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#### REVIEW

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# Exploring the efficacy of natural products in alleviating Alzheimer's disease

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### Abstract

Alzheimer's disease (hereafter AD) is a progressive neurodegenerative disorder that affects the central nervous system. There are multiple factors that cause AD, viz., accumulation of extracellular Amyloid-beta 42 plaques, intracellular hyper-phosphorylated Tau tangles, generation of reactive oxygen species due to mitochondrial dysfunction and genetic mutations. The plaques and tau tangles trigger aberrant signaling, which eventually cause cell death of the neurons. As a result, there is shrinkage of brain, cognitive defects, behavioral and psychological problems. To date, there is no direct cure for AD. Thus, scientists have been testing various strategies like screening for the small inhibitor molecule library or natural products that may block or prevent onset of AD. Historically, natural products have been used in many cultures for the treatment of various diseases. The research on natural products have gained importance as the active compounds extracted from them have medicinal values with reduced side effects, and they are bioavailable. The natural products may target the proteins or members of signaling pathways that get altered in specific diseases. Many natural products are being tested in various animal model systems for their role as a potential therapeutic target for AD, and to address questions about how these natural products can rescue AD or other neurodegenerative disorders. Some of these products are in clinical trials and results are promising because of their neuroprotective, anti-inflammatory, antioxidant, anti-amyloidogenic, anticholinesterase activities and easy availability. This review summarizes the use of animal model systems to identify natural products, which may serve as potential therapeutic targets for AD.

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# Full Text

#### Introduction

In 1906, Dr. Alois Alzheimer, first described shrinkage of the brain in the autopsy of the patient who suffered from increasing short-term memory loss, paranoia and deteriorating cognitive abilities. In 1968, the damaged brain tissue in elderly subjects was measured using the newly developed cognitive measurement scales (Tomlinson et al., 1968). Alzheimer's disease (AD), a senile dementia, is an age-related neurodegenerative disorder. Based on the time and cause of onset of AD, epidemiologists have clinically divided AD into: the early onset before 65 years, late-onset group consist of patients with the age 65 or above and familial AD (Rajan et al., 2018). The prevalence of AD in 2017 is 44 million worldwide. It is estimated to be around 75 million worldwide in 2030 (Selkoe and Hardy, 2016). AD is the sixth leading cause of mortality. It is estimated that AD will be a global epidemic by 2050, when the number of individuals affected by AD will soar to 135 million (Alzheimer's Association, 2018). AD manifests in the patients as declining cognitive and mental abilities due to the loss of neurons in the hippocampus and the cortex (Hardy, 2009; Sarkar et al., 2016). Neurons communicate with each other via synapses, which makes it plausible to recall the memories. The loss of synapses in AD results in loss of learning and memory. Clinically, AD patients face difficulty in short term memory, word finding, and language, which results in memory loss and slow progression of cognitive impairment (Albert et al., 2011).

AD is caused by multiple mechanisms including excessive accumulation of extracellular amyloid-beta 42 (AB42) plaques, intracellular hyper-phosphorylated tau neurofibrillary tangles in the brain, oxidative stress due to mitochondrial dysfunction, and/ or genetic as well as the environmental factors [Figure 1] and [Figure 2] (Selkoe and Podlisny, 2002; Hardy, 2009; Sarkar et al., 2016; Selkoe and Hardy, 2016; Sierra-Fonseca and Gosselink, 2018). Genetically the mutation in apolipoprotein E, which regulates the lipid homeostasis and carries the cholesterol to neurons, is a risk factor for the late-onset of AD. Oxidative stress results from generation of reactive oxygen species (ROS) (Chen et al., 2012; Sarkar et al., 2016). The Aβ42 plaques can form due to inappropriate cleavage of Amyloid precursor protein (APP), a transmembrane glycoprotein, encoded by the APP gene on chromosome 21. Generally, APP is cleaved proteolytically by  $\alpha$ -secretase and  $\gamma$ -secretase to form 40 amino acid long peptide, however, when miscleaved by  $\beta$ -secretase and  $\gamma$ -secretase results in A $\beta$ 42 polypeptides [Figure 3]A (O'Brien and Wong, 2010; Selkoe and Hardy, 2016). These misfolded Aβ42 polypeptides accumulate to form amyloid plaques, which triggers neurodegeneration by activating aberrant signaling [Figure 3]B (Shankar et al., 2008; O'Brien and Wong, 2010; Fernandez-Funez et al., 2013; Sarkar et al., 2016; Selkoe and Hardy, 2016). Hyper-phosphorylation of tau, a protein involved in the stability of microtubules, decreases tau's association to microtubules, results in accumulation of toxic insoluble intracellular neurofibrillary tangles [Figure 1] (Gendron and Petrucelli, 2009; Sierra-Fonseca and Gosselink, 2018). The existing clinical regimen for AD offer transient symptomatic improvement but offer little or no benefit in terms of changing the overall course of disease. Lately, there have been disappointing results from AD drug trials (Goldman et al., 2018). These widespread AD drug trial failures are due to lack of diversity in novel targets and a lack of reliable and effective preclinical models. It is imperative that there is a need for a more robust, diversified drug development pipeline and screening animal models to find cures for AD.{Figure 1}{Figure 2}{Figure 3}

#### **Animal Models**

The genetic machinery is highly conserved across the species. As a result, several animal models for AD have been developed to understand the mechanism of disease progression, and to test the potential targets of AD (Pandey and Nichols, 2011; Fernandez-Funez et al., 2013; Saito and Saido, 2014; Sarkar et al., 2016). Among vertebrates, the mouse model has been used extensively since they have similarities with the human brain (Di Carlo, 2012; Sasaguri et al., 2017). Both loss-of-function and gain-of-function mouse models have been generated using the genes responsible for AD. Early AD mouse model was generated where mutant APP was expressed that progressively manifested AD neuropathology (Games et al., 1995). Later, a combination type model system including Tau and Psen1 was generated in APP mouse background (Oddo et al., 2003). It was modified further where human sequence was introduced into the APP mouse to better understand AD pathology in humans (Saito et al., 2014). These model systems helped in discerning the learning and memory defects associated with AD by using behavioral assays. One of the major challenges was lack of concordance between preclinical models and human clinical trials (LaFerla and Green, 2012). As a result the drugs, which successfully inhibited the amyloid plaques in mice, failed to cure AD in humans (King, 2018). Majority of the data generated in mouse was from young animals, when their immune system is robust unlike the old ones. Thus, mouse models reflects early stages of AD. The reason clinical trial are failing because drugs that target AB42 in mouse model most probably treat early plaque build-up process rather than late, which is unlike the stage when AD is detected in humans. Thus, those drugs might fit prevention than treatment paradigm (King, 2018).

