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# Hydrocyclones for single-use perfusion application

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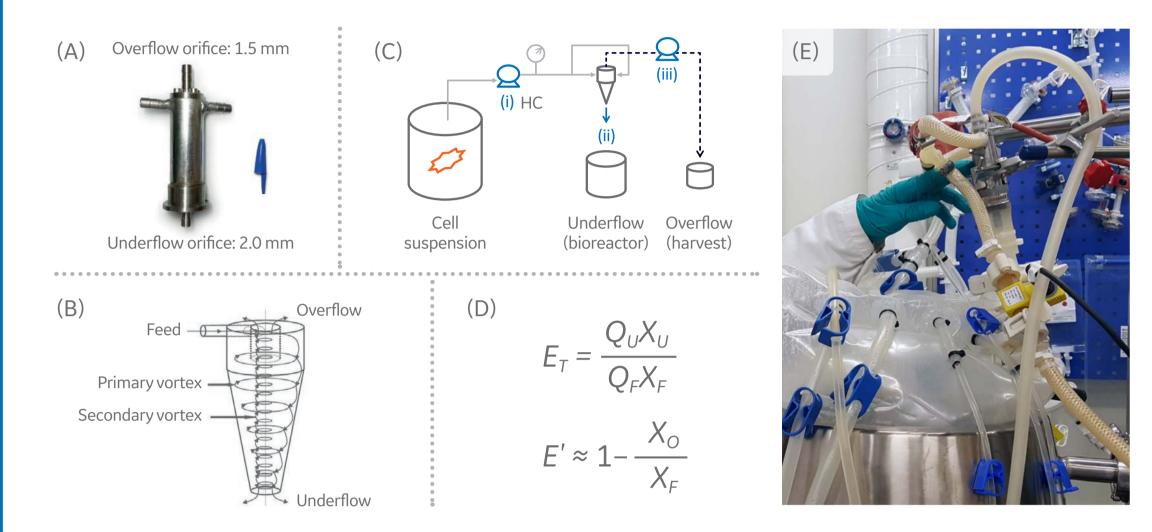
# Why is a hydrocyclone a suitable cell retention device for perfusion applications?

- No clogging
- Compatibility with single-use technology
- No product retention Possibility of 3D printing
- No moving parts

# Perfusion runs with the hydrocyclone coupled to a 50-L single-use bioreactor bag

- **Perfusion #1:** HC was installed in a late-stage fed-batch culture, but not with an optimal configuration. Nevertheless, recovery of viability and cell growth were successfully achieved (Fig 5A).
- **Perfusions #2 and #3:** bioreactor bags were customized with a ReadyMate<sup>™</sup> TC port on top of the bag wide enough to enable the umbrella-type underflow discharge (Fig 1E). Perfusion runs achieved high cell viabilities with cell-specific perfusion rates of 50 down to 15 pL/cell/d (Figs 5B–C).
- Perfusion #3: E<sub>T</sub> up to 96% and E' of 79% were achieved at ΔP > 2 bar (Fig 5D). A natural cell bleed with diluted cells occurred through the overflow orifice. Additionally, there was a preferential retention of viable cells, since non-viable cells and debris with smaller sizer were eliminated through the overflow, contributing to a healthier culture environment (Fig 5E).

