Supplementary material for the article:

Dinić, J.; Novaković, M.; Podolski-Renić, A.; Vajs, V.; Tešević, V.; Isaković, A.; Pešić, M. Structural Differences in Diarylheptanoids Analogues from Alnus Viridis and Alnus Glutinosa Influence Their Activity and Selectivity towards Cancer Cells. *Chemico-Biological Interactions* **2016**, *249*, 36–45. <u>https://doi.org/10.1016/j.cbi.2016.02.019</u> Supporting information

Structural differences in diarylheptanoids analogues from *Alnus viridis* and *Alnus glutinosa* influence their activity and selectivity towards cancer cells

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Spectroscopic data for new compound v4 isolated from A. viridis.

(3*S*)-1,7-di(4-hydroxyphenyl)-heptan-3-*O*-α-L-arabinofuranosyl(1→6)-β-Dglucopyranoside (**v4**): brown amorphous solid; $[\alpha]_{D}^{22}$ -51.0 (c 0.1, MeOH); UV (MeOH) λ_{max} (log ε) 224 (3.83), 278 (3.55) nm; IR (MeOH) ν_{max} 3357, 2929, 1614, 1513, 1450, 1361, 1233, 1076, 1042 cm⁻¹; ¹H-NMR and ¹³C-NMR data, see Table S1; HR-ESI-MS *m*/*z* 593,2617 [M - H]⁻ (calcd. for C₃₀H₄₂O₁₂-H, 593.2604).

Experimental procedure for isolation and structure elucidation. Optical rotations were measured on a Rudolph Research Analytical AUTOPOL IV automatic polarimeter. UV spectra were recorded using GBC Cintra 40 UV/VIS spectrometer. IR spectra were recorded at a ThermoScientific Nicolet 6700 FT-IR spectrometer using capilar film technique. All NMR spectra (¹H, ¹³C, HSQC, HMBC) were recorded on a Bruker Avance III 500 spectrometer at 500.26 for ¹H and 125.80 MHz for ¹³C, with CD₃OD as solvent and TMS as reference. Mass spectral (HR-ESI-MS) data were obtained from Agilent Technologies 6210 time-of-flight LC/MS system. For column chromatography (CC) silicagel 60 (SiO₂; under 0.063 mm, Merck) was used. Analytical TLC was carried out on silicagel 60 GF₂₅₄ 20 × 20 cm plates, layer thickness 0.25 mm (Merck). Semipreparative HPLC was performed on an Agilent 1100 Series instrument equipped with DAD and using a Zorbax Eclipse XDB C-18 column (250 mm × 9.4 mm, 5 µm). All solvents used for column chromatography were freshly distilled and solvents for HPLC analysis were chromatographic grade.

Plant Material. The plant material was collected at Stara planina in south-eastern Serbia in July 2010 and was identified by Professor Petar Marin, Faculty of Biology, University of Belgrade, Serbia. A voucher specimen No. 16681 was deposited at the Herbarium of the Institute of Botany and Botanical Garden 'Jevremovac', Faculty of Biology, University of Belgrade (BEOU), Serbia.

Extraction and isolation. The air-dried bark (150.0 g) was powdered and extracted with CHCl₃/MeOH 1:1 (4 × 1 l, 24 h) at room temperature, with the use of an ultrasonic bath in the last hour of extraction. The yield of the extraction was 22 %. The half of the crude extract (16.5 g) was fractionated by gradient CC (60 cm length and 5 cm diameter column size), starting elution with 100% CH₂Cl₂ and gradually increasing

polarity by addition of MeOH (up to 40% MeOH) to yield 200 fractions of approximately 15 ml. Similar fractions were combined after TLC (carried out with different CH₂Cl₂/MeOH solvent systems) and further fractionated by semipreparative HPLC-DAD into pure compounds using 0.025% HCOOH/MeCN elution system with flow rate of 4 ml/min and gradient program: 0-20 min, 15-25% MeCN; 20-25 min, 25-40% MeCN; 25-28 min, 40-70% MeCN; 28-31 min, 70-100%. The detection wavelength was 280 nm.





Figure S1. Chemical structures of compounds 4v and 7v from *A. viridis*.



Figure S2. Heptane parts of the 13 C NMR spectra (CD₃OD) of 4v (up) and 7v (down).



