Invasive American bullfrogs and African clawed frogs in South America high suitability of occurrence in biodiversity hotspots

Barbosa, Fabiana G.; Both, Camila; Bastos, Miguel

Published in: Zoological Studies

DOI: 10.6620/ZS.2017.56-28

Publication date: 2017

Document version Publisher's PDF, also known as Version of record

Document license: Unspecified

Citation for published version (APA): Barbosa, F. G., Both, C., & Bastos, M. (2017). Invasive American bullfrogs and African clawed frogs in South America: high suitability of occurrence in biodiversity hotspots. *Zoological Studies*, *56*, [28]. https://doi.org/10.6620/ZS.2017.56-28



Open Access

Invasive American bullfrogs and African Clawed Frogs in South America: High Suitability of Occurrence in Biodiversity Hotspots

Fabiana G. Barbosa^{1,*}, Camila Both², and Miguel B. Araújo^{1,3}

¹Departmento de Biogeografía y Cambio Global, Museo Nacional de Ciencias Naturales, Consejo Superior de Investigaciones Cientificas (CSIC), Calle José Gutiérrez Abascal, 2, 28006, Madrid, Spain

- ²Centro de Ciências Naturais e Exatas, Universidade Federal de Santa Maria, Avenida Roraima, 1000, 97105-9000, Santa Maria, RS, Brazil
- ³Center for Macroecology, Evolution and Climate, Natural History Museum of Denmark, University of Copenhagen, Copenhagen, Denmark

(Received 14 June 2017; Accepted 10 September 2017; Communicated by Benny K.K. Chan)

Fabiana G. Barbosa, Camila Both, and Miguel B. Araújo (2017) Invasion of protected areas by non-native species is currently one of the main threats to global biodiversity. Using an ensemble of bioclimatic envelope models we quantify the degree of exposure of South American protected areas to invasion by two invasive amphibian species. We focus on protected areas that coincide with global biodiversity hotspots. The species modeled, *Lithobates catesbeianus* and *Xenopus laevis*, have been reported to threaten local faunas in several non-native areas that they invaded, including areas in Asia, Europe, North America, and South America. We show that 87.5% of the protected areas within the Atlantic Forest may be most at risk of invasion by *L. catesbeianus* and *X. laevis* under current climate conditions, followed by areas in the Cerrado (51.7), Tropical Andes (37.6%), Tumbes-Choco-Magdalena (22.5%), and Chilean Winter Rainfall and Valdivian Forests (20.5%). Conservation plans for these regions should, therefore, consider latent threats from multiple sources including invasion by highly competitive non-native species such as the ones modeled in our study.

Key words: Biological invasions, Bioclimatic envelope models, *Lithobates catesbeianus*, Protected areas, *Xenopus laevis*.

BACKGROUND

Protected areas (henceforth PAs) are one of the cornerstones of spatial conservation planning for biodiversity (Margules and Pressey 2000). While the total coverage of PAs has been increasing worldwide (Jenkins and Joppa 2009). PAs are facing increasing pressures such as unsustainable levels of tourism, financial shortfalls, poaching and other extractive activities, deforestation and habitat fragmentation, encroachment by urbanization and other human activities, and invasion by non-native invasive species (Ervin 2003).

Invasions by non-native amphibian species have been reported in several parts

of the world (Kraus 2009). In South America, two invasive amphibian species have received particular attention from researchers: *Lithobates catesbeianus* (Shaw 1802) and *Xenopus laevis* (Daudin 1802) (e.g., Lobos and Jaksic 2005; Laufer et al. 2008; Lobos et al. 2013). *Lithobates catesbeianus*, known as the American bullfrog, is native to eastern North America and its global distribution includes 41 countries (Ervin 2003; Frost 2013). In South America, *L. catesbeianus* is present in ten countries (Ervin 2003; Frost 2013) with populations established in a variety of environments (e.g., Laufer et al. 2008; Lobos et al. 2013; Ferreira and Lima 2012), including PAs (Lucas and Fortes 2008; Schüttler and Karez

^{*}Correspondence: E-mail: fabibarbos@gmail.com

2008; Both et al. 2011; Madalozzo et al. 2016). Invasive populations of L. catesbeianus in new environments have caused substantial damage to native fauna through competition (e.g., Kupferberg 1997; Pearl et al. 2004), predation (e.g., Silva et al. 2011; Boelter et al. 2012), or acoustic interference (e.g., Both and Grant 2012). For these reasons, L. catesbeianus has been listed among the "100 worst invasive species" by the International Union for Conservation of Nature (IUCN) (Lowe et al. 2000). Xenopus laevis, known as the African clawed frog, is native to Mediterranean zone of the Cape Region of South Africa (see Lobos et al. 2013 for details), but has invaded extensive areas on several continents (see Measey et al. 2012 for review). In South America, X. laevis is present in natural and disturbed environments in Chile (Lobos and Jaksic 2005; Lobos et al. 2013), but to our knowledge there are no records in PAs. Damage documented for X. laevis includes consumption of native prey (Lafferty and Page 1997) and reducing reproductive rates of native amphibians (Lillo et al. 2011). Lithobates catesbeianus and X. laevis are both recognized as vectors of the fungus Batrachochytrium dendrobatidis, which is lethal to many other amphibians and has been linked with amphibian declines (e.g., Garner et al. 2006; Solis et al. 2010).

