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Preliminary evaluation of associated viruses in production systems of cape gooseberry, purple passion fruit, and rose

Evaluación preliminar de virus asociados a sistemas productivos de uchuva, gulupa y rosa

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Fruit deformation and leaf blistering in purple passion fruit (*Passiflora edulis* Sims).

Photo: J. Cutler

ABSTRACT

Plant viruses may pose a threat to crops in Colombia. To evaluate the potential risk of yield losses due to plant virus infection, a literature analysis followed by a first field study was carried out focusing on purple passion fruit (*Passiflora edulis* Sims), cape gooseberry (*Physalis peruviana* L.), and ornamental rose (*Rosa* sp.), which are important Colombian exports. Over the past three years, plant material was collected from 21 farms in

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Cundinamarca and Boyacá, Colombia, two regions that are in close proximity to El Dorado International Airport, the country's largest air freight terminal. Plants were visually inspected and subsequently tested by bioassay and serological methods. Overall, in the samples investigated by the two diagnostic methods, plant viruses were detected. Detected viruses belong to the genus *Poty-*, *Tobamo-*, *Nepo-*, *Ilar-*, and *Tospovirus*. The extent of the distribution and occurrence of these viruses in each crop has to be determined in a representative field study. Such a monitoring program could be supported by a standardized farmer interview. The development of suitable plant virus diagnostic and management tools is the focus of a cooperation project between German and Colombian universities, the Colombian Agricultural Institute (ICA), the Colombian Corporation of Agricultural Investigation (AGROSAVIA) and the International Center for Tropical Agriculture (CIAT).

Additional key words: *Physalis peruviana*, *Rosa* sp., *Passiflora edulis* Sims, crop protection, plant disease diagnosis.

RESUMEN

Los virus representan una amenaza para las plantas cultivadas en Colombia, y algunos estudios indican que las pérdidas económicas causadas por estos problemas fitosanitarios podrían evitarse mediante procedimientos preventivos estándar. Como objeto de estudio, se seleccionaron tres importantes productos de Colombia, como lo son: rosa ornamental (*Rosa* sp.), uchuva (*Physalis peruviana* L.) y gulupa (*Passiflora edulis* Sims). En el presente artículo, se muestran los primeros desarrollos resultantes del monitoreo de virus en estas plantas, para determinar el estado de las virosis en fincas colombianas. Se realizaron análisis serológicos de 21 fincas en Cundinamarca y Boyacá, Colombia, en 2016-18. El objetivo de esta investigación fue el desarrollo de un protocolo piloto para el diagnóstico rutinario, que se pueda aplicar en un programa de certificación de material vegetal para la determinación de la presencia de virus, en varios productos hortícolas colombianos. Con base en este protocolo, se pueden determinar los riesgos que representan ciertos virus y se puede considerar la necesidad de certificar el material de siembra evaluado según la presencia de virus. Se están desarrollando herramientas de diagnóstico confiables y prácticas, para la detección de los virus más relevantes, en un proyecto conjunto entre universidades alemanas y colombianas, el Instituto Colombiano de Agropecuario (ICA), la Corporación Colombiana de Investigación Agropecuaria (AGROSAVIA) y el Centro Internacional de Agricultura Tropical (CIAT).

Palabras clave adicionales: *Physalis peruviana*, *Rosa* sp., *Passiflora edulis* Sims, protección de cultivos, diagnóstico de enfermedades de plantas.

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INTRODUCTION

Colombia's agricultural export sector has advanced and in parallel the need for phytosanitary control has become more important. There is knowledge about the presence of pests, fungi, and bacterial diseases, but only little about plant viruses and their potential impact on plant or fruit quality and yield quantity. We focused on cape gooseberry (*Physalis peruviana*), rose (*Rosa* sp.), and purple passion fruit (*Passiflora edulis* Sims), three important plant cultivars for Colombia's export market.

