3778, C(12)•••P_{benzene} = 3.3745(12), C(5)•••C(12) = 4.2910(15), C(12)–H(10)•••C_{benzene} = 173.74, C(2)–C(5)–C(6) = 113.63(8), C(4)–C(12)–H(10) = 106.0.

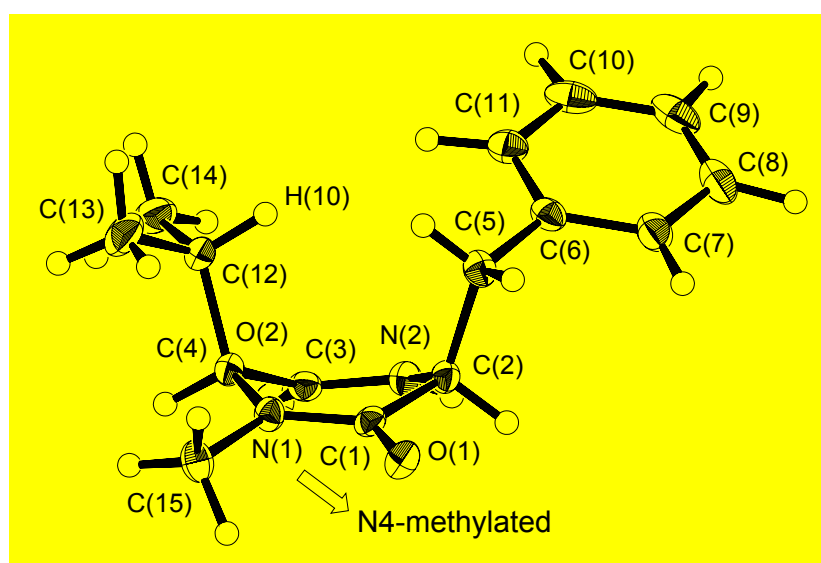


Fig. 3. ORTEP drawing of N4-methylated 2,5-DKP (*S,S*)-**7a** with 50% probability ellipsoids. Selected distances (Å) and angles (°) are: C(5)•••C(12) = 3.7541(16) , C(2)–C(5)–C(6) = 112.48(9), C(4)–C(12)–H(10) = 107.2.

3.3. ¹H NMR analysis of N-methylated and *N,N'*-dimethylated 2,5-DKPs

Next, we investigated the conformational aspects of N1-methylated 2,5-DKPs (*S,S*)-**5a–c**, N4-methylated 2,5-DKPs (*S,S*)-**7a–c**, and *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a–c** in solution by ¹H NMR spectroscopy. Selected chemical shifts of (*S,S*)-**5a–c**, (*S,S*)-**7a–c**, and (*S,S*)-**8a–c** in ¹H NMR spectra (500 MHz, CDCl₃) are given in Figure 4. Protons of the *cis*-substituent (on the same side of the 2,5-DKP ring as the benzyl moiety) of (*S,S*)-**5a–c** were found to appear at higher magnetic fields than those of (*S,S*)-**7a–c**. In particular, the isopropyl protons (H_A and Me_A) of (*S,S*)-**5a**, isobutyl protons (H_C and H_D) of (*S,S*)-**5b**, and methyl protons (Me_E) of (*S,S*)-**5c** exhibited remarkable upfield chemical shifts. The *trans*-protons H_B (on the opposite side of the 2,5-DKP ring with the benzyl moiety) of (*S,S*)-**5a–c** and (*S,S*)-**7a–c** did not show large differences in chemical shifts. The upfield chemical shifts of protons of the *cis*-substituent of (*S,S*)-**5a–c** could be attributed to the strong shielding effect of the benzene ring of the benzyl moiety. Therefore, it was suggested that N1-methylated 2,5-DKPs (*S,S*)-**5a–c** adopted a folded conformation and the protons of the *cis*-substituent were close to the benzene ring in solution. On the other hand, N4-methylated 2,5-DKPs (*S,S*)-**7a–c** seemed to prefer an extended conformation because the signals of protons of the *cis*-substituent were not observed in higher magnetic fields such as that of (*S,S*)-**5a–c**. However, *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a–c** showed upfield chemical shifts of protons of the *cis*-substituent compared to those of (*S,S*)-**7a–c**, which suggests the predominance of the folded conformation in (*S,S*)-**8a–c** relative to (*S,S*)-**7a–c**.