Eradication of *L. catesbeianus* and *X. laevis* is difficult once populations are established in nonnative ranges (Fouquet and Measey 2006; Adams and Pearl 2007). Preventive action is therefore required to prevent or slow the rate of spread of these invasive species in PAs. Of particular interest is the identification of areas where both species are likely to invade, as these are areas where the effects of these two species could be additive.

In the last decades, bioclimatic envelope models (henceforth BEMs) have become one of the main tools for predicting the distribution of suitable climatic areas for the establishment of invasive species (Jiménez-Valverde et al. 2011; Barbosa et al. 2012). Such models are built using modelling methods that establish correlations between the geographic distribution of a species and a set of predictor variables (typically climatic) to identify conditions that are suitable for maintaining viable populations (see Araújo and Peterson 2012 for review on BEMs). Currently, multiple methods are now available for modelling bioclimatic (see Peterson et al. 2011 for review). Many studies have demonstrated that different methods calibrated using the same dataset can provide different predictions (e.g., Elith et al.

2006; Tsoar et al. 2007), raising the possibility that no single "best" method exists (Pearson et al. 2006). To handle such algorithmic uncertainties, Araújo and New (2007) advocate the fitting of ensembles of BEMs. In some instances, BEMs used to predict the distribution of suitable climatic areas of invasive species were fitted using data from the native range of the invasive species (e.g., Giovanelli et al. 2008), implicitly assuming that the present distribution of the species is at equilibrium with the current climate (Araújo and Pearson 2005). However, some models using data from the native ranges of invasive species have been able to accurately predict areas of introduction, but not necessarily the total area of invasion (e.g., Broennimann et al. 2007; Barbosa et al. 2013). This failure to predict the extent of invasions has been attributed to a low degree of equilibrium with climate in their native range (Peterson 2011). To reduce such biases in estimates of a species' potential distribution it has been suggested that, whenever possible, data from the entire range (native and invaded) should be used when building models (Broennimann and Guisan 2008; Beaumont et al. 2009a).

In this context, we quantified the degree of exposure of South American PAs to invasion by L. catesbeianus and X. laevis under current climate conditions. We focused specifically on PAs that coincide with biodiversity hotspots in South America (Atlantic Forest, Tropical Andes, Chilean Winter Rainfall and Valdivian Forests, and Tumbes-Choco-Magdalena), as they are likely to be of greater importance for conservation. First, for each invasive species, we built an ensemble of BEMs using data from both its native and invaded ranges in order to estimate a broad range of climatically suitable areas for establishment of the species in South America. Second, we combined these predictions to build a map that shows the areas climatically suitable for the establishment of the two invasive species in South America. Third, we overlaid the PAs with the map of climatic suitability for the two invasive species modeled in order to assess the degree of exposure of PAs to invasion. Finally, we calculated the potential surface of each biodiversity hotspot in South America that may be most at risk of invasion by both species.

MATERIALS AND METHODS

Species data and bioclimatic variables

We obtained worldwide native and invasive presence records for *L. catesbeianus* and *X. laevis* from the Global Biodiversity Information Facility online database (GBIF; http://data.gbif.org) and from other published sources (see Supplementary 1). A total of 2,140 presence records were compiled for *L. catesbeianus*, 1,263 from within its native range (eastern North America) and 877 from invaded areas in Asia, Europe, western North America. For *X. laevis*, 175 records were obtained, 93 from its native range (Mediterranean zone of the Cape Region of South Africa) and 82 from invaded areas in Asia, Europe, North America, and South America.

We extracted 19 current bioclimatic variables from the WorldClim database (Hijmans et al. 2005; see http://www.worldclim.org for description of the variables) with a spatial resolution of 10 arcminutes. We then selected six bioclimatic variables: (i) mean temperature diurnal range (mean of the monthly difference of the maximum and minimum temperatures), (ii) temperature seasonality, (iii) mean temperature of wettest quarter, (iv) annual precipitation, (v) precipitation of driest month, and (vi) precipitation seasonality. These bioclimatic variables were selected based on the basis of studies carried with the two invasive species (Nori et al. 2011a; Loyola et al. 2012, Lobos et al. 2013), excluding highly correlated variables (pairwise r_{Pearson} < 0.75; Braunish et al. 2013) to reduce multicollinearity and subsequent model over-fitting (Jiménez-Valverde et al. 2011).

We built two equal-area grids (0.16×0.16) degrees of latitude/longitude). The first grid covered the study area for L. catesbeianus (the full extent of its native range in eastern North America and in invaded areas), and the second covered the study area for X. laevis (in its native range in Mediterranean zone of the Cape Region of South Africa as well as in invaded areas). We then mapped all the presence records of each species into its study area to generate a grid cell matrix. We also resampled the values of each bioclimatic variable in each grid cell. The grids were generated using the software Spatial Analysis in Macroecology (SAM v.4.0; Rangel et al. 2010). All analyses below were performed using the data obtained from these grid cell matrices.