The other common animal models to study underlying mechanisms of AD are Drosophila melanogaster (fruit fly), Caenorhabditis elegans (worm), Danio rerio (zebrafish), and sea urchins (lijima and lijima-Ando, 2008; lijima-Ando and lijima, 2010; Pandey and Nichols, 2011; Singh, 2012; Singh and Irvine, 2012; Fernandez-Funez et al., 2013; Prussing et al., 2013; Saito and Saido, 2014; Sarkar et al., 2016). Caenorhabditis elegans, a nematode, has short life-span, and has been established as a simple genetic model system (Di Carlo, 2012; LaFerla and Green, 2012). Transgenic lines were generated in nematode to study  $A\beta$  toxicity by using learning and behavior assays (Steinkraus et al., 2008). Zebrafish is a good genetic model system due to its optical clarity during developmental process of embryogenesis (Di Carlo, 2012), and genome is sequenced. This zebrafish possess genes orthologous to those mutated in familial AD, which makes it a useful model system to study AD (Newman et al., 2014).

Drosophila melanogaster, an invertebrate, has developed into an excellent model for human disease. The entire genome of Drosophila is sequenced, and the genetic machinery is highly conserved between flies and humans (Lenz et al., 2013). The fly genome has less genetic redundancy as compared to the humans (Bier, 2005; Lenz et al., 2013). Nearly 70% of fly genes are closely related to the human disease genes (Bier, 2005; Singh and Irvine, 2012). Therefore, using fruit fly to model human disease is practically beneficial. In addition, they are smaller in size and thus can be stored efficiently, and are cost effective. Drosophila can produce two generations in a month as the life cycle is short, and has a repertoire of genetic tools (Sarkar et al., 2016). In addition, the transgenic system is well established, which allows introduction of genes from other organism in the flies. It makes Drosophila an ideal model for studying age related progressive diseases like AD. Several models of AD have been developed in Drosophila. These include transgenic flies misexpressing human A $\beta$ 42, human A $\beta$ 40, human APP,  $\beta$ -site APP-cleaving enzyme,  $\beta$ -secretase and Presenilin, and Familial AD (Cauchi and van den Heuvel, 2006; Cao et al., 2008; lijima and lijima-Ando, 2008; Tare et al., 2011; Fernandez-Funez et al., 2013). These flies display several aspects of clinical AD neuropathology and symptoms including the generation of amyloid aggregates, external morphological abnormalities, dramatic neuroanatomical changes and defects in motor reflex behavior and memory. As a proof of concept, treatment with an inhibitor of y-secretase can suppress these neurodegenerative phenotypes. Drosophila has also been used to model other neurodegenerative diseases like Parkinson's disease, Huntington's disease, fronto-temporal lobar degeneration, and amyotrophic lateral sclerosis (Bier, 2005; Pandey and Nichols, 2011; Singh and Irvine, 2012; Fernandez-Funez et al., 2013).

Lately, Drosophila eye has also been extensively used for modeling human disease (Singh and Irvine, 2012; Moran et al., 2013; Steffensmeier et al., 2013). Drosophila has a fully functional nervous system where function of vision, olfaction, learning and memory can be separated. Furthermore, the genes involved in eye development are

structurally and functionally similar between flies and humans. The adult compound eye of Drosophila [Figure 4]A, develops from a mono-layer epithelium housed in the larva and referred to as the eye-antennal imaginal disc (Ready et al., 1976; Singh and Choi, 2003; Singh et al., 2005a, 2012; Kumar, 2011; Tare et al., 2013a). The larval eye imaginal disc metamorphose into a highly organized compound eye comprised of photoreceptor cells, cone cells, and pigment cells (Ready et al., 1976; Kumar, 2011, 2013; Singh et al., 2012; Tare et al., 2013a). The precise nature of the structure of Drosophila eye makes it highly sensitive to genetic manipulations that can be easily screened using a stereo microscope (Tare et al., 2013b; Singh et al., 2005b), which allows quick screening of large sample size (Singh and Choi, 2003; Singh et al., 2005a, b, 2006; Tare et al., 2013a). The Drosophila eye allows direct visualization of the cellular and developmental defects. Therefore, Drosophila eye can be used to mimic many neurodegenerative disorders including AD (Bier, 2005; Tare et al., 2011; Singh and Irvine, 2012; Moran et al., 2013; Cutler et al., 2015; Sarkar et al., 2016). Since it is easy to visualize the phenotypes in Drosophila eyes caused by neurodegeneration (Tare et al., 2013a), the Drosophila eye model can be used to study the different signaling pathways causing AD or other neurodegenerative disorders, and to screen for the therapeutic targets.{Figure 4}

The Drosophila model has a large repertoire of tools that allows expression of foreign genes along spatial and temporal axes. Using Gal4/UAS (upstream activating sequence) transgenic systems (Brand and Perrimon, 1993), human Aβ42 was misexpressed in the differentiating photoreceptor neurons of the developing eye (Tare et al., 2011; Moran et al., 2013; Steffensmeier et al., 2013). It resulted in accumulation of extracellular amyloid plaques, as evidenced by immuno-histochemical approaches. These human Aβ42 expressing eyes exhibited progressive neurodegenerative phenotype in the Drosophila eye [Figure 4]B as compared to the wild-type adult compound eye [Figure 4]A (Tare et al., 2011; Sarkar et al., 2016). This Aβ42 mediated neurodegeneration phenocopies AD like neuropathology in terms of progression of phenotype [Figure 4]A & [Figure 4]B. Thus, Drosophila eye model of Aβ42 mediated neurodegeneration can be exploited for genome wide genetic screens or for screening chemical libraries for potential therapeutic targets.