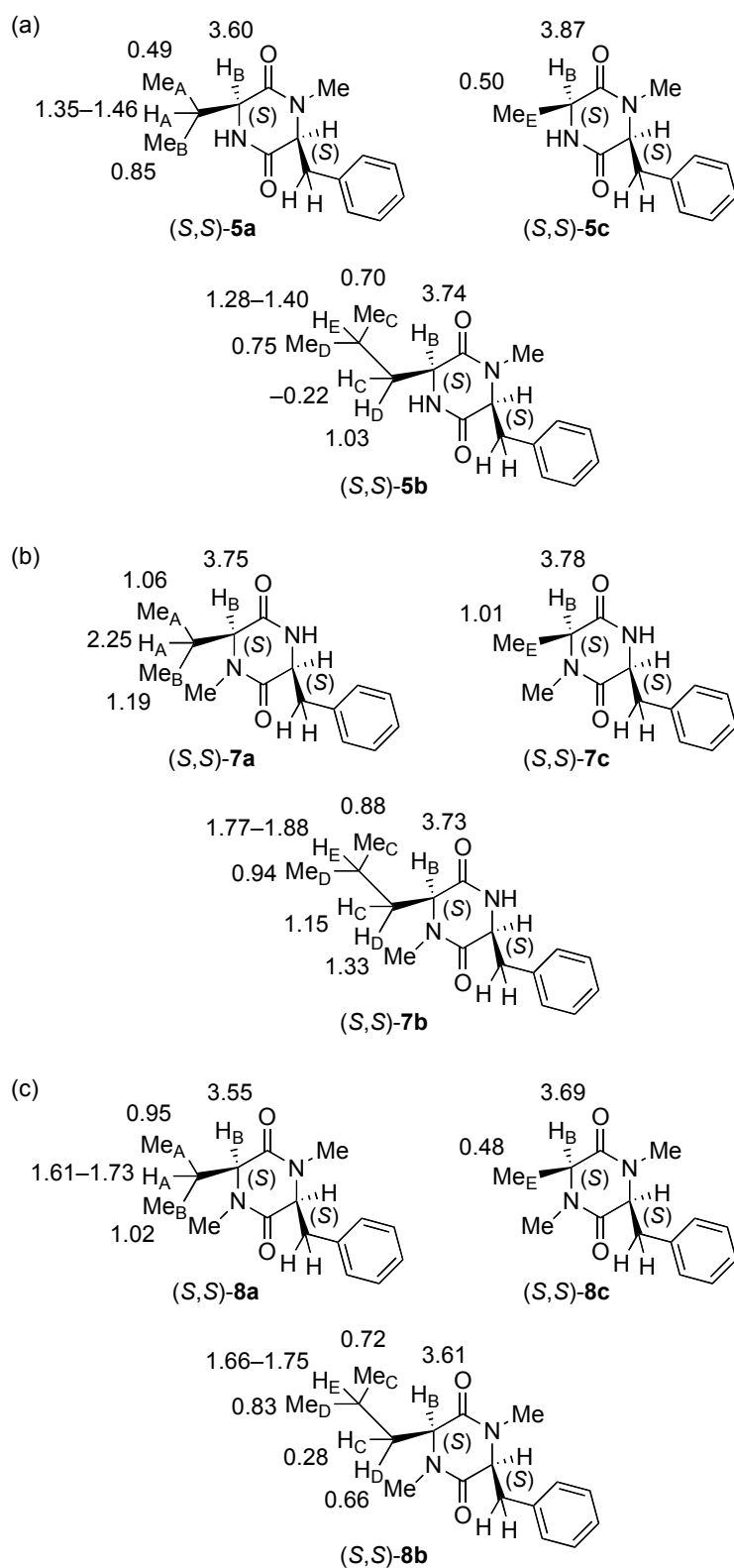


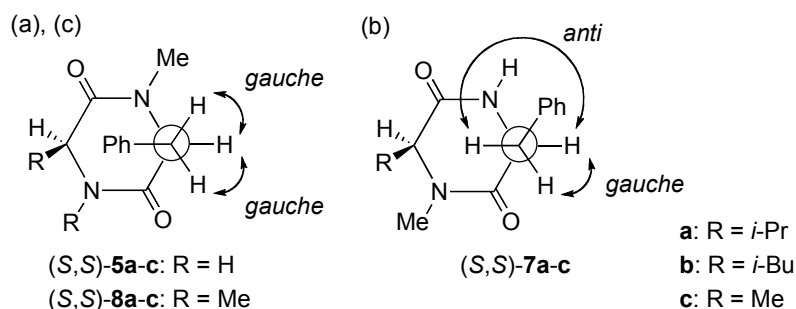
Fig. 4. Selected chemical shifts of (a) N1-methylated 2,5-DKPs (*S,S*)-**5a–c**, (b) N4-methylated 2,5-DKPs (*S,S*)-**7a–c**, and (c) *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a–c** in ^1H NMR (500 MHz, CDCl_3) analysis.