Bioclimatic envelope modelling

For each species, we built an ensemble of the BEM (Araújo and New 2007) using four presenceonly methods. Models were fitted using the **BIOENSEMBLES** software for computer-intensive modelling of species potential distributions (Diniz-Filho et al. 2009). BIOENSEMBLES, userfriendly software, is an integrated framework for ensemble forecasting of species distributions that has modeling methods with different strategies of adjustment and type of input data (Terribile et al. 2012), such as presence-absence records methods (e.g., Generalized Linear Model), presence-background methods (e.g., Maxent), presence-pseudoabsence methods (e.g., GARP), and presence-only methods (e.g., BIOCLIM) (Peterson et al. 2011). Herein, the methods used were BIOCLIM (e.g., Busby 1991), Euclidean Distance (e.g., Farber and Kadmon 2003), Gower Distance (e.g., Carpenter et al. 1993), and Mahalanobis Distance (e.g., Farber and Kadmon 2003). Presence-only methods were chosen because absence records do not always imply a lack of climatic suitability (see Bradley 2009 for more details).

We obtained BEMs as follows. First, for each method, BEMs were built using a calibration subset of 70% of all the presence records (i.e., native and invasive range) selected at random and then evaluated with the remaining 30%; this process was repeated 50 times. We use 63 possible combinations $(2^n - 1, where "n")$ is the number of variables) of the six bioclimatic variables (Diniz-Filho et al. 2009; Collevatti et al. 2013). Thus, for each species and modeling method used, our modeling procedure yielded 3150 BEMs (63 combinations of bioclimatic variables × 50 cross-validation). Then continuous predictions of the models were converted into a binary vector of 0/1, indicating forecasted presences and absences in each grid cell using a cut-off point. True Skill Statistics (TSS) (Allouche et al. 2006), varying between -1 and 1, was used to evaluate the performance of the models. Finally, all models outputs with TSS > 0.5 were combined (an average weighted by the TSS value of each model) to generate a map of the frequency of BEM supporting the occurrence of species in each grid cell (Diniz-Filho et al. 2009; Collevatti et al. 2013). Herein we use these frequencies of occurrence of a species as a proxy of climatic suitability for the establishment of the invasive species in each grid cell.

We then built a map of climatic suitability for each species using the outputs of the BEMs, interpreted as an index of the potential for establishment for the species in South America. We then overlaid the maps of species to obtain a combined estimate of the potential for establishment of the two invasive species in South America. All maps were built using the software ArcGIS 10 (ESRI 2010).

Protected areas

We identified PAs in South America that may be most at risk of invasion by L. catesbeianus and X. laevis under current climate conditions, focusing on PAs that coincide with global biodiversity hotspots. Firstly, we downloaded the polygons of the biodiversity hotspots in South America from Conservation International (http:// www.conservation.org/) as well as the polygons of the PAs from the World Database on Protected Areas (http://www.protectedplanet.net/). Second, we filtered the PAs to retain only those PAs that overlapped with biodiversity hotspots in South America. This resulted in 593 PAs within biodiversity hotspots in South America. Finally, we overlaid the PAs within biodiversity hotspots that coincided with the map of modeled climatic suitability for the two invasive species in order to assess the degree of exposure of PAs to risk of invasion. In addition, we also calculated the potential surface of each biodiversity hotspot in South America that may be most at risk of invasion by both species.

All spatial analyses were performed using the software ArcGIS 10. We only considered strict PAs (PA categories I – IV, International Union for Conservation of Nature (IUCN)) (UNEP-WCMC 2012). We used five classes of risk of invasion according to the climatic suitability of the total area in the PA (0 to 20, 20 to 40, 40 to 60, 60 to 80, and 80 to 100%), which we designate as presenting no, low, medium, high, and critical invasion risk, respectively.

RESULTS

Current climatically suitable areas coincided strongly with areas in which the invasive species have already been recorded in South America (Fig. 1a and 1b). Furthermore, low climatic suitability for L. catesbeianus was predicted in central eastern parts of Brazil, southeastern and northwestern portions of Argentina, northern Chile, southwestern Bolivia, the coast of Peru, the eastern portion of Colombia, and the southern and northern portions of Venezuela (Fig. 1a). For X. laevis, low climatic suitability was predicted in major portions of the northern and central eastern parts of the continent, southwest Bolivia, a southern portion of Peru, the northern and southern portions of Chile, and the northwest, southern and central western portions of Argentina (Fig. 1b). In parallel the combination of the current climatically suitable areas of both species (Fig. 1a and 1b) showed a large overlap in the potential distribution of *L. catesbeianus* and *X.* laevis in South America (Fig. 1c).