#### **Current Therapeutic Targets for AD**

Presently, Food and Drug Administration approved drug regimen for AD help to treat the symptoms but do not cure the disease itself. The anti-inflammatory drugs (Aisen, 2000), like atorvastatin (Sparks et al., 2005), aspirin (AD2000 Collaborative Group et al., 2008), and rosiglitazone (Harrington et al., 2011) have not shown the complete cure in improving the cognitive decline in AD patients. The AD patients have less acetylcholine neurotransmitter in the brain that results in loss of neurons. Therefore, cholinesterase inhibitors have been approved by Food and Drug Administration for AD treatment regimen (Gomes et al., 2018). But these drugs have side-effects like nausea, vomiting, diarrhea, muscle cramps, fatigue indigestion, weight loss, confusion and headache, and do not sure AD (Colovic et al., 2013). Hence, there is a need of medication, which has less side effects and can treat the disease by preventing the neuronal cell death. The available medicines for AD ameliorate the symptoms but cannot cure the disease altogether. Lately, there have been disappointing results from AD drug trials due to widespread drug failures and lack of diversity of novel targets (Goldman et al., 2018). Efforts are directed towards search for alternative treatment approaches, and among them screening for natural product library for potential therapeutic targets is on the forefront.

#### Natural Products: Potential Targets for AD

The traditional herbal medicines that are compounds extracted from natural plant products can be less toxic, may have less side-effects and inexpensive than the synthetic drugs. Some plant products have medicinal properties for neurodegenerative diseases like anticholinesterase activities, anti-inflammatory, antioxidant, and neuroprotective function. Natural products like Lunasin, Polyphenols, Flavonoids, Alkaloids, and Tannins are potential therapeutic

candidates for the AD (Zhang et al., 2008; Williams et al., 2011; Pany et al., 2014; Sarkar et al., 2018). Here we discuss some of the natural products that have demonstrated a potential in preventing neurodegeneration observed in AD in cell lines or in animal model systems.

#### Lunasin

Lunasin was first extracted in 1987 in Japan from soybean (Glycine max) seed, a staple in the diets of many cultures, while screening for the protease inhibitors. It is also found in other plants like wheat, barley, rye, amaranth and triticale. Lunasin, a thermostable protein, which should be ingested at optimum amount (Liu et al., 2014). Lunasin has antioxidant, anti-inflammatory and anti-cancer properties (Jones and Srivastava, 2014). Lunasin is the smaller subunit of the soybean 2S albumin protein [Figure 4]D. The gene GM2S-1 encodes methionine rich protein, signal peptide, a linker peptide and Lunasin which are post-translationally processed (Jones and Srivastava, 2014). It has 43 amino acid residues, a molecular weight of 5.5 kDa, and eight negatively charged aspartate residue sequence at the carboxyl end. Lunasin has four motifs: N-terminal domain, predicted helical region, arginine-glycine-aspartate (RGD) domain and poly-D tail [Figure 4]E. The function of N-terminal domain of Lunasin is unknown. The predicted helical domain binds to the chromatin. The poly D sequence acts as H3-H4 histone acetylation inhibitor. The amino acids preceded the poly D stretch are RGD, which helps in internalization of Lunasin into nucleus and in binding of Lunasin to the core histone proteins (Liu et al., 2014). The function of other peptides in Lunasin has not been defined.

Lunasin can inhibit mitosis by increasing the expression of cyclin dependent kinases inhibitors like p21 and p27 [Figure 5] (Dia and Gonzalez de Mejia, 2011; Jones and Srivastava, 2014). The cyclin dependent kinases play an important role in cell cycle and their misexpression results in cell cycle deregulation. Tumor suppressor genes regulate the cell cycle progression and are often downregulated in cancers. PTEN, a Tensin homolog and a tumor suppressor phosphatase, is mutated in cancer. Lunasin is found to increase the levels of PTEN in human breast cancer cells (MCF-7) inducing the apoptosis [Figure 5] (Pabona et al., 2012). Lunasin represses the expression of genes, which are involved in tumor suppression (Hsieh et al., 2017). Animal studies suggests that Lunasin has anti-inflammatory effects [Table 1] and [Figure 5] by inhibiting cytokines such as tumor necrosis factor  $\alpha$  and interleukin-6. It also inhibits the release of nitric oxide (Liu and Pan, 2010). Hence, Lunasin may be potential therapeutic target for the disease where inflammation is observed.{Figure 5}{Table 1}

Drosophila eye model system has been used to study the effects of Lunasin in transgenic flies expressing high levels of human Aβ42 in differentiating retinal neurons of the eye (Sarkar et al., 2018). Targeted misexpression of Aβ42 in the developing retinal neurons of the flies showed glazed eye appearance and necrotic spots suggesting neurodegeneration (Sarkar et al., 2018). This study suggests that misexpression of Lunasin in the differentiating retinal neurons of the developing Drosophila eye can rescue Aβ42 mediated neurodegeneration [Figure 2] (Sarkar et al., 2018). Previously, it has been shown that activation of evolutionarily conserved Jun-N terminal kinase signaling pathway is involved in AD. Sarkar et al. (2018) reported that Lunasin downregulates Jun-N terminal kinase signaling to rescue Aβ42 mediated neurodegeneration. AD a neurodegenerative disorder exhibits neuro-inflammation due to production of the ROS [Figure 5]. Lunasin can be used as a potential therapeutic target for AD and further studies on vertebrate models will be helpful to discern its medical properties on mammals.

#### Polyphenols

Polyphenols derived from different plants are part of our daily diet, and have been reported to improve cognitive activities and reduce the neuropathology in animal models of AD (Wang et al., 2014). They have antioxidant property, which helps in scavenging ROS (Malar and Devi, 2014). Polyphenols can prevent formation of neurofibrillary tangles by preventing tau hyper-phosphorylation, which makes it potential target for AD [Figure 2] and [Table 1].

Curcumin, a polyphenol, extracted from Curcuma longa and has been used in Ayurveda medicines. Curcumin has been shown to inhibit the generation of Aβ42 oligomers and the plaques [Figure 2] and [Table 1] (Williams et al., 2011). Administration of curcumin in AD animal model prevented neuro-inflammation, tau hyper-phosphorylation and protein kinase B (Akt)/glycogen synthase kinase 3 signaling pathway (Hoppe et al., 2013). It also helps in decreasing

ROS by regulating pathways mediated neurodegeneration [Figure 2] (Doggui et al., 2013).