Cape gooseberry, known as uchuva in Colombia, belongs to the family *Solanaceae* and genus *Physalis*. Colombia is the largest producer of uchuva followed by South Africa (Mazorra *et al.*, 2006). Uchuva

started to have commercial importance in Colombia in 1985 (Chaves, 2006). Colombia is the country with the largest area of cultivation, from 800 to 1,000 ha producing between 15 to 18 t ha⁻¹ (Fischer *et al.*, 2014). The quality of domestic and exportable postharvest uchuva is largely correlated with proper plant pathogen management. Cape gooseberry is affected by pests and several fungal, bacterial and viral pathogens. Increasing occurrence may be due to uncontrolled distribution of primary planting material among the farmers. Fungal pathogens such as *Fusarium oxysporum*, *Alternaria* spp., *Cladosporium* spp., *Phytophthora infestans*, *Cercospora* spp., and *Phoma* spp., the bacterial pathogen *Ralstonia solanacearum*, and plant viruses belonging to the genus *Alfamovirus*,

Bigeminivirus, *Comovirus*, *Cucumovirus*, *Fabavirus*, *Furovirus*, *Hybrigeminivirus*, *Ilarvirus*, *Luteovirus*, *Nepovirus*, *Orthotospovirus*, *Potexvirus*, *Potyvirus*, *Tobamovirus*, and *Tymovirus* have been reported to occur in cape gooseberry (Rodríguez *et al.*, 2016; Aguirre-Ráquira *et al.*, 2014). But only few studies present deeper research into the complete genomic sequence and the complex relationship between genotype and phenotype of such plant viruses. For instance, Gutiérrez *et al.* (2015) present the complete genome sequence of an isolate of *Potato virus X* (PVX) infecting cape gooseberry in Colombia.

Purple passion fruit, known as gulupa in Colombia, belongs to the family *Passifloraceae* and genus *Passiflora*. The main diseases infecting gulupa in Colombia were summarized by Gutiérrez *et al.* (2015). They include the bacteria *Xanthomonas axonopodis* and the viruses *Soybean mosaic virus* (SMV) and *Cucumber mosaic virus* (CMV). On a global level further viruses have been described: *Passion fruit ringspot virus* (PFRSV), *Passiflora virus Y* (PaVY), *East Asian passiflora virus* (EAPV), *Okra mosaic virus* (OkMV), *Maracuja mosaic virus* (MarMV), *Passion fruit crinkle virus* (PCV), *Passion fruit rhabdovirus* (PRV), *Passiflora latent virus* (PLV), *Passion fruit mottle virus* (PFMoV), *Purple granadilla mosaic virus* (Gil *et al.*, 2017), and *Passion fruit yellow mosaic virus* (PFYMV) (Morales *et al.*, 2002).

Ornamental roses are perennial flowering plants of the genus *Rosa*, in the family *Rosaceae*. The floral industry is dominated by Holland, which produces 83% of the world's cut flowers, followed by Colombia, the second largest exporter worldwide (PMA, 2015). The main exported flower from Colombia is the rose which totals 365 million USD, and 70% of its cut roses are exported to the USA (PMA, 2015). Global studies have reported the following viruses to infect rose: *Prunus necrotic ringspot virus* (PNRSV) (Fajardo *et al.*, 2015), *Apple mosaic virus* (ApMV) (Sertkaya, 2010; Abtahi, 2015), *Arabidopsis mosaic virus* (ArMV) (Abtahi, 2015), *Tobacco ringspot virus* (TRSV) (McDaniel *et al.*, 1971), *Tomato ringspot virus* (ToRSV) (Halliwell and Milbrath, 1962), *Raspberry ringspot virus* (RRSV) (Von Bargaen *et al.*, 2015), *Rose yellow leaf virus* (RYLV) (Mollov *et al.*, 2014), and *Rose rosette virus* (RRV) (Laney *et al.*, 2011).