According to the models, up to 87.5% of the PAs within the Atlantic Forest may be at risk of invasion by *L. catesbeianus* and *X. laevis* under current climate conditions, followed by the Cerrado (51.7%), Tropical Andes (37.6%), Tumbes-Choco-Magdalena (22.4%), and Chilean Winter Rainfall and Valdivian Forests (20.5%) (Table 1). Additionally, 74.72% of the surface of the Atlantic

Table 1. Percentage of Protected Areas suitable (*i.e.*, Percentage by number of Protected Areas suitable) within each biodiversity hotspot in South America that may be most at each class of risk from invasions of both non-native invasive species (*Lithobates catesbeianus* and *Xenopus laevis*) under current climate conditions

Biodiversity hotspot	Class of risk				
	Critically 1 - 0.8	High 0.8 - 0.6	Medium 0.6 - 0.4	Low 0.4 -0.2	None 0.2 - 0
Atlantic Forest	87.5	0.0	0.0	0.0	12.5
Cerrado	51.7	0.0	2.5	0.0	45.8
Chilean Winter Rainfall and Valdivian Forests	20.5	0.0	2.6	10.2	66.7
Tropical Andes	37.6	1.5	3.8	0.7	30.8
Tumbes-Choco-Magdalena	22.5	0.0	2.8	15.4	59.3



Fig. 1. Map of climatic suitability for the establishment of *Lithobates catesbeianus* (a), *Xenopus laevis* (b), and of both species (c) in South America. Back dots indicate presence records of the species and blue are Protected Areas within biodiversity hotspots in South America.

Forest was projected as a suitable climatic area for the establishment of both non-native invasive amphibian species, followed by the Cerrado (28.04%), Tropical Andes (21.23%), Chilean Winter Rainfall and Valdivian Forests (15.15%), and Tumbes-Choco-Magdalena (1.64%) (Fig. 2).

DISCUSSION

Prediction in the early stages, with subsequent prevention, is the most effective management strategy for invasive species, and is often the most efficient too (Richardson and Thuiller 2007). Bioclimatic envelope models are one potentially useful tool to predict climatically suitable areas for the establishment of invasive species in non-native areas (Jiménez-Valverde et al. 2011), as well as in PAs (e.g., Beaumont et al. 2009b; Vicente et al. 2013). For instance, in the case of L. catesbeianus, two studies modeled the potential distribution of the species in PAs in South America. Nori et al. (2011b) modeled the potential distribution of L. catesbeianus in PAs (six IUCN categories) in South America, concluding that over 30% of the surface of all the IUCN PAs in South America are climatically suitable for establishment of the species under current conditions. On the other hand, Loyola et al. (2012) modeled the potential distribution of L. catesbeianus in PAs



Fig. 2. Percentage of suitable surface within each biodiversity hotspot in South America that may be most at risk from invasions of both non-native invasive species (*Lithobates catesbeianus* and *Xenopus laevis*) under current climate conditions. AF = Atlantic Forest, TA = Tropical Andes, CWRVF = Chilean Winter Rainfall and Valdivian Forests, TCM = Tumbes-Choco-Magdalena.

(IUCN categories I - IV with PAs > 5000 hectares) in the Brazilian Atlantic Forest and showed that several PAs are suitable for establishment of the species under current conditions. In contrast, to our knowledge no studies have quantified the degree of exposure of PAs to invasion by X. laevis in South America using BEMs. Here, we quantified, for the first time, the percentage of suitable PAs within each biodiversity hotspot in South America that may be most at risk of invasion by L. catesbeianus and X. laevis (Fig. 2a) under current climate conditions. Our results show that several PAs within biodiversity hotspots in South America may be at risk of invasion by L. catesbeianus and X. laevis under current climate conditions (Table 1). Although there were differences in the percentage of suitable PAs among biodiversity hotspots, our results indicate that all biodiversity hotspots in South America have PAs that may be at risk of invasion by both species (Fig. 2) (also see Supplementary - figure A1, which provides results for each invasive species).

The biodiversity of amphibian species peaks in tropical regions, especially in biodiversity hotspots in South America (Myers et al. 2000), making this a critical region for amphibian conservation. However, a large amount of the native habitat in biodiversity hotspots in South America has been transformed into numerous smaller isolated patches (e.g., Silva and Bates 2002; Armenteras et al. 2003; Ribeiro et al. 2009; Sarkar et al. 2009) and, consequently, many current PAs are increasingly isolated, especially PAs within the Atlantic Forest and the Cerrado (e.g., Silva and Bates 2002; Ribeiro et al. 2009), which in turn make the PAs more susceptible to invasion. Additionally, amphibians are thought to be the most threatened vertebrate group and many authors consider that they are declining globally (e.g., Stuart et al. 2004). Invasive species have been suggested as a possible extrinsic cause of amphibian decline (Kats and Ferrer 2003) together with climate change, changes in land use and disease (Hof et al. 2011). In invaded areas, L. catesbeianus and X. laevis have caused substantial ecological damage to native organisms, especially to native species of amphibian (e.g., Kraus 2009; Lillo et al. 2011). In the case of L. catesbeianus. the decline of some native amphibian species has been associated with its establishment worldwide (e.g., Kupferberg 1997); but see also Pearl et al. 2004 and Kats and Ferrer 2003), although evidence that the species is negatively affecting local amphibian communities in South America is still lacking (Both et al. 2014). In addition, *L. catesbeianus* and *X. laevis* are recognized as vectors of the fungus *Batrachochytrium dendrobatidis* (e.g., Solis et al. 2010; Schloegel et al. 2010), the agent responsible for chytridiomycosis, a disease considered one of the main causes of decline and extinction in amphibian populations worldwide (Daszak et al. 2003; Skerratt et al. 2007). Therefore, the presence of *L. catesbeianus* and *X. laevis* in South American PAs is dangerous.

