



## Termination of Vernakalant-Resistant Atrial Fibrillation by Inhibition of Small-Conductance Ca<sup>2+</sup>-Activated K<sup>+</sup> Channels in Pigs

Diness, Jonas Goldin; Skibsbye, Lasse; Simó-Vicens, Rafel; Louro Larupa dos Santos, Joana; Lundegaard, Pia Rengtved; Citerni, Carlotta; Sauter, Daniel Rafael Peter; Bomholtz, Sofia Hammami; Svendsen, Jesper Hastrup; Olesen, Søren-Peter; Sørensen, Ulrik S; Jespersen, Thomas; Grønnet, Morten; Bentzen, Bo Hjorth

*Published in:*

Circulation: Arrhythmia and Electrophysiology

*DOI:*

[10.1161/CIRCEP.117.005125](https://doi.org/10.1161/CIRCEP.117.005125)

*Publication date:*

2017

*Document version*

Publisher's PDF, also known as Version of record

*Document license:*

[CC BY](#)

*Citation for published version (APA):*

Diness, J. G., Skibsbye, L., Simó-Vicens, R., Louro Larupa dos Santos, J., Lundegaard, P. R., Citerni, C., ... Bentzen, B. H. (2017). Termination of Vernakalant-Resistant Atrial Fibrillation by Inhibition of Small-Conductance Ca<sup>2+</sup>-Activated K<sup>+</sup> Channels in Pigs. *Circulation: Arrhythmia and Electrophysiology*, 10, [e005125]. <https://doi.org/10.1161/CIRCEP.117.005125>



# Termination of Vernakalant-Resistant Atrial Fibrillation by Inhibition of Small-Conductance Ca<sup>2+</sup>-Activated K<sup>+</sup> Channels in Pigs

See Editorial by Heijman and Dobrev

**BACKGROUND:** Evidence has emerged that small-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> (SK) channels constitute a new target for treatment of atrial fibrillation (AF). SK channels are predominantly expressed in the atria as compared with the ventricles. Various marketed antiarrhythmic drugs are limited by ventricular adverse effects and efficacy loss as AF progresses.

**METHODS AND RESULTS:** A total of 43 pigs were used for the studies. AF reversion in conscious long-term tachypaced pigs: Pigs were subjected to atrial tachypacing (7 Hz) until they developed sustained AF that could not be reverted by vernakalant 4 mg/kg (18.8±3.3 days of atrial tachypacing). When the SK channel inhibitor AP14145 was tested in these animals, vernakalant-resistant AF was reverted to sinus rhythm, and reinduction of AF by burst pacing (50 Hz) was prevented in 8 of 8 pigs. Effects on refractory period and AF duration in open chest pigs: The effects of AP14145 and vernakalant on the effective refractory periods and acute burst pacing-induced AF were examined in anaesthetized open chest pigs. Both vernakalant and AP14145 significantly prolonged atrial refractoriness and reduced AF duration without affecting the ventricular refractoriness or blood pressure in pigs subjected to 7 days atrial tachypacing, as well as in sham-operated control pigs.

**CONCLUSIONS:** SK currents play a role in porcine atrial repolarization, and pharmacological inhibition of these with AP14145 demonstrates antiarrhythmic effects in a vernakalant-resistant porcine model of AF. These results suggest SK channel blockers as potentially interesting anti-AF drugs.

Jonas Goldin Diness, PhD\*  
 Lasse Skibsbye, PhD  
 Rafel Simó-Vicens, MSc  
 Joana Larupa Santos, MSc  
 Pia Lundegaard, PhD  
 Carlotta Citerni, DMV  
 Daniel Rafael Peter Sauter, PhD  
 Sofia Hammami Bomholtz, PhD  
 Jesper Hastrup Svendsen, MD  
 Søren-Peter Olesen, MD, DMSc  
 Ulrik S. Sørensen, PhD  
 Thomas Jespersen, PhD, DMSc  
 Morten Grunnet, PhD, DrSc  
 Bo Hjorth Bentzen, PhD\*

\*Drs Diness and Bentzen contributed equally to this work.

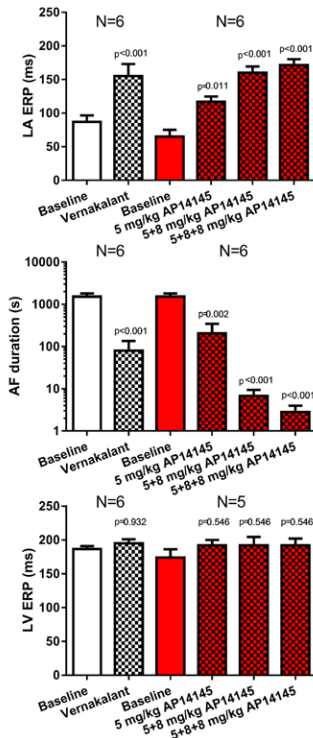
**Correspondence to:** Jonas Goldin Diness, PhD, Acesion Pharma, Blegdamsvej 3B, DK-2200, Copenhagen, Denmark. E-mail jgd@acesionpharma.com

**Key Words:** atrial fibrillation  
 ■ blood pressure ■ heart atria  
 ■ swine ■ vernakalant

© 2017 The Authors. *Circulation: Arrhythmia and Electrophysiology* is published on behalf of the American Heart Association, Inc., by Wolters Kluwer Health, Inc. This is an open access article under the terms of the [Creative Commons Attribution License](#), which permits use, distribution, and reproduction in any medium, provided that the original work is properly cited.

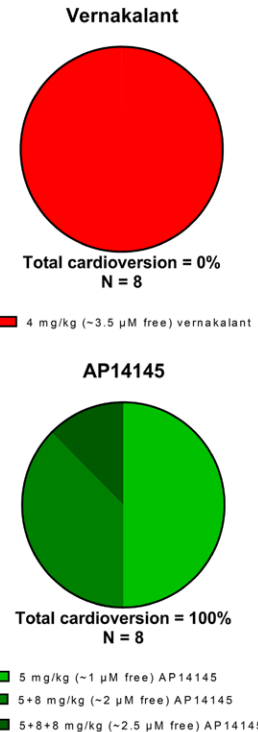
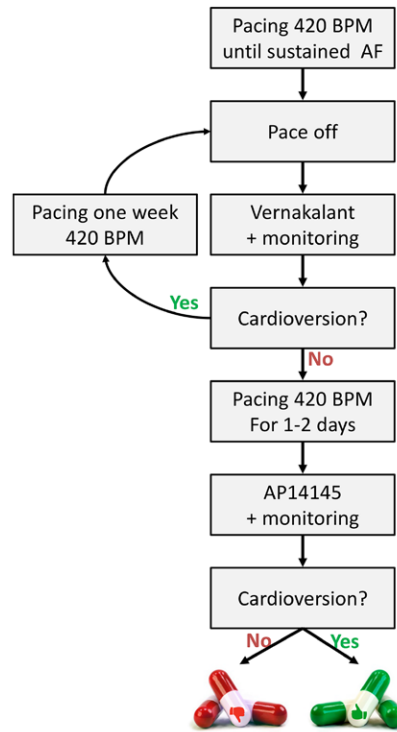
1-week tachypaced pigs

- Both vernakalant and AP14145 act antiarrhythmically by increasing the atrial effective refractory period and decreasing AF duration
- Neither vernakalant nor AP14145 have effects on ventricular repolarization



Pig model of vernakalant-resistant AF

- 10-38 days of tachypacing
- Vernakalant does not cardiovert AF
- AP14145 cardioverts vernakalant-resistant AF



Effective, safe, and tolerable pharmacological treatment for atrial fibrillation (AF) remains an unmet need. The latest medication to reach the market for intravenous cardioversion was the combined sodi-

um and potassium channel inhibitor vernakalant, however not yet available in the United States. Vernakalant terminated ~50% of episodes of AF lasting <7 days in randomized controlled studies, with its highest conversion rate during the first few days while after 8 to 45 days of AF, the conversion rate was <10%, which was not statistically different from that of placebo.<sup>1,2</sup>

WHAT IS KNOWN?

- Small-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> (SK) channels are predominantly expressed in the atria as compared with the ventricles.
- Various marketed antiarrhythmic drugs are limited by ventricular adverse effects and efficacy loss as atrial fibrillation progresses.

WHAT THE STUDY ADDS?

- Data from a newly developed porcine model of vernakalant-resistant atrial fibrillation showing that SK channel currents play a role in porcine atrial repolarization.
- Data showing that pharmacological inhibition of SK channel current with AP14145 demonstrates antiarrhythmic effects in a vernakalant-resistant porcine model of atrial fibrillation.

If lasting for >24 hours, AF promotes further progression of the disease—a phenomenon described as AF begets AF.<sup>3</sup> If atrial remodeling continues, AF often progresses to more sustained forms and becomes more resistant to both pharmacological and nonpharmacological treatments, including ablation.<sup>4-6</sup> Among contributing factors, an increased influx of calcium seems to promote fibrosis development and remodeling.<sup>7</sup>

Three subtypes of small-conductance calcium-activated K<sup>+</sup> channels, SK1, SK2, and SK3, are represented in the heart. They are all exclusively activated by intracellular Ca<sup>2+</sup>, and the current conducted by these channels is recognized as I<sub>KCa</sub>. Originally, SK channels were cloned from the central nervous system where they contribute to the afterhyperpolarization of the neuronal action potential.<sup>8</sup> With the finding of SK channels in cardiac tissue, it was demonstrated that these channels are

also important for the repolarization of cardiac action potentials.<sup>8–11</sup> Confirmation of the role of SK channels in human AF has since been provided in several genome-wide association studies concluding that common variants in the genes encoding SK2 and SK3 are associated with AF.<sup>12–14</sup> We demonstrated that  $I_{KCa}$  inhibition can terminate AF or protect against its induction in models of AF in isolated perfused heart preparations from rat, guinea pig, and rabbit, as well as in in vivo models of AF in rat, dog, and horse.<sup>8–11,15</sup> We also obtained evidence that SK channels play a functional role in right atrial (RA) appendage tissue, but not ventricular tissue, from patients in sinus rhythm (SR), as well as patients with >6 months of AF, and that inhibition of  $I_{KCa}$  with the SK channel inhibitor NS8593 prolonged refractory periods in atrial tissue from these patients.<sup>16</sup> Preliminary evidence suggests that the  $I_{KCa}$  is increased in patients with paroxysmal AF compared with patients in SR, and  $I_{KCa}$  inhibition by the peptide apamin was associated with a 4-fold increased action potential prolongation in cells from patients in paroxysmal AF.<sup>17</sup>

Significant antiarrhythmic effects in normal and remodeled atria, without adverse effects in the ventricles, have long been a desirable, but nonachievable, therapeutic goal. This seems possible by inhibition of the SK channels, which may, therefore, qualify as a promising target for the development of a novel agent for treatment of AF.

In a preclinical canine model of vagotonic AF, vernakalant cardioverted 7 of 7 dogs at a dose of 4 mg/kg.<sup>18</sup>

To obtain a preclinical model better simulating clinical AF, we performed long-lasting atrial tachypacing (AT) in pigs to develop sustained AF until it was no longer possible to convert with a clinically relevant dose of vernakalant. In this model of sustained vernakalant-resistant AF, we tested whether an SK channel inhibitor, the tool compound AP14145, could successfully convert AF to SR and protect against reinduction of AF.

## METHODS

### In Vitro Electrophysiology

Effects of AP14145 on hSK3 (KCa2.3) and hERG (hKV11.1) were investigated by automated patch clamp (Qpatch; Biolin Scientific, Sophion, Denmark) as previously described.<sup>19</sup> Likewise, the effect on rNa<sub>v</sub>1.5 was evaluated as earlier described<sup>20</sup> but with a modified voltage protocol so that rNa<sub>v</sub>1.5 currents were elicited with 1-second interval by a 50-ms depolarizing step to –20 mV from a holding potential of –120 mV (a total of 80 pulses). Effect of AP14145 on L-type calcium channels was investigated by automated patch clamping (Qpatch; Biolin Scientific) using a hCa<sub>v</sub>1.2 stable cell line ( $\alpha$ 1C,  $\beta$ 2C, and  $\alpha$ 2delta; SB Drug Discovery, Glasgow, United Kingdom).