Resveratrol, a polyphenolic compound, has found to play role in activation of SIRT1, SIRT2 and SIRT3 (Sirtuins), which plays role in neuronal cell survival and longevity (Bastianetto et al., 2000; Rahman et al., 2006; Schirmer et al., 2012). As a result, Sirtuins reduce the  $A\beta$  plaque aggregation in transgenic model of AD [Table 1] (Karuppagounder et al., 2009). Hyper-activation of Microglia triggers neuronal death. Resveratrol exhibits antioxidant effects in microglial cell line in hypoxia injury model. Thus, resveratrol may exhibit neuroprotective function by regulating microglia in brain (Song et al., 2014). Polyphenols have also been reported to lessen the mitochondrial dysfunction thereby preventing onset or delaying progression of neurodegeneration.

Combination of polyphenol treatment has been found to be the novel treatment target for AD (Auti and Kulkarni, 2018). A combination of three polyphenols like grape seed extract, resveratrol and Concord grape juice extract have been tested in mouse model of AD. They found that combination treatment proved to be better than the individual treatment in reducing amyloid content in the brain and thereby improving the cognitive impairment in AD mouse model (Wang et al., 2014).

#### Flavonoids

Flavonoids are plant secondary metabolites, which are anti-oxidant, anti-amyloidogenic, anti-inflammatory and modulate cell-signaling pathways (Baptista et al., 2014; Bakhtiari et al., 2017). Approximately 128 flavonoids have been reported. They have acetylcholinesterase inhibitory activity proving to be a promising drug for AD (Auti and Kulkarni, 2018). Moreover, these phytochemicals can cross the blood-brain barrier, which has direct effect on the brain (de Andrade Teles et al., 2018). Flavonoids affect phosphatidylinositol 3-kinase/Akt pathway and mitogen activated protein kinase signaling pathway, which regulate expression of transcription factors involved in survival (Bakhtiari et al., 2017). Flavonoids reduce the toxic A $\beta$  aggregation by inhibiting  $\beta$ -secretase and activate the- secretase activity [Figure 2] (Baptista et al., 2014). Additionally, it also prevents the formation of tau hyper-phosphorylation (Baptista et al., 2014). Another flavonoid, Quercitin (3,3',4',5,7-pentahydroxyflavone), can decrease the levels of ROS and inhibits apoptosis in young rats [Figure 2] (de Andrade Teles et al., 2018). Another study shows that quercitin blocks nuclear factor kB levels and hence reduces the inflammation (Pany et al., 2014). Catechin, derived from catechu, is a plant secondary metabolite. Catechins prevent A $\beta$ 42 accumulation and oxidative stress [Figure 2] and [Table 1] (Rezai-Zadeh et al., 2005; Pervin et al., 2018). Kaempferol, a natural flavonoid, can be derived from variety of plants. Effect of Kaempferol on transgenic AD flies was found to decrease the oxidative stress and acetylcholinesterase activity [Table 1] (Beg et al., 2018).

#### Alkaloids

Alkaloids are organic compounds that contain basic nitrogen atoms. Most of them are acetyl-cholinesterase and butylcholinesterase inhibitors (Konrath et al., 2013). Huperzine A, isolated from Huperzia serrate, has been used for memory enhancement in ancient Chinese medicine. Huperzine A, a specific and selective acetylcholinesterase inhibitor, showed increase in acetylcholine in rat brain (Zangara, 2003). It is also known to reduce Aβ aggregates in brain (Zhang et al., 2008). Additionally, Huperzine A, an N-methyl-D-aspartic acid receptor antagonist, reduces glutamate neurotoxicity. Hence, it reduces the synaptic loss and cell death of neurons [Table 1] (Auti and Kulkarni, 2018).

Nicotine, 3-(1-methyl-2-pyrrolidinyl) pyridine, is obtained from Nicotiana tobacum. Nicotine patches as well as injections have reported to improve the learning abilities in animals as well as AD patients. Since nicotine has adverse effects, it is important to assess the efficacy in AD patients [Table 1] (Levin and Simon, 1998; Nordberg et al., 2002; Newhouse et al., 2012; Auti and Kulkarni, 2018). Berberine, isoquinoline, have been used in Chinese and Ayurvedic medicine regimens. It is anti-inflammatory, anti-oxidant, anti-cancer, anti-depressants, and prevents  $A\beta 42$  plaque formation [Figure 2], [Table 1] (Durairajan et al., 2012; Cai et al., 2016).

Terpenes

Terpenes, aromatic molecules, are group of hydrocarbons that consist of repeats of isoprene units. Terpenes inhibit acetylcholinesterase activity and block the advanced glycation end product formation (Auti and Kulkarni, 2018). Triterpenes have anti-oxidative and anti-inflammatory activities. Gensenoside, which promotes health and longevity, have been studied for their effects on AD. Gensenoside prevents the A $\beta$ 42 levels by increasing Neprilysin gene expression [Figure 2] and [Table 1] (Yang et al., 2009). Moreover, Ginseng improves learning ability in aging brains of rodents.

Ginkgolides, cyclic diterpenes, isolated from Ginkgo biloba. It has been shown that it promotes cell survival of neurons from synaptic damage by Synaptophysin, a presynaptic marker (Bate et al., 2008). It has been reported that Ginkgolide B can rescue  $A\beta$  induced apoptosis by promoting the synthesis of neurotrophic factor and reduces the cell death of neurons in the AD rats brains [Figure 2] (Serruys et al., 2011).

A triterpenoid, Platycodin D, which is present in the root extract of Platycodon Grandiflorus (balloon flower or Chinese bellflower) inhibits the Aβ42 plaques mediated neuronal loss [Figure 2]. Using rat PC12 cell lines (derived from phenochromocytoma of the rat adrenal medulla cells), it was shown that Platycodon Grandiflorus extract from root contains Platycodin D, can increase synaptogenesis in the hippocampus by activating mitogen activated protein kinase/extracellular regulated protein kinase pathway. Platycodon Grandiflorus extract from root can be studied further on AD models to test its therapeutic potential efficiency (Kim et al., 2017).

#### Future of Natural Products As a Therapeutic Medicine

The use of medicinal plants as source of natural products to cure disease is a result of man experimenting by trial and error for hundreds of centuries, searching for available foods for the treatment of diseases. The natural products have been used as medicines throughout history in the form of traditional medicines, remedies, potions and oils. Before the discovery of chemical based drug formulations, majority of drug regimens were natural product based across the civilizations. The earliest records dates as early as Mesopotamia (2600 B.C.) where oils from Cupressus semperviviens (Cypress) and Commiphos species (myrrh) were used to treat cough and colds. These treatments are still in use today. In India, Curcurmin or Azadirachta indica (Neem), a member of Meliaceae family that has rich source of antioxidants has been extensively used in Ayurveda medicine. These natural products have been extensively used in Indian, Chinese and Unani medicine because of their property of scavenging ROS, and their pivotal role in anticancer management and other diseases. It has been shown to modulate various growth regulatory pathways including p53, phosphate and tension homology deleted on chromsome ten (pTEN), phosphatidylinositol 3-kinase/Akt, etc. (Alzohairy, 2016).