A range of preventive measures for L. catesbeianus and X. laevis in South America has been recommended in previous studies (Giovanelli et al. 2008; Nori et al. 2011a, b; Urbina-Cardona et al. 2011; Loyola et al. 2012; Lobos et al. 2013). We agree with these studies and reinforce the urgency in implementing effective prevention programs in South American PAs. Another issue to consider is that L. catesbeianus and X. laevis have already established populations in some South American hotspots (Both et al. 2011; Sanabria et al. 2011), including PAs in the case of L. catesbeianus (Lucas and Fortes 2008; Schüttler and Karez 2008; Both et al. 2011). Populations of L. catesbeianus and X. laevis are difficult to eradicate once established (Fouquet and Measey 2006; Adams and Pearl 2007) and, according to Leung et al. (2002), eradication programs are less cost-effective than prevention programs. However, several methods have been proposed to eradicate and/or control L. catesbeianus in North America (see Louette et al. 2012 for some proposed methods). Additionally, for both invasive species, authors have recommended interventions to reduce the permanency of lentic systems, as an eradication and/or control measure (e.g., Fuller et al. 2011; Lobos et al. 2013; Peterson et al. 2013). Water permanency is an environmental gradient well known to structure lentic communities (see Wellborn et al. 1996). Thus, such measures should be carefully and particularly considered, since they can have more negative impacts upon native fauna than the invasive species itself. Therefore, in parallel with prevention programs. we also recommend strict control of activities that represent high-risk pathways for accidental and/or intentional introduction of these species in natural environments (e.g., frog farms and the pet trade).

CONCLUSIONS

In summary, our study is the first to quantify the degree of exposure of South American PAs to invasion by *L. catesbeianus* and *X. laevis*, and such information provides a valuable baseline for implementing management programs. Moreover, our results showed a large overlap in the potential distribution of *L. catesbeianus* and *X. laevis* in South America (Fig. 1c). Therefore, we suggest that future studies should assess the potential impact of co-occurrence of these invasive species upon native fauna (*e.g.*, Araújo et al. 2011). Knowledge about the effects of interactions among competing species could provide additional support to decision making on eradication and/or control and prevention priorities.

Acknowledgments: We thank Dora Neto for the support in Geographic Information Systems. This work was supported by the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) (FGB, grant number 246048/2012-3).

Authors' contributions: FGB and MBA designed the study. FGB analyzed the data. FGB, CB, and MBA wrote the manuscript. All authors participated in revising the manuscript. All authors read and approved the final manuscript.

Competing interests: FGB, MBA, and CB declare that they have no conflict of interest. FGB has received research grants from the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) (grant number 246048/2012-3).

Availability of data and materials: Additional data is provided on supplementary files. Other data and materials will be provided by the corresponding author on request.

Consent for publication: Not applicable.

Ethics approval consent to participate: Not applicable.

REFERENCES

Adams MJ, Pearl CA. 2007. Problems and opportunities managing invasive bullfrogs. Is there any hope? *In*: Gherardi F (ed) Biological invaders in inland waters profiles, distribution and threats, Springer, Dordrecht, pp.

page 8 of 10

679-693.

- Allouche O, Tsoar A, Kadmon R. 2006. Assessing the accuracy of species distribution models, prevalence, kappa and the true skill statistic (TSS). J Appl Ecol **43:**1223-1232.
- Araújo MB, New M. 2007. Ensemble forecasting of species distributions. Trends Ecol Evol 22:42-47.
- Araújo MB, Pearson RG. 2005. Equilibrium of species' distributions with climate. Ecography 28:693-695.
- Araújo MB, Peterson AT. 2012. Uses and misuses of bioclimatic envelope modeling. Ecology 93:1527-1539.
- Araújo MB, Rozenfeld A, Rahbek C, Marquet PA. 2011. Using species co-occurrence networks to assess the impacts of climate change. Ecography 34:897-908.
- Armenteras D, Gast F, Villareal H. 2003. Andean forest fragmentation and the representativeness of protected natural areas in the eastern Andes, Colombia. Biol Conserv **113**:245-256.
- Barbosa FG, Pillar DV, Palmer AR, Melo AS. 2013. Predicting the current distribution and potential spread of the exotic grass *Eragrostis plana* Nees in South America and identifying a bioclimatic niche shift during invasion. Austral Ecol 38:260-267.
- Barbosa FG, Schneck F, Melo AS. 2012. Use of ecological niche models to predict the distribution of invasive species: a scientometric analysis. Braz J Biol **72**:821-829.
- Beaumont LJ, Gallagher RV, Downey PO, Thuiller W, Leishman MR, Hughes L. 2009b. Modelling the impact of *Hieracium* spp. on protected areas in Australia under future climates. Ecography **32**:757-764.
- Beaumont LJ, Gallagher RV, Thuiller W, Downey PO, Leishman MR, Hughes L. 2009a. Different climatic envelopes among invasive populations may lead to underestimations of current and future biological invasions. Divers Distrib 5:409-420.
- Boelter RA, Kaefer IL, Both C, Cechin S. 2012. Invasive bullfrogs as predators in a Neotropical assemblage: what frog species do they eat? Anim Biol **62:**397-408.
- Both C, Grant T. 2012. Biological invasions and the acoustic niche: the effect of bullfrog calls on the acoustic signals of white-banded tree frogs. Biol Lett **8**:714-716.
- Both C, Lingnau R, Santos-Jr A, Madalazzo B, Lima LP, Grant T. 2011. Widespread occurrence of the American bullfrog, *Lithobates catesbeianus* (Shaw, 1802) (Anura: Ranidae), in Brazil. South Am J Herp **6**:127-134.
- Both C, Madalozzo M, Lingnau R, Grant T. 2014. Amphibian richness patterns in Atlantic Forest areas invaded by American bullfrogs. Austral Ecol **39**:864-878.
- Bradley BA. 2009. Regional analysis of the impacts of climate change on cheatgrass invasion shows potential risk and opportunity. Glob Chang Biol **15**:196-208.
- Braunish V, Coppes J, Arlettaz R, Suchant R, Schmid H, Bollmann K. 2013. Selecting from correlated climate variables: a major source of uncertainty for predicting species distributions under climate change. Ecography 36:971-983.
- Broennimann O, Guisan A. 2008. Predicting current and future biological invasions: both native and invaded ranges matter. Biol Lett **4**:585-589.
- Broennimann O, Treier UA, Müller-Schärer H, Thuiller W, Peterson AT, Guisan A. 2007. Evidence of climatic niche shift during biological invasion. Ecol Lett **10**:701-709.
- Busby JR. 1991. BIOCLIM a bioclimate analysis and prediction system. Plant Prot Q 6:8-9.
- Carpenter G, Gillison AN, Winter J. 1993. DOMAIN: a flexible