Off-target effects on Kir3.1/Kir3.4 ( $I_{KACH}$ ), K<sub>v</sub>1.5 ( $I_{Kur}$ ), K<sub>v</sub>7.1/KCNE1 ( $I_{Ks}$ ), K<sub>v</sub>4.3/KChIP2 ( $I_{to}$ ), and Kir2.1 ( $I_{K1}$ ) were investigated using 2-electrode voltage-clamp experiments on *Xenopus laevis* oocytes expressing the relevant channels.<sup>19</sup> For further details, please see [Data Supplement](#).

## Animal Models

All animal studies were performed under a license from the Danish Ministry of Environment and Food (license No. 2012-15-2934-00083) and in accordance with the Danish guidelines for animal experiments according to the European Commission Directive 86/609/EEC. A total of 43 Danish landrace pigs (from Kringsagergård w. Henrik Larsen, Gilleleje, Denmark; 12–13 weeks old, 30–35 kg gilts) were studied. The pigs were divided into 4 main groups: (1) open chest surgery (n=21), (2) cardioversion in conscious pigs (n=8), (3) expression studies (n=12), and (4) pharmacokinetic studies (n=2). The open chest surgery group consisted of pigs subjected to 1-week AT and sham-operated time matched controls (CTRL). The group for conscious cardioversion studies consisted of long-term AT pigs with persistent AF that could not be converted by a clinically relevant dose of vernakalant. The group for expression studies was divided into long-term AT pigs and control pigs.

All pigs underwent the following procedure: After pre-medication with zoletil pig mixture (250 mg dry tiletamin+zolazepam, 6.5 mL xylazine 20 mg/mL, 1.25 ketamine 100 mg/mL, 2.5 mL butorphanol 10 mg/mL, and 2 mL methadone 10 mg/mL) 0.1 mL/kg given intramuscularly, the pig was given an intravenous infusion of propofol and fentanyl (15 mg/kg per hour and 50  $\mu$ g/kg per hour, respectively) and intubated and ventilated with a tidal volume of 10 mL/kg and a respiration frequency of 12 to 14 per minutes. During surgery arterial partial pressure of carbon dioxide (Paco<sub>2</sub>), blood pressure and ECG were monitored, and the pig was given 6 mL/kg per hour isotonic saline solution. Under aseptic conditions and fluoroscopic guidance, a bipolar pacing-electrode was inserted into the RA appendage and connected to a neurostimulator (Medtronic Synergy versitrel or Itriel 3) implanted subcutaneously in the neck region. The pigs in the long-term AT group received a central venous catheter in the internal jugular vein with an exit in the dorsal neck region.

### Open Chest Surgery

In the 1-week AT pigs, after 7 days recovery after pacemaker implantation, the pacemakers were turned on and the RA was paced 420 beats per minute (BPM) for 7 days. To avoid symptoms of heart failure caused by a high ventricular rate because of pacing, the pigs were medicated with digoxin (250  $\mu$ g/d) starting 4 days before turning on the pacemaker and discontinued 2 days before open chest surgery. The same procedure was used in the CTRL pigs except that the pacemakers remained turned off in this group. Sixteen hours before study days, the pacemaker was deactivated. On study days, pigs were anesthetized as described for the pacemaker implantation. After median sternotomy, custom-made hook-shaped teflon-coated stainless steel electrodes for recording and stimulation were inserted into the left atrial (LA) appendage and the left ventricle (LV). If the placement of the electrodes gave rise to AF, which was the case in all of the 1-week AT pigs, the AF was DC cardioverted (Zoll M Series, ZOLL Medical, Cheshire, United Kingdom) with the electrodes placed on the LA and RA using 5 to 30 J as appropriate. After conversion to SR, the heart was allowed 30 minutes of stabilization.

A programmable stimulator (Hugo Sachs, Germany) was used to deliver 2-ms double-rheobase current pulses, and the LA effective refractory period (ERP), the LV ERP, and the mean AF duration after burst pacing were measured. The LV ERP and LA ERP were measured at a basic cycle length of 400 ms. The ERP was determined as the longest S1-S2 failing to capture, with 10 basic stimuli (S1) followed by a premature extra stimulus (S2) with 5 ms increments. AF was induced by burst pacing at 50 Hz and 10 V. The mean AF duration was based on 10 AF inductions in each pig or 5 AF inductions if the mean AF duration of the first 5 AF durations was >1 minute and <5 minutes or as many AF inductions as time allowed (usually 1 or 2) within 30 minutes if the AF durations lasted >5 minutes. In 12 of 14 1-week AT pigs AF was induced by the first S2 stimulus after the LA ERP and was sustained for at least 30 minutes after which drug treatment was started.

Before the experiments, each pig was randomly assigned to receive either AP14145 or vernakalant 30 minutes after induction of the first AF episode.

Vernakalant was given as an infusion of 0.12 mg/kg per minute (60 minutes) followed by a maintenance infusion of 0.08 mg/kg per minute to produce a steady plasma concentration of  $\approx 4000$  ng/mL as described in ref<sup>21</sup>, equivalent to the maximal plasma concentration measured in patients in the ACT I clinical trial (Arrhythmia Conversion Trial).<sup>22</sup> Recordings after baseline in the groups receiving vernakalant were all conducted at steady state plasma concentration of vernakalant. AP14145 was given as 3 consecutive bolus injections for 1 to 2 minutes of 5, 8, and 8 mg/kg with 30 minutes intervals. Recordings in the groups receiving AP14145 were conducted a few minutes after each bolus dose.

### Cardioversion in Conscious Pigs

In the long-term AT pigs, after 7 days of recovery, the pacemakers were turned on and the RA was paced at 420 BPM (Figure 1). The pigs were medicated with digoxin (250  $\mu$ g/d) starting 4 days before turning on the pacemaker. After 7 days pacing, the pigs were Holter-monitored (Televet 100; Engel Engineering Service GmbH, Heusenstamm, Germany) while the pacemaker was turned off. Sustained AF was defined as AF that was sustained after the pacemaker was turned off during >10 minutes monitoring and an additional 10 minutes with infusion of saline (1 mL/kg) followed by 30 minutes monitoring, a total of 50 minutes of AF. When sustained AF had been confirmed, vernakalant (4 mg/kg) was infused for 10 minutes, and the pig was monitored for an additional 30 minutes. If AF converted at any point, the pig was paced 420 BPM for another 7 days, and the procedure was repeated. When AF could no longer be converted to SR by vernakalant, the pig was paced 420 BPM for another 24 to 48 hours to allow vernakalant to be cleared from the system (half-life, 2–3 hours). After this, the pacemaker was turned off under Holter-monitoring. After having observed 10 minutes of AF, 1 mL/kg vehicle was injected over 2 minutes, and the ECG was monitored for 30 minutes. When AF did not convert to SR within this period, 5 mg/kg AP14145 was injected over 2 minutes, and the ECG was monitored for 30 minutes. If AF converted within 30 minutes, burst pacing with 50 Hz was given thrice. If AF did not convert or if a single AF episode lasting for >10 minutes could be

induced with burst pacing, an additional 8 mg/kg AP14145 was injected over 2 minutes and the ECG was monitored for 30 minutes. If AF converted within 30 minutes, burst pacing with 50 Hz for 5 to 15 seconds was given thrice. If AF did not convert or an AF episode of at least 10 minutes duration could be induced with burst pacing, a final dose of 8 mg/kg AP14145 was injected over 2 minutes and the ECG was monitored for another 30 minutes. If at any point AP14145 injection gave rise to unexpected adverse effects, the pig was sedated by a small dose of zoletil while the experiment was completed. If at any point AF was converted and could not be reinduced with burst pacing, the experiment was terminated. Cardioversion was considered successful if AF was converted. Protection against reinduction of AF was considered successful if AF lasting >10 minutes could not be reinduced by burst pacing.

### Pharmacokinetic Measurements

Two pigs underwent pharmacokinetic studies and were pre-medicated, anaesthetized, intubated, and ventilated as described above.  $Paco_2$ , temperature, and ECG were monitored. Invasive blood pressure was measured in the right femoral artery, and the pig was given 6 mL/kg per hour isotonic salt solution in an ear vein. At T=0, 5 mg/kg AP14145 was infused in the left external jugular vein, and blood samples were taken from the right femoral vein at (in minutes) T=-10, 5, 10, 15, 20, 30, 40, 60, 80, 100, 140, and 180 and stored in heparin treated tubes. Blood samples were centrifuged at 2G for 5 minutes at 4°C, and the plasma samples analyzed for AP14145 concentration at CiToxLAB ScanTox AS, Ejby, Denmark.

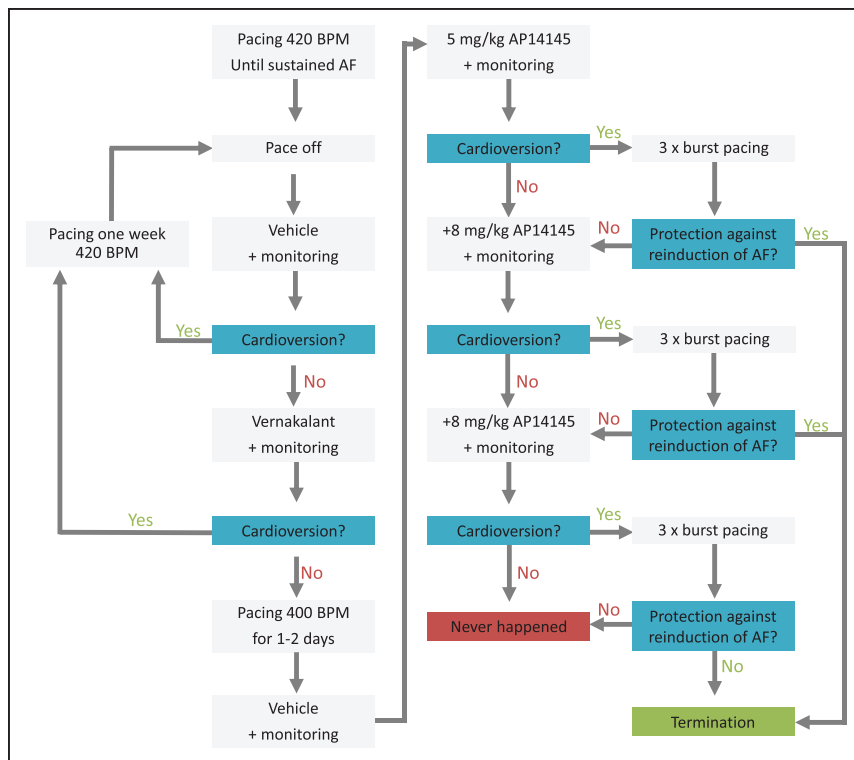
### KCNX Expression in Pig Hearts

Cardiac tissue from each of the 4 heart chambers was obtained from long-term AT pigs (n=6) or healthy controls (HC; n=6). After pre-medication with zoletil pig mixture (250 mg dry tiletamin+zolazepam, 6.5 mL xylazine 20 mg/mL, 1.25 ketamine 100 mg/mL, 2.5 mL butorphanol 10 mg/mL, and 2 mL methadone 10 mg/mL) 0.1 mL/kg given intramuscularly, the pigs were euthanized by intravenous injection of pentobarbital 200 mg/mL, and the hearts were excised and placed in ice-cold cardioplegic solution (NaCl 110.0, KCl 16.0,  $MgCl_2$  16.0,  $CaCl_2$  1.2, and  $NaHCO_3$  10.0 mmol/L). Total RNA from tissue samples was extracted according to manufacturer's instruction (miRNeasy Mini Kit, Qiagen, CA). RNA was reverse transcribed to cDNA using the nanoscript2 kit (Primerdesign, United Kingdom). Expression profiling of the SK channels (*KCNN1*, *KCNN2*, and *KCNN3*) was performed using quantitative real-time quantitative polymerase chain reaction with Taqman double dye probes (Primerdesign, United Kingdom). For further details, please see [Data Supplement](#).