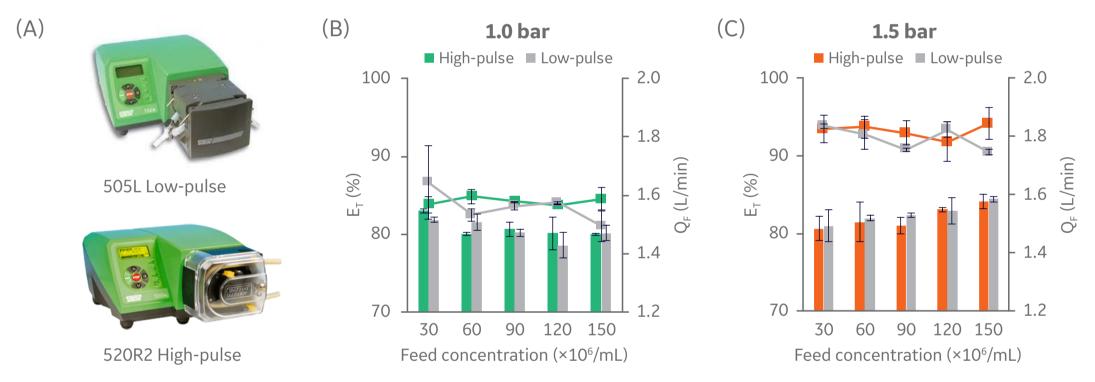
# Characterization of the hydrocyclone



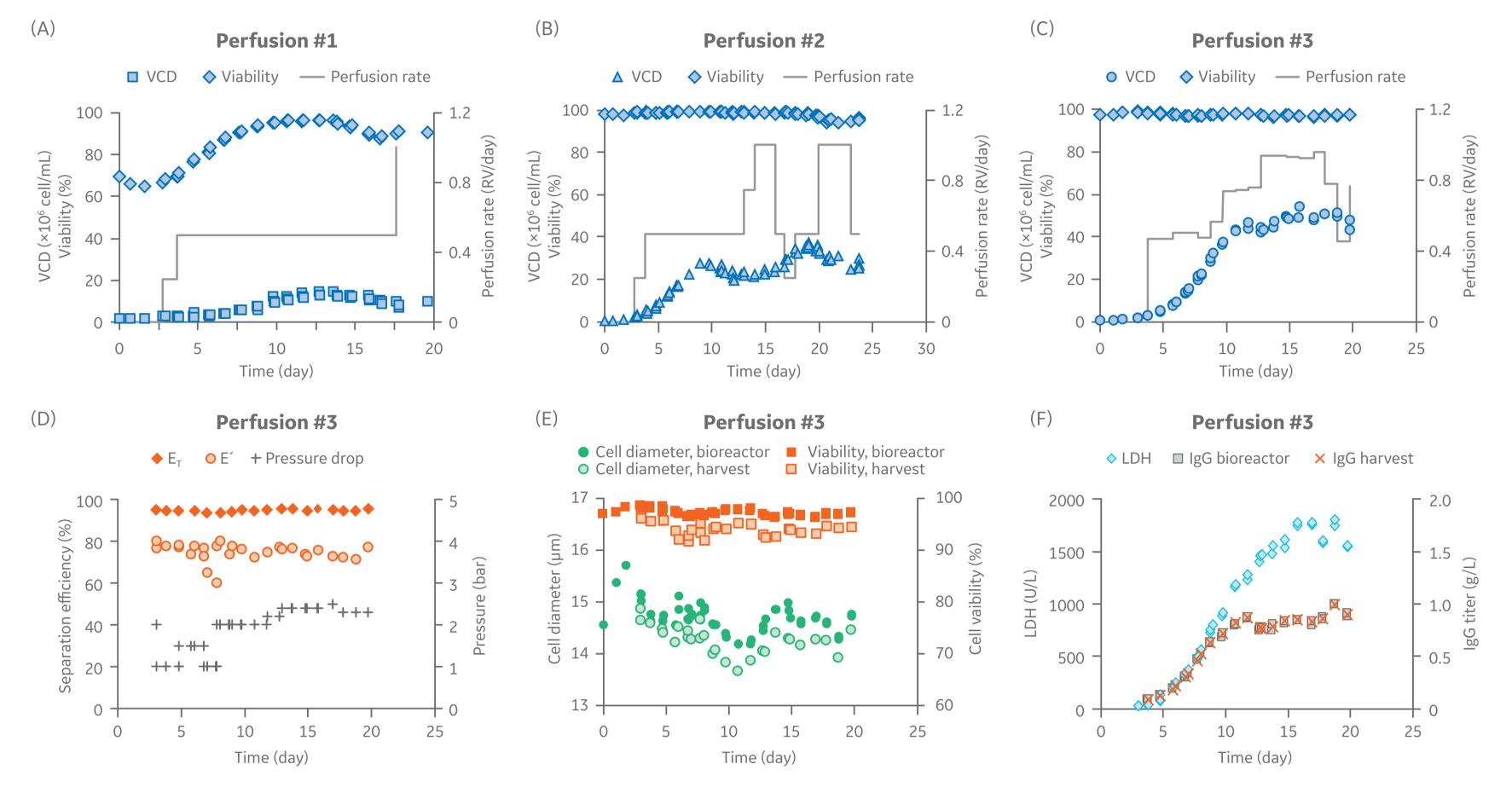
**Fig 1.** (A) Stainless-steel prototype HC2015; (B) Fluid flow in the HC; (C) Test rig for batch separations to investigate variables relevant to HC attachment (depicted in blue): (i) feed pump, (ii) tubing leaving the underfow, and (iii) overflow pump; (D) Total ( $E_T$ ) and reduced (E') separation efficiencies calculated to evaluate cell retention performance; (E) HC2015 placed on top of a 50-L single-use bioreactor bag.

