COFACTOR SWITCH: DEVELOPMENT OF A NAD*-DEPENDENT CASCADE FOR THE PRODUCTION OF URSODEOXYCHOLIC ACID (UDCA)

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The employment of alcohol dehydrogenases in cascade reactions is often limited by the different cofactor specificity of the enzymes involved: the employment of additional cofactor regeneration systems and the excess amount of sacrificial substrates frequently increase the environmental impact and the costs of biocatalytic processes. Additionally, a NADP+-dependent process is generally less desirable, inasmuch this cofactor is more expensive, unstable and less naturally available than NAD+, leading to an increase of the process costs. Nowadays, protein engineering offers the possibility to switch the cofactor dependency of enzymes introducing few targeted mutation¹.

We applied this methodologies for the development of the first fully NAD⁺ mediated cascade for the production of Ursodeoxycholic acid (UDCA), a widely used pharmaceutical ingredient for the clinical treatment of cholesterol gallstones and liver diseases². The enzymatic epimerization of CDCA into UDCA, can be carried out with two specific hydroxysteroid dehydrogenases (HSDH): the 7a-OH group is firstly oxidized to the ketone by



7α-HSDH and subsequently rereduced with opposite stereochemistry (7 β -OH) by 7 β -HSDH. All procedures up to now require a set of a respectively NAD⁺ and NADP⁺ dependent enzymes. Nevertheless, gene sequences of NAD+-dependent 7β-HSDH were not reported. In order to obtain a full-NAD+-dependent process, the NADP⁺ dependent 7β-HSDH from Clostridium absonum was engineered. Employing a semi-rational mutagenesis approach, we obtained a variant with shifted cofactor preference. Importantly, this study allows to identify the residues responsible of the cofactor recognition in other 7β-HSDH homologues and, thus, to the identification of the gene coding for a wild-type NADH-dependent homologue from Lactobacillus

spicheri (Ls7β-HSDH). These novel NAD⁺-dependent 7β-HSDH enzymes in combination with 7α-HSDH from Stenotrophomonas maltophilia permitted the redox-neutral biotransformations of CA and CDCA in the presence of catalytic amounts of NAD+, resulting in high yields (>90 %) of UCA and UDCA3.

Further studies are underway to develop a flow process for this industrially relevant biotransformation.

Cahn, J. K.; Werlang, C. A.; Baumschlager, A.; Brinkmann-Chen, S.; Mayo, S. L.; Arnold, F. H., A (1)general tool for engineering the NAD/NADP cofactor preference of oxidoreductases. ACS Synthetic Biology 2016, 6 (2), 326-333.

Tonin, F.; Arends, I. W., Latest development in the synthesis of ursodeoxycholic acid (UDCA): a critical (2)review. Beilstein J. Org. Chem. 2018, 14, 470.

(3)Tonin, F.; Otten, L. G.; Arends, I. W., NAD+-Dependent Enzymatic Route for the Epimerization of Hydroxysteroids. ChemSusChem 2018, DOI: 10.1002/cssc.201801862.