To extend the knowledge on both the occurrence of plant viruses in cape gooseberry, purple passion fruit and ornamental roses and their impact on crop yield and quality, we conducted a first field study. Biological and serological laboratory investigations completed the visual inspection of selected fields in the main growing area of these crops.

MATERIAL AND METHODS

Forty-four fields on 21 farms were inventoried for the incidence of virus-suspected symptoms in cape gooseberry, purple passion fruit, and rose in one of the main exporting Colombian cultivation areas (Fig. 1). Visual inspection is always the first step to identify the cause of plant damage. On the basis of the observed symptoms, laboratory investigations can be initiated to confirm the particular suspicion. We focused on virus-suspected symptoms such as chlorotic discoloration, leaf blistering, necrosis, stunting, and partial decline. Mechanical inoculation of the putative viral pathogen with the diseased tissue on indicator plants was applied to verify the infectivity of the agent. Different enzyme-linked immune assays (ELISA) were used to identify the viral pathogens.

Sampling

The field survey was carried out from 2016 to 2018 and comprised 21 national and export production sites in the Cundinamarca and Boyaca departments of Colombia (Fig. 1). The distribution of the sites visited were 10 cultivated farms of cape gooseberry, five of purple passion fruit and six of ornamental roses. Initial assessment of the health status of the plants in the field was conducted with a stratified random sampling pattern according to Delp *et al.* (1986) (Fig. 1). The distribution of the sites visited were 10 cultivated farms of cape gooseberry, five of purple passion fruit and six of ornamental roses. This pattern divides the field into regions of higher and lower disease risk. The disease foci were established by the general managers of each farm. As recommended, samples were collected from disease foci, as well as along other parts of the fields (Mavrodieva, 2004). Percent error is reduced using this sampling design compared to systematic entire field X and W shaped patterns or diagonal or partial field sampling patterns (Fletcher, 2006). To represent the impacted area, 10 to 20 samples were taken based on farm size, if procurable. Samples were collected in BIOREBA® extraction bags, transported in cooler bags and stored at 4°C before being investigated.

Mechanical inoculation

Bioassays are used to diagnose a virus by its reactions in a variety of plant species, to test the infectivity of an isolate using local lesion hosts, and to propagate viruses. According to Hull (2009) plant homogenate was rubbed onto the surface of the leaf of the test plant to break the surface cells without causing too

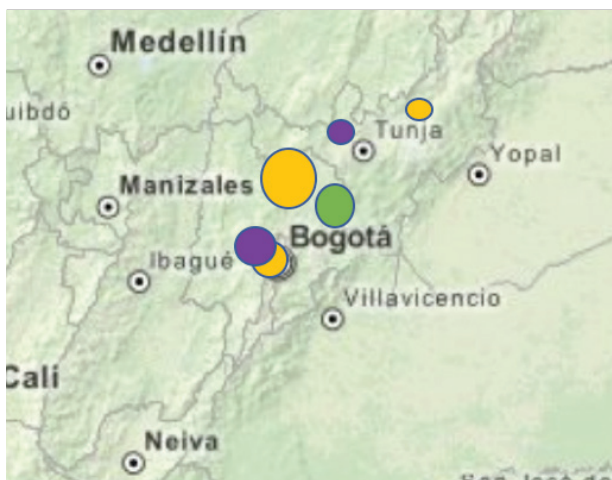


Figure 1. Sampling sites in Cundinamarca and Boyacá, Colombia: cape gooseberry (orange), purple passion fruit (purple), ornamental rose (green).

much mechanical damage. *Chenopodium quinoa*, *Nicotiana benthamiana*, *Nicotiana clevelandii*, *Nicotiana tabacum* L. cv. Samsun-NN, and *Cucumis sativus* were used as test plants.