#### (i) Is it important to have a low pulsation pump to feed the HC?

• No, we achieved comparable performance between a low- and high pulse peristaltic pump.



- An increase in pressure drop up to 2.2 bar for HC operation did not negatively affect cell viability. Increase of LDH levels over time correlated with the viability profile (Fig 5F).
- No IgG retention inside the bioreactor was observed (Fig 5F).

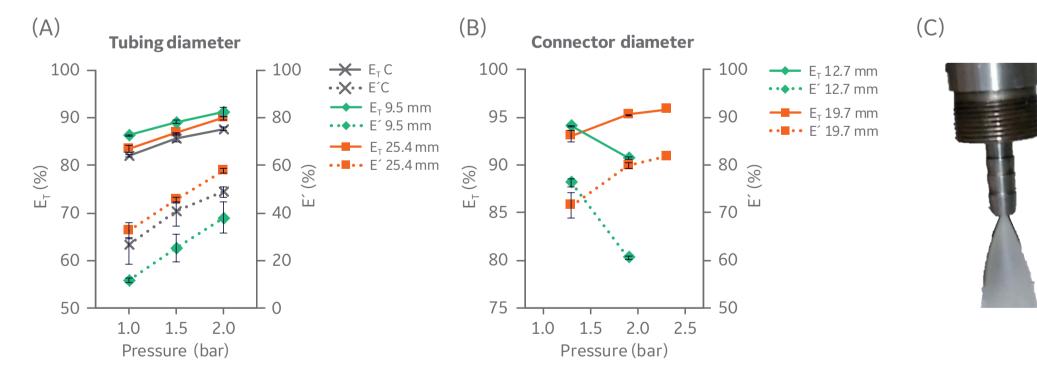


**Fig 5.** (A–C) Viable cell density (VCD), viability, and perfusion rate in 3 perfusion runs with HC2015. In run #3, optimal operational conditions for the HC separation with no restriction in underflow tubing, coupled to feeding strategy with supplemented ActiPro<sup>™</sup> medium resulted in up to 50 × 10<sup>6</sup> cells/mL. (D) Separation efficiencies (E<sub>T</sub> and E') obtained in run #3. (E) Viabilities and cell diameter measured in the bioreactor and in the harvest for run #3. (F) Lactate dehydrogenase (LDH) activity and product titer for run #3.

**Fig 2.** (A) Watson-Marlow pump heads in the feed peristaltic pump. Total separation efficiencies ( $E_{T}$ , bars) and feed flow rates ( $Q_{F}$ , lines) obtained with both pump head models at (B) 1 bar and (C) 1.5 bar.

#### (ii) How important is the design of the underflow geometry?

- Flow restrictions imposed by means of a 9.5-mm tubing or a 12.7-mm reducer had a negative impact on reduced separation efficiency (Figs 3A–B).
- Recirculation loop should be large enough to enable the umbrella-pattern discharge in the underflow line (Fig 3C).



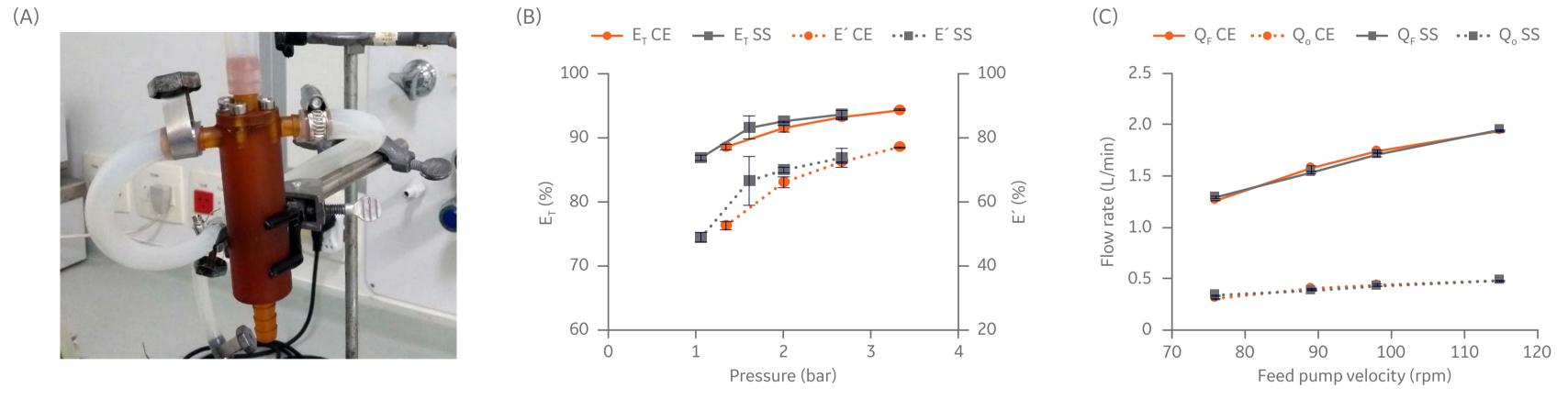
**Fig 3.** (A) Effect of tubing diameter on separation of Superose<sup>™</sup> 6 beads (model for cells) at 5 × 10<sup>6</sup> beads/mL: open underflow discharge (C), 9.5-mm tubing attached to underflow nozzle, or 25.4-mm tubing attached to HC cylindrical body. (B) Effect of presence or absence of 12.7-mm ID reducer in 19.7-mm ReadyMate devices on separation of CHO cells at ~ 20 × 10<sup>6</sup> cells/mL. (C) Typical umbrella-pattern underflow discharge.