It is evident that discovery of natural products will be beneficial on multiple levels. Many of these bioactive natural products are still unidentified. Natural plant products are available in everyday diet and they are the part of the traditional medicines in Indian Ayurveda and Chinese medicines. The fruits, vegetables and grains prevent the onset or delay the onset of the disease and promote healthy aging. These natural products also reduce the risk of AD (Howes et al., 2003). The bioactive compounds with pharmacological effects have been extensively studied to cure various neurological diseases. These phytochemicals have better pharmacological effects and can cross the blood-brain barrier and high bioavailability. They are safe and effective as compared to other available drugs in the market. As a result, they might be considered to be an effective alternative to the current drugs.

#### Conclusions

The natural phytochemicals with pharmacological effects can serve as potential therapeutic treatments for AD. These

natural products may reduce ROS, or prevent the toxic A $\beta$ 42 plaque production. Alternatively, these products may act downstream of A $\beta$ 42 plaque accumulation and modulate the aberrant signaling events that are triggered due to A $\beta$ 42 plaques in AD. Studies in animal model system strongly suggest that some of these natural products can cure and prevent AD effectively [Figure 2] and [Table 1].

As the age expectancy will increase due to advancement in medical science, the prevalence of the neurodegenerative diseases will increase in aging population. It has been seen that changes in lifestyle, change in food habits, excessive stress due to work or environmental conditions have aided to increase in frequency of neurodegenerative disease like AD. The management or finding cures for AD is an issue to target multiple causative agents of the disease (Goldman et al., 2018). Understanding the molecular mechanism of the disease by using animal model system in vivo studies would be helpful in identifying the potential therapeutic targets. Various animal models are being used to study the mechanism of action of these products at molecular level. Scientists have turned to natural products for drug development for neurodegenerative diseases like AD, because of their medicinal properties. The phytochemicals seem to be promising and potential therapeutic targets for neurodegenerative diseases like AD. The strength of usage of natural products lie in their bioavailability. Thus, following a strict diet regimen with emphasis on natural products with potential medicinal properties have promising expectation for finding cures for AD.

Interestingly, in humans by the time AD is detected, the disease has progressed significantly. At this stage, a critical number of neurons are already lost, which manifests as the loss of cognitive functions in the patient. Since many of the neurons are post mitotic, they cannot be replaced by normal cell division. Therefore, efforts are being directed towards approaches to recover and regenerate the damaged or lost neurons to restore the lost cognitive functions (Felsenstein et al., 2014). Blocking A $\beta$ 42 mediated neurodegeneration at the time when AD is detected, will only prevent the further loss of neurons after that stage. However, it will not recover dead neurons and as a result, there will be no recovery of cognitive functions lost due to neurodegeneration. Thus, there is a need to utilize these established animal models to screen for potential therapeutic targets which can generate regeneration response in the neuronal population of AD patients.[96]

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## References

- 1 AD2000 Collaborative Group, Bentham P, Gray R, Sellwood E, Hills R, Crome P, Raftery J (2008) Aspirin in Alzheimer's disease (AD2000): a randomised open-label trial. Lancet Neurol 7:41-49.
- 2 Aisen PS (2000) Anti-inflammatory therapy for Alzheimer's disease: implications of the prednisone trial. Acta Neurol Scand Suppl 176:85-89.
- 3 Albert MS, DeKosky ST, Dickson D, Dubois B, Feldman HH, Fox NC, Gamst A, Holtzman DM, Jagust WJ, Petersen RC, Snyder PJ, Carrillo MC, Thies B, Phelps CH (2011) The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement 7:270-279.
- 4 Alzheimer's Association (2018) Alzheimer's disease facts and figures. Alzheimers Dement 14:367-429.
- 5 Alzohairy MA (2016) Therapeutics role of Azadirachta indica (Neem) and their active constituents in diseases prevention and treatment. Evid Based Complement Alternat Med 2016:7382506.
- 6 Auti ST, Kulkarni YA (2018) A systematic review on the role of natural products in modulating the pathways in Alzheimer's disease. Int J Vitam Nutr Res doi: 10.1024/0300-9831/a000405.
- 7 Bakhtiari M, Panahi Y, Ameli J, Darvishi B (2017) Protective effects of flavonoids against Alzheimer's diseaserelated neural dysfunctions. Biomed Pharmacother 93:218-229.
- 8 Baptista FI, Henriques AG, Silva AM, Wiltfang J, da Cruz e Silva OA (2014) Flavonoids as therapeutic compounds targeting key proteins involved in Alzheimer's disease. ACS Chem Neurosci 5:83-92.
- 9 Bastianetto S, Zheng WH, Quirion R (2000) Neuroprotective abilities of resveratrol and other red wine constituents against nitric oxide-related toxicity in cultured hippocampal neurons. Br J Pharmacol 131:711-720.
- 10 Bate C, Tayebi M, Williams A (2008) Ginkgolides protect against amyloid-beta1-42-mediated synapse damage in vitro. Mol Neurodegener 3:1.
- 11 Beg T, Jyoti S, Naz F, Rahul, Ali F, Ali SK, Reyad AM, Siddique YH (2018) Protective effect of kaempferol on the transgenic Drosophila model of Alzheimer's disease. CNS Neurol Disord Drug Targets 17:421-429.
- 12 Bier E (2005) Drosophila, the golden bug, emerges as a tool for human genetics. Nat Rev Genet 6:9-23.
- 13 Brand AH, Perrimon N (1993) Targeted gene expression as a means of altering cell fates and generating dominant phenotypes. Development 118:401-415.
- 14 Cai Z, Wang C, Yang W (2016) Role of berberine in Alzheimer's disease. Neuropsychiatr Dis Treat 12:2509-2520.
- 15 Cao W, Song HJ, Gangi T, Kelkar A, Antani I, Garza D, Konsolaki M (2008) Identification of novel genes that modify phenotypes induced by Alzheimer's beta-amyloid overexpression in Drosophila. Genetics 178:1457-1471.
- 16 Cauchi RJ, van den Heuvel M (2006) The fly as a model for neurodegenerative diseases: is it worth the jump? Neurodegener Dis 3:338-356.
- 17 Chen XP, Guo CY, Kong JM (2012) Oxidative stress in neurodegenerative diseases. Neural Regen Res 7:376-385.
- 18 Colovic MB, Krstic DZ, Lazarevic-PaÅ<sub>i</sub>ti TD, Bondzic AM, Vasic VM (2013) Acetylcholinesterase inhibitors: pharmacology and toxicology. Curr Neuropharmacol 11:315-335.
- 19 Cutler T, Sarkar A, Moran M, Steffensmeier A, Puli OR, Mancini G, Tare M, Gogia N, Singh A (2015) Drosophila eye model to study neuroprotective role of CREB binding protein (CBP) in Alzheimer's disease. PLoS One 10:e0137691.
- 20 de Andrade Teles RB, Diniz TC, Costa Pinto TC, de Oliveira Junior RG, Gama ESM, de Lavor EM, Fernandes AWC, de Oliveira AP, de Almeida Ribeiro FPR, da Silva AAM, Cavalcante TCF, Quintans Junior LJ, da Silva Almeida JRG (2018) Flavonoids as therapeutic agents in Alzheimer's and Parkinson's diseases: a systematic review of preclinical evidences. Oxid Med Cell Longev 2018:7043213.
- 21 Di Carlo M (2012) Simple model systems: a challenge for Alzheimer's disease. Immun Ageing 9:3.