The vicinal coupling constants (J_1 and J_2) between the benzylic protons and the adjacent methine proton of N1-methylated 2,5-DKPs (*S,S*)-**5a–c**, N4-methylated 2,5-DKPs (*S,S*)-**7a–c**, and *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a–c** are listed in Table 1. The results showed that similar values of the vicinal coupling

constants ($J_1 = 3.7\text{--}4.7$ Hz, $J_2 = 4.6\text{--}5.0$ Hz) were observed for (*S,S*)-**5a–c**. Therefore, a *gauche* relationship was presumed to exist in the CDCl₃ solution between the two benzylic hydrogens and the adjacent methine hydrogen, indicating that the N1-methylated 2,5-DKPs (*S,S*)-**5a–c** adopted the folded conformation [14]. On the other hand, it was found that J_1 and J_2 were quite different ($J_1 = 3.2\text{--}3.9$ Hz, $J_2 = 7.5\text{--}11.4$ Hz) in N4-methylated 2,5-DKPs (*S,S*)-**7a–c**. The large difference in the vicinal coupling constants suggested that one of the two benzylic hydrogens and the adjacent methine hydrogen could be in a *gauche* relationship, while the other benzylic hydrogen and adjacent methine hydrogen were in an *anti*-relationship [14]. Therefore, it was strongly suggested that the extended conformation of (*S,S*)-**7a–c** was predominant in CDCl₃ solution. The values of the vicinal coupling constants for *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a–c** were similar to those for (*S,S*)-**5a–c**, and the preference for the folded conformation of (*S,S*)-**8a–c** was estimated. It is remarkable that differences of the vicinal coupling constants between (*S,S*)-**5a–c** and (*S,S*)-**7a–c** were correlated to the bulkiness of 3-alkyl substituents (methyl, isopropyl, and isobutyl groups) of the corresponding 2,5-DKPs. In the case of (*S,S*)-**5a** and (*S,S*)-**7a**, a methine carbon is directly connected to the 3-position of the 2,5-DKP ring and a larger difference of vicinal coupling constants ($J_1 = 3.2$ Hz, $J_2 = 11.4$ Hz) is observed. Thus, the ¹H NMR spectra of (*S,S*)-**5a** and (*S,S*)-**7a** showed a good agreement with the conformations in the solid state established by X-ray crystallography, as depicted in Figures 2 and 3.

Table 1

Vicinal coupling constants (J_1 and J_2) between the benzylic protons and the adjacent methine proton (a) N1-methylated 2,5-DKPs (*S,S*)-**5a–c**, (b) N4-methylated 2,5-DKPs (*S,S*)-**7a–c**, and (c) *N,N'*-dimethylated 2,5-DKPs (*S,S*)-**8a–c** in ¹H NMR (500 MHz, CDCl₃) analysis.



Compounds	R	J_1	J_2
(<i>S,S</i>)- 5a	<i>i</i> -Pr	4.7	5.0
(<i>S,S</i>)- 5b	<i>i</i> -Bu	3.7	4.6
(<i>S,S</i>)- 5c	Me	3.7	4.6
(<i>S,S</i>)- 7a	<i>i</i> -Pr	3.2	11.4
(<i>S,S</i>)- 7b	<i>i</i> -Bu	3.7	8.7
(<i>S,S</i>)- 7c	Me	3.9	7.5
(<i>S,S</i>)- 8a	<i>i</i> -Pr	4.4	7.7
(<i>S,S</i>)- 8b	<i>i</i> -Bu	4.6	4.9
(<i>S,S</i>)- 8c	Me	4.2	4.4

4. Conclusions

A series of novel N1- and/or N4-methylated 2,5-DKPs [(*S,S*)-**5a–c**, (*S,S*)-**7a–c**, and (*S,S*)-**8a–c**] have been prepared and fully characterized. Their conformational aspects have been confirmed by ¹H NMR spectroscopy in solution. Single crystal X-ray structural analysis of (*S,S*)-**5a** and (*S,S*)-**7a** has also been performed. In conclusion, N1-methylation and N1,N4-dimethylation have no influence on the folded conformation of (*S,S*)-**1a–c**. However, we have succeeded in changing the folded conformation of (*S,S*)-**1a–c** to the extended conformation by N4-methylation of the 2,5-DKP ring. This simple chemical modification could be applied to access novel functional molecules such as organocatalysts and pharmaceuticals based on the 2,5-DKP ring.

Acknowledgements

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Appendix A. Supplementary data

Deposition number CCDC-1429971 for compound (*S,S*)-**5a** and CCDC-1429972 for compound (*S,S*)-**7a** contains the supplementary crystallographic data. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

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Highlights

- A series of novel N-methylated and N,N'-dimethylated 2,5-DKPs were prepared.
- Conformational analysis of prepared N-methylated and N,N'-dimethylated 2,5-DKPs based on ¹H NMR spectroscopy were performed.
- Single crystal X-ray structural analysis of prepared N1-methylated and N4-methylated 2,5-DKPs were performed.
- N4-methylation of mono-benzylated 2,5-DKPs, which overcome the intramolecular CH/π interaction, was found to change their conformation from folded to extended.