### Data and Statistical Analyses

GraphPad Prism software (GraphPad Software, San Diego, CA) and Chart 7 software (ADInstruments) were used for data analyses and figures. Continuous data other than AF durations were summarized using the mean $\pm$ SEM. AF durations were log-transformed to pass a





**Figure 1. Flowchart for cardioversion of vernakalant-resistant atrial fibrillation (AF) in conscious pigs.** BPM indicates beats per minute.

Kolmogorov–Smirnov normality test. In the text, AF duration was given as a back transformation the mean of log AF duration ( $10^{\text{mean of log AF duration}}$ ) followed by the interval ( $10^{\text{mean of log AF duration} - \text{SEM}}$ – $10^{\text{mean of log AF duration} + \text{SEM}}$ ) in parentheses. In figures, the AF duration is shown as the raw AF duration on a logarithmic scale. Expression data were log-transformed to obtain a more Gaussian distribution. In figures, the expression data are shown as the raw values on a logarithmic scale.

Wilcoxon matched pairs signed-rank test was used to compare pooled expression data from AF and HC pigs for LA versus LV and RA versus right ventricle. Mann–Whitney test was used to compare the differences in expression of KCNN2 and KCNN3 in AF versus HC, as well as LA ERP, LV ERP, and AF durations of CTRL and 1-week AT groups. Fisher exact test was used to compare the number of pigs with AF durations >30 minutes in the CTRL and 1-week AT groups. Ordinary 1-way ANOVA and Sidak correction for multiple comparisons test were used to compare mean values of heart rates in 1-week AT before AT, right after AT, and at vernakalant-resistant AF, as well as for comparing LA ERP, LV ERP, and log AF duration by concentration of AP14145 and vernakalant as compared with the respective baseline values.

P values are given with 3 significant digits, and values of  $P < 0.05$  were considered statistically significant.

## Compounds and Solution

Vernakalant (Brinavess) was purchased from a Danish pharmacy as a 20-mg/mL solution for intravenous infusion. Before infusion, it was diluted in sterile isotonic saline to yield a final concentration of 4 mg/mL. AP14145 is an Acesion Pharma proprietary compound and was synthesized at Syngene International, Bangalore, India. AP14145 was dissolved in

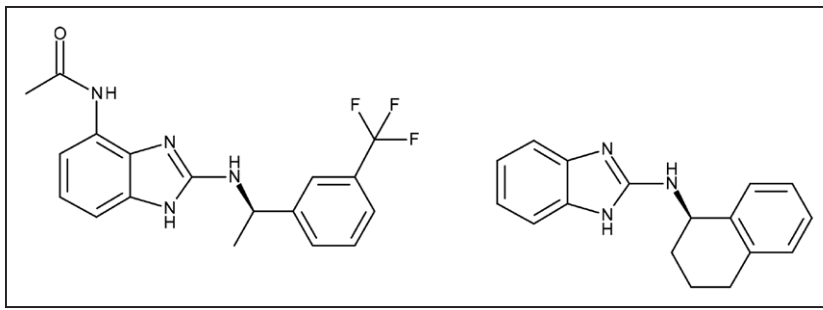
dimethyl sulfoxide for cellular electrophysiology or saline and 30% hydroxypropylbetacyclodextrin (Roquette, France) for pig experiments. The solution was sterile filtered before use.

## RESULTS

### AP14145: a New Small Molecule SK Channel Inhibitor

The small molecule compound AP14145 (*N*-(2-((1*R*)-1-[3-(trifluoromethyl)phenyl]ethyl)amino)-1*H*-1,3-benzodiazol-4-yl) acetamide (Figure 2) was developed as a novel potent SK3 channel inhibitor and has structural similarities to the earlier published SK channel inhibitor NS8593.<sup>23</sup>

AP14145 demonstrated an  $IC_{50}$  in whole-cell patch clamp on the human SK3 channel of  $1.3 \pm 0.4 \mu\text{mol/L}$  with a Hill-slope of  $1.4 \pm 0.7$  showing no cooperativity (Figure 3, left). To investigate the selectivity of AP14145, the effect of the compound was tested on a panel of cardiac ion channels (Figure 1 in the [Data Supplement](#)). AP14145 inhibited hERG ( $K_v11.1$ ) with an  $IC_{50}$  of  $71.8 \pm 0.5 \mu\text{mol/L}$  and  $K_{ir}3.1/K_{ir}3.4$  ( $K_{KACH}$ ) with an  $IC_{50}$  of  $9.3 \pm 0.4 \mu\text{mol/L}$  (Figure 3, left) and did not produce any significant effects on  $K_v1.5$  ( $I_{Kur}$ ),  $K_v7.1/KCNE1$  ( $I_{Ks}$ ),  $K_v4.3/KChIP2$  ( $I_{to}$ ), and  $K_{ir}2.1$  ( $I_{K1}$ ) in  $30 \mu\text{mol/L}$  or on  $Na_v1.5$  ( $15 \mu\text{mol/L}$ ;  $I_{Na}$ ; Figure 3, middle). Concentrations from 1 to  $10 \mu\text{mol/L}$  AP14145 produced no significant block of  $Ca_v1.2$ , whereas AP14145 at  $30 \mu\text{mol/L}$  changed the baseline significantly more than dimethyl sulfoxide ( $54 \pm 3\%$  versus  $28 \pm 7\%$ ; Figure 3, right).



**Figure 2.** Small-conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channel inhibitors AP14145 (left) and NS8593 (right).

## SK2 and SK3 Are Expressed in Pig Atria

The expression of SK channels in porcine heart has not been previously described. To further substantiate that effects of AP14145 on atrial electrophysiology were mediated by SK channels, we investigated the expression profile of *KCNN1*, *KCNN2*, and *KCNN3* (SK1–3) in the hearts of pigs with sustained AF, as well as controls. *KCNN1* was only found in diminutive amounts in all chambers of both HC and AF pigs (data not shown).

*KCNN2* was expressed in both atria and ventricles (Figure 4, left). The expression levels of *KCNN2* were not found to be significantly affected by tachypacing and the presence of AF in any of the chambers. A higher level ( $\approx 2$ -fold) of *KCNN2* was found in LA compared with LV ( $P=0.005$ ) and in RA compared with right ventricle ( $P=0.002$ ) in HC and AF pigs.

*KCNN3* was expressed in both atria and ventricles (Figure 4, right). The expression levels of *KCNN3* were not found to be significantly affected by tachypacing and the presence of AF in any of the chambers. No statistically significant difference between LA compared with LV ( $P=0.424$ ) or RA compared with right ventricle ( $P=0.077$ ) was observed.

## Pharmacokinetic Results

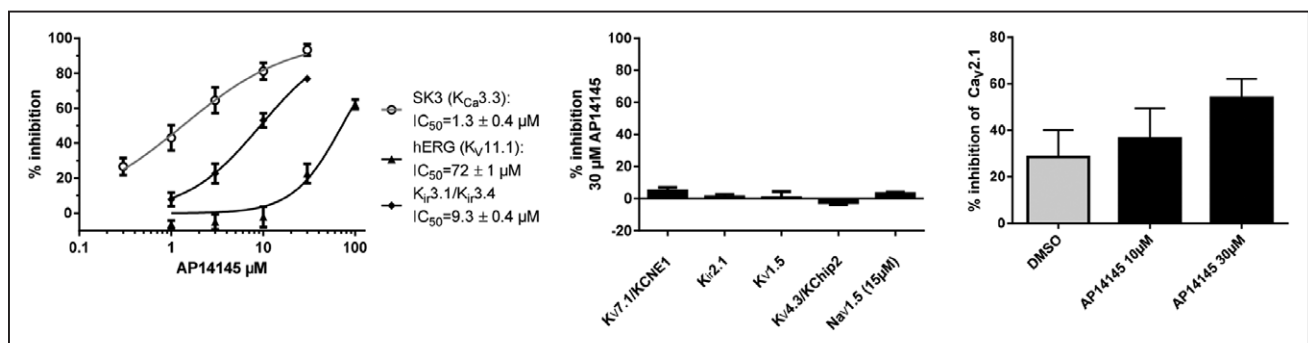
An exponential 1-phase decay model fitted the observations made with regard to plasma concentration of

AP14145 over time with  $R^2=0.97$ ,  $C_{\max}=8355$  nmol/L,  $t_{1/2}=24.3$  minutes, and  $\tau=35$  minutes (Figure 5, left). Using these values and the formula  $C=C_{\max}e^{-t/\tau}$ , as well as a plasma protein binding from minipig plasma of 88%, the free plasma concentrations of AP14145 at any given time during the experiments can be calculated for the dosing regimen used (Figure 5, right). According to these calculations, the maximal free plasma concentrations of AP14145 after each bolus injection were 1.0, 2.0, and 2.5  $\mu\text{mol/L}$  (Figure 5, right), corresponding to 43%, 54%, and 67%  $I_{\text{KCa}}$ -inhibition, assuming Michaelis–Menten kinetics.

## Open Chest Experiments: 1-Week AT and CTRL Pigs

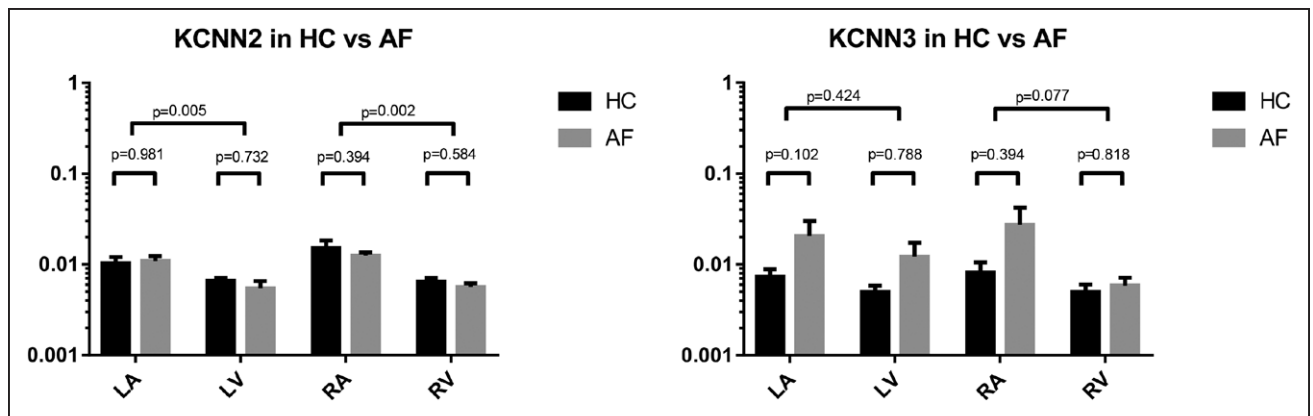
### Effects of 1-Week AT

Neither the LA ERP of  $69\pm 8$  ms nor the LV ERP of  $183\pm 12$  ms in the 1-week AT pigs was significantly different from their CTRL counterparts of  $75\pm 8$  and  $181\pm 6$  ms, respectively ( $P=0.517$  for LA ERP and  $P=0.859$  for LV ERP). However, the 1-week AT gave rise to significantly longer AF durations as compared with the CTRL with mean AF durations of 1130 seconds (822–1553 seconds) and 175 seconds (93–329 seconds), respectively ( $P=0.004$ ). Significantly, more pigs in the 1-week AT group (10 of 12) had AF durations  $>30$  minutes than in the CTRL group (2 of 9;



**Figure 3.** Effect of AP14145 on heterologously expressed ion channels.

**Left,** Concentration response curves of AP14145 effects on hSK3 ( $n=11$ ), hERG ( $n=5$ ), and  $K_{\text{v}3.1}/K_{\text{v}3.4}$  ( $n=6$ ). **Middle,** % Inhibition of  $K_{\text{v}7.1}/\text{KCN}E1$  ( $n=9$ ),  $K_{\text{v}2.1}$  ( $n=5$ ),  $K_{\text{v}1.5}$  ( $n=5$ ), and  $K_{\text{v}4.3}/\text{KChIP}2$  ( $n=5$ ) by 30  $\mu\text{mol/L}$  AP14145 and on  $\text{NaV}1.5$  by 15  $\mu\text{mol/L}$  AP14145 ( $n=3$ ). **Right,** % Inhibition of h $\text{CaV}2.1$  by dimethyl sulfoxide ( $n=3$ ) or AP14145 by 10 and 30  $\mu\text{mol/L}$  ( $n=6$ ). hERG,  $\text{NaV}1.5$ ,  $\text{CaV}1.2$ , and SK3 are measured by automated patch clamping and the remaining by 2-electrode voltage-clamp.