Figure S3. Aromatic parts of the 13 C NMR spectra (CD₃OD) of 4v (up) and 7v (down).



Figure S4. Heptane parts of the ¹³C NMR spectra (pyridine-d5) of **4v** (up) and **7v** (down) used for comparing with (-)-centrolobol literature data.

С/Ц		4v	7v			
C/H	$\delta_{\rm C}$, tip	$\delta_{\rm H}(J { m u} { m Hz})$	$\delta_{\rm C}$, tip	$\delta_{\rm H} \left(J { m u} { m Hz} ight)$		
1	31,5; CH ₂	2,61 m	31,9; CH ₂	2,60 m		
2	37,6; CH ₂	1,77 m	38,3; CH ₂	1,75 m		
3	80,4; CH	3,67 t (6,0)	80,2; CH	3,66 m		
4	36,0; CH ₂	1,59 m	35,1; CH ₂	1,55 m; 1,61 m		
5	26,0; CH ₂	1,40 m	25,8; CH ₂	1,39 m		
6	33,1; CH ₂	1,59 m	33,2; CH ₂	1,54 m		
7	36,1; CH ₂	2,51 t (7,0)	36,1; CH ₂	2,50 t (7,5)		
1'	135,1; C	-	135,0; C	-		
2'	130,5; CH	7,02 d (8,5)	130,6; CH	7,02 d (8,5)		
3'	116,2; CH	6,68 d (8,5)	116,2; CH	6,69 d (8,5)		
4'	156,4; C	-	156,3; C	-		
5'	116,2; CH	6,68 d (8,5)	116,2; CH	6,69 d (8,5)		
6'	130,5; CH	7,01 d (8,5)	130,6; CH	7,02 d (8,5)		
1"	134,9; C	-	135,0; C	-		
2"	130,5; CH	7,01 d (8,5)	130,4; CH	6,97 d (8,5)		
3"	116,2; CH	6,68 d (8,5)	116,2; CH	6,68 d (8,5)		
4"	156,3; C	-	156,3; C	-		
5"	116,2; CH	6,68 d (8,5)	116,2; CH	6,68 d (8,5)		
6"	130,5; CH	6,99 d (8,5)	130,4; CH	6,97 d (8,5)		
Glcp						
1g	103,9; CH	4,29 d (8,0)	103,7; CH	4,29 d (8,0)		
2g	75,5; CH	3,18 dd (8,0; 1,5)	75,4; CH	3,18 dd (8,0; 1,5)		
3g	78,3; CH	3,34 m	78,2; CH	3,34 m		
4g	72,3; CH	3,34 m	72,1; CH	3,34 m		
5g	76,6; CH	3,36 m	76,6; CH	3,39 m		
6g	68.3: CH ₂	3,61 m	68.2: CH ₂	3,64 dd (11,0; 5,5)		
~0		3,99 m	,	4,03 dd (11,0; 2,0)		
Arabf						
1a	110,0; CH	4,96 d (1,0)	110,0; CH	4,97 d (1,0)		
2a	83,2; CH	4,00 m	83,2; CH	4,01 dd (3,5; 1,5)		
3a	79,1; CH	3,84 m	79,0; CH	3,84 dd (5,5; 3,5)		
4a	86,2; CH	3,96 m	86,1; CH	3,96 m		
5a	63,1; CH ₂	3,58 m 3,72 dd (12,0; 3,5)	63,0; CH ₂	3,59 dd (12,0; 5,5) 3,69 dd (12,0; 3,5)		

Table S1.¹³C and ¹H NMR data (CD₃OD) for **4v** and **7v**.