Enzyme-linked immunosorbent assay (ELISA)

The detection of plant viruses using the ELISA was introduced to plant virology in the late 1970s. This method was selected among the diagnostic techniques due to its affordability, extreme sensitivity, and potential for measurement (Clark and Adams, 1977). ELISA was conducted with leaf material at Humboldt University (Berlin, Germany) and BIOREBA laboratories (Reinach, Switzerland) using available antibodies of previously described known viruses: *Cucumber Mosaic Virus* (CMV), *Potato virus Y* (PVY), *Tobacco mosaic virus* (TMV), and *Tomato spotted wilt virus* (TSWV) in passion fruit and cape gooseberry, *Prunus necrotic ringspot virus* (PNRSV), *Tobacco streak virus* (TSV), *Arabis mosaic virus* (ArMV), and *Tomato ringspot virus* (ToRSV) in rose, and *Soybean mosaic virus*

(SMV) in passion fruit. In addition to these plant virus species specific tests carried out in the Double Antibody sandwich-ELISA format, a Plate-Trapped Antibody (PTA)-ELISA using a *Potyvirus*-group specific antibody was performed to test plant material of all three cultivars. All tests were carried out according to protocols recommended by the supplier.

RESULTS AND DISCUSSION

Virus suspected symptoms were observed in all three crops: cape gooseberry, purple passion fruit and ornamental roses (Fig. 2). Plants showed mild to extreme yellowing, browning, stunting, spots, mosaics, curling, blistering, mottling, chlorosis, ringspots, deformation, discoloration, vein clearing, and variegation in leaves, fruits, and flowers. A total of 320 samples of 113 cape gooseberry, 112 purple passion fruit, and 95 ornamental roses were collected and examined for different globally known viruses in the laboratory. After specific viruses were not detectable in our samples [e.g. Potyvirus-group, PVY, TMV, CMV (in all cultivars), ArMV (in rose), and ToRSV, TSWV (in physalis)], testing of these viruses was discontinued. The SMV antibody was only tested for samples collected in 2018, as it was only first developed for passion fruit by BIOREBA at this time. Overall, various viruses have been detected by ELISA so far, yet none of the detected viruses could be transmitted by mechanical inoculation to herbaceous test plants.

All passion fruit farms had plants with symptoms of wilting, yellowing in leaves, distorted, decayed, and/or misshapen fruit with a similar scattered distribution in high and low areas. In three of the five farms inspected, passion fruit showed unique zebra striped mottling or blistering symptoms of leaves and fruits. Only four out of 112 samples tested were infected with *Soybean mosaic virus* (Tab. 1). Applying the ELISA this infection was detected first by a genus-specific

Table 1. Detection of particular plant viruses in fruit and leaf samples of diseased cape gooseberry, purple passion fruit and ornamental roses (ArMV: *Arabis mosaic virus*, PNRSV: *Prunus necrotic ringspot virus*, PVY: *Potato virus Y*, TMV: *Tobacco mosaic virus*, ToRSV: *Tomato ringspot virus*, TSV: *Tomato streak virus*, TSWV: *Tomato spotted wilt virus*, SMV: *Soybean mosaic virus*). Numbers: virus infected plants/tested plants/collected plants; nt: not tested).

Crop	ArMV	PNRSV	Potyvirus	PVY	TMV	ToRSV	TSV	TSWV	SMV
Purple passion fruit	nt	nt	4//101/112	0/68/112	0/68/112	nt	nt	0/33/112	4//64/112
Cape gooseberry	nt	nt	11//113/113	3/113/113	0/89/113	nt	nt	0/51/113	nt
Ornamental roses	0/82/95	47/95/95	1/59/95	nt	nt	0//49/95	3/95/95	nt	nt



Figure 2. Characteristic symptoms associated with plant virus infection. Row 1: purple passion fruit with discoloration (left), leaf with mosaic (right top) and blistering (right below). Row 2: cape gooseberry with yellow and stunting (left) and vein necrosis (right). Row 3: ornamental rose leaf with oak leaf pattern (left) and mottling (right).

antibody and confirmed to be *Soybean mosaic virus* by using a species specific antibody which was recently developed by BIOREBA to be applied in passion fruit. The four samples, each representing an individual plant, came from two different farms.