**Figure 4.** Expression of KCNN2 (left) and KCNN3 (right) in tissue from the 4 heart chambers isolated from healthy control (HC) pigs (n=6) and pigs with sustained atrial fibrillation (AF; n=6).

LA indicates left atrial; and LV, left ventricle; RA, right atrial; and RV, right ventricle.

$P=0.009$ ). Despite treatment with digoxin, 1-week AT increased the mean ventricular rate from  $174 \pm 6$  to  $258 \pm 19$  BPM right after initiation of AT ( $P < 0.003$ ), and the mean ventricular rate remained elevated ( $221 \pm 13$  BPM;  $P=0.028$ ) also after long-term AT when vernakalant-resistant AF was present.

#### Effects of Compounds in CTRL Pigs

In the CTRL pigs, both vernakalant and the low dose of AP14145 failed to significantly increase the LA ERP (Table 1; Figure 6, top). The higher doses of AP14145 caused a significant increase of LA ERP. The increase in LA ERP with AP14145 was dose dependent. Concomitant with the increase in LA ERP, the AF duration was decreased by AP14145. Again, the effect of AP14145 on AF duration was dose dependent. Neither vernakalant nor AP14145 changed the LV ERP significantly.

#### Effects of Compounds in 1-Week AT Pigs

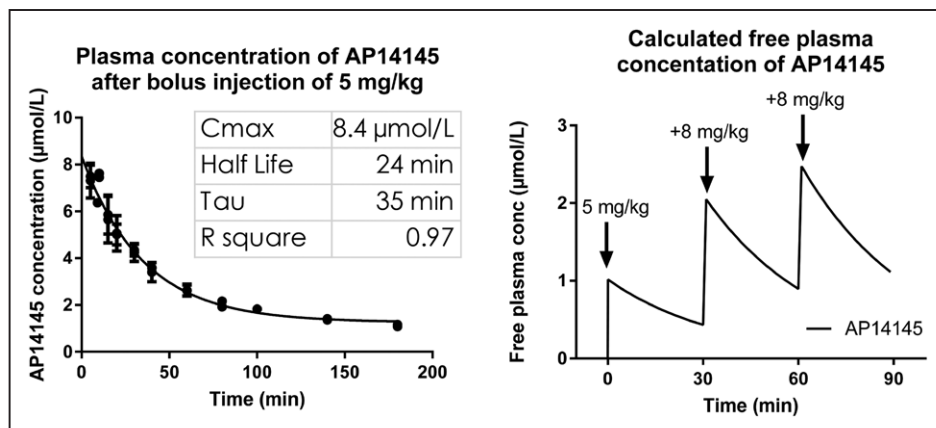
The effects of vernakalant and AP14145 were comparable in both 1-week AT and CTRL groups; both com-

pounds increased LA ERP and decreased AF durations with no significant changes of LV ERP (Table 2; Figure 6, bottom). The effects of AP14145 on LA ERP and AF duration were dose dependent.

In 5 of 6 pigs assigned to receive vernakalant and in 5 of 6 pigs assigned to receive AP14145, the AF that was induced during baseline recording lasted for >30 minutes, and the infusion of the compounds was initiated while AF was still ongoing. Both vernakalant and the lowest dose of AP14145 terminated the AF in 5 of 6 pigs in each 1-week AT group. The time to conversion was  $13.8 \pm 5.5$  minutes in the vernakalant group and  $3.6 \pm 1.8$  minutes in the AP14145 group.

#### Cardioversion in Conscious Pigs

The average time for the development of sustained AF was  $15.6 \pm 1.7$  days of AT (Figure 7). On confirming sustained AF, the pigs received an infusion of 4 mg/kg vernakalant for 10 minutes and were observed for 30 minutes. In 2 of 8 pigs, AF was converted to SR, where-



**Figure 5.** Plasma concentrations of AP14145 in pig.

**Left,** Measured total plasma concentration of AP14145 in pigs after bolus injection of 5 mg/kg AP14145 fitted with an exponential 1-phase decay model. **Right,** Calculated free plasma concentration of AP14145 based on a plasma protein binding of 88% after 3 injections of 5, 8, and 8 mg/kg of AP14145 with 30-min intervals.



**Table 1.** Effects of Vernakalant and AP14145 in CTRL Pigs

	LA ERP, ms	LA ERP (% of Baseline)	AF Duration, s	LV ERP, ms	LV ERP (% of Baseline)
Vernakalant baseline	81±14	100±0	31 (26–39)	203±13	100±0
Vernakalant	130±20	169±27	8 (5–12)	193±9	95±3
AP14145 baseline	59±9	100±0	690 (382–1246)	163±9	100±0
5 mg/kg AP14145	110±14	191±13	9 (5–16)	177±13	108±6
5+8 mg/kg AP14145	140±13	249±24	3 (2–4)	173±17	106±7
5+8+8 mg/kg AP14145	159±15	281±23	2 (1–3)	173±17	106±7

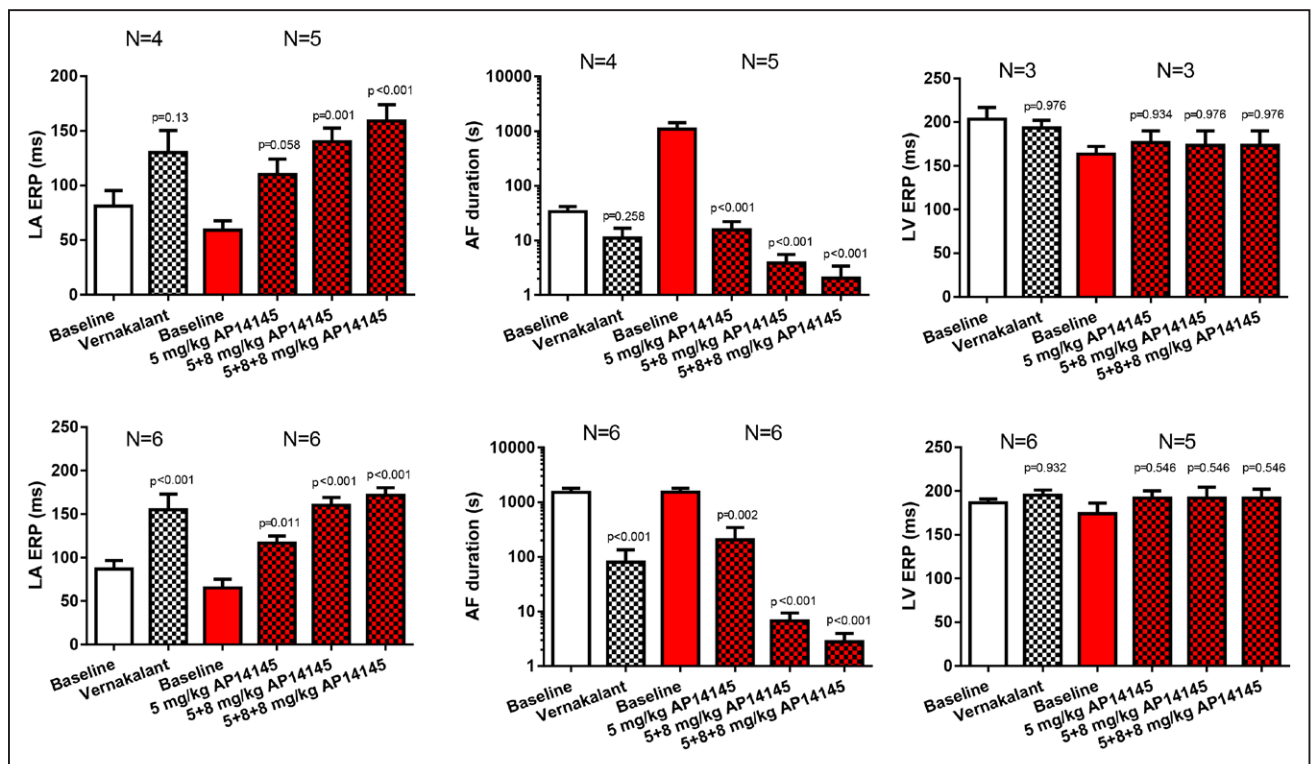
AF indicates atrial fibrillation; CTRL, controls; ERP, effective refractory period; LA, left atrial; and LV, left ventricle.

as AF remained unchanged in 6 of 8 pigs. The 2 pigs with conversion were tachypaced for an additional 1 and 2 weeks, respectively, before they became resistant to vernakalant. On average, vernakalant-resistant AF was obtained after pacing for 18.8±3.3 days. One to 2 days after having observed vernakalant-resistant AF, 8 of 8 pigs still had sustained AF when the pacemaker was turned off. Vernakalant-resistant, sustained AF was cardioverted to normal SR by 5 mg/kg of AP14145 in 4 of the 8 pigs, by an additional 8 mg/kg in 3 of the remaining 4 pigs, and with yet an additional dose of 8 mg/kg AP14145 in the last pig (Figure 8, left). Sustained AF (>10 minutes) could not be reinduced by burst pacing in any of the pigs 2 to 20 minutes after SR was restored. The time to conversion decreased with increasing doses of AP14145, from 16.3±7.9 min-

utes after 5 mg/kg AP14145 to 7.3±2.7 minutes after 8+5 mg/kg AP14145 to 5.0 minutes after 5+8+8 mg/kg AP14145 (Figure 8, right).

## DISCUSSION

We present the first results from a model of vernakalant-resistant AF in conscious pigs showing that SK channel inhibition by AP14145 is capable of both converting vernakalant-resistant AF to SR and protect against reinduction of AF. Complementary molecular, biophysical, and molecular methods were used and showed that (1) SK2 and SK3 channels are present in the porcine atria in both HC and in pigs with vernakalant-resistant AF, (2) AP14145 is a reasonably potent and selective inhibitor of the SK3 current, and (3) SK channel inhibition by



**Figure 6.** Effects of vernakalant and AP14145 on left atrial (LA) effective refractory period (ERP), atrial fibrillation (AF) duration, and left ventricle (LV) ERP.

**Top,** Effects in control pigs. **Bottom,** Effects in 1-wk atrial tachypacing pigs.

**Table 2. Effects of Vernakalant and AP14145 in 1-Week AT Pigs**

	LA ERP, ms	LA ERP (% of Baseline)	AF Duration, s	LV ERP, ms	LV ERP (% of Baseline)
Vernakalant baseline	87±10	100±0	1057 (620–1801)	187±4	100±0
Vernakalant	155±18	184±15	21 (10–46)	195±6	105±3
AP14145 baseline	65±10	100±0	1208 (810–1801)	174±12	100±0
5 mg/kg AP14145	117±8	202±35	44 (20–100)	192±8	112±5
5+8 mg/kg AP14145	160±9	278±48	5 (3–7)	192±12	111±5
5+8+8 mg/kg AP14145	172±9	297±46	1 (1–2)	192±10	111±4

AF indicates atrial fibrillation; AT, atrial tachypacing; ERP, effective refractory period; LA, left atrial; and LV, left ventricle.

AP14145 selectively prolongs the refractory period in porcine LA but not in the LV.