modelling procedure for mapping potential distributions of plants and animals. Biodivers Conserv **2**:667-680.

- Collevatti RG, Terribile LC, de Oliveira G, Lima-Ribeiro MS, Nabout JC, Rangel TF, Diniz-Filho JAF. 2013. Drawbacks to palaeodistribution modelling: the case of South American seasonally dry forests. J Biogeogr **40**:345-358.
- Daszak P, Cunningham AA, Hyatt AD. 2003. Infectious disease and amphibian population declines. Divers Distrib **9**:141-150.
- Diniz-Filho JAF, Bini LM, Rangel TF, Loyola RD, Hof C, Nogués-Bravo D, Araújo M B. 2009. Partitioning and mapping uncertainties in ensembles of forecasts of species turnover under climate change. Ecography **32**:897-906.
- Elith J, Graham CH, Anderson RP, Dudík M, Ferrier S, Guisan A, Hijmans RJ, Huettmann F, Leathwick JR, Lehmann A, Li J, Lohmann LG, Loiselle BA, Manion G, Moritz C, Nakamura M, Nakazawa Y, Overton JM, Peterson AT, Phillips SJ, Richards K, Scachetti-Pereira R, Schapire RE, Soberón J, Williams S, Wisz MS, Zimmermann NE. 2006. Novel methods improve prediction of species' distributions from occurrence data. Ecography **29**:129-151
- Ervin J. 2003. Rapid assessment of protected area management effectiveness in four countries. BioScience 53:833-841.
- ESRI. 2010. ESRI ArcGis 9.3. Environmental Systems Research Institute Inc.
- Farber O, Kadmon R. 2003. Assessment of alternative approaches for bioclimatic modeling with special emphasis on the Mahalanobis distance. Ecol Modell **160:**115-130.
- Ferreira RB, Lima CS. 2012. Anuran hotspot at Brazilian Atlantic rainforest invaded by the non-native *Lithobates catesbeianus* Sahw, 1802 (Anura: Ranidae). N West J Zool **8:**386-389.
- Fouquet A, Measey GJ. 2006. Plotting the course of an African clawed frog invasion in Western France. Anim Biol **56**:95-102.
- Frost DR. 2013. Amphibian species of the world: an online reference. Version 6.0. American Museum of Natural History, New York, USA. http://research.amnh.org/ herpetology/amphibia/index.html. Accessed 29 April 2013.
- Fuller TE, Pope KL, Ashton DT, Welsh Jr HH. 2011. Linking the distribution of an invasive amphibian (*Rana catesbeiana*) to habitat conditions in a managed river system in northern California. Restor Ecol **19**:204-213.
- Garner TWJ, Perkins MW, Govindarajulu P, Seglie D, Walker S, Cunningham AA, Fisher MC. 2006. The emerging amphibian pathogen *Batrachochytrium dendrobatidis* globally infects introduced populations of the North American bullfrog, *Rana catesbeiana*. Biol Lett 2:455-459.
- Giovanelli JGR, Haddad CFB, Alexandrino J. 2008. Predicting the potential distribution of the alien invasive American bullfrog (*Lithobates catesbeianus*) in Brazil). Biol Invasions **10**:585-590.
- Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A. 2005. Very high resolution interpolated climate surfaces for global land areas. Int J Climatol **25**:1965-1978.
- Hof C, Araújo MB, Jetz W, Rahbek C. 2011. Additive threats from pathogens, climate and land-use change for global amphibian diversity. Nature **480**:516-519.
- Jenkins CN, Joppa L. 2009. Expansion of the global terrestrial protected area system. Biol Conserv **142**:2166-2174.
- Jiménez-Valverde A, Peterson AT, Soberón J, Overton JM, Aragón P, Lobo JM. 2011. Use of niche models in invasive

species risk assessments. Biol Invasions 13:2785-2797.