22

Dia VP, Gonzalez de Mejia E (2011) Lunasin induces apoptosis and modifies the expression of genes associated with extracellular matrix and cell adhesion in human metastatic colon cancer cells. Mol Nutr Food Res 55:623-634.

- 23 Doggui S, Belkacemi A, Paka GD, Perrotte M, Pi R, Ramassamy C (2013) Curcumin protects neuronal-like cells against acrolein by restoring Akt and redox signaling pathways. Mol Nutr Food Res 57:1660-1670.
- 24 Durairajan SS, Liu LF, Lu JH, Chen LL, Yuan Q, Chung SK, Huang L, Li XS, Huang JD, Li M (2012) Berberine ameliorates beta-amyloid pathology, gliosis, and cognitive impairment in an Alzheimer's disease transgenic mouse model. Neurobiol Aging 33:2903-2919.
- 25 Felsenstein KM, Candelario KM, Steindler DA, Borchelt DR (2014) Regenerative medicine in Alzheimer's disease. Transl Res 163:432-438.
- 26 Fernandez-Funez P, Sanchez-Garcia J, Ribncon-Limas DE (2013) Unraveling the Basis of Neurodegeneration using the Developing Eye. New York: Springer.
- 27 Games D, Adams D, Alessandrini R, Barbour R, Berthelette P, Blackwell C, Carr T, Clemens J, Donaldson T, Gillespie F, et al. (1995) Alzheimer-type neuropathology in transgenic mice overexpressing V717F beta-amyloid precursor protein. Nature 373:523-527.
- 28 Gendron TF, Petrucelli L (2009) The role of tau in neurodegeneration. Mol Neurodegener 4:13.
- 29 Goldman DP, Fillit H, Neumann P (2018) Accelerating Alzheimer's disease drug innovations from the research pipeline to patients. Alzheimers Dement 14:833-836.
- 30 Gomes RC, Sakata RP, Almeida WP, Coelho F (2018) Spirocyclohexadienones as an Uncommon Scaffold for Acetylcholinesterase Inhibitory Activity. Med Chem doi: 10.2174/1573406414666181109114214.
- 31 Hardy J (2009) The amyloid hypothesis for Alzheimer's disease: a critical reappraisal. J Neurochem 110:1129-1134.
- 32 Harrington C, Sawchak S, Chiang C, Davies J, Donovan C, Saunders AM, Irizarry M, Jeter B, Zvartau-Hind M, van Dyck CH, Gold M (2011) Rosiglitazone does not improve cognition or global function when used as adjunctive therapy to AChE inhibitors in mild-to-moderate Alzheimer's disease: two phase 3 studies. Curr Alzheimer Res 8:592-606.
- 33 Hoppe JB, Coradini K, Frozza RL, Oliveira CM, Meneghetti AB, Bernardi A, Pires ES, Beck RC, Salbego CG (2013) Free and nanoencapsulated curcumin suppress beta-amyloid-induced cognitive impairments in rats: involvement of BDNF and Akt/GSK-3beta signaling pathway. Neurobiol Learn Mem 106:134-144.
- 34 Howes MJ, Perry NS, Houghton PJ (2003) Plants with traditional uses and activities, relevant to the management of Alzheimer's disease and other cognitive disorders. Phytother Res 17:1-18.
- 35 Hsieh CC, Martinez-Villaluenga C, de Lumen BO, Hernandez-Ledesma B (2017) Updating the research on the chemopreventive and therapeutic role of the peptide lunasin. J Sci Food Agric 98:2070-2079.
- 36 lijima-Ando K, lijima K (2010) Transgenic Drosophila models of Alzheimer's disease and tauopathies. Brain Struct Funct 214:245-262.
- 37 lijima K, lijima-Ando K (2008) Drosophila models of Alzheimer's amyloidosis: the challenge of dissecting the complex mechanisms of toxicity of amyloid-beta 42. J Alzheimers Dis 15:523-540.
- 38 Jones G, Srivastava A (2014) Understanding Lunasin's biology and potential as a cancer therapeutic by utilizing Drosophila genetics. Exp Biol Med (Maywood) 239:519-528.
- 39 Karuppagounder SS, Pinto JT, Xu H, Chen HL, Beal MF, Gibson GE (2009) Dietary supplementation with resveratrol reduces plaque pathology in a transgenic model of Alzheimer's disease. Neurochem Int 54:111-118.
- 40 Kim JI, Jeon SG, Kim KA, Kim JJ, Song EJ, Jeon Y, Kim E, Lee KB, Kwak JH, Moon M (2017) Platycodon grandiflorus root extract improves learning and memory by enhancing synaptogenesis in mice hippocampus. Nutrients 9:E794.
- 41 King A (2018) The search for better animal models of Alzheimer's disease. Nature 559:S13-S15.
- 42 Konrath EL, Passos Cdos S, Klein LC, Jr., Henriques AT (2013) Alkaloids as a source of potential anticholinesterase inhibitors for the treatment of Alzheimer's disease. J Pharm Pharmacol 65:1701-1725.
- 43 Kumar J (2013) Catching the next wave: patterning of the Drosophila eye by the morphogenetic furrow. In: Molecular Genetics of Axial Patterning, Growth and Disease in the Drosophila Eye (Singh A and Kango-Singh M, eds), pp 75-97. New York, USA: Springer.