SK channels are predominantly expressed in the atria compared with the ventricles in the normal heart of several species, including humans, with no differences between LA and RA.<sup>11,24–26</sup> Functionally, SK channels play a greater role in the atria compared with the ventricles when using effect criteria, such as ERP prolongation, action potential durations, and QT intervals.<sup>8,11,16</sup> These findings have led to the notion that SK channels might be an interesting target for development of novel treatment for AF where more atrial selective pharmacotherapy has long been sought for. Significant increases of both LA ERP and RA ERP without similar effects in the LV ERP or ventricular repolarization are promising results relating to both the efficacy and safety of the treatment.

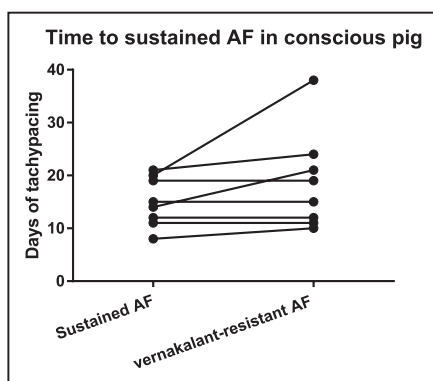
## Expression of Porcine Cardiac SK Channels

We present, for the first time, evidence that SK2 and SK3 but not SK1 subunits are expressed in porcine atria

and ventricles in HC pigs and in pigs with vernakalant-resistant AF. Skibsbjerg et al<sup>16</sup> found that the SK1 subunit was minimally expressed in RA appendage tissue from patients in SR, as well as patients with >6 months in AF, but both the SK2 and SK3 channel subunits were present in SR tissue as well as chronic AF tissue, although to a significantly lower extent in the latter. Considering the SK channel subtypes present in the atria, the pig seems to be more similar to man than, for example, the dog, where all 3 SK channel subtypes are present in the atria.<sup>11</sup> According to the quantitative polymerase chain reaction data from the current study, SK channel mRNA was expressed in slightly higher amounts in both atria compared with the ventricles of both the control pigs and pigs with long-term AT. Although a tendency toward an increased SK3 expression in pigs with long-term AT compared with HC pigs was observed in tissue from LA, RA, and to a lesser extent LV, this apparent upregulation of SK3 was not statistically significant.

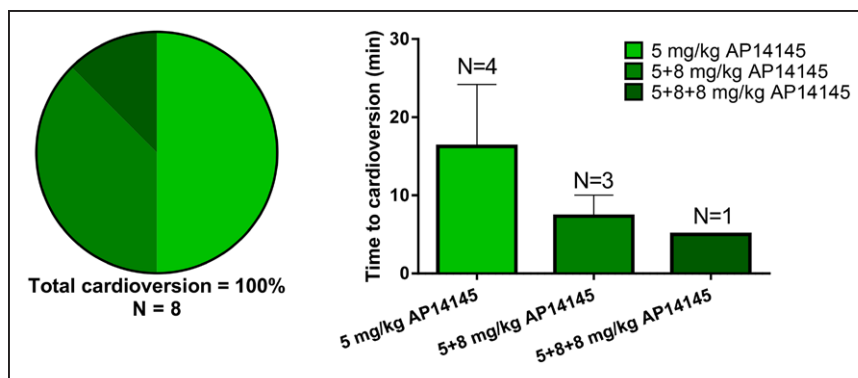
## Effects of AP14145 in Pigs

AP14145 is a new SK channel inhibitor that demonstrated a reasonable selectivity toward cardiac SK channels compared with the other cardiac ion channels tested. The functional data from our open chest experiments suggest an atrial selective effect of AP14145. The LA ERP was concentration-dependently increased almost 3-fold by AP14145 while the LV ERP remained unchanged in both control pigs and pigs that underwent 1 week of AT. One explanation for this atrial selectivity of AP14145 could be differences in expression levels. As described above, SK channel mRNA was expressed in slightly higher amounts in the LA compared with the LV of both CTRL pigs and pigs with vernakalant-resistant AF. Other potentially contributing explanations for this apparent atrial selective effect include Ca<sup>2+</sup> dynamics in ventricles versus atria, location of the channels and neighboring L-type Ca<sup>2+</sup> channels, transportation to the membrane, and redundancy of other repolarizing currents in the ventricles that renders SK current relatively less important in the ventricles. These possibilities were not investigated in the present study.



**Figure 7. Time to sustained atrial fibrillation (AF) in conscious pig.**

Sustained AF was defined as AF that was sustained after the pacemaker was turned off during >10 min monitoring and an additional 10 min with infusion of saline (1 mL/kg) followed by 30 min monitoring, a total of 50 min of AF. Vernakalant-resistant AF was defined as sustained AF where vernakalant (4 mg/kg) infused for 10 min did not cardiovert AF to sinus rhythm during a monitoring phase of an additional 30 min.



**Figure 8. Cardioversion and protection against reinduction of vernakalant-resistant atrial fibrillation (AF) with AP14145.**

**Left**, Increasing doses of AP14145 cardioverts vernakalant-resistant AF and protects against reinduction of AF in 8 of 8 vernakalant-resistant AF pigs. **Right**, Time to cardioversion of AF by increasing doses of AP14145 in vernakalant-resistant AF pigs.

The calculated peak free plasma concentration of AP14145 was  $\approx 1$ ,  $\approx 2$ , and  $\approx 2.5$   $\mu\text{mol/L}$  after each injection, whereas the infusion regimen of vernakalant that was used should produce a steady state free unbound concentration of  $4.8$   $\mu\text{mol/L}$  based on a total plasma concentration of  $4000$   $\text{ng/mL}$ ,<sup>21</sup> a molar weight of  $349.5$   $\text{g/mol}$ , and assuming a plasma protein binding of  $58\%$  based on the human plasma protein binding of  $53\%$  to  $63\%$ .<sup>27</sup> As expected, vernakalant increased the LA ERP and reduced AF duration in the open chest experiments in both CTRL and 1-week AT pigs. In these settings of no or little remodeling, the efficacy of vernakalant ( $4.8$   $\mu\text{mol/L}$  free) and the lowest dose of AP14145 ( $1$   $\mu\text{mol/L}$  free) on AF duration and LA ERP were comparable. The highest dose of AP14145 had more prominent effects on LA ERP and AF duration even though the maximal free plasma concentration of AP14145 was only half of that of vernakalant.

In the long-term AT conscious pigs, vernakalant was administered as an infusion of  $4$   $\text{mg/kg}$  given over 10 minutes. In the clinical phase I study of vernakalant, this dose resulted in a  $C_{\text{max}}$  of  $2.0$  to  $2.2$   $\mu\text{g/mL}$  which corresponds to a free unbound  $C_{\text{max}}$  of  $\approx 3.5$   $\mu\text{mol/L}$  which is slightly more than the maximal plasma concentration of AP14145 that was used. The results of vernakalant and AP14145 in the long-term AT pigs were different. AP14145 converted the vernakalant-resistant AF to SR in all 8 pigs tested. We cannot make firm conclusions on the mechanism behind the vernakalant-resistance, but it is likely not because of reduced expression of its main ion channel targets as we found that the expressions of  $K_{\text{v}1.5}$  and  $\text{Na}_{\text{v}1.5}$  were not affected by AT (data not shown). This is in concordance with other studies suggesting that the loss of cardioversion efficacy of vernakalant is likely not due at decreased effect of vernakalant on its target ionic currents but rather because of other AF stabilizing processes.<sup>28,29</sup>

### AT and Cardiac Remodeling

The atrial selective effect observed in this study might be changed by certain pathophysiological conditions,

such as myocardial infarction and congestive heart failure. Under these conditions, some studies have shown that ventricular SK current can be upregulated and that blocking SK current can exhibit both proarrhythmic<sup>30,31</sup> and antiarrhythmic<sup>32,33</sup> ventricular effects, depending on the experimental setup.

The main focus of this study has been to examine whether SK channel inhibition could be beneficial in a setting of relatively early persistent AF with remodeling to a degree that renders pharmacotherapy for recent onset AF ineffective. Even though all the pigs undergoing AT received digoxin to prevent symptoms of heart failure, there might be a component of ventricular remodeling in our pig model of AF because the ventricular rate was increased during tachypacing. However, no symptoms of heart failure were observed. Also no signs of ventricular proarrhythmia in the presence of AP14145 were observed in any of the tested pigs. Whether SK channel inhibition can have any ventricular effects in pigs under pathophysiological conditions, such as myocardial infarction and congestive heart failure, thus remains an open question.

In patients with permanent (>6 months) AF, the atrial effective refractory period (AERP) is shortened, but in RA tissue from patients with paroxysmal AF, this does not seem to be the case.<sup>34-37</sup> A recent study has revealed a correlation of the degree of left ventricular dysfunction to prolongation of atrial refractoriness, showing that decreased left ventricular function is associated with relative action potential duration prolongation in patients with SR, paroxysmal AF, as well as permanent AF.<sup>37</sup>

In our short-term AT group, 1 week of pacing increased the susceptibility to AF but changed the refractory periods in neither atria nor ventricles compared with CTRL.

The absent change in AERP after short-term AT is different from what has been observed in dogs, goats, and sheep, where short-term tachypacing has been shown to significantly decrease atrial refractoriness.<sup>3,11,38</sup> This discrepancy could be caused by species differences or ventricular remodeling resulting in decreased left ventricular function. In the animal models demonstrating

abbreviated AERP, no signs of decreased left ventricular function were reported. In the dog and sheep models of AF, the atrioventricular node had been ablated, and in goats, AF does not seem to cause ventricular dysfunction.<sup>39</sup> In a dog model of AF, in the presence of congestive heart failure caused by ventricular tachypacing, the decrease in AERP was smaller (24%) than in a model of AF with no congestive heart failure (42%).<sup>40</sup> Likewise, in a pig model of AF with congestive heart failure induced by AT, no reduction of AERP was reported after 7 days of pacing—on the contrary, a significant AERP prolongation was observed.<sup>41</sup> We cannot exclude a component of ventricular dysfunction in our model. Even though the pigs received digoxin to prevent symptoms of heart failure, the ventricular rate was significantly increased by AT, and the unchanged AERP value could therefore be speculated to result from the combination of tachypaced induced shortening of AERP and concomitant prolongation associated with ventricular dysfunction.

### AP14145 and Adverse Events

In the present study, we did not see any significant effects of AP14145 on the heart rate, the blood pressure, the ventricular repolarization (LV ERP), or the QRS interval. In all the conscious long-term AT pigs, adverse effects, such as vomiting, were seen 20 to 30 minutes after injection of AP14145. A dose of zoletil (a mixture of tiletamine hydrochloride and zolazepam hydrochloride) was used to remedy these effects but in effect left the pigs unconscious. Two of the 8 vernakalant-resistant pigs cardioverted on AP14145 before zoletil was injected, but in 6 of 8 pigs zoletil had been injected 2 to 40 minutes before cardioversion was obtained. We cannot exclude that AP14145 had been better tolerated if it had been given as an infusion over longer time.

### Limitations

After the higher doses of AP14145, cardioversion was obtained within 10 minutes in all pigs, but we cannot rule out a possible cardiac impact of zoletil in 6 of the 8 pigs. We cannot exclude that other effects of AP14145 than SK channel inhibition could have influenced our findings. However, AP14145 did not affect a panel of relevant cardiac ion channels (Figure 3) and had a 65- and 8-fold higher  $IC_{50}$  on hERG and  $IK_{ACh}$  as compared with SK3. Moreover, based on the measured free plasma concentrations of AP14145 in our experiments, we do not anticipate a significant contribution from inhibition of hERG or  $IK_{ACh}$ . The small sample sizes for the quantitative polymerase chain reaction experiments and for the open chest CTRL pigs increase the chance of a type II error.