- Kats LB, Ferrer RP. 2003. Alien predators and amphibian declines: review of two decades of science and the transition to conservation. Divers Distrib **9**:99-110.
- Kraus F. 2009. Alien reptiles and amphibians: a scientific compendium and analysis. Springer Series in Invasion Ecology vol.4. Springer Science & Business Media B.V.
- Kupferberg SJ. 1997. Bullfrog (*Rana catesbeiana*) invasion of a California river: the role of larval competition. Ecology 78:1736-1751.
- Lafferty KD, Page CJ. 1997. Predation on the endangered tidewater goby, *Eucyclobius newberryi*, by the introduced African clawed frog, *Xenopus laevis*, with notes on the frog's parasite. Copeia **1**:589-592.
- Laufer G, Canavero A, Nuñez D, Maneyro R. 2008. Bullfrog (*Lithobates catesbeianus*) invasion in Uruguay. Biol Invasions **10:**1183-1189.
- Leung B, Lodge DM, Finnoff D, Shogren JF, Lewis MA, Lamberti G. 2002. An ounce of prevention or a pound of cure: bioeconomic risk analysis of invasive species. P Roy Soc B-Biol Sci **9**:2407-2413.
- Lillo F, Faraone FP, Valvo ML. 2011. Can the introduction of *Xenopus laevis* affect native amphibian populations? Reduction of reproductive occurrence in presence of the invasive species. Biol Invasions **13**:1533-1541.
- Lobos G, Cattan P, Estades C, Jaksic FM. 2013. Invasive African clawed frog *Xenopus laevis* in southern South America: key factors and predictions. Stud Neotrop Fauna E **48:**1-12.
- Lobos G, Jaksic FM. 2005. The ongoing invasion of African clawed frogs (*Xenopus laevis*) in Chile: causes of concern. Biodivers Conserv **14**:429-439.
- Lowe S, Browne M, Boudjelas S, De Poorter M. 2000. 100 of the Word's worst invasive alien species: a selection from the Global Invasive Species Database. IUCN/SSC Invasive Species Specialist Group, Auckland.
- Louette G, Devisscher S, Adriaens T. 2012. Control of invasive American bullfrog *Lithobathes catesbeianus* in small shallow water bodies. Eur J Wildlife Res **9:**105-114.
- Loyola RD, Nabout JC, Trindade-Filho J, Lemes P, Urbina-Cardona JN, Dobrovolski R, Sagnori MD, Diniz-Filho JAF. 2012. Climate change might drive species into reserves: a case study of the American bullfrog in the Atlantic Forest Biodiversity Hotspot. Alytes **29**:61-74.
- Lucas EM, Fortes VB. 2008. Frog diversity in the Floresta Nacional de Chapecó, Atlantic Forest of Southern Brazil. Biota Neotrop **8**:51-61.
- Madalozzo B, Both C, Cechin S. 2016. Can protected areas with agricultural edges avoid invasions? The case of bullfrogs in the Southern Atlantic Rainforest in Brazil. Zool Stud **55**:51. doi:10.6620/ZS.2016.55-51.
- Margules CR, Pressey RL. 2000. Systematic conservation planning. Nature **405**:243-253.
- Measey GJ, Rodder D, Green SL, Kobayashi R, Lillo F, Lobos G, Rebelo R, Thirion JM. 2012. Ongoing invasions of the African clawed frog, *Xenopus laevis*: a global review. Biol Invasions **14**:2255-2270.
- Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GAB, Kent J. 2000. Biodiversity hotspots for conservation priorities. Nature **403**:853-858.
- Nori J, Akmentins MS, Ghirardi R, Frutos N, Leynaud GC. 2011b. American bullfrog invasion in Argentina: where should we take urgent measures? Biodivers Conserv **20**:1125-1132.