#### (iii) Is it beneficial to use a perfusate pump in the overflow line?

• No. The very high  $E_{T}$  values (Fig 4A) were the result of a very high flow ratio  $(Q_{U}/Q_{F})$ , and not a result of centrifugal action (lower E<sup>'</sup>). Therefore, a less clarified harvest

# Can hydrocyclones be 3-D printed?

• Yes, preliminary tests with the 3-D printed plastic prototype shown in Fig 6A showed comparable performance, confirming that geometrical proportions inside the hydrocyclone play a key role in separation efficiency.

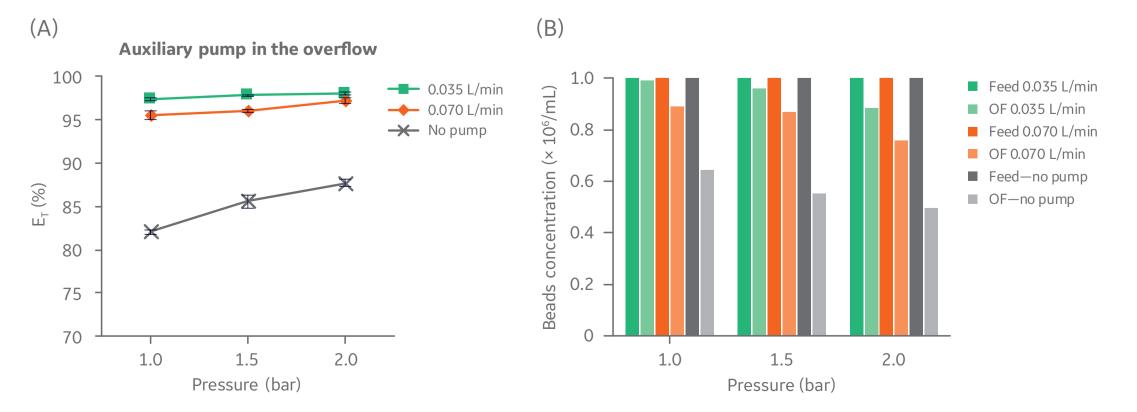


**Fig 6.** (A) 3-D printed HC2015, made of cyanate ester (CE). Comparison of plastic and stainless steel (SS) prototypes at the same feed pump velocity: (B) separation efficiencies. (C) flow rates in the feed, Q<sub>F</sub>, and in the overflow, Q<sub>o</sub>.

# Conclusion

 For the first time, a hydrocyclone set-up is reported to enable perfusion processes at cell densities in the range of 20 to 50 × 10<sup>6</sup> cells/mL for 20 to 25 days.

## was obtained through the overflow (Fig 4B).



**Fig 4.** Use of a 520R2 peristaltic pump controlling Q<sub>o</sub> at 0.035 (green) or 0.070 L/min (red), or no pump. These Q<sub>o</sub> values refer to 1 and 2 RV/d, respectively, for a bioreactor working volume of 50 L. (A) Total separation efficiency. (B) Concentration in the feed and overflow. Superose beads were used as model particles in these tests.

#### • Also for the first time, a HC is operated attached to a single-use bioreactor.

- Pressure drops in the HC higher than 1 bar promoted high separation efficiencies and did not affect cell viability, LDH level, and mAb production.
- An intermittent perfusion at working volume of 40 L bioreactor was successfully operated at perfusion rates up to 1 RV/d. The time intervals of feed pump on and feed pump off can be easily manipulated by means of a timer to increase or decrease the medium exchange per day.
- When continuously operated, this HC can process over 500 L/d of perfusate.

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