## Summary and Conclusions

In healthy pigs and pigs that had been subjected to 1-week AT, both vernakalant and AP14145 prolonged the AERP and reduced the duration of acutely induced AF while not significantly prolonging the LV ERP or showing other signs of ventricular effects. In an advanced pig model of long-term AT-induced sustained AF where clinically relevant doses of vernakalant could no longer convert the AF to SR, AP14145 was able to convert the pigs to SR and protect against reinduction of AF. Vomiting needing attention occurred after all doses of AP14145 that produced conversion to SR in pigs. The capability of SK channel inhibition to prolong atrial but not ventricular refractoriness and to convert vernakalant-resistant AF, as well as protecting from reinduction of AF, was confirmed in pigs with the tool compound AP14145. SK channel inhibition is an interesting concept and a promising target for development of new treatment of AF.

## AFFILIATIONS

From the Acesion Pharma, Copenhagen, Denmark (J.G.D., R.S.-V., C.C., D.R.P.S., S.H.B., U.S.S., M.G., B.H.B.); Department of Biomedical Sciences, Faculty of Health and Medical Sciences, University of Copenhagen, Denmark (L.S., J.L.S., P.L., D.R.P.S., S.-P.O., T.J., M.G., B.H.B.); and the Heart Centre, Rigshospitalet, Copenhagen, Denmark (J.H.S.).

## ACKNOWLEDGMENTS

We thank Amer Mujezinovic and Lea Abildgaard for expert technical assistance.

## SOURCES OF FUNDING

The study was supported by Innovation Fund Denmark, the Carlsberg Foundation, the Wellcome Trust (award reference no. 100406/Z/12/Z), and the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement no. 675351.

## DISCLOSURES

Drs Diness, Sørensen, Grunnet, and Bentzen are used by and have interests in Acesion Pharma and are inventors of Acesion Pharma patents within the field of SK channels. The other authors report no conflicts.

## FOOTNOTES

Received February 6, 2017; accepted August 21, 2017.

The Data Supplement is available at <http://circep.ahajournals.org/lookup/suppl/doi:10.1161/CIRCEP.117.005125/-/DC1>.

*Circ Arrhythm Electrophysiol* is available at <http://circep.ahajournals.org>.



## REFERENCES

- Roy D, Pratt CM, Torp-Pedersen C, Wyse DG, Toft E, Juul-Møller S, Nielsen T, Rasmussen SL, Stiell IG, Couto B, Ip JH, Pritchett EL, Camm AJ; Atrial Arrhythmia Conversion Trial Investigators. Vernakalant hydrochloride for rapid conversion of atrial fibrillation: a phase 3, randomized, placebo-controlled trial. *Circulation*. 2008;117:1518–1525. doi: 10.1161/CIRCULATIONAHA.107.723866.
- Pratt CM, Roy D, Torp-Pedersen C, Wyse DG, Toft E, Juul-Møller S, Retyk E, Drenning DH; Atrial Arrhythmia Conversion Trial (ACT-III) Investigators. Usefulness of vernakalant hydrochloride injection for rapid conversion of atrial fibrillation. *Am J Cardiol*. 2010;106:1277–1283. doi: 10.1016/j.amjcard.2010.06.054.
- Wijffels MC, Kirchhof CJ, Dorland R, Allesie MA. Atrial fibrillation begets atrial fibrillation. A study in awake chronically instrumented goats. *Circulation*. 1995;92:1954–1968.
- Oral H, Knight BP, Tada H, Ozaydin M, Chugh A, Hassan S, Scharf C, Lai SW, Greenstein R, Pelosi F Jr, Strickberger SA, Morady F. Pulmonary vein isolation for paroxysmal and persistent atrial fibrillation. *Circulation*. 2002;105:1077–1081.
- Eijsbouts S, Ausma J, Blaauw Y, Schotten U, Duytschaever M, Allesie MA. Serial cardioversion by class IC Drugs during 4 months of persistent atrial fibrillation in the goat. *J Cardiovasc Electrophysiol*. 2006;17:648–654. doi: 10.1111/j.1540-8167.2006.00407.x.
- Haissaguerre M, Hocini M, Denis A, Shah AJ, Komatsu Y, Yamashita S, Daly M, Amraoui S, Zellerhoff S, Picat MQ, Quotb A, Jesel L, Lim H, Ploux S, Bordachar P, Attuel G, Meillet V, Ritter P, Derval N, Sacher F, Bernus O, Cochet H, Jais P, Dubois R. Driver domains in persistent atrial fibrillation. *Circulation*. 2014;130:530–538. doi: 10.1161/CIRCULATIONAHA.113.005421.
- Nattel S, Harada M. Atrial remodeling and atrial fibrillation: recent advances and translational perspectives. *J Am Coll Cardiol*. 2014;63:2335–2345. doi: 10.1016/j.jacc.2014.02.555.
- Diness JG, Sørensen US, Nissen JD, Al-Shahib B, Jespersen T, Grunnet M, Hansen RS. Inhibition of small-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels terminates and protects against atrial fibrillation. *Circ Arrhythm Electrophysiol*. 2010;3:380–390. doi: 10.1161/CIRCEP.110.957407.
- Diness JG, Skibsbjerg L, Jespersen T, Bartels ED, Sørensen US, Hansen RS, Grunnet M. Effects on atrial fibrillation in aged hypertensive rats by Ca<sup>2+</sup>-activated K<sup>+</sup> channel inhibition. *Hypertension*. 2011;57:1129–1135. doi:10.1161/HYPERTENSIONAHA.111.170613.
- Skibsbjerg L, Diness JG, Sørensen US, Hansen RS, Grunnet M. The duration of pacing-induced atrial fibrillation is reduced *in vivo* by inhibition of small conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels. *J Cardiovasc Pharmacol*. 2011;57:672–681. doi: 10.1097/FJC.0b013e318217943d.
- Qi XY, Diness JG, Brundel BJ, Zhou XB, Naud P, Wu CT, Huang H, Harada M, Aflaki M, Dobrev D, Grunnet M, Nattel S. Role of small-conductance calcium-activated potassium channels in atrial electrophysiology and fibrillation in the dog. *Circulation*. 2014;129:430–440. doi: 10.1161/CIRCULATIONAHA.113.003019.
- Ellinor PT, Lunetta KL, Glazer NL, Pfeufer A, Alonso A, Chung MK, Sinner MF, de Bakker PI, Mueller M, Lubitz SA, Fox E, Darbar D, Smith NL, Smith JD, Schnabel RB, Soliman EZ, Rice KM, Van Wagoner DR, Beckmann BM, van Noord C, Wang K, Ehret GB, Rotter JJ, Hazen SL, Steinbeck G, Smith AV, Launer LJ, Harris TB, Makino S, Nelis M, Milan DJ, Perz S, Esko T, Köttgen A, Moebus S, Newton-Cheh C, Li M, Möhlenkamp S, Wang TJ, Kao WH, Vasani RS, Nöthen MM, MacRae CA, Stricker BH, Hofman A, Uitterlinden AG, Levy D, Boerwinkle E, Metspalu A, Topol EJ, Chakravarti A, Gudnason V, Psaty BM, Roden DM, Meitinger T, Wichmann HE, Witteman JC, Barnard J, Arking DE, Benjamin EJ, Heckbert SR, Kääb S. Common variants in KCNN3 are associated with lone atrial fibrillation. *Nat Genet*. 2010;42:240–244. doi: 10.1038/ng.537.
- Ellinor PT, Lunetta KL, Albert CM, Glazer NL, Ritchie MD, Smith AV, Arking DE, Müller-Nurasyid M, Krijthe BP, Lubitz SA, Bis JC, Chung MK, Dörr M, Ozaki K, Roberts JD, Smith JG, Pfeufer A, Sinner MF, Lohman K, Ding J, Smith NL, Smith JD, Rienstra M, Rice KM, Van Wagoner DR, Magnani JW, Wakili R, Clauss S, Rotter JJ, Steinbeck G, Launer LJ, Davies RW, Borkovitch M, Harris TB, Lin H, Völker U, Völzke H, Milan DJ, Hofman A, Boerwinkle E, Chen LY, Soliman EZ, Voight BF, Li G, Chakravarti A, Kubo M, Tedrow UB, Rose LM, Ridker PM, Conen D, Tsuruda T, Furukawa T, Sotoodehnia N, Xu S, Kamatani N, Levy D, Nakamura Y, Parvez B, Mahida S, Furie KL, Rosand J, Muhammad R, Psaty BM, Meitinger T, Perz S, Wichmann HE, Witteman JC, Kao WH, Kathiresan S, Roden DM, Uitterlinden AG, Rivadeneira F, McKnight B, Sjögren M, Newman AB, Liu Y, Gollub MH, Melander O, Tanaka T, Stricker BH, Felix SB, Alonso A, Darbar D, Barnard J, Chasman DI, Heckbert SR, Benjamin EJ, Gudnason V, Kääb S. Meta-analysis identifies six new susceptibility loci for atrial fibrillation. *Nat Genet*. 2012;44:670–675. doi: 10.1038/ng.2261.
- Christophersen IE, Rienstra M, Roselli C, Yin X, Geelhoed B, Barnard J, Lin H, Arking DE, Smith AV, Albert CM, Chaffin M, Tucker NR, Li M, Klarin D, Bihlmeyer NA, Low SK, Weeke PE, Müller-Nurasyid M, Smith JG, Brody JA, Niemeijer MN, Dörr M, Trompet S, Huffman J, Gustafsson S, Schurmann C, Kleber ME, Lytikäinen LP, Seppälä I, Malik R, V R Horimoto A, Perez M, Sinisalo J, Aeschbacher S, Thériault S, Yao J, Radmanesh F, Weiss S, Teumer A, Choi SH, Weng LC, Clauss S, Deo R, Rader DJ, Shah SH, Sun A, Hopewell JC, Debette S, Chauhan G, Yang Q, Worrall BB, Paré G, Kamatani Y, Hagemeijer YP, Verweij N, Siland JE, Kubo M, Smith JD, Van Wagoner DR, Bis JC, Perz S, Psaty BM, Ridker PM, Magnani JW, Harris TB, Launer LJ, Shoemaker MB, Padmanabhan S, Haessler J, Bartz TM, Waldenberger M, Lichtner P, Arendt M, Krieger JE, Kähönen M, Risch L, Mansur AJ, Peters A, Smith BH, Lind L, Scott SA, Lu Y, Bottinger EB, Hernessniemi J, Lindgren CM, Wong JA, Huang J, Eskola M, Morris AP, Ford I, Reiner AP, Delgado G, Chen LY, Chen YI, Sandhu RK, Li M, Boerwinkle E, Eisele L, Lannfelt J, Rost N, Anderson CD, Taylor KD, Campbell A, Magnusson PK, Porteous D, Hocking LJ, Vlachopoulou E, Pedersen NL, Nikus K, Orholm-Melander M, Hamsten A, Heeringa J, Denny JC, Kriebel J, Darbar D, Newton-Cheh C, Shaffer C, Macfarlane PW, Heilmann-Heimbach S, Almgren P, Huang PL, Sotoodehnia N, Soliman EZ, Uitterlinden AG, Hofman A, Franco OH, Völker U, Jöckel KH, Sinner MF, Lin HJ, Guo X, Dichgans M, Ingelsson E, Kooperberg C, Melander O, J F Loos R, Laurikka J, Conen D, Rosand J, van der Harst P, Lokki ML, Kathiresan S, Pereira A, Jukema JW, Hayward C, Rotter JJ, März W, Lehtimäki T, Stricker BH, Chung MK, Felix SB, Gudnason V, Alonso A, Roden DM, Kääb S, Chasman DI, Heckbert SR, Benjamin EJ, Tanaka T, Lunetta KL, Lubitz SA, Ellinor PT; METASTROKE Consortium of the ISGC, Neurology Working Group of the CHARGE Consortium; AFGen Consortium. Erratum: large-scale analyses of common and rare variants identify 12 new loci associated with atrial fibrillation. *Nat Genet*. 2017;49:1286. doi: 10.1038/ng0817-1286c.
- Haugaard MM, Hesselkilde EZ, Pehrson S, Carstensen H, Flethøj M, Præstegaard KF, Sørensen US, Diness JG, Grunnet M, Buhl R, Jespersen T. Pharmacologic inhibition of small-conductance calcium-activated potassium (SK) channels by NS8593 reveals atrial antiarrhythmic potential in horses. *Heart Rhythm*. 2015;12:825–835. doi: 10.1016/j.hrthm.2014.12.028.
- Skibsbjerg L, Poulet C, Diness JG, Bentzen BH, Yuan L, Kappert U, et al. Small-conductance calcium-activated potassium (SK) channels contribute to action potential repolarization in human atria. *Cardiovasc Res*. 2014;103:156–167. doi: 10.1093/cvr/cvu121.
- Zhou X, Qun S, Voigt N, Heijman J, Lang S, Borggrefe, Wieland, Nattel S, Dobrev D. ESC 2014 Abstracts. *Eur Heart J*. 2014;35(suppl 1):173–512. doi:10.1093/eurheartj/ehu323.
- Nattel S, De Blasio E, Beatch GN, Wang WQ. RSD1235: a novel antiarrhythmic agent with a unique electrophysiological profile that terminates AF in dogs. *Eur Heart J*. 2001;22:448–448.
- Skarsfeldt MA, Jepps TA, Bomholtz SH, Albigdaard L, Sørensen US, Gregers E, Svendsen JH, Diness JG, Grunnet M, Schmitt N, Olesen SP, Bentzen BH. pH-dependent inhibition of K<sub>v</sub>3.1 prolongs atrial refractoriness in whole hearts. *Pflugers Arch*. 2016;468:643–654. doi: 10.1007/s00424-015-1779-0.
- Skibsbjerg L, Wang X, Axelsen LN, Bomholtz SH, Nielsen MS, Grunnet M, Bentzen BH, Jespersen T. Antiarrhythmic mechanisms of SK channel inhibition in the rat atrium. *J Cardiovasc Pharmacol*. 2015;66:165–176. doi: 10.1097/FJC.0000000000000259.
- Becharad J, Gibson JK, Killingsworth CR, Wheeler JJ, Schneidkraut MJ, Huang J, Ideker RE, McAfee DA. Vernakalant selectively prolongs atrial refractoriness with no effect on ventricular refractoriness or defibrillation threshold in pigs. *J Cardiovasc Pharmacol*. 2011;57:302–307. doi: 10.1097/FJC.0b013e3182073c94.
- Mao ZL, Wheeler JJ, Clohs L, Beatch GN, Keirns J. Pharmacokinetics of novel atrial-selective antiarrhythmic agent vernakalant hydrochloride injection (RSD1235): influence of CYP2D6 expression and other factors. *J Clin Pharmacol*. 2009;49:17–29. doi: 10.1177/0091270008325148.
- Strøbaek D, Hougaard C, Johansen TH, Sørensen US, Nielsen EØ, Nielsen KS, Taylor RD, Pedarzani P, Christophersen P. Inhibitory gating modulation of small conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels by the synthetic compound @-N-(benzimidazol-2-yl)-1,2,3,4-tetrahydro-1-naphthylamine (NS8593) reduces afterhyperpolarizing current in hippocampal CA1 neurons. *Mol Pharmacol*. 2006;70:1771–1782. doi: 10.1124/mol.106.027110.