Glcp – glucopyranosyl group; Arabf – arabinofuranosyl group

	NCI-H460												
	1v	2v	3v	4 v	5v	6v	7v	8v	9v	3g	5g	8g	9g
1v	/	****	****	****	****	****	****	****	****	****	****	****	****
2v	****	/	****	****	****	****	*	****	****	****	****	****	****
3v	****	****	/	****	ns	****	****	ns	ns	ns	ns	ns	ns
4 v	****	****	****	/	****	****	****	****	****	****	****	****	****
5v	****	****	ns	****	/	****	****	*	ns	ns	*	ns	ns
6v	****	****	****	****	****	/	****	****	****	****	****	****	****
7v	****	*	****	****	****	****	/	****	****	****	****	****	****
8v	****	****	ns	****	*	****	****	/	****	ns	ns	ns	ns
9v	****	****	ns	****	ns	****	****	****	/	ns	***	**	ns
3g	****	****	ns	****	ns	****	****	ns	ns	/	ns	ns	ns
5g	****	****	ns	****	*	****	****	ns	***	ns	/	ns	ns
8g	****	****	ns	****	ns	****	****	ns	**	ns	ns	/	ns
9g	****	****	ns	****	ns	****	****	ns	ns	ns	ns	ns	/
						Ha	nCaT						
	1v	2v	3v	4 v	5v	6v	7v	8v	9v	3g	5g	8g	9g
1v	/	ns	****	ns	****	****	****	****	****	****	***	****	****
2v	ns	/	****	ns	****	ns	**	****	****	****	ns	****	****
3v	****	****	/	****	ns	****	****	****	ns	****	****	ns	****
4 v	ns	ns	****	/	****	**	***	****	****	****	ns	****	****
5v	****	****	ns	****	/	****	****	****	ns	***	****	ns	****
6v	****	ns	****	**	****	/	ns	****	****	****	ns	****	****
7v	****	**	****	***	****	ns	/	****	****	****	ns	****	****
8v	****	****	****	****	****	****	****	/	****	ns	****	****	ns
9v	****	****	ns	****	ns	****	****	****	/	***	****	ns	****
3g	****	****	****	****	***	****	****	ns	***	/	****	***	ns
5g	***	ns	****	ns	****	ns	ns	****	****	****	/	****	****
8g	****	****	ns	****	ns	****	****	****	ns	***	****	/	****
9g	****	****	****	****	****	****	****	ns	****	ns	****	****	/

Table S2. Statistical analysis of growth inhibition activity of A. viridis and A. glutinosa diarylheptanoids in NCI-H460 and HaCaT cells.

* p<0.05

** p<0.00 *** p<0.001 **** p<0.0001 ***** p<0.0001

ns = non significant

Table	S3 .	Statistical	analysis	of	cell	death	inducing	activity	of A.	viridis	and A.	glutinosa
diarylh	lepta	noids in N	CI-H460	and	d Ha	.CaT c	ells.					

NCI-H460											
		5v 15 µM	5v 45 µM	5g 15 µM	5g 45 µM	9v 15 μM	9v 45 μM	9g 15 μM	9g 45 μM		
viable cells		ns	****	ns	****	*	****	ns	****		
early apoptosis	untreated	ns	**	ns	ns	ns	ns	ns	***		
late apoptosis		ns	ns	ns	*	ns	****	ns	**		
necrosis		ns	ns	ns	*	ns	ns	ns	*		
HaCaT											
		5v 15 µM	5v 45 µM	5g 15 µM	5g 45 µM	9v 15 μM	9v 45 μM	9g 15 μM	9g 45 μM		
viable cells		***	****	ns	**	****	****	ns	****		
early apoptosis		ns									
late apoptosis	untreated	ns	****	ns	ns	*	****	ns	****		
necrosis		ns	****	ns	ns	ns	*	ns	***		

* p<0.05 ** p<0.01 *** p<0.001 **** p<0.0001 ns = non significant

Table S4.	Statistical	analysis	of cel	cycle	arresting	activity	of A.	viridis	and A.	glutinosa
diarylhepta	anoids in N	CI-H460	and H	aCaT c	ells.					

NCI-H460												
		5v 15 µM	5v 45 µM	5g 15 µM	5g 45 µM	9v 15 μM	9v 45 μM	9g 15 μM	9g 45 μM			
subG ₀		ns	**	ns	ns	*	*	ns	ns			
G ₀ /G ₁	untreated	ns	****	ns	ns	***	****	ns	ns			
S		ns										
G ₂ /M		ns										
HaCaT												
		5v 15 µM	5v 45 µM	5g 15 µM	5g 45 µM	9v 15 μM	9v 45 μM	9g 15 μM	9g 45 μM			
subG ₀		*	*	ns	ns	ns	ns	ns	ns			
G ₀ /G ₁	untreated	ns	**	ns	ns	**	*	ns	**			
S		ns										
G ₂ /M		*	ns									

* p<0.05 ** p<0.01 *** p<0.001 **** p<0.0001

ns = non significant