Cape gooseberry plants from all farms exhibited similar symptoms of yellowing, leaf spots, mosaic, and distorted fruits. But only 11 out of 113 samples proved to be infected with one of the tested plant viruses (Tab. 1) and originated from seven farms. Those plants are infected with a potyvirus, leaf material of three of them reacted with an antibody against PVY, the type member of the genus potyvirus. Further studies should identify and characterize the Potyvirus infecting cape gooseberry. It might be a distinct strain of PVY, or a potyvirus not known in cape gooseberry so far or even not at all.

In all of the six rose farms that were involved in the field survey, there was at least one sample containing a plant virus in a scattered distribution in rows. An infection with PNRSV could be detected in about 50% of tested roses, 3 out of 95 plants tested plants revealed an infection with TSV, one with a *Potyvirus* (Tab. 1). Most plants that tested positive for PNRSV in ELISA showed a characteristic chlorotic line pattern, named oak leaf pattern. In four farms, all symptomatic leaf samples taken from the variety Freedom were PNRSV-infected independent on the farm they were taken. Samples of the varieties Topaz and Blues were infected with that virus in two of the six farms. This distribution and the fact that no insect vector is known, in spite of extensive surveys suggests that the virus was introduced to the farm with the propagation material, rootstocks or scions.

Significant research on viruses in Colombia has been carried out in recent years. A novel ilarvirus infecting purple passion fruit was detected in Cundinamarca by RT-PCR in the Humboldt University in 2018. Vaca-Vaca *et al.* (2017) characterized the complete genome sequence of a begomovirus infecting yellow passion fruit (*Passiflora edulis*) in Valle del Cauca. In Antioquia, Gallo-Garcia *et al.* (2018) have reported the genome sequence of a novel ilarvirus infecting physalis. It is advisable that further research on the presence of viruses in ornamental roses in Colombia be carried out.

RECOMMENDATIONS AND CONCLUSIONS FROM THIS STUDY

Our results are a preliminary investigation of the occurrence of plant viruses in Cape gooseberry, purple passion fruit and ornamental roses in Colombia and do not give a reliable countrywide survey in terms of region, climatic conditions, mode of cultivation and

cultivars. In this first field survey few plant viruses could be detected in the 320 samples. The damage of the plants are therefore either due to other harmful organisms, or caused by previously untested or unknown viruses. A state of the art high-throughput sequencing (NGS) approach should be applied to discover more viruses. With such an approach novel viruses and/or virus complexes can be determined or excluded as the causes of the observed damage.

On the basis of an intensive monitoring and screening at different Colombian production sites the occurrence and impact of particular plant viruses can be determined and economic thresholds can be estimated. With this research, management programs can be recommended.

Such programs would involve training of farmers in recognizing early symptoms of a disease, on-site diagnostic tools to be used by consultants, sanitation measures like disinfection of knives while grafting, and certification of propagation material. Certification of mother seeds, rootstocks and scions to be virus-tested can prevent the transmission and distribution of plant viruses in the field and country, respectively. Farmers propagating their own plants should be aware that even mother plants infected with a virus can infect generative or vegetative relatives. Depending on the virus, this can result in high losses. Training schools and routine diagnostic tools are being developed for a national agricultural certification program that could be established in a common project between German and Colombian universities, the Colombian Agricultural Institute (ICA), the Colombian Corporation of Agricultural Investigation (AGROSAVIA) and the International Center for Tropical Agriculture (CIAT).

An increased attention to the use of virus-free plant material will support Colombian agricultural products domestically and in international export markets (Cutler *et al.*, 2017). A certification of virus-tested plant material can contribute to better quantity and quality in production and contributes to better trade policy decision-making. Many stakeholders in the Colombian supply chain would gain from the knowledge and practical experience in controlling plant pathogens, from small farmers to the major worldwide distributors of Colombia's agricultural and horticultural products.

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