24. Tuteja D, Xu D, Timofeyev V, Lu L, Sharma D, Zhang Z, Xu Y, Nie L, Vázquez AE, Young JN, Glatzer KA, Chiamvimonvat N. Differential expression of small-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels SK1, SK2, and SK3 in mouse atrial and ventricular myocytes. *Am J Physiol Heart Circ Physiol*. 2005;289:H2714–H2723. doi: 10.1152/ajpheart.00534.2005.
25. Xu Y, Tuteja D, Zhang Z, Xu D, Zhang Y, Rodriguez J, Nie L, Tuxson HR, Young JN, Glatzer KA, Vázquez AE, Yamoah EN, Chiamvimonvat N. Molecular identification and functional roles of a Ca(2+)-activated K+ channel in human and mouse hearts. *J Biol Chem*. 2003;278:49085–49094. doi: 10.1074/jbc.M307508200.
26. Yu T, Deng C, Wu R, Guo H, Zheng S, Yu X, Shan Z, Kuang S, Lin Q. Decreased expression of small-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels SK1 and SK2 in human chronic atrial fibrillation. *Life Sci*. 2012;90:219–227. doi: 10.1016/j.lfs.2011.11.008.
27. Cheng JW, Rybak I. Pharmacotherapy options in atrial fibrillation: focus on vernakalant. *Clin Med Insights Ther*. 2009;1:215.
28. van Hunnik A, Lau DH, Zeemering S, Kuiper M, Verheule S, Schotten U. Antiarrhythmic effect of vernakalant in electrically remodeled goat atria is caused by slowing of conduction and prolongation of postrepolarization refractoriness. *Heart Rhythm*. 2016;13:964–972. doi: 10.1016/j.hrthm.2015.12.009.
29. Wettwer E, Christ T, Endig S, Rozmaritsa N, Matschke K, Lynch JJ, Pourrier M, Gibson JK, Fedida D, Knaut M, Ravens U. The new antiarrhythmic drug vernakalant: ex vivo study of human atrial tissue from sinus rhythm and chronic atrial fibrillation. *Cardiovasc Res*. 2013;98:145–154. doi: 10.1093/cvr/cvt006.
30. Chang PC, Hsieh YC, Hsueh CH, Weiss JN, Lin SF, Chen PS. Apamin induces early afterdepolarizations and torsades de pointes ventricular arrhythmia from failing rabbit ventricles exhibiting secondary rises in intracellular calcium. *Heart Rhythm*. 2013;10:1516–1524. doi: 10.1016/j.hrthm.2013.07.003.
31. Bonilla IM, Long VP 3rd, Vargas-Pinto P, Wright P, Belevych A, Lou Q, Mowrey K, Yoo J, Binkley PF, Fedorov VV, Györke S, Janssen PM, Kilic A, Mohler PJ, Carnes CA. Calcium-activated potassium current modulates ventricular repolarization in chronic heart failure. *PLoS One*. 2014;9:e108824. doi: 10.1371/journal.pone.0108824.
32. Chua SK, Chang PC, Maruyama M, Turker I, Shinohara T, Shen MJ, Chen Z, Shen C, Rubart-von der Lohe M, Lopshire JC, Ogawa M, Weiss JN, Lin SF, Ai T, Chen PS. Small-conductance calcium-activated potassium channel and recurrent ventricular fibrillation in failing rabbit ventricles. *Circ Res*. 2011;108:971–979. doi: 10.1161/CIRCRESAHA.110.238386.
33. Gui L, Bao Z, Jia Y, Qin X, Cheng ZJ, Zhu J, Chen QH. Ventricular tachyarrhythmias in rats with acute myocardial infarction involves activation of small-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels. *Am J Physiol Heart Circ Physiol*. 2013;304:H118–H130. doi: 10.1152/ajpheart.00820.2011.
34. Voigt N, Heijman J, Wang Q, Chiang DY, Li N, Karck M, Wehrens XHT, Nattel S, Dobrev D. Cellular and molecular mechanisms of atrial arrhythmogenesis in patients with paroxysmal atrial fibrillation. *Circulation*. 2014;129:145–156. doi: 10.1161/CIRCULATIONAHA.113.006641.
35. Schmidt C, Wiedmann F, Voigt N, Zhou XB, Heijman J, Lang S, Albert V, Kallenberger S, Ruhparwar A, Szabó G, Kallenbach K, Karck M, Borggreffe M, Biliczki P, Ehrlich JR, Baczkó I, Lugenbiel P, Schweizer PA, Donner BC, Katus HA, Dobrev D, Thomas D. Upregulation of K(2P)3.1 K+ current causes action potential shortening in patients with chronic atrial fibrillation. *Circulation*. 2015;132:82–92. doi: 10.1161/CIRCULATIONAHA.114.012657.
36. Ford J, Milnes J, El Haou S, Wettwer E, Loose S, Matschke K, Tyl B, Round P, Ravens U. The positive frequency-dependent electrophysiological effects of the IKur inhibitor XEN-D0103 are desirable for the treatment of atrial fibrillation. *Heart Rhythm*. 2016;13:555–564. doi: 10.1016/j.hrthm.2015.10.003.
37. Schmidt C, Wiedmann F, Zhou XB, Heijman J, Voigt N, Ratte A, Lang S, Kallenberger SM, Campana C, Weymann A, De Simone R, Szabo G, Ruhparwar A, Kallenbach K, Karck M, Ehrlich JR, Baczkó I, Borggreffe M, Ravens U, Dobrev D, Katus HA, Thomas D. Inverse remodelling of K2P3.1 K+ channel expression and action potential duration in left ventricular dysfunction and atrial fibrillation: implications for patient-specific antiarrhythmic drug therapy. *Eur Heart J*. 2017;38:1764–1774. doi: 10.1093/eurheartj/ehw559.
38. Anné W, Willems R, Holemans P, Beckers F, Roskams T, Lenaerts I, Ector H, Heidbüchel H. Self-terminating AF depends on electrical remodeling while persistent AF depends on additional structural changes in a rapid atrially paced sheep model. *J Mol Cell Cardiol*. 2007;43:148–158. doi: 10.1016/j.yjmcc.2007.05.010.
39. Dosdall DJ, Ranjan R, Higuchi K, Kholmovski E, Angel N, Li L, Macleod R, Norlund L, Olsen A, Davies CJ, Marrouche NF. Chronic atrial fibrillation causes left ventricular dysfunction in dogs but not goats: experience with dogs, goats, and pigs. *Am J Physiol Heart Circ Physiol*. 2013;305:H725–H731. doi: 10.1152/ajpheart.00440.2013.
40. Cha TJ, Ehrlich JR, Zhang L, Nattel S. Atrial ionic remodeling induced by atrial tachycardia in the presence of congestive heart failure. *Circulation*. 2004;110:1520–1526. doi: 10.1161/01.CIR.0000142052.03565.87.
41. Lugenbiel P, Wenz F, Govorov K, Schweizer PA, Katus HA, Thomas D. Atrial fibrillation complicated by heart failure induces distinct remodeling of calcium cycling proteins. *PLoS One*. 2015;10:e0116395. doi: 10.1371/journal.pone.0116395.

## Termination of Vernakalant-Resistant Atrial Fibrillation by Inhibition of Small-Conductance $Ca^{2+}$ -Activated $K^{+}$ Channels in Pigs

Jonas Goldin Diness, Lasse Skibsbye, Rafel Simó-Vicens, Joana Larupa Santos, Pia Lundegaard, Carlotta Citerni, Daniel Rafael Peter Sauter, Sofia Hammami Bomholtz, Jesper Hastrup Svendsen, Søren-Peter Olesen, Ulrik S. Sørensen, Thomas Jespersen, Morten Grunnet and Bo Hjorth Bentzen

*Circ Arrhythm Electrophysiol.* 2017;10:  
doi: 10.1161/CIRCEP.117.005125

*Circulation: Arrhythmia and Electrophysiology* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231

Copyright © 2017 American Heart Association, Inc. All rights reserved.  
Print ISSN: 1941-3149. Online ISSN: 1941-3084

The online version of this article, along with updated information and services, is located on the World Wide Web at:

<http://circep.ahajournals.org/content/10/10/e005125>  
Free via Open Access

Data Supplement (unedited) at:

<http://circep.ahajournals.org/content/suppl/2017/10/09/CIRCEP.117.005125.DC1>

**Permissions:** Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Circulation: Arrhythmia and Electrophysiology* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the [Permissions and Rights Question and Answer](#) document.

**Reprints:** Information about reprints can be found online at:  
<http://www.lww.com/reprints>

**Subscriptions:** Information about subscribing to *Circulation: Arrhythmia and Electrophysiology* is online at:  
<http://circep.ahajournals.org/subscriptions/>

## Supplemental material

### In vitro electrophysiology

#### ***Automated patch clamp experiments***

SK, hERG, NaV1.5 and CaV1.2 experiments were performed using a QPatch 16 HT system and single-hole Qplates (Biolin Scientific, Sophion, Denmark). The Qpatch generates giga sealing, whole-cell formation, compound application and recording of current automatically.