- Nori J, Urbina-Cardona JN, Loyola RD, Lescano JN, Leynaud GC. 2011a. Climate change and American bullfrog invasion: what could we expect in South America? PLoS ONE 6:e25718.
- Pearl CA, Adams MJ, Bury RB, McCreary B. 2004. Asymmetrical effects of introduced bullfrogs (*Rana catesbeiana*) on native ranid frogs in Oregon. Copeia 1:11-20.
- Pearson RG, Thuiller W, Araújo MB, Martinez-Meyer E, Brotons L, McClean C, Miles L, Segurado P, Dawson TP, Lees DC. 2006. Model-based uncertainty in species range prediction. J Biogeogr 33:1704-1711.
- Peterson AC, Richgels KLD, Johnson PTJ, McKenzie VJ. 2013. Investigating the dispersal routes used by an invasive amphibian, *Lithobates catesbeianus*, in human-dominated lanscapes. Biol Invasions **15**:2179-2191.
- Peterson AT. 2011. Ecological niche conservatism: a timestructured review of evidence. J Biogeogr **38**:817-827.
- Peterson AT, Soberón J, Pearson RG, Anderson RP, Martinez-Meyer E, Nakamura M, Araújo MB. 2011. Ecological Niches and Geographic Distributions. Princeton University Press.
- Rangel TF, Diniz-Filho JAF, Bini LM. 2010. SAM: a comprehensive application for spatial analysis in macroecology. Ecography **33**:46-50.
- Ribeiro CM, Metzger JP, Martensen AC, Ponzoni FJ, Hirota MM. 2009. The Brazilian Atlantic Forest: how much is left, and how is the remaining forest distributed? Implications for conservation. Biol Conserv **142**:1141-1153.
- Richardson DM, Thuiller W. 2007. Home away from home objective of high-risk source areas for plant introductions. Divers Distrib 13:299-312.
- Sanabria EA, Ripoll Y, Jordan M, Quiroga L, Ariza M, Guillemain M, Pérez M, Chávez H. 2011. A new record for American Bullfrog (*Lithobates catesbeianus*) in San Juan, Argentina. Revista Mexicana de Biodiversidad 82:311-313.
- Sarkar S, Sánchez-Cordero V, Londoño MC, Fuller T. 2009. Systematic conservation assessment for the Mesoamerica, Chocó, and Tropical Andes biodiversity hotspots: a preliminary analysis. Biodivers Conserv 18:1793-1828.
- Silva JM, Bates JM. 2002. Biogeographic patterns and conservation in the South American Cerrado: a tropical savanna hotspot. Bioscience **52**:225-233.
- Silva ET, Ribeiro Filho OP, Feio RN. 2011. Predation of native anurans by invasive bullfrogs in southeastern Brazil: spatial variation and effect of microhabitat use by prey. S Am J Herpetol **6**:1-10.
- Schloegel LM, Ferreira CM, James TY, Hipolito M, Longcore JE, Hyatt D, Yabsley M, Martins AMCRPF, Mazzoni R, Davies AJ, Daszak P. 2010. The North American bullfrog as a reservoir for the spread of *Batrachochytrium dendrobatidis* in Brazil. Anim Conserv **13**:53-61.
- Schüttler E, Karez CS. 2008. Especies exóticas invasoras en las Reservas de Biosfera de América Latina y el Caribe. Un informe técnico para fomentar el intercambio de experiencias entre las Reservas de Biosfera y promover el manejo efectivo de las invasions biológicas. UNESCO, Montevideo, Uruguay.
- Skerratt LF, Berger L, Speare R, Cashins S, McDonald KR, Phillott AD, Hines HB, Kenyon N. 2007. Spread of chytridiomycosis has caused the rapid global decline and extinction of frogs. EcoHealth 4:125-134.
- Solis R, Lobos G, Walker SF, Fisher M, Bosch J. 2010.

Presence of *Batrachochytrium dendrobatidis* in feral populations of *Xenopus laevis* in Chile. Biol Invasions **12**:1641-1646.

- Stuart SN, Chanson JS, Cox NA, Young BE, Rodrigues AS, Fischman DL, Waller RW. 2004. Status and trends of amphibian declines and extinctions worldwide. Science **306**:1783-1786.
- Terribile LC, Lima-Ribeiro MS, Araújo MB, Bizão N, Collevatti RG, Dobrovolski R, Franco AA, Guilhaumon F, Lima JS, Murakami DM, Nabout JC, de Oliveira G, Oliveira LK, Rabelo SG, Rangel TF, Simon LM, Soares TN, Telles MPC, Diniz-Filho JAF. 2012. Areas of climate stability of species ranges in the Brazilian Cerrado: disentangling uncertainties through time. Natureza & Conservação 10:152-159.
- Tsoar A, Allouche O, Steinitz O, Rotem D, Kadmon R. 2007. A comparative evaluation of presence-only methods for modelling species distribution. Divers Distrib **13**:397-405.
- UNEP-WCMC. 2012. Data Standards for the World Database on Protected Areas. UNEP-WCMC.
- Urbina-Cardona JN, Nori J, Castro F. 2011. Áreas vulnerables a la invasión actual y futura de la rana toro (*Lithobates catesbeianus*) em Colombia: estratégias propuestas para su manejo y control. Biota Colomboniana **12:**23-34.
- Vicente JR, Fernandes RF, Randin CF, Broennimann O, Gonçalves J, Marcos B, Pôças I, Alves P, Guisan A, Honrado JP. 2013. Will climate change drive alien invasive plants into areas of high protection value? An improved model-based regional assessment to prioritize the management of invasions. J Environ Manage **131**:185-195.
- Wellborn GA, Skelly DK, Werner EE. 1996. Mechanisms creating community structure across a freshwater habitat gradient. Annu Rev Ecol Evol Syst **27**:337-363.

Supplementary 1. Sources of presence records for *L. catesbeianus* and *X. laevis*. (download)

Supplementary - Fig. A1. Percentage of suitable surface within each biodiversity hotspots in South America that may be most at risk from invasions of *Lithobates catesbeianus* and *Xenopus laevis* under current climate conditions. AF = Atlantic Forest, TA = Tropical Andes, CWRVF = Chilean Winter Rainfall and Valdivian Forests, TCM = Tumbes-Choco-Magdalena. (download)