- 44 Kumar JP (2011) My what big eyes you have: how the Drosophila retina grows. Dev Neurobiol 71:1133-1152.
- 45 LaFerla FM, Green KN (2012) Animal models of Alzheimer disease. Cold Spring Harb Perspect Med 2:a006320.
- 46 Lenz S, Karsten P, Schulz JB, Voigt A (2013) Drosophila as a screening tool to study human neurodegenerative diseases. J Neurochem 127:453-460.
- 47 Levin ED, Simon BB (1998) Nicotinic acetylcholine involvement in cognitive function in animals. Psychopharmacology (Berl) 138:217-230.
- 48 Liu CF, Pan TM (2010) Recombinant expression of bioactive peptide lunasin in Escherichia coli. Appl Microbiol Biotechnol 88:177-186.
- 49 Liu J, Jia SH, Kirberger M, Chen N (2014) Lunasin as a promising health-beneficial peptide. Eur Rev Med Pharmacol Sci 18:2070-2075.
- 50 Malar DS, Devi KP (2014) Dietary polyphenols for treatment of Alzheimer's disease--future research and development. Curr Pharm Biotechnol 15:330-342.
- 51 Moran MT, Tare M, Kango-Singh M, Singh A (2013) Homeotic Gene teashirt (tsh) has a neuroprotective function in amyloid-beta 42 mediated neurodegeneration. PLoS One 8:e80829.
- 52 Newhouse P, Kellar K, Aisen P, White H, Wesnes K, Coderre E, Pfaff A, Wilkins H, Howard D, Levin ED (2012) Nicotine treatment of mild cognitive impairment: a 6-month double-blind pilot clinical trial. Neurology 78:91-101.
- 53 Newman M, Ebrahimie E, Lardelli M (2014) Using the zebrafish model for Alzheimer's disease research. Front Genet 5:189.
- 54 Nordberg A, Hellstrom-Lindahl E, Lee M, Johnson M, Mousavi M, Hall R, Perry E, Bednar I, Court J (2002) Chronic nicotine treatment reduces beta-amyloidosis in the brain of a mouse model of Alzheimer's disease (APPsw). J Neurochem 81:655-658.
- 55 O'Brien RJ, Wong PC (2010) Amyloid precursor protein processing and Alzheimers disease. Annu Rev Neurosci 34:185-204.
- 56 Oddo S, Caccamo A, Shepherd JD, Murphy MP, Golde TE, Kayed R, Metherate R, Mattson MP, Akbari Y, LaFerla FM (2003) Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Abeta and synaptic dysfunction. Neuron 39:409-421.
- 57 Pabona JM, Dave B, Su Y, Montales MT, de Lumen BO, de Mejia EG, Rahal OM, Simmen RC (2012) The soybean peptide lunasin promotes apoptosis of mammary epithelial cells via induction of tumor suppressor PTEN: similarities and distinct actions from soy isoflavone genistein. Genes Nutr 8:79-90.
- 58 Pandey UB, Nichols CD (2011) Human disease models in Drosophila melanogaster and the role of the fly in therapeutic drug discovery. Pharmacol Rev 63:411-436.
- 59 Pany S, Pal A, Sahu PK (2014) Neuroprotective effect of quercetin in neurotoxicity induced rats: Role of neuroinflammation in neurodegeneration. Asian J Pharm Clin Res 7:152-156.
- 60 Pervin M, Unno K, Ohishi T, Tanabe H, Miyoshi N, Nakamura Y (2018) Beneficial effects of green tea catechins on neurodegenerative diseases. Molecules 23:E1297.
- 61 Prussing K, Voigt A, Schulz JB (2013) Drosophila melanogaster as a model organism for Alzheimer's disease. Mol Neurodegener 8:35.
- 62 Rahman I, Biswas SK, Kirkham PA (2006) Regulation of inflammation and redox signaling by dietary polyphenols. Biochem Pharmacol 72:1439-1452.
- 63 Rajan KB, Weuve J, Barnes LL, Wilson RS, Evans DA (2018) Prevalence and incidence of clinically diagnosed Alzheimer's disease dementia from 1994 to 2012 in a population study. Alzheimers Dement 15:1-7.
- 64 Ready DF, Hanson TE, Benzer S (1976) Development of the Drosophila retina, a neurocrystalline lattice. Dev Biol 53:217-240.
- 65 Rezai-Zadeh K, Shytle D, Sun N, Mori T, Hou H, Jeanniton D, Ehrhart J, Townsend K, Zeng J, Morgan D, Hardy J, Town T, Tan J (2005) Green tea epigallocatechin-3-gallate (EGCG) modulates amyloid precursor protein cleavage and reduces cerebral amyloidosis in Alzheimer transgenic mice. J Neurosci 25:8807-8814.
- 66 Saito A, Saido T (2014) Animal models of Alzheimer's disease. Nihon Yakurigaku Zasshi 144:250-252.
- 67

Saito T, Matsuba Y, Mihira N, Takano J, Nilsson P, Itohara S, Iwata N, Saido TC (2014) Single App knock-in mouse models of Alzheimer's disease. Nat Neurosci 17:661-663.