#### ***SK assay***

Automated whole-cell patch-clamping was performed on HEK-293 cells stably expressing hK<sub>Ca</sub>2.3, in symmetrical K<sup>+</sup> solutions, with an extracellular solution consisting of (in mM): KCl 150; CaCl<sub>2</sub> 0.1; MgCl<sub>2</sub> 3; HEPES 10; Glucose 10, pH=7.4 with KOH, and an intracellular solution consisting of (in mM): KCl 108; KOH/EGTA 31.25/10; CaCl<sub>2</sub> 8.1; MgCl<sub>2</sub> 1.2; HEPES 10; KOH 15, pH adjusted to pH=7.2 with HCl. The free calcium concentration was calculated to 400 nM. The cells were clamped at 0 mV and hK<sub>Ca</sub>2.3 currents were continuously elicited every 5th second by a linear voltage ramp from -80 mV to +80 mV (200 ms in duration). The compound application protocol consisted of 9 recording periods lasting from 50-200 s: 1) Baseline recordings in extracellular solution; 2) Application of the positive control methyl-bicuculline (100µM) (Sigma), which is characterized by full efficacy, fast on- and off-rate; 3-4) Wash-out; 5-9) Increasing concentrations of test compound to establish an IC<sub>50</sub> value. Data were sampled at 10 kHz, 4th order Bessel filter, cut-off frequency 3 kHz. Currents were compensated for run-down. Potency was quantified as the concentration needed to inhibit half of the SK channel activity recorded at -80 mV and reported as an IC<sub>50</sub> value. All drug effects were normalized to the observed full inhibitory effect of methyl-bicuculline.

#### ***hERG Assay***

Recordings were performed on CHO-K1 cells stably expressing hKV11.1 with an intracellular solution containing (mM): CaCl<sub>2</sub> 5.4; MgCl 1.75; KOH/EGTA 31.25/10; KCl 120; HEPES 10; Na<sub>2</sub>ATP 4, pH pH=7.2 with KOH, and an extracellular solution (mM): CaCl<sub>2</sub> 2; MgCl<sub>2</sub> 1; NaCl 145; KCl 4, HEPES 10; Glucose 10, pH=7.4 with KOH. The cells were kept in voltage clamp and held at -90 mV. Currents were elicited every 7th 's second by depolarizing the membrane potential to +20 mV for 2 s followed by a 2 s voltage step to -50 mV in order to record tail currents. The application protocol used was: baseline recordings in extracellular solution followed by application of increasing concentration of AP14145 (1, 3, 10, 30, 100 µM). Finally 100µM Dofetilide was added as a positive control. Effect of AP14145 on tail current amplitude was

recorded at baseline and following drug application, normalized to full block by dofetilide and used to establish the  $IC_{50}$ . Data were sampled at 10 kHz, 8th order Bessel filter, cut-off frequency 3 kHz, and 80%  $R_s$  compensation. Only experiments with a whole cell seal of  $> 500 M\Omega$  were used.

### *Nav1.5 assay*

The effect of AP14145 was examined using HEK293 cells stably expressing rNav1.5. Nav1.5 currents were recorded with an intracellular solution that contained (in mM) 135 CsF, 1/5 EGTA/CsOH, 10 HEPES, 10 NaCl, 4 Na-ATP, pH 7.3, 300 mOsm and an extracellular solution containing (in mM): 2  $CaCl_2$ , 1  $MgCl_2$ , 10 HEPES, 4 KCl, 145 NaCl, 10 glucose, pH 7.2, 310 mOsm (adjusted with sucrose). rNav1.5 currents were elicited every 1000 milliseconds by a 50 ms depolarizing step to -20 mV from a holding potential of -120 mV (a total of 80 pulses). The application protocol used was baseline recordings in standard extracellular solution followed by application of AP14145 (15  $\mu$ M). The current amplitude at the 80<sup>th</sup> pulse was used for analysis. Data were sampled at 25 kHz, eighth-order Bessel filter, cutoff frequency 3 kHz. Only experiments with a whole-cell seal of  $>500 M[\text{ohm}]$  were used.

### *Calcium assay*

Effect of AP14145 on L-type calcium channels was investigated by automated patch clamping (Qpatch, Sophion) using a hCaV 1.2 stable cell line (Alpha1C, Beta2C and Alpha2delta) (SB Drug Discovery). The extracellular solution contained (in mM): 145 NaCl, 10  $BaCl_2$ , 4 KCl, 10 HEPES and 10 Glucose (pH 7.4). The intracellular solution contained (in mM) 27 CsF, 112 CsCl, 2 NaCl, 8.2 EGTA, 10 HEPES and 4 MgATP; (pH 7.25). The stimulus regime consisted of an initial 50 ms pulse at -120 mV, followed by a 300 ms test pulse at 0mV, and then returned back to -120 mV for another 50 ms. Four cumulative concentrations of compound was applied to the cell (1, 3, 10, 30 (in  $\mu$ M)), followed by the positive control, Nifedipine (10  $\mu$ M).

### **Two-electrode voltage-clamp**

cRNA for injection was prepared according to from linearized plasmids using the mMESSAGE mMACHINE T7 kit (Ambion, TX, USA). RNA concentrations and quality were assessed by UV spectroscopy (NanoDrop, Thermo Scientific,

Wilmington, USA) and gel electrophoresis. The following concentrations of cRNA were used ( $\mu\text{g}/\mu\text{L}$ ):  $\text{K}_v7.1+\text{KCNE1}$  0.08 + 0.02;  $\text{K}_{ir}2.1$  0.4;  $\text{K}_v1.5$  0,1;  $\text{K}_v4.3+\text{KChIP2}$  0.01+0.01;  $\text{K}_{ir}3.1+\text{K}_{ir}3.4$  0.04+0.04. 50 nL cRNA was injected in *Xenopus laevis* oocytes (EcoCyte Bioscience, Castrop-Rauxel, Germany), and currents were recorded after 2-3 days of incubation ( $19^\circ\text{C}$ ) using a two-electrode voltage-clamp amplifier (Dagan CA-1B; IL, USA). Borosilicate glass recording electrodes (0.5–1  $\text{M}\Omega$  when filled with 2 M KCl) (Module Ohm, Herlev, Denmark) were pulled on a DMZ-Universal Puller (Zeitz Instruments, Martinsried, Germany). Oocytes were superfused with Kulori solution (in mM: NaCl 90, KCl 4,  $\text{MgCl}_2$  1,  $\text{CaCl}_2$  1, 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid 5,  $\text{pH}=7.4$  with NaOH, room temperature). After stabilization the solution was changed for a solution containing AP14145 (30 $\mu\text{M}$ ). For Kir3.1/kir3.4 four concentrations of AP14145 were tested (1, 3, 10, 30  $\mu\text{M}$ ) in order to establish an  $\text{IC}_{50}$  value.

Oocytes were voltage clamped at -80 mV.  $\text{K}_v7.1/\text{KCNE1}$  currents were elicited by depolarizing the membrane potential to +20 mV for 5 s followed by a 1 s step to -30 mV.  $\text{K}_v1.5$  and  $\text{K}_v4.3/\text{KChIP2}$  currents were evoked by changing the membrane potential to +20 mV or 0 mV for 2 s respectively. Steady state current amplitudes ( $\text{K}_v7.1/\text{KCNE1}$  and  $\text{K}_v1.5$ ) and peak current amplitudes ( $\text{K}_v4.3/\text{KChIP2}$ ) were used for analysis. Kir3.1/Kir3.4 and Kir2.1 currents were elicited by a ramp protocol from -100 mV to +80 mV (5 s duration). Current amplitudes at -100 mV were used for analysis.

Data acquisition was performed with the Pulse software (HEKA Elektronik, Lambrecht/Pfalz, Germany).

## KCNNx expression in pig hearts

### Tissue samples

After premedication with zoletil pig mixture (250 mg dry tiletamin+zolazepam, 6.5 ml xylazine 20 mg/ml, 1.25 ketamine 100 mg/ml, 2.5 ml butorphanol 10 mg/ml, and 2 ml methadone 10 mg/ml) 1 ml/10 kg given IM, the pigs were euthanized by intravenous injection of pentobarbital 200 mg/ml. The hearts were immediately excised and placed in ice-cold cardioplegic solution (NaCl 110.0 mM, KCl 16.0 mM,  $\text{MgCl}_2$  16.0 mM,  $\text{CaCl}_2$  1.2 mM,  $\text{NaHCO}_3$  10.0 mM). Cardiac tissue from each of the four chambers was obtained from long-term AT pigs (n=6) or controls (n=6) and rapidly snap-frozen in liquid nitrogen. Tissue was kept at  $-80^\circ\text{C}$  for later RNA extraction.



Approximately 40mg of atrial and ventricular tissue specimens were lysed in QIAzol reagent (QIAGEN, Maryland, USA) using a Precellys 24 (Bertin Technologies, Montigny-le-Bretonneux, France). Total RNA including small RNAs was purified according to the manufacturer's instruction using the miRNeasy Mini kit (QIAGEN, Hilden, Germany). RNA samples were treated with DNase. The RNA concentration and purity was determined by spectrophotometry (NanoDrop2000, Thermo Scientific, Wilmington, USA) using the absorbance ratio of A260/A280.

### **Expression profiling**

Reverse transcription (RT) reactions were performed following the manufacturer's instructions using the Precision nanoScript2 Reverse Transcription kit (PrimerDesign Ltd., Southampton, UK). Each reaction had a final volume of 20  $\mu$ L containing 1 $\mu$ g of total RNA. The experiment included a minus reverse transcriptase control (-RT) to check for genomic contamination and other amplification artifacts. The reverse transcription steps were conducted in a PTC-200 Peltier Thermal Cycler (Struers KEBO Lab, Albertslund, DK) as follows: 25°C for 5 minutes, 42°C for 20 minutes and 75°C for 10 minutes. To assess the levels of mRNAs from the genes KCNN1, KCNN2 and KCNN3 in the samples, quantitative real-time polymerase chain reaction (qPCR) was carried out in duplicates using Custom real-time PCR assays with double dye probe (Taqman style) and PrecisionPLUS MasterMix with ROX (PrimerDesign Ltd., Southampton, UK) according to manufacturer's instruction. *GAPDH* and *GPI* were used as reference genes for qPCR normalization and no template controls (NTCs) were run simultaneously to assess contamination. The qPCR steps were conducted with Precision BrightWhite real-time PCR 96-well plates (PrimerDesign Ltd., Southampton, UK) on a CFX Connect Real-Time System (BIO-RAD, Hertfordshire, UK) as follows: 95°C for 2min followed by 40 cycles of 95°C for 15 seconds and 60°C for 1 minute with FAM dye fluorescence read at the end of each cycle. Threshold cycle ( $C_t$ ) values were obtained using Bio-Rad CFX96 Managed 3.0 software and assuming a single threshold mode. The data was transferred to a spreadsheet for calculation of  $\Delta C_t$ s and the relative expression of the genes in both groups was calculated using the  $2^{-\Delta C_t}$  method.

## Primer sequences

Gene	Forward primer	Reverse primer	Product length (bp)	Accession number
<i>KCNN1</i>	5'- CTCCTGGATCATTGCTGCCT -3'	5'- TTGCTGGTCACTTCCTGCTT -3'	81	XM_005654898
<i>KCNN2</i>	5'- CGCCGCTTCGTCCTCAG -3'	5'- CCGCCTCCTCCAGTGCT -3'	111	XM_005661582
<i>KCNN3</i>	5'- ATCAGTCTGTCCACCATCATCC -3'	5'- CTCGTAGGTCATGGCTATCCG -3'	117	NM_213985

Reference Genes	Species	Accession number	Anchor Nucleotide	Product length (bp)
<i>GAPDH</i>	Sus scrofa	AF017079	918	188
<i>GPI</i>	Sus scrofa	NM_214330	685	105