- 68 Sarkar A, Irwin M, Singh A, Riccetti M, Singh A (2016) Alzheimer's disease: the silver tsunami of the 21(st) century. Neural Regen Res 11:693-697.
- 69 Sarkar A, Gogia N, Glenn N, Singh A, Jones G, Powers N, Srivastava A, Kango-Singh M, Singh A (2018) A soy protein Lunasin can ameliorate amyloid-beta 42 mediated neurodegeneration in Drosophila eye. Sci Rep 8:13545.
- 70 Sasaguri H, Nilsson P, Hashimoto S, Nagata K, Saito T, De Strooper B, Hardy J, Vassar R, Winblad B, Saido TC (2017) APP mouse models for Alzheimer's disease preclinical studies. EMBO J 36:2473-2487.
- 71 Schirmer H, Pereira TC, Rico EP, Rosemberg DB, Bonan CD, Bogo MR, Souto AA (2012) Modulatory effect of resveratrol on SIRT1, SIRT3, SIRT4, PGC1alpha and NAMPT gene expression profiles in wild-type adult zebrafish liver. Mol Biol Rep 39:3281-3289.
- 72 Selkoe DJ, Podlisny MB (2002) Deciphering the genetic basis of Alzheimer's disease. Annu Rev Genomics Hum Genet 3:67-99.
- 73 Selkoe DJ, Hardy J (2016) The amyloid hypothesis of Alzheimer's disease at 25 years. EMBO Mol Med 8:595-608.
- 74 Serruys PW, Onuma Y, Dudek D, Smits PC, Koolen J, Chevalier B, de Bruyne B, Thuesen L, McClean D, van Geuns RJ, Windecker S, Whitbourn R, Meredith I, Dorange C, Veldhof S, Hebert KM, Sudhir K, Garcia-Garcia HM, Ormiston JA (2011) Evaluation of the second generation of a bioresorbable everolimus-eluting vascular scaffold for the treatment of de novo coronary artery stenosis: 12-month clinical and imaging outcomes. J Am Coll Cardiol 58:1578-1588.
- 75 Shankar GM, Li S, Mehta TH, Garcia-Munoz A, Shepardson NE, Smith I, Brett FM, Farrell MA, Rowan MJ, Lemere CA, Regan CM, Walsh DM, Sabatini BL, Selkoe DJ (2008) Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. Nat Med 14:837-842.
- 76 Sierra-Fonseca JA, Gosselink KL (2018) Tauopathy and neurodegeneration: a role for stress. Neurobiol Stress 9:105-112.
- 77 Singh A (2012) Neurodegeneration- a means to an end. J Cell Sci Ther 3:e107.
- 78 Singh A, Choi KW (2003) Initial state of the Drosophila eye before dorsoventral specification is equivalent to ventral. Development 130:6351-6360.
- Singh A, Irvine KD (2012) Drosophila as a model for understanding development and disease. Dev Dyn 241:1 2.
- 80 Singh A, Shi X, Choi KW (2006) Lobe and serrate are required for cell survival during early eye development in Drosophila. Development 133:4771-4781.
- 81 Singh A, Chan J, Chern JJ, Choi KW (2005a) Genetic interaction of Lobe with its modifiers in dorsoventral patterning and growth of the Drosophila eye. Genetics 171:169-183.
- 82 Singh A, Tare M, Puli OR, Kango-Singh M (2012) A glimpse into dorso-ventral patterning of the Drosophila eye. Dev Dyn 241:69-84.
- 83 Singh A, Lim J, Choi KW (2005b) Dorso-ventral boundary is required for organizing growth and planar polarity in the Drosophila eye. In: Planar Cell Polarization during Development: Advances in Developmental Biology and Biochemistry (Mlodzik M, ed), pp 59-91. Elsevier Science & Technology Books.
- 84 Song J, Cheon SY, Jung W, Lee WT, Lee JE (2014) Resveratrol induces the expression of interleukin-10 and brain-derived neurotrophic factor in BV2 microglia under hypoxia. Int J Mol Sci 15:15512-15529.
- 85 Sparks DL, Sabbagh MN, Connor DJ, Lopez J, Launer LJ, Petanceska S, Browne P, Wassar D, Johnson-Traver S, Lochhead J, Ziolkowski C (2005) Atorvastatin therapy lowers circulating cholesterol but not free radical activity in advance of identifiable clinical benefit in the treatment of mild-to-moderate AD. Curr Alzheimer Res 2:343-353.
- 86 Steffensmeier AM, Tare M, Puli OR, Modi R, Nainaparampil J, Kango-Singh M, Singh A (2013) Novel neuroprotective function of apical-basal polarity gene crumbs in amyloid beta 42 (abeta42) mediated neurodegeneration. PLoS One 8:e78717.

Steinkraus KA, Smith ED, Davis C, Carr D, Pendergrass WR, Sutphin GL, Kennedy BK, Kaeberlein M (2008) Dietary restriction suppresses proteotoxicity and enhances longevity by an hsf-1-dependent mechanism in Caenorhabditis elegans. Aging Cell 7:394-404.

- 88 Tare M, Puli OR, and, Singh A (2013a) Molecular Genetic Mechanisms of Axial Patterning: Mechanistic Insights into Generation of Axes in the Developing Eye. In: Molecular Genetics of Axial Patterning, Growth and Disease in the Drosophila Eye (Singh A, Kango-Singh M, eds), pp 37-75. New York, USA: Springer.
- 89 Tare M, Puli OR, Moran MT, Kango-Singh M, Singh A (2013b) Domain specific genetic mosaic system in the Drosophila eye. Genesis 51:68-74.
- <sup>90</sup> Tare M, Modi RM, Nainaparampil JJ, Puli OR, Bedi S, Fernandez-Funez P, Kango-Singh M, Singh A (2011) Activation of JNK signaling mediates amyloid-ß-dependent cell death. PLoS One 6:e24361.
- 91 Tomlinson BE, Blessed G, Roth M (1968) Observations on the brains of non-demented old people. J Neurol Sci 7:331-356.
- 92 Wang J, Bi W, Cheng A, Freire D, Vempati P, Zhao W, Gong B, Janle EM, Chen TY, Ferruzzi MG, Schmeidler J, Ho L, Pasinetti GM (2014) Targeting multiple pathogenic mechanisms with polyphenols for the treatment of Alzheimer's disease-experimental approach and therapeutic implications. Front Aging Neurosci 6:42.
- 93 Williams P, Sorribas A, Howes MJ (2011) Natural products as a source of Alzheimer's drug leads. Nat Prod Rep 28:48-77.
- 94 Yang L, Hao J, Zhang J, Xia W, Dong X, Hu X, Kong F, Cui X (2009) Ginsenoside Rg3 promotes beta-amyloid peptide degradation by enhancing gene expression of neprilysin. J Pharm Pharmacol 61:375-380.
- 95 Zangara A (2003) The psychopharmacology of huperzine A: an alkaloid with cognitive enhancing and neuroprotective properties of interest in the treatment of Alzheimer's disease. Pharmacol Biochem Behav 75:675-686.
- 96 Zhang HY, Zheng CY, Yan H, Wang ZF, Tang LL, Gao X, Tang XC (2008) Potential therapeutic targets of huperzine A for Alzheimer's disease and vascular dementia. Chem Biol Interact 175